

**Research Paper** 



# Association between NEFL Gene Polymorphisms and Neuroblastoma Risk in Chinese Children: A Two-Center Case-Control Study

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#### Abstract

Neuroblastoma is a lethal tumor that mainly occurs in children. To date, the genetic etiology of sporadic neuroblastoma remains obscure. A previous study identified three neuroblastoma susceptibility loci (rs11994014 G>A, rs2979704 T>C, rs1059111 A>T) in *neurofilament light (NEFL)* gene. Here, we attempted to evaluate the contributions of these three single nucleotide polymorphisms to neuroblastoma susceptibility in Chinese children. We genotyped these three polymorphisms using subjects from Guangdong province (256 cases and 531 controls) and Henan province (118 cases and 281 controls). Logistic regression models were performed to generate odds ratios and 95% confidence intervals to access the association of these three polymorphisms with neuroblastoma risk. Overall, we failed to provide any evidence supporting the association between these three polymorphisms and neuroblastoma susceptibility, either in single center population or in the combined population. Moreover, such null association was also observed when the samples were stratified by age, gender, tumor sites, and clinical stages. In the future, larger samples from different ethnicities are needed to clarify the role of *NEFL* gene polymorphisms in neuroblastoma risk.

Key words: neuroblastoma; NEFL; polymorphism; susceptibility

## Introduction

Neuroblastoma, one the most common solid cancer in children, is characterized by various clinical phenotypes [1]. It accounts for about 7% of all cancer types in children under 15 years of age [2, 3]. The prevalence of neuroblastoma is higher in the United States [4] than that in China [5] (8-14 cases per million versus about 7.7 per million). Neuroblastoma can be categorized into three different groups, low-risk, intermediate-risk, and high-risk groups. A large

proportion of the affected patients have a favorable prognosis, as they bear localized benign tumors that can regress spontaneously [6]. The survival rate for low-risk patients is generally greater than 95%, no matter treated with minimal chemotherapy or not. Patients with intermediate-risk neuroblastoma constitute about 15% of all neuroblastoma patients, and their survival rate is larger than 80% after multimodal therapy. Moreover, nearly 50% of patients are classified as high-risk neuroblastoma, with survival rate approximately less than 35% even after` receiving comprehensive treatments [7, 8]. Such poor prognosis might be attributed to the widespread dissemination of tumor to bone and bone marrow at the time of diagnosis.

By now, the etiology of familial neuroblastoma is mainly attributed to the ALK [9, 10] and PHOX2B gene mutations. However, [11, 12] familial of all neuroblastoma only accounts for 1% neuroblastoma cases [13]. The explicit causes of most common sporadic neuroblastoma still remain obscure. Previous studies have paid attention to the role of environmental risk factors in influencing susceptibility to neuroblastoma [14, 15]. However, environmental etiology alone cannot explain the fact that only a small portion of children finally developed neuroblastoma after parental exposure to risk environmental factors [16]. Mounting evidence has that genetic factors and suggested gene-environmental interactions might influence the susceptibility to neuroblastoma [17, 18].

Early genome-wide association studies (GWASs) of neuroblastoma have identified a number of neuroblastoma susceptibility loci in the genes including BARD1 [19], DUSP12 [20], DDX4 [20], HSD17B12 [20], IL31RA [20], LIN28B [21], HACE1 [21], and LMO1 [22]. Moreover, several single nucleotide polymorphisms (SNPs) in lncRNA also contributed to the susceptibility to neuroblastoma and other cancers [23, 24]. However, these gene loci only account for a small fraction of neuroblastoma heritability. Some potential functional gene loci might be ruled out as a result of the introduction of the multiple testing corrections in GWAS [25]. To discover more hidden true-positive loci, a variety of novel methods have been presented. The methods include but not limit to: adopting gene- or pathway-based approaches, using larger GWAS samples, replicating previously reported GWAS signals in a larger cohort, conducting a meta-analysis of GWAS datasets, and performing imputation and epistasis analysis [26]. For instance, using larger GWAS samples, rare variants of TP53 were found to be associated with neuroblastoma predisposition [27].

In a previous proteomic study on differentiated neuroblastoma cell line, Capasso et al. [28] found that eight proteins were differentially expressed during neuroblastoma differentiation. To discover more variants predisposing to neuroblastoma, Capasso et al. [29] further investigated whether the genes encoding these eight proteins were related to neuroblastoma development. Using the discovery set (2,101 patients and 4,202 control subjects) and the replication set (459 cases and 809 controls), they successfully identified three common neuroblastoma predisposing variants in the *neurofilament light* (*NEFL*) gene (rs11994014 G>A, rs2979704 T>C, rs1059111 A>T).

Here, we tested the association between the three putative SNPs in *NEFL* gene and neuroblastoma risk in a two-center, case-control study in Chinese population. The aim of our study was to reveal the contribution of these SNPs to the neuroblastoma susceptibility in Chinese children.

# Materials and methods

## **Study subjects**

This study consisted of two populations, one including 256 cases and 531 controls from Guangdong province, and the other including 118 cases and 281 controls from Henan province (**Supplemental Table 1**) [30]. The selection details and criteria of the subjects were provided in our previously published articles [31-36]. At recruitment, informed written consent was obtained from all participants or their guardians. The study protocols were approved by the Institutional Review Boards of Guangzhou Women and Children's Medical Center and the First Affiliated Hospital of Zhengzhou University.

## SNP selection and genotyping

Three SNPs (rs11994014 G>A, rs2979704 T>C, rs1059111 A>T) in the *NEFL* gene by Capasso et al. [29] were selected. Genomic DNA was mainly purified from venous blood collected from the subjects, using a TIANamp Blood DNA Kit (TianGen Biotech Co. Ltd., Beijing, China). Genotyping was performed using TaqMan real-time PCR by standard methods, with details reported elsewhere [37-39]. For quality control, we randomly selected approximately 10% of the samples to perform duplicate analyses. 100% concordance rate was observed for each SNP among duplicate sets.

## Statistical analysis

Deviation from Hardy-Weinberg equilibrium (HWE) for each SNP was assessed in the control subjects, using the goodness-of-fit  $\chi^2$  test. The chi-squared test was employed to evaluate differences in the demographic variables and allele frequencies between the cases and controls. Odds ratios and corresponding 95% confidence intervals were calculated to determine the relative cancer risk, adjusting for age and gender. A two-sided *P* value < 0.05 was set as the statistical significance level. All statistical analyses were conducted using SAS statistical package, version 9.1 (SAS Institute, Cary, NC).

#### Results

## Association between NEFL gene polymorphisms and neuroblastoma susceptibility

Genotyping were successfully conducted in the Guangdong population (255 cases and 531 controls) and Henan population (118 cases and 281 controls). The genotype frequencies and association results of these three NEFL gene polymorphisms (rs11994014 G>A, rs2979704 T>C, and rs1059111 A>T) were presented for Guangdong, Henan, and combined population separately in Table 1. All of these three polymorphisms were agreed with HWE (P=0.513 for rs11994014 G>A, P=0.498 for rs2979704 T>C, and P=0.535 for rs1059111 A>T in the controls of combined population). No significant association between these three polymorphisms and neuroblastoma risk was observed in either study population. We then validated such association in the combined population, aiming to obtain a robust conclusion. Likely, no association were found polymorphisms between these three and neuroblastoma risk (Table 1).

## Stratification analysis for the association between NEFL gene polymorphisms and neuroblastoma susceptibility for combined subjects

Stratified analysis was also performed in the combined population by age, gender, sites of origin, and clinical stages. Consistently, we found no significant associations for all these three polymorphisms (**Table 2**).

Herein, we investigated the association of *NEFL* gene polymorphisms with neuroblastoma risk in Chinese population. To the best of our knowledge, this is the first study to replicate the previously reported associations between *NEFL* gene polymorphisms and neuroblastoma risk in a two-center case-control study of Chinese children.

NEFL gene is located at chromosome 8p21.2. This gene encodes the light subunit of neurofilaments, which play an important role in maintaining normal function of nerve cells [40, 41]. More than 10 mutations in the NEFL gene are found to be associated with Charcot-Marie-Tooth disease types 2, or distal nerve demyelination [42]. In addition to the definite role of NEFL gene in neuron disease, NEFL gene also has been implicated in cancer initiation and progression. First, the chromosome 8p21 is enriched for loss of heterozygosity (LOH), which was involved in various cancers, including breast cancer [43, 44], lung cancer [45], prostate cancer [46], head and neck cancer [47]. It was reported that more than 40% of head and neck cancer patients harbored LOH at the NEFL locus. Second, several functional molecular targets related to cancer-associated pathways have been shown to interact with NEFL gene [48]. Furthermore, NEFL could also alter the cancer cell resistance to chemotherapy. Previous study has pointed out that restoring expression of NEFL in head and neck cancer cell lines could increase sensitivity of the cells to the cisplatin [48]. Last, aberrant expression and methylation of NEFL gene was detected in several cancers [49-51].

Table 1. Association between NEFL gene polymorphisms and neuroblastoma susceptibility

Genotype	Guangdong	g province		Henan prov	vince		Combined subjects					
	Cases (N=255)	Controls (N=531)	Adjusted OR (95% CI) ª	P a	Cases (N=118)	Controls (N=281)	Adjusted OR (95% CI) ª	P <sup>a</sup>	Cases (N=373)	Controls (N=812)	Adjusted OR (95% CI) ª	P a
rs11994014 G	>A (HWE=0.51	13 for combine	ed subjects)				,				,	
GG	83 (32.55)	186 (35.03)	1.00		47 (39.83)	115 (40.93)	1.00		130 (34.85)	301 (37.07)	1.00	
AG	127 (49.80)	267 (50.28)	1.06 (0.76-1.48)	0.747	60 (50.85)	127 (45.20)	1.13 (0.72-1.79)	0.595	187 (50.13)	394 (48.52)	1.10 (0.84-1.44)	0.496
AA	45 (17.65)	78 (14.69)	1.28 (0.81-2.00)	0.287	11 (9.32)	39 (13.88)	0.70 (0.33-1.48)	0.351	56 (15.01)	117 (14.41)	1.11 (0.76-1.62)	0.597
Additive			1.12 (0.90-1.39)	0.324			0.92 (0.67-1.28)	0.632			1.06 (0.89-1.27)	0.510
Dominant	172 (67.45)	345 (64.97)	1.11 (0.81-1.52)	0.533	71 (60.17)	166 (59.07)	1.03 (0.66-1.60)	0.890	243 (65.15)	511 (62.93)	1.10 (0.85-1.42)	0.465
Recessive	210 (82.35)	453 (85.31)	1.24 (0.83-1.85)	0.302	107 (90.68)	242 (86.12)	0.65 (0.32-1.33)	0.240	317 (84.99)	695 (85.59)	1.05 (0.74-1.48)	0.784
rs2979704 T>	C (HWE=0.498	for combined	l subjects)									
TT	83 (32.55)	184 (34.65)	1.00		48 (40.68)	115 (40.93)	1.00		131 (35.12)	299 (36.82)	1.00	
CT	126 (49.41)	268 (50.47)	1.04 (0.74-1.45)	0.835	60 (50.85)	127 (45.20)	1.11 (0.70-1.75)	0.670	186 (49.87)	395 (48.65)	1.07 (0.82-1.41)	0.602
CC	46 (18.04)	79 (14.88)	1.28 (0.82-2.00)	0.286	10 (8.47)	39 (13.88)	0.62 (0.29-1.34)	0.225	56 (15.01)	118 (14.53)	1.08 (0.74-1.58)	0.680
Additive			1.11 (0.89-1.39)	0.338			0.89 (0.64-1.23)	0.465			1.05 (0.88-1.26)	0.609
Dominant	172 (67.45)	347 (65.35)	1.09 (0.79-1.50)	0.592	70 (59.32)	166 (59.07)	0.99 (0.64-1.54)	0.968	242 (64.88)	513 (63.18)	1.08 (0.83-1.39)	0.574
Recessive	209 (81.96)	452 (85.12)	1.25 (0.84-1.86)	0.275	108 (91.53)	242 (86.12)	0.59 (0.28-1.22)	0.155	317 (84.99)	694 (85.47)	1.04 (0.74-1.47)	0.827
rs1059111 A>	T (HWE=0.535	for combined	l subjects)									
AA	84 (32.94)	185 (34.84)	1.00		48 (40.68)	115 (40.93)	1.00		132 (35.39)	300 (36.95)	1.00	
AT	122 (47.84)	267 (50.28)	1.00 (0.72-1.40)	0.999	60 (50.85)	127 (45.20)	1.11 (0.70-1.75)	0.670	182 (48.79)	394 (48.52)	1.05 (0.80-1.37)	0.729
TT	49 (19.22)	79 (14.88)	1.35 (0.87-2.10)	0.181	10 (8.47)	39 (13.88)	0.62 (0.29-1.34)	0.225	59 (15.82)	118 (14.53)	1.14 (0.78-1.65)	0.501
Additive			1.13 (0.91-1.41)	0.256			0.89 (0.64-1.23)	0.465			1.06 (0.89-1.27)	0.506
Dominant	171 (67.06)	346 (65.16)	1.08 (0.79-1.48)	0.634	70 (59.32)	166 (59.07)	0.99 (0.64-1.54)	0.968	241 (64.61)	512 (63.05)	1.07 (0.83-1.38)	0.608
Recessive	206 (80.78)	452 (85.12)	1.35 (0.91-2.00)	0.134	108 (91.53)	242 (86.12)	0.59 (0.28-1.22)	0.155	314 (84.18)	694 (85.47)	1.11 (0.79-1.55)	0.560

 Table 2. Stratification analysis for the association between NEFL gene polymorphisms and neuroblastoma susceptibility for combined subjects

Variables	rs11994014 G>A				rs29797(	)4 T>C			rs1059111 A>T			
	GG	AG/AA	Adjusted OR a	P a	TT	CT/CC	Adjusted OR a	P a	AA AT/T	□ Adjusted OR ª	P a	
	(Cases/Controls)		(95% CI)		(Cases/	Controls)	(95% CI)		(Cases/Control	s) (95% CI)		
Age, month												
≤18	48/120	76/185	1.03 (0.67-1.58)	0.903	48/117	76/188	0.99 (0.64-1.51)	0.947	48/118 76/182	1.00 (0.65-1.53)	0.997	
>18	82/181	167/326	1.13 (0.82-1.56)	0.451	83/182	166/325	1.12 (0.81-1.54)	0.487	84/182 165/32	1.10 (0.80-1.51)	0.559	
Gender												
Females	50/135	107/207	1.40 (0.94-2.09)	0.099	50/131	107/211	1.33 (0.89-1.99)	0.161	50/132 107/21	.0 1.35 (0.90-2.01)	0.143	
Males	80/166	136/304	0.93 (0.66-1.30)	0.662	81/168	135/302	0.93 (0.66-1.30)	0.657	82/168 134/30	02 0.91 (0.65-1.27)	0.574	
Sites of origin												
Adrenal gland	49/301	85/511	1.02 (0.70-1.49)	0.918	50/299	84/513	0.98 (0.67-1.43)	0.918	50/300 84/512	2 0.98 (0.67-1.44)	0.933	
Retroperitoneal	24/301	63/511	1.53 (0.93-2.50)	0.092	24/299	63/513	1.52 (0.93-2.48)	0.098	25/300 62/512	2 1.44 (0.89-2.34)	0.142	
Mediastinum	39/301	70/511	1.07 (0.70-1.62)	0.764	39/299	70/513	1.05 (0.69-1.59)	0.822	39/300 70/512	1.06 (0.70-1.60)	0.797	
Others	14/301	21/511	0.88 (0.44-1.76)	0.719	14/299	21/513	0.87 (0.44-1.74)	0.696	14/300 21/512	2 0.88 (0.44-1.75)	0.709	
Clinical stages												
I+II+4s	53/301	111/511	1.25 (0.87-1.78)	0.230	55/299	109/513	1.16 (0.81-1.65)	0.412	55/300 109/51	.2 1.17 (0.82-1.66)	0.392	
III+IV	70/301	119/511	1.01 (0.72-1.40)	0.978	70/299	119/513	1.00 (0.72-1.39)	0.985	70/300 119/51	.2 1.00 (0.72-1.39)	0.998	

OR, odds ratio; CI, confidence interval. <sup>a</sup> Adjusted for age and gender, omitting the corresponding stratification factor.

Given the common implication of NEFL in carcinogenesis as well as the relative paucity of investigation into its role in neuroblastoma risk, we were motivated to explore the association between the three NEFL polymorphisms and neuroblastoma risk in a two-center case-control study of Chinese children. Unexpectedly, both overall analysis and stratified analysis indicated that there was no significant association of these studied NEFL polymorphisms with neuroblastoma risk in the study populations. Such negative association results might be ascribed to the following possible reasons. First, relatively small sample size might result in inadequate power to detect moderate effects. Second, most of the common SNPs only have weak effects on cancer risk [24, 52]. Thus, effects of NEFL polymorphisms might be overridden by environmental factors or other genetic variations. Third, lack of association between these three NEFL polymorphisms and neuroblastoma in Chinese children may be attributed to ethnic differences in allele frequencies of the SNPs (Supplemental Table 2).

Although this study has some merits, limitations are inevitable. First, inherent bias could not be avoided in this case-control study, since all participants were enrolled from hospitals. Second, statistical power might be limited due to the moderate sample size. It is challenging to recruit more cases, due to the relatively low morbidity rate of neuroblastoma in China. Third, we only replicated three SNPs in NEFL gene in this study. There are other potentially functional SNPs that might modify neuroblastoma susceptibility. Thus, more SNPs are needed to be explored in the future. Fourth, details on environmental factors, including dietary intake, living environment, and parental exposures, were not accessible, which disables us to analyze

gene-environmental interaction. Fifth, though this is a two-center study, all the subjects were unrelated Chinese Han ethnicity. It should be cautious to extrapolate the results to other ethnic groups.

In all, we for the first time replicated the association between the *NEFL* gene polymorphisms and neuroblastoma risk using two independent populations in China. No significant association with neuroblastoma risk was detected. More epidemiological studies in different ethnic group are warranted before fully ascertaining the contribution of the *NEFL* gene polymorphisms to neuroblastoma susceptibility.

## Abbreviations

GWAS, genome-wide association study; SNP, single nucleotide polymorphism; NEFL, neurofilament light; LOH, loss of heterozygosity.

# Supplementary Material

Supplementary tables. http://www.jcancer.org/v09p0535s1.pdf

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## **Competing Interests**

The authors have declared that no competing interest exists.

#### References

- Maris JM, Hogarty MD, Bagatell R, Cohn SL. Neuroblastoma. Lancet. 2007; 369: 2106-20.
- Maris JM, Mosse YP, Bradfield JP, Hou C, Monni S, Scott RH, et al. Chromosome 6p22 locus associated with clinically aggressive neuroblastoma. N Engl J Med. 2008; 358: 2585-93.
- Campbell K, Gastier-Foster JM, Mann M, Naranjo AH, Van Ryn C, Bagatell R, et al. Association of MYCN copy number with clinical features, tumor biology, and outcomes in neuroblastoma: A report from the Children's Oncology Group. Cancer. 2017; 123: 4224-35.
- Moreno F, Lopez Marti J, Palladino M, Lobos P, Gualtieri A, Cacciavillano W. Childhood Neuroblastoma: Incidence and Survival in Argentina. Report from the National Pediatric Cancer Registry, ROHA Network 2000-2012. Pediatr Blood Cancer. 2016; 63: 1362-7.
- Bao PP, Li K, Wu CX, Huang ZZ, Wang CF, Xiang YM, et al. [Recent incidences and trends of childhood malignant solid tumors in Shanghai, 2002-2010]. Zhonghua Er Ke Za Zhi. 2013; 51: 288-94.
- Maris JM. Recent advances in neuroblastoma. N Engl J Med. 2010; 362: 2202-11.
- Matthay KK, Villablanca JG, Seeger RC, Stram DO, Harris RE, Ramsay NK, et al. Treatment of high-risk neuroblastoma with intensive chemotherapy, radiotherapy, autologous bone marrow transplantation, and 13-cis-retinoic acid. Children's Cancer Group. N Engl J Med. 1999; 341: 1165-73.
- Berthold F, Boos J, Burdach S, Erttmann R, Henze G, Hermann J, et al. Myeloablative megatherapy with autologous stem-cell rescue versus oral maintenance chemotherapy as consolidation treatment in patients with high-risk neuroblastoma: a randomised controlled trial. Lancet Oncol. 2005; 6: 649-58.
- Osajima-Hakomori Y, Miyake I, Ohira M, Nakagawara A, Nakagawa A, Sakai R. Biological role of anaplastic lymphoma kinase in neuroblastoma. Am J Pathol. 2005; 167: 213-22.
- Devoto M, Specchia C, Laudenslager M, Longo L, Hakonarson H, Maris J, et al. Genome-wide linkage analysis to identify genetic modifiers of ALK mutation penetrance in familial neuroblastoma. Hum Hered. 2011; 71: 135-9.
- Mosse YP, Laudenslager M, Khazi D, Carlisle AJ, Winter CL, Rappaport E, et al. Germline PHOX2B mutation in hereditary neuroblastoma. Am J Hum Genet. 2004; 75: 727-30.
- Bourdeaut F, Trochet D, Janoueix-Lerosey I, Ribeiro A, Deville A, Coz C, et al. Germline mutations of the paired-like homeobox 2B (PHOX2B) gene in neuroblastoma. Cancer Lett. 2005; 228: 51-8.
- Cheung NK, Dyer MA. Neuroblastoma: developmental biology, cancer genomics and immunotherapy. Nat Rev Cancer. 2013; 13: 397-411.
- De Roos AJ, Olshan AF, Teschke K, Poole C, Savitz DA, Blatt J, et al. Parental occupational exposures to chemicals and incidence of neuroblastoma in offspring. Am J Epidemiol. 2001; 154: 106-14.
- De Roos AJ, Teschke K, Savitz DA, Poole C, Grufferman S, Pollock BH, et al. Parental occupational exposures to electromagnetic fields and radiation and the incidence of neuroblastoma in offspring. Epidemiology. 2001; 12: 508-17.
- Patton T, Olshan AF, Neglia JP, Castleberry RP, Smith J. Parental exposure to medical radiation and neuroblastoma in offspring. Paediatr Perinat Epidemiol. 2004; 18: 178-85.
- Oldridge DA, Wood AC, Weichert-Leahey N, Crimmins I, Sussman R, Winter C, et al. Genetic predisposition to neuroblastoma mediated by a LMO1 super-enhancer polymorphism. Nature. 2015; 528: 418-21.
- McDaniel LD, Conkrite KL, Chang X, Capasso M, Vaksman Z, Oldridge DA, et al. Common variants upstream of MLF1 at 3q25 and within CPZ at 4p16 associated with neuroblastoma. PLoS Genet. 2017; 13: e1006787.
- Capasso M, Devoto M, Hou C, Asgharzadeh S, Glessner JT, Attiyeh EF, et al. Common variations in BARD1 influence susceptibility to high-risk neuroblastoma. Nat Genet. 2009; 41: 718-23.
- Nguyen le B, Diskin SJ, Capasso M, Wang K, Diamond MA, Glessner J, et al. Phenotype restricted genome-wide association study using a gene-centric approach identifies three low-risk neuroblastoma susceptibility Loci. PLoS Genet. 2011; 7: e1002026.
- Diskin SJ, Capasso M, Schnepp RW, Cole KA, Attiyeh EF, Hou C, et al. Common variation at 6q16 within HACE1 and LIN28B influences susceptibility to neuroblastoma. Nat Genet. 2012; 44: 1126-30.
- Wang K, Diskin SJ, Zhang H, Attiyeh EF, Winter C, Hou C, et al. Integrative genomics identifies LMO1 as a neuroblastoma oncogene. Nature. 2011; 469: 216-20.
- Russell MR, Penikis A, Oldridge DA, Alvarez-Dominguez JR, McDaniel L, Diamond M, et al. CASC15-S Is a Tumor Suppressor IncRNA at the 6p22 Neuroblastoma Susceptibility Locus. Cancer Res. 2015; 75: 3155-66.
- He BS, Sun HL, Xu T, Pan YQ, Lin K, Gao TY, et al. Association of Genetic Polymorphisms in the LncRNAs with Gastric Cancer Risk in a Chinese Population. J Cancer. 2017; 8: 531-6.
- Gao X. Multiple testing corrections for imputed SNPs. Genet Epidemiol. 2011; 35: 154-8.
- Cantor RM, Lange K, Sinsheimer JS. Prioritizing GWAS results: A review of statistical methods and recommendations for their application. Am J Hum Genet. 2010; 86: 6-22.
- Diskin SJ, Capasso M, Diamond M, Oldridge DA, Conkrite K, Bosse KR, et al. Rare variants in TP53 and susceptibility to neuroblastoma. J Natl Cancer Inst. 2014; 106: dju047.

- Cimmino F, Spano D, Capasso M, Zambrano N, Russo R, Zollo M, et al. Comparative proteomic expression profile in all-trans retinoic acid differentiated neuroblastoma cell line. J Proteome Res. 2007; 6: 2550-64.
- Capasso M, Diskin S, Cimmino F, Acierno G, Totaro F, Petrosino G, et al. Common genetic variants in NEFL influence gene expression and neuroblastoma risk. Cancer Res. 2014; 74: 6913-24.
- Zhang J, Lin H, Wang J, He J, Zhang D, Qin P, et al. LMO1 polymorphisms reduce neuroblastoma risk in Chinese children: a two-center case-control study. Oncotarget. 2017; 8: 65620-6.
- He J, Zhong W, Zeng J, Zhu J, Zhang R, Wang F, et al. LMO1 gene polymorphisms contribute to decreased neuroblastoma susceptibility in a Southern Chinese population. Oncotarget. 2016; 7: 22770-8.
- He J, Yang T, Zhang R, Zhu J, Wang F, Zou Y, et al. Potentially functional polymorphisms in the LIN28B gene contribute to neuroblastoma susceptibility in Chinese children. J Cell Mol Med. 2016; 20: 1534-41.
- He J, Wang F, Zhu J, Zhang R, Yang T, Zou Y, et al. Association of potentially functional variants in the XPG gene with neuroblastoma risk in a Chinese population. J Cell Mol Med. 2016; 20: 1481-90.
- He J, Wang F, Zhu J, Zhang Z, Zou Y, Zhang R, et al. The TP53 gene rs1042522 C>G polymorphism and neuroblastoma risk in Chinese children. Aging (Albany NY). 2017; 9: 852-9.
- Zhang Z, Zhang R, Zhu J, Wang F, Yang T, Zou Y, et al. Common variations within HACE1 gene and neuroblastoma susceptibility in a Southern Chinese population. Onco Targets Ther. 2017; 10: 703-9.
- Zhang Z, Zou Y, Zhu J, Zhang R, Yang T, Wang F, et al. HSD17B12 gene rs11037575 C>T polymorphism confers neuroblastoma susceptibility in a Southern Chinese population. Onco Targets Ther. 2017; 10: 1969-75.
- Lou J, Gong J, Ke J, Tian J, Zhang Y, Li J, et al. A functional polymorphism located at transcription factor binding sites, rs6695837 near LAMC1 gene, confers risk of colorectal cancer in Chinese populations. Carcinogenesis. 2017; 38: 177-83.
- Li J, Zou L, Zhou Y, Li L, Zhu Y, Yang Y, et al. A low-frequency variant in SMAD7 modulates TGF-beta signaling and confers risk for colorectal cancer in Chinese population. Mol Carcinog. 2017; 56: 1798-807.
- He J, Qiu LX, Wang MY, Hua RX, Zhang RX, Yu HP, et al. Polymorphisms in the XPG gene and risk of gastric cancer in Chinese populations. Hum Genet. 2012; 131: 1235-44.
- Previtali SC, Zerega B, Sherman DL, Brophy PJ, Dina G, King RH, et al. Myotubularin-related 2 protein phosphatase and neurofilament light chain protein, both mutated in CMT neuropathies, interact in peripheral nerve. Hum Mol Genet. 2003; 12: 1713-23.
- Arrambide G, Espejo C, Eixarch H, Villar LM, Alvarez-Cermeno JC, Picon C, et al. Neurofilament light chain level is a weak risk factor for the development of MS. Neurology. 2016; 87: 1076-84.
- Mersiyanova IV, Perepelov AV, Polyakov AV, Sitnikov VF, Dadali EL, Oparin RB, et al. A new variant of Charcot-Marie-Tooth disease type 2 is probably the result of a mutation in the neurofilament-light gene. Am J Hum Genet. 2000; 67: 37-46.
- Kerangueven F, Essioux L, Dib A, Noguchi T, Allione F, Geneix J, et al. Loss of heterozygosity and linkage analysis in breast carcinoma: indication for a putative third susceptibility gene on the short arm of chromosome 8. Oncogene. 1995; 10: 1023-6.
- Seitz S, Werner S, Fischer J, Nothnagel A, Schlag PM, Scherneck S. Refined deletion mapping in sporadic breast cancer at chromosomal region 8p12-p21 and association with clinicopathological parameters. Eur J Cancer. 2000; 36: 1507-13.
- Kurimoto F, Gemma A, Hosoya Y, Seike M, Takenaka K, Uematsu K, et al. Unchanged frequency of loss of heterozygosity and size of the deleted region at 8p21-23 during metastasis of lung cancer. Int J Mol Med. 2001; 8: 89-93.
- Schmidt H, Semjonow A, Csiszar K, Korsching E, Brandt B, Eltze E. [Mapping of a deletion interval on 8p21-22 in prostate cancer by gene dosage PCR]. Verh Dtsch Ges Pathol. 2007; 91: 302-7.
- Huang Z, Zhuo Y, Shen Z, Wang Y, Wang L, Li H, et al. The role of NEFL in cell growth and invasion in head and neck squamous cell carcinoma cell lines. J Oral Pathol Med. 2014; 43: 191-8.
- Chen B, Chen J, House MG, Cullen KJ, Nephew KP, Guo Z. Role of neurofilament light polypeptide in head and neck cancer chemoresistance. Mol Cancer Res. 2012; 10: 305-15.
- Revill K, Wang T, Lachenmayer A, Kojima K, Harrington A, Li J, et al. Genome-wide methylation analysis and epigenetic unmasking identify tumor suppressor genes in hepatocellular carcinoma. Gastroenterology. 2013; 145: 1424-35 e1-25.
- Kim MS, Chang X, LeBron C, Nagpal JK, Lee J, Huang Y, et al. Neurofilament heavy polypeptide regulates the Akt-beta-catenin pathway in human esophageal squamous cell carcinoma. PLoS One. 2010; 5: e9003.
- Dubrowinskaja N, Gebauer K, Peters I, Hennenlotter J, Abbas M, Scherer R, et al. Neurofilament Heavy polypeptide CpG island methylation associates with prognosis of renal cell carcinoma and prediction of antivascular endothelial growth factor therapy response. Cancer Med. 2014; 3: 300-9.
- Tan H, Bao J, Zhou X. A novel missense-mutation-related feature extraction scheme for 'driver' mutation identification. Bioinformatics. 2012; 28: 2948-55.