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Estrogen and estrogen receptors in kidney diseases

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ABSTRACT

Acute kidney injury (AKI) and chronic kidney disease (CKD) are posing great threats to global health within this century. Studies have suggested that estrogen and estrogen receptors (ERs) play important roles in many physiological processes in the kidney. For instance, they are crucial in maintaining mitochondrial homeostasis and modulating endothelin-1 (ET-1) system in the kidney. Estrogen takes part in the kidney repair and regeneration *via* its receptors. Estrogen also participates in the regulation of phosphorus homeostasis *via* its receptors in the proximal tubule. The ER α polymorphisms have been associated with the susceptibilities and outcomes of several renal diseases. As a consequence, the altered or dysregulated estrogen/ERs signaling pathways may contribute to a variety of kidney diseases, including various causes-induced AKI, diabetic kidney disease (DKD), lupus nephritis (LN), IgA nephropathy (IgAN), CKD complications, etc. Experimental and clinical studies have shown that targeting estrogen/ERs signaling pathways might have protective effects against certain renal disorders. However, many unsolved problems still exist in knowledge regarding the roles of estrogen and ERs in distinct kidney diseases. Further research is needed to shed light on this area and to enable the discovery of pathway-specific therapies for kidney diseases.

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Introduction

Although estrogen is classically regarded as a reproductive hormone in mammalian species, it also plays an important role in many other physiological processes (e.g., cell growth, development and differentiation, lipid and glucose homeostasis, renal endocrine function, immune function, etc.) [1–5]. The effects of estrogen in physiological and pathophysiological conditions are mediated by two different structural receptor classes, namely, estrogen receptor (ER) α/β and G proteincoupled estrogen receptor (GPER) [6]. Estrogen and its receptors are implicated in the development and progression of various diseases, including cancer, osteoporosis, endometriosis, neurodegenerative disorders, as well as cardiovascular, metabolic, and autoimmune diseases [7–12].

Acute kidney injury (AKI) and chronic kidney disease (CKD) are posing substantial threats to global health. The interdependent relationship between AKI and CKD further adds complexity to the clinical picture [13]. Recent experimental and clinical studies have suggested that estrogen and ERs serve pathophysiological roles in kidney diseases, including AKI, diabetic kidney disease, lupus nephritis, IgA nephropathy, complications of CKD, etc. This review mainly examines the roles of estrogen and its receptors in certain kidney diseases. We also discuss the structures, functions, mechanisms, and modulation of ERs, providing the basis for potential therapeutic interventions.

Structures and functions of ERs

As members of the nuclear receptor family, ERs are found mainly in the nucleus, but also in the cytoplasm and mitochondria [14]. The classical ER subfamily mainly consists of ER α and ER β [15]. Both of them consist of six functional domains A–F. The NH₂-terminal A/ B domain contains a ligand-independent transactivation function-1 (AF-1). It's noteworthy that the transactivation potency of AF-1 varies in a ligand-, cell type-, and promoter-specific manner [3]. The C domain (DNA binding domain) binds to DNA motifs called estrogen response elements (EREs). It also plays a part, to a minor extent, in the stability of ER dimerization [3]. The D domain is a hinge region between the C and E domains. It is involved in ER conformational changes,

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interaction with other transcription factors, nuclear translocation, and posttranslational modifications [16-18]. The E domain is identified as the ligand-binding domain (LBD) and the principal dimerization interface of the receptor, which contains a liganddependent activation function AF-2 [19,20]. The F domain is the least conserved region with high variability, and many nuclear receptors do not contain such a region [3]. ER α and ER β share a high degree of conservation in their C and E domains, while the other domains are more divergent [21]. The synergistic effect of AF1 and AF2 is required in the transcriptional regulation mediated by both receptors [22,23]. However, it is still unclear how AF-1 and AF-2 activities are regulated cooperatively by ligands. Recent studies have shown that the AF-2 contains an AF-1 suppression function element and that AF-1 is regulated in an AF-2-dependent manner [24,25].

ERα

In humans, ER α is encoded by the gene ESR1, located on chromosome 6, locus 6q25.1 [26]. ER α is primarily expressed in sex organs (breast, uterus and ovary, testes, epididymis, prostatic stroma), bone, liver, adipose tissue, cardiovascular, and central nervous system (CNS) [27,28]. The classic full-length 67 kDa ER α includes a DNA-binding domain, a ligand-binding domain, and two transcriptional activation functions (AF-1 and AF-2) [3]. In addition, two shorter isoforms (46 kDa ER α 46 and 36 kDa ER α 36) have been identified. ER α 46 lacks the N-terminal region harboring AF-1, whereas ER α 36 lacks both AF-1 and AF-2 and encodes a unique 29 amino acid sequence [29,30].

The functional role of ER α was first discovered from a clinical situation where a man bearing a mutation in the ER α gene developed a premature and severe metabolic syndrome [31]. It is now recognized that ER α is a key regulator of energy homeostasis and glucose metabolism and that the ER α pathway might represent a potential therapeutic target for the prevention or treatment of insulin resistance, type 2 diabetes mellitus, and non-alcoholic fatty liver diseases [16,32]. On the other hand, ER α is linked with a variety of cancers and metastases, including breast cancer, cervical cancer, lung carcinoma, and prostate cancer [33–36].

ERβ

In humans, ER β is encoded by the gene ESR2, located on chromosome 14 (14q23–24), and has five isoforms (ER β 1-5) [37]. These five isoforms exist as a result of alternative splicing of the last coding exon (exon 8). It is noteworthy that ER β 1 is the only full-function isoform with the native LBD and that the other isoforms do not have innate activities in their homodimeric forms but can heterodimerize with ER β 1 and enhance ER β 1induced transactivation in a ligand-dependent manner [38,39]. ER β and its isoforms have wider tissue distribution than ER α and they are expressed primarily during embryonic development and in the prostatic epithelium, bladder, ovary, colon, lung, adipose tissue, immune system, cardiovascular system, and CNS [27,40,41].

Similar to ER α , ER β is involved in cellular differentiation, mitochondrial bioenergetics, lipid and glucose metabolisms, energy expenditure, etc. [42–44]. ER β is generally thought to be a tumor suppressor gene and its expression is dysregulated in different cancers [45–47]. There is an increasing awareness that selective targeting of ER β signaling pathways might be useful in the treatment of several inflammatory and proliferative diseases [48,49]. Studies support the idea that ER β agonists can reasonably be used in hormone replacement therapy, early stage prostate and colon cancers, suppression of the immune system without negative effects on bone, and neuroprotection [50].

GPER

In the last decades, studies have suggested that apart from the classical steroid receptors $ER\alpha$ and $ER\beta$, the G protein-coupled estrogen receptor (GPER, formerly known as GPR30) also mediates the effect of estrogen in a rapid signaling pathway [51-53]. As a member of the G-protein coupled receptor superfamily, GPER is localized predominately within intracellular membranes in most cell types [54]. It is widely expressed in numerous tissues and organs, including the vessels, skeletal muscle, brain, heart, kidney, pancreas, and reproductive organs [55,56]. Studies have shown that GPER is involved in many physiological responses, including maintenance of vascular tonicity and blood pressure, reproduction, lipid and glucose metabolisms, immune and inflammatory responses, etc. [57-60]. For instance, mice lacking GPER exhibited metabolic syndrome, such as obesity, impaired glucose tolerance, or dyslipidemia [59,61]. Pharmacological modulation of GPER could promote pancreatic cell survival and improve glucose tolerance [62,63]. The effects of GPER are mediated via multiple signaling pathways, including the activation of adenylyl cyclase (AC)/protein kinase A (PKA), epidermal growth factor receptor (EGFR), PI3 kinases, as well as extracellular signal-regulated kinase (ERK) pathways and G protein-coupled pathways [64].



Figure 1. Classical (genomic) and non-genomic estrogen signaling pathways. E: estrogen; ERs: estrogen receptors; P: phosphorylation; ERE: estrogen receptor elements.

Mechanisms of estrogen action

The mechanisms of estrogen action are categorized into classical (genomic) and rapid (non-genomic) ones. In the classical pathway, estrogen binds to the ERs in the cytoplasm, leading to ER dimerization and translocation to the nucleus, where the estrogen-ER complex interacts with ERE sequences in target genes [16]. This process typically occurs within hours [65]. In recent decades, however, rapid or 'non-genomic' effects of estrogen (also termed non-nuclear or membrane initiated steroid signaling) has been reported [66]. This occurs through the ER located in or adjacent to the plasma membrane, or through other non-ER plasma membrane-associated estrogen-binding proteins, which usually takes seconds or minutes [67]. GPER has been identified as one of the main estrogen-sensitive receptors responsible for the rapid non-genomic action of estrogen [68]. The classical (genomic) and non-genomic estrogen signaling pathways are illustrated in Figure 1 [22,69].

Modulation of ERs

Selective estrogen receptor modulators (SERMs) are antiestrogens designed to compete with estrogen and modulate ER activity in a tissue-specific manner [70,71]. For instance, tamoxifen can exhibit antagonistic effect on mammary tissue, whereas it can have agonistic effects on other tissues such as the uterus, bone, and heart [72]. Raloxifene acts as an estrogen agonist in bone and an estrogen antagonist in uterine and breast tissues [73]. Similarly, bazedoxifene functions as a pure antagonist in the breast and an agonist in the bone [74]. Since ERs are nuclear transcription factors involved in the regulation of a variety of physiological and pathological processes in humans, modulation of the receptors either by SERMs or by agonists/antagonists might be beneficial for the prevention and treatment of various diseases [27].

Estrogen and ERs in kidney diseases

Acute kidney injury (AKI)

Gender differences in AKI epidemiology

The incidence of AKI has been steadily increasing, particularly among elderly hospitalized patients [75]. Epidemiological studies suggested that the mortality rates of AKI patients in hospital settings (including intensive care units) ranged from 17.5% to 64.7% [75,76]. As a broad clinical syndrome encompassing different etiologies, AKI is characterized by an abrupt decline of glomerular filtration rate, which is associated with high morbidity and mortality [77]. Various etiologies include pre-renal azotemia, acute tubular necrosis, acute glomerular/interstitial nephritis, acute vasculitic renal diseases, acute post-renal obstructive nephropathy, and mixed forms [78].

Clinical studies have shown that sex disparities might influence the susceptibility, progression, and therapeutic response to AKI [79-81] and that female sex might have a protective effect on the development of AKI [79,82]. Though a recent retrospective cohort study of AKI complicating acute myocardial infarction-related cardiogenic shock suggested that female sex was independently associated with higher in-hospital mortality, it did mention that women with AKI were older $(74 \pm 12 \text{ years})$, which highlighted the potential role of estrogen in AKI [83]. Another interesting phenomenon is that compared with females, males were endowed with lower mitochondrial respiratory capacity and poor antioxidant defense system, exhibiting fragmented and smaller mitochondria [84,85]. Given that the disrupted mitochondrial homeostasis plays a prominent role in the pathogenesis of AKI [86], this might partially explain the sex disparities in AKI. The gender dimorphism was also observed in animal experiments where male mice/ rats exhibited more severe AKI manifestations than their female counterparts through different mechanisms [87-91].

The role of silent mating type information regulator 2 homolog 1 (SIRT1) in mediating the renoprotective effects of estrogen on AKI has been systematically reviewed recently [92]. Experimental studies have indicated that SIRT1 exerts a protective effect against AKI through regulating oxidative stress, mitochondrial biogenesis, energy metabolism, inflammation and apoptosis [93–97]. The functional interaction between estrogen/ER and SIRT1 has been investigated in different disease models. Estrogen exerts protective effects against inflammation and mitochondrial dysfunction *via* ER α /SIRT1 pathway [98,99]. SIRT1 acts as an ER α co-activator and is required for modulation of ER α -signaling pathways [100]. In this regard, it is proposed that estrogen through SIRT1 might protect against AKI [92].

Estrogen and ERs in AKI settings

Renal ischemia–reperfusion injury (IRI). Renal IRI is characterized by a temporary shortage and subsequent restoration of blood supply and oxygen delivery to the kidney, initiating a cascade of deleterious cellular responses leading to ROS generation, inflammation and tubular cell death, leading to AKI [101,102]. As one of the leading causes of AKI, renal IRI often occurs with kidney transplantation, postoperative hypotension, traumatic hemorrhage, cardiovascular surgery, cardiac arrest, and cardiopulmonary resuscitation [103–108].

Clinical studies regarding the association of gender with kidney allograft outcome have mixed results. While some clinical observations suggested that female recipients generally had a better graft function and survival than male counterparts, others indicated that the risk of developing graft failure varied in women stratified by age and donor sex [109–112]. Although the effect of gender disparity in the susceptibility and pathogenesis of IRI in the transplanted kidney is less well established in humans, the majority of experimental studies indicate that females had better outcome of kidney transplantation than males due to increased IRI tolerance [113]. This sex-related differences in IRIinduced AKI might be attributed to the depression of renal sympathetic nervous system with endogenous estrogen and the subsequent reduced regional noradrenaline level [89,114,115]. As expected, the supplementation of estrogen prior to the induction of IRI protected kidney function in female mice and neutered males. Studies also revealed that estrogen administration after cardiac arrest and resuscitation ameliorated AKI in both male and female mice [116,117]. In contrast, testosterone enhances kidney susceptibility to IRI through inhibiting the activation of nitric oxide (NO) synthases and Akt signaling [118]. Notably, renal IRI was exacerbated in female $ER\alpha$ knockout mice [113].

The roles of estrogen and ERs in the signaling mechanisms of IRI-induced AKI have been reported. Estrogen attenuated renal IRI through the activation of peroxisome proliferator-activated receptor γ (PPAR γ), a nuclear receptor which plays a vital role in the maintenance of renal metabolic homeostasis [119-121]. The overproduction of renal endothelin-1 (ET-1), which plays a critical role in the initiation of AKI and the subsequent transition to CKD through its two receptors ET_A and ET_B, was suppressed with the administration of estrogen in rats challenged with IRI [122-125]. This effect was probably mediated by ER β and GPER1 in the kidney [126,127]. Estrogen also reduced inflammation and accelerated injured tubular cell regeneration in male rats after IRI-induced AKI [128]. In the uninephrectomized rat model of IRI, the renoprotective effect of estrogen was mediated by the activation of PI3K/Akt pathway followed by increased endothelial nitric oxide synthase (eNOS) phosphorylation in the affected kidneys [129]. Another mechanism might be estrogenmediated antagonism on N-methyl-D-aspartate receptors (NMDAR) that were expressed in nephrons [130]. The activation of GPER1 also protected the kidney from IRI through enhancing glomerular endothelial barrier function and regulating the content of NO in renal interlobular artery smooth muscle and endothelial cells

[131,132]. Renal Na⁺/K⁺-ATPase is a well-recognized energy-dependent sodium pump which helps maintain the electrolyte and fluid balance via manipulating the transport of certain solutes [133,134]. Under the circumstance of renal IRI, the impaired activity of Na⁺/K⁺-ATPase contributed to abnormal hydroelectrolyte status [135,136]. A sex disparity in the alterations of this enzyme was observed following renal IRI, favoring the protective role of estrogen in the stability and modulaof Na⁺/K⁺-ATPase tion upon various renal insults [137,138].

Together, these results demonstrate that there exists a sex dimorphism in IRI-induced AKI. However, the discrepancy between clinical observations and experimental findings regarding this aspect remains to be investigated. Estrogen and ERs might have potential therapeutic implications in the treatment of renal IRI, but recent studies have pointed out that the renoprotective effect of exogenous estrogen becomes evident only when administered at a dose above the physiological level [139]. It has also been doubted that whether the sequelae of renal IRI (i.e., subsequent kidney fibrosis) could actually be brought to a halt by estrogen. Therefore, the therapeutic effect of estrogen and its analogues derived from the existing animal studies needs to be cautiously interpreted when applied to human patients.

Septic AKI. Sepsis-induced AKI is the leading cause of AKI in the intensive care unit (ICU) and accounts for nearly half of all AKI events, which is associated with increased mortality and morbidity [106,140]. A multicenter prospective study including 1177 ICU patients with sepsis in 24 European countries reported a 51.2% incidence of AKI with a mortality rate of 41.2% [141]. A multicenter retrospective study of [146,148]. Chinese hospitalized adults found AKI in 47.1% of sepsis cases [142].

Studies regarding the role of estrogen in septic AKI have conflicting results. Feng et al. found that serum estrogen levels were correlated with the severity of renal dysfunction and that increased estrogen levels predicted the development of newly onset AKI within a month [143]. A recent post-hoc analysis of patients with sepsis from the Randomized Evaluation of Normal versus Augmented Level renal replacement therapy (RENAL) trial suggested that female sex was associated with improved survival in septic patients with severe AKI [82]. Trentzsch et al. found that female patients had lower rates of sepsis and renal failure than males following traumatic injury and hemorrhagic shock [144]. Animal studies also demonstrated that estrogen and its

selective modulator raloxifene had protective effects on renal function in ovariectomized rats with sepsis through the activation of autophagy [145,146]. Several preclinical studies indicated that the activation of ERs (mainly ER α and ER β) could regulate inflammatory responses and control sepsis-induced multiorgan dysfunction [147–149].

The exact mechanisms responsible for the roles of estrogen and ERs in septic AKI remain to be elucidated. Further studies are needed to uncover the potential roles of estrogen and ERs in mediating the gender differences in septic AKI. Though a majority of experimental studies have demonstrated the salutary effects of estrogen and ER agonists on the outcome of sepsisinduced multiple organ failure including AKI, clinical investigations have not reached a consensus in this respect. The remaining gap between the bench and the bedside prompts us to take into consideration the different clinical study protocols. For example, the hormonal status or the level of estrogen of a patient at the time of sepsis may have an effect on the clinical outcome [150,151]. Studies providing information regarding hormonal status (oral contraceptives, menstrual cycle, and hormone replacement therapy) at the time of patient enrollment are encouraged. It is important to note that kidney is just one of the many involved organs in sepsis and that the overall effects of estrogen in other organs or systems need addressing. Also, fluid resuscitation and antibiotics are still the mainstay of treatment for patients with sepsis and, therefore, estrogen-related therapies may be developed for the best interests of the individual patient.

Drug-induced AKI. Drug-induced AKI accounts for 0.7% to 26% of cases with AKI among hospitalized patients [152–154]. Various drugs that are metabolized in the kidney and cleared *via* glomerular filtration and/ or tubular secretion may induce AKI from a variety of mechanisms [155]. In the proximal tubules, apical transport of the aminoglycosides and basolateral transport of cisplatin increase the risk for AKI [156]. Acute interstitial nephritis is another form of drug-induced AKI, which typically develops from medications that incite an allergic reaction (e.g., antibiotics, proton pump inhibitors, non-steroidal anti-inflammatory drugs, etc.) [157–159].

Studies regarding the sex differences in the susceptibility to drug-induced AKI have conflicting results. While some suggested that females were more vulnerable to aminoglycoside- and cisplatin-induced nephrotoxicity, others found no such gender disparities [160–162]. At least in the field of drug-induced acute interstitial nephritis, epidemiological characteristics showed a female preponderance [159,163]. This might be attributed to gender differences in the expression of drug transporters on proximal renal tubules [164]. However, the issue of gender differences in the susceptibility to nephrotoxins is far more nuanced. There is actually no impact of gender on the protein levels of some key drug transporters in human kidney samples, as revealed by liquid chromatography-tandem mass spectrometry-based targeted proteomics [165,166]. On the other hand, sex differences in the expression of kidney genes responsible for the metabolism of certain drugs have been confirmed in experimental studies [164,167]. For example, females could better handle mercuric chloride-induced tubular injury because of their correspondingly increased renal expression of organic anion transporter 1/3 (OAT1/3) and multidrug resistance-associated protein 2, leading to higher excretion of mercury and less likelihood of AKI [168]. A sexhormone dependent pattern of the expression of OAT2 was also observed in mouse and rat kidneys (i.e., OAT2 could be upregulated by estrogen and progesterone) [169].

The roles of estrogen and ERs in drug-induced AKI have been investigated. Preclinical studies demonstrated that young females exhibited decreased susceptibility to cisplatin-induced AKI, highlighting the potential role of estrogen [154,170,171]. Moreover, estrogen and GPER1 agonist G-1 had protective effects against human tubular epithelial cell injury induced by methotrexate [172]. In a rat model of heavy metal-induced AKI, tamoxifen prevented mercury-induced toxicity on mitochondrial energy-dependent functions in the kidney [173]. In contrast, fulvestrant, an ER α down-regulator, exacerbated kidney injury in rats with AKI induced by gentamicin [174,175]. Further studies are warranted to determine the roles of estrogen and ERs in drug-induced AKI.

It is well established that renal tubules are very susceptible to various insults including a range of commonly used drugs [176]. However, there exist distinct differences regarding the vulnerability to certain drugs among different segments of tubules or regions of the kidney. Whether or not estrogen or ERs confer comprehensive protection within the kidney is still unknown. Besides, drug-induced nephropathy can also be accompanied by other organ disorders such as hepatotoxicity and thrombocytopenia [177,178]. Management of these comorbidities, therefore, is equally important. A physician should have a high index of suspicion for the risk factors associated with drug-induced AKI in that the underlying health conditions of the patients may have a great impact on the clinical outcome. Correcting risk factors and treating underlying diseases should be emphasized.

Chronic kidney disease (CKD)

Gender differences in CKD epidemiology

According to the 2017 KDIGO guidelines, CKD is defined as abnormalities of kidney structure or function, present for more than 3 months, with implications for health [179]. Major causes of CKD include diabetic nephropathy, IgA nephropathy, lupus nephritis, and membranous nephropathy [180,181].

Epidemiological studies suggested that females experienced slower renal function decline than males, possibly owing to the protective effect of estrogen or the damaging effect of testosterone [182,183]. Studies also revealed that premenopausal women (particularly those less than 45 years old) who underwent bilateral oophorectomy were at higher risk of developing CKD [184], highlighting the renoprotective effect of estrogen [185,186]. On the other hand, CKD itself is associated with hypothalamic-pituitary-ovarian dysfunction, which results in the earlier onset of menopause in uremic women [187]. The estrogen-based hormone replacement therapy seems to ameliorate renal dysfunction and delay CKD progression in postmenopausal patients, which might be partially due to the increased renal NO production and reduced oxidative stress [188–190]. Similarly, animal studies also indicated that the progression of kidney disease was slower in females.

Estrogen and ERs in CKD settings

Diabetic kidney disease (DKD). As a major microvascular complication of diabetes mellitus, DKD is the most common cause of end-stage renal disease (ESRD) in developed countries. DKD has severe individual and societal consequences, owing to its high morbidity, mortality, and health-care costs [191]. Clinical studies regarding the sex-specific differences in the prevalence and progression of DKD are inconsistent. While some studies reported a higher incidence of DKD and subsequent ESRD in male population, others suggested a female predominance, or no such differences [192–194]. Different study protocols (e.g., age, race, types of diabetes, etc.) and confounding factors might explain this inconclusive link between sex and DKD [195]. The role of sex hormones including estrogen in the setting of DKD has not yet been determined. Nevertheless, a majority of studies have shown that an imbalance of sex hormones does exist among both

The roles of estrogen and ERs in DKD have been investigated. It was reported that the deficiency of estrogen (ovariectomy) could exacerbate renal pathological manifestations (e.g., glomerulosclerosis and tubulointerstitial fibrosis) in rats with diabetes induced by streptozotocin (STZ). The supplementation of estrogen or raloxifene attenuated these changes through reducing lipid peroxidation and oxidative stress [199,201,202]. Similarly, in male diabetic mice with or without orchiectomy, estrogen effectively inhibited DKD progression (e.g., reduced glomerulosclerosis, albuminuria and glomerular hyperfiltration) [198]. In a model of spontaneous DKD, the estrogen treatment attenuated mesangial expansion and glomerular basement membrane (GBM) thickening but failed to ameliorate proteinuria and glomerulosclerosis in male Otsuka-Long-Evans-Tokushima-Fatty rats [203]. Mechanistic studies showed that estrogen exerted its renoprotective effect on DKD through upregulating matrix metalloproteinase (MMP)-2 and MMP-9 to accelerate the degradation of extracellular matrix (ECM) [204]. In addition, estrogen attenuated albuminuria and ECM deposition by regulating the expression of transforming growth factor- β 1 (TGF- β 1) and its downstream signaling pathway [205]. Moreover, both estrogen and raloxifene ameliorated albuminuria and mesangial expansion in ovariectomized db/db mice, possibly via inhibiting TGF-β1-induced fibronectin transcription and activator protein-1 (AP-1) activity [206].

ER α gene polymorphisms have been associated with the risks of developing CKD with type 2 diabetes mellitus in the African American population and of developing DKD in girls with type 1 diabetes [207,208]. ER α and its splice variants exhibited protective effects against DKD induced by STZ in female mice, as evidenced by reduced glomerular size, hyperfiltration, macrophage infiltration, and proteinuria [209]. The ER α signaling was proposed as one of the renal signaling pathways involved in the pathogenesis of DKD and might be a promising target for the treatment of nephropathy in diabetic patients [210].

Since glomerular mesangial cell (MC) and podocyte injuries are involved in the progression of proteinuria and DKD, quite a few studies have focused on the effects of estrogen and ERs on modulating these cellular processes [211]. Both ER α and ER β are expressed in human/murine MCs and podocytes [212,213]. It was demonstrated that estrogen played a protective role in the regulation of proliferation and apoptosis of these

cells via its receptors. For example, estrogen could increase the degradation of ECM to slow the progression of DKD through upregulating MMP-9 in MCs [213]. It also inhibited podocyte apoptosis through binding to ER β , which was associated with the activation of the JAK2/STAT3 signaling pathway [214]. Moreover, estrogen or tamoxifen could improve albumin excretion, reduce glomerular size, and decrease matrix accumulation *via* upregulating the expression of ER β and downregulating TGF- β in podocytes from db/db mice [215]. Podocytes isolated from these mice had a higher level of F-actin and lower level of caspase-9, indicating that estrogen might protect against podocyte injury in DKD through regulating both actin cytoskeleton and apoptosis [216]. In ERa knockout mice, podocyte injury (increased desmin and decreased nephrin and Wilms tumor-1) and apoptosis developed, and the estrogen treatment could prevent these changes via the activation of extracellular signal-regulated protein kinase (ERK) signaling pathway [217,218].

Studies regarding the role of GPER1 in DKD are limited. GPER1 agonist icariin was reported to exert protective effects against oxidative stress and fibrosis in male rats with DKD induced by STZ [219]. GPER1 was crucial in regulating MC migration and ECM production in response to TGF- β 1 [220]. It was through GPER1 that icariin reduced the deposition of type IV collagen and fibronectin induced by high glucose in human/rat MCs by inhibiting TGF- β /Smad and ERK1/2 signaling pathways [221]. The activation of GPER1 also inhibited high glucose-induced podocyte apoptosis by modulating Bcl-2 expression and mitochondrial translocation [222].

The altered renin–angiotensin system (RAS) plays a crucial role in the context of DKD. Male STZ-induced diabetic mice administered with angiotensin II had more prominent albuminuria, glomerular hypertrophy and mesangial expansion than females [223]. Female rats had lower levels of albuminuria and renal angiotensinogen (AOGEN) mRNA compared with male rats in the development of STZ-induced DKD [224]. This indicates the sex dimorphism regarding RAS in DKD, highlighting the potential role of estrogen in this disease.

In brief, a majority of experimental studies back up the idea that estrogen might play a protective role in DKD through attenuating glomerular MC and podocyte injuries *via* its receptors and through its sex dimorphism in RAS. However, many controversies and questions remain regarding the roles of sex hormones in the pathophysiology and progression of DKD. The imbalance of sex steroids (e.g., estrogen versus testosterone) has been confirmed in the setting of diabetes from clinical trials, which suggests that restoration of hormonal homeostasis is far more important than just supplying estrogen as is done in most of the animal studies. Diabetes mellitus, especially type 2, is recognized as the cardiovascular disease equivalent and is associated with other end-organ complications such as retinopathy and neuropathy. The potential role of estrogen or ERs in these aspects has yet to be investigated. Moreover, the complexity and diversity of signaling networks involved in diabetes require more drugs developed to specifically target these dysregulated pathways.

Lupus nephritis (LN). As one of the most severe manifestations of systemic lupus erythematosus (SLE), LN is characterized by proteinuria, hematuria and progressive renal dysfunction. It affects over half of all patients diagnosed with SLE and is a major risk factor for overall morbidity and mortality [225]. The female to male incidence of SLE mounts to nine during the fourth decade of life and declines subsequently until the seventh or eighth decade, disproportionately affecting women of reproductive age [226].

The predominance of females among patients with SLE underlines a pathogenic role for female hormones including estrogen [227]. Though mixed results exist regarding whether exogenous estrogen (either oral contraceptives or hormone replacement therapy) may increase the risk for SLE, endogenous estrogen and ERs have been confirmed to be associated with the modulation of both innate and adaptive immune responses in SLE [228-230]. The development and function of immune cells such as T/B cells and plasmacytoid dendritic cells (DCs), as demonstrated by a large body of literature, can be directly influenced by estrogen [231,232]. For instance, estrogen can interfere with the normal tolerance of naive DNA-reactive B cells and amplify the activation of autoreactive B cells [233,234]. Clinical studies further revealed that a baseline sexbiased difference of differentially expressed genes (DEGs) in B cells existed between healthy females and males, and that some of the DEGs were relevant to estrogen-induced type I interferon (IFN)/B cell activator factor (BAFF) signaling pathway [235,236]. Moreover, estrogen could stimulate the expression of CD40 ligand, a molecule involved in the pathogenesis of LN, in peripheral T cells isolated from patients with SLE [232,237]. The detrimental impact of estrogen on SLE was also confirmed by its inhibitory effect on T cell apoptosis, thus allowing for the persistence of autoreactive cells [231]. One randomized controlled trial evaluating the efficacy of fulvestrant in SLE patients found that selective ERs blocking could reduce disease activity as revealed by decreased expression of T cell activation

markers and improved SLE disease activity index (SLEDAI) [238]. Interestingly, studies conducted from different nations have shown that male SLE patients had more severe disease patterns and worse prognoses compared with females. For instance, renal involvement, especially type IV LN with nephrotic syndrome, was more frequent in males [239–247].

In murine models of lupus, female mice were more susceptible to the development of glomerulonephritis [248] and the estrogen treatment accelerated the progression of LN [249]. This sex discrepancy suggests a potential role for estrogen in this autoimmune disease. A majority of animal studies concluded that estrogen might play a detrimental role in the pathogenesis of LN. The estrogen treatment could induce a lupus phenotype with kidney damage in wild-type mice and resulted in more immune cells expressing $ER\alpha$ (e.g., CD4⁺ and CD8⁺ T cells, dendritic cells, and macrophages) in autoimmune-prone mice [250]. The administration of estrogen to lupus-prone mice increased the levels of BAFF as well as anti-C1g and anti-dsDNA antibodies, and expanded the population of self-reactive idiotypic B and T cells, thereby accelerating the progression of glomerulonephritis in lupus [249,251]. Through binding to ERs, environmental estrogens (e.g., diethylstilbestrol and bisphenol-A) that mimic estrogenic activity also enhanced autoantibody production and IgG immune complex deposition in the kidney in lupus-prone mice after orchiectomy [252].

Studies of ERs in the pathogenesis of LN and other immune-mediated glomerular diseases have mixed results. Those in favor of the detrimental role of ERs argued that $ER\alpha$ acted in a B cell-intrinsic manner to promote B cell activation specifically in female lupusprone mice, and that its deletion in B cells attenuated autoantibody production and glomerular immune complexes [253]. Furthermore, ERa knockout female mice developed less severe nephritis induced by nephrotoxic serum while manifesting normal autoimmune humoral response [12]. Similarly, other studies indicated that ERa deficiency conferred protection against proteinuria and tubular injury in female lupus-prone mice [254,255]. However, Scott et al. pointed out that previously reported 'ER α knockout mice' were not in fact $ER\alpha$ null but expressed an N-terminally truncated $ER\alpha$ (ERa short, similar to an endogenously expressed ERa46 variant), and that these mice were protected from LN after ovariectomy only if supplemented with estrogen [256,257]. This suggested that the ER α short protein had a protective role in LN and that endogenous $ER\alpha$ variants might represent a potential therapeutic target. Likewise, Shim et al. suggested that mice lacking $ER\alpha$

rather than lacking ER β developed autoimmune glomerulonephritis, proteinuria, and plasma cell infiltration in the kidney without antigen challenge [258]. Clinical data have shown that ER α polymorphism is associated with SLE susceptibility in the overall and Asian populations as well as the renal and cutaneous involvements [259–261].

Mechanistic studies regarding the roles of estrogen and ERs in LN and immune-mediated glomerular injury are limited. Estrogen was reported to enhance the activation of conventional DCs through modulating IFNdependent and independent pathways in both wildtype and lupus-prone mice [262]. ER α was required for Toll like receptor (TLR)-induced inflammation and the generation of both plasmacytoid DCs and interleukin-17 (IL-17) producing cells, all of which were implicated in SLE [263,264]. Another possible mechanism was that it was through ER α that the estrogen induced the expression of tumor necrosis factor-like weak inducer of apoptosis (TWEAK) and aggravated LN [265].

In conclusion, epidemiological studies and the majority of animal studies highlight the possibility of the involvement of estrogen/ERs in the pathophysiology of LN. The specific mechanisms underlying its role require further investigation. A recent study revealed that aberrant posttranslational modification of steroid receptors including ER α in T cells contributed to the gender-bias of SLE and that targeting ER α could improve the symptoms of lupus patients [266]. Since there are limited clinical trials investigating the efficacy of ER α antagonism in these patients, this pioneering finding may provide a novel molecular basis for future precision treatment.

IgA nephropathy (IgAN). IgAN is the most common glomerulonephritis globally leading to ESRD [267]. Mesangial hypercellularity with deposition of IgA-containing immune complexes is one of the key pathological features.

Male gender was an independent risk factor for developing ESRD in IgAN and male patients presented with worse clinicopathologic manifestations than females [268,269]. Consistently, an animal study suggested that trichothecene vomitoxin (VT)-induced IgAN had a male predilection in B6C3F1 mice and that these males had more severe disease patterns [270]. To further investigate the role of estrogen in IgAN, a study found that castration of female B6C3F1 mice increased the severity of VT-induced IgAN, but supplementation with estrogen did not attenuate this effect but rather increased disease severity [271].

The polymorphism of $ER\alpha$ gene might be associated with the pathogenesis of IgAN [272]. Among several

pathways regulating the proliferation of MCs in IgAN, ER α acted as a hub protein that could affect a set of proteins and transcription factors involved in the disease process [273]. A recent study using bioinformatics analysis based on the Gene Expression Omnibus database found that some of the key genes upregulated in IgAN were linked with the estrogen signaling pathway [274]. Another study reported that the expression of glomerular ER α in IgAN renal tissue decreased with the increasing severity of disease, proposing ER α as an independent factor affecting the prognosis of patients with IgAN [275]. Further studies are needed to elucidate the roles of estrogen and its receptors in the pathogenesis of IgAN.

Other CKD models. In animal models of CKD, estrogen was reported to exert protective effects against renal pathologies (e.g., glomerular hypertrophy, atrophic tubules, loss of podocytes, etc.) through attenuating oxidative stress and inflammation [276-278]. In the aging Dahl salt-sensitive (DSS) rats with ovariectomy, estrogen could attenuate glomerulosclerosis and tubulointerstitial fibrosis [279]. Spontaneously hypertensive stroke-prone (SHRSP) rats with uninephrectomy treated with estrogen had reduced albuminuria, glomerulosclerosis, and tubulointerstitial fibrosis [276]. Estrogen also preserved the contralateral kidney function in rats with unilateral ureteral obstruction (UUO), as revealed by reduced expressions of TGF- β and α -smooth muscle actin (α -SMA) [280]. As a SERM, tamoxifen conferred protection against kidney fibrosis induced by UUO via modulation of ER α -mediated TGF- β 1/Smad pathway [281]. Its anti-fibrotic effect was also confirmed in a model of chronic nephropathy (hypertensive nephrosclerosis by chronic inhibition of NO synthesis) where treated rats had less albuminuria, glomerulosclerosis and interstitial fibrosis than untreated ones [282].

The role of ERs has also been investigated. In an adenine-induced CKD model, male rats developed more severe kidney damage than female ones, which might be associated with decreased renal expression of ER α [283]. However, a study using cotton rats to simulate the spontaneous CKD in the elderly population showed that both ER α and ER β were strongly present in the renal tubules, which correlated with renal cystic lesions, inflammation and fibrosis [284]. Renal tubular epithelial cell regeneration and proliferation contribute to chronic kidney fibrosis, which ultimately leads to CKD [285,286]. Estrogen participated in the renal tubular regeneration process by modulating cell proliferation through the activation of both ER α and GPER1 receptor [287]. The role of ER α in kidney regeneration

and growth was further confirmed in an animal model, where $ER\alpha$ knockout female mice exhibited reduced compensatory kidney growth [288].

Taken together, a majority of the studies suggest that estrogen might exert protective effects against CKD through ameliorating renal fibrosis and that SERMs might be a group of drugs of interest for CKD. However, pharmacokinetics in CKD should be taken into account because of the altered renal and non-renal drug clearance in this population [289]. SERMs such as tamoxifen and raloxifene have been confirmed to be associated with the risks of deep vein thrombosis and pulmonary embolism [290,291]. The long-term safety and efficacy of these agents need to be investigated. Besides, little progress has been made regarding the prevention and reversal of kidney fibrosis up to now. Whether these drugs confer protection to CKD patients is still unclear.

Complications of CKD

Chronic kidney disease-mineral and bone disorder (CKD-MBD)

CKD-MBD is a major complication of CKD characterized by biochemical abnormalities, bone disorders, and vascular/soft tissue calcification, contributing to cardiovascular disease and mortality [292]. The abnormal metabolisms of calcium, phosphorus, parathyroid hormone (PTH), and vitamin D are key disturbances in CKD-MBD [293].

Physiologically, estrogen takes part in the regulation of phosphorus homeostasis by activating $ER\alpha/ER\beta$ mediated phosphate transporter NaPi-IIa in kidney proximal tubules [294]. Studies regarding the impact of estrogen on phosphate metabolism are mainly confined to the general population. The endogenous estrogen levels in older men were inversely associated with serum phosphorus levels [295]. Consistently, postmenopausal women receiving estrogen treatments had lower serum levels of phosphorus by increasing the urinary excretion of phosphorus [296]. Another SERM, bazedoxifene, could reduce serum phosphorus levels, thereby improving renal functions in postmenopausal osteoporotic women without severe renal insufficiency [297].

CKD-associated osteoporosis is more prominent in postmenopausal uremic patients, highlighting the potential role of estrogen in the disease process [298,299]. For instance, raloxifene was effective in improving bone mineral density (BMD) in postmenopausal osteoporotic women with CKD [300–303]. Further studies revealed that ER α gene polymorphism might dictate the different outcomes of BMD in postmenopausal hemodialysis patients who received raloxifene [304]. Animal studies showed that the deficiency of estrogen (ovariectomy) contributed to the impaired fixation of titanium implants in the femurs and further bone loss in the mandibles of uremic mice (5/6 nephrectomy) [305,306]. In an animal model of progressive CKD (male Cy/+ rats with autosomal dominant cystic kidney disease), raloxifene improved skeletal material properties (those independent of bone mass) and structural properties [307]. Mechanistic studies revealed that estrogen could inhibit PTH-stimulated osteoclast-like cell formation through blocking both the cAMP-dependent protein kinase (PKA) and the calcium/ PKC pathway [308,309].

Vascular calcification is highly prevalent in the CKD population, which is associated with major adverse cardiovascular events. Hyperphosphatemia and the abnormality of renin-angiotensin-aldosterone system are contributing factors for this phenomenon [310,311]. Studies focusing on the effects of estrogen on phosphate/angiotensin II-induced vascular smooth muscle cells (VSMCs) proliferation revealed that estrogen and raloxifene prevented the mineralization of VSMCs [312]. Further analysis revealed that this protective effect was mediated by the transactivation of growth arrest-specific gene 6 (Gas6) and upregulation of liver kinase B1 (LKB1) in the presence of ERa [313,314]. A randomized controlled trial suggested that estrogen, which had a favorable effect on oxidative stress, might protect against atherosclerotic cardiovascular disease in postmenopausal women with ESRD on hemodialysis [315].

In conclusion, these results suggest that estrogen and SERMs are of clinical significance in the management of CKD-MBD through attenuating biochemical abnormalities, bone disorders, and vascular calcification. Potential adverse effects of these agents are still to be alerted, especially in hemodialysis patients. Although there were no reported breast cancer or thromboembolism associated with the use of raloxifene by the aforementioned studies, the long-term safety and efficacy of SERMs need to be confirmed by larger prospective studies with longer follow-ups. Moreover, the mainstays of therapy for CKD-MBD are dietary phosphorus restriction, modification of dialysis prescriptions, phosphate binders, calcitriol or vitamin D analogs, and calcimimetics [316]. Parathyroidectomy (PTX) is considered when patients develop drug resistance or side effects [317]. Recent studies also confirmed the feasibility and efficacy of microwave ablation treatment for patients ineligible for PTX [318,319]. Therefore, more studies are warranted to evaluate whether there is an

additional benefit of adding estrogen or SERMs to the established treatment regimen.

Uremic coagulopathy

Epidemiological studies revealed an increased susceptibility to major bleeding events in dialysis patients [320,321]. Potential etiologies include platelet dysfunction, dysfunctional von Willebrand factor, decreased production of thromboxane, uremic toxins, and anemia [322,323]. The bleeding tendency in uremic patients was further complicated or exacerbated by their comorbidities and medications (e.g., hypertension, atrial fibrillation with warfarin therapy, cerebrovascular disease with anticoagulants) [318,324-326]. For patients who were refractory to routine interventions for uremic bleeding, conjugated estrogens might be another option [327-329]. Overall, there lacks concrete data regarding the efficacy of estrogen in the prevention and treatment of bleeding in CKD patients. Most of the studies investigating the hemostatic effect of estrogen were confined to the general population, particularly in the setting of recurrent bleeding from arteriovenous malformations, and still there were conflicting results [330,331]. Several case reports have suggested that estrogen, as part of the hormonal therapy, might be effective in the management of recurrent bleeding or hematoma in uremic patients [332-334]. An animal study suggested that estrogen could shorten the prolonged bleeding time in uremic rats and that this hemostatic effect was neutralized by ER antagonists [335]. A possible hypothesis was that the impaired Larginine-NO signaling under uremic conditions led to acute endogenous deprivation of estrogen and resulted in inhibited platelet aggregation and adhesion [336-338].

The long-term effects of estrogen as a component of hormone therapy in postmenopausal women have been well described by the Women's Health Initiative trial and related studies, and a consensus has been reached that women who start hormone therapy within 10 years of menopause onset may have less risk of thromboembolic complications [339,340]. However, this notion should be taken prudently in CKD population, as suggested by KDOQI guidelines and several studies [341-343]. CKD is an established risk factor for venous thromboembolism [344,345]. Patients with CKD are at high risk for thromboembolic events due to endothelial dysfunction and retention of indolic solutes [346,347]. Estrogen can cause thrombosis through elevating the levels of prothrombotic factors and decreasing the levels of protein S and antithrombin [348]. Because of the

altered drug pharmacokinetics in patients with CKD, exogenous estrogen treatment might be associated with an increased risk of arteriovenous access thrombosis [348–350]. Likewise, an animal study suggested that the estrogen supplementation in rats with renal impairments produced thrombotic microangiopathic lesions in the kidneys [351].

The dual effect of estrogen in uremic coagulopathy needs further investigation. Clinical application of estrogenic agents in the treatment of uremic bleeding requires more explorations. The risk-benefit ratio of estrogen in postmenopausal women with advanced CKD is still unknown and clinical trials addressing this issue are needed.

Conclusions and research gaps

Beyond its conventional role in the reproductive system, estrogen also functions in diverse developmental and physiological processes through its different receptors. Estrogen and ERs are crucial in maintaining mitochondrial homeostasis and modulating ET-1 system in the kidney, which points forward to their indispensable roles in normal kidney function. In kidney proximal tubules, estrogen takes part in the regulation of phosphorus homeostasis *via* its receptors. Estrogen and modulation of ERs have been shown to exert anti-oxidative stress and anti-fibrosis effects in CKD models. On the other hand, studies have revealed the involvement of estrogen and ERs signaling pathways in some autoimmune kidney disorders such as LN.

In light of these findings, the altered or dysregulated estrogen/ERs signaling pathways contribute to a variety of diseases (Table 1). Moreover, given the fact that sex dimorphism exists in certain renal disease entities revealed by epidemiological studies, the roles of estrogen and ERs cannot be overemphasized in the pathogenesis and prognosis of kidney diseases. Indeed, clinical and experimental studies have shown that the level of estrogen or ER α gene polymorphism influences the susceptibilities or dictates the outcomes of several renal diseases. Based on the evidence provided in this review, we propose the concept that targeting estrogen/ERs signaling pathways, either by agonizing or by antagonizing, might works in patients with certain kidney diseases.

However, many gaps exist in knowledge regarding the roles of estrogen and ERs in distinct kidney diseases and further research is necessary to address these areas. The gender differences in the setting of AKI does not necessarily mean this is the case in every single cause for AKI. Likewise, the functional role of estrogen or ERs

Kidney diseases	Animal/cell models	Key estrogen/ER signaling pathways	Potential drugs
ΑKI	IRI AKI after cardiac arrest and	Estrogen-renal sympathetic nervous system (↓)-regional noradrenaline	Raloxifene Tamoxifen
	cardiopulmonary resuscitation	(1)	
	Sepsis-associated AKI	Estrogen-PPARγ (+)	
	Mercury/cisplatin/gentamicin-	Estrogen-ERβ/GPER-ET1 (↓)	
	induced AKI	Estrogen-PI3K/Akt (+)-eNOS	
	Methotrexate-induced tubular	phosphorylation (↑)	
	epithelial cell injury	Estrogen-NMDAR (—)	
		GPER-NO	
		Estrogen-Na+/K+-ATPase	
		Estrogen-OAT2 (↑)	
DN	STZ-induced DN	Estrogen-lipid peroxidation/	Raloxifene
	db/db mice	oxidative stress (↓)	Tamoxifen
	Otsuka-Long-Evans-Tokushima-Fatty	Estrogen-MMP2/9 (+)-ECM (\downarrow)	Icariin
	rats	Estrogen-TGF β 1 (—)-FN/AP-1 (—)	
	High glucose-induced mesangial	Estrogen-ERK (+)	
	cell/podocyte injuries	ER β -JAK2/STAT3 (+)	
		GPER1-TGF β /Smad (—)-ERK1/2 (—)	
LN	MRL/Ipr mice	Estrogen-autoreactive B cells (↑)/T	Fulvestrant
	NZB/WF1 mice	cell apoptosis (↓)	
	Chronic GVHD-induced LN	Estrogen/ERa-BAFF (+)	
		ERα-TLR (+)	
		Estrogen-IFN dependent/	
		independent pathways	
		Estrogen/ERa-TWEAK	
IgAN	VT-induced IgAN	ERα-MCs proliferation	
ĊĸD	Aging DSS rats	Estrogen-renal NO (↑)-oxidative	Tamoxifen
	SHRSP rats with uninephrectomy	stress (↓)	
	UUO-induced kidney fibrosis	Estrogen-TGFβ/α-SMA (—)	
	Hypertensive nephrosclerosis	ERα-TGFβ/Smad	
	induced by inhibition of NO	ERα/β-cystic lesions/inflammation/	
	synthesis	fibrosis	
	Adenine-induced CKD	Estrogen-ERa/GPER-kidney	
		regeneration and growth	
CKD-MBD	5/6 nephrectomy	Estrogen-ER α /ER β -NaPi-IIa (+)	Bazedoxifene
	Male Cy/+ rats with autosomal	Estrogen-cAMP dependent PKA	Raloxifene
	dominant cystic kidney disease	(–)-calcium/PKC (–)	
	PTH-stimulated osteoclast-like cell	Estrogen/ERa-ERa-GAS6	
	formation	(+)-LKB1 (↑)	
	Phosphate/Angiotensin II-induced		
	VSMC proliferation		
Uremic coagulopathy	Uremic rats	Impaired L-arginine-NO signaling $ ightarrow$	Conjugated estrogens
		estrogen $(\downarrow) \rightarrow$ platelet	
		aggregation/adhesion (-)	
		Estrogen-protein S/antithrombin (1)	

 Table 1. Summary of key estrogen/ER signaling pathways and potential drugs in kidney diseases.

AKI: acute kidney injury; IRI: ischemia-reperfusion injury; PPAR: peroxisome proliferator-activated receptor; ET1: endothelin-1; eNOS: endothelial nitric oxide synthase; NMDAR: N-methyl-D-aspartate receptors; NO: nitric oxide; OAT2: organic anion transporter 2; DN: diabetic nephropathy; STZ: streptozotocin; MMP: matrix metalloproteinase; ECM: extracellular matrix; TGF β : transforming growth factor β ; FN: fibronectin; AP-1: activator protein-1; ERK: extracellular signal-regulated protein kinase; JAK2/STAT3: Janus kinase 2/signal transducer and activator of transcription 3; LN; lupus nephritis; GVHD: graft-versus-host disease; BAFF: B cell activating factor; TLR: toll-like receptor; IFN: interferon; TWEAK: tumor necrosis factor-like weak inducer of apoptosis; IgAN: IgA nephropathy; VT: trichothecene vomitoxin; MCs: mesangial cells; CKD: chronic kidney disease; DSS: Dahl salt-sensitive; SHRSP: spontaneously hypertensive stroke-prone; UUO: unilateral ureteral obstruction; α -SMA: α -smooth muscle actin; CKD-MBD: chronic kidney disease-mineral and bone disorder; PTH: parathyroid hormone; VSMC: vascular smooth muscle cell; cAMP: cyclic adenosine monophosphate; PKA: protein kinase A; PKC: protein kinase C; GAS6: growth arrest-specific gene 6; LKB1: liver kinase B1.

confined to one disease entity cannot readily be replicated in another and thus should be analyzed separately. Besides, evidence regarding the exact roles of estrogen and ERs is mixed, particularly in some autoimmune kidney disorders. Although modulation of ERs or supplementation of estrogen has renoprotective effects in several experimental models, there is still a long way to go before it can be applied to clinical trials. The benefits of SERMs have been witnessed in patients with CKD, however, adverse events and long-term outcomes should be assessed.

The therapeutic potential of targeting estrogen/ERs signaling pathways remains to be tested. Moreover, by no means does targeting these mechanisms fully tackle renal disorders, considering the complex signaling networks involved. Further experimental and clinical studies are warranted to comprehensively understand the roles of estrogen and ERs in different kidney diseases.

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References

- [1] Barros RP, Gustafsson J. Estrogen receptors and the metabolic network. Cell Metab. 2011;14(3):289–299.
- [2] Kitajima Y, Ono Y. Estrogens maintain skeletal muscle and satellite cell functions. J Endocrinol. 2016;229(3): 267–275.
- [3] Arnal J-F, Lenfant F, Metivier R, et al. Membrane and nuclear estrogen receptor alpha actions: from tissue specificity to medical implications. Physiol Rev. 2017; 97(3):1045–1087.
- [4] El-Gendy AA, Elsaed WM, Abdallah HI. Potential role of estradiol in ovariectomy-induced derangement of renal endocrine functions. Ren Fail. 2019;41(1): 507–520.
- [5] Dogan E, Erkoc R, Demir C, et al. Effect of hormone replacement therapy on CD4+ and CD8+ numbers, CD4+/CD8+ ratio, and immunoglobulin levels in hemodialysis patients. LRNF. 2005;27(4):421–424.
- [6] Jacenik D, Beswick EJ, Krajewska WM, et al. G protein-coupled estrogen receptor in colon function, immune regulation and carcinogenesis. World J Gastroenterol. 2019;25(30):4092–4104.
- [7] Hsu LH, Chu NM, Kao SH. Estrogen, estrogen receptor and lung cancer. Int J Mol Sci. 2017;18(8):1713.
- [8] Shang DP, Lian HY, Fu DP, et al. Relationship between estrogen receptor 1 gene polymorphisms and postmenopausal osteoporosis of the spine in Chinese women. Genet Mol Res. 2016;15(2):gmr8106.
- [9] Yang J, Han R, Chen M, et al. Associations of estrogen receptor alpha gene polymorphisms with Type 2 diabetes mellitus and metabolic syndrome: a systematic review and meta-analysis. Horm Metab Res. 2018;50(06):469–477.
- [10] Tang Y, Min Z, Xiang X-J, et al. Estrogen-related receptor alpha is involved in Alzheimer's disease-like pathology. Exp Neurol. 2018;305:89–96.
- [11] Muka T, Vargas KG, Jaspers L, et al. Estrogen receptor β actions in the female cardiovascular system: a systematic review of animal and human studies. Maturitas. 2016;86:28–43.
- [12] Corradetti C, Jog NR, Cesaroni M, et al. Estrogen receptor α signaling exacerbates immune-mediated nephropathies through alteration of metabolic activity. J Immunol. 2018;200(2):512–522.
- [13] Kline J, Rachoin JS. Acute kidney injury and chronic kidney disease: it's a two-way street. Ren Fail. 2013; 35(4):452–455.
- [14] Chen JQ, Delannoy M, Cooke C, et al. Mitochondrial localization of ERalpha and ERbeta in human MCF7

cells. Am J Physiol Endocrinol Metab. 2004;286(6): E1011–E1022.

- [15] Razzaque MA, Masuda N, Maeda Y, et al. Estrogen receptor-related receptor gamma has an exceptionally broad specificity of DNA sequence recognition. Gene. 2004;340(2):275–282.
- [16] Guillaume M, Montagner A, Fontaine C, et al. Nuclear and membrane actions of estrogen receptor alpha: contribution to the regulation of energy and glucose homeostasis. Adv Exp Med Biol. 2017;1043:401–426.
- [17] Kim K, Thu N, Saville B, et al. Domains of estrogen receptor alpha (ERalpha) required for ERalpha/Sp1mediated activation of GC-rich promoters by estrogens and antiestrogens in breast cancer cells. Mol Endocrinol. 2003;17(5):804–817.
- [18] Sentis S, Le Romancer M, Bianchin C, et al. Sumoylation of the estrogen receptor alpha hinge region regulates its transcriptional activity. Mol Endocrinol. 2005;19(11):2671–2684.
- [19] Brzozowski AM, Pike AC, Dauter Z, et al. Molecular basis of agonism and antagonism in the oestrogen receptor. Nature. 1997;389(6652):753–758.
- [20] Moras D, Gronemeyer H. The nuclear receptor ligand-binding domain: structure and function. Curr Opin Cell Biol. 1998;10(3):384–391.
- [21] Mosselman S, Polman J, Dijkema R. ER beta: identification and characterization of a novel human estrogen receptor. FEBS Lett. 1996;392(1):49–53.
- [22] Fuentes N, Silveyra P. Estrogen receptor signaling mechanisms. Adv Protein Chem Struct Biol. 2019;116: 135–170.
- [23] Hamilton KJ, Hewitt SC, Arao Y, et al. Estrogen hormone biology. Curr Top Dev Biol. 2017;125:109–146.
- [24] Arao Y, Coons LA, Zuercher WJ, et al. Transactivation function-2 of estrogen receptor α contains transactivation function-1-regulating element. J Biol Chem. 2015;290(28):17611–17627.
- [25] Arao Y, Hamilton KJ, Goulding EH, et al. Transactivating function (AF) 2-mediated AF-1 activity of estrogen receptor α is crucial to maintain male reproductive tract function. Proc Natl Acad Sci USA. 2012;109(51):21140–21145.
- [26] Gosden JR, Middleton PG, Rout D. Localization of the human oestrogen receptor gene to chromosome 6q24–q27 by in situ hybridization. Cytogenet Cell Genet. 1986;43(3–4):218–220.
- [27] Paterni I, Granchi C, Katzenellenbogen JA, et al. Estrogen receptors alpha (ER α) and beta (ER β): sub-type-selective ligands and clinical potential. Steroids. 2014;90:13–29.
- [28] Chen C, Gong X, Yang X, et al. The roles of estrogen and estrogen receptors in gastrointestinal disease. Oncol Lett. 2019;18(6):5673–5680.
- [29] Wang Z, Zhang X, Shen P, et al. Identification, cloning, and expression of human estrogen receptoralpha36, a novel variant of human estrogen receptor-alpha66. Biochem Biophys Res Commun. 2005; 336(4):1023–1027.
- [30] Chantalat E, Boudou F, Laurell H, et al. The AF-1-deficient estrogen receptor ERα46 isoform is frequently expressed in human breast tumors. Breast Cancer Res. 2016;18(1):123.

- [31] Smith EP, Boyd J, Frank GR, et al. Estrogen resistance caused by a mutation in the estrogen-receptor gene in a man. N Engl J Med. 1994;331(16):1056–1061.
- [32] Riant E, Waget A, Cogo H, et al. Estrogens protect against high-fat diet-induced insulin resistance and glucose intolerance in mice. Endocrinology. 2009; 150(5):2109–2117.
- [33] Yu P, Wang Y, Li C, et al. Protective effects of downregulating estrogen receptor alpha expression in cervical cancer. Anticancer Agents Med Chem. 2018; 18(14):1975–1982.
- [34] Pelekanou V, Anastasiou E, Bakogeorgou E, et al. Estrogen receptor-alpha isoforms are the main estrogen receptors expressed in non-small cell lung carcinoma. Steroids. 2019;142:65–76.
- [35] Stein RA, Gaillard S, McDonnell DP. Estrogen-related receptor alpha induces the expression of vascular endothelial growth factor in breast cancer cells. J Steroid Biochem Mol Biol. 2009;114(1–2):106–112.
- [36] Mishra S, Tai Q, Gu X, et al. Estrogen and estrogen receptor alpha promotes malignancy and osteoblastic tumorigenesis in prostate cancer. Oncotarget. 2015;6(42):44388–44402.
- [37] Enmark E, Pelto-Huikko M, Grandien K, et al. Human estrogen receptor beta-gene structure, chromosomal localization, and expression pattern. J Clin Endocrinol Metab. 1997;82(12):4258–4265.
- [38] Liu J, Sareddy GR, Zhou M, et al. Differential effects of estrogen receptor β isoforms on glioblastoma progression. Cancer Res. 2018;78(12):3176–3189.
- [39] Leung Y-K, Mak P, Hassan S, et al. Estrogen receptor (ER)-beta isoforms: a key to understanding ER-beta signaling. Proc Natl Acad Sci USA. 2006;103(35): 13162–13167.
- [40] Fitts JM, Klein RM, Powers CA. Tamoxifen regulation of bone growth and endocrine function in the ovariectomized rat: discrimination of responses involving estrogen receptor α /estrogen receptor β , G proteincoupled estrogen receptor, or estrogen-related receptor γ using fulvestrant (ICI 182780). J Pharmacol Exp Ther. 2011;338(1):246–254.
- [41] Younes M, Honma N. Estrogen receptor β. Arch Pathol Lab Med. 2011;135(1):63–66.
- [42] Ponnusamy S, Tran QT, Harvey I, et al. Pharmacologic activation of estrogen receptor β increases mitochondrial function, energy expenditure, and brown adipose tissue. Faseb J. 2017;31(1): 266–281.
- [43] Varshney MK, Inzunza J, Lupu D, et al. Role of estrogen receptor beta in neural differentiation of mouse embryonic stem cells. Proc Natl Acad Sci USA. 2017; 114(48): E10428–E10437.
- [44] Efstathiadou ZA, Sakka C, Polyzos SA, et al. Associations of estrogen receptor alpha and Beta gene polymorphisms with lipid levels and insulin resistance in men. Metabolism. 2015;64(5):611–617.
- [45] Edvardsson K, Ström A, Jonsson P, et al. Estrogen receptor β induces antiinflammatory and antitumorigenic networks in colon cancer cells. Mol Endocrinol. 2011;25(6):969–979.

- [46] Liu J, Viswanadhapalli S, Garcia L, et al. Therapeutic utility of natural estrogen receptor beta agonists on ovarian cancer. Oncotarget. 2017;8(30):50002–50014.
- [47] Yu C-P, Ho J-Y, Huang Y-T, et al. Estrogen inhibits renal cell carcinoma cell progression through estrogen receptor- β activation. PLOS One. 2013;8(2): e56667.
- [48] Xiao L, Luo Y, Tai R, et al. Estrogen receptor β suppresses inflammation and the progression of prostate cancer. Mol Med Rep. 2019;19(5):3555–3563.
- [49] Song P, Li Y, Dong Y, et al. Estrogen receptor β inhibits breast cancer cells migration and invasion through CLDN6-mediated autophagy. J Exp Clin Cancer Res. 2019;38(1):354.
- [50] Gustafsson JA, Strom A, Warner M. Update on ERbeta. J Steroid Biochem Mol Biol. 2019;191:105312.
- [51] Prossnitz ER, Barton M. Estrogen biology: new insights into GPER function and clinical opportunities. Mol Cell Endocrinol. 2014;389(1–2):71–83.
- [52] Sharma G, Mauvais-Jarvis F, Prossnitz ER. Roles of G protein-coupled estrogen receptor GPER in metabolic regulation. J Steroid Biochem Mol Biol. 2018;176: 31–37.
- [53] Barton M, Filardo EJ, Lolait SJ, et al. Twenty years of the G protein-coupled estrogen receptor GPER: historical and personal perspectives. J Steroid Biochem Mol Biol. 2018;176:4–15.
- [54] Gaudet HM, Cheng SB, Christensen EM, et al. The Gprotein coupled estrogen receptor, GPER: the inside and inside-out story. Mol Cell Endocrinol. 2015;418 Pt 3(Pt 3):207–219.
- [55] Olde B, Leeb-Lundberg LM. GPR30/GPER1: searching for a role in estrogen physiology. Trends Endocrinol Metab. 2009;20(8):409–416.
- [56] Zimmerman MA, Budish RA, Kashyap S, et al. GPERnovel membrane oestrogen receptor. Clin Sci. 2016; 130(12):1005–1016.
- [57] Meyer MR, Prossnitz ER, Barton M. GPER/GPR30 and regulation of vascular tone and blood pressure. Immunol Endocr Metab Agents Med Chem. 2011; 11(4):255–261.
- [58] Krejčířová R, Maňasová M, Sommerová V, et al. G protein-coupled estrogen receptor (GPER) in adult boar testes, epididymis and spermatozoa during epididymal maturation. Int J Biol Macromol. 2018;116: 113–119.
- [59] Sharma G, Hu C, Brigman JL, et al. GPER deficiency in male mice results in insulin resistance, dyslipidemia, and a proinflammatory state. Endocrinology. 2013;154(11):4136–4145.
- [60] Torres-López L, Maycotte P, Liñán-Rico A, et al. Tamoxifen induces toxicity, causes autophagy, and partially reverses dexamethasone resistance in Jurkat T cells. J Leukoc Biol. 2019;105(5):983–998.
- [61] Haas E, Bhattacharya I, Brailoiu E, et al. Regulatory role of G protein-coupled estrogen receptor for vascular function and obesity. Circ Res. 2009;104(3): 288–291.
- [62] Kumar R, Balhuizen A, Amisten S, et al. Insulinotropic and antidiabetic effects of 17β-estradiol and the GPR30 agonist G-1 on human pancreatic islets. Endocrinology. 2011;152(7):2568–2579.

- [63] Balhuizen A, Kumar R, Amisten S, et al. Activation of G protein-coupled receptor 30 modulates hormone secretion and counteracts cytokine-induced apoptosis in pancreatic islets of female mice. Mol Cell Endocrinol. 2010;320(1–2):16–24.
- [64] Feldman RD, Limbird LE. GPER (GPR30): a nongenomic receptor (GPCR) for steroid hormones with implications for cardiovascular disease and cancer. Annu Rev Pharmacol Toxicol. 2017;57:567–584.
- [65] Vrtačnik P, Ostanek B, Mencej-Bedrač S, et al. The many faces of estrogen signaling. Biochem Med. 2014;24(3):329–342.
- [66] Wu Q, Chambliss K, Umetani M, et al. Non-nuclear estrogen receptor signaling in the endothelium. J Biol Chem. 2011;286(17):14737–14743.
- [67] Moriarty K, Kim KH, Bender JR. Minireview: estrogen receptor-mediated rapid signaling. Endocrinology. 2006;147(12):5557–5563.
- [68] Alexander A, Irving AJ, Harvey J. Emerging roles for the novel estrogen-sensing receptor GPER1 in the CNS. Neuropharmacology. 2017;113(Pt B):652–660.
- [69] Björnström L, Sjöberg M. Mechanisms of estrogen receptor signaling: convergence of genomic and nongenomic actions on target genes. Mol Endocrinol. 2005;19(4):833–842.
- [70] Deroo BJ, Korach KS. Estrogen receptors and human disease. J Clin Invest. 2006;116(3):561–570.
- [71] Nilsson S, Koehler KF. Oestrogen receptors and selective oestrogen receptor modulators: molecular and cellular pharmacology. Basic Clin Pharmacol Toxicol. 2005;96(1):15–25.
- [72] Jordan VC. Tamoxifen: catalyst for the change to targeted therapy. Eur J Cancer. 2008;44(1):30–38.
- [73] Moen MD, Keating GM. Raloxifene: a review of its use in the prevention of invasive breast cancer. Drugs. 2008;68(14):2059–2083.
- [74] Wardell SE, Nelson ER, Chao CA, et al. Bazedoxifene exhibits antiestrogenic activity in animal models of tamoxifen-resistant breast cancer: implications for treatment of advanced disease. Clin Cancer Res. 2013;19(9):2420–2431.
- [75] Negi S, Koreeda D, Kobayashi S, et al. Acute kidney injury: epidemiology, outcomes, complications, and therapeutic strategies. Semin Dial. 2018;31(5): 519–527.
- [76] Fujii T, Uchino S, Takinami M, et al. Subacute kidney injury in hospitalized patients. Clin J Am Soc Nephrol. 2014;9(3):457–461.
- [77] Kellum JA, Lameire N, KDIGO AKI Guideline Work Group. Diagnosis, evaluation, and management of acute kidney injury: a KDIGO summary (Part 1). Crit Care. 2013;17(1):204.
- [78] Koza Y. Acute kidney injury: current concepts and new insights. J Inj Violence Res. 2016;8(1):58–62.
- [79] Neugarten J, Golestaneh L. Female sex reduces the risk of hospital-associated acute kidney injury: a meta-analysis. BMC Nephrol. 2018;19(1):314.
- [80] Neugarten J, Sandilya S, Singh B, et al. Sex and the risk of AKI following cardio-thoracic surgery: a metaanalysis. CJASN. 2016;11(12):2113–2122.
- [81] Diptyanusa A, Phumratanaprapin W, Phonrat B, et al. Characteristics and associated factors of acute kidney

injury among adult dengue patients: a retrospective single-center study. PLOS One. 2019;14(1):e0210360.

- [82] O'Brien Z, Cass A, Cole L, et al. Sex and mortality in septic severe acute kidney injury. J Crit Care. 2019; 49:70–76.
- [83] Vallabhajosyula S, Ya'Qoub L, Dunlay SM, et al. Sex disparities in acute kidney injury complicating acute myocardial infarction with cardiogenic shock. ESC Heart Fail. 2019;6(4):874–877.
- [84] Cardinale DA, Larsen FJ, Schiffer TA, et al. Superior intrinsic mitochondrial respiration in women than in men. Front Physiol. 2018;9:1133.
- [85] Borrás C, Sastre J, García-Sala D, et al. Mitochondria from females exhibit higher antioxidant gene expression and lower oxidative damage than males. Free Radic Biol Med. 2003;34(5):546–552.
- [86] Bhargava P, Schnellmann RG. Mitochondrial energetics in the kidney. Nat Rev Nephrol. 2017;13(10): 629–646.
- [87] Reed DK, Arany I. p66shc and gender-specific dimorphism in acute renal injury. In Vivo. 2014;28(2): 205–208.
- [88] Kang KP, Lee JE, Lee AS, et al. Effect of gender differences on the regulation of renal ischemia-reperfusion-induced inflammation in mice. Mol Med Rep. 2014;9(6):2061–2068.
- [89] Tanaka R, Yazawa M, Morikawa Y, et al. Sex differences in ischaemia/reperfusion-induced acute kidney injury depends on the degradation of noradrenaline by monoamine oxidase. Clin Exp Pharmacol Physiol. 2017;44(3):371–377.
- [90] Hodeify R, Megyesi J, Tarcsafalvi A, et al. Gender differences control the susceptibility to ER stressinduced acute kidney injury. Am J Physiol Renal Physiol. 2013;304(7):F875–F882.
- [91] Woodman AG, Mah R, Keddie D, et al. Prenatal iron deficiency causes sex-dependent mitochondrial dysfunction and oxidative stress in fetal rat kidneys and liver. Faseb J. 2018;32(6):3254–3263.
- [92] Morigi M, Perico L, Benigni A. Sirtuins in renal health and disease. J Am Soc Nephrol. 2018;29(7):1799–1809.
- [93] Xu S, Gao Y, Zhang Q, et al. SIRT1/3 activation by resveratrol attenuates acute kidney injury in a septic rat model. Oxid Med Cell Longev. 2016;2016:1–12.
- [94] Ouyang J, Zeng Z, Fang H, et al. SIRT3 inactivation promotes acute kidney injury through elevated acetylation of SOD2 and p53. J Surg Res. 2019;233: 221–230.
- [95] Ugur S, Ulu R, Dogukan A, et al. The renoprotective effect of curcumin in cisplatin-induced nephrotoxicity. Ren Fail. 2015;37(2):332–336.
- [96] Wei S, Gao Y, Dai X, et al. SIRT1-mediated HMGB1 deacetylation suppresses sepsis-associated acute kidney injury. Am J Physiol Renal Physiol. 2019;316(1): F20–F31.
- [97] Gao Q, Zhu H. The overexpression of Sirtuin1 (SIRT1) alleviated lipopolysaccharide (LPS)-induced Acute Kidney Injury (AKI) via inhibiting the activation of nucleotide-binding oligomerization domain-like receptors (NLR) family pyrin domain containing 3

(NLRP3) inflammasome. Med Sci Monit. 2019;25: 2718–2726.

- [98] Khan M, Ullah R, Rehman SU, et al. 17β-estradiol modulates SIRT1 and halts oxidative stress-mediated cognitive impairment in a male aging mouse model. Cells. 2019;8(8):928.
- [99] Guo J-M, Shu H, Wang L, et al. SIRT1-dependent AMPK pathway in the protection of estrogen against ischemic brain injury. CNS Neurosci Ther. 2017;23(4): 360–369.
- [100] Yao Y, Li H, Gu Y, et al. Inhibition of SIRT1 deacetylase suppresses estrogen receptor signaling. Carcinogenesis. 2010;31(3):382–387.
- [101] Malek M, Nematbakhsh M. Renal ischemia/reperfusion injury; from pathophysiology to treatment. J Renal Inj Prev. 2015;4(2):20–27.
- [102] Fu Y, Tang C, Cai J, et al. Rodent models of AKI-CKD transition. Am J Physiol Renal Physiol. 2018;315(4): F1098–F1106.
- [103] Packialakshmi B, Stewart IJ, Burmeister DM, et al. Large animal models for translational research in acute kidney injury. Ren Fail. 2020;42(1):1042–1058.
- [104] Pegues MA, McCrory MA, Zarjou A, et al. C-reactive protein exacerbates renal ischemia-reperfusion injury. Am J Physiol Renal Physiol. 2013;304(11): F1358–F1365.
- [105] Philipponnet C, Aniort J, Garrouste C, et al. Ischemia reperfusion injury in kidney transplantation: a case report. Medicine. 2018;97(52):e13650.
- [106] Bellomo R, Kellum JA, Ronco C, et al. Acute kidney injury in sepsis. Intensive Care Med. 2017;43(6): 816–828.
- [107] Sandroni C, Dell'anna AM, Tujjar O, et al. Acute kidney injury after cardiac arrest: a systematic review and meta-analysis of clinical studies. Minerva Anestesiol. 2016;82(9):989–999.
- [108] Fu Z-Y, Wu Z-J, Zheng J-H, et al. The incidence of acute kidney injury following cardiac arrest and cardiopulmonary resuscitation in a rat model. Ren Fail. 2019;41(1):278–283.
- [109] Redfield RR, Scalea JR, Zens TJ, et al. Predictors and outcomes of delayed graft function after living-donor kidney transplantation. Transpl Int. 2016;29(1):81–87.
- [110] Damodaran S, Bullock B, Ekwenna O, et al. Risk factors for delayed graft function and their impact on graft outcomes in live donor kidney transplantation. Int Urol Nephrol. 2021;53(3):439–446.
- [111] Lepeytre F, Dahhou M, Zhang X, et al. Association of sex with risk of kidney graft failure differs by age. J Am Soc Nephrol. 2017;28(10):3014–3023.
- [112] Zeier M, Döhler B, Opelz G, et al. The effect of donor gender on graft survival. J Am Soc Nephrol. 2002; 13(10):2570–2576.
- [113] Aufhauser DD, Wang Z, Murken DR Jr, et al. Improved renal ischemia tolerance in females influences kidney transplantation outcomes. J Clin Invest. 2016;126(5):1968–1977.
- [114] Tanaka R, Tsutsui H, Ohkita M, et al. Sex differences in ischemia/reperfusion-induced acute kidney injury are dependent on the renal sympathetic nervous system. Eur J Pharmacol. 2013;714(1–3):397–404.

- [115] Tanaka R, Tsutsui H, Kobuchi S, et al. Protective effect of 17β-estradiol on ischemic acute kidney injury through the renal sympathetic nervous system. Eur J Pharmacol. 2012;683(1–3):270–275.
- [116] Ikeda M, Swide T, Vayl A, et al. Estrogen administered after cardiac arrest and cardiopulmonary resuscitation ameliorates acute kidney injury in a sex- and age-specific manner. Crit Care. 2015;19(1):332.
- [117] Hutchens MP, Nakano T, Kosaka Y, et al. Estrogen is renoprotective via a nonreceptor-dependent mechanism after cardiac arrest in vivo. Anesthesiology. 2010;112(2):395–405.
- [118] Park KM, Kim JI, Ahn Y, et al. Testosterone is responsible for enhanced susceptibility of males to ischemic renal injury. J Biol Chem. 2004;279(50):52282–52292.
- [119] Singh AP, Singh N, Pathak D, et al. Estradiol attenuates ischemia reperfusion-induced acute kidney injury through PPAR-γ stimulated eNOS activation in rats. Mol Cell Biochem. 2019;453(1–2):1–9.
- [120] Singh AP, Singh N, Singh Bedi PM. Estrogen attenuates renal IRI through PPAR-γ agonism in rats. J Surg Res. 2016;203(2):324–330.
- [121] Liu B, Tan P. PPAR γ/TLR4/TGF-β1 axis mediates the protection effect of erythropoietin on cyclosporin Ainduced chronic nephropathy in rat. Ren Fail. 2020; 42(1):216–224.
- [122] Žeravica R, Čabarkapa V, Ilinčić B, et al. Plasma endothelin-1 level, measured glomerular filtration rate and effective renal plasma flow in diabetic nephropathy. Ren Fail. 2015;37(4):681–686.
- [123] Zager RA, Johnson ACM, Andress D, et al. Progressive endothelin-1 gene activation initiates chronic/end-stage renal disease following experimental ischemic/reperfusion injury. Kidney Int. 2013;84(4): 703–712.
- [124] Arfian N, Emoto N, Vignon-Zellweger N, et al. ET-1 deletion from endothelial cells protects the kidney during the extension phase of ischemia/reperfusion injury. Biochem Biophys Res Commun. 2012;425(2): 443–449.
- [125] Takaoka M, Yuba M, Fujii T, et al. Oestrogen protects against ischaemic acute renal failure in rats by suppressing renal endothelin-1 overproduction. Clin Sci (Lond). 2002;103 Suppl 48(Suppl 48):434S–437S.
- [126] Ba ZF, Chaudry IH. Role of estrogen receptor subtypes in estrogen-induced organ-specific vasorelaxation after trauma-hemorrhage. Am J Physiol Heart Circ Physiol. 2008;295(5):H2061–H2067.
- [127] Gohar EY, Daugherty EM, Aceves JO, et al. Evidence for G-protein-coupled estrogen receptor as a pronatriuretic factor. J Am Heart Assoc. 2020;9(10): e015110.
- [128] Wu C-C, Chang C-Y, Chang S-T, et al. 17β-Estradiol accelerated renal tubule regeneration in male rats after ischemia/reperfusion-induced acute kidney injury. Shock. 2016;46(2):158–163.
- [129] Satake A, Takaoka M, Nishikawa M, et al. Protective effect of 17beta-estradiol on ischemic acute renal failure through the PI3K/Akt/eNOS pathway. Kidney Int. 2008;73(3):308–317.
- [130] Singh AP, Singh N, Bedi PMS. Estradiol mitigates ischemia reperfusion-induced acute renal failure

through NMDA receptor antagonism in rats. Mol Cell Biochem. 2017;434(1–2):33–40.

- [131] Hutchens MP, Fujiyoshi T, Komers R, et al. Estrogen protects renal endothelial barrier function from ischemia-reperfusion in vitro and in vivo. Am J Physiol Renal Physiol. 2012;303(3):F377–F385.
- [132] Chang Y, Han Z, Zhang Y, et al. G protein-coupled estrogen receptor activation improves contractile and diastolic functions in rat renal interlobular artery to protect against renal ischemia reperfusion injury. Biomed Pharmacother = Biomed Pharmacother. 2019; 112:108666.
- [133] Jørgensen PL. Structure, function and regulation of Na,K-ATPase in the kidney. Kidney Int. 1986;29(1): 10–20.
- [134] Zhang L-M, Jiang L-J, Zhao Z-G, et al. Mesenteric lymph duct ligation after hemorrhagic shock enhances the ATP level and ATPase activity in rat kidneys. Ren Fail. 2014;36(4):593–597.
- [135] Molinas SM, Trumper L, Serra E, et al. Evolution of renal function and Na+, K +-ATPase expression during ischaemia-reperfusion injury in rat kidney. Mol Cell Biochem. 2006;287(1–2):33–42.
- [136] Kumaş M, Eşrefoğlu M, Karataş E, et al. Investigation of dose-dependent effects of berberine against renal ischemia/reperfusion injury in experimental diabetic rats. Nefrologia. 2019;39(4):411–423.
- [137] Fekete A, Vannay A, Vér A, et al. Sex differences in the alterations of Na(+), K(+)-ATPase following ischaemia-reperfusion injury in the rat kidney. J Physiol. 2004;555(Pt 2):471–480.
- [138] Gracelli JB, Souza-Menezes J, Barbosa CML, et al. Role of estrogen and progesterone in the modulation of CNG-A1 and Na/K+-ATPase expression in the renal cortex. Cell Physiol Biochem. 2012;30(1): 160–172.
- [139] Buléon M, Cuny M, Grellier J, et al. A single dose of estrogen during hemorrhagic shock protects against kidney injury whereas estrogen restoration in ovariectomized mice is ineffective. Sci Rep. 2020;10(1): 17240.
- [140] Sun J, Zhang J, Tian J, et al. Mitochondria in sepsisinduced AKI. J Am Soc Nephrol. 2019;30(7): 1151–1161.
- [141] Vincent J-L, Sakr Y, Sprung CL, et al. Sepsis in European intensive care units: results of the SOAP study. Crit Care Med. 2006;34(2):344–353.
- [142] Xu X, Nie S, Liu Z, et al. Epidemiology and clinical correlates of AKI in Chinese hospitalized adults. Clin J Am Soc Nephrol. 2015;10(9):1510–1518.
- [143] Feng J-Y, Liu K-T, Abraham E, et al. Serum estradiol levels predict survival and acute kidney injury in patients with septic shock-a prospective study. PLoS One. 2014;9(6):e97967.
- [144] Trentzsch H, Nienaber U, Behnke M, et al. Female sex protects from organ failure and sepsis after major trauma haemorrhage. Injury. 2014;45(Suppl 3): S20–S28.
- [145] Chung M-T, Lee Y-M, Shen H-H, et al. Activation of autophagy is involved in the protective effect of 17β-oestradiol on endotoxaemia-induced multiple

organ dysfunction in ovariectomized rats. J Cell Mol Med. 2017;21(12):3705–3717.

- [146] Shen H-H, Huang S-Y, Cheng P-Y, et al. Involvement of HSP70 and HO-1 in the protective effects of raloxifene on multiple organ dysfunction syndrome by endotoxemia in ovariectomized rats. Menopause. 2017;24(8):959–969.
- [147] Zhong L, Zhou X-L, Liu Y-S, et al. Estrogen receptor α mediates the effects of notoginsenoside R1 on endotoxin-induced inflammatory and apoptotic responses in H9c2 cardiomyocytes. Mol Med Rep. 2015;12(1):119–126.
- [148] Yuk J-M, Kim TS, Kim SY, et al. Orphan nuclear receptor ERRα controls macrophage metabolic signaling and A20 expression to negatively regulate TLRinduced inflammation. Immunity. 2015;43(1):80–91.
- [149] Christaki E, Opal SM, Keith JC, et al. Estrogen receptor beta agonism increases survival in experimentally induced sepsis and ameliorates the genomic sepsis signature: a pharmacogenomic study. J Infect Dis. 2010;201(8):1250–1257.
- [150] Bösch F, Angele MK, Chaudry IH. Gender differences in trauma, shock and sepsis. Mil Med Res. 2018;5(1): 35.
- [151] Kawasaki T, Chaudry IH. The effects of estrogen on various organs: therapeutic approach for sepsis, trauma, and reperfusion injury. Part 2: liver, intestine, spleen, and kidney. J Anesth. 2012;26(6):892–899.
- [152] Srisawat N, Kulvichit W, Mahamitra N, et al. The epidemiology and characteristics of acute kidney injury in the Southeast Asia intensive care unit: a prospective multicentre study. Nephrol Dial Transplant. 2019; 35(10):1729–1738.
- [153] Goswami S, Pahwa N, Vohra R, et al. Clinical spectrum of hospital acquired acute kidney injury: a prospective study from Central India. Saudi J Kidney Dis Transpl. 2018;29(4):946–955.
- [154] Mehta RL, Pascual MT, Soroko S, et al. Spectrum of acute renal failure in the intensive care unit: the PICARD experience. Kidney Int. 2004;66(4):1613–1621.
- [155] Perazella MA. Pharmacology behind common drug nephrotoxicities. Clin J Am Soc Nephrol. 2018;13(12): 1897–1908.
- [156] Perazella MA. Drug-induced acute kidney injury: diverse mechanisms of tubular injury. Curr Opin Crit Care. 2019;25(6):550–557.
- [157] Hosohata K. Role of oxidative stress in drug-induced kidney injury. Int J Mol Sci. 2016;17(11): 1826.
- [158] Muriithi AK, Leung N, Valeri AM, et al. Biopsy-proven acute interstitial nephritis, 1993–2011: a case series. Am J Kidney Dis. 2014;64(4):558–566.
- [159] Muriithi AK, Leung N, Valeri AM, et al. Clinical characteristics, causes and outcomes of acute interstitial nephritis in the elderly. Kidney Int. 2015;87(2): 458–464.
- [160] Sweileh WM. Gender differences in aminoglycoside induced nephrotoxicity: a prospective, hospital-based study. Curr Clin Pharmacol. 2009;4(3):229–232.
- [161] Chen W-Y, Hsiao C-H, Chen Y-C, et al. Cisplatin nephrotoxicity might have a sex difference. An analysis based on women's sex hormone changes. J Cancer. 2017;8(19):3939–3944.

- [162] Neugarten J, Golestaneh L. The effect of gender on aminoglycoside-associated nephrotoxicity. Clin Nephrol. 2016;86(10):183–189.
- [163] Zajjari Y, Montasser D, Sobhi A, et al. Acute interstitial nephritis in the military hospital of Morocco: clinical features and renal outcomes. Saudi J Kidney Dis Transpl. 2019;30(6):1407–1414.
- [164] Joseph S, Nicolson TJ, Hammons G, et al. Expression of drug transporters in human kidney: impact of sex, age, and ethnicity. Biol Sex Differ. 2015;6(1):4.
- [165] Oswald S, Müller J, Neugebauer U, et al. Protein abundance of clinically relevant drug transporters in the human kidneys. Int J Mol Sci. 2019;20(21):5303.
- [166] Li CY, Hosey-Cojocari C, Basit A, et al. Optimized renal transporter quantification by using Aquaporin 1 and Aquaporin 2 as anatomical markers: application in characterizing the ontogeny of renal transporters and its correlation with hepatic transporters in paired human samples. Aaps J. 2019;21(5):88.
- [167] Kwekel JC, Desai VG, Moland CL, et al. Sex differences in kidney gene expression during the life cycle of F344 rats. Biol Sex Differ. 2013;4(1):14.
- [168] Hazelhoff MH, Bulacio RP, Chevalier A, et al. Renal expression of organic anion transporters is modified after mercuric chloride exposure: gender-related differences. Toxicol Lett. 2018;295:390–396.
- [169] Ljubojević M, Balen D, Breljak D, et al. Renal expression of organic anion transporter OAT2 in rats and mice is regulated by sex hormones. Am J Physiol Renal Physiol. 2007;292(1): F361–F372.
- [170] Nematbakhsh M, Ebrahimian S, Tooyserkani M, et al. Gender difference in Cisplatin-induced nephrotoxicity in a rat model: greater intensity of damage in male than female. Nephro Urol Mon. 2013;5(3):818–821.
- [171] Boddu R, Fan C, Rangarajan S, et al. Unique sex- and age-dependent effects in protective pathways in acute kidney injury. Am J Physiol Renal Physiol. 2017;313(3): F740–F755.
- [172] Kurt AH, Bozkus F, Uremis N, et al. The protective role of G protein-coupled estrogen receptor 1 (GPER-1) on methotrexate-induced nephrotoxicity in human renal epithelium cells. Ren Fail. 2016;38(5):686–692.
- [173] Hernández-Esquivel L, Zazueta C, Buelna-Chontal M, et al. Protective behavior of tamoxifen against Hg2+-induced toxicity on kidney mitochondria: in vitro and in vivo experiments. J Steroid Biochem Mol Biol. 2011;127(3–5):345–350.
- [174] Abd El-Lateef SM, El-Sayed E-SM, Mansour AM, et al. The protective role of estrogen and its receptors in gentamicin-induced acute kidney injury in rats. Life Sci. 2019;239: 117082.
- [175] Lee CI, Goodwin A, Wilcken N. Fulvestrant for hormone-sensitive metastatic breast cancer. Cochrane Database Syst Rev. 2017;1(1):CD011093.
- [176] Ghane Shahrbaf F, Assadi F. Drug-induced renal disorders. J Renal Inj Prev. 2015;4(3):57–60.
- [177] Katarey D, Verma S. Drug-induced liver injury. Clin Med (Lond). 2016;16(Suppl 6):s104-s109.
- [178] Danese E, Montagnana M, Favaloro EJ, et al. Druginduced thrombocytopenia: mechanisms and laboratory diagnostics. Semin Thromb Hemost. 2020;46(03): 264–274.

- [179] Kidney Disease: Improving Global Outcomes (KDIGO) CKD Work Group. KDIGO 2012 clinical practice guideline for the evaluation and management of chronic kidney disease. Kidney Int Suppl. 2013;3(1):1–150.
- [180] Chen L, Luo M, Dong C, et al. Pathological spectrum of glomerular disease in patients with renal insufficiency: a single-center study in Northeastern China. Ren Fail. 2019;41(1):473–480.
- [181] Xie Y, Chen X. Epidemiology, major outcomes, risk factors, prevention and management of chronic kidney disease in China. Am J Nephrol. 2008;28(1):1–7.
- [182] Halbesma N, Brantsma AH, Bakker SJL, et al. Gender differences in predictors of the decline of renal function in the general population. Kidney Int. 2008; 74(4):505–512.
- [183] Eriksen BO, Ingebretsen OC. The progression of chronic kidney disease: a 10-year population-based study of the effects of gender and age. Kidney Int. 2006;69(2):375–382.
- [184] Kattah AG, Smith CY, Gazzuola Rocca L, et al. CKD in patients with bilateral oophorectomy. CJASN. 2018; 13(11):1649–1658.
- [185] Hecking M, Bieber BA, Ethier J, et al. Sex-specific differences in hemodialysis prevalence and practices and the male-to-female mortality rate: the Dialysis Outcomes and Practice Patterns Study (DOPPS). PLoS Med. 2014;11(10):e1001750.
- [186] Carrero JJ, Hecking M, Chesnaye NC, et al. Sex and gender disparities in the epidemiology and outcomes of chronic kidney disease. Nat Rev Nephrol. 2018;14(3):151–164.
- [187] Matuszkiewicz-Rowinska J, Skorzewska K, Radowicki S, et al. Endometrial morphology and pituitarygonadal axis dysfunction in women of reproductive age undergoing chronic haemodialysis-a multicentre study. Nephrol Dial Transplant. 2004;19(8):2074–2077.
- [188] Park YJ, Kim JM. Klotho and postmenopausal hormone replacement therapy in women with chronic kidney disease. J Menopausal Med. 2018;24(2):75–80.
- [189] Ji H, Pesce C, Zheng W, et al. Sex differences in renal injury and nitric oxide production in renal wrap hypertension. Am J Physiol Heart Circ Physiol. 2005; 288(1):H43–H47.
- [190] Ji H, Zheng W, Menini S, et al. Female protection in progressive renal disease is associated with estradiol attenuation of superoxide production. Gend Med. 2007;4(1):56–71.
- [191] Flyvbjerg A. The role of the complement system in diabetic nephropathy. Nat Rev Nephrol. 2017;13(5): 311–318.
- [192] Möllsten A, Svensson M, Waernbaum I, et al. Cumulative risk, age at onset, and sex-specific differences for developing end-stage renal disease in young patients with type 1 diabetes: a nationwide population-based cohort study. Diabetes. 2010;59(7): 1803–1808.
- [193] Goñi MJ, Forga L, Ibañez B, et al. Incidence and risk factors involved in the development of nephropathy in patients with Type 1 diabetes mellitus: follow up since onset. Can J Diabetes. 2016;40(3):258–263.
- [194] Costacou T, Orchard TJ. Cumulative kidney complication risk by 50 years of Type 1 diabetes: the effects

of sex, age, and calendar year at onset. Diabetes Care. 2018;41(3):426-433.

- [195] Maric-Bilkan C. Sex differences in diabetic kidney disease. Mayo Clin Proc. 2020;95(3):587–599.
- [196] Maric C, Forsblom C, Thorn L, et al. Association between testosterone, estradiol and sex hormone binding globulin levels in men with type 1 diabetes with nephropathy. Steroids. 2010;75(11):772–778.
- [197] Salonia A, Lanzi R, Scavini M, et al. Sexual function and endocrine profile in fertile women with type 1 diabetes. Diabetes Care. 2006;29(2):312–316.
- [198] Inada A, Inada O, Fujii NL, et al. Adjusting the 17βestradiol-to-androgen ratio ameliorates diabetic nephropathy. J Am Soc Nephrol. 2016;27(10): 3035–3050.
- [199] Mankhey RW, Bhatti F, Maric C. 17beta-Estradiol replacement improves renal function and pathology associated with diabetic nephropathy. Am J Physiol Renal Physiol. 2005;288(2):F399–F405.
- [200] Riazi S, Maric C, Ecelbarger CA. 17-beta Estradiol attenuates streptozotocin-induced diabetes and regulates the expression of renal sodium transporters. Kidney Int. 2006;69(3):471–480.
- [201] Dixon A, Wells CC, Singh S, et al. Renoprotective effects of a selective estrogen receptor modulator, raloxifene, in an animal model of diabetic nephropathy. Am J Nephrol. 2007;27(2):120–128.
- [202] Ulas M, Cay M. 17β-Estradiol and vitamin E modulates oxidative stress-induced kidney toxicity in diabetic ovariectomized rat. Biol Trace Elem Res. 2011; 144(1–3):821–831.
- [203] Tomiyoshi Y, Sakemi T, Aoki S, et al. Different effects of castration and estrogen administration on glomerular injury in spontaneously hyperglycemic Otsuka Long-Evans Tokushima Fatty (OLETF) rats. Nephron. 2002;92(4):860–867.
- [204] Mankhey RW, Wells CC, Bhatti F, et al. 17beta-Estradiol supplementation reduces tubulointerstitial fibrosis by increasing MMP activity in the diabetic kidney. Am J Physiol Regul Integr Comp Physiol. 2007;292(2):R769–R77.
- [205] Dixon A, Maric C. 17beta-Estradiol attenuates diabetic kidney disease by regulating extracellular matrix and transforming growth factor-beta protein expression and signaling. Am J Physiol Renal Physiol. 2007;293(5):F1678–F1690.
- [206] Chin M, Isono M, Isshiki K, et al. Estrogen and raloxifene, a selective estrogen receptor modulator, ameliorate renal damage in db/db mice. Am J Pathol. 2005;166(6):1629–1636.
- [207] Keene KL, Mychaleckyj JC, Smith SG, et al. Comprehensive evaluation of the estrogen receptor alpha gene reveals further evidence for association with type 2 diabetes enriched for nephropathy in an African American population. Hum Genet. 2008; 123(4):333–341.
- [208] Ryba-Stanisławowska M, Rybarczyk-Kapturska K, Brandt A, et al. IVS1 -397T > C estrogen receptor α polymorphism is associated with low-grade systemic inflammatory response in type 1 diabetic girls. Mediators Inflamm. 2014;2014:839585.

- [209] Irsik DL, Romero-Aleshire MJ, Chavez EM, et al. Renoprotective impact of estrogen receptor-α and its splice variants in female mice with type 1 diabetes. Am J Physiol Renal Physiol. 2018;315(3):F512–F520.
- [210] Wang XX, Wang D, Luo Y, et al. FXR/TGR5 dual agonist prevents progression of nephropathy in diabetes and obesity. J Am Soc Nephrol. 2018;29(1):118–137.
- [211] Tung C-W, Hsu Y-C, Shih Y-H, et al. Glomerular mesangial cell and podocyte injuries in diabetic nephropathy. Nephrology (Carlton). 2018;23 (Suppl 4):32–37.
- [212] Catanuto P, Xia X, Pereira-Simon S, et al. Estrogen receptor subtype ratio change protects against podocyte damage. Curr Trends Endocinol. 2017;9: 19–29.
- [213] Potier M, Elliot SJ, Tack I, et al. Expression and regulation of estrogen receptors in mesangial cells: influence on matrix metalloproteinase-9. J Am Soc Nephrol. 2001;12(2):241–251.
- [214] Ren W, Yi H, Bao Y, et al. Oestrogen inhibits PTPRO to prevent the apoptosis of renal podocytes. Exp Ther Med. 2019;17(3):2373–2380.
- [215] Catanuto P, Doublier S, Lupia E, et al. 17 beta-estradiol and tamoxifen upregulate estrogen receptor beta expression and control podocyte signaling pathways in a model of type 2 diabetes. Kidney Int. 2009;75(11):1194–1201.
- [216] Catanuto P, Fornoni A, Pereira-Simon S, et al. In vivo 17β-estradiol treatment contributes to podocyte actin stabilization in female db/db mice. Endocrinology. 2012;153(12):5888–5895.
- [217] Kummer S, Jeruschke S, Wegerich LV, et al. Estrogen receptor alpha expression in podocytes mediates protection against apoptosis in-vitro and in-vivo. PLoS One. 2011;6(11):e27457.
- [218] Doublier S, Lupia E, Catanuto P, et al. Testosterone and 17β-estradiol have opposite effects on podocyte apoptosis that precedes glomerulosclerosis in female estrogen receptor knockout mice. Kidney Int. 2011; 79(4):404–413.
- [219] Qi MY, Chen K, Liu HR, et al. Protective effect of lcariin on the early stage of experimental diabetic nephropathy induced by streptozotocin via modulating transforming growth factor β1 and type IV collagen expression in rats. J Ethnopharmacol. 2011; 138(3):731–736.
- [220] Li Y-C, Ding X-S, Li H-M, et al. Role of G proteincoupled estrogen receptor 1 in modulating transforming growth factor- β stimulated mesangial cell extracellular matrix synthesis and migration. Mol Cell Endocrinol. 2014;391(1–2):50–59.
- [221] Li Y-C, Ding X-S, Li H-M, et al. Icariin attenuates high glucose-induced type IV collagen and fibronectin accumulation in glomerular mesangial cells by inhibiting transforming growth factor-β production and signalling through G protein-coupled oestrogen receptor 1. Clin Exp Pharmacol Physiol. 2013;40(9): 635–643.
- [222] Qiao C, Ye W, Li S, et al. Icariin modulates mitochondrial function and apoptosis in high glucose-induced glomerular podocytes through G protein-coupled

estrogen receptors. Mol Cell Endocrinol. 2018;473: 146–155.

- [223] Clotet-Freixas S, Soler MJ, Palau V, et al. Sex dimorphism in ANGII-mediated crosstalk between ACE2 and ACE in diabetic nephropathy. Lab Invest. 2018;98(9):1237–1249.
- [224] de Alencar Franco Costa D, Todiras M, Campos LA, et al. Sex-dependent differences in renal angiotensinogen as an early marker of diabetic nephropathy. Acta Physiol. 2015;213(3):740–746.
- [225] Almaani S, Meara A, Rovin BH. Update on lupus nephritis. Clin J Am Soc Nephrol. 2017;12(5):825–835.
- [226] Margery-Muir AA, Bundell C, Nelson D, et al. Gender balance in patients with systemic lupus erythematosus. Autoimmun Rev. 2017;16(3):258–268.
- [227] Guéry JC. Why is systemic lupus erythematosus more common in women? Joint Bone Spine. 2019;86(3): 297–299.
- [228] Rojas-Villarraga AJV, Torres-Gonzalez M, Ruiz-Sternberg Á. Safety of hormonal replacement therapy and oral contraceptives in systemic lupus erythematosus: a systematic review and meta-analysis. PLoS One. 2014;9(8):e104303.
- [229] Kovats S. Estrogen receptors regulate innate immune cells and signaling pathways. Cell Immunol. 2015; 294(2):63–69.
- [230] Moulton VR. Sex hormones in acquired immunity and autoimmune disease. Front Immunol. 2018;9: 2279.
- [231] Kim W-U, Min S-Y, Hwang S-H, et al. Effect of oestrogen on T cell apoptosis in patients with systemic lupus erythematosus. Clin Exp Immunol. 2010;161(3): 453–458.
- [232] Rider V, Jones S, Evans M, et al. Estrogen increases CD40 ligand expression in T cells from women with systemic lupus erythematosus. J Rheumatol. 2001; 28(12):2644–2649.
- [233] Grimaldi CM, Jeganathan V, Diamond B. Hormonal regulation of B cell development: 17 beta-estradiol impairs negative selection of high-affinity DNA-reactive B cells at more than one developmental checkpoint. J Immunol. 2006;176(5):2703–2710.
- [234] Grimaldi CM. Sex and systemic lupus erythematosus: the role of the sex hormones estrogen and prolactin on the regulation of autoreactive B cells. Curr Opin Rheumatol. 2006;18(5):456–461.
- [235] Fan H, Dong G, Zhao G, et al. Gender differences of B cell signature in healthy subjects underlie disparities in incidence and course of SLE related to estrogen. J Immunol Res. 2014;2014:814598.
- [236] Fan H, Zhao G, Ren D, et al. Gender differences of B cell signature related to estrogen-induced IFI44L/ BAFF in systemic lupus erythematosus. Immunol Lett. 2017;181:71–78.
- [237] Ramanujam M, Steffgen J, Visvanathan S, et al. Phoenix from the flames: rediscovering the role of the CD40-CD40L pathway in systemic lupus erythematosus and lupus nephritis. Autoimmun Rev. 2020; 19(11):102668.
- [238] Abdou NI, Rider V, Greenwell C, et al. Fulvestrant (Faslodex), an estrogen selective receptor downregulator, in therapy of women with systemic lupus

erythematosus. clinical, serologic, bone density, and T cell activation marker studies: a double-blind placebo-controlled trial. J Rheumatol. 2008;35(5):797.

- [239] Ding Y, He J, Guo J-P, et al. Gender differences are associated with the clinical features of systemic lupus erythematosus. Chin Med J (Engl). 2012; 125(14):2477–2481.
- [240] Faezi ST, Hosseini Almodarresi M, Akbarian M, et al. Clinical and immunological pattern of systemic lupus erythematosus in men in a cohort of 2355 patients. Int J Rheum Dis. 2014;17(4):394–399.
- [241] Alonso MD, Martínez-Vázquez F, Riancho-Zarrabeitia L, et al. Sex differences in patients with systemic lupus erythematosus from Northwest Spain. Rheumatol Int. 2014;34(1):11–24.
- [242] Zhang S, Su J, Li X, CSTAR Co-authors, et al. Chinese SLE Treatment and Research group (CSTAR) registry:
 V. gender impact on Chinese patients with systemic lupus erythematosus. Lupus. 2015;24(12):1267–1275.
- [243] Rastin M, Mahmoudi M, Sahebari M, et al. Clinical & immunological characteristics in systemic lupus erythematosus patients. Indian J Med Res. 2017; 146(2):224–229.
- [244] Urrestarazú A, Otatti G, Silvariño R, et al. Lupus nephritis in males: clinical features, course, and prognostic factors for end-stage renal disease. Kidney Int Rep. 2017;2(5):905–912.
- [245] Santamaría-Alza Y, Motta JZN, Fajardo-Rivero JE, et al. Systemic lupus erythematosus, gender differences in Colombian patients. Clin Rheumatol. 2018; 37(9):2423–2428.
- [246] De Oliveira NT, Silva NG, Dos Santos TAFG, Department of Medicine, Mackenzie Evangelical School of Medicine, Curitiba, Brazil, et al. Clinical and autoantibody profile in male and female patients with systemic lupus erythematosus: a retrospective study in 603 Brazilian patients. Eur J Rheumatol. 2020;7(4):164–168.
- [247] Hwang J, Lee J, Ahn JK, et al. Clinical characteristics of male and female Korean patients with systemic lupus erythematosus: a comparative study. Korean J Intern Med. 2015;30(2):242–249.
- [248] Van Griensven M, Bergijk EC, Baelde JJ, et al. Differential effects of sex hormones on autoantibody production and proteinuria in chronic graft-versushost disease-induced experimental lupus nephritis. Clin Exp Immunol. 1997;107(2):254–260.
- [249] Bassi N, Luisetto R, Ghirardello A, et al. 17-β-estradiol affects BLyS serum levels and the nephritogenic autoantibody network accelerating glomerulonephritis in NZB/WF1 mice. Lupus. 2015;24(4–5):382–391.
- [250] Feng F, Nyland J, Banyai M, et al. The induction of the lupus phenotype by estrogen is via an estrogen receptor-alpha-dependent pathway. Clin Immunol. 2010;134(2):226–236.
- [252] Yurino H, Ishikawa S, Sato T, et al. Endocrine disruptors (environmental estrogens) enhance

autoantibody production by B1 cells. Toxicol Sci. 2004;81(1):139–147.

- [253] Tabor DE, Gould KA. Estrogen receptor alpha promotes lupus in (NZB × NZW)F1 mice in a B cell intrinsic manner. Clin Immunol. 2017;174:41–52.
- [254] Bynoté KK, Hackenberg JM, Korach KS, et al. Estrogen receptor-alpha deficiency attenuates autoimmune disease in (NZB x NZW)F1 mice. Genes Immun. 2008;9(2):137–152.
- [255] Svenson JL, EuDaly J, Ruiz P, et al. Impact of estrogen receptor deficiency on disease expression in the NZM2410 lupus prone mouse. Clin Immunol. 2008; 128(2):259–268.
- [256] Cunningham MA, Richard ML, Wirth JR, et al. Novel mechanism for estrogen receptor alpha modulation of murine lupus. J Autoimmun. 2019;97:59–69.
- [257] Scott JL, Wirth JR, Eudaly J, et al. Complete knockout of estrogen receptor alpha is not directly protective in murine lupus. Clin Immunol. 2017;183:132–141.
- [258] Shim G-J, Kis LL, Warner M, et al. Autoimmune glomerulonephritis with spontaneous formation of splenic germinal centers in mice lacking the estrogen receptor alpha gene. Proc Natl Acad Sci U S A. 2004; 101(6):1720–1724.
- [259] Xie Q-M, Hu H-Q, Li S-S, et al. Association of oestrogen receptor alpha gene polymorphisms with systemic lupus erythematosus risk: an updated metaanalysis. Microb Pathog. 2019;127:352–358.
- [260] Drehmer MN, Andrade D, Pereira IA, et al. Estrogen receptor alpha gene (ESR1) polymorphism can contribute to clinical findings in systemic lupus erythematosus patients. Lupus. 2017;26(3):294–298.
- [261] Liu Z-H, Cheng Z-H, Gong R-J, et al. Sex differences in estrogen receptor gene polymorphism and its association with lupus nephritis in Chinese. Nephron. 2002;90(2):174–180.
- [262] Lee MH, Chakhtoura M, Sriram U, et al. Conventional DCs from male and female lupus-prone B6.NZM Sle1/Sle2/Sle3 mice express an IFN signature and have a higher immunometabolism that are enhanced by estrogen. J Immunol Res. 2018;2018:1601079.
- [263] Cunningham MA, Naga OS, Eudaly JG, et al. Estrogen receptor alpha modulates Toll-like receptor signaling in murine lupus. Clin Immunol. 2012;144(1):1–12.
- [264] Svenson J, Cunningham M, Dasgupta S, et al. Estrogen receptor alpha modulates mesangial cell responses to toll-like receptor ligands. Am J Med Sci. 2014;348(6):492–500.
- [265] Xue L, Liu Z, Hu J, et al. Estrogen-induced expression of tumor necrosis factor-like weak inducer of apoptosis through ERα accelerates the progression of lupus nephritis. Rheumatology (Oxford). 2016;55(10): 1880–1888.
- [266] Rider V, Abdou NI, Kimler BF, et al. Gender bias in human systemic lupus erythematosus: a problem of steroid receptor action? Front Immunol. 2018;9:611.
- [267] Rodrigues JC, Haas M, Reich HN. IgA nephropathy. Clin J Am Soc Nephrol. 2017;12(4):677–686.
- [268] Huang PP, Shu DH, Su Z, et al. Association between lifestyle, gender and risk for developing end-stage renal failure in IgA nephropathy: a case-control study within 10 years. Ren Fail. 2019;41(1):914–920.

- [269] Deng W, Tan X, Zhou Q, et al. Gender-related differences in clinicopathological characteristics and renal outcomes of Chinese patients with IgA nephropathy. BMC Nephrol. 2018;19(1):31.
- [270] Greene DM, Azcona-Olivera JI, Pestka JJ. Vomitoxin (deoxynivalenol)-induced IgA nephropathy in the B6C3F1 mouse: dose response and male predilection. Toxicology. 1994;92(1–3):245–260.
- [271] Greene DM, Azcona-Olivera JI, Murtha JM, et al. Effects of dihydrotestosterone and estradiol on experimental IgA nephropathy induced by vomitoxin. Fundam Appl Toxicol. 1995;26(1):107–116.
- [272] Yamamoto R, Nagasawa Y, Shoji T, et al. A candidate gene approach to genetic contributors to the development of IgA nephropathy. Nephrol Dial Transplant. 2012;27(3):1020–1030.
- [273] Mirfazeli ES, Marashi SA, Kalantari S. In silico prediction of specific pathways that regulate mesangial cell proliferation in IgA nephropathy. Med Hypotheses. 2016;97:38–45.
- [274] Hu S-L, Wang D, Yuan F-L, et al. Identification of key genes and pathways in IgA nephropathy using bioinformatics analysis. Medicine (Baltimore). 2020; 99(30):e21372.
- [275] Yu W, Zhao B, Zhong H, et al. Estrogen receptor alpha expression in renal tissue and its relationship with prognosis in immunoglobulin A nephropathy. Int J Clin Exp Pathol. 2020;13(9):2319–2325.
- [276] Gross ML. Beneficial effects of estrogens on indices of renal damage in uninephrectomized SHRsp rats. J Am Soc Nephrol. 2004;15(2):348–358.
- [277] Mercantepe T, Unal D, Selli J, et al. Protective effects of estrogen and bortezomib in kidney tissue of postmenopausal rats: an ultrastructural study. Ren Fail. 2016;38(7):1129–1135.
- [278] Kasimay O, Sener G, Cakir B, et al. Estrogen protects against oxidative multiorgan damage in rats with chronic renal failure. Ren Fail. 2009;31(8):711–725.
- [279] Maric C, Sandberg K, Hinojosa-Laborde C. Glomerulosclerosis and tubulointerstitial fibrosis are attenuated with 17beta-estradiol in the aging Dahl salt sensitive rat. J Am Soc Nephrol. 2004;15(6): 1546–1556.
- [280] Mao S, Xu HUA, Zou L, et al. Estrogen preserves split renal function in a chronic complete unilateral ureteral obstruction animal model. Exp Ther Med. 2014;7(6):1555–1562.
- [281] Kim D, Lee AS, Jung YJ, et al. Tamoxifen ameliorates renal tubulointerstitial fibrosis by modulation of estrogen receptor α -mediated transforming growth factor- β 1/Smad signaling pathway. Nephrol Dial Transplant. 2014;29(11):2043–2053.
- [282] Dellê H, Rocha JRC, Cavaglieri RC, et al. Antifibrotic effect of tamoxifen in a model of progressive renal disease. J Am Soc Nephrol. 2012;23(1):37–48.
- [283] Diwan V, Small D, Kauter K, et al. Gender differences in adenine-induced chronic kidney disease and cardiovascular complications in rats. Am J Physiol Renal Physiol. 2014;307(11):F1169–F1178.
- [284] Ichii O, Nakamura T, Irie T, et al. Close pathological correlations between chronic kidney disease and reproductive organ-associated abnormalities in

female cotton rats. Exp Biol Med (Maywood). 2018; 243(5):418–427.

- [285] Moonen L, D'Haese PC, Vervaet BA. Epithelial cell cycle behaviour in the injured kidney. Int J Mol Sci. 2018;19(7): 2038.
- [286] Kramann R, Kusaba T, Humphreys BD. Who regenerates the kidney tubule? Nephrol Dial Transplant. 2015;30(6):903–910.
- [287] Sánchez DS, Fischer Sigel LK, Azurmendi PJ, et al. Estradiol stimulates cell proliferation via classic estrogen receptor-alpha and G protein-coupled estrogen receptor-1 in human renal tubular epithelial cell primary cultures. Biochem Biophys Res Commun. 2019; 512(2):170–175.
- [288] Sun J, Langer WJ, Devish K, et al. Compensatory kidney growth in estrogen receptor-alpha null mice. Am J Physiol Renal Physiol. 2006;290(2):F319–F323.
- [289] Tieu A, House AA, Urquhart BL. Drug disposition issues in CKD: implications for drug discovery and regulatory approval. Adv Chronic Kidney Dis. 2016; 23(2):63–66.
- [290] Lin H-F, Liao K-F, Chang C-M, et al. Correlation of the tamoxifen use with the increased risk of deep vein thrombosis and pulmonary embolism in elderly women with breast cancer: a case-control study. Medicine (Baltimore). 2018;97(51):e12842.
- [291] Adomaityte J, Farooq M, Qayyum R. Effect of raloxifene therapy on venous thromboembolism in postmenopausal women. A meta-analysis. Thromb Haemost. 2008;99(02):338–342.
- [292] Isakova T, Nickolas TL, Denburg M, et al. KDOQI US commentary on the 2017 KDIGO Clinical Practice Guideline update for the diagnosis, evaluation, prevention, and treatment of Chronic Kidney Disease-Mineral and Bone Disorder (CKD-MBD). Am J Kidney Dis. 2017;70(6):737–751.
- [293] Chen H, Han X, Cui Y, et al. Parathyroid hormone fragments: new targets for the diagnosis and treatment of chronic kidney disease-mineral and bone disorder. Biomed Res Int. 2018;2018:9619253.
- [294] Webster R, Sheriff S, Faroqui R, et al. Klotho/fibroblast growth factor 23- and PTH-independent estrogen receptor-α-mediated direct downregulation of NaPi-Ila by estrogen in the mouse kidney. Am J Physiol Renal Physiol. 2016;311(2):F249–F259.
- [295] Meng J, Ohlsson C, Laughlin GA, Osteoporotic Fractures in Men (MrOs) Study Group, et al. Associations of estradiol and testosterone with serum phosphorus in older men: the Osteoporotic Fractures in Men study. Kidney Int. 2010;78(4): 415–422.
- [296] Ix JH, Chonchol M, Laughlin GA, et al. Relation of sex and estrogen therapy to serum fibroblast growth factor 23, serum phosphorus, and urine phosphorus: the Heart and Soul Study. Am J Kidney Dis. 2011; 58(5):737–745.
- [297] Masaki H, Imanishi Y, Naka H, et al. Bazedoxifene improves renal function and increases renal phosphate excretion in patients with postmenopausal osteoporosis. J Bone Miner Metab. 2020;38(3): 405–411.

- [298] Suzuki H, Kondo K. Chronic kidney disease in postmenopausal women. Hypertens Res. 2012;35(2): 142–147.
- [299] Khairallah P, Nickolas TL. Management of osteoporosis in CKD. Clin J Am Soc Nephrol. 2018;13(6): 962–969.
- [300] Haghverdi F, Farbodara T, Mortaji S, et al. Effect of raloxifene on parathyroid hormone in osteopenic and osteoporotic postmenopausal women with chronic kidney disease stage 5. Iran J Kidney Dis. 2014;8(6):461–466.
- [301] Saito O, Saito T, Asakura S, et al. The effects of raloxifene on bone turnover markers and bone mineral density in women on maintenance hemodialysis. Clin Exp Nephrol. 2011;15(1):126–131.
- [302] Nagatoya K, Nishimoto K, Shibahara N, Hokusetsu Renal Osteodystrophy Study Group, et al. Effects of raloxifene on bone metabolism in postmenopausal women on chronic hemodialysis. Clin Exp Nephrol. 2015;19(5):939–946.
- [303] Ishani A, Blackwell T, Jamal SA, et al. The effect of raloxifene treatment in postmenopausal women with CKD. JASN. 2008;19(7):1430–1438.
- [304] Heilberg IP, Hernandez E, Alonzo E, et al. Estrogen receptor (ER) gene polymorphism may predict the bone mineral density response to raloxifene in postmenopausal women on chronic hemodialysis. LRNF. 2005;27(2):155–161.
- [305] Zhang S, Guo Y, Zou H, et al. Effect of estrogen deficiency on the fixation of titanium implants in chronic kidney disease mice. Osteoporos Int. 2015;26(3): 1073–1080.
- [306] Guo Y, Sun N, Duan X, et al. Estrogen deficiency leads to further bone loss in the mandible of CKD mice. PLoS One. 2016;11(2):e0148804.
- [307] Newman CL, Creecy A, Granke M, et al. Raloxifene improves skeletal properties in an animal model of cystic chronic kidney disease. Kidney Int. 2016;89(1): 95–104.
- [308] Kanatani M, Sugimoto T, Takahashi Y, et al. Estrogen via the estrogen receptor blocks cAMP-mediated parathyroid hormone (PTH)-stimulated osteoclast formation. J Bone Miner Res. 1998;13(5):854–862.
- [309] Liu B-Y, Wu P-W, Bringhurst FR, et al. Estrogen inhibition of PTH-stimulated osteoclast formation and attachment in vitro: involvement of both PKA and PKC. Endocrinology. 2002;143(2):627–635.
- [310] Romagnani P, Remuzzi G, Glassock R, et al. Chronic kidney disease. Nat Rev Dis Primers. 2017;3:17088.
- [311] Voelkl J, Lang F, Eckardt K-U, et al. Signaling pathways involved in vascular smooth muscle cell calcification during hyperphosphatemia. Cell Mol Life Sci. 2019;76(11):2077–2091.
- [312] Rzewuska-Lech E, Jayachandran M, Fitzpatrick LA, et al. Differential effects of 17beta-estradiol and raloxifene on VSMC phenotype and expression of osteoblast-associated proteins. Am J Physiol Endocrinol Metab. 2005;289(1):E105–E112.
- [313] Nanao-Hamai M, Son B-K, Hashizume T, et al. Protective effects of estrogen against vascular calcification via estrogen receptor α-dependent growth

arrest-specific gene 6 transactivation. Biochem Biophys Res Commun. 2016;480(3):429–435.

- [314] Kim SA, Lee KY, Kim J-R, et al. Estrogenic compound attenuates angiotensin Il-induced vascular smooth muscle cell proliferation through interaction between LKB1 and estrogen receptor α. J Pharmacol Sci. 2016; 132(1):78–85.
- [315] Chang SP, Yang WS, Lee KS, et al. Effects of hormonal replacement therapy on oxidative stress and total antioxidant capacity in postmenopausal hemodialysis patients. Ren Fail. 2002;24(1):49–57.
- [316] Kendrick J, Chonchol M. Novel therapeutic options for the treatment of mineral metabolism abnormalities in end stage renal disease. Semin Dial. 2015; 28(6):610–619.
- [317] National Kidney Foundation. K/DOQI clinical practice guidelines for bone metabolism and disease in chronic kidney disease. Am J Kidney Dis. 2003;42(4 Suppl 3):S1–S201.
- [318] Ma H, Ouyang C, Huang Y, et al. Comparison of microwave ablation treatments in patients with renal secondary and primary hyperparathyroidism. Ren Fail. 2020;42(1):66–76.
- [319] Yu M-A, Yao L, Zhang L, et al. Safety and efficiency of microwave ablation for recurrent and persistent secondary hyperparathyroidism after parathyroidectomy: a retrospective pilot study. Int J Hyperthermia. 2016;32(2):180–186.
- [320] Lin X-H, Lin C-C, Wang Y-J, et al. Risk factors of the peptic ulcer bleeding in aging uremia patients under regular hemodialysis. J Chin Med Assoc. 2018;81(12): 1027–1032.
- [321] Wakasugi M, Matsuo K, Kazama JJ, et al. Higher mortality due to intracerebral hemorrhage in dialysis patients: a comparison with the general population in Japan. Ther Apher Dial. 2015;19(1):45–49.
- [322] Sohal AS, Gangji AS, Crowther MA, et al. Uremic bleeding: pathophysiology and clinical risk factors. Thromb Res. 2006;118(3):417–422.
- [323] Hunt BJ. Bleeding and coagulopathies in critical care. N Engl J Med. 2014;370(9):847–859.
- [324] Hussain S, Siddiqui AN, Baxi H, et al. Warfarin use increases bleeding risk in hemodialysis patients with atrial fibrillation: a meta-analysis of cohort studies. J Gastroenterol Hepatol. 2019;34(6):975–984.
- [325] Ishii M, Ogawa H, Unoki T, et al. Relationship of hypertension and systolic blood pressure with the risk of stroke or bleeding in patients with atrial fibrillation: the Fushimi AF Registry. Am J Hypertens. 2017;30(11):1073–1082.
- [326] Yamashita Y, Takagi D, Hamatani Y, et al. Clinical characteristics and outcomes of dialysis patients with atrial fibrillation: the Fushimi AF Registry. Heart Vessels. 2016;31(12):2025–2034.
- [327] Gonzalez J, Bryant S, Hermes-DeSantis ER. Transdermal estradiol for the management of refractory uremic bleeding. Am J Health Syst Pharm. 2018; 75(9):e177–e183.
- [328] Pei J, Harakalova M, den Ruijter H, et al. Cardiorenal disease connection during post-menopause: the protective role of estrogen in uremic toxins induced

microvascular dysfunction. Int J Cardiol. 2017;238: 22–30.

- [329] Hedges SJ, Dehoney SB, Hooper JS, et al. Evidencebased treatment recommendations for uremic bleeding. Nat Clin Pract Nephrol. 2007;3(3):138–153.
- [330] Szilagyi A, Ghali MP. Pharmacological therapy of vascular malformations of the gastrointestinal tract. Can J Gastroenterol. 2006;20(3):171–178.
- [331] Muftah M, Mulki R, Dhere T, et al. Diagnostic and therapeutic considerations for obscure gastrointestinal bleeding in patients with chronic kidney disease. Ann Gastroenterol. 2019;32(2):113–123.
- [332] Mosconi G, Mambelli E, Zanchelli F, et al. Severe gastrointestinal bleeding in a uremic patient treated with estrogen-progesterone therapy. Int J Artif Organs. 1999;22(5):313–316.
- [333] Hermans C, Goffin E, Horsmans Y, et al. Watermelon stomach. An unusual cause of recurrent upper Gl tract bleeding in the uraemic patient: efficient treatment with oestrogen-progesterone therapy. Nephrol Dial Transplant. 1996;11(5):871–874.
- [334] Bali A, Hix JK, Kouides P. Safe and effective use of chronic transdermal estradiol for life-threatening uremic bleeding in a patient with coronary artery disease. Nephron Extra. 2014;4(2):134–137.
- [335] Viganò G, Zoja C, Corna D, et al. 17 beta-estradiol is the most active component of the conjugated estrogen mixture active on uremic bleeding by a receptor mechanism. J Pharmacol Exp Ther. 1990;252(1):344–348.
- [336] Virdis A, Ghiadoni L, Pinto S, et al. Mechanisms responsible for endothelial dysfunction associated with acute estrogen deprivation in normotensive women. Circulation. 2000;101(19):2258–2263.
- [337] Brunini TMC, da Silva CD, Siqueira MAS, et al. Uremia, atherothrombosis and malnutrition: the role of L-arginine-nitric oxide pathway. Cardiovasc Hematol Disord Drug Targets. 2006;6(2):133–140.
- [338] Brunini TMC, Mendes-Ribeiro AC, Ellory JC, et al. Platelet nitric oxide synthesis in uremia and malnutrition: a role for L-arginine supplementation in vascular protection? Cardiovasc Res. 2007;73(2):359–367.
- [339] Gurney EP, Nachtigall MJ, Nachtigall LE, et al. The Women's Health Initiative trial and related studies: 10 years later: a clinician's view. J Steroid Biochem Mol Biol. 2014;142:4–11.
- [340] Rossouw JE, Manson JE, Kaunitz AM, et al. Lessons learned from the Women's Health Initiative trials of menopausal hormone therapy. Obstet Gynecol. 2013; 121(1):172–176.
- [341] K/DOQI Workgroup. K/DOQI clinical practice guidelines for cardiovascular disease in dialysis patients. Am J Kidney Dis. 2005;45(4 Suppl 3):S1–S153.
- [342] Kramer HM, Curhan GC, Singh A, Hemodialysis and Estrogen Levels in Postmenopausal Patients Study Group. Permanent cessation of menses and postmenopausal hormone use in dialysis-dependent women: the HELP study. Am J Kidney Dis. 2003;41(3): 643–650.
- [343] Vellanki K, Hou S. Menopause in CKD. Am J Kidney Dis. 2018;71(5):710–719.
- [344] Dobrowolski C, Clark EG, Sood MM. Venous thromboembolism in chronic kidney disease: epidemiology,

the role of proteinuria, CKD severity and therapeutics. J Thromb Thrombolysis. 2017;43(2):241–247.

- [345] Lu HY, Liao KM. Increased risk of deep vein thrombosis in end-stage renal disease patients. BMC Nephrol. 2018;19(1):204.
- [346] Huang M-J, Wei R-b, Wang Y, et al. Blood coagulation system in patients with chronic kidney disease: a prospective observational study. BMJ Open. 2017; 7(5):e014294.
- [347] Shashar M, Belghasem ME, Matsuura S, et al. Targeting STUB1-tissue factor axis normalizes hyperthrombotic uremic phenotype without increasing bleeding risk. Sci Transl Med. 2017;9(417): eaam8475.
- [348] DeLoughery TG. Estrogen and thrombosis: controversies and common sense. Rev Endocr Metab Disord. 2011;12(2):77–84.
- [349] Ahmed SB, Ramesh S. Sex hormones in women with kidney disease. Nephrol Dial Transplant. 2016;31(11): 1787–1795.
- [350] Andreoli SP. Hormone replacement therapy in postmenopausal women with end-stage renal disease (1). Kidney Int. 2000;57(1):341–342.
- [351] Hoshi-Fukushima R, Nakamoto H, Imai H, et al. Estrogen and angiotensin II interactions determine cardio-renal damage in Dahl salt-sensitive rats with heart failure. Am J Nephrol. 2008;28(3):413–423.