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CCAAT/enhancer-binding protein β promotes receptor activator of nuclear factor-kappa-B ligand (RANKL) expression and osteoclast formation in the synovium in rheumatoid arthritis

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Abstract

Introduction: CCAAT/enhancer-binding protein β (C/EBP β) is a transcription factor that is activated in the synovium in rheumatoid arthritis (RA) and promotes expression of various matrix metalloproteinases. In this study, we examined whether C/EBP β mediates the expression of receptor activator of nuclear factor-kappa-B ligand (RANKL) and drives osteoclast formation in primary fibroblast-like synoviocytes (FLS) from RA patients. The cooperation of C/EBP β and activation transcription factor-4 (ATF4) in the regulation of the RANKL promoter was also investigated.

Methods: Immunofluorescence staining was performed for C/EBP β , RANKL, and ATF4 in synovium from RA patients. Adenovirus expression vectors for two major isoforms, C/EBP β -liver-enriched activator protein (LAP) and -liver-enriched inhibitory protein (LIP), or small interfering RNA for C/EBP β , were used to manipulate C/EBP β expression in RA-FLS. RA-FLS over-expressing C/EBP β were co-cultured with peripheral blood mononuclear cells (PBMCs) to test osteoclast formation by tartrate-resistant acid phosphatase (TRAP) staining. A promoter assay for RANKL, a chromatin immunoprecipitation (ChIP) assay and an immunoprecipitation (IP) assay were also performed.

Results: Immunofluorescence staining showed colocalization of C/EBP β , ATF4 and RANKL in RA synovium. Western blotting revealed the expression of C/EBP β -LAP and -LIP in RA-FLS. Over-expression of either C/EBP β -LAP or -LIP significantly increased the expression of RANKL mRNA, while C/EBP β -LIP down-regulated osteoprotegerin (OPG) mRNA. The RANKL/OPG mRNA ratio was significantly increased by C/EBP β -LIP over-expression. Knockdown of C/EBP β with siRNA decreased the expression of RANKL mRNA. The number of TRAP-positive multinucleated cells was increased in co-cultures of PBMCs and FLS over-expressing either C/EBP β -LAP or -LIP, but was more significant with LIP. C/EBP β -LIP does not have a transactivation domain. However, promoter assays showed that C/EBP β -LIP and ATF4 synergistically transactivate the RANKL promoter. ChIP and IP assays revealed the cooperative binding of C/EBP β and ATF4 on the RANKL promoter.

Conclusions: We demonstrated that C/EBP β , especially C/EBP β -LIP in cooperation with ATF4, is involved in osteoclast formation by regulating RANKL expression in RA-FLS. These findings suggest that C/EBP β plays a crucial role in bone destruction in RA joints.

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Introduction

Cartilage degeneration and bone destruction are the main features of rheumatoid arthritis (RA) [1]. Inflammation pathways are involved in the catabolic processes of articular cartilage and bone degeneration in RA. Inflammatory cytokines such as IL-1 β , TNF- α , IL-6, and IL-17 play significant roles in mediating inflammation and joint destruction. These cytokines are expressed in arthritic joints in RA and induce expression of receptor activator of nuclear factor kappa B ligand (RANKL) in the synovium [2]. RANKL is an essential factor for osteoclast differentiation [3,4]. Osteoprotegerin (OPG) is a decoy receptor that inhibits RANKL activation of osteoclastogenesis and reduces bone resorption [5]. RA synovium-induced RANKL stimulates osteoclast differentiation at sites where bone and RA synovial membranes contact each other.

Inflammatory cytokines in RA joints activate numerous transcription factors including nuclear factor-kappa-B (NF- κ B), activator protein-1 (AP-1), janus kinase-signal transducer and activator of transcription (JAK-STAT) and the CCAAT/enhancer-binding protein (C/EBP) family. The C/EBP family consists of six members: C/EBP α , β , δ , ϵ , γ , and ζ [6]. C/EBP β is an intron-less gene and has three major isoforms: 38 kD (liver-enriched activator protein Star (LAP*)), 36 kD (LAP) and 20 kD (liver-enriched inhibitory protein (LIP)) [7]. The isoforms, LAP* and LAP, each contain an N-terminal transactivation domain (TAD) and a chromatin remodeling domain. The LIP isoform lacks the TAD, although it retains DNA binding capability, and is generally recognized to be a dominant negative isoform.

Recent studies indicated that C/EBP β is involved in differentiation of osteoblasts and osteoclasts both physiologically and pathologically. C/EBP β activates osteocalcin gene transcription and promotes osteoblast differentiation [8-10]. For osteoclast differentiation, the C/EBP β isoform ratio in mononuclear cells regulates osteoclastogenesis through V-maf musculoaponeurotic fibrosarcoma oncogene homolog B (MafB) [11]. C/EBP β and RANKL are upregulated in GCT. C/EBP β induces RANKL promoter activity in GCT stromal cells, which causes osteolysis [12]. In inflammatory chronic diseases such as RA, C/EBP β is strongly induced in response to inflammatory stimulation. C/EBP β is expressed in synovial tissues and chondrocytes of RA [13,14]. C/EBP β plays a crucial role in cartilage degradation along with proteolytic enzymes such as matrix metalloproteinase-1 (MMP-1), MMP-3, MMP-13, and aggrecanase-2 (a disintegrin and metalloproteinase with thrombospondin motifs-5: ADAMTS-5) in inflammatory arthritis. Hence, we hypothesized that an imbalance of C/EBP β isoforms may upset skeletal integrity in RA by being involved in both cartilage and bone destruction.

In this paper, we investigated whether C/EBP β mediates the expression of RANKL in RA synovium and

consequently, whether it induces osteoclast formation. In addition, we analyzed the mechanism of RANKL and OPG expression by the C/EBP β isoforms, C/EBP β -LAP and -LIP, and by cooperation with activation transcription factor-4 (ATF4). Determining the mechanisms related to the regulation of RANKL expression and bone resorption by C/EBP β may provide new insights into the development of potential therapies for RA patients.

Methods

Clinical samples

Tissue samples of synovium were obtained from patients with RA at the time of total knee arthroplasty (TKA) or synovectomy. Patients signed informed consent for providing tissue samples for this study. Subjects included seven RA patients (mean age, 60.3 \pm 11.3 years), who fulfilled the 2010 American College of Rheumatology (ACR) and the European League Against Rheumatism (EULAR) diagnostic criteria for RA [15]. All studies were performed under the approval of the Institutional Ethics Board of Kyushu University (approval number: 22-99) and in accordance with the tenets of the Declaration of Helsinki.

Isolation of human fibroblast-like synoviocytes

Human fibroblast-like synoviocytes were isolated from the synovium of RA patients (RA-FLS). Synovial tissues were minced into small pieces and digested with 2 mg/ml collagenase L (Wako, Osaka, Japan) for 90 minutes at 37°C. The collected cells were resuspended in DMEM supplemented with 10% FBS (Gibco, Gaithersburg, MD, USA). Adherent cells were used after three to five passages.

As a control for RA-FLS, human fibroblast-like synoviocytes (HFLS, Cell Applications, San Diego, California, USA), which is a cell line derived from normal synovial tissue, were also cultured in DMEM supplemented with 10% FBS.

Immunofluorescence staining

Specimens were incubated overnight at 4°C with primary rabbit polyclonal anti-C/EBP β antibodies (C-19; Santa Cruz Biotechnology, Santa Cruz, CA, USA) diluted 1:100, mouse monoclonal anti-RANKL antibodies (ab45039; Abcam, Cambridge, England) diluted 1:50, gout polyclonal anti-OPG antibodies (sc-8468; Santa Cruz Biotechnology) diluted 1:100, rabbit polyclonal anti-ATF4 antibodies (sc-200; Santa Cruz Biotechnology) or normal rabbit IgG (sc-2027; Santa Cruz Biotechnology) diluted 1:100, respectively. RA-FLS plated on glass coverslips were transfected with adenovirus expression vectors for C/EBP β -LAP, -LIP or LacZ control [16] for 24 hours and then replaced with fresh medium. After 48 hours, immunofluorescence staining was performed.

Treatment of cells with cytokines

Confluent cultures of RA-FLS were subjected to serum-free medium for 24 hours. This medium was replaced with fresh medium containing cytokines as follows: IL-1 β (R&D Systems, Minneapolis, MN, USA) at a concentration of 2 ng/ml, TNF- α (Sigma-Aldrich, St Louis, MO, USA) at 10 ng/ml, IL-6 (R&D Systems) at 10 ng/ml, and IL-17 (R&D Systems) at 100 ng/ml. Cells were cultured for a further 48 hours. Concentrations of cytokines were determined based on previous literature [17-19].

Western blotting

Nuclear proteins were isolated using Nuclear and Cytoplasmic Extraction Reagent (NE-PER; Pierce, Rockford, IL, USA). Protein samples were transferred onto nitrocellulose membranes and were treated overnight at 4°C with primary antibodies.

RNA extraction and real-time reverse transcription (RT)-PCR

Quantitative RT-PCR was performed with the LightCycler 2.0 system (Roche, Basel, Switzerland) using SYBR Premix Ex Taq (Takara Bio, Ohtsu, Japan). The primers were as follows: for C/EBP β , 5'-AGTACAAGATCCGGCGCGAG-3' (sense) and 5'-TGCTTGAACAAGTTCCGCAG-3' (antisense); for RANKL, 5'-ATGAACTCCTTCTCCAC AAGCG-3' (sense) and 5'-CTCCTTCTCAGGGCTG AG-3' (antisense; purchased from Takara Bio; oligo name HA137381F and R); for OPG, 5'-GCTTGAAACATA GGAGCTG-3' (sense) and 5'-GTTTACTTT GGT GCCAGG-3' (antisense); for ATF4, 5'-TCAAACCTCAT GGGTTCTCC-3' (sense) and 5'-GTGTCATCCAACGT GGTGAG-3' (antisense); and for GAPDH, 5'-GGTGAA GGTCCGAGTCAACGGA-3' (sense) and 5'-GAGGGAT CTCGCTCTGGAAGA-3' (antisense). Data were normalized to the expression of GAPDH.

Osteoclast formation in a peripheral blood mononuclear cell (PBMC) and RA-FLS co-culture system

Peripheral blood was obtained from healthy donors. Isolated PBMCs (2×10^5 cells/well) were resuspended in α -minimum essential medium (α -MEM) containing 10% FBS and 50 ng/ml macrophage colony-stimulating factor (M-CSF; R&D Systems) and then seeded in 96-well tissue culture plates. Three days later, adherent cells were used for the co-culture system.

Isolated FLS were transfected with adenovirus expression vectors for 24 hours and then fresh medium containing 10% FBS was added. After 48 hours, FLS were added into the 96-well plate with cultured PBMCs in α -MEM containing 10% FBS and 50 ng/ml M-CSF. After 72 hours of co-culture, wells were stained for tartrate-resistant acid phosphatase (TRAP) (Primary Cell Co, Hokkaido, Japan). Osteoclasts were identified as TRAP-positive multinucleated cells that contained more than three nuclei.

Gene knockdown in RA-FLS

Pre-designed small interference RNA (siRNA) for C/EBP β (C/EBP β siRNA-1 target sequence, 5'-CCCACGUGUAA CUGUCAGCt-3' (sense) and 5'-GCUGACAGUUACAC GUGGGt-3' (antisense)) or negative-control siRNA was purchased (Ambion, Austin, TX, USA). Transfection mixes were prepared using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA). RA-FLS cells were cultured for 24 hours after transfection and then treated with 10 ng/ml IL-1 β for 72 hours.

Human RANKL promoter reporter constructs

Promoter constructs for human RANKL were sub-cloned into the pGL-4.10 (luc2) vector (Promega, Madison, WI, USA). The 5'-upstream region (-1591 bp) of the human RANKL gene was prepared using human genomic DNA as a template (p-full). There are four putative binding sites for C/EBP β between -1591 bp and +12 bp. A 2-bp mutation (AA to CC) was made at one site on the p-full construct using the QuickChange site-directed mutagenesis kit (Stratagene, La Jolla, CA, USA).

Plasmid transfection and luciferase assay

HeLa cells seeded in 12-well plates were co-transfected with 0.5 μ g/well RANKL promoter constructs and various concentrations of pCMV-LAP, an expression vector of rat C/EBP β -LAP directed by a cytomegalovirus promoter [20], or pCI-neo-LIP, an expression vector of rat C/EBP β -LIP [21], or pCMV6-AC-GFP-tagged ATF4 (OriGene, Rockville, MD, USA), an expression vector of human ATF4, using Lipofectamine LTX (Invitrogen). pRL-SV40 (Promega) was used as an internal control. Luciferase activity was then assayed using the Dual-luciferase Reporter Assay System (Promega).

Chromatin immunoprecipitation (ChIP) assay

RA-FLS cells were transfected with the adenovirus vector C/EBP β -LIP and incubated for 72 hours. A ChIP assay was performed with a ChIP Assay kit (Upstate Biotechnology, Lake Placid, NY, USA). The primers used in the PCR for RANKL promoter sequences were as follows: 5'-GAGGGCGAAAG GAAGGAAGGGGAG-3' (sense) and 5'-GGCGTTGGA GAGCCCTGGCCTCGG -3' (antisense), which amplified between -125 bp and +26 bp. For a negative control, sequence between -1727 bp and -1487 bp was used. The PCR products were amplified for 33 cycles.

Immunoprecipitation (IP)

Nuclear proteins were isolated from RA-FLS transfected with adenovirus vector C/EBP β -LIP for 72 hours. The IP protocol used Dynabeads Protein A (Invitrogen). Anti-C/EBP β antibodies, anti-ATF4 antibodies or normal rabbit IgG and Dynabeads-complex, respectively, were added to antigen-containing lysates. Proteins were separated by SDS-PAGE and immunoblotted using specific antibodies.

Statistical analyses

For *in vitro* investigations, nonparametric comparisons were performed using the Mann-Whitney *U*-test. *P*-values less than 0.05 were considered significant.

Results

Co-localization of C/EBP β and RANKL in the synovium from RA patients

We initially examined C/EBP β and RANKL expression by immunofluorescence staining in erosive areas of synovial tissue from RA patients. C/EBP β and RANKL were expressed in RA synovial tissue (Figure 1A). The distribution patterns of C/EBP β and RANKL were similar and both were strongly expressed in the synovial lining layer rather than in the sub-lining layer. The co-localization of C/EBP β and RANKL in RA synovium suggests that C/EBP β is involved in the regulation of RANKL expression.

Expression of C/EBP β in RA-FLS after treatment with pro-inflammatory cytokines

Primary cultures of FLS were established and C/EBP β expression was examined by western blotting. C/EBP β -LAP* (38 kDa), -LAP (36 kDa) and -LIP (20 kDa) were detected with LIP showing dominant expression. There was a varying degree of C/EBP β expression (Figure 1B). The difference in expression levels of C/EBP β may depend on the history of the patients such as degree of inflammation at the time of sample collection, disease duration, or therapies. Human FLS from normal articular joints lacks C/EBP β protein expression.

Next, we set out to determine whether pro-inflammatory cytokines could promote C/EBP β protein in FLS. Western blots revealed that stimulation with IL-1 β (2 ng/ml), TNF- α (10 ng/ml), IL-6 (5 ng/ml), or IL-17 (100 ng/ml) increased the expression of both LAP and LIP isoforms in nuclear extracts, whereas the samples without any treatment did not show expression of C/EBP β protein (Figure 1C). Interestingly, the expression of LIP was higher than that of LAP as shown in experiments of primary cultured RA-FLS.

Overexpression of C/EBP β regulates expression of RANKL and OPG in RA-FLS

RA-FLS cells were transfected with adenovirus expression vectors expressing C/EBP β -LAP, -LIP or LacZ control. Western blots confirmed the exogenous overexpression of LAP or LIP in whole protein extracts isolated from transfected cells (Figure 2A). RANKL mRNA expression was examined by quantitative RT-PCR. The overexpression of LAP induced RANKL mRNA expression up to 80-fold compared to the LacZ control in a time-dependent manner. In RA-FLS transfected with the LIP vector, RANKL mRNA expression was increased approximately 6-fold (Figure 2A). We also investigated the expression of OPG.

Expression of OPG mRNA was upregulated by LAP in RA-FLS, whereas LIP significantly reduced OPG mRNA. Consequently, the RANKL-OPG ratio was highly upregulated in RA-FLS transfected with LIP (Figure 2B).

In addition, we examined whether C/EBP β induced RANKL expression at the protein level by cell fluorescent immunostaining in a time-course experiment. The stimulated expression of C/EBP β was observed in the nucleus of RA-FLS at 24 hours (Figure 2C). RANKL protein was localized in the cell cytoplasm of FLS over-expressing LAP or LIP at 72 hours. Similarly, the expression of OPG was also examined in a different series of experiments. The expression of OPG was stimulated by C/EBP β -LAP, but not by C/EBP β -LIP in RA-FLS at 72 hours.

C/EBP β knockdown by siRNA reduced RANKL expression in RA-FLS

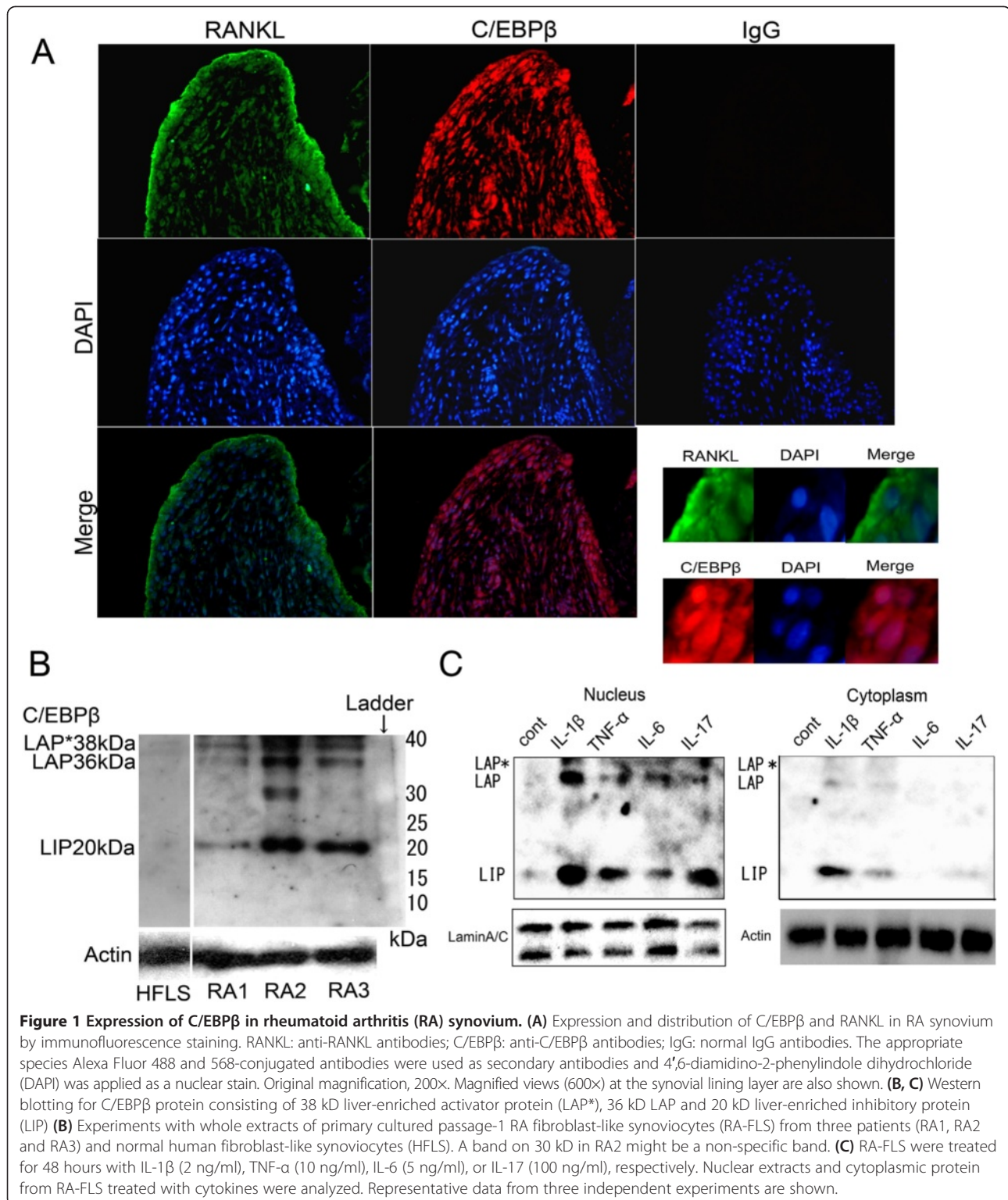
We assessed the effect of C/EBP β knockdown on RANKL mRNA expression using siRNAs targeting C/EBP β mRNA. Transfected cells were cultured with IL-1 β . C/EBP β knockdown significantly reduced RANKL mRNA expression by 50% after IL-1 β treatment in RA-FLS (Figure 2D).

C/EBP β induced osteoclast formation through RANKL expression in RA-FLS

To investigate whether C/EBP β -induced RANKL expression stimulated osteoclast formation, we co-cultured RA-FLS, which were transfected with adenovirus vectors expressing C/EBP β -LAP, -LIP or LacZ control, and PBMCs, which were isolated and stimulated with M-CSF. After three days of co-culture, formation of TRAP-positive multinucleated cells was observed in co-cultures of RA-FLS over-expressing LAP or LIP, but not in control cells. Interestingly, more multinucleated cells were induced in the co-cultures of RA-FLS over-expressing LIP than with LAP transfected cells (Figure 3A). As a negative control, we performed monotype cell cultures of RA-FLS over-expressing C/EBP β , or PBMC over-expressing C/EBP β . These cells did not form osteoclasts (Figure 3B).

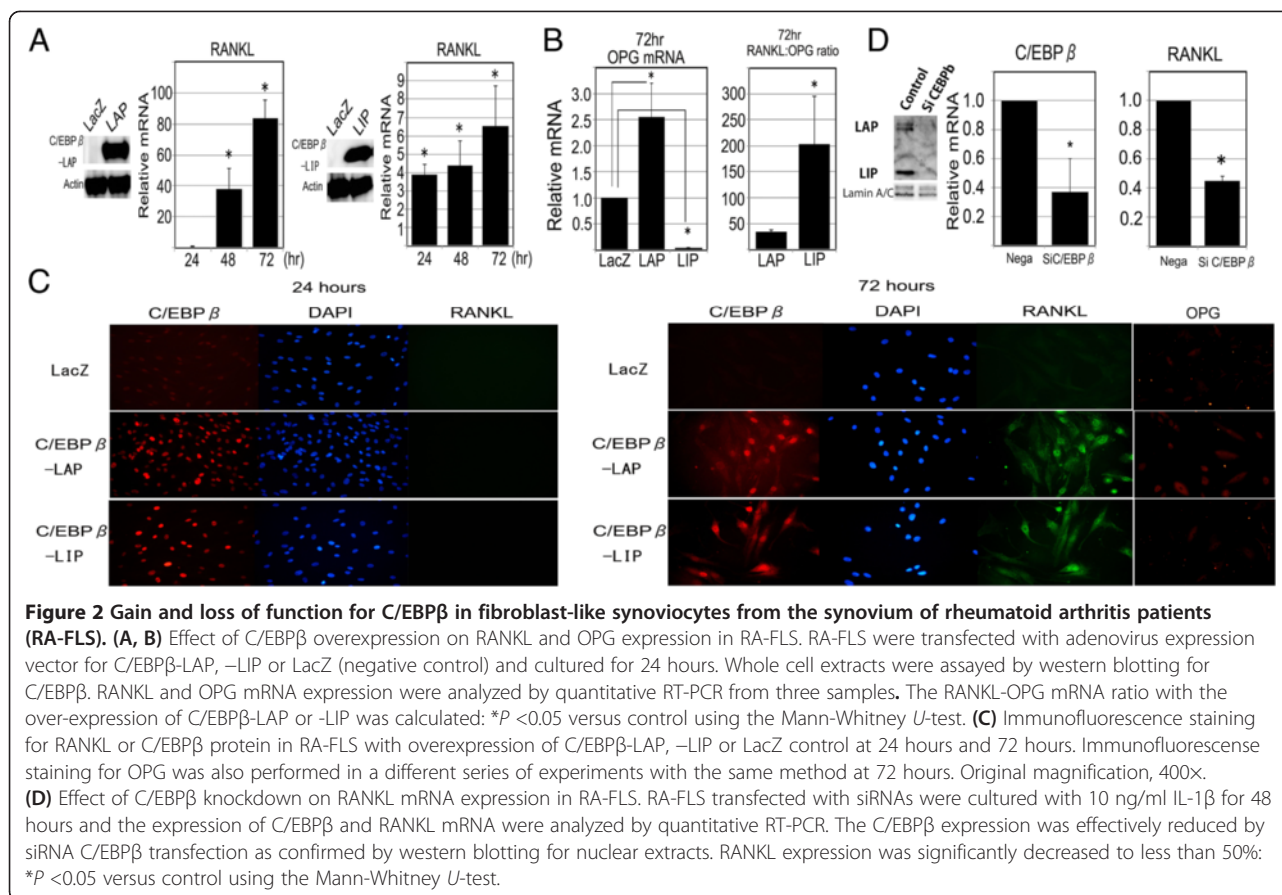
C/EBP β functions as an activator of the human RANKL promoter

We further analyzed the *in vitro* promoter activity of human RANKL using HeLa cells. A luciferase reporter gene construct containing -1591 bp of the RANKL promoter was co-transfected with the expression vectors for C/EBP β -LAP (pCMV-LAP) or C/EBP β -LIP (pCI-neo-LIP) into HeLa cells. RANKL promoter activity was upregulated in a dose-dependent manner with either LAP or LIP (Figure 4A). To identify the C/EBP β responsive element in the RANKL promoter, mutation analysis was performed using site-directed mutagenesis. We created four single mutation constructs in the RANKL promoter: single mutation 1 (mut-1), mutation 2 (mut-2), mutation 3 (mut-3),



and mutation 4 (mut-4). Luciferase activities of mut-1, mut-2, and mut-3 reporter constructs were equally increased with pCMV-LAP, while mut-4 reporter construct decreased luciferase activity by 30% (Figure 4B). Similarly, using the C/EBP β -LIP expression vector, mut-2 and mut-3

did not show difference of activity from full reporter construct, while mut-4 decreased to 25%. These results showed that the putative C/EBP β binding site is located between -59 bp and -52 bp in the RANKL promoter. The rationale for the increased activity of mut-1 with C/EBP β -



LIP expression vector has not been discovered yet as we failed to show the direct binding of C/EBP β on this site.

C/EBP β -LIP and ATF4 synergistically stimulate RANKL expression

C/EBP β -LIP does not have an activation domain and is considered to be a dominant negative isoform. However, our results suggest that LIP is involved in RANKL expression in RA-FLS and plays a role in induction of osteoclast formation. We hypothesized that some transcriptional co-factors may cooperate with C/EBP β -LIP to activate transcription of the RANKL promoter. ATF4 is known to stimulate RANKL expression in osteoblasts [22]. Additionally, ATF4 has been shown previously to interact with C/EBP β , which activates various downstream factors such as osteocalcin and discoidin domain receptor tyrosine kinase (DDR2) [23]. Thus, we considered that a similar mechanism might exist for the regulation of RANKL gene expression. A luciferase assay showed that ATF4 slightly activated the RANKL promoter (Figure 4C). RANKL promoter activity was significantly enhanced following co-transfection of LIP and ATF4. Mut-4 abrogated the responsiveness of the RANKL promoter to the combination of LIP and ATF4. A ChIP assay was performed using RA-FLS over-expressing LIP and primers

constructed from the human RANKL promoter sequence, which amplify sites including the C/EBP β consensus site-4. This analysis indicated that LIP binds to the RANKL promoter region containing CS-4 and that ATF4 also binds in the same region (Figure 4D). IP and immunoblotting demonstrated that ATF4 bound to over-expressed LIP in RA-FLS (Figure 4E). Collectively, these results suggest that C/EBP β -LIP cooperates with ATF4 in activating RANKL gene expression.

ATF4 constitutively exists in RA synovium

We then examined the localization of ATF4 in RA synovium. ATF4 was observed in erosive areas of RA synovium by immunofluorescence staining (Figure 5A). Western blotting showed that ATF4 was expressed in whole cell extracts of RA-FLS (Figure 5B). Next, we examined whether ATF4 expression was affected by C/EBP β in RA-FLS transfected with adenovirus expression vectors in time-course experiments. ATF4 mRNA expression was not significantly changed by C/EBP β (Figure 5C). In addition, we performed organ cultures using RA synovium tissue. In RA synovium transfected with adenovirus LacZ control, ATF4 mainly localized in the cytoplasm of cells. Interestingly, in RA synovium over-expressing LIP, ATF4 was mainly located in the nucleus (Figure 5D). The results suggest that ATF4

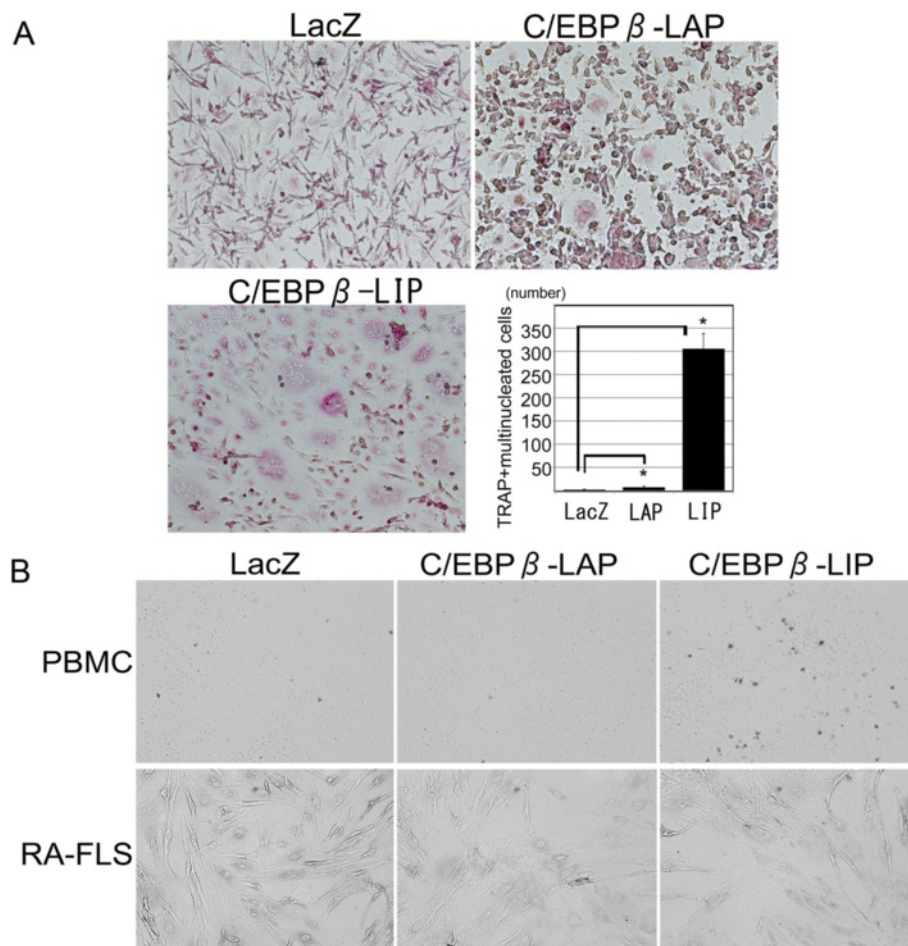


Figure 3 Osteoclast formation from peripheral blood mononuclear cells (PBMC) is promoted by the enhanced expression of C/EBPβ in fibroblast-like synoviocytes from the synovium of rheumatoid arthritis patients (RA-FLS). **(A)** Co-cultures of PBMCs and RA-FLS transfected with adenovirus vectors expressing C/EBPβ-LAP, -LIP or LacZ control for 72 hours. Number of TRAP-positive multinucleated cells recognized in the co-culture system. Osteoclasts were identified as TRAP-positive multinucleated cells that contained more than three nuclei. Original magnification, 100×. * $P < 0.05$ versus control using the Mann-Whitney U -test. **(B)** Negative control for the co-culture experiments. PBMC or RA-FLS over-expressing C/EBPβ-LAP, -LIP or LacZ control, respectively, were cultured for 72 hours. No TRAP-positive multinucleated cells were observed.

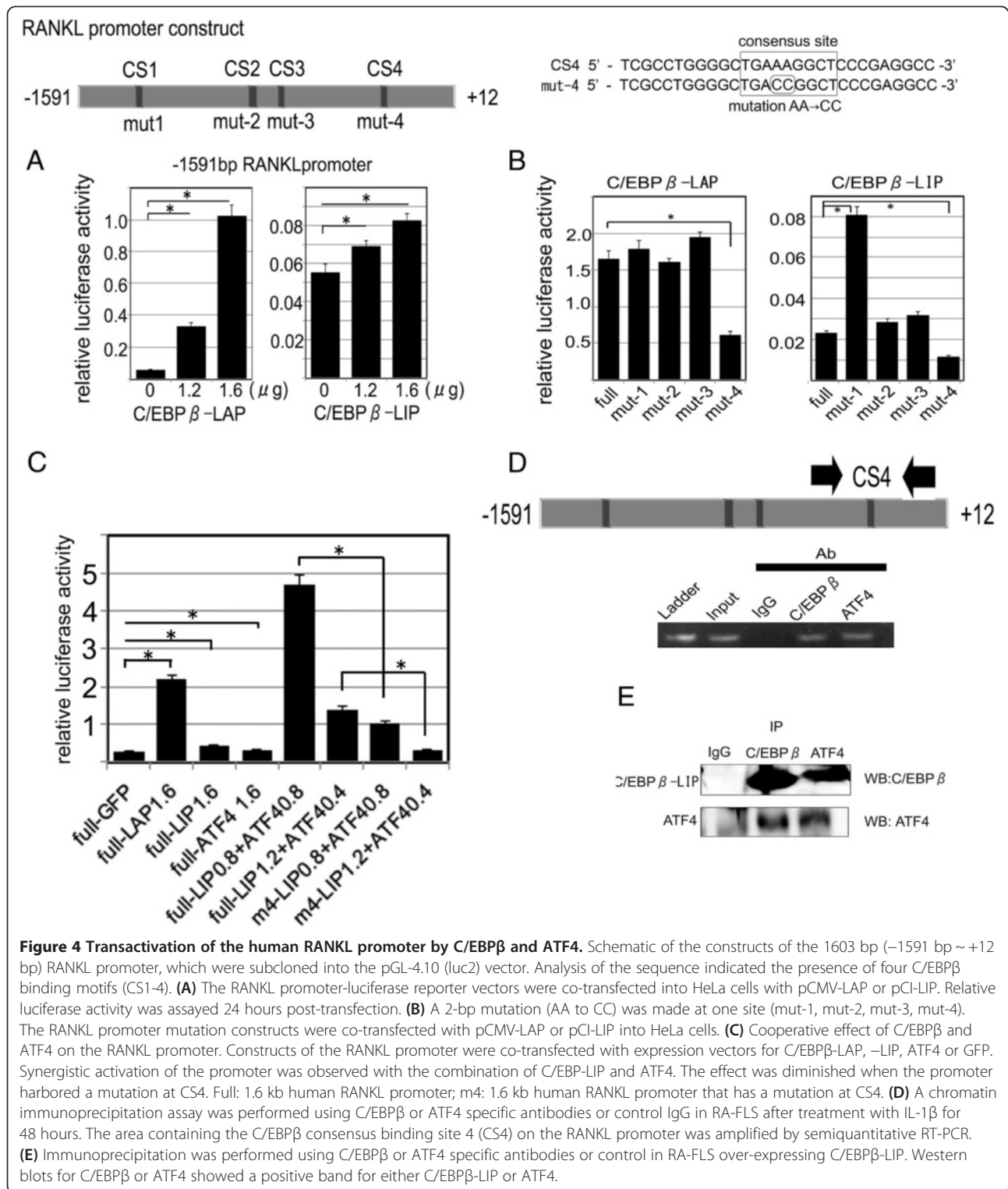
is translocated from the cytoplasm into the nucleus in the RA synovium overexpressing C/EBPβ-LIP although the mechanisms are not clear.

Discussion

In this study, we have shown that the transcription factor C/EBPβ promotes the expression of RANKL in RA synovium. C/EBPβ-induced RANKL in synovium could induce the formation of osteoclasts. This paper demonstrates that RA-FLS expressed the C/EBPβ-LIP isoform more dominantly than the C/EBPβ-LAP isoform in response to pro-inflammatory cytokines. C/EBPβ-LIP stimulated RANKL expression even though C/EBPβ-LIP lacks the transactivating domain. Recruitment of ATF4, which is constitutively expressed in the cytoplasm of RA-FLS, to the RANKL promoter might be the mechanism by which

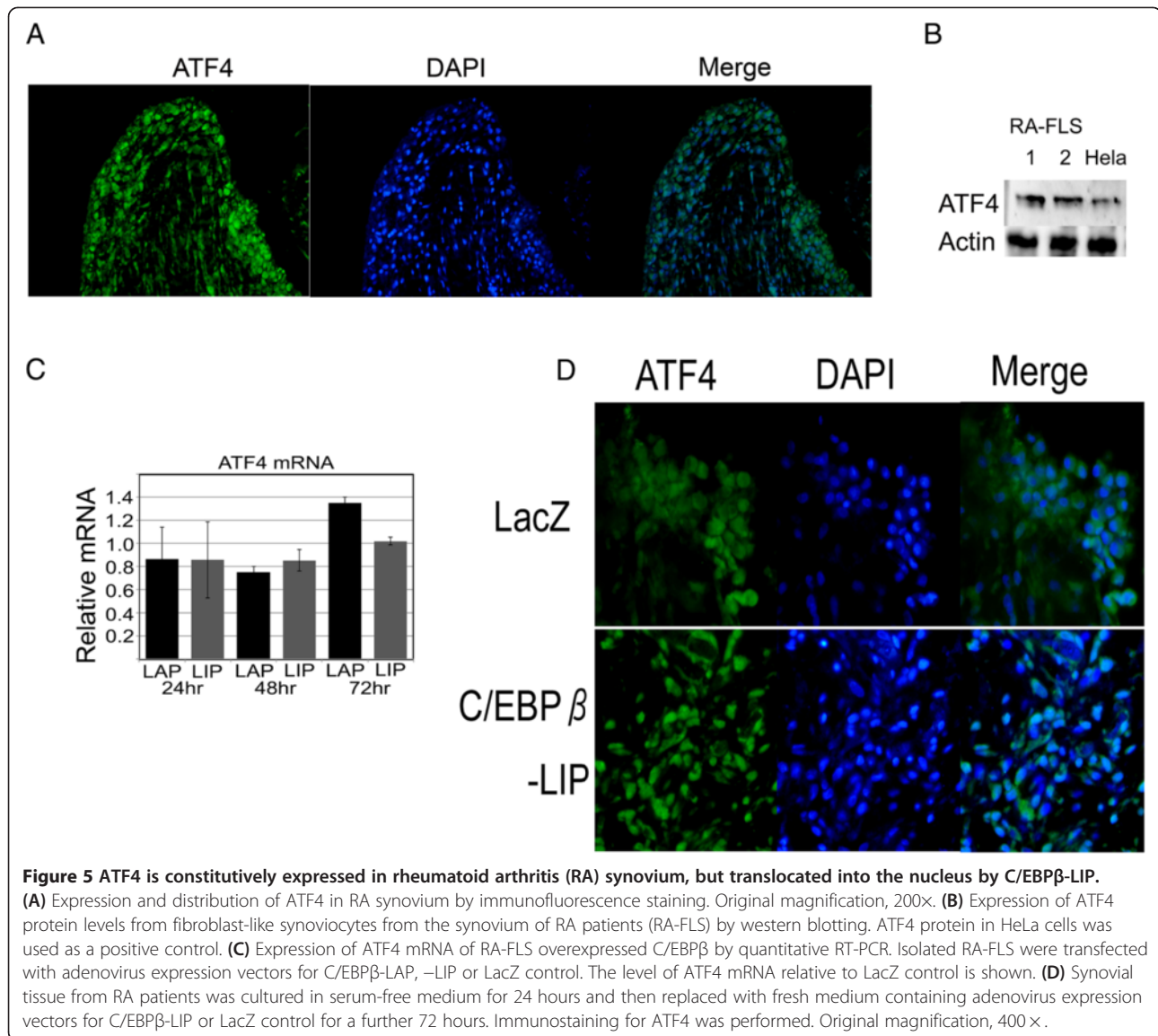
C/EBPβ-LIP activates the promoter. More interestingly, the effect of C/EBPβ-LIP in osteoclast formation is stronger than that of C/EBPβ-LAP. The lower expression of OPG might be involved in this mechanism. These results suggest that C/EBPβ-LIP is one of the key regulators of inflammation-induced osteoclast formation. As C/EBPβ is also involved in cartilage degradation [19], C/EBPβ may play a crucial role in joint destruction in RA.

Understanding the mechanisms that mediate RANKL gene expression may aid development of new therapies for reducing bone resorption in RA. We showed that LAP and LIP directly bind a site located between -59 bp and -52 bp of the RANKL promoter. LIP is increased in the RA synovium and forms a complex with constitutively expressed ATF4. This complex may activate transcription of RANKL by binding to the C/EBPβ binding



motif of the RANKL promoter. ATF4 belongs to the ATF/cAMP responsive element binding protein (CREB) family, which contains a basic leucine zipper region, and is one of the major regulators of osteoblast differentiation [24]. Moreover, ATF4 regulates RANKL expression [22].

This study showed that ATF4 is expressed in RA synovium and is involved in RANKL expression. Interestingly, in *ex vivo* experiments (Figure 5D), ATF4 was mainly expressed in the cytoplasm of FLS transfected with the LacZ adenovirus vector (control), while ATF4 tended to



be located in the nucleus of FLS that overexpressed LIP. ATF4 in cooperation with C/EBP β might be a crucial regulator of RANKL expression in mediating synovium-induced bone resorption in RA. Other transcription factors, such as NF- κ B, AP-1, STAT3 and Runt-related transcription factor-2 (Runx2), may also interface with C/EBP β . Runx2 and C/EBP β cooperatively promote the expression of Indian Hedgehog in hypertrophic chondrocytes [25]. STAT3 is induced by IL-1 β , TNF- α , and IL-6 and increases the expression of IL-6 and RANKL. A positive feedback loop, via IL-6 and STAT3, enhanced RANKL expression and osteoclastogenesis in inflammatory arthritis [26]. C/EBP β was previously known as nuclear factor for IL-6 expression (NF-IL6) [27]. IL-6 induced by C/EBP β regulates C/EBP β gene transcription

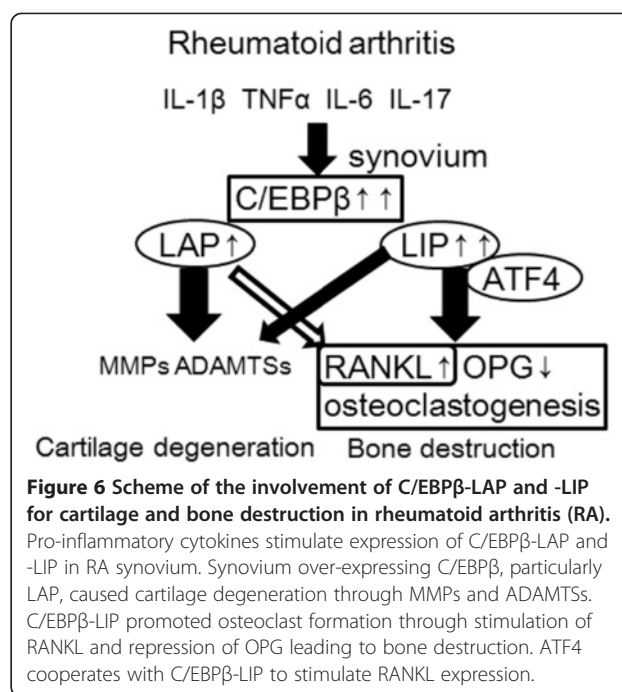
with recruitment of STAT3 to the promoter of the C/EBP β gene, especially in hepatocytes [28]. Therefore, we consider that positive feedback loops involving pro-inflammatory cytokines, IL-6, STAT3 and C/EBP β , might strongly increase RANKL expression in joints with RA. Indeed, STAT3 is essential for stimulation of RANKL and its binding element is located at -82 bp of the RANKL promoter, which is next to the C/EBP β responsive motifs. C/EBP β -LIP may exist as an anchor to form a complex with other transcription factors mediated by inflammatory pathways.

The C/EBP β isoform ratio can alter in response to cellular processes [7,29,30]. Of note, the LAP-LIP ratio is significant for osteoclastogenesis in PBMC through the mTOR pathway [11]. These papers indicate that an

appropriate LAP-LIP ratio results in higher transcriptional activation of the target gene, which is very important in proliferation and differentiation. We showed that LIP protein is more highly expressed in RA-FLS than LAP. This imbalance of the LAP-LIP ratio caused a concomitant change in OPG mRNA expression. OPG is an endogenous inhibitor of RANKL-RANK interaction and is produced in synovial cells of patients with RA. The balance between levels of RANKL and OPG (RANKL-OPG ratio) is correlated with the extent of bone resorption in RA joints [31]. The current study demonstrates that LIP drastically increased the RANKL-OPG ratio in RA-FLS, which subsequently induced significant osteoclast formation.

Previous studies showed that C/EBP β -LAP is a key regulator of cartilage degradation in inflammatory arthritis. C/EBP β -LAP plays a crucial role in cartilage degradation along with proteolytic enzymes such as MMP-1, MMP-3, MMP-13, and aggrecanase-2 (ADAMTS-5) in chondrocytes and FLS in inflammatory arthritis [19,32,33]. The role of LIP is not well investigated in inflammatory arthritis. Our unpublished data revealed that overexpression of LIP in FLS increased MMP-1, MMP-3, MMP-9, MMP-13, and ADAMTS-4 mRNA similar to the overexpression of LAP. The data presented here suggest that LAP and LIP coordinate in enhancing expression of RANKL, MMPs, and ADAMTSs, which may result in cartilage degradation and bone destruction of RA joints. C/EBP β may be a common regulator, which can be stimulated in response to pro-inflammatory cytokines and upregulated in RA synovium. Therefore, selective blockage of C/EBP β expression may be one potential strategy for preventing inflammation and bone resorption in arthritis.

This study has several limitations. First, double staining for C/EBP β and RANKL did not work well on the RA synovium sections, although the reason was unclear. Therefore, we stained C/EBP β and RANKL separately. However, the distribution of these molecules overlapped each other, suggesting that C/EBP β and RANKL are co-expressed. Next, the promoter assays were performed in HeLa cells. We could not obtain reliable results of the promoter assay in RA FLS probably because of low transfection efficiency. However, the purpose of these experiments was to see the effect of various transcription factors that were exogenously introduced by expression vectors. Therefore, the influence of cell characteristics on the results is considered to be limited. Thirdly, the promoter that harbors a mutation in CS1 showed increased activity with C/EBP β -LIP (Figure 4B). This result suggests that LIP act as a repressor in CS1. However, ChIP assay for CS1 sequences did not show binding of C/EBP β on CS1 (data not shown). Therefore, the function of CS1 remains unclear at the moment.



Conclusions

In conclusion, C/EBP β increased RANKL expression in RA-FLS and induced osteoclastogenesis (Figure 6). Pro-inflammatory cytokines significantly induced C/EBP β -LIP, which strongly induced osteoclastogenesis by increasing the RANKL-OPG ratio in RA-FLS. LIP possesses transactivation activity for the RANKL promoter by recruiting ATF4, which constitutively exists in the cytoplasm of RA-FLS, to the C/EBP binding site in the RANKL promoter. In pathological inflammatory arthritis, C/EBP β is a crucial factor in damaging cartilage and bone in joints.

Abbreviations

α -MEM: α -minimum essential medium; AP-1: activator protein-1; ATF-4: activation transcription factor 4; bp: base pairs; C/EBP: CCAAT/enhancer binding protein; ChIP: chromatin immunoprecipitation; DMEM: Dulbecco's modified Eagle's medium; FBS: fetal bovine serum; FLS: fibroblast-like synoviocytes; GCT: giant cell tumor; IL: interleukin; JAK-STAT: janus kinase-signal transducer and activator of transcription; LAP: liver-enriched activator protein; LIP: liver-enriched inhibitory protein; M-CSF: macrophage colony-stimulating factor; MMP: matrix metalloproteinase; NF- κ B: nuclear factor-kappa-B; OPG: osteoprotegerin; PBMC: peripheral blood mononuclear cell; RA: rheumatoid arthritis; RA-FLS: fibroblast-like synoviocytes from the synovium of RA patients; RANKL: receptor activator of nuclear factor kappa B ligand; RT: reverse transcription; siRNA: small interference RNA; TNF: tumor necrosis factor; TAD: N-terminal transactivation domain; TRAP: tartrate-resistant acid phosphatase.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

HT participated in all the experiments and drafted the manuscript. KO conceived of the study, and participated in its design and coordination and helped to draft the manuscript. KI participated in the western blotting for RA-FLS and immunofluorescence staining of cells for OPG and helped to revise the manuscript. TU participated in the knockdown experiment with

siRNA and the luciferase assays and helped to draft the manuscript. YI participated in the design of the study and involved in revising the manuscript critically for important intellectual content. All authors read and approved the final manuscript.

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