

RESEARCH ARTICLE

Effects of urbanization on host-pathogen interactions, using *Yersinia* in house sparrows as a model

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Abstract

Urbanization strongly affects biodiversity, altering natural communities and often leading to a reduced species richness. Yet, despite its increasingly recognized importance, how urbanization impacts on the health of individual animals, wildlife populations and on disease ecology remains poorly understood. To test whether, and how, urbanization-driven ecosystem alterations influence pathogen dynamics and avian health, we use house sparrows (*Passer domesticus*) and *Yersinia* spp. (pathogenic for passerines) as a case study. Sparrows are granivorous urban exploiters, whose western European populations have declined over the past decades, especially in highly urbanized areas. We sampled 329 house sparrows originating from 36 populations along an urbanization gradient across Flanders (Belgium), and used isolation combined with ‘matrix-assisted laser desorption ionization- time of flight mass spectrometry’ (MALDI-TOF MS) and PCR methods for detecting the presence of different *Yersinia* species. *Yersinia* spp. were recovered from 57.43% of the sampled house sparrows, of which 4.06%, 53.30% and 69.54% were identified as *Y. pseudotuberculosis*, *Y. enterocolitica* and other *Yersinia* species, respectively. Presence of *Yersinia* was related to the degree of urbanization, average daily temperatures and the community of granivorous birds present at sparrow capture locations. Body condition of suburban house sparrows was found to be higher compared to urban and rural house sparrows, but no relationships between sparrows’ body condition and presence of *Yersinia* spp. were found. We conclude that two determinants of pathogen infection dynamics, body condition and pathogen occurrence, vary along an urbanization gradient, potentially mediating the impact of urbanization on avian health.

Introduction

With growing human populations, cities are expanding rapidly and urbanization represents one of the most intense anthropogenic modifications of natural systems, strongly affecting species, communities and ecosystems [1,2]. The direction and strength of responses of bird species to urbanization is function of their life-history strategies [3]. This has led to the 'biotic homogenization' of urban bird communities [4], i.e. whereby the latter become gradually dominated by a limited number of 'urban exploiter' species, such as house sparrows (*Passer domesticus*) [5]. Studies focussing on how avian communities respond to urbanization find that bird species richness [4,6,7] and population densities [6] are often highest at intermediate levels of urbanization. However, although several authors have addressed the effects of urbanization on avian stress levels and body condition (e.g. [8–11]), how individuals of urban exploiters successfully cope with urban environments, remains poorly understood.

How urbanization affects disease ecology, wildlife-pathogen interactions and animal health remains particularly underexplored, despite its potential effect on ecological and evolutionary mechanisms driving population dynamics [12–16]. In addition, wildlife is increasingly being recognized as an important vector, or potentially even reservoir, for various human diseases [17], such as yersiniosis, the third most commonly reported bacterial zoonotic disease in Europe in 2013 [18]. In humans, yersiniosis is most frequently caused by *Yersinia enterocolitica* biotype (BT) 1B and 2–5 and to a lesser extent by *Y. pseudotuberculosis* [18,19]. In passerines, the facultative pathogen *Y. pseudotuberculosis* is the most probable etiologic agent of yersiniosis, which typically has an acute enteric disease progression [20–22], but has on several occasions been isolated from apparently healthy birds [23,24]. Although it is possible that these birds were in the incubation phase of the disease, it has been speculated that wild-ranging birds maintain the bacteria at low level, developing acute disease when subjected to stressful conditions [24]. Yet, the potential existence of subclinical effects on avian health and body condition remains a gap in our knowledge.

So far only few studies have focused on the combination of differential pathogen exposure along urbanization gradients and the effects on the body condition of their avian hosts (e.g. [15,25,26]). With respect to *Yersinia*, their psychrotolerant nature [27] potentially renders these bacteria susceptible to microclimate differences (e.g. heat island effect) between urbanized and rural areas [28]. In addition, the distinct metabolic flexibility of various *Yersinia* species [29] may affect environmental survival and persistence, enhancing the survival of the less pathogenic environmental strains with higher metabolic capacity compared to the more pathogenic strains which are metabolically more constrained and are more dependent on the presence of suitable hosts. Depending on the pathogen-suitability of the hosts, higher host diversity or density may both reduce or amplify the bacterium-load in the environment [12], and hence, the faeco-oral transmission of pathogenic *Yersinia* species. Not only can *Yersinia* affect birds' health, but vice versa, avian health, related to stress and estimated by body condition [30], could affect the excretion of pathogens in the environment [31,32].

In order to gain more insights into urban wildlife-disease ecology, we assessed the prevalence of an important zoonotic and avian pathogen (i.e. *Yersinia* spp.) in house sparrows along an urbanization gradient. House sparrows constitute an adequate study species as they inhabit rural, suburban and urban areas, they are considered to be very sedentary, and they have experienced severe population declines over the last decades, especially in urban centres [33–36]. We evaluated how urbanization and the local community of granivorous birds impact on house sparrows' body condition and on the presence of *Yersinia* spp. in their faeces, in combination with the two-way host-pathogen interaction, taking into account temperature and time of sparrow capture during sampling.

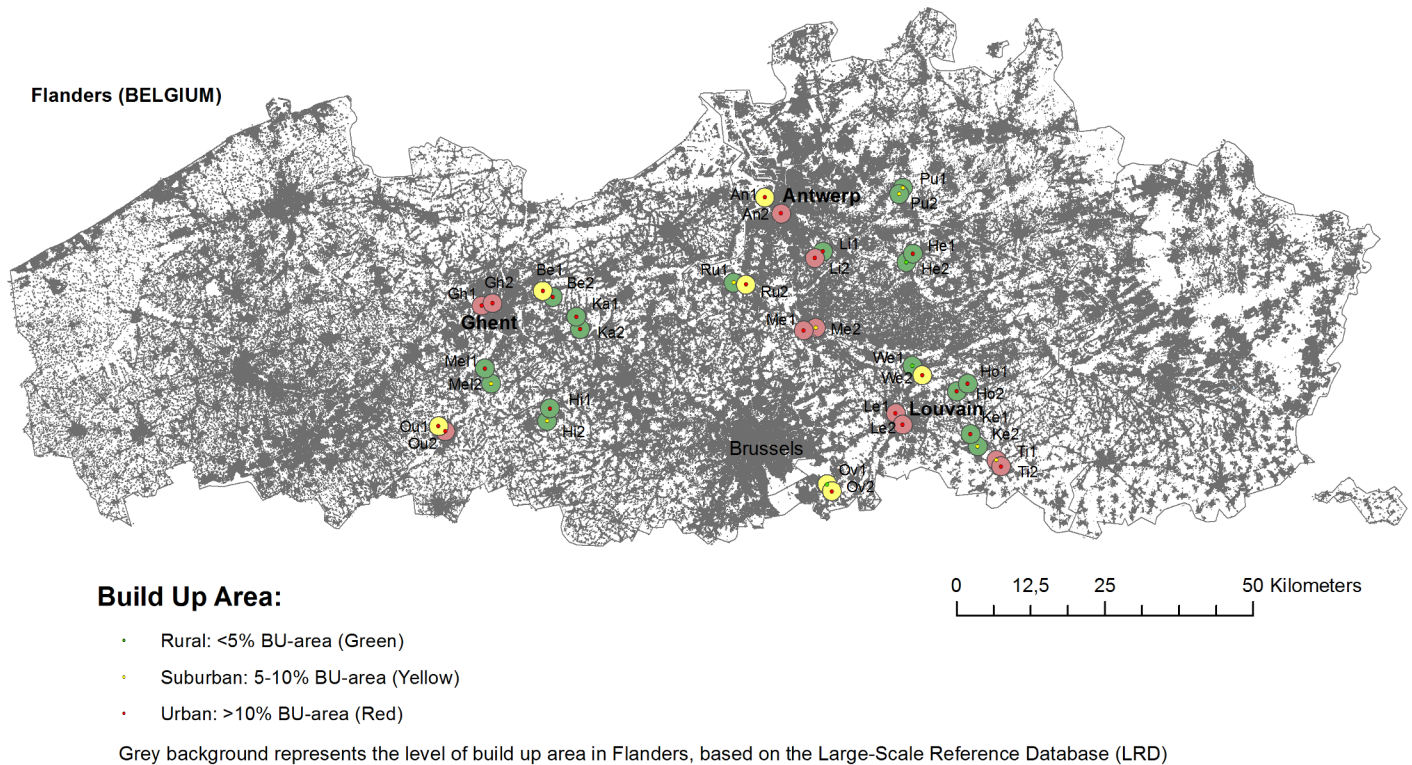


Fig 1. House sparrow populations clustered around the cities of Ghent, Antwerp and Leuven.

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Material and methods

House sparrow sampling and environmental data

Since disease outbreaks most often occur during winter [21,23,37], faecal samples from 329 house sparrows were collected during two consecutive sampling periods, i.e. 3 October till 20 December 2013 (‘autumn’) and 10 January until 28 March 2014 (‘winter-early spring’), respectively. Sampled house sparrows originated from 36 populations located in 11 ‘urban’, 7 ‘suburban’ and 18 ‘rural’ regions (details on urbanization levels are given in the supporting information: [S1 Table](#)) clustered pairwise around the Flemish cities of Ghent, Antwerp and Leuven (Fig 1), every population being sampled at least once per sampling period.

The sampling protocol is as described in [38]. Upon capture, each individual was ringed, sexed, weighed (± 0.01 g; digital balance) and their tarsus length was measured (± 0.01 mm; digital calliper). To quantify sparrows’ body condition, we applied the scaled-mass index (SMI), which adjusts the mass of all individuals to that which they would have obtained if they all had the same body size, using the equation of the linear regression of \ln -body mass on \ln -tarsus length estimated by type-II (standardized major axis; SMA) regression [30]. Two outliers were present in the data (i.e. $|\text{standardized residuals}| > 3$), these two observations were not considered for deriving the SMI relationship. The regression slope was 1.50 and average tarsus length was 18.8 mm. We thus calculated the SMI as $\text{body mass} \times (18.8/\text{tarsus length})^{1.50}$ [30]. House sparrows are considered species of Least Concern on the ‘IUCN Red List of Threatened Species’ [39] and all people involved in the sampling were holders of a scientific ringing certificate issued annually by the Agency for Nature and Forest. The sparrows were captured on private land for which oral permission was granted by the respective land owners.

All trapping and sampling protocols were approved by the Ethical Committee VIB Ghent site (EC2013-027).

As environmental predictors, we considered the degree of urbanization, the average air temperatures at the day of sampling and the presence of other granivorous birds. In order to quantify the degree of urbanization at sampling sites, the level of built-up area (BU) was calculated in circular plots around each trapping site based on the very high resolution (i.e. 0.15m pixels) 'Large-scale Reference Database' (LRD) GIS layers [40,41], both at a local 'home-range' scale (using a 100 m radius around the capture site) and at a 'landscape' scale (using a 1600 m radius around capture site, thereby excluding the 100 m radius of the home range scale) [35,42]. The extent of the home-range scale was based on radio-telemetric observations of habitat use by Flemish house sparrows [8] and represents the extent of daily foraging movements. The landscape scale was based on population genetic estimates [35] and reflects the average distance at which sparrow populations can genetically be considered independent from each other. To ensure a more natural environment for the lowest urbanization class, we only selected plots comprising >20% of ecologically valuable areas, as described by the Flemish Governments' Biological Valuation Map [43]. Urbanization at the home-range scale was modelled as a continuous variable (range 1.72–55.04% BU area), while at the landscape scale, it was modeled as class variable, i.e. 'rural' (<5% built-up area), 'suburban' (5–10%) or 'urban' (>10%) [44]. Average daily temperatures were derived from the nearest located weather station and were provided by the Belgian Royal Meteorological Institute (RMI). For every house sparrow population under study, a granivore-index was calculated, i.e. indicating the degree to which a local bird assemblage is dominated by granivorous species which could, through similar foraging strategies, have a higher potential of exchanging enteropathogenic bacteria through the faeco-oral transmission route [45–48]. Since conducting bird surveys during sampling was not feasible because of logistic reasons, we relied on data collected during the most recent Flemish breeding bird atlas [49] whereby the Flemish region was divided in a grid of 5km x 5km. Within each of these squares, bird surveyors were instructed to carry out two one-hour long visits to sets of eight fixed 1km x 1km squares in order to arrive at a list of breeding bird species (see [50] for details). For each sparrow sampling site, we determined the closest (5x5 km) grid cell sampled by the breeding bird atlas (using Euclidean distance) and extracted the species list for that grid cell. Each bird species present was assigned a 'granivore score', varying from 0 to 1, based on bird diets as mentioned in [51]. Following [3], scoring was as follows: 0 = no grains, 0.1 = occasionally grains, 0.5 = frequently grains, 1 = almost exclusively grains. In order to obtain an overall 'granivore-index' for each sparrow sampling site, we summed the granivore-scores of all birds present in a grid cell and divided this sum by the total number of bird species present.

Yersinia isolation and identification

Faecal samples were subjected to a cold enrichment procedure in combination with an alkali (KOH) treatment as described in [52]. This isolation method has previously been demonstrated to be the most successful method for the isolation of *Y. pseudotuberculosis* and *Y. enterocolitica*, even when only small numbers of bacteria are present in a sample [24,53]. All the colonies suspicious for *Yersinia* were biochemically tested at 30°C using Kligler (Oxoid, Ltd), Aesculine (Oxoid, Ltd.) and Urea (Oxoid, Ltd), before performing MALDI-TOF MS (Matrix-Assisted Laser Desorption Ionization-Time of Flight Mass Spectrometry) at the Department of Clinical Microbiology, Laboratory Medicine, AZ Sint-Lucas in Ghent. Every MALDI-TOF assigned-*Y. enterocolitica* and *Y. pseudotuberculosis* was subjected to virulence PCR on chromosomal- (*ail*, *ystA*, *ystB*, *inv*) and plasmid-borne- (*virF*) virulence genes, according to the

PCR-protocol and primers used by [19]. *Yersinia pseudotuberculosis* (22.36a), human pathogenic *Y. enterocolitica* 4/O:3 (75.55b) and *Y. enterocolitica* BT1A (FAVV208) were used as positive controls. If virulence genes were detected, *Y. pseudotuberculosis* isolates were serotyped at the National Reference Center *Yersinia* (IREC).

Although PCR on the combination of chromosomal- and plasmid-borne virulence genes and MALDI-TOF MS has previously been used for the identification of (enteropathogenic) *Y. enterocolitica* and *Y. pseudotuberculosis* [19,52,54–57], the accurate species identification of the latter technique is highly dependent on the validation of the reference library used to identify the bacterial isolates, resulting in high sensitivity and specificity for the validated species [55,56,58]. This validation was performed for *Y. pseudotuberculosis* and *Y. enterocolitica* on the Bruker Daltonik MALDI Biotyper at the Department of Clinical Microbiology [59], but not for other *Yersinia* species. As such, the *Yersinia* species other than *Y. enterocolitica* and *Y. pseudotuberculosis* were not identified up to species level and are included in the statistics as “*Yersinia* species”.

Statistical analyses

First, in order to test whether *Yersinia* spp. prevalence was related to the degree of urbanization and presence of possible host species (expressed by the granivore-index), we applied Generalized Linear Mixed Models (GLMM) [60,61] with a binomial error distribution, using the R ‘lme4’, ‘lmerTest’, ‘Hmisc’, ‘plyr’ and ‘effects’ packages [62–66]. Degree of urbanization at home-range and landscape scales (and the two-factor interaction), granivore-index, daily average temperature, sex and host SMI were modelled as fixed effects, while sampling period was modeled as a random effect using the glmer command (S1 Protocol; S1 and S2 Datasets). To account for possible spatial autocorrelation in *Yersinia* prevalence, latitude and longitude of sampling locations were included as fixed effects [67]. Separate models were run to identify factors influencing the distribution of “*Y. enterocolitica*”, “*Y. pseudotuberculosis*”, “*Yersinia* spp. other than *Y. enterocolitica* and *Y. pseudotuberculosis*”. We applied a model selection procedure based on Akaike’s Information Criterion AIC [68] and calculated AICc values for all possible models, using the R MuMIn package [69]. Models were ranked based on their AICc values, and the relative importance of variables was assessed by summing the AICc weights of all models in which the variable under consideration was included. Important variables are characterized by a high AICc weight (i.e. >0.5) and model-averaged estimates that are higher than their standard errors [70].

Second, to test whether host SMI was impacted by *Yersinia* spp. along the urbanization level, we applied a linear mixed model (LMM) using a Gaussian error distribution, including presence/absence of *Y. enterocolitica*, *Y. pseudotuberculosis* or other *Yersinia* spp., degree of urbanization at home-range and landscape scales (and two-factor interaction), sex, granivore-index, daily average temperature and time (hour) of capture as fixed effects, and sampling period as random effect, using the same packages as for the GLMM, and the lmer-function (S1 Protocol; S1 and S2 Datasets). Model residuals were normally distributed (Shapiro-Wilk $W > 0.95$). Since the AIC-weight of the two-way interaction (see higher) was low (<0.5) for all the GLMM and LMM analyses, models were rerun without interaction to obtain final AIC-weights. All analyses were conducted in R [71].

Results

In total, 329 house sparrows (143 females, 186 males) were captured from rural (51%), suburban (14%) and urban habitats (35%) (S1 Table). All individuals, with the exception of one bird which was diagnosed with pox-virus [38], were apparently healthy. *Yersinia* species were

Table 1. Best models using AIC-based model selection for *Y. pseudotuberculosis*, *Y. enterocolitica*, other *Yersinia* species and Scaled Mass Index as respective response variables.

Response variable: explanatory variables	Log(L)	AIC	ΔAIC	weight
<i>Y. pseudotuberculosis</i> : Granivore-index, Urbanization (landscape level)	-32.21	78.76	0.00	0.64
<i>Y. enterocolitica</i> : Average temperature, Granivore-index, Urbanization (home range level), Urbanization (landscape level)	-185.47	389.50	0.00	0.42
Other <i>Yersinia</i> species: Average temperature, Urbanization (home range level)	-218.75	449.75	0.00	0.60
SMI: Time of capture, Urbanization (landscape level)	-465.15	946.74	0.00	0.55

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isolated from 59% (193/329) of the examined hosts with *Y. enterocolitica* being the most commonly isolated *Yersinia* species, isolated from 31% (103/329) of the individuals (S1 Table). Except for the *ystB*-gene, identified in 92 (89%) of the *Y. enterocolitica* isolates, none of the isolates harbored the examined virulence genes. *Y. pseudotuberculosis* was recovered from 2% (8/329) of the hosts (S1 Table). With four isolates, serotype I was the most encountered serotype. Two isolates were identified as serotype II and two as serotype III and V respectively. All the isolates, apart from both serotype II isolates, originated from different house sparrow populations. Except for serotype III and V, which did not possess the *virF* plasmid-borne virulence gene, both the *inv*- and *virF*-gene were detected in the different serotypes. *Yersinia* species, other than *Y. enterocolitica* and *Y. pseudotuberculosis* were isolated from 41% (134/329) of the house sparrows. In total 51 house sparrows harbored multiple *Yersinia* species in their faeces.

When testing for drivers of different *Yersinia* spp. presence in house sparrow faeces, AIC-based model averaging appointed different variables as important explanatory variables, depending on the *Yersinia* species tested (Table 1). Presence of *Y. pseudotuberculosis* was best explained by the granivore-index, for which a positive relationship was observed (AIC-weight: 0.90, estimate ± standard error: 1.18±0.59; Tables 2 and 3). In addition, landscape-level urbanization influences *Y. pseudotuberculosis* distribution: compared to rural habitats, this species tends to be most prevalent in suburban habitats, and to a lesser extent in urban habitats (AIC-weight: 0.61, estimate: 2.83±1.35 and 1.95±1.08 resp.; Tables 2 and 3). No strong evidence for an effect of host SMI, sex, daily average temperature and home-range level factor on presence of *Y. pseudotuberculosis* was evident (AIC-weights <0.5; Table 2). Variables best explaining the presence of *Y. enterocolitica* were, in order of importance, daily average temperature, the granivore-index, the percentage of built-up area at the home-range scale and, to a lesser extent, at

Table 2. Variable importance after model-averaging in order to explain the presence of *Y. pseudotuberculosis*, *Y. enterocolitica* and other *Yersinia* species and the SMI of the host.

	<i>Y. pseudotuberculosis</i>	<i>Y. enterocolitica</i>	Other <i>Yersinia</i> species	SMI
Granivore-index	0.90	0.92	0.32	0.49
Urbanization (landscape level)	0.61	0.59	0.16	0.64
Urbanization (home range level)	0.38	0.75	0.67	0.48
Average temperature	0.39	1.00	0.92	0.27
Scaled Mass Index	0.26	0.30	0.35	NA
Sex	0.44	0.26	0.44	0.38
Time of Capture	NA	NA	NA	0.76
<i>Y. pseudotuberculosis</i>	NA	NA	NA	0.26
<i>Y. enterocolitica</i>	NA	NA	NA	0.39
Other <i>Yersinia</i> species	NA	NA	NA	0.38

NA (not applicable)

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Table 3. Parameter estimates and standard deviation for response variables: *Y. pseudotuberculosis*, *Y. enterocolitica*, other *Yersinia* species and SMI (shown in Table 1).

Parameters for <i>Y. pseudotuberculosis</i>	Estimate ± SE
Granivore-index	1.18±0.59
Urbanization landscape (Suburban) ^a	2.83±1.35
Urbanization landscape (Urban) ^a	1.95±1.08
Parameters for <i>Y. enterocolitica</i>	
Average temperature	-0.68±0.17
Granivore-index	-0.39±0.15
Urbanization home range	-0.32±0.16
Urbanization landscape (Urban) ^b	0.96±0.48
Urbanization landscape (Rural) ^b	0.79±0.49
Parameters for other <i>Yersinia</i> species	
Average temperature	-0.31±0.12
Urbanization home range	-0.21±0.12
Parameters for SMI	
Time of capture	0.06±0.03
Urbanization landscape (Urban) ^b	-0.43±0.18
Urbanization landscape (Rural) ^b	-0.27±0.17

^a Urbanization within 1600m radius is compared to the Rural habitat

^b Urbanization within 1600m radius is compared to the Suburban habitat

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the landscape scale. *Yersinia enterocolitica* was negatively correlated to daily average temperatures (AIC-weight: 1.00, estimate: -0.68±0.17), to the granivore-index (AIC-weight: 0.92, estimate: -0.39±0.15) and to the percentage of built-up area at home-range level (AIC-weight: 0.75, estimate: -0.32±0.16) (Tables 2 and 3). At the landscape level, the prevalence of *Y. enterocolitica* tends to be lower in suburban house sparrows, compared to the urban and (to a lesser degree) to the rural birds (AIC-weight: 0.59, estimate: 0.96±0.48 and 0.79±0.49 resp.; Table 3). Nor the SMI, nor the sex influenced *Y. enterocolitica* prevalence (AIC-weight: <0.5; Table 2). Presence of other *Yersinia* species was best explained by the average daily temperature (AIC-weight: 0.92, estimate: -0.31±0.12), to which it was negatively related, and by the home-range level (AIC-weight: 0.67, estimate: -0.21±0.12), as *Yersinia* species tended to be less prevalent in more urbanized core habitats (Tables 2 and 3). After accounting for the effect of time of capture (AIC weight: 0.76, 0.06±0.03), we found that sparrow body condition (i.e. SMI) was correlated to landscape-level urbanization (AIC weight: 0.64) (Tables 1–3). The SMI was generally higher for suburban house sparrows compared to either urban (estimate: -0.43±0.18) or rural house sparrows (estimate: -0.27±0.17) (Table 3). Specifically, suburban sparrows were on average 3% heavier than urban birds and 2% than rural sparrows. Presence of *Y. enterocolitica*, *Y. pseudotuberculosis* or other *Yersinia* species, average daily temperatures, sex, granivore-index or home-range level urbanization did not affect hosts SMI (all variable AIC-weights <0.5; Table 2).

Discussion

A high prevalence of *Yersinia* was demonstrated in the faeces of the examined house sparrows, of which most isolates belonged to *Y. enterocolitica* and only a small percentage to *Y. pseudotuberculosis*. These results are in agreement with previous reports using cold enrichment methods [24,31,37]. Apart from the *ystB*-gene, which was demonstrated in most of the *Y.*

enterocolitica isolates and is associated with biotype 1A [19,54], no human-pathogenic *Y. enterocolitica* biotype was recovered from our house sparrows. In humans, controversy exist regarding the pathogenicity of *Y. enterocolitica* BT1A [57,72], in birds however no case-reports related to disease caused by BT1A were found. This could either be an indication that *Y. enterocolitica* BT1A does not tend to be pathogenic in birds, or that only limited research has been conducted on the pathogenicity of *Y. enterocolitica* BT1A in birds.

On the contrary, all recovered serotypes of *Y. pseudotuberculosis*, with serotype I being the most encountered serotype in Europe [24,73,74], have been implicated in yersiniosis cases and outbreaks in birds and mammals, including humans [37,73–78], but have also been isolated from apparently healthy birds and mammals [23,24,37,79,80]. The absence of the *virF* plasmid-borne virulence gene in serotype III and V is potentially an indication of a decrease in virulence [80,81], however, loss of the pYV virulence plasmid during the isolation/purification-procedure cannot be ruled out [19,74]. Since none of the positive birds in our study were recaptured, no inference can be made whether these house sparrows were temporary carriers with the potential of eliminating the pathogen, whether the passerines were in the incubation phase of the disease or actually presented a wildlife-reservoir of *Y. pseudotuberculosis*.

The dominant feeding strategy of the local bird assemblage affected the presence of *Y. pseudotuberculosis* and *Y. enterocolitica* in opposite ways. As for the pathogenic *Y. pseudotuberculosis*, higher prevalence of these bacteria was detected when the local bird populations were dominated by granivorous species, such as the highly susceptible Fringillidae [22,82], which, by using similar foraging strategies could enhance faeco-oral transmission [45–48]. On the other hand, *Y. enterocolitica* BT1A was negatively influenced by the degree of granivory of local bird communities, which could be an indication that, at least for this *Yersinia* species, granivorous birds are less suitable hosts/carriers than birds with other feeding patterns [83,84]. With respect to the other *Yersinia* species, no relation with granivory was demonstrated, suggesting that other, potentially more abiotic factors drive the distribution and prevalence of these *Yersinia* species [29]. However, since the group “*Yersinia* species” most likely comprises various species, the effect of granivores on the species-group could be neutralized due to counteracting effects on the separate *Yersinia* species. We should also keep in mind that for all analyses, the density of the different bird species was not taken into account, nor were other animals that could potentially act as a reservoir, which could likewise alter disease-ecology [12].

The prevalence of *Yersinia enterocolitica* and other *Yersinia* species was highly affected by the average daily temperature, being more prevalent when temperature was lower. As was previously observed when comparing *Yersinia*-survival in soil and water at different temperatures [27], the increased survival at colder temperatures potentially increases the bacteria-load in the environment and subsequently the prevalence in faeces. No such an effect was observed for *Y. pseudotuberculosis*, however the low prevalence likely decreased the power of the statistical analyses and potentially obscured potential relationships between temperature and prevalence.

The amount of built-up area had various effects on the presence of *Yersinia*. At the landscape scale, *Y. pseudotuberculosis* tended to be more prevalent in suburban hosts, and to a lesser extent in urban ones, compared to rural individuals. Although not investigated in our study, previous research has demonstrated higher densities of urban exploiters in suburban and urban regions [2,6], which could enhance the pathogen transmission in these habitats. On the contrary, *Y. enterocolitica* BT1A tends to be less prevalent in suburban house sparrows. The higher prevalence observed in the more urban areas could, similarly as for *Y. pseudotuberculosis*, be related to the higher density of other urban exploiters [6,85]. In rural areas on the other hand, other animals such as rodents, hares and livestock [52,86,87], possibly contribute to an increased occurrence of *Y. enterocolitica* BT1A in the examined house sparrows.

Nevertheless, further investigations are warranted, including different taxa, and taking densities of all potential host species into account.

At the scale of individual home ranges, *Y. enterocolitica* and other *Yersinia* species were shown to be less prevalent in more urbanized habitats. This could be explained by the lower permeability of the surfaces in the more urbanized habitats, from which water excess is lost through run-off and as such dry-up relatively faster compared to actual soil substance [27,28]. Since *Yersinia* species are known to have a higher survival in wet to damp soil [27] the prevalence will likely be higher in less urbanized local habitats. The SMI did not have an influence on the presence of *Y. pseudotuberculosis*, *Y. enterocolitica* or other *Yersinia* species, neither did these *Yersinia* isolates affect the SMI of the house sparrows. With respect to *Y. enterocolitica* BT1A and the environmental *Yersinia* spp. it has been suggested that these *Yersinia* species are part of the normal avian microbiota [24,31], which could explain the lack of effect on house sparrows SMI. Nevertheless, only limited research has been performed on the pathogenicity of *Y. enterocolitica* BT1A and environmental *Yersinia* species in birds. *Yersinia pseudotuberculosis* on the other hand is known to be pathogenic for Passerines, and as such, a bi-directional effect of SMI and *Y. pseudotuberculosis* was expected. The lack of effect in either direction could be due to the low prevalence of *Y. pseudotuberculosis* in our house sparrow populations. However, as [24] previously suggested, wild birds potentially are able to sustain *Y. pseudotuberculosis* at low levels, without clinical signs, developing acute disease when exposed to stressful conditions.

The SMI was observed to increase from the morning to the afternoon, probably related to overnight fasting [26,88] although this observation is not always apparent [89]. Regarding the effect of urbanization on house sparrow body condition, most studies have compared strongly urbanized with rural habitats, disregarding the suburban areas (e.g. [9–11,89]). In this study, no significant differences were observed between populations from rural and strongly urbanized habitats, however, individuals from suburban populations had a higher SMI compared to urban populations (and to a lesser extent rural ones). Body condition has earlier been associated with stress response and overall health [9,30], though environmental factors such as habitat coverage [8], predictability of food supply and quality [10], presence of predators [8] have been hypothesized to influence the body condition of the birds. Suburban habitats in Flanders are typically characterized by strongly connected hedges and bushes, which are generally considered good habitat for house sparrows, allowing for a higher foraging efficiency compared to more fragmented highly urbanized or rural habitats. Indeed, [8] found that suitable foraging and shelter sites are highly scattered in urban areas. In rural areas, shelter sites are more connected than in highly urbanized areas, but the presence of intensive-agricultural fields forces sparrows to occupy larger home-ranges, increasing the energy expenditure when patrolling the entire home range, and thus potentially decreasing the body condition [8].

In conclusion, we here show that the urbanization gradient affects body condition and pathogen occurrence, two determinants of pathogen infection dynamics, suggesting a potential impact of urbanization on avian health. When assessing the impact of urbanization on animal health and pathogen dynamics, information regarding the presence/absence and preferably also the density of other suitable hosts, the two-way interaction between pathogen and host, and various levels of urbanization including the suburban habitat is required in order to have a better understanding of how urbanization can have an impact on urban wildlife health and diseases.

Supporting information

S1 Table. Sampled house sparrow populations with indication of the percentage of Build-Up-area in the local and landscape scale and provision of information regarding presence

or absence of *Y. pseudotuberculosis* and *Y. enterocolitica*. Abbreviations similar as in Fig 1. (DOCX)

S1 Protocol. (g)lmer codes in R.
(XLSX)

S1 Dataset. Dataset comprising the variables included in the (g)lmer.
(CSV)

S2 Dataset. Dataset comprising the variables used to calculate the SMI of the house sparrows.
(CSV)

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References

1. Grimm NB, Grove M, Pickett STA, Redman CL. Integrated approaches to long-term studies of urban ecological systems. *Bioscience*. 2000; 50(7): 571–584
2. Evans KL, Newson SE, Gaston KJ. Habitat influences on urban avian assemblages. *Ibis*. 2009; 151: 19–39. <https://doi.org/10.1111/j.1474-919X.2008.00898.x>
3. Sol D, González-Lagos C, Moreira D, Maspons J, Lapiedra O. Urbanisation tolerance and the loss of avian diversity. *Ecol Lett*. 2014; 17(8): 942–950. <https://doi.org/10.1111/ele.12297> PMID: 24835452
4. McKinney ML. Effects of urbanization on species richness: a review of plants and animals. *Urban Ecosyst*. 2008; 11: 161–176. <https://doi.org/10.1007/s11252-007-0045-4>
5. McKinney ML. Urbanization, Biodiversity, and Conservation. *BioScience*. 2002; 52(10): 883–890.
6. Blair RB. Land use and avian species diversity along an urban gradient. *Ecol Appl*. 1996; 6(2): 506–519. <http://www.jstor.org/stable/2269387>
7. Chace JF, Walsh JJ. Urban effects on native avifauna: a review. *Landsc Urban Plan*. 2006; 74: 46–69. <https://doi.org/10.1016/j.landurbplan.2004.08.007>
8. Vangestel C, Braeckman BP, Matheve H, Lens L. Constraints on Home Range Behaviour Affect Nutritional Condition in Urban House Sparrows (*Passer domesticus*). *Biol J Linn Soc Lond*. 2010; 101: 41–50. <https://doi.org/10.1111/j.1095-8312.2010.01493.x>
9. Bókony V, Seress G, Nagy S, Lendvai Á, Liker A. Multiple indices of body condition reveals no negative effect of urbanization in adult house sparrows. *Landsc Urban Plan*. 2012; 104: 75–84. <https://doi.org/10.1016/j.landurbplan.2011.10.006>
10. Salleh Hudin N, Strubbe D, Teyssier A, De Neve L, White J, Janssens GPJ, et al. Predictable food supplies induce plastic shifts in avian scaled body mass. *Behavioral Ecology*. 2016; 27(6): 1833–1840. <https://doi.org/10.1093/beheco/arw108>
11. Meillère A, Brischox F, Henry P-Y, Michaud B, Garcin R, Angelier F. Growing in a city: Consequences on body size and plumage quality in an urban dweller, the house sparrow (*Passer domesticus*). *Landsc Urban Plan*. 2017; 160: 127–138. <https://doi.org/10.1016/j.landurbplan.2016.12.014>
12. Keesing F, Holt RD, Ostfeld RS. Effects of species diversity on disease risk. *Ecol Lett*. 2006; 9: 485–498. <https://doi.org/10.1111/j.1461-0248.2006.00885.x> PMID: 16623733
13. Bradley CA, Altizer S. Urbanization and the ecology of wildlife diseases. *TRENDS Ecol Evol*. 2007; 22(2): 95–102. <https://doi.org/10.1016/j.tree.2006.11.001> PMID: 17113678
14. Evans KL, Gaston KJ, Sharp SP, McGowan A, Simeoni M, Hatchwell BJ. Effects of urbanisation on disease prevalence and age structure in blackbird *Turdus merula* populations. *Oikos*. 2009; 118: 774–782. <https://doi.org/10.1111/j.1600-0706.2008.17226.x>
15. Delgado-V CA, French K. Parasite-bird interactions in urban areas: Current evidence and emerging questions. *Landsc Urban Plan*. 2009; 105: 5–14. <https://doi.org/10.1016/j.landurbplan.2011.12.019>
16. Hamer SA, Lehrer E, Magle SB. Wild birds as sentinels for multiple zoonotic pathogens along an urban to rural gradient in Greater Chicago, Illinois. *Zoonoses Public Health*. 2012; 59: 355–364. <https://doi.org/10.1111/j.1863-2378.2012.01462.x> PMID: 22353581
17. Artois M, Delahay R, Guberti V, Cheeseman C. Control of infectious diseases of wildlife in Europe. *Vet J*. 2001; 162(2): 141–152. <https://doi.org/10.1053/tvjl.2001.0601> PMID: 11531398
18. EFSA and ECDC. The European Union summary report on trends and sources of zoonoses, zoonotic agents and food-borne outbreaks in 2014. *EFSA Journal*. 2015; 13(12): 4329–4520. <https://doi.org/10.2903/j.efsa.2015.4329>
19. Thoerner P, Bin Kingombe CI, Bögli-Stubler K, Bissig-Choisat B, Wassenaar TM, Frey J, et al. PCR detection of virulence genes in *Yersinia enterocolitica* and *Yersinia pseudotuberculosis* and investigation of virulence gene distribution. *Appl Environ Microbiol*. 2003; 69(3): 1810–1816. <https://doi.org/10.1128/AEM.69.3.1810-1816.2003> PMID: 12620874
20. Clark GM, Locke LN. Case Report: Observations on Pseudotuberculosis in common grackles. *Avian Dis*. 1962; 6(4): 506–510. <http://www.jstor.org/stable/1587927>
21. Mair NS. Yersiniosis in wildlife and its public health implications. *J Wildl Dis*. 1973; 9(1): 64–71. <https://doi.org/10.7589/0090-3558-9.1.64> PMID: 4571715
22. Cork SC, Collins-Emerson JM, Alley MR, Fenwick SG. Visceral lesions caused by *Yersinia pseudotuberculosis*, serotype II, in different species of bird. *Avian Pathol*. 1999; 28(4): 393–399. <https://doi.org/10.1080/03079459994669> PMID: 26905497
23. Mackintosh CG, Henderson T. Potential wildlife sources of *Yersinia pseudotuberculosis* for farmed red deer (*Cervus elaphus*). *N Z Vet J*. 1984; 32(12): 208–210. <https://doi.org/10.1080/00480169.1984.35123> PMID: 16031024

24. Niskanen T, Waldenström J, Fredriksson-Ahomaa M, Olsen B, Korkeala H. *VirF*-Positive *Yersinia pseudotuberculosis* and *Yersinia enterocolitica* found in migratory birds in Sweden. *Appl Environ Microbiol.* 2003; 69(8): 4670–4675. <https://doi.org/10.1128/AEM.69.8.4670-4675.2003> PMID: 12902256
25. Bichet C, Scheiffler R, Coeurdassier M, Julliard R, Sorci G, Loiseau C. Urbanization, trace metal pollution, and malaria prevalence in the house sparrow. *PLOS ONE.* 2013; 8(1): e53866. <https://doi.org/10.1371/journal.pone.0053866> PMID: 23342022
26. Galbraith JA, Stanley MC, Jones DN, Beggs JR. Experimental feeding regime influences urban bird disease dynamics. *J Avian Biol.* 2017; 48: 001–014. <https://doi.org/10.1111/jav.01076>
27. Tashiro K, Kubokura Y, Kato Y, Kaneko K, Ogawa M. Survival of *Yersinia enterocolitica* in soil and water. *J Vet Med Sci.* 1991; 53(1): 23–27. PMID: 1830776
28. Trusilova K, Jung M, Churkina G, Karstens U, Heimann M, Claussen M. Urbanization impacts on the climate in Europe: Numerical experiments by the PSU-NCAR Mesoscale Model (MM5). *Journal of Applied Meteorology and Climatology.* 2008; 47: 1442–1455. <https://doi.org/10.1175/2007JAMC1624.1>
29. Reuter S, Connor TR, Barquist L, Walker D, Feltwell T, Harris SR, et al. Parallel independent evolution of pathogenicity with the genus *Yersinia*. *PNAS.* 2014; 111(18): 6768–6773. <https://doi.org/10.1073/pnas.1317161111> PMID: 24753568
30. Peig J, Green AJ. New perspectives for estimating body condition from mass/length data the scaled mass index as an alternative method. *Oikos.* 2009; 118(12): 1883–1891. <https://doi.org/10.1111/j.1600-0706.2009.17643.x>
31. Kisková J, Hrehová Z, Aniga MJ, Lukán M, Haas M, Jurcovicová M. *Yersinia* Species in the Dunnock (*Prunella modularis*) in Sub-alpine Habitats of the Western Carpathians. *Pol J Microbiol.* 2011; 60(1): 79–83 PMID: 21630578
32. Verbrugghe E, Boyen F, Gaastra W, Bekhuis L, Leyman B, Van Parys A, et al. The complex interplay between stress and bacterial infections in animals. *Vet Microbiol.* 2012; 155: 115–127. <https://doi.org/10.1016/j.vetmic.2011.09.012> PMID: 21963418
33. De Laet J, Summers-Smith JD. The Status of the Urban House Sparrow *Passer domesticus* in North-Western Europe: a Review. *J Ornithol.* 2007; 148(2): S275–278. <https://doi.org/10.1007/s10336-007-0154-0>
34. Everaert J, Bauwens D. A Possible Effect of Electromagnetic Radiation from Mobile Phone Base Stations on the Number of Breeding House Sparrows (*Passer domesticus*). *Electromagn Biol Med.* 2007; 26: 63–72. <https://doi.org/10.1080/15368370701205693> PMID: 17454083
35. Vangestel C, Mergeay J, Dawson DA, Vandomme V, Lens L. Spatial heterogeneity in genetic relatedness among house sparrows along an urban-rural gradient as revealed by individual-based analysis. *Mol Ecol.* 2011; 20: 4643–4653. <https://doi.org/10.1111/j.1365-294X.2011.05316.x> PMID: 22004175
36. De Coster G, De Laet J, Vangestel C, Adriaensen F, Lens L. Citizen science in action—Evidence for long-term, region-wide house sparrow declines in Flanders, Belgium. *Landsc Urban Plan.* 2015; 134: 139–146. <https://doi.org/10.1016/j.landurbplan.2014.10.020>
37. Cork SC, Marshall RB, Madie P, Fenwick SG. The role of wild birds and the environment in the epidemiology of *Yersinia* in New Zealand. *N Z Vet J.* 1995; 43(5): 169–174. <https://doi.org/10.1080/00480169.1995.35883> PMID: 16031843
38. Rouffaer L, Lens L, Haesendonck R, Teyssier A, Salleh Hudin N, Strubbe D, et al. House Sparrows Do Not Constitute a Significant *Salmonella* Typhimurium Reservoir across Urban Gradients in Flanders, Belgium. *PLOS ONE.* 2016; 11(5): e0155366. <https://doi.org/10.1371/journal.pone.0155366> PMID: 27168186
39. BirdLife International. 2016. *Passer domesticus*. The IUCN Red List of Threatened Species 2016: e.T103818789A87943000. Accessed on: 13 April 2017
40. AGIV. 2013a. Large Scale Reference Database (LRD): <https://overheid.vlaanderen.be/producten-diensten/basiskaart-vlaanderen-grb>.
41. AGIV. 2013b. Gebouw aan de grond (Gbg): <https://www.agiv.be/producten/grb/objectcatalogus/entiteiten/gbg-gebouw-aan-de-grond>
42. Vriens L, Bosch H, De Knijf G, De Saeger S, Oosterlynck P, Guelinckx R, et al. De Biologische Waarderingskaart. Biotopen en hun verspreiding in Vlaanderen en het Brussels Hoofdstedelijk Gewest, van het Instituut voor Natuur-en Bosonderzoek INBO. 2011. Mededelingen
43. Teyssier A, Rouffaer LO, Salleh Hudin N, Strubbe D, Lens L, White J. Inside the guts of the city: Urban-induced alterations of the gut microbiota in a wild passerine. *Sci Total Environ.* 2018; 612: 1276–1286. <https://doi.org/10.1016/j.scitotenv.2017.09.035> PMID: 28898933
44. Melles S, Glenn S, Martin K. Urban bird diversity and landscape complexity: Species—environment associations along a multiscale habitat gradient. *Conservation Ecology.* 2003; 7(1): 5. <http://www.consecol.org/vol7/iss1/art5>

45. Brittingham MC, Temple SA. Avian Disease and Winter Bird Feeding. The Passenger Pigeon. 1988; 50 (3): 195–203
46. Pennycott TW, Cinderey RN, Park A, Mather HA, Foster G. *Salmonella enterica* subspecies *enterica* serotype Typhimurium and *Escherichia coli* O86 in wild birds at two garden sites in south-west Scotland. Vet Rec. 2002; 151: 563–567. <https://doi.org/10.1136/vr.151.19.563> PMID: 12452355
47. Refsum T, Vikøren T, Handeland K, Kapperud G, Holstad G. Epidemiologic and Pathologic Aspects of *Salmonella* Typhimurium Infection in Passerine Birds in Norway. J Wildl Dis. 2003; 39(1): 64–72. <https://doi.org/10.7589/0090-3558-39.1.64> PMID: 12685069
48. Perkins AJ, Anderson G, Wilson JD. Seed food preferences of granivorous farmland passerines. Bird Study. 2007; 54(1): 46–53. <https://doi.org/10.1080/00063650709461455>
49. Vermeersch G, Anselin A, Devos K, Herremans M, Stevens J, Gabriëls J et al. Atlas van de Vlaamse broedvogels: 2000–2002. Mededelingen van het Instituut voor Natuurbehoud: Brussel. 2004a; 498p. ISBN 90-403-0215-4
50. Vermeersch G, Anselin A, Devos K, Herremans M, Stevens J, Gabriëls J, et al. Atlas of the Breeding birds in Flanders: 2000–2002. Bird Census News 1/2: Chapter 2: 14–33; 2004b.
51. Cramp S, Perrins CM. Handbook of the birds of Europe, the Middle East and North Africa: The birds of the western Palearctic; Oxford University Press: Oxford; 1977–1994
52. Rouffaer LO, Baert K, Van den Abeele A-M, Cox I, Vanantwerpen G, De Zutter L, et al. Low prevalence of human enteropathogenic *Yersinia* spp. in brown rats (*Rattus norvegicus*) in Flanders. PLOS ONE. 2017; 12(4): e0175648. <https://doi.org/10.1371/journal.pone.0175648> PMID: 28403184
53. Niskanen T, Fredriksson-Ahoma M, Korkeala H. *Yersinia pseudotuberculosis* with limited genetic diversity is a common finding in tonsils of fattening pigs. J Food Prot. 2002; 65: 540–545. <https://doi.org/10.4315/0362-028X-65.3.540> PMID: 11899054
54. Singh I, Virdi JS. Production of *Yersinia* stable toxin (YST) and distribution of *yst* genes in biotype 1A strains of *Yersinia enterocolitica*. J Med Microbiol. 2004; 53: 1065–1068. <https://doi.org/10.1099/jmm.0.45527-0> PMID: 15496381
55. Ayyadurai S, Flaudrops C, Raoult D, Drancourt M. Rapid identification and typing of *Yersinia pestis* and other *Yersinia* species by matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry. BMC Microbiol. 2010; 10: 285. <https://doi.org/10.1186/1471-2180-10-285> PMID: 21073689
56. Stephan R, Cernela N, Ziegler D, Pflüger V, Tonolla M, Ravasi D, et al. Rapid species specific identification and subtyping of *Yersinia enterocolitica* by MALDI-TOF Mass spectrometry. J Microbiol Methods. 2011; 87: 150–153. <https://doi.org/10.1016/j.mimet.2011.08.016> PMID: 21914454
57. Stephan R, Joutsen S, Hofer E, Säde E, Björkroth J, Ziegler D, et al. Characteristics of *Yersinia enterocolitica* biotype 1A strains isolated from patients and asymptomatic carriers. Eur J Clin Microbiol Infect Dis. 2013; 32(7): 869–875. <https://doi.org/10.1007/s10096-013-1820-1> PMID: 23354676
58. Seng P, Drancourt M, Gouriet F, La Scola B, Fournier P-E, Rolain JM, et al. Ongoing revolution in bacteriology: Routine identification of bacteria by Matrix-Assisted Laser Desorption Ionization Time-of-Flight Mass Spectrometry. Clin Infect Dis. 2009; 49: 543–551. <https://doi.org/10.1086/600885> PMID: 19583519
59. CLSI. 2015. Verification of Commercial Microbial Identification and Antimicrobial Susceptibility Testing Systems. 1st ed. CLSI guideline M52. Wayne PA. Clinical and Laboratory Standards Institute
60. Bates D, Mächler M, Bolker BM, Walker SC. Fitting Linear Mixed-Effects Models using lme4. J Stat Softw. 2015; 67(1). <https://doi.org/10.18637/jss.v067.i01>
61. Bates D, Mächler M, Bolker BM, Walker SC. Linear Mixed-Effects Models using ‘Eigen’ and S. R package version 1.1–12; 2016: <http://cran.r-project.org/package=lme4>
62. Harrell FE. Harrell Miscellaneous. R package version 4.0–2; 2016: <http://cran.r-project.org/package=Hmisc>
63. Bates D, Mächler M, Bolker B, Walker S, et al. Linear Mixed-Effects Models using ‘Eigen’ and S4. R package version 1.1–12; 2016: <http://cran.r-project.org/package=lme4>
64. Kuznetsova A, Brockhoff PB, Christensen RHB. Tests in Linear Mixed Effects Models. R package version 2.0–33; 2016: <http://cran.r-project.org/package=lmerTest>
65. Wickham H. Tools for Splitting, Applying and Combining Data. R package version 1.8.4.; 2016: <http://cran.r-project.org/package=plyr>
66. Fox J, Weisberg S, Friendly M, Hong J, et al. Effect Displays for Linear, Generalized Linear, and Other Models. R package version 3.1–2; 2016: <http://cran.r-project.org/package=Effects>

67. Dormann CF, McPherson JM, Araújo MB, Bivand R, Bolliger J, Carl G, et al. Methods to account for spatial autocorrelation in the analysis of species distributional data: a review. *Ecography*. 2007; 30: 609–628. <https://doi.org/10.1111/j.2007.0906-7590.05171.x>
68. Burnham KP, Anderson DR. *Model Selection and Multimodel Inference: A Practical Information-Theoretic Approach* (Second Edition). Springer: Verlag New York Berlin Heidelberg; 2002
69. Barton K. Multi-Model Inference. R package version 1.15.6; 2015: <http://cran.r-project.org/package=MuMIn>
70. Anderson DR. *Model-based Inference in the Life Sciences: a primer on evidence*. Springer: New York; 2008.
71. R Development Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria; 2015: ISBN 3-900051-07-0, URL: <http://www.R-project.org>. R version 3.2.3 (2015-12-10)—"Wooden Christmas-Tree"
72. Tennant SM, Grant TH, Robins-Browne RM. Pathogenicity of *Yersinia enterocolitica* biotype Ia. *FEMS Immunol Med Microbiol*. 2003; 38(2): 127–137. [https://doi.org/10.1016/S0928-8244\(03\)00180-9](https://doi.org/10.1016/S0928-8244(03)00180-9) PMID: 13129647
73. EFSA. Scientific Opinion of the Panel on BIOHAZ on a request from EFSA on monitoring and identification of human enteropathogenic *Yersinia* spp. *The EFSA Journal*. 2007; 595: 1–30.
74. Niskanen T, Laukkanen R, Murros A, Björkroth J, Skurnik M, Korkeala H, et al. Characterisation of non-pathogenic *Yersinia pseudotuberculosis*-like strains isolated from food and environmental samples. *Int J Food Microbiol*. 2009; 129: 150–156. <https://doi.org/10.1016/j.ijfoodmicro.2008.11.015> PMID: 19095324
75. Bradley JM, Skinner JL. Isolation of *Yersinia pseudotuberculosis* serotype V from the blood of a patient with sickle-cell anaemia. *J Med Microbiol*. 1974; 7: 383–386. <https://doi.org/10.1099/00222615-7-3-383> PMID: 4607637
76. Nakano T, Kawaguchi H, Nakao K, Maruyama T, Kamiya H, Sakurai M. Two outbreaks of *Yersinia pseudotuberculosis* 5a infection in Japan. *Scand J Infect Dis*. 1989; 21: 175–179. <https://doi.org/10.3109/00365548909039966> PMID: 2658021
77. Fukushima H, Gomyoda M, Ishikura S, Nishio T, Moriki S, Endo J, et al. Cat-contaminated environmental substances lead to *Yersinia pseudotuberculosis* infection in children. *J Clin Microbiol*. 1989; 27(12): 2706–2709. PMID: 2687319
78. Nuorti JP, Niskanen T, Hallanvuo S, Mikkola J, Kela E, Hatakka M, et al. A widespread outbreak of *Yersinia pseudotuberculosis* O:3 infection from iceberg lettuce. *J Infect Dis*. 2004; 189: 766–774. <https://doi.org/10.1086/381766> PMID: 14976592
79. Hamasaki S-i, Hayashidani H, Kaneko K-i, Ogawa M, Shigeta Y. A survey for *Yersinia pseudotuberculosis* in migratory birds in coastal Japan. *J Wildl Dis*. 1989; 25(3): 401–403. <https://doi.org/10.7589/0090-3558-25.3.401> PMID: 2761013
80. Fukushima H, Sato T, Nagasako R, Takeda I. Acute mesenteric lymphadenitis due to *Yersinia pseudotuberculosis* lacking a virulence plasmid. *J Clin Microbiol*. 1991; 29(6): 1271–1275 PMID: 1864948
81. Nagano T, Kiyohara T, Suzuki K, Tsubokura M, Otsuki K. Identification of pathogenic strains within serogroups of *Yersinia pseudotuberculosis* and the presence of non-pathogenic strains isolated from animals and the environment. *J Vet Med Sci*; 1997; 59(3): 153–158 PMID: 9101473
82. Sandmeier P, Coutteel P. Chapter 39: Management of Canaries, Finches and Mynahs. In: *Clinical Avian Medicine* (Volume II): Spix Publishing; Editors: Harrison GJ and Lightfoot TL; 2005
83. Novotný M, Fečková M, Janiga M, Lukáš M, Novotná M, Kovalčíková Z. High incidence of *Yersinia enterocolitica* (Enterobacteriaceae) in Alpine Accentors *Prunella collaris* of the Tatra Mountains. *Acta Ornithologica*. 2007; 42(2): 137–143
84. Benskin C McW H., Wilson K, Jones K, Hartley IR. Bacterial Pathogens in Wild Birds: a Review of the Frequency and Effects of Infection. *Biol Rev*. 2009; 84: 349–373. <https://doi.org/10.1111/j.1469-185X.2008.00076.x> PMID: 19438430
85. McKinney ML. Urbanization as a major cause of biotic homogenization. *Biol Conserv*. 2006; 127: 247–260. <https://doi.org/10.1016/j.biocon.2005.09.005>
86. Frandölich K, Wisser J, Schmäuser H, Fehlberg U, Neubauer H, Grunow R, et al. Epizootiologic and ecologic investigations of european brown hares (*Lepus europaeus*) in selected populations from Schleswig-Holstein, Germany. *J Wildl Dis*. 2003; 39(4):751–761. <https://doi.org/10.7589/0090-3558-39.4.751> PMID: 14733269
87. Vanantwerpen G, Van Damme I, De Zutter L, Houf K. Within-batch prevalence and quantification of human pathogenic *Yersinia enterocolitica* and *Y. pseudotuberculosis* in tonsils of pigs at slaughter. *Vet Microbiol*. 2014; 169: 223–227. <https://doi.org/10.1016/j.vetmic.2013.12.019> PMID: 24472227

88. Milenkaya O, Weinstein N, Legge S, Walters JR. Variation in body condition indices of crimson finches by sex, breeding stage, age, time of day, and year. *Conserv Physiol*. 2013; 1(1):cot020. <https://doi.org/10.1093/conphys/cot020> PMID: 27293604
89. Meillère A, Brischoux F, Parenteau C, Angelier F. Influence of urbanization on body size, condition and physiology in an urban exploiter: a multi-component approach. *PLOS ONE*. 2015; 10(8): e0135685. <https://doi.org/10.1371/journal.pone.0135685> PMID: 26270531