Original Research

Factors Influencing Clinical and MRI Outcomes of Mesenchymal Stem Cell Implantation With Concomitant High Tibial Osteotomy for Varus Knee Osteoarthritis

Yong Sang Kim,* MD, Dong Suk Suh,* MD, Dae Hyun Tak,* MD, Pill Ku Chung,* MD, Yoo Beom Kwon,* MD, Tae Yong Kim,* MD, and Yong Gon Koh,*[†] MD

Investigation performed at Yonsei Sarang Hospital, Seoul, Republic of Korea

Background: Cartilage repair procedures using mesenchymal stem cells (MSCs) can provide superior cartilage regeneration in the medial compartment of the knee joint when high tibial osteotomy (HTO) is performed for varus knee osteoarthritis (OA). However, few studies have reported the factors influencing the outcomes of MSC implantation with concomitant HTO.

Purpose: To investigate the outcomes of MSC implantation with concomitant HTO and to identify the prognostic factors that are associated with the outcomes.

Study Design: Case series; Level of evidence, 4.

Methods: A total of 71 patients (75 knees) were retrospectively evaluated after MSC implantation with concomitant HTO. Clinical and radiological outcomes were evaluated, and magnetic resonance imaging (MRI) was used to assess cartilage regeneration. Statistical analyses were performed to determine the effect of different factors on clinical, radiographic, and MRI outcomes.

Results: Clinical and radiographic outcomes improved significantly from preoperatively to final follow-up (P < .001 for all), and overall cartilage regeneration was encouraging. Significant correlations were found between clinical and MRI outcomes. However, radiographic outcomes were not significantly correlated with clinical or MRI outcomes. Patient age and number of MSCs showed significant correlations with clinical and MRI outcomes. On multivariate analyses, patient age and number of MSCs showed high prognostic significance with poor clinical outcomes.

Conclusion: MSC implantation with concomitant HTO provided feasible cartilage regeneration and satisfactory clinical outcomes for patients with varus knee OA. Patient age and number of MSCs were important factors that influenced the clinical and MRI outcomes of MSC implantation with concomitant HTO for varus knee OA.

Keywords: mesenchymal stem cells; high tibial osteotomy; osteoarthritis; varus knee

The Orthopaedic Journal of Sports Medicine, 9(2), 2325967120979987 DOI: 10.1177/2325967120979987 © The Author(s) 2021

Primary knee osteoarthritis (OA) is caused by a combination of risk factors, including increasing age and obesity, knee malalignment, increased biomechanical loading of joints, and/or genetics.³⁷ Among these risk factors, varus knee malalignment can lead to increased loads on the medial compartment and induce progressive cartilage degeneration, leading to medial compartmental knee OA.⁵ For patients who have medial compartmental knee OA, high tibial osteotomy (HTO), which corrects the limb deformity by shifting the mechanical axis to the lateral side and decreasing contact pressure on the affected medial cartilage, can provide the adequate mechanical environment for preventing further degeneration of articular cartilage.^{2,33,51} Although authors have reported encouraging short-term and midterm outcomes of HTO,^{3,48,50} satisfactory long-term outcomes of HTO are questionable until

[†]Address correspondence to Yong Gon Koh, MD, Department of Orthopaedic Surgery, Yonsei Sarang Hospital, 10 Hyoryeong-ro, Seocho-gu, Seoul, Republic of Korea (email: yonggonkoh@gmail.com).

^{*}Center for Stem Cell & Arthritis Research, Department of Orthopaedic Surgery, Yonsei Sarang Hospital, Seoul, Republic of Korea.

Final revision submitted August 9, 2020; accepted August 11, 2020.

The authors declared that there are no conflicts of interest in the authorship and publication of this contribution. AOSSM checks author disclosures against the Open Payments Database (OPD). AOSSM has not conducted an independent investigation on the OPD and disclaims any liability or responsibility relating thereto.

Ethical approval for this study was obtained from Yonsei Sarang Hospital (project No. 19-E003-005).

This open-access article is published and distributed under the Creative Commons Attribution - NonCommercial - No Derivatives License (https://creativecommons.org/ licenses/by-nc-nd/4.0/), which permits the noncommercial use, distribution, and reproduction of the article in any medium, provided the original author and source are credited. You may not alter, transform, or build upon this article without the permission of the Author(s). For article reuse guidelines, please visit SAGE's website at http://www.sagepub.com/journals-permissions.



Figure 1. Flow diagram of patient involvement in the study. MRI, magnetic resonance imaging.

adequate cartilage regeneration in the medial compartment of the knee joint is accomplished.^{48,52} Therefore, several studies on additional cartilage repair procedures with concomitant HTO have emerged in an attempt to obtain more adequate regeneration of cartilage in the medial compartment of the knee joint.^{13,15,44}

Mesenchymal stem cells (MSCs) have been assessed as a potential cell-based therapy for the treatment of OA because of their capacity for differentiation into chondrocytes and their immunomodulatory properties.^{16,29} It has been reported that the addition of MSC treatment is useful in achieving greater cartilage remodeling and better clinical outcomes in patients undergoing HTO, 24,25,28,49,55 which indicates that the outcomes of this combined procedure should be different from those of HTO alone. However, to date, no studies have assessed factors that influence the outcomes of MSC implantation in combination with HTO for varus knee OA. The identification of factors associated with favorable and unfavorable outcomes would provide patients with accurate expectations of this treatment. Accordingly, this study aimed to investigate the clinical, radiographic, and magnetic resonance imaging (MRI) outcomes of MSC implantation in combination with HTO in patients with varus knee OA and to identify the prognostic factors that are associated with the outcomes.

METHODS

Patient Enrollment

This study was reviewed and approved by the institutional review board at our institution, and informed consent was obtained from all participants before study enrollment. We retrospectively reviewed the data of 83 consecutive patients (91 knees) who underwent MSC implantation in combination with HTO for varus knee

 TABLE 1

 Patient Demographics and Characteristics^a

Variable	Value
Age, y	$60.2 \pm 6.1 \ (48-73)$
Sex, male/female, n	35/40
Side of involvement, right/left/both, n	37/34/2
Body mass index, kg/m ²	$25.8 \pm 3.0 \ (19.3-32.9)$
Follow-up period, mo Lesion size, cm ²	$26.8 \pm 3.1 \; (24\text{-}34)$
Femoral condyle Tibial plateau	$7.1 \pm 1.0 (4.8-9.1)$ $6.2 \pm 0.9 (4.0-8.3)$
No. of MSCs	$1.19 \times 10^7 \pm 3.72 \times 10^6 (7.21 \times 10^6 \text{-} 1.98 \times 10^7)$

 aValues are expressed as mean \pm SD (range) unless otherwise indicated. MSC, mesenchymal stem cell.

OA between July 2017 and November 2018. The inclusion criteria, determined by medical records, plain radiographs, and MRI scans, were symptomatic knee pain unresponsive to nonoperative treatment, radiographs showing medial compartmental knee OA classified as Kellgren-Lawrence grade 3 or 4,²¹ and varus deformity between the tibial and femoral mechanical axes as measured on a hip-to-ankle standing anteroposterior (AP) radiograph.⁴⁰ The exclusion criteria were previous surgery, cartilage lesions in the lateral or patellofemoral compartment, and other pathological diseases including rheumatoid arthritis, hemophilia, active knee infections, chronic anterior/posterior cruciate ligament instability, or meniscal tears. Of the 83 patients (91 knees), 12 patients (16 knees) were excluded: 9 patients (13 knees) did not meet the inclusion criteria, 2 patients (2 knees) were lost to follow-up, and 1 patient (1 knee) refused to undergo follow-up MRI. A total of 71 patients (75 knees) were ultimately included in the analysis (Figure 1). The baseline demographics of the patients are summarized in Table 1.

Preparation of MSCs

Subcutaneous adipose tissue samples were obtained through liposuction from the gluteal regions of the patients 1 day before MSC implantation with concomitant HTO. The liposuction material was aspirated by gentle suction, the gluteal fat pad was collected, and separation of the stromal vascular fraction (SVF) through centrifugation was performed in accordance with a previously reported method.⁵⁶ Stem cells were isolated from the lipoaspirate by enzymatic digestion and cultured to determine the characteristics of the adipose-derived stem cells. The adipose-derived stem cell immunophenotype was investigated using cell markers by analytical flow cytometry, 35 and the differentiation potentials of adipose-derived stem cells into adipogenic, osteogenic, and chondrogenic cell lineages were also assessed using specific inductive culture media.³⁵ These isolation and characterization procedures determined that the SVF contained adiposederived MSCs, which made up 9.8% of this fraction.

Consequently, a mean of 1.21×10^8 cells in the SVF, which contained a mean of 1.19×10^7 stem cells (9.8% of 1.21×10^8 cells in the SVF; range, 7.21×10^6 to 1.98×10^7), were used for MSC implantation.

Surgical Procedure

The surgical procedure was identical in all patients and was performed as described previously.²³ The patients were placed in the supine position on the operating table, and thigh tourniquets were applied. Before MSC implantation, accurate debridement of all unstable and damaged cartilage in the lesion was performed. The prepared MSCs were loaded into the fibrin glue product (Greenplast Kit W; Green Cross), which was used as a scaffold for MSC implantation. After the arthroscopic fluid was extracted, MSCs mixed with fibrin glue were implanted into the lesion site under arthroscopic guidance. Next, the applied MSCs mixed with fibrin glue were manipulated using a probe to evenly cover the surface of the cartilage lesion. No marrow stimulation procedures such as microfracture, subchondral drilling, or abrasion arthroplasty were performed before this procedure. After the arthroscopic procedure, HTO was performed. During preoperative planning, the desired correction angle and wedge size were calculated using hip-to-ankle standing AP radiographs, with the aim of mild overcorrection.¹¹ Openwedge HTO was performed with the angular-stable TomoFix plate (Synthes), and the osteotomy site was filled with a β -tricalcium phosphate wedge (Synthes), which is a synthetic resorbable substitute possessing compressive strength similar to that of cancellous bone, in compliance with the open space.

As a postoperative rehabilitation exercise, active and passive range of motion was initiated on the first postoperative day. The patients were allowed to move their knee from 0° to 90° after 2 weeks. Toe-touch weightbearing was allowed at 2 weeks after surgery, followed by partial weightbearing for 2 weeks. Full weightbearing was allowed at 4 weeks, after radiographic evidence of bone consolidation at the osteotomy site was confirmed. Sports and intense levels of activity were allowed at 3 months after surgery.

Clinical and Radiographic Evaluations

All patients were evaluated clinically and radiographically before surgery and postoperatively at 4 weeks, 3 months, 6 months, 1 year, and annually until the last follow-up visit (mean, 26.8 months [range, 24-34 months]). For the clinical evaluation, the Lysholm score²⁶ and the Knee injury and Osteoarthritis Outcome Score (KOOS)⁴⁷ were used to determine joint function and the ability to perform sports activities. Clinical outcome scores were collected by a special nurse who was blinded to the intention of this study. Radiographs of the knee joints were obtained and included AP views, true-lateral views at 30° of knee flexion, and AP long-leg weightbearing views. The femorotibial angle (the angle between the femoral and tibial shaft axes on AP radiographs)⁴⁰ and posterior tibial slope (the angle formed by the tangent to the medial tibial plateau and the line perpendicular to the tangent at the anterior tibial cortex) with the knee in 30° of flexion³⁹ were measured to analyze the mechanical effects of HTO.

MRI Evaluation

Preoperative and follow-up MRI was performed using a 3.0-T MRI scanner (Achieva 3.0-T SE; Philips) with a dedicated 8-channel knee coil. Follow-up MRI was performed at a mean of 18.5 months postoperatively. The following sequences were utilized: (1) proton density (PD) spectral presaturation with inversion recovery (SPIR) transversal image (repetition time/echo time [TR/TE], 4000/15 milliseconds; field of view [FOV], 150×150 mm; matrix, 308×249; slice thickness [SL], 3.5 mm with 0.35-mm gap), (2) PD SPIR coronal image (TR/TE, 3500/15 milliseconds; FOV, 150×150 mm; matrix, 260×240; SL, 3.0 mm with 0.5-mm gap), (3) T2 SPIR sagittal image (TR/TE, 3200/70 milliseconds; FOV, 150×150 mm; matrix, 240×192; SL, 3.0 mm with 0.3-mm gap), and (4) turbo spin echo T1-weighted sagittal image (TR/TE, 600/20 milliseconds; FOV, 150×150; matrix, 240×240; SL, 3.0 mm with 0.3-mm gap). To avoid potential bias, an independent observer, who was a musculoskeletal radiologist not involved in the care of patients and blinded to the intention of this study, evaluated the MRI scans. The size of the cartilage lesion was measured using preoperative MRI. On follow-up MRI, the MOCART (magnetic resonance observation of cartilage repair tissue) scoring system was used for the evaluation of repaired cartilage (Table 2).³⁶

Statistical Analysis

The principal dependent variables were the Lysholm score and KOOS at the final follow-up (clinical outcomes) as well as the postoperative MOCART score (MRI outcomes). Data are reported as means \pm SDs unless otherwise indicated. The Wilcoxon signed-rank test was used to evaluate differences between the preoperative and final follow-up values. To assess the association of patients' demographic characteristics with clinical and MRI outcomes, each category was divided into groups: age (<50, 50-59, 60-69, and \geq 70 years), sex (male/female), side of involvement (right/left), body mass index (BMI; <20.0, 20.0-24.9, 25.0-29.9, and $>30.0 \text{ kg/m}^2$), cartilage lesion size (femoral condyle: <6.0, 6.0-6.9, 7.0-7.9, and \geq 8.0 cm²; tibial plateau: <5.0, 5.0-5.9, 6.0-6.9, and \geq 7.0 cm²), and number of MSCs (<1.0×10⁷, 1.0×10⁷ to 1.5×10^7 , and $\geq 1.6 \times 10^7$). Differences between the groups were analyzed using the Mann-Whitney U test or the Kruskal-Wallis test for multiple comparisons.

The Spearman rank-order correlation test was used to evaluate the potential bivariate associations between the different factors to assess whether a statistically significant correlation existed. Multivariate logistic regression

TABLE 2 MOCART Scoring System^a

Variable	Score
Degree of defect repair and filling of defect	
Complete	20
Hypertrophy	15
Incomplete	
$\geq\!50\%$ of adjacent cartilage	10
${<}50\%$ of adjacent cartilage	5
Subchondral bone exposed	0
Integration to border zone	
Complete	15
Incomplete	
Demarcating border visible	10
Defect visible	
${<}50\%$ of length of repair tissue	5
$\geq 50\%$ of length of repair tissue	0
Surface of repair tissue	
Surface intact	10
Surface damaged	
<50% of repair tissue depth or total degeneration	5
\geq 50% of repair tissue depth or total degeneration	0
Structure of repair tissue	
Homogeneous	5
Inhomogeneous or cleft formation	0
Signal intensity of repair tissue	
Normal (identical to adjacent cartilage)	30
Nearly normal (slight areas of signal alteration)	15
Abnormal (large areas of signal alteration)	0
Subchondral lamina	
Intact	5
Not intact	0
Subchondral bone	
Intact	5
Not intact	0
Adhesions	
No	5
Yes	0
Effusion	
No	5
Yes	0
Total	100

 $^a\mathrm{MOCART},$ magnetic resonance observation of cartilage repair tissue.

analyses were used to assess the factors that were independently associated with unsatisfactory clinical outcomes. We defined unsatisfactory clinical outcomes as a Lysholm score of <83 or KOOS subscores of <74 (Pain), 72 (Symptoms), 75 (Activities of Daily Living [ADL]), 54 (Sports and Recreation [Sports/Rec]), or 62 (Quality of Life [QoL]) because the mean Lysholm score at the final follow-up was 82.9, and the mean KOOS subscores were 73.4 for pain, 71.4 for symptom, 74.1 for ADL, 53.2 for Sports/Rec, and 61.1 for QoL, respectively. We calculated odds ratios and 95% CIs relative to a chosen reference group for the logistic regression models. Linear regression analyses were also used to evaluate the correlation between number of MSCs and MOCART score. Statistical analyses were performed using SPSS (Version 13.0; IBM), with significance defined as P < .05.

RESULTS

Clinical, Radiographic, and MRI Outcomes

Before surgery, the mean clinical outcome scores were 55.1 ± 3.7 (Lysholm), 43.2 ± 2.4 (KOOS-Pain), 41.4 ± 2.4 (KOOS-Symptoms), 50.8 ± 2.2 (KOOS-ADL), 24.0 ± 4.4 (KOOS-Sports/Rec), and 39.2 ± 4.1 (KOOS-QoL). At the final follow-up, all mean scores improved significantly to $82.9 \pm$ 3.8 (Lysholm), 73.4 ± 3.4 (KOOS-Pain), 71.4 ± 3.4 (KOOS-Symptoms), 74.1 ± 4.6 (KOOS-ADL), 53.2 ± 3.2 (KOOS-Sports/Rec), and 61.1 ± 4.5 (KOOS-QoL) (P < .001 for all). In addition, no clinically significant adverse event was noted. Radiographic outcomes at the final follow-up showed that knee joint alignment had been corrected. The mean femorotibial angle and posterior tibial slope were significantly changed from varus $3.3^{\circ} \pm 1.9^{\circ}$ and $10.0^{\circ} \pm 1.0^{\circ}$ to valgus $8.8^{\circ} \pm 2.4^{\circ}$ and $10.3^{\circ} \pm 2.7^{\circ}$, respectively (P < .001 for both). Follow-up MRI was performed at a mean of 18.5 months postoperatively (Figure 2). Before surgery, the mean MOCART score was 35.7 ± 9.3 for the femoral condyle and 35.2 ± 9.7 for the tibial plateau. On follow-up MRI, the mean MOCART score improved significantly to 74.2 ± 8.6 for the femoral condyle and 74.1 ± 7.5 for the tibial plateau (P < .001 for both).

Correlations Among Clinical, Radiographic, and MRI Outcomes

Significant correlations were found between the final clinical and MRI outcomes (all P < .05) (Table 3). The Lysholm and KOOS scores improved significantly as the level of repaired cartilage improved. However, the radiographic outcomes were not significantly correlated with clinical outcomes at the final follow-up. In addition, no significant correlation between the postoperative clinical, radiographic, and MRI outcomes was observed (Table 4).

Association Between Patient Demographics and Clinical and MRI Outcomes

Significant differences were found in clinical outcomes and MOCART scores between the different age groups, with the exception of the KOOS-ADL and KOOS-QoL (Table 5). Significant differences were also found in clinical outcomes and MOCART scores with respect to the number of MSCs (Table 6). However, no significant differences were found in clinical outcomes and MOCART scores with respect to patient sex, side of involvement, BMI, or lesion size (Appendix Tables A1-A5).

Multivariate logistic regression analyses were used to assess the factors that were independently associated with unsatisfactory clinical outcomes. All multivariate models were adjusted for differences in age and number of MSCs (Table 7). Patient age and number of MSCs were found to be independent predictors of clinical outcomes of MSC implantation with concomitant HTO (P = .014 and P = .037, respectively). Compared with patients aged <50 years, those aged 50-59 years were 2.211 times more likely to have unsatisfactory clinical outcomes (95% CI, 0.223-21.942),



Figure 2. (A) Preoperative anteroposterior (AP) long-leg weightbearing view. Varus malalignment of the knee joint is observed. (B, C) Preoperative fat-saturated proton density coronal and sagittal images of the right knee of a 64-year-old female patient. Cartilage loss is observed in the medial femoral condyle and tibial plateau. (D) Follow-up AP long-leg weightbearing view shows the correction of varus malalignment of the knee joint. (E, F) Follow-up AP fat-saturated proton density coronal and sagittal images at 18 months after surgery. Cartilage regeneration along with favorable integration with adjacent native cartilage is observed.

TABLE 3 Correlation Between Clinical and MRI Outcomes a			
	MOCART	Score	
	Femoral Condyle	Tibial Platea	

	Femoral Condyle		Tibial	Plateau
	$r_{ m S}$	Р	$r_{\rm S}$	Р
Lysholm score KOOS	0.324	.005	0.392	<.001
Pain Symptoms Activities of Daily Living Sports and Recreation Quality of Life	0.477 0.320 0.568 0.579 0.332	< .001 .005 < .001 < .001 .004	$\begin{array}{c} 0.415 \\ 0.312 \\ 0.484 \\ 0.457 \\ 0.235 \end{array}$	<.001 .006 <.001 <.001 .043
Quality of Life	0.002	.004	0.250	.045

^aCalculated using the Spearman rank-order test. KOOS, Knee injury and Osteoarthritis Outcome Score; MOCART, magnetic resonance observation of cartilage repair tissue; MRI, magnetic resonance imaging.

those aged 60-69 years were 10.599 times more likely to have unsatisfactory clinical outcomes (95% CI, 1.051-106.886), and those aged \geq 70 years were 18.167 times more likely to have unsatisfactory clinical outcomes (95% CI, 0.771-428.211) after MSC implantation. Compared with patients with \geq 1.6×10⁷ MSCs, those with MSCs between 1.0×10⁷ and 1.5×10⁷ were 2.819 times more likely to have unsatisfactory clinical outcomes (95% CI, 0.813-9.773), and

TABLE 4 Correlation Between Clinical, Radiographic, and MRI $Outcomes^a$

	Femorotibial Angle		Posterior Tibia Slope	
	$r_{\rm S}$	Р	$r_{\rm S}$	Р
Lysholm score	-0.155	.184	0.038	.745
KOOS				
Pain	-0.199	.655	-0.163	.163
Symptoms	0.110	.087	0.006	.956
Activities of Daily Living	0.048	.346	0.109	.351
Sports and Recreation	-0.053	.683	0.109	.350
Quality of Life	-0.040	.732	0.049	.678
MOCART score				
Femoral condyle	-0.056	.633	0.034	.774
Tibial plateau	-0.112	.340	0.010	.935

^{*a*}Calculated using the Spearman rank-order test. KOOS, Knee injury and Osteoarthritis Outcome Score; MOCART, magnetic resonance observation of cartilage repair tissue; MRI, magnetic resonance imaging.

those with $<1.0\times10^7$ MSCs were 5.153 times more likely to have unsatisfactory clinical outcomes (95% CI, 1.118-23.753) after surgery. The bivariate correlation analysis showed a statistically significant association between age and number of MSCs ($r_{\rm S} = -0.479$; P < .001).

Clinical and Mrki Outcomes Stratified by Age					
	$<\!50 \ y \ (n=8)$	50-59 y $(n = 30)$	$60-69 \ y \ (n=31)$	$\geq 70 \ y \ (n=6)$	P Value ^{b}
Lysholm score	85.2 ± 3.8	84.1 ± 4.0	82.4 ± 3.1	79.6 ± 3.9	.022
KOOS					
Pain	76.2 ± 4.1	74.6 ± 2.8	72.4 ± 3.1	70.6 ± 3.5	.002
Symptoms	74.5 ± 3.5	72.2 ± 3.2	70.7 ± 3.0	68.8 ± 2.8	.005
Activities of Daily Living	77.0 ± 5.1	75.1 ± 4.2	72.8 ± 4.7	73.0 ± 3.6	.120
Sports and Recreation	56.0 ± 3.8	53.6 ± 3.2	53.0 ± 2.9	50.5 ± 1.9	.015
Quality of Life	64.5 ± 3.8	61.2 ± 4.5	60.8 ± 4.5	59.4 ± 3.9	.143
MOCART score					
Femoral condyle	80.8 ± 11.1	76.8 ± 7.5	71.8 ± 7.3	68.1 ± 10.0	.006
Tibial plateau	80.0 ± 8.4	76.8 ± 6.4	71.5 ± 7.1	69.4 ± 6.2	.002

TABLE 5 Clinical and MRI Outcomes Stratified by Age^a

^aValues are expressed as mean ± SD. KOOS, Knee injury and Osteoarthritis Outcome Score; MOCART, magnetic resonance observation of cartilage repair tissue; MRI, magnetic resonance imaging.

^bKruskal-Wallis test.

TABLE 6
Clinical and MRI Outcomes Stratified by Number of MSCs ^a

	$\substack{< 1.0 \times 10^7 \\ (n = 29)}$	$\begin{array}{c} 1.0{\times}10^7 \text{ to} \\ 1.5{\times}10^7 \\ (n=27) \end{array}$	$\begin{array}{c} \geq \! 1.6 {\times} 10^7 \\ (n=19) \end{array}$	PValue ^b
Lysholm score KOOS	81.3 ± 2.6	82.1 ± 3.3	84.2 ± 5.7	.034
Pain	71.2 ± 2.5	73.9 ± 3.0	76.0 ± 3.2	< .001
Symptoms	70.3 ± 2.8	71.1 ± 2.8	73.6 ± 3.9	.003
Activities of Daily Living	71.8 ± 3.7	74.4 ± 4.2	77.1 ± 4.6	.001
Sports and Recreation	51.2 ± 1.9	53.3 ± 2.5	56.3 ± 3.2	<.001
Quality of Life MOCART score	59.7 ± 4.2	60.9 ± 4.0	63.6 ± 4.6	.013
Femoral condyle	66.0 ± 5.7	76.5 ± 3.6	83.4 ± 5.3	<.001
Tibial plateau	67.8 ± 5.1	75.7 ± 4.5	81.6 ± 5.5	<.001

^{*a*}Values are expressed as mean \pm SD. KOOS, Knee injury and Osteoarthritis Outcome Score; MOCART, magnetic resonance observation of cartilage repair tissue; MRI, magnetic resonance imaging; MSC, mesenchymal stem cell.

^bKruskal-Wallis test.

As illustrated in Figure 3, the number of MSCs was 1.2×10^7 for the mean postoperative MOCART score (74.2 for the femoral condyle and 74.1 for the tibial plateau). According to the theoretical extrapolation of the correlation, the predicted number of MSCs for the highest possible MOCART score of 100 points was 2.5×10^7 for the femoral condyle and 2.8×10^7 for the tibial plateau.

DISCUSSION

The most important finding of this study was that patient age and number of MSCs were the principal prognostic factors that were significantly associated with the outcomes of MSC implantation with concomitant HTO (P = .014 and

TABLE 7
Association of Patient Age and Number of MSCs
With Clinical $Outcomes^a$

	n (%)	Unsatisfactory Clinical Outcomes, Odds Ratio (95% CI)	<i>P</i> Value
Age group, y			.014
$<\!\!50$	8 (8.0)	1.000	
50-59	30 (41.3)	2.211(0.223-21.942)	
60-69	31 (40.0)	10.599 (1.051-106.886)	
≥ 70	6 (10.7)	$18.167\ (0.771-428.211)$	
No. of MSCs			.037
${<}1.0{ imes}10^{7}$	29 (38.7)	5.153(1.118 - 23.753)	
$1.0{ imes}10^7$ to $1.5{ imes}10^7$	27(36.0)	2.819 (0.813-9.773)	
$\geq \! 1.6 { imes} 10^7$	$19\ (25.3)$	1.000	

^aMSC, mesenchymal stem cell.

P = .037, respectively). Additionally, a statistically significant association was found between patient age and number of MSCs ($r_{\rm S} = -0.479$; P < .001). Although the application of MSCs is known to be beneficial for better cartilage regeneration, with improved clinical outcomes in patients undergoing concomitant HTO for varus knee OA,^{4,25,28,49,55} we understand little about the contribution of known influential preoperative factors on the outcomes of MSC implantation with concomitant HTO. To our knowledge, this is the first study to assess the effect of patient demographic characteristics on the clinical and MRI outcomes after such a procedure.

Patient age is the main risk factor of OA.^{37,42,45} In their study of 3,266,826 participants, Prieto-Alhambra et al⁴⁵ reported that the incidence rates of knee OA increased progressively with age, with the steepest slope in those aged 50-70 years, which then slowed down and peaked in those aged 75-85 years, with a slight decrease in the final years of life (\geq 85 years). Advanced age is not only a significant risk factor for OA, but it also affects the quality of MSCs.¹⁰ Several studies have described an age-dependent effect on



Figure 3. Correlation between the number of mesenchymal stem cells (MSCs) and postoperative MOCART (magnetic resonance observation of cartilage repair tissue) score for the (A) femoral condyle and (B) tibial plateau.

the properties of MSCs.^{6,7} Chang et al⁶ compared the number and function of MSCs in articular cartilage among human fetuses, healthy adults (aged 28-45 years), and elderly adults (aged 60-75 years), and they found that MSCs accounted for 94.69%, 4.85%, and 6.33% of the cells in articular cartilage, respectively (P < .001). They also reported that a lower chondrogenic differentiation of MSCs derived from elderly patients might be associated with the development of OA.⁶ Choudhery et al⁷ investigated the expansion and in vitro differentiation potential of MSCs in younger (<30 years), adult (35-50 years), and older (>60 years) patients, and they found that older MSCs displayed more senescent features than MSCs that were isolated from younger donors, concomitant with reduced viability, proliferation, and differentiation potentials. In the present study, significant differences were found in clinical and MRI outcomes among the 4 age groups, with the exception of KOOS-ADL and KOOS-QoL subscores (Table 5). In addition, we found that age was an independent predictor of clinical outcomes of MSC implantation (P = .014)(Table 7), and a significant association between patient age and number of MSCs was also found ($r_{\rm S}=-0.479; P<.001$). We consider that these findings are a consequence of the less favorable quality of the MSCs in older patients.

Obesity is a well-established risk factor for the development and progression of OA, especially in weightbearing joints.^{10,37} Furthermore, obesity is known to reduce the capability of MSCs. Louwen et al³⁴ demonstrated that MSCs in obese patients have impaired functionalities including differentiation, proangiogenesis, motility, and immunomodulation, accompanied by alterations in their own undifferentiated state and metabolism. Roldan et al⁴⁶ examined MSCs from obese patients and showed a reduced proliferation rate, greater cell senescence, and reduced differentiation to multiple lineages, including chondrogenesis. In the present study, we divided the patients into 4 groups according to BMI, and we found no significant differences in the clinical and MRI outcomes among the BMI groups (Appendix Table A3). According to the 1998 "Clinical Guidelines on the Identification, Evaluation, and Treatment of Overweight and Obesity in Adults,"8 overweight is defined as a BMI of 25.0 to 29.9 kg/m² and obesity as a BMI of >30.0 kg/m². In the present study, only 4 patients were in the obesity group, which means that MSCs from obese patients would not have influenced the outcomes; thus, no significant differences in the clinical and MRI outcomes were found among the BMI groups. Further studies comparing the outcomes between different BMI groups, including more patients with a BMI of \geq 30 kg/m², are needed to evaluate the independent effect of BMI on clinical and MRI outcomes of MSC implantation with concomitant HTO.

Although several clinical studies have focused on the use of MSCs as a cell-based treatment for OA, only a few have reported cartilage lesion size as a prognostic factor in using MSCs to treat knee OA.^{22,23,27} Koh et al²⁷ performed MSC implantation under arthroscopic guidance and found that large lesion size ($\geq 5.4 \text{ cm}^2$) was a factor that resulted in incomplete cartilage repair and poor clinical outcomes. Kim et al²³ evaluated 54 patients (56 knees) who underwent second-look arthroscopic surgery after MSC implantation for cartilage lesions in their OA knees and reported that large lesion size $(\geq 5.7 \text{ cm}^2)$ was a significant predictor of poor clinical and arthroscopic outcomes. Kim et al²² evaluated 49 patients (55 knees) who underwent MSC implantation for knee OA and suggested that a lesion size of 6.0 cm² was the upper limit for obtaining encouraging outcomes of MSC implantation.

In the present study, we found no significant differences in clinical and MRI outcomes between lesion size groups (Appendix Tables A4 and A5). In this study, the mean lesion sizes were 7.1 cm² for the femoral condyle and 6.2 cm² for the tibial plateau, which were larger than those in previous studies.^{22,23,27} Furthermore, the distribution boundary of the lesion size was narrow compared with that of previous studies,^{22,23,27} in which most lesion sizes were $\geq 6.0 \text{ cm}^2$ (61/75 knees) for the femoral condyle and $\geq 5.0 \text{ cm}^2$ (69/75 knees) for the tibial plateau—definitely large lesion sizes considering the medial tibiofemoral joint contact area reported by Tummala et al.⁵³ Therefore, we speculate that lesion size did not influence the clinical and MRI outcomes of surgery in the current study.

Several studies have documented the biomechanical improvements in the knee joint that can be achieved by restoring joint orientation and axial alignment, with satisfactory clinical outcomes of HTO for varus knee OA.^{3,17,31,48} Although cartilage regeneration in the medial compartment of the knee joint has been reported to be achieved after HTO, ^{19,51} the quality of the regenerated cartilage was mostly insufficient for the load-bearing functions of articular cartilage.²⁰ In addition, a positive correlation between cartilage regeneration and clinical outcomes after HTO has been reported,^{28,30,43} indicating that the biochemical properties of the regenerated cartilage are as important as its biomechanical properties. In this regard, conventional cartilage repair procedures with concomitant HTO have been performed to obtain better cartilage regeneration^{15,44}; however, such procedures have only been shown to stimulate the growth of fibrocartilage, which cannot withstand mechanical stresses as well as native hyaline cartilage.^{32,38} Thus, several recent studies have suggested the application of MSCs for superior cartilage regeneration, reporting satisfactory clinical outcomes of concurrent MSC therapy with HTO.^{4,24,25,28,49,55} Moreover, Saw et al⁴⁹ performed a histological evaluation after chondrogenesis with stem cells in combination with HTO, and they found that the regenerated cartilage closely resembled native articular cartilage. From this viewpoint, we believe that MSC implantation plays a role in the restoration of the essential biomechanical and biochemical properties of diseased cartilage in the medial compartment of the knee joint.

A review of the literature revealed that the number of MSCs used for the treatment of OA is another important prognostic factor of the outcomes.^{1,16} According to Afizah and Hui,¹ who reviewed the use of bone marrow-derived MSCs for OA treatment, studies used the following numbers of MSCs: 8×10^6 as the lowest number⁹ and 1.46×10^7 , ⁵⁵ 1.3×10^{7} , 5^{4} 2.0×10^{7} to 2.4×10^{7} , 1^{12} and 4×10^{7} as the highest numbers, ⁴¹ and markedly better results were observed in most studies^{12,41,54,55} except for the study that used the lowest number of MSCs (8×10^6) .⁹ Considering these results, Afizah and Hui concluded that $>1 \times 10^7$ of bone marrow-derived MSCs are required to achieve significantly better repair. In terms of adipose-derived MSCs, Kim and Koh²⁵ reported improved cartilage regeneration with better clinical outcomes after injection of adiposederived MSCs (4.26×10^6) with concurrent HTO for varus knee OA. Jo et al 18 performed intra-articular injection of adipose-derived MSCs with 3 different amounts of MSCs $(1 \times 10^7, 5 \times 10^7, \text{ and } 1 \times 10^8)$ for knee OA and found that only

patients injected with 1×10^8 MSCs demonstrated a decrease in articular cartilage defects by the regeneration of hyaline-like articular cartilage.

In the present study, 1.19×10^7 cells were used, on average, for MSC implantation, and we assessed whether the number of MSCs influenced the clinical and MRI outcomes of MSC implantation. Accordingly, we found significant differences in clinical and MRI outcomes among groups according to the number of MSCs (Table 6). Furthermore, we found that number of MSCs was an independent predictor of clinical outcomes (Table 7). Although the optimal number of MSCs to be applied remains unknown, we believe that a larger number is required to obtain adequate cartilage regeneration. We used bivariate correlation analysis to predict the required number of MSCs for a MOCART score of 100 points, and our results suggested that an approximate minimum of 2.5×10^7 cells is necessary to obtain encouraging cartilage regeneration after MSC implantation with concomitant HTO (see Figure 3).

This study had some limitations. First, the small number of study participants may have limited the power to detect other prognostic factors. For example, given our sample size, we cannot conclude that there is no relationship between lesion size or BMI and our primary clinical and MRI outcomes. However, given that no similar studies of this size have been published, we believe that these data are valuable for the assessment of prognostic factors influencing the outcomes of MSC implantation with concomitant HTO. In addition, this ongoing study can be strengthened in the future, as the number of patients who underwent this treatment will increase over time. Second, because of the retrospective nature of this study, specific patient information that may have improved the quality of the data reported, such as underlying diabetes, smoking status, and the use of anti-inflammatory medication, could not be assessed.

A third limitation was that we did not conduct a histological evaluation to assess the quality of regenerated cartilage. Because MSCs are a heterogeneous population of cells with variable growth potentials and distinct morphological and functional characteristics,¹⁴ the quality of MSCs needed to achieve adequate cartilage regeneration should be identified to predict the outcomes of MSC implantation. In this study, we found that number of MSCs was a prognostic factor that influenced clinical outcomes, and the optimal number of MSCs is expected to obtain better cartilage regeneration. However, a future study that estimates other characteristics of MSCs that influence the clinical outcomes of MSC implantation is needed to more accurately assess influential prognostic factors. Finally, follow-up MRI was performed at about 18 months postoperatively. It is unknown how regenerated cartilage will behave over time, and changes in the influential factors after 18 months cannot be predicted.

CONCLUSION

The study results showed improved clinical and MRI outcomes of MSC implantation with concomitant HTO for varus knee OA. Furthermore, patient age and number of MSCs were important factors that influenced the clinical and MRI outcomes. Identifying these factors may provide a more accurate screening tool for surgeons to better assess which patients are good candidates for MSC implantation with concomitant HTO.

REFERENCES

- 1. Afizah H, Hui JH. Mesenchymal stem cell therapy for osteoarthritis. *J Clin Orthop Trauma*. 2016;7(3):177-182.
- Amendola A, Bonasia DE. Results of high tibial osteotomy: review of the literature. *Int Orthop.* 2010;34(2):155-160.
- Bode G, von Heyden J, Pestka J, et al. Prospective 5-year survival rate data following open-wedge valgus high tibial osteotomy. *Knee Surg Sports Traumatol Arthrosc.* 2015;23(7):1949-1955.
- Cavallo M, Sayyed-Hosseinian SH, Parma A, Buda R, Mosca M, Giannini S. Combination of high tibial osteotomy and autologous bone marrow derived cell implantation in early osteoarthritis of knee: a preliminary study. *Arch Bone Joint Surg.* 2018;6(2):112-118.
- Cerejo R, Dunlop DD, Cahue S, Channin D, Song J, Sharma L. The influence of alignment on risk of knee osteoarthritis progression according to baseline stage of disease. *Arthritis Rheum.* 2002; 46(10):2632-2636.
- Chang HX, Yang L, Li Z, Chen G, Dai G. Age-related biological characterization of mesenchymal progenitor cells in human articular cartilage. *Orthopedics*. 2011;34(8):e382-e388.
- Choudhery MS, Badowski M, Muise A, Pierce J, Harris DT. Donor age negatively impacts adipose tissue-derived mesenchymal stem cell expansion and differentiation. *J Transl Med.* 2014;12:8.
- Clinical guidelines on the identification, evaluation, and treatment of overweight and obesity in adults: the evidence report. National Institutes of Health. Obes Res. 1998;6(suppl_2):51S-209S.
- Davatchi F, Abdollahi BS, Mohyeddin M, Shahram F, Nikbin B. Mesenchymal stem cell therapy for knee osteoarthritis: preliminary report of four patients. *Int J Rheum Dis.* 2011;14(2):211-215.
- Diekman BO, Guilak F. Stem cell-based therapies for osteoarthritis: challenges and opportunities. *Curr Opin Rheumatol.* 2013;25(1): 119-126.
- Dugdale TW, Noyes FR, Styer D. Preoperative planning for high tibial osteotomy: the effect of lateral tibiofemoral separation and tibiofemoral length. *Clin Orthop Relat Res.* 1992;274:248-264.
- Emadedin M, Aghdami N, Taghiyar L, et al. Intra-articular injection of autologous mesenchymal stem cells in six patients with knee osteoarthritis. Arch Iran Med. 2012;15(7):422-428.
- Ferruzzi A, Buda R, Cavallo M, Timoncini A, Natali S, Giannini S. Cartilage repair procedures associated with high tibial osteotomy in varus knees: clinical results at 11 years' follow-up. *Knee*. 2014;21(2): 445-450.
- Harrell CR, Markovic BS, Fellabaum C, Arsenijevic A, Volarevic V. Mesenchymal stem cell-based therapy of osteoarthritis: current knowledge and future perspectives. *Biomed Pharmacother*. 2019; 109:2318-2326.
- Harris JD, McNeilan R, Siston RA, Flanigan DC. Survival and clinical outcome of isolated high tibial osteotomy and combined biological knee reconstruction. *Knee*. 2013;20(3):154-161.
- Im GI. Tissue engineering in osteoarthritis: current status and prospect of mesenchymal stem cell therapy. *BioDrugs*. 2018;32(3): 183-192.
- Jacquet C, Gulagaci F, Schmidt A, et al. Opening wedge high tibial osteotomy allows better outcomes than unicompartmental knee arthroplasty in patients expecting to return to impact sports. *Knee Surg Sports Traumatol Arthrosc.* 2020;28(12):3849-3857.
- Jo CH, Lee YG, Shin WH, et al. Intra-articular injection of mesenchymal stem cells for the treatment of osteoarthritis of the knee: a proofof-concept clinical trial. *Stem Cells*. 2014;32(5):1254-1266.

- Jung WH, Takeuchi R, Chun CW, et al. Second-look arthroscopic assessment of cartilage regeneration after medial opening-wedge high tibial osteotomy. *Arthroscopy*. 2014;30(1):72-79.
- Kanamiya T, Naito M, Hara M, Yoshimura I. The influences of biomechanical factors on cartilage regeneration after high tibial osteotomy for knees with medial compartment osteoarthritis: clinical and arthroscopic observations. *Arthroscopy*. 2002;18(7):725-729.
- 21. Kellgren JH, Lawrence JS. Radiological assessment of osteo-arthrosis. *Ann Rheum Dis.* 1957;16(4):494-502.
- Kim YS, Choi YJ, Koh YG. Mesenchymal stem cell implantation in knee osteoarthritis: an assessment of the factors influencing clinical outcomes. *Am J Sports Med.* 2015;43(9):2293-2301.
- Kim YS, Choi YJ, Suh DS, et al. Mesenchymal stem cell implantation in osteoarthritic knees: is fibrin glue effective as a scaffold? *Am J Sports Med.* 2015;43(1):176-185.
- Kim YS, Chung PK, Suh DS, Heo DB, Tak DH, Koh YG. Implantation of mesenchymal stem cells in combination with allogenic cartilage improves cartilage regeneration and clinical outcomes in patients with concomitant high tibial osteotomy. *Knee Surg Sports Traumatol Arthrosc.* 2020;28(2):544-554.
- Kim YS, Koh YG. Comparative matched-pair analysis of open-wedge high tibial osteotomy with versus without an injection of adiposederived mesenchymal stem cells for varus knee osteoarthritis: clinical and second-look arthroscopic results. *Am J Sports Med*. 2018;46(11): 2669-2677.
- Kocher MS, Steadman JR, Briggs KK, Sterett WI, Hawkins RJ. Reliability, validity, and responsiveness of the Lysholm knee scale for various chondral disorders of the knee. *J Bone Joint Surg Am*. 2004;86(6):1139-1145.
- Koh YG, Choi YJ, Kwon OR, Kim YS. Second-look arthroscopic evaluation of cartilage lesions after mesenchymal stem cell implantation in osteoarthritic knees. Am J Sports Med. 2014;42(7):1628-1637.
- Koh YG, Kwon OR, Kim YS, Choi YJ. Comparative outcomes of openwedge high tibial osteotomy with platelet-rich plasma alone or in combination with mesenchymal stem cell treatment: a prospective study. *Arthroscopy*. 2014;30(11):1453-1460.
- Kong L, Zheng LZ, Qin L, Ho KKW. Role of mesenchymal stem cells in osteoarthritis treatment. J Orthop Translat. 2017;9:89-103.
- Koshino T, Wada S, Ara Y, Saito T. Regeneration of degenerated articular cartilage after high tibial valgus osteotomy for medial compartmental osteoarthritis of the knee. *Knee*. 2003;10(3):229-236.
- Kunze KN, Beletsky A, Hannon CP, et al. Return to work and sport after proximal tibial osteotomy and the effects of opening versus closing wedge techniques on adverse outcomes: a systematic review and meta-analysis. *Am J Sports Med.* 2020;48(9):2295-2304.
- LaPrade RF, Bursch LS, Olson EJ, Havlas V, Carlson CS. Histologic and immunohistochemical characteristics of failed articular cartilage resurfacing procedures for osteochondritis of the knee: a case series. *Am J Sports Med*. 2008;36(2):360-368.
- LaPrade RF, Spiridonov SI, Nystrom LM, Jansson KS. Prospective outcomes of young and middle-aged adults with medial compartment osteoarthritis treated with a proximal tibial opening wedge osteotomy. *Arthroscopy*. 2012;28(3):354-364.
- Louwen F, Ritter A, Kreis NN, Yuan J. Insight into the development of obesity: functional alterations of adipose-derived mesenchymal stem cells. *Obes Rev.* 2018;19(7):888-904.
- Marchal JA, Picon M, Peran M, et al. Purification and long-term expansion of multipotent endothelial-like cells with potential cardiovascular regeneration. *Stem Cells Dev.* 2012;21(4):562-574.
- Marlovits S, Singer P, Zeller P, Mandl I, Haller J, Trattnig S. Magnetic resonance observation of cartilage repair tissue (MOCART) for the evaluation of autologous chondrocyte transplantation: determination of interobserver variability and correlation to clinical outcome after 2 years. *Eur J Radiol.* 2006;57(1):16-23.
- Martel-Pelletier J, Barr AJ, Cicuttini FM, et al. Osteoarthritis. Nat Rev Dis Primers. 2016;2:16072.
- Mithoefer K, Williams RJ 3rd, Warren RF, et al. The microfracture technique for the treatment of articular cartilage lesions in the knee:

a prospective cohort study. *J Bone Joint Surg Am*. 2005;87(9): 1911-1920.

- Moore TM, Harvey JP Jr. Roentgenographic measurement of tibialplateau depression due to fracture. *J Bone Joint Surg Am.* 1974;56(1): 155-160.
- Ogata K, Yoshii I, Kawamura H, Miura H, Arizono T, Sugioka Y. Standing radiographs cannot determine the correction in high tibial osteotomy. J Bone Joint Surg Br. 1991;73(6):927-931.
- Orozco L, Munar A, Soler R, et al. Treatment of knee osteoarthritis with autologous mesenchymal stem cells: a pilot study. *Transplantation*. 2013;95(12):1535-1541.
- Palazzo C, Nguyen C, Lefevre-Colau MM, Rannou F, Poiraudeau S. Risk factors and burden of osteoarthritis. *Ann Phys Rehabil Med*. 2016;59(3):134-138.
- Parker DA, Beatty KT, Giuffre B, Scholes CJ, Coolican MR. Articular cartilage changes in patients with osteoarthritis after osteotomy. *Am J Sports Med.* 2011;39(5):1039-1045.
- Pascale W, Luraghi S, Perico L, Pascale V. Do microfractures improve high tibial osteotomy outcome? Orthopedics. 2011;34(7):e251-e255.
- 45. Prieto-Alhambra D, Judge A, Javaid MK, Cooper C, Diez-Perez A, Arden NK. Incidence and risk factors for clinically diagnosed knee, hip and hand osteoarthritis: influences of age, gender and osteoarthritis affecting other joints. *Ann Rheum Dis.* 2014;73(9):1659-1664.
- Roldan M, Macias-Gonzalez M, Garcia R, Tinahones FJ, Martin M. Obesity short-circuits stemness gene network in human adipose multipotent stem cells. *FASEB J*. 2011;25(12):4111-4126.
- Roos EM, Roos HP, Lohmander LS, Ekdahl C, Beynnon BD. Knee Injury and Osteoarthritis Outcome Score (KOOS): development of a self-administered outcome measure. J Orthop Sports Phys Ther. 1998;28(2):88-96.

- Salzmann GM, Ahrens P, Naal FD, et al. Sporting activity after high tibial osteotomy for the treatment of medial compartment knee osteoarthritis. *Am J Sports Med.* 2009;37(2):312-318.
- Saw KY, Anz A, Jee CS, Ng RC, Mohtarrudin N, Ragavanaidu K. High tibial osteotomy in combination with chondrogenesis after stem cell therapy: a histologic report of 8 cases. *Arthroscopy*. 2015;31(10): 1909-1920.
- Schuster P, Geßlein M, Schlumberger M, et al. Ten-year results of medial open-wedge high tibial osteotomy and chondral resurfacing in severe medial osteoarthritis and varus malalignment. *Am J Sports Med.* 2018;46(6):1362-1370.
- Sterett WI, Steadman JR, Huang MJ, Matheny LM, Briggs KK. Chondral resurfacing and high tibial osteotomy in the varus knee: survivorship analysis. *Am J Sports Med.* 2010;38(7):1420-1424.
- 52. Tsukada S, Wakui M. Is overcorrection preferable for repair of degenerated articular cartilage after open-wedge high tibial osteotomy? *Knee Surg Sports Traumatol Arthrosc.* 2017;25(3):785-792.
- Tummala S, Nielsen M, Lillholm M, Christiansen C, Dam EB. Automatic quantification of tibio-femoral contact area and congruity. *IEEE Trans Med Imaging*. 2012;31(7):1404-1412.
- Wakitani S, Imoto K, Yamamoto T, Saito M, Murata N, Yoneda M. Human autologous culture expanded bone marrow mesenchymal cell transplantation for repair of cartilage defects in osteoarthritic knees. Osteoarthritis Cartilage. 2002;10(3):199-206.
- 55. Wong KL, Lee KB, Tai BC, Law P, Lee EH, Hui JH. Injectable cultured bone marrow-derived mesenchymal stem cells in varus knees with cartilage defects undergoing high tibial osteotomy: a prospective, randomized controlled clinical trial with 2 years' follow-up. *Arthroscopy*. 2013;29(12):2020-2028.
- Zuk PA, Zhu M, Mizuno H, et al. Multilineage cells from human adipose tissue: implications for cell-based therapies. *Tissue Eng.* 2001; 7(2):211-228.

APPENDIX

TABLE A1 Clinical and MRI Outcomes Stratified by Patient Sex^{α}

	$\begin{array}{c} Male \\ (n=35) \end{array}$	$\begin{array}{c} Female \\ (n=40) \end{array}$	P Value ^{b}
Lysholm score	82.5 ± 3.9	83.5 ± 3.8	.301
KOOS			
Pain	73.1 ± 3.8	73.7 ± 3.1	.477
Symptoms	71.0 ± 3.5	71.8 ± 3.2	.290
Activities of Daily Living	74.1 ± 5.1	74.2 ± 4.1	.898
Sports and Recreation	53.6 ± 3.0	52.9 ± 3.8	.351
Quality of Life	61.2 ± 4.2	61.0 ± 4.7	.613
MOCART score			
Femoral condyle	73.7 ± 8.9	74.6 ± 8.4	.562
Tibial plateau	73.0 ± 8.2	75.1 ± 6.7	.184

 aValues are expressed as mean \pm SD. KOOS, Knee injury and Osteoarthritis Outcome Score; MOCART, magnetic resonance observation of cartilage repair tissue; MRI, magnetic resonance imaging.

^bMann-Whitney U test.

 TABLE A2

 Clinical and MRI Outcomes

 Stratified by Side of Involvement^a

	Right(n=39)	Left(n=36)	P Value ^{b}
Lysholm score	82.9 ± 3.5	83.2 ± 4.2	.701
KOOS			
Pain	73.4 ± 3.7	73.4 ± 3.0	.919
Symptoms	71.9 ± 3.7	71.0 ± 3.0	.458
Activities of Daily	74.1 ± 4.3	74.1 ± 4.9	.844
Living			
Sports and Recreation	53.1 ± 3.4	53.3 ± 3.0	.781
Quality of Life	60.8 ± 4.5	61.4 ± 4.5	.456
MOCART score			
Femoral condyle	73.2 ± 8.8	75.1 ± 8.4	.211
Tibial plateau	73.6 ± 7.4	74.6 ± 7.6	.343

"Values are expressed as mean \pm SD. KOOS, Knee injury and Osteoarthritis Outcome Score; MOCART, magnetic resonance observation of cartilage repair tissue; MRI, magnetic resonance imaging.

^bMann-Whitney U test.

	$<\!\!20.0 \text{ kg/m}^2(n=4)$	$20.0\text{-}24.9 \text{ kg/m}^2 (n=32)$	$25.0\text{-}29.9 \text{ kg/m}^2 \ (n=35)$	$\geq\!\!30.0 \text{ kg/m}^2(n=4)$	P Value ^{b}		
Lysholm score	87.7 ± 2.5	83.1 ± 3.9	82.9 ± 3.8	82.7 ± 3.8	.225		
KOOS							
Pain	75.3 ± 1.2	72.7 ± 3.7	73.7 ± 3.2	74.3 ± 3.4	.361		
Symptoms	72.0 ± 3.2	70.8 ± 3.3	72.1 ± 3.6	70.6 ± 4.0	.444		
Activities of Daily Living	80.7 ± 3.2	73.7 ± 4.9	73.9 ± 4.1	74.4 ± 4.0	.129		
Sports and Recreation	56.3 ± 4.7	52.9 ± 2.9	53.3 ± 3.5	53.3 ± 2.0	.503		
Quality of Life	62.0 ± 3.2	60.9 ± 4.6	61.2 ± 4.6	61.3 ± 4.6	.879		
MOCART score							
Femoral condyle	83.3 ± 2.9	73.9 ± 8.6	73.9 ± 8.3	72.9 ± 8.6	.205		
Tibial plateau	80.0 ± 7.2	73.9 ± 7.7	73.8 ± 7.6	74.3 ± 7.3	.450		

 $\label{eq:TABLE A3} TABLE \ A3 Clinical \ and \ MRI \ Outcomes \ Stratified \ by \ Body \ Mass \ Index^a$

 a Values are expressed as mean \pm SD. KOOS, Knee injury and Osteoarthritis Outcome Score; MOCART, magnetic resonance observation of cartilage repair tissue; MRI, magnetic resonance imaging.

^bKruskal-Wallis test.

	${<}6.0\ cm^2\ (n=14)$	$6.0-6.9 \text{ cm}^2 (n=23)$	$7.0-7.9 \ cm^2 \ (n=23)$	$\geq\!\!8.0\ cm^2\ (n=15)$	P Value ^{b}
Lysholm score	82.3 ± 4.5	83.4 ± 3.3	83.4 ± 3.0	83.2 ± 3.9	.887
KOOS					
Pain	74.2 ± 3.1	73.0 ± 3.4	73.7 ± 3.6	72.7 ± 3.5	.881
Symptoms	73.4 ± 2.8	71.2 ± 3.6	71.0 ± 3.6	70.2 ± 2.4	.081
Activities of Daily Living	75.3 ± 5.1	72.4 ± 4.7	75.6 ± 4.3	73.4 ± 3.4	.052
Sports and Recreation	55.2 ± 3.9	52.4 ± 3.0	53.2 ± 2.6	52.6 ± 2.9	.095
Quality of Life	62.7 ± 4.5	61.4 ± 5.0	60.8 ± 4.2	59.3 ± 3.7	.172
MOCART score					
Femoral condyle	77.7 ± 8.3	72.2 ± 8.0	75.2 ± 9.6	72.1 ± 7.5	.166
Tibial plateau	76.3 ± 8.3	73.5 ± 6.5	74.1 ± 8.6	72.9 ± 6.1	.439

 $\label{eq:TABLEA4} TABLE \mbox{ A4} Clinical and MRI \mbox{ Outcomes Stratified by Lesion Size of the Femoral Condyle}^a$

 a Values are expressed as mean \pm SD. KOOS, Knee injury and Osteoarthritis Outcome Score; MOCART, magnetic resonance observation of cartilage repair tissue; MRI, magnetic resonance imaging.

^bKruskal-Wallis test.

TABLE A5 Clinical and MRI Outcomes Stratified by Lesion Size of the Tibial Plateau a

	${<}5.0\ cm^2\ (n=6)$	$5.0\text{-}5.9\ \text{cm}^2\ (n=25)$	$6.0\text{-}6.9\ \text{cm}^2\ (n=32)$	${\geq}7.0~cm^2~(n=12)$	P Value ^{b}
Lysholm score	83.5 ± 3.3	82.6 ± 4.1	83.5 ± 4.1	83.3 ± 3.9	.765
KOOS					
Pain	74.5 ± 3.0	73.6 ± 3.3	73.3 ± 3.4	72.6 ± 4.1	.966
Symptoms	74.0 ± 2.8	72.1 ± 3.6	70.9 ± 3.2	70.2 ± 3.1	.061
Activities of Daily Living	74.3 ± 4.3	73.8 ± 5.3	74.7 ± 4.2	72.9 ± 4.5	.386
Sports and Recreation	57.5 ± 3.0	53.1 ± 3.6	53.4 ± 2.8	51.5 ± 2.3	.056
Quality of Life	62.8 ± 5.9	61.8 ± 4.8	61.5 ± 4.2	58.7 ± 2.6	.053
MOCART score					
Femoral condyle	78.8 ± 4.8	73.8 ± 8.9	74.3 ± 9.1	73.2 ± 7.5	.549
Tibial plateau	72.5 ± 6.5	74.6 ± 7.3	73.7 ± 8.2	74.2 ± 7.5	.899

 a Values are expressed as mean \pm SD. KOOS, Knee injury and Osteoarthritis Outcome Score; MOCART, magnetic resonance observation of cartilage repair tissue; MRI, magnetic resonance imaging.

^bKruskal-Wallis test.