

Review



Clinical Impact of Tumor-Infiltrating Lymphocytes and PD-L1-Positive Cells as Prognostic and Predictive Biomarkers in Urological Malignancies and Retroperitoneal Sarcoma

Makito Miyake ^{1,*}, Shunta Hori ¹, Takuya Owari ¹, Yuki Oda ¹, Yoshihiro Tatsumi ¹, Yasushi Nakai ¹, Tomomi Fujii ² and Kiyohide Fujimoto ¹

- ¹ Department of Urology, Nara Medical University, Nara 634-8522, Japan; horimaus@naramed-u.ac.jp (S.H.); tintherye@naramed-u.ac.jp (T.O.); K197610@naramed-u.ac.jp (Y.O.); takuro.birds.nest@gmail.com (Y.T.); nakaiyasusiuro@live.jp (Y.N.); kiyokun@naramed-u.ac.jp (K.F.)
- ² Department of Diagnostic Pathology, Nara Medical University, Nara 634-8522, Japan; fujiit@naramed-u.ac.jp
- * Correspondence: makitomiyake@yahoo.co.jp; Tel.: +81-744-22-3051; Fax: +81-744-22-9282

Received: 16 September 2020; Accepted: 24 October 2020; Published: 27 October 2020



Simple Summary: Two host-dependent biological characteristics, "avoiding immune destruction" and "tumor-promoting inflammation" have been added to cancer hallmarks in 2011. The interaction and cross-talk among tumor cells and several immune cells in a tumor microenvironment are dynamic and complex processes. The purpose of this review is to discuss the prognostic impact of tumor-infiltrating lymphocytes and predictive biomarkers for immune checkpoint inhibitors in four urological solid tumors, the urothelial carcinoma, renal cell carcinoma, prostate cancer, and retroperitoneal sarcoma, through summarizing the findings of observation studies and clinical trials.

Abstract: Over the past decade, an "immunotherapy tsunami", more specifically that involving immune checkpoint inhibitors (ICIs), has overtaken the oncological field. The interaction and cross-talk among tumor cells and several immune cells in the tumor microenvironment are dynamic and complex processes. As immune contexture can vary widely across different types of primary tumors and tumor microenvironments, there is still a significant lack of clinically available definitive biomarkers to predict patient response to ICIs, especially in urogenital malignancies. An increasing body of evidence evaluating urological malignancies has proven that tumor-infiltrating lymphocytes (TILs) are a double-edged sword in cancer. There is an urgent need to shed light on the functional heterogeneity in the tumor-infiltrating immune system and to explore its prognostic impact following surgery and other treatments. Notably, we emphasized the difference in the immunological profile among urothelial carcinomas arising from different primary origins, the bladder, renal pelvis, and ureter. Significant differences in the density of FOXP3-positive TILs, CD204-positive tumor-infiltrating macrophages, PD-L1-positive cells, and colony-stimulating factors were observed. This review discusses two topics: (i) the prognostic impact of TILs and (ii) predictive biomarkers for ICIs, to shed light on lymphocyte migration in four solid tumors, the urothelial carcinoma, renal cell carcinoma, prostate cancer, and retroperitoneal sarcoma.

Keywords: tumor-infiltrating lymphocyte; immune cells; tumor microenvironment; prognosis; immune checkpoint inhibitor; treatment response; urothelial carcinoma; renal cell carcinoma; prostate cancer; retroperitoneal sarcoma

1. Introduction

The interaction and cross-talk among tumor cells and several immune cells in a tumor microenvironment are dynamic and complex processes [1,2]. In 2011, Hanahan and Weinberg defined "avoiding immune destruction" and "tumor-promoting inflammation" as emerging cancer hallmarks [3], which are host-dependent biological characteristics play crucial roles in the immune cell-mediated orchestration of tumor proliferation, progression, angiogenesis, epithelial-to-mesenchymal transition (EMT), invasion, and metastasis [4,5]. Although inflammation caused by the innate immune system was originally designed to fight infections and heal wounds, "tumor-promoting inflammation" can inadvertently contribute to multiple cancer hallmark capabilities by supplying active molecules to the tumor microenvironment [3,6–8]. "Avoiding immune destruction" allows tumor cells to escape immunosurveillance; the main role of T lymphocytes, B lymphocytes, macrophages, natural killer (NK) cells, neutrophils, and dendritic cells [3,9,10].

An increasing body of oncological research has provided evidence to suggest that tumor-infiltrating lymphocytes (TILs), including T lymphocytes such as CD8+ T cells and regulatory T cells, and NK cells such as tumor-associated NK cells and tumor-infiltrating NK cells, are double-edged swords in cancer [9,11,12]. The density, composition, and types of TILs vary greatly across tumor stage [10] and tumor entity [13]. Moreover, these features display significant heterogeneity between patients with the same type of tumors [14,15]. Lymphocyte migration to neoplastic lesions is mainly controlled by chemotactic factors, including chemokines and small cytokines, which are secreted from immune cells and tumor cells [16]. There are three types of immunological profiles: (1) immunologically "tumors" present with a high degree of T cell infiltration (e.g., melanoma, non-small-cell lung carcinoma, and renal cell carcinoma (RCC)) [17,18]; (2) immunologically "cold tumors" present with scarce immune infiltrates (e.g., prostate cancer (PCa) and pancreatic cancer) [19,20]; (3) immunologically "altered tumors" in which peri-tumoral sites are densely inflamed with immune cells which lack the capability to infiltrate the tumor [21]. Classification of tumors according to their immune phenotype can help predict responses to immune checkpoint inhibitors (ICIs), such as anti-programmed cell death 1 (PD-1) inhibitors, pembrolizumab, and nivolumab. Higher levels of immune cell infiltration and interferon signature (a T-cell-inflamed phenotype) are associated with a positive response to ICIs [19]. However, many other solid tumors fail to respond to ICIs due to limited immunogenicity, unfavorable tumor microenvironments with scarce immune infiltrates, and excessive accumulation of regulatory T cells [17]. Therefore, the potential to develop new therapeutic approaches that can convert immunologically 'cold' or 'altered' environments to 'hot' environments has recently attracted increasing attention. [22,23].

As immune contexture can vary widely across types of tumor and tumor microenvironment, there exists a significant lack of clinically available definitive biomarkers that provide accurate predictions for treatment responses, especially in urogenital malignancies. Additionally, in this review, we will discuss the functional heterogeneity in the tumor-infiltrating immune system to explore its prognostic impact after surgery and other treatments. This review focuses on two main topics: (i) the prognostic impact of TILs and (ii) predictive biomarker for ICIs, to shed light on lymphocyte migration in four solid tumors which are urothelial carcinoma (UC), RCC, PCa, and retroperitoneal sarcoma (RSar) (Figure 1).



Currently available PD-L1 companion diagnostic assays predicting response

_	Nivolumab
_	Pembrolizumab
_	Atezolizumab
-	Durvalumab
	- - -

Figure 1. Two topics discussed in this review. Urologists should handle several malignancies arising from different organs, including the bladder, renal pelvis, ureter, kidney, prostate, and tissues of the retroperitoneal cavity. In this review, we discuss two topics: (1) the prognostic impact of tumor-infiltrating leukocytes (TILs) and (2) predictive markers for immune checkpoint inhibitors to shed light on lymphocyte migration in four solid tumors, the urothelial carcinoma, renal cell carcinoma, PCa, and retroperitoneal sarcoma. Currently available PD-L1 companion diagnostic assays predicting response to immune checkpoint inhibitors are shown.

2. Methods

2.1. Literature Search

We searched for relevant papers published before 30 August 2020, by using the PubMed database with the following terms: "urothelial carcinoma" or "renal cell carcinoma" or "prostate cancer" or "sarcoma", "tumor-infiltrating lymphocyte/TIL", "tumor microenvironment", "immune checkpoint inhibitor", and "programmed death-ligand 1/PD-L1". The inclusion criteria were as follows: studies which addressed the relevance between clinical outcomes including the treatment response and TILs, other immune cells, and PD-L1 positive cells. Any of immunohistochemical (IHC) staining analyses, hematoxylin and eosin (HE) staining analysis, and flow cytometry (FCM) analysis, and dataset analysis were allowed as methods for assessment of immune cells. Various types of retrospective studies and prospective clinical trials were included. If the same study was reported more than once, we selected the most recent publication with updated information.

2.2. Immunohistochemical Staining and Quantification in Tumor Tissues of UC Diseases Arising from Different Origins

IHC staining using paraffin-embedded, formalin-fixed tissue blocks was performed as previously described [16]. Bladder, renal pelvis, and ureter are three common primary origins of UC and compared in terms of the immune profile. The patients' background is depicted in Table S1. The primary/secondary antibodies used in this study and the conditions are available in Table S2. A Histofine Simple Stain[™] MAX PO (MULTI) kit (Nichirei Corporation, Tokyo, Japan) was used for peroxidase color development according to the manufacturer's instructions. The staining results were evaluated by two investigators (Y.T. and Y.N.) who were blinded to any clinicopathological data. Data were expressed by box-and-whisker plots, in which the outliers are indicated as dots, and compared using the Kruskal-Wallis test, followed by the post hoc test (Dunn test). This observational study was approved by the ethics committee of the Nara Medical University, and informed consent from the participants was obtained in the form of written signature or opt-out on the web-site (reference ID: 1256).

3. Urothelial Carcinoma (UC)

3.1. Epidemiology and Current Issues of UC

UC is a histopathological type of cancer that typically arises from the urothelium of the bladder, renal pelvis, ureter, or urethral. UC of the bladder is the most common malignancy involving the urinary tract and is the sixth most common cancer in the United States [24]. Surgical resection is a standard treatment strategy for UC and other solid tumors. However, unresectable/metastatic UC is strongly associated with poor prognosis and requires multidisciplinary therapy, including chemotherapy, ICIs, and palliative radiotherapy. Wide use of pembrolizumab (KEYTRUDA[®], Merck & Co. Inc., Kenilworth, NJ, USA) has clearly improved survival in selected patients with advanced UC [25]. As there exists a lack of prognostic markers after radical surgery and predictive markers for response to ICIs, we focused on the potential of TILs as a clinically available biomarker.

3.2. Clinical Impact of TILs in Patients with UC

Table 1 summarizes previous studies addressing the clinical relevance of TILs in patients with UC [26–37]. There are several types of treatment settings, including transurethral resection of bladder tumor (TURBT) with or without intravesical Bacillus Calmette-Guérin (BCG) for non-muscle invasive bladder cancer (NMBIC), radical cystectomy for muscle-invasive bladder cancer (MIBC), radical nephroureterectomy for upper urinary tract urothelial cancer (UTUC), and systemic chemotherapy for metastatic UC. Our group previously reported that a higher number of tumor-infiltrating regulatory T cells (Tregs) was associated with a higher risk of treatment failure [26]. This result was reasonable in terms of the hypothesis that pre-BCG baseline status of Th1/Th2 balance and Treg recruitment in the tumor microenvironment, or both, could influence the response to BCG [38]. Wahlin et al. demonstrated that CD8+ and FoxP3+ TILs in cystectomy specimens were independently associated with better outcomes, which disagrees with our previous data [26,32]. Additionally, this association was only observed in CD8+ TILs in TURBT specimens [32]. However, we understand that the data can be affected by the type of specimen and patient background, including treatment and race. Due to accessibility, most studies have utilized IHC staining analyses using archival paraffin-embedded tissues. Kawashima et al. evaluated nine extracellular surface markers measured by flow cytometry in freshly resected UC specimens [33]. In their study, Kawashima et al. classified tumors into the CD4 T-cell-dominant group and the immunologically activated group according to the immunologic condition, concluding that the latter group showed significantly poorer outcomes in patients with MIBC. Evaluation with freshly resected UC specimens is costly, time consuming, and requires effort; however, the results obtained are often robust and reliable. Overall, aside from the method of evaluation and interpretation, a consensus has not yet been reached regarding the prognostic value of TILs in patients with UC.

Types of Tumor	No. of Patients	Treatment	Makers or Assessment Assay		Clinical Relevance	Reference No.
Urothelial Ca	rcinoma (UC)					
NMIBC	154	TURBT followed by intravesical BCG	FOXP3, CD204	IHC	High Tregs and tumor-associated macrophages were associated with a high risk of intravesical recurrence.	[26]
NMIBC	115	TURBT	CD3, CD4, CD8, CD20, CD56, CD68, granzyme B	IHC	Low CD3+ TILs and CD8+ TILs were associated with a high risk of intravesical recurrence.	[27]
NMIBC	131	TURBT	CD4	IHC	High CD4+ TILs were associated with poor OS.	[28]
NMIBC	102	TURBT	CD8, CD66b	IHC	High tumor-infiltrating neutrophils and NLR were associated with poor OS. High TILs were related to longer OS.	[29]
MIBC	67	Radical cystectomy	CD3, CD8	IHC	High CD8+ TILs and CD3+ TILs in the invasion margin were associated with better DFS and OS.	[30]
MIBC	406	Radical cystectomy	CD3D, CD4, CD8A	D3D, CD4, CD8A mRNA (TCGA dataset) High CD3D/CD4 ratio was associated with improved survival. The power was stronger in basal-squamous tumors.		[31]
MIBC	145	Radical cystectomy	CD8, FOXP3, CD20, PD-1, PD-L1	IHC	High density of CD8, FOXP3, CD20, and PD-1 was associated with a low risk of recurrence.	[32]
Bladder cancer and UTUC	52 and 18	Surgical resection Nine extracellular surface markers		FCM	The immunologically activated group showed poorer PFS and CSS compared that in to the CD4+ T-cell-dominant group in bladder cancer. However, there was no significant difference in UTUC.	[33]
UTUC	162	Radical nephroureterectomy	PD-L1	IHC	High PD-L1 expression in tumor cells was associated with shorter CSS. High PD-L1 expression on TILs was associated with longer CSS.	[34]
UTUC	423	Radical nephroureterectomy	PD-1, PD-L1	IHC	High PD-1 level was associated with poor CSS and OS. In patients with organ-confined disease (pT2≤, N0/xM0), high PD-L1 was associated with a high risk of recurrence and poor OS.	[35]

Table 1. Studies for the clinical relevance of tumor-infiltrating lymphocytes (TILs) in urological malignancies and retroperitoneal sarcoma.

Types of Tumor	No. of Patients	Treatment	Makers or Assessment	Assay	Clinical Relevance	Reference No.
UTUC	88	Radical nephroureterectomy	CD4, CD8, CD20, APE1, NTH1, OGG1, XRCC1, polβ, STING, IRF3, PD-L1, PD-L2	IHC	High CD8+ TILs were associated with poor DFS.	[36]
Metastatic UC	259	Platinum-based chemotherapy	Recommendations by an International TILs Working Group 2014	mendations by an rnational TILs eosin staining chemotherapy both in bladder ca ing Group 2014		[37]
Renal cell carc	inoma (RCC)					
ccRCC	43	Untreated stage III/IV disease	CD4, CD45RA, CD8, CD11, HLA-DR, CD3, CD16, CD57	FCM	An increase in CD8+/CD11- and a decrease in CD4+/CD45RA- cells were observed along with the aggravation of tumor stage and grade.	[39]
ccRCC	473	Previously treated	Th17, CTL, Tregs, Th2	mRNA (TCGA dataset)	Long-lived patients have high levels of Th17 and CD8+ T cells, while short-lived patients have high levels of Tregs and Th2.	[40]
RCC 891		Untreated	M1 macrophages, M2 macrophages, memory CD4+ T, $\gamma\delta$ T, CD8+ T, Tregs, naïve CD4+ T, NK cell, mast cells, B cells, DC, monocytes, plasma cells, neutrophils, eosinophils	CIBERSORT	CD8+ T cells were associated with prolonged OS. A higher proportion of regulatory T cells was associated with a poorer outcome. M1 macrophages were associated with a favorable outcome, while M2 macrophages indicated a poorer outcome.	[41]
Metastatic ccRCC	167	Previously treated	CD8, PD-1, TIM-3, LAG-3	IHC	A high percentage of CD8+/PD-1+/TIM-3-/LAG-3- cells correlated with high levels of T-cell activation and were associated with longer median irPFS and higher irORR.	[42]
ccRCC	199	Previously treated	PD-1, FOXP3	IHC	PD1-positive or FOXP3-positive lymphocytes can be used as significant prognostic indicators, and PD1 positivity could be very helpful in the prediction of latent distant metastasis.	[43]

Table 1. Cont.

Types of Tumor	No. of Patients Treatment		Makers or Assessment	Assay	Clinical Relevance	Reference No.
Metastatic ccRCC	58	interleukin-2-based immunotherapy	FOXP3	IHC	Intratumoral FOXP3-positive regulatory immune cells significantly increased during interleukin-2-based immunotherapy, and high numbers of on-treatment FOXP3-positive cells were correlated with poor prognosis.	[44]
ccRCC	125	Radical nephrectomy or nephron- sparing surgery	CD4, FOXP3	CD4, FOXP3 IHC Increased peritumoral Tregs a poorer progno		[45]
ccRCC	Radical nephrectomy RCC 170 or nephron- sparing surgery		CD4, CD25, FOXP3	IHC	Increased number of CD4+CD25+Foxp3+ T cells was not associated with RCC death. In contrast, CD4+CD25+Foxp3- T cells, which may represent a unique set of Tregs or activated helper T cells, were significantly associated with the outcome.	[46]
RCC	97	Previously treated	CD45, CD3, CD4, CD8, CD45RA, ICOS, Tim3, CD25, PD-1, FOXP3	FCM	Tumor grade significantly correlated with dysfunction of both CD4+ and CD8+ TILs and the efficacy of nivolumab treatment.	[47]
Radical nephr Localized ccRCC 40 or nephro sparing sur		Radical nephrectomy or nephron- sparing surgery	CD3, CD4, CD8, CD45RA, CCR7, CD69, CD38, CD40L, ICOS, GITR, PD-1, TIM-3, CTLA-4, LAG-3, CD127, CD25	FCM	Infiltration with CD8+PD-1+Tim-3+Lag-3+ exhausted TILs and ICOS+ Tregs identified patients with deleterious prognosis who could benefit from adjuvant therapy with TME-modulating agents and checkpoint blockade.	[48]
Metastatic RCC 231 Tyrosine kina inhibitors		Tyrosine kinase inhibitors	CD8, PD-1, PD-L1	IHC	Increased numbers of CD8+ T cells are significantly associated with improved survival in patients with mRCC treated with TKIs. PD-1 could be used as a predictive and prognostic factor.	[49]
	Prostate cancer (PC	Ca)				
Localized PCa	126	Radical prostatectomy	CD8, FOXP3	IHC	High CD8+ TILs were significantly associated with good DFS, whereas FOXP3+Treg tumor infiltration was significantly correlated with poor DFS.	[50]
Localized PCa	535	Radical prostatectomy	CD8	IHC	A high density of CD8+ TILs is an independent negative prognostic factor for biochemical failure-free survival.	[51]

Table 1. Cont.

Types of Tumor	No. of Patients	Treatment	Makers or Assessment	Assay	Clinical Relevance	Reference No.
Biochemical recurence after radical prostatectomy	22	Salvage radiotherapy	PD-1, FOXP3	IHC	High PD-1 and FOXP3+ Treg tumor infiltration was significantly associated with short PFS.	[52]
Localized PCa	75	Radical prostatectomy	CCR4	IHC	CCR4+ Tregs are highly infiltrated in the prostate tissue with poor prognosis, with a strong potential to progress to CRPC.	[53]
Ret	roperitoneal sarcom	a (RSar)				
RSar (various types)	51	Surgical resection	PD-1, PD-L1, PD-L2, Ki-67	IHC	The prognostic value of PD-L1, PD-L2, and PD-1 expression was evaluated, and only high expression of PD-1 was a possible predictor of postoperative recurrence.	[54]
RSar (WDLPS)	6	Surgical resection	CD4, CD8, CD20	IHC	CD8+ T cells were mostly seen in scattered gout of the tumor. CD4+ T cells were observed in clusters and follicles. CD20+ cells (B cells) were found almost exclusively in cluster and forming immature follicles.	[55]
RSar (WDLPS/DDLPS)	8	Surgical resection	CD3, CD4,CD8, PD-1, 4-1BB	IHC FCM	Cytotoxic CD8+ T cells accounted for 20% of CD3+ T cells. Notably, 65% of CD8+ T cells were positive for PD-1. Immune cell aggregates evaluated by IHC were associated with poorer prognosis in both well-differentiated and dedifferentiated retroperitoneal liposarcoma.	[56]
RSar (WDLPS/DDLPS/ MLPS/PLPS)	56	Surgical resection	CD4, CD8, FOXP3, CD20, PD-1, PD-L1	IHC	Higher FOXP3+ Treg or PD-1/PD-L1+ cells tended to be associated with poor prognosis. Heterogeneous TIL distribution was found in 50% of patients and tended to correlate with favorable disease-free survival.	[57]

Table 1. Cont.

UC, urothelial carcinoma; NMIBC, non-muscle invasive bladder cancer; MIBC, muscle invasive bladder cancer; TURBT, transurethral resection of bladder tumor; IHC, immunohistochemical staining; TIL, tumor-infltrating lymphocyte; FCM, flow cytometry; Treg, regulatory T cell; UTUC, upper urinary tract urothelial cancer; NLR, neutrophil-to-leukocyte ration; CSS, cancer-specific survival; OS, overall survival; DFS, disease-specific survival; PFS, progression-free survival; NA, not available; ccRCC, clear cell type RCC; CRPC, castration resistant prostate cancer; WDLPS, well differentiated liposarcoma; DDLPS, dedifferentiated liposarcoma; MLPS, myxoid/round cell liposarcoma; PLPS, pleomorphic liposarcoma; PD-L1, programmed cell death ligand-1; PD-L2, programmed cell death ligand-2.

3.3. Predictive Biomarkers for Response to ICIs in UC

As of August 2020, five PD-1/programmed death-ligand 1 (PD-L1) inhibitors have been approved for the treatment of locally advanced or mUC. The U.S. Food and Drug Administration and the European Medicines Agency require IHC PD-L1 testing prior to first-line use with pembrolizumab and atezolizumab in platinum-ineligible patients [58]. Table 2 summarizes key clinical trials of ICIs for patients with UC refractory to chemotherapy [25,59–63]. There is a substantial proportion of patients who are less likely to benefit from ICIs, and the majority of the trials have so far explored the potential of PD-L1-related biomarkers to predict responses to ICIs in mUC. However, differences in antibodies, staining platforms, scoring algorithms and cut-off systems between trials have raised questions about interchangeability and comparability for diagnostic use, which could lead most pathologists to non-workable situation [58]. Used PD-L1 testing system in clinical trials are the Dako 28-8 for nivolumab, the Dako 22C3 for pembrolizumab, Ventana SP142 for atezolizumab, and the Ventana SP263 assays for durvalumab. Trials for pembrolizumab, atezolizumab, and durvalumab included combined assessments of PD-L1 staining of tumor cells and tumor-infiltrating immune cells or a single assessment of tumor-infiltrating immune cells, while the trials for nivolumab (CheckMate) included only PD-L1 expression in tumor cells [25,59,61–63]. CheckMate 032 did not check the expression of cytotoxic T-lymphocyte antigen 4 (CTLA-4) expression [60]. Expression of immune checkpoint proteins such as PD-1, PD-L1, and CTLA-4 can vary widely among primary and metastatic lesions and are affected by intratumor heterogeneity. A significant lack of correlation between the expression of these proteins and clinical outcomes/responses hampers the establishment of a Schottker-driven immunotherapy strategy.

Types of Tumor	No. of	Treatment (Phase) -	Outcomes and Response		Δεεαν	Maulau Assessment	Clinical Relevance of		
Types of Tumor	Patients	Treatment (Phase)	OS	PFS	ORR	- Assay	Marker Assessment	Maker Assessment	Keference No.
Urothel	ial carcinoma	a (UC)							
Advanced Muc (JAVELIN Solid Tumor)	249	Second-line avelumab (Phase I)	6.5 months	1.7 months	17%	NA	NA	NA	[59]
Advanced mUC (CheckMate 032)	274	Platinum-pretreated nivolumab ± ipilimumab (Phase I/II)	10.4 months in NIVO3 7.4 months in NIVO3+IPI1 27.6 months in NIVO1+IPI3	2.8 months in NIVO3 2.6 months in NIVO3+IPI1 4.9 months in NIVO1+IPI3	26% in NIVO3 27% in NIVO3+IPI1 38% in NIVO1+IPI3	IHC Dako 28-8	PD-L1 expression in tumor cells	PD-L1 expression was not associated with ORR. High PD-L1 expression was associated with longer mOS.	[60]
Advanced mUC	191	Durvalumab (Phase I/II)	18.2 months	1.5 months	18%	IHC Ventana SP263	PD-L1 (combined assessment of PD-L1 staining of tumor cells and immune cells)	Tumor response to durvalumab was not associated with PD-L1 staining.	[61]
Advanced mUC (CheckMate 275)	270	Second-line Nivolumab (Phase II)	8.7 months	2.0 months	20%	IHC Dako 28-8	PD-L1 expression in tumor cells	OR was observed in 28% of patients with PD-L1 expression of 5% or greater, 24% of patients with PD-L1 expression of 1% or greater, and 16% of patients with PD-L1 expression of less than 1%.	[62]
Advanced mUC (KEYNOTE-045)	542	Second-line Pembrolizuma vs. chemotherapy (Phase III)	10.3 vs. 7.4 months HR: 0.73 p value: 0.002	2.1 vs. 3.3 months HR: 0.98 <i>p</i> value: 0.42	21% vs. 11%	IHC Dako 22C3	PD-L1 combined positive score (CPS; the percentage of PD-L1-expressing tumor and infiltrating immune cells relative to the total number of tumor cells)	Treatment response was similar in patients with a CPS of 10% or more.	[25]
Advanced mUC (IMvigor211)	931	Second-line Atezolizumab vs. chemotherapy (Phase III)	11.1 vs. 10.6 months HR:0.87 <i>p</i> value: 0.41	2.4 vs. 4.2 months HR: 1.01	23% vs. 22%	IHC Ventana SP142	PD-L1 expression on <1% [IC0], 1% to <5% [IC1], and ³ 5% of tumor-infiltrating immune cells [IC2/3]	Atezolizumab was not associated with longer OS than chemotherapy in patients with IC2/3.	[63]

Table 2. Key trials of immune checkpoint inhibitors and association between treatment response and maker assessment in advanced urogenital malignancies.

Tunos of Tumor	No. of	True true and (Discov)	Out	comes and Respo	onse	Access		Clinical Relevance of	
Types of Tumor	Patients	Treatment (Phase)	OS	PFS	ORR	- Assay	Marker Assessment	Maker Assessment	Keference No.
Renal ce	ell carcinoma	(RCC)							
Metastatic RCC (BTCRC-GU14-003)	61	Second or Third-line pembrolizumab plus bevacizumab (Phase Ib/II)	NA at the median follow-up of 28.3 months	20.7 months	60.90%	IHC Dako 22C3	PD-L1 expression in tumor cells	Patients with tumors overexpressing PD-L1 > 0 showed a trend toward better PFS after 20 months, but there was no statistical difference in overall PFS.	[64]
Metastatic RCC	30	Second or Third-line lenvatinib plus pembrolizumab (Phase Ib/II)	NA	19.8 months	70%	NA	NA	NA	[65]
Advanced RCC (KEYNOTE-426)	861	First-line pembrolizumab plus axitinib vs. sunitinib (Phase III)	HR: 0.53 <i>p</i> value: <0.0001	15.1 vs. 11.1 months HR = 0.69 <i>p</i> value: <0.001	59.7% vs. 35.7%	IHCDako 22C3	PD-L1 combined positive score (the percentage of PD-L1+ tumor and infiltrating immune cells/the total tumor cells)	The benefit of pembrolizumab plus axitinib was observed in patients with tumors expressing PD-L1 expression and those with tumors without PD-L1 expression.	[66]
Advanced RCC (JAVELIN Renal 101)	886	First-line avelumab plus axitinib vs. sunitinib (Phase III)	12.0 and 11.5 months HR: 0.78 p value: 0.14	13.8 vs. 8.4 months HR = 0.69 <i>p</i> value: <0.001	51.4% vs. 25.7%	IHC Ventana SP263	PD-L1 expression in tumor cells	ORR among patients with PD-L1–positive tumors who received avelumab plus axitinib was twice as that in patients who received sunitinib (55.2% vs. 25.5%, respectively).	[67]
Advanced or Metastatic RCC (CheckMate 214)	1096	First-line nivolumab plus ipilimumab vs. sunitinib (Phase III)	NR and 26.6 months HR: 0.66 <i>p</i> value: <0.0001	8.2 vs. 8.3 months HR = 0.77 <i>p</i> value: 0.0014	42% vs. 29%	IHC Dako 28-8	PD-L1 expression in tumor cells	Partial responders and complete responders to nivolumab plus ipilimumab both had higher baseline tumor PD-L1 expression than that in non-responders.	[68]

Marker Assessment	Clinical Relevance of Maker Assessment	Reference No.
	In the PD-L1 positive population, the median	

	-	-
Tabla	<u>_</u>	Cont
Idule	Z .	Com.

Tunes of Tumor	No. of	Treatment (Phase)	Outcomes and Response			- Accaw	Markar Assassment	Clinical Relevance of	Potoron co No
Types of Tumor	Patients		OS	PFS	ORR	ASSay	Warker Assessment	Maker Assessment	Kererence No.
Advanced or Metastatic RCC (IMmotion151 trial)	915	First-line atezolizumab plus bevacizumab vs. sunitinib (Phase III)	33.6 and 34.9 months HR = 0.93 <i>p</i> value: 0.48	11.2 and 8.4 months HR = 0.83 <i>p</i> value: 0.022	37% vs. 33%	IHC Ventana SP142	PD-L1 expression in tumor cells	In the PD-L1 positive population, the median progression-free survival in the atezolizumab plus bevacizumab group was significantly longer than that in the sunitinib group. PD-L1 can be used as a supporting tool for treatment selection.	[69]
Advanced or Metastatic RCC (CheckMate 025)	821	Second or Third-line Nivolumab vs. everolimus (Phase III)	25.0 and 19.6 months HR: 0.73 <i>p</i> value: 0.002	4.6 vs. 4.4 months HR = 0.88 <i>p</i> value: 0.11	25% vs. 5%	IHC Dako 28-8	PD-L1 expression in tumor cells	Higher levels of PD-L1 expression are associated with poorer survival, while it does not support PD-L1 as a marker of treatment benefit.	[70]
Pros	state cancer (PCa)/castration resistant	PCa (CRPC)						
Metastatic CRPCKEYNOTE-199	258	Pembrolizumab after docetaxel or ARATs	9.5, 7.9, and 14.1 months in cohort 1 (PD-L1 positive), cohort 2 (PD-L1 negative), and cohort 3 (Bone- predominant)	2.1, 2.1, and 3.7 months in cohort 1 (PD-L1 positive), cohort 2 (PD-L1 negative), and cohort 3 (Bone- predominant)	7% and 2 % in cohort 1 (PD-L1 positive) and cohort 2 (PD-L1 negative)	IHC Dako 22C3	PD-L1 expression and aberrations of homologous recombination repair (HRR) gene in tumor cells	There were no significant differences in the response to pemblolizumab between the PD-L1-positive and -negative groups.	[71]
Metastatic CRPC	28	Pemblolizumab and enzalutamide	22.2 months	3.7 months (PSA-PFS)	18%	IHC and FCM	PD-L1 expression in tumor cells	The frequency of granzyme B+ CD8+ and perforin+CD8+ T cells were higher in responders those that in non-responders.	[72]
Retroperit	oneal sarcom	na (RSar)							
STS and BS (SARC028)	40 and 40	Pembrolizumab (phase II)	12.3 months (95% CI, 8.5–18.3)	4.2mounths (95% CI, 2.0–5.3)	NA	IHC Dako 22C3	Score was expressed as percentage of tumour cells positive for PD-L1. A tumour was considered positive for PD-L1 expression if more than 1% of its cells showed membranous staining.	PD-L1 expression was observed in only 5% of samples; both were UPS and responded to therapy. Pembrolizumab showed encouraging activity in patients with UPS or DDLPS.	[73]

Trues of Trues	No. of	Transformers (Disc.)	Out	Outcomes and Response				Clinical Relevance of	
Types of Tumor	Patients	Treatment (Phase)	os	PFS	ORR	- Assay	Marker Assessment	Maker Assessment	Reference No.
Advanced or metastaic STS and BS (Alliance A091401)	43 and 42	Nivolumab vs. Nivolumab+ipilimmab (phase II)	10.7 and 14.3 months (95% CI, 5.5–15.4 and NA)	1.7 and 4.1 months (95% CI, 1.4–4.3 and 1.4–4.7)	NA	NA	NA	Treatment with nivolumab plus ipilimumab in an unselected cohort of heavily treated patients with advanced sarcoma, achieved a proportion of 16% of 38 patients with confirmed objective responses, which is similar to the results obtained with standard chemotherapy.	[74]
Locally advanced or metastaic sarcoma	20	T-VEC plus pemblolizumab (phase II)	18.7 months (95% CI, 12.3–NA)	4.3 months (95% CI, 3.2–NA)	30%	IHC	PD-L1 tumor membrane expression and CD3+/CD8+ TILs at the infiltrating edge of the tumor. The patients underwent pretreatment and posttreatment tumor biopsies.	The data show that 64% of the posttreatment tumors were PD-L1 positive and 55% of patients converted from PD-L1 negative to PD-L1 positive after treatment.	[75]
Advanced sarcomas including alveolar soft-part sarcoma	33	Axitinib plus pembrolizumab (phase II)	18.7 months (95% CI, 12.0–NA)	4.7 months (95% CI, 3.0–9.4)	25%	IHC	PD-L1 expression in sarcoma cells	PD-L1 expression was positive in 52% of patients with evaluable tumor biopsy samples. Neither PD-L1 positivity nor increased TIL score correlated with progression-free survival of longer than 6 months or with achieving a partial response.	[76]

Table 2. Cont.

OS, overall survival; PFS, progression-free survival; ORR, objective response rate; UC, urothelial carcinoma; HR, hazard risk; IHC, immunohistochemical staining; PD-L1, programmed cell death ligand-1; CPS, combined positive score; RCC, renal cell carcinoma; NR, not reached; RSar, retroperitoneal sarcoma; STS, soft-tissue sarcoma; BS, bone sarcoma; CI, confidence interval; NA, not available; FCM, flow cytometry; Treg, regulatory T cell; NLR, neutrophil-to-leukocyte ration; NA, not available; TIL, tumor-infiltrating leukocytes.

3.4. The Immunological Profile in the Tumor Microenvironment of UC Arises from Different Primary Origins

Bladder, renal pelvis, and ureter are three common primary origins of UC. Although bladder UC, renal pelvic UC, and ureteral UC have many similarities, there are anatomical, biological, and molecular differences that should be considered as three distinct urothelium-derived malignancies [77]. Yates et al. reported significant differences in the genetic (microsatellite instability) and epigenetic (hypermethylation) backgrounds of bladder UC and UTUC [78]. Moreover, we have previously demonstrated that the subsequent NMIBC after radical nephroureterectomy for primary UTUC had a poorer prognosis after intravesical BCG compared to that in primary NMIBC [79]. This finding suggested that the primary origin is associated with an inherently poor response to BCG. However, all clinical trials of ICIs for metastatic UC clustered with UC diseases arising from different origins. Figure 2 shows representative images of IHC staining of seven immune-related markers using surgically resected UC specimens. Our previous works have presented the oncogenic or anti-tumoral effect of endogenous and exogenous colony-stimulating factors (CSFs) in UC [80-82]. In addition, accumulating evidence revealed that CD204, but not CD163, positive tumor-associated macrophages are associated with the aggressive behavior of various cancers including UC [16,83]. Thus, we included three CSFs and CD204 in the IHC analysis. The patients' background and primary/secondary antibodies used in this study are depicted in Tables S1 and S2. Significant differences in the density of FOXP3-positive TILs, CD204-positive tumor-infiltrating macrophages, PD-L1-positive cells, granulocyte-macrophage colony-stimulating factor (GM-CSF), and macrophage colony-stimulating factor (M-CSF) were observed in the quantitative analysis. In the immunotherapy era, we may need to consider differences in the immunological profile among the disparate triplets.



Figure 2. Difference in tumor immune microenvironments in urothelial carcinoma: comparison of primary tumor origins. The left panels are representative images of immunohistochemical staining of seven immune-related markers using surgically resected formalin-fixed paraffin-embedded specimens. Data were expressed by box-and-whisker plots, in which the outliers are indicated as dots, and compared using the Kruskal-Wallis (KW) test, followed by the post hoc test (Dunn test). * p < 0.05, *** p < 0.01, n.s., not significant. Abbreviations: UC, urothelial carcinoma; TIL, tumor-infiltrating leukocyte; HPF, high power field (magnification, ×400); PD-L1, programmed death-ligand 1; G-CSF, granulocyte colony-stimulating factor; GM-CSF, granulocyte-macrophage colony-stimulating factor; M-CSF, macrophage colony-stimulating factor.

4. Renal Cell Carcinoma (RCC)

4.1. Epidemiology and Current Issues of RCC

RCC accounts for approximately 2.2% of all cancers and is estimated to cause over 170,000 annual deaths globally [84]. RCC has been regarded as an immunogenic tumor, known as "hot tumor", and is thought to weaken host immunity in order to enhance tumor growth. This feature has encouraged urologists to use immunotherapies including interleukin-2, interferon-alpha, and ICIs [81,82]. In contrast, approximately 30% of patients with RCC present with metastases, and recurrent disease develops in approximately 40% of patients previously treated for localized RCC [83]. Recently, the development of ICIs has significantly improved prognosis in advanced or metastatic RCC [64–66,84–87]. As there are still a large number of patients with advanced RCC who fail in these treatments, there is an urgent clinical need to identify predictive markers to improve treatment efficacy.

4.2. Clinical Impact of TILs in Patients with RCC

Table 1 summarizes previous studies addressing the clinical relevance of TILs in patients with RCC [39–49]. Most of these studies were intended for advanced or metastatic RCC and several types of treatment settings, including radical nephrectomy and systemic treatment with molecular targeted drugs. The clinical significance of TILs in patients with RCC has been reported over the past three decades.

In 1992, Igarashi et al. showed that CD8+/CD11- cells, which inhibit antibody production, increased and CD4+/CD45RA- cells, which introduced antibody-producing cells, decreased along with tumor progression [39]. Subsequent studies using IHC and flow cytometry analyses revealed that the complex interactions between tumor cells and TILs such as cytotoxic T cells, Tregs, exhausted T cells, and B cells were associated with the acceleration or suppression of tumor growth, thereby affecting the prognosis of RCC patients [40-48]. With regard to Tregs, migration of CD4+Foxp3+ T cells to the tumor microenvironment is an independent factor for poor prognosis [43–45]. However, Siddiqui et al. have shown that CD4+CD25+FoxP3+ T cells are not associated with prognosis, while an increase in CD4+CD25+FoxP3- T cells was significantly associated with poor prognosis [46]. Similar to Tregs, there are several subtypes of CD8+ TILs. An increase in the number of CD8+ TILs in the tumor microenvironment was associated with prolonged prognosis in RCC [41,43,49], while the increase in exhausted T cells, which belong to CD8+ T cells, was associated with poor prognosis in RCC [47,48]. There are various subtypes of CD4+ T cells and CD8+ T cells, and the difference in one cell-surface marker can be a switch for a completely opposite function in tumor progression. These complex mechanisms make it difficult to understand the various roles of each immune cell type. In contrast, ICIs use these complex mechanisms and enable the switch to escape from antitumor suppression. Overall, the clinical significance of TILs has not yet been fully elucidated. However, TILs may have the potential to improve the clinical outcome in patients treated with molecular targeted drugs and ICIs.

4.3. Predictive Biomarkers for Response to ICIs in RCC

As of August 2020, five PD-1/PD-L1/CTLA-4 inhibitors have been approved for the treatment of locally advanced or metastatic RCC. Table 2 summarizes key clinical trials (limited to phase II or III) of ICIs for advanced/metastatic RCC [64–70]. These clinical trials showed that ICIs were superior to conventional molecular targeted drugs such as tyrosine kinase inhibitors (TKIs) and mammalian target of rapamycin inhibitors (mTORis). Moreover, most of these clinical trials evaluated the efficacy of combination therapy with one ICI plus one targeted drug compared to that of TKI or mTORi, which was different from the one used in the trial arm. Combination therapies have shown a higher objective response rate (ORR). Careful interpretation of outcomes such as progression-free survival (PFS) and overall survival (OS) is needed. Additionally, most of these clinical trials have investigated the association between PD-L1 expression in tumor cells and outcomes. The CheckMate 025, KEYNOTE-426, JAVELIN Renal 101, CheckMate 214, BTCRC-GU14-003, and IMmotion151 trials investigated the

association between PD-L1 expression in tumor cells and prognosis as a subgroup analysis, and these trials showed that high PD-L1 expression was associated with a treatment benefit for ICIs [64,66–70]. Interestingly, CheckMate 214 revealed that high expression of PD-L1 had a high rate of mortality in patients who received sunitinib [68]. This indicates the potential benefit of sequential treatment with ICIs and TKI/mTORis, which could create a favorable immune microenvironment. In contrast, there has been limited evidence for predictive biomarkers for ICI responses. One major limitation is that PD-L1 expression in tumor cells is the only immune checkpoint protein investigated in clinical trials. Thus, further research is necessary to establish a novel biomarker-driven immunotherapy strategy.

5. Prostate Cancer (PCa)

5.1. Epidemiology and Current Issues of PCa

PCa is the most common malignancy in men. The estimated lifetime risk of prostate cancer diagnosis is 13%, with a mortality to incidence ratio of 20% [88,89]. Prostatectomy and radiotherapy are standard curative treatments for localized PCa. Moreover, androgen deprivation therapy (ADT) is the standard primary therapy for metastatic PCa, because it is initially androgen-dependent and has a good response to ADT. However, some aggressive subsets progress to an androgen independent state, resulting in castration-resistant PCa (CRPC).

Various treatments, including androgen receptor-axis-targeted (ARAT) agents such as abiraterone [90] and enzalutamide [91], chemotherapy such as docetaxel [71] and cabazitaxel [72], a cellular vaccine called sipuleucel-T [92], and a radiopharmaceutical radium-223 [93] are available for patients with metastatic CRPC. Although the latest ICIs have shown strong antitumor activity in many tumors types, ICIs for advanced PCa and metastatic CRPC remain challenging [94–97].

5.2. Clinical Impact of TILs in Patients with PCa

As TILs are not abundant in the microenvironment of primary PCa [98,99] and PD-L1 expression in PCa cells is scarce [100], PCa is thought to be an immunologically "cold tumor". This feature may limit the success of ICI trials for advanced PCa and metastatic CRPC. However, two studies demonstrated that sufficient TILs were detected in post-ADT tumors [50,101], suggesting that CRPC could be a potential target of ICIs. Sorrentino et al. demonstrated that ADT induced abundant T cell infiltration in both benign glands and tumor tissues in human prostates [101]. The authors also showed that an increased number of Foxp3+CD25+CD127- Tregs was detected after ADT [50]. Previous studies addressing the correlation between TILs and post-treatment survival in patients with PCa are listed in Table 1 [50–53]. A high CD8+/Foxp3+ ratio in prostate tissues treated with neoadjuvant ADT was identified as a good prognostic factor after prostatectomy [50]. On the other hand, Ness et al. demonstrated that high density of CD8+ lymphocyte infiltration, especially in the tumor epithelial area, was identified as an independent poor prognostic factor for biochemical-failure survival after radical prostatectomy [51], which differs from the data reported by Sorrentino et al. [50]. Nardone et al. revealed that increased expression of CD8+ and CCR7+ TILs is associated with longer PFS and OS, whereas increased PD-1 and Foxp3+ Treg expression was associated with longer PFS and OS in patients treated with salvage radiotherapy after radical prostatectomy [52]. Overall, there is a lack of sufficient evidence regarding the prognostic value of TILs in patients with prostate cancer.

5.3. Predictive Biomarkers for Response to ICIs in PCa

Unfortunately, there are no randomized control trials that demonstrated the benefit of ICIs for metastatic CRPC. Two studies analyzed biomarkers predicting the response to ICIs in patients with metastatic CRPC. The KEYNOTE-199 phase II study analyzed the response to pembrolizumab for 258 patients with metastatic CRPC who had previously treated with docetaxel or ARATs [71]. PSA response rates were 6% in cohort 1 (PD-L1-positive group), 8% in cohort 2 (PD-L1-negative group), and 2% in cohort 3 (bone-predominant disease, regardless of PD-L1 expression), respectively.

Median OS were 9.5 months in cohort 1, 7.9 months in cohort 2, and 14.1 months in cohort 3, respectively. Moreover, this study analyzed correlation between the status of the homologous recombination repair genes and the response to pembrolizumab. Response duration was 4.4 months in patients with an ATM mutation and more than 21.8 months in patients with a BRCA2 mutation. However, it is difficult to draw a conclusion regarding predictive biomarkers of response to ICIs because of the low number of responders in this study. Another single-arm phase II trial analyzed the efficacy of pembrolizumab plus enzalutamide for 28 patients with metastatic CRPC [53]. Five (18%) of 28 patients had PSA decline of 50 % or greater. For entire cohort, the median OS was 22 months. The median OS for 5 responders was 41.7 months. Thirteen-patients could be identified tumors by base line biopsy for metastatic lesions. In one responder and two non-responders, PD-L1 was detected on TILs. The frequency of granzyme B+ CD8+ T cell and perforin+ CD8+ T cell was higher in responder than in non-responder, suggesting that the clinical benefit of pembrolizumab plus enzalutamide might require the pre-exiting tumor-reactive CD8+ T cell. The frequency of PD-L1 expression in patients with metastatic CRPC was 31.6% in previously publication [102], which was higher than in this study. Based on these findings, there are a significant lack on evidence regarding correlation between TILs (i.e., PD-1 and PD-L1) and outcome/response to ICIs in patients with metastatic CRPC because of the nature of prostate cancer, considering scarce TILs and low PD-L1 expressions. Therefore, further studies were warranted to demonstrate the anti-tumor activity of immunotherapy including not only PD-1 inhibitor monotherapy but also combination-therapy with CTLA-4 inhibitor.

6. Retroperitoneal Sarcoma (RSar)

6.1. Epidemiology and Current Issues of RSar

Soft tissue sarcomas (STSs) are rare tumors with the incidence less than 1% of all adult solid malignancies [103]. STSs can occur in any anatomic region and those arising from the retroperitoneal cavity are classified "RSar" and a urological malignancy, accounting for only 12–15% of all STSs [104]. Extended surgical resection of primary tumor and the surrounding vital organs is the mainstay of the treatment of RSar [105]. Although perioperative radiotherapy and chemotherapy are available to improve the rate of complete resection, it is complicated to define the optimal treatment strategies due to the disease heterogeneity and variety of tumor subtype.

Recurrent, unresectable, and metastatic RSars are indicated for systemic therapy. Chemotherapeutic drugs such as doxorubicin plus dacarbazine, doxorubicin plus ifosfamide, or doxorubicin alone has been administered as the first-line treatment for advanced sarcomas [106]. Recently, accumulating evidences have demonstrated the potential benefit of immunotherapies, especially ICIs (Table 2) [73–76,107,108]. Because the response to chemotherapy and immunotherapy is not expected in all the patients and the predictive markers are not available, there are still many limitations in the clinical management of advanced RSar. Here, we summarize the potential of TILs and PD-L1 expression as a clinically available biomarker for RSar.

6.2. Clinical Impact of TILs in Patients with RSar

We previously investigated the clinical significance and prognostic implications of intratumoral PD-L1, PD-L2, PD-1, and Ki-67 expression in patients with RSar [54]. Among these markers, only high expression of PD-1 in the TILs was a possible predictor of postoperative recurrence. Interestingly, observation of several clinicopathological parameters showed that high levels of serum lactate dehydrogenase (LDH) were significantly correlated with high intratumoral expressions of PD-L1 and PD-L2 [54]. This novel finding implies that elevated levels of serum LDH might be associated with response to the treatment of PD-1/PD-L1 blockade. Tseng et al. analyzed the intratumoral adaptive immune response in well differentiated- and dedifferentiated-retroperitoneal liposarcomas using isolation of TILs from surgically resected tumors followed by flow cytometry [74,75]. Although the majority of TILs were CD4+ T cells, cytotoxic CD8+ T cells accounted for 20% of CD3+ T cells.

Notably, 65% of CD8+ T cells were positive for the PD-1. Immune cell aggregates evaluated by IHC was associated with worse prognosis in both well differentiated and dedifferentiated retroperitoneal liposarcoma, suggesting that an adaptive immune response was present in the liposarcomas but may be hindered by Immune cell aggregates among other possible microenvironmental factors [56]. Moreover, Yan et al. investigated the density of TILs in various types of retroperitoneal liposarcomas [57]. The proportion of TILs was the highest in the dedifferentiated retroperitoneal liposarcoma and the lowest in pleomorphic liposarcoma. The authors demonstrated that patients with higher FOXP3+ Treg or PD-1/PD-L1+ cells tended to be a poor prognostic factor. Heterogeneous TILs distribution was found in 50% patients and tended to correlate with favorable disease-free-survival [56]. In spite of the accumulating evidence, the clinical impact of TILs in RSar still remains uncertain, especially in other types of RSar such as leiomyosarcoma and undifferentiated pleomorphic sarcoma, formally known as malignant fibrous histiocytoma.

6.3. Predictive Biomarkers for Response to ICIs in RSar

Because RSar is an extremely rare tumor, no clinical trial for the treatment with ICIs has evaluated the clinical benefit only in advanced RSar. Majority of clinical trials included the STS and bone sarcoma (BS). The SARC028 phase II study was the first clinical trial evaluating ICIs for advanced sarcomas [73], in which 40 patients in each disease cohort were treated pembrolizumab. A tumor was considered positive for PD-L1 expression if more than 1% of its cells showed membranous staining. PD-L1 expression was observed in only 5% of patients; both were undifferentiated pleomorphic sarcoma and responded to pembrolizumab treatment. Another combination therapy with talimogene laherparepvec (T-VEC) plus pembrolizumab for locally advanced or metastatic sarcomas has shown promising results [75]. T-VEC is a biopharmaceutical drug and shows anti-tumor effect by increase in tumor-specific immune activation via augmenting antigen presentation and T-cell priming. Of all patients showing the treatment response, pre-treatment tumor sample had aggregates of CD3+/CD8+ TILs at the infiltrating tumor edge. However, only 1 patient demonstrated PD-L1 positive at baseline; this patient achieved a partial response. In contrast, 64% of the posttreatment tumor were PD-L1 positive and 55% of patients converted from PD-L1 negative to positive after treatment [75]. The phase II study of the combination of axitinib plus pembrolizumab for advanced or metastatic sarcomas was published in 2019 [76]. PD-L1 expression was positive in 52% of patients with evaluable tumor biopsy samples. However, neither PD-L1 positivity nor increased tumour-infiltrating lymphocyte score correlated with progression-free survival of longer than 6 months or achieving a partial response. Currently, there is no predictive biomarker available for the treatment of ICIs in advanced sarcomas including RSar.

7. Limitations and Current Perspective Regarding the Assessment of TILs

We focused exclusively on the potential of TILs as a prognostic or predictive marker. Relevant studies have been performed in a retrospective manner and in relatively small cohorts. The definitions of TILs, such as inclusion of intratumoral TILs and/or stromal TILs, and the scoring methodology varied among studies. These inconsistencies hinder comparisons across studies and extrapolation of findings to clinical practice. Large studies investigating the potential prognostic value of TILs as assessed on HE staining are lacking. International Immuno-Oncology Biomarker Working Group on Breast Cancer has developed the international guidelines regarding the assessement of TILs on HE-stained slides without any specific staining [109]. The purpose of this group is to develop standards on the assessment of immuno-oncology biomarkers to aid pathologists, clinicians and researchers in their research and daily clinical practice. International Guidelines on TIL-assessment in solid tumors Part 2 provided the recommendation in assessment of genitourinary carcinomas. According to the guidelines [109], separate reporting of intratumoral TILs and stromal TILs is recommended—this is based on the context of atezolizumab treatment in mUC, where the PD-L1 "immune cell" score is derived from the stromal TILs score [110]. In addition, special care should be taken to avoid areas of

tumor zones with necrosis, coagulation artifact, and previous biopsy sites, which is a common finding in resected specimens of bladder tumor. However, detailed tutorial for RCC, PCa, and sarcoma is not available because of insufficient data to make specific recommendations.

Early data showed the presence of TILs in UC was associated with a favorable prognosis [111]. As to early data of RCC, increased TILs, both CD4-positive and CD8-positive T cells, appear to related with high risk of post-nephrectomy recurrence and poor prognosis [112–116]. However, accumulating evidence, largely based on IHC quantification of different TIL subsets, have somehow turned conflicting results on the prognostic relevance of TILs (Table 1). Majority of reports on TILs in PCa have focused on the prognostic value of TILs, while few studies investigating the potential to predicting response to drug therapies. Most reports have shown the evidence for a relationship between the high TILs and increased risk of recurrence [117–119], metastasis [120], and poor cancer specific survival [121]. The result on the composition of TILs in are heterogeneous and sometimes conflicting, and the relationship between TILs and survival is still unclear in PCa [109]. Overall, we emphasis on the importance of uniform assessment of TILs and uniform comparison of study results in research practices.

8. Conclusions

As, an "immunotherapy tsunami", in particular ICIs, has overtaken the oncological field in this decade, it is mandatory for Physicians to deepen knowledge about cancer immunity and tumor immune microenvironment. This review highlights comprehensively the following two topics: (i) prognostic impact of TILs and (ii) predictive maker for ICIs in four urological solid tumors: UC, RCC, PCa, and Sar. Although there is accumulating evidence that the density of TILs can serve as a prognostic biomarker and/or predictive biomarker for immunotherapies, inconsistency of TIL evaluation and interpretation for the results seems to hinder its clinical application. Unfortunately, across different solid malignancies, the response rate and predictive markers for ICIs may vary significantly. Multiple biomarkers including tumor-infiltrating immune cells, PD-L1 expression, other immune checkpoint protein expression, mRNA gene expression analysis, mismatch-repair deficiency, and tumor mutational burden may need to overcome disease heterogeneity and complex tumor immunity. Both identification of positive or negative predictive biomarkers of ICIs and development promising combination are required urgently to refine the clinical management of advanced urological malignancies. Further studies with large-scale cohorts and long follow-up periods to prove the clinical impact of novel prognostic/predictive biomarkers, followed by their adoption in clinical practice.

Supplementary Materials: The following are available online at http://www.mdpi.com/2072-6694/12/11/3153/s1, Table S1: The patients' background used for immunohistochemical staining analysis, Table S2: Primary and secondary antibodies used for immunohistochemical staining analysis.

Author Contributions: Conceptualization, M.M.; Funding acquisition, M.M. and K.F.; Methodology, Y.T.; Writing—original draft preparation, M.M., S.H., T.O., Y.O., Y.N., and T.F.; Writing—review and editing, M.M. and K.F.; supervision, K.F. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by JSPS KAKENHI Grant Numbers 16K20159 (M.M.) and 26861290 (K.F.), and Fiscal Years. 2015–2016 Nara Medical University Grant-in-Aid for the Collaborative Research Projects (K.F. and M.M.).

Acknowledgments: The authors would like to thank Kenta Ohnishi, Kota Iida, and Sayuri Ohnish for technical assistance and materials used for experiments.

Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. Chen, D.S.; Mellman, I. Oncology meets immunology: The cancer-immunity cycle. *Immunity* **2013**, *39*, 1–10. [CrossRef] [PubMed]
- Chen, D.S.; Mellman, I. Elements of cancer immunity and the cancer-immune set point. *Nature* 2017, 541, 321–330. [CrossRef] [PubMed]

- 3. Hanahan, D.; Weinberg, R.A. Hallmarks of cancer: The next generation. *Cell* **2011**, *144*, 646–674. [CrossRef] [PubMed]
- Bruno, A.; Pagani, A.; Magnani, E.; Rossi, T.; Noonan, D.M.; Cantelmo, A.R.; Albini, A. Inflammatory angiogenesis and the tumor microenvironment as targets for cancer therapy and prevention. *Cancer Treat. Res.* 2014, 159, 401–426. [CrossRef] [PubMed]
- 5. Piotrowski, I.; Kulcenty, K.; Suchorska, W. Interplay between inflammation and cancer. *Rep. Pract. Oncol. Radiother.* **2020**, *25*, 422–427. [CrossRef]
- 6. Frost, F.G.; Cherukuri, P.F.; Milanovich, S.; Boerkoel, C.F. Pan-cancer RNA-seq data stratifies tumours by some hallmarks of cancer. *J. Cell. Mol. Med.* **2020**, *24*, 418–430. [CrossRef]
- 7. Grivennikov, S.I.; Greten, F.R.; Karin, M. Immunity, inflammation, and cancer. *Cell* **2010**, *140*, 883–899. [CrossRef] [PubMed]
- 8. Qian, B.Z.; Pollard, J.W. Macrophage diversity enhances tumor progression and metastasis. *Cell* **2010**, 141, 39–51. [CrossRef]
- 9. Shembrey, C.; Huntington, N.D.; Hollande, F. Impact of Tumor and Immunological Heterogeneity on the Anti-Cancer Immune Response. *Cancers* **2019**, *11*, 1217. [CrossRef]
- Bindea, G.; Mlecnik, B.; Tosolini, M.; Kirilovsky, A.; Waldner, M.; Obenauf, A.C.; Angell, H.; Fredriksen, T.; Lafontaine, L.; Berger, A.; et al. Spatiotemporal dynamics of intratumoral immune cells reveal the immune landscape in human cancer. *Immunity* 2013, *39*, 782–795. [CrossRef]
- 11. Lakshmi Narendra, B.; Eshvendar Reddy, K.; Shantikumar, S.; Ramakrishna, S. Immune system: A double-edged sword in cancer. *Inflamm. Res.* **2013**, *62*, 823–834. [CrossRef] [PubMed]
- Bassani, B.; Baci, D.; Gallazzi, M.; Poggi, A.; Bruno, A.; Mortara, L. Natural Killer Cells as Key Players of Tumor Progression and Angiogenesis: Old and Novel Tools to Divert Their Pro-Tumor Activities into Potent Anti-Tumor Effects. *Cancers* 2019, *11*, 461. [CrossRef] [PubMed]
- Gentles, A.J.; Newman, A.M.; Liu, C.L.; Bratman, S.V.; Feng, W.; Kim, D.; Nair, V.S.; Xu, Y.; Khuong, A.; Hoang, C.D.; et al. The prognostic landscape of genes and infiltrating immune cells across human cancers. *Nat. Med.* 2015, 21, 938–945. [CrossRef] [PubMed]
- 14. Doucette, T.; Rao, G.; Rao, A.; Shen, L.; Aldape, K.; Wei, J.; Dziurzynski, K.; Gilbert, M.; Heimberger, A.B. Immune Heterogeneity of Glioblastoma Subtypes: Extrapolation from the Cancer Genome Atlas. *Cancer Immunol. Res.* **2013**, *1*, 112–122. [CrossRef]
- Reuben, A.; Gittelman, R.; Gao, J.; Zhang, J.; Yusko, E.C.; Wu, C.J.; Emerson, R.; Zhang, J.; Tipton, C.; Li, J.; et al. TCR Repertoire Intratumor Heterogeneity in Localized Lung Adenocarcinomas: An Association with Predicted Neoantigen Heterogeneity and Postsurgical Recurrence. *Cancer Discov.* 2017, 7, 12–17. [CrossRef]
- Miyake, M.; Hori, S.; Morizawa, Y.; Tatsumi, Y.; Nakai, Y.; Anai, S.; Torimoto, K.; Aoki, K.; Tanaka, N.; Shimada, K.; et al. CXCL1-Mediated Interaction of Cancer Cells with Tumor-Associated Macrophages and Cancer-Associated Fibroblasts Promotes Tumor Progression in Human Bladder Cancer. *Neoplasia* 2016, 18, 636–646. [CrossRef]
- 17. Rodallec, A.; Sicard, G.; Fanciullino, R.; Benzekry, S.; Lacarelle, B.; Milano, G.; Ciccolini, J. Turning cold tumors into hot tumors: Harnessing the potential of tumor immunity using nanoparticles. *Expert Opin. Drug Metab. Toxicol.* **2018**, *14*, 1139–1147. [CrossRef]
- Kawashima, A.; Kanazawa, T.; Goto, K.; Matsumoto, M.; Morimoto-Okazawa, A.; Iwahori, K.; Ujike, T.; Nagahara, A.; Fujita, K.; Uemura, M.; et al. Immunological classification of renal cell carcinoma patients based on phenotypic analysis of immune check-point molecules. *Cancer Immunol. Immunother.* 2018, 67, 113–125. [CrossRef]
- 19. Vareki, S.M. High and low mutational burden tumors versus immunologically hot and cold tumors and response to immune checkpoint inhibitors. *J. Immunother. Cancer* **2018**, *6*, 157. [CrossRef]
- 20. Strasner, A.; Karin, M. Immune infiltration and prostate Cancer. Front. Oncol. 2015, 5, 128. [CrossRef]
- Camus, M.; Tosolini, M.; Mlecnik, B.; Pagè, F.; Kirilovsky, A.; Berger, A.; Costes, A.; Bindea, G.; Charoentong, P.; Bruneval, P.; et al. Coordination of Intratumoral Immune Reaction and Human Colorectal Cancer Recurrence. *Cancer Res.* 2009, 69, 2685–2693. [CrossRef] [PubMed]
- 22. Prendergast, G.C.; Mondal, A.; Dey, S.; Laury-Kleintop, L.D.; Muller, A.J. Inflammatory Reprogramming with IDO1 Inhibitors: Turning Immunologically Unresponsive 'Cold' Tumors 'Hot'. *Trends Cancer* **2018**, *4*, 38–58. [CrossRef] [PubMed]

- 23. Galluzzi, L.; Buqué, A.; Kepp, O.; Zitvogel, L.; Kroemer, G. Immunological Effects of Conventional Chemotherapy and Targeted Anticancer Agents. *Cancer Cell* **2015**, *28*, 690–714. [CrossRef] [PubMed]
- 24. Miyake, M.; Owari, T.; Hori, S.; Nakai, Y.; Fujimoto, K. Emerging biomarkers for the diagnosis and monitoring of urothelial carcinoma. *Res. Rep. Urol.* **2018**, *10*, 251–261. [CrossRef]
- Fradet, Y.; Bellmunt, J.; Vaughn, D.J.; Lee, J.L.; Fong, L.; Vogelzang, N.J.; Climent, M.A.; Petrylak, D.P.; Choueiri, T.K.; Necchi, A.; et al. Randomized phase III KEYNOTE-045 trial of pembrolizumab versus paclitaxel, docetaxel, or vinflunine in recurrent advanced urothelial cancer: Results of >2 years of follow-up. *Ann. Oncol.* 2019, 30, 970–976. [CrossRef] [PubMed]
- 26. Miyake, M.; Tatsumi, Y.; Gotoh, D.; Ohnishi, S.; Owari, T.; Iida, K.; Ohnishi, K.; Hori, S.; Morizawa, Y.; Itami, Y.; et al. Regulatory T Cells and Tumor-Associated Macrophages in the Tumor Microenvironment in Non-Muscle Invasive Bladder Cancer Treated with Intravesical Bacille Calmette-Guérin: A Long-Term Follow-Up Study of a Japanese Cohort. *Int. J. Mol. Sci.* 2017, *18*, 2186. [CrossRef]
- Krpina, K.; Babarović, E.; Jonjić, N. Correlation of tumor-infiltrating lymphocytes with bladder cancer recurrence in patients with solitary low-grade urothelial carcinoma. *Virchows Arch.* 2015, 467, 443–448. [CrossRef]
- Zhang, Q.; Hao, C.; Cheng, G.; Wang, L.; Wang, X.; Li, C.; Qiu, J.; Ding, K. High CD4⁺ T cell density is associated with poor prognosis in patients with non-muscle-invasive bladder cancer. *Int. J. Clin. Exp. Pathol.* 2015, *8*, 11510–11516.
- 29. Liu, K.; Zhao, K.; Wang, L.; Sun, E. The prognostic values of tumor-infiltrating neutrophils, lymphocytes and neutrophil/lymphocyte rates in bladder urothelial cancer. *Pathol. Res. Pract.* **2018**, 214, 1074–1080. [CrossRef]
- Yu, A.; Mansure, J.J.; Solanki, S.; Siemens, D.R.; Koti, M.; Dias, A.B.T.; Burnier, M.M.; Brimo, F.; Kassouf, W. Presence of lymphocytic infiltrate cytotoxic T lymphocyte CD3⁺, CD8⁺, and immunoscore as prognostic marker in patients after radical cystectomy. *PLoS ONE* 2018, *13*, e0205746. [CrossRef]
- 31. Shi, M.J.; Meng, X.Y.; Wu, Q.J.; Zhou, X.H. High CD3D/CD4 ratio predicts better survival in muscle-invasive bladder cancer. *Cancer Manag. Res.* 2019, *11*, 2987–2995. [CrossRef] [PubMed]
- 32. Wahlin, S.; Nodin, B.; Leandersson, K.; Boman, K.; Jirström, K. Clinical impact of T cells, B cells and the PD-1/PD-L1 pathway in muscle invasive bladder cancer: A comparative study of transurethral resection and cystectomy specimens. *Oncoimmunology* **2019**, *8*, e1644108. [CrossRef] [PubMed]
- Kawashima, A.; Kanazawa, T.; Jingushi, K.; Kato, T.; Ujike, T.; Nagahara, A.; Fujita, K.; Morimoto-Okazawa, A.; Iwahori, K.; Uemura, M.; et al. Phenotypic Analysis of Tumor Tissue-Infiltrating Lymphocytes in Tumor Microenvironment of Bladder Cancer and Upper Urinary Tract Carcinoma. *Clin. Genitourin. Cancer* 2019, 17, 114–124. [CrossRef]
- Zhang, B.; Yu, W.; Feng, X.; Zhao, Z.; Fan, Y.; Meng, Y.; Hu, S.; Cui, Y.; He, Q.; Zhang, H.; et al. Prognostic significance of PD-L1 expression on tumor cells and tumor-infiltrating mononuclear cells in upper tract urothelial carcinoma. *Med. Oncol.* 2017, 34, 94. [CrossRef] [PubMed]
- Krabbe, L.M.; Heitplatz, B.; Preuss, S.; Hutchinson, R.C.; Woldu, S.L.; Singla, N.; Boegemann, M.; Wood, C.G.; Karam, J.A.; Weizer, A.Z.; et al. Prognostic Value of PD-1 and PD-L1 Expression in Patients with High Grade Upper Tract Urothelial Carcinoma. *J. Urol.* 2017, *198*, 1253–1262. [CrossRef] [PubMed]
- Wang, L.A.; Yang, B.; Rao, W.; Xiao, H.; Wang, D.; Jiang, J. The correlation of BER protein, IRF3 with CD8+ T cell and their prognostic significance in upper tract urothelial carcinoma. *Onco. Targets Ther.* 2019, 12, 7725–7735. [CrossRef] [PubMed]
- 37. Huang, H.S.; Su, H.Y.; Li, P.H.; Chiang, P.H.; Huang, C.H.; Chen, C.H.; Hsieh, M.C. Prognostic impact of tumor infiltrating lymphocytes on patients with metastatic urothelial carcinoma receiving platinum based chemotherapy. *Sci. Rep.* **2018**, *8*, 7485. [CrossRef]
- Pichler, R.; Fritz, J.; Zavadil, C.; Schäfer, G.; Culig, Z.; Brunner, A. Tumor-infiltrating immune cell subpopulations influence the oncologic outcome after intravesical Bacillus Calmette-Guérin therapy in bladder cancer. *Oncotarget* 2016, 7, 39916–39930. [CrossRef]
- 39. Igarashi, T.; Murakami, S.; Takahashi, H.; Matsuzaki, O.; Shimazaki, J. Changes on distribution of CD4+/CD45RA- and CD8+/CD11- cells in tumor-infiltrating lymphocytes of renal cell carcinoma associated with tumor progression. *Eur. Urol.* **1992**, *22*, 323–328. [CrossRef]
- 40. Huang, Y.; Wang, J.; Jia, P.; Li, X.; Pei, G.; Wang, C.; Fang, X.; Zhao, Z.; Cai, Z.; Yi, X.; et al. Clonal architectures predict clinical outcome in clear cell renal cell carcinoma. *Nat. Commun.* **2019**, *10*, 1245. [CrossRef]

- 41. Zhang, S.; Zhang, E.; Long, J.; Hu, Z.; Peng, J.; Liu, L.; Tang, F.; Li, L.; Ouyang, Y.; Zeng, Z. Immune infiltration in renal cell carcinoma. *Cancer Sci.* **2019**, *110*, 1564–1572. [CrossRef] [PubMed]
- Pignon, J.C.; Jegede, O.; Shukla, S.A.; Braun, D.A.; Horak, C.E.; Wind-Rotolo, M.; Ishii, Y.; Catalano, P.J.; Grosha, J.; Flaifel, A.; et al. irRECIST for the Evaluation of Candidate Biomarkers of Response to Nivolumab in Metastatic Clear Cell Renal Cell Carcinoma: Analysis of a Phase II Prospective Clinical Trial. *Clin. Cancer Res.* 2019, 25, 2174–2184. [CrossRef] [PubMed]
- Kang, M.J.; Kim, K.M.; Bae, J.S.; Park, H.S.; Lee, H.; Chung, M.J.; Moon, W.S.; Lee, D.G.; Jang, K.Y. Tumor-infiltrating PD1-Positive Lymphocytes and FoxP3-Positive Regulatory T Cells Predict Distant Metastatic Relapse and Survival of Clear Cell Renal Cell Carcinoma. *Transl. Oncol.* 2013, *6*, 282–289. [CrossRef] [PubMed]
- 44. Jensen, H.K.; Donskov, F.; Nordsmark, M.; Marcussen, N.; von der Maase, H. Increased intratumoral FOXP3-positive regulatory immune cells during interleukin-2 treatment in metastatic renal cell carcinoma. *Clin. Cancer Res.* **2009**, *15*, 1052–1058. [CrossRef]
- 45. Li, J.F.; Chu, Y.W.; Wang, G.M.; Zhu, T.Y.; Rong, R.M.; Hou, J.; Xu, M. The prognostic value of peritumoral regulatory T cells and its correlation with intratumoral cyclooxygenase-2 expression in clear cell renal cell carcinoma. *BJU Int.* **2009**, *103*, 399–405. [CrossRef]
- Siddiqui, S.A.; Frigola, X.; Bonne-Annee, S.; Mercader, M.; Kuntz, S.M.; Krambeck, A.E.; Sengupta, S.; Dong, H.; Cheville, J.C.; Lohse, C.M.; et al. Tumor-infiltrating Foxp3⁻CD4⁺CD25⁺ T cells predict poor survival in renal cell carcinoma. *Clin. Cancer Res.* 2007, *13*, 2075–2081. [CrossRef]
- 47. Kawashima, A.; Kanazawa, T.; Kidani, Y.; Yoshida, T.; Hirata, M.; Nishida, K.; Nojima, S.; Yamamoto, Y.; Kato, T.; Hatano, K.; et al. Tumour grade significantly correlates with total dysfunction of tumour tissue-infiltrating lymphocytes in renal cell carcinoma. *Sci. Rep.* **2020**, *10*, 6220. [CrossRef]
- 48. Giraldo, N.A.; Becht, E.; Vano, Y.; Petitprez, F.; Lacroix, L.; Validire, P.; Sanchez-Salas, R.; Ingels, A.; Oudard, S.; Moatti, A.; et al. Tumor-Infiltrating and Peripheral Blood T-cell Immunophenotypes Predict Early Relapse in Localized Clear Cell Renal Cell Carcinoma. *Clin. Cancer Res.* 2017, 23, 4416–4428. [CrossRef]
- 49. Yao, J.; Xi, W.; Zhu, Y.; Wang, H.; Hu, X.; Guo, J. Checkpoint molecule PD-1-assisted CD8⁺ T lymphocyte count in tumor microenvironment predicts overall survival of patients with metastatic renal cell carcinoma treated with tyrosine kinase inhibitors. *Cancer Manag. Res.* **2018**, *10*, 3419–3431. [CrossRef]
- 50. Sorrentino, C.; Musiani, P.; Pompa, P.; Cipollone, G.; Di Carlo, E. Androgen deprivation boosts prostatic infiltration of cytotoxic and regulatory T lymphocytes and has no effect on disease-free survival in prostate cancer patients. *Clin. Cancer Res.* **2011**, *17*, 1571–1581. [CrossRef]
- 51. Ness, N.; Andersen, S.; Valkov, A.; Nordby, Y.; Donnem, T.; Al-Saad, S.; Busund, L.T.; Bremnes, R.M.; Richardsen, E. Infiltration of CD8+ lymphocytes is an independent prognostic factor of biochemical failure-free survival in prostate cancer. *Prostate* **2014**, *74*, 1452–1461. [CrossRef]
- 52. Nardone, V.; Botta, C.; Caraglia, M.; Martino, E.C.; Ambrosio, M.R.; Carfagno, T.; Tini, P.; Semeraro, L.; Misso, G.; Grimaldi, A.; et al. Tumor infiltrating T lymphocytes expressing FoxP3, CCR7 or PD-1 predict the outcome of prostate cancer patients subjected to salvage radiotherapy after biochemical relapse. *Cancer Biol. Ther.* **2016**, *17*, 1213–1220. [CrossRef]
- 53. Watanabe, M.; Kanao, K.; Suzuki, S.; Muramatsu, H.; Morinaga, S.; Kajikawa, K.; Kobayashi, I.; Nishikawa, G.; Kato, Y.; Zennami, K.; et al. Increased infiltration of CCR4-positive regulatory T cells in prostate cancer tissue is associated with a poor prognosis. *Prostate* **2019**, *79*, 1658–1665. [CrossRef]
- 54. Makito, M.; Oda, Y.; Nishimura, N.; Morizawa, Y.; Ohnishi, S.; Hatakeyama, K.; Fujii, T.; Hori, S.; Gotoh, D.; Nakai, Y.; et al. Integrative assessment of clinicopathological parameters and the expression of PD-L1, PD-L2 and PD-1 in tumor cells of retroperitoneal sarcoma. *Oncol. Lett.* **2020**, *20*, 190. [CrossRef]
- 55. Tseng, W.W.; Demicco, E.G.; Lazar, A.J.; Lev, D.C.; Pollock, R.E. Lymphocyte composition and distribution in inflammatory, well-differentiated retroperitoneal liposarcoma: Clues to a potential adaptive immune response and therapeutic implications. *Am. J. Surg. Pathol.* **2012**, *36*, 941–944. [CrossRef]
- 56. Tseng, W.W.; Malu, S.; Zhang, M.; Chen, J.; Sim, G.C.; Wei, W.; Ingram, D.; Somaiah, N.; Lev, D.C.; Pollock, R.E.; et al. Analysis of the intratumoral adaptive immune response in well differentiated and dedifferentiated retroperitoneal liposarcoma. *Sarcoma* **2015**, *2015*, 547460. [CrossRef]
- Yan, L.; Wang, Z.; Cui, C.; Guan, X.; Dong, B.; Zhao, M.; Wu, J.; Tian, X.; Hao, C. Comprehensive immune characterization and T-cell receptor repertoire heterogeneity of retroperitoneal liposarcoma. *Cancer Sci.* 2019, 110, 3038–3048. [CrossRef]

- 58. Gevaert, T.; Cimadamore, A.; Montironi, R.; Eckstein, M. PD-L1 Testing for Urothelial Carcinoma: Interchangeability, Reliability and Future Perspectives. *Curr. Drug Targets* **2020**, *9*. [CrossRef]
- 59. Patel, M.R.; Ellerton, J.; Infante, J.R.; Agrawal, M.; Gordon, M.; Aljumaily, R.; Britten, C.D.; Dirix, L.; Lee, K.W.; Taylor, M.; et al. Avelumab in metastatic urothelial carcinoma after platinum failure (javelin Solid Tumor): Pooled results from two expansion cohorts of an open-label, phase 1 trial. *Lancet Oncol.* 2018, 19, 51–64. [CrossRef]
- 60. Sharma, P.; Siefker-Radtke, A.; de Braud, F.; Basso, U.; Calvo, E.; Bono, P.; Morse, M.A.; Ascierto, P.A.; Lopez-Martin, J.; Brossart, P.; et al. Nivolumab Alone and with Ipilimumab in Previously Treated Metastatic Urothelial Carcinoma: CheckMate 032 Nivolumab 1 mg/kg Plus Ipilimumab 3 mg/kg Expansion Cohort Results. *J. Clin. Oncol.* **2019**, *37*, 1608–1616. [CrossRef]
- Powles, T.; O'Donnell, P.H.; Massard, C.; Arkenau, H.T.; Friedlander, T.W.; Hoimes, C.J.; Lee, J.L.; Ong, M.; Sridhar, S.S.; Vogelzang, N.J.; et al. Efficacy and safety of durvalumab in locally advanced or metastatic urothelial carcinoma: Updated results from a phase 1/2 open-label study. *JAMA Oncol.* 2017, *3*, e172411. [CrossRef]
- 62. Sharma, P.; Retz, M.; Siefker-Radtke, A.; Baron, A.; Necchi, A.; Bedke, J.; Plimack, E.R.; Vaena, D.; Grimm, M.O.; Bracarda, S.; et al. Nivolumab in metastatic urothelial carcinoma after platinum therapy (CheckMate 275): A multicentre, single-arm, phase 2 trial. *Lancet Oncol.* **2017**, *18*, 312–322. [CrossRef]
- 63. Powles, T.; Durán, I.; van der Heijden, M.S.; Loriot, Y.; Vogelzang, N.J.; De Giorgi, U.; Oudard, S.; Retz, M.M.; Castellano, D.; Bamias, A.; et al. Atezolizumab versus chemotherapy in patients with platinum-treated locally advanced or metastatic urothelial carcinoma (IMvigor211): A multicentre, open-label, phase 3 randomised controlled trial. *Lancet* **2018**, *391*, 748–757. [CrossRef]
- 64. Dudek, A.Z.; Liu, L.C.; Gupta, S.; Logan, T.F.; Singer, E.A.; Joshi, M.; Zakharia, Y.N.; Lang, J.M.; Schwarz, J.K.; Al-Janadi, A.; et al. Phase Ib/II Clinical Trial of Pembrolizumab With Bevacizumab for Metastatic Renal Cell Carcinoma: BTCRC-GU14-003. *J. Clin. Oncol.* **2020**, *38*, 1138–1145. [CrossRef]
- 65. Taylor, M.H.; Lee, C.H.; Makker, V.; Rasco, D.; Dutcus, C.E.; Wu, J.; Stepan, D.E.; Shumaker, R.C.; Motzer, R.J. Phase IB/II Trial of Lenvatinib Plus Pembrolizumab in Patients With Advanced Renal Cell Carcinoma, Endometrial Cancer, and Other Selected Advanced Solid Tumors. J. Clin. Oncol. 2020, 38, 1154–1163. [CrossRef]
- Rini, B.I.; Plimack, E.R.; Stus, V.; Gafanov, R.; Hawkins, R.; Nosov, D.; Pouliot, F.; Alekseev, B.; Soulières, D.; Melichar, B.; et al. Pembrolizumab plus Axitinib versus Sunitinib for Advanced Renal-Cell Carcinoma. *N. Engl. J. Med.* 2019, *380*, 1116–1127. [CrossRef]
- Motzer, R.J.; Penkovm, K.; Haanen, J.; Rini, B.; Albiges, L.; Campbell, M.T.; Venugopal, B.; Kollmannsberger, C.; Negrier, S.; Uemura, M.; et al. Avelumab plus Axitinib versus Sunitinib for Advanced Renal-Cell Carcinoma. *N. Engl. J. Med.* 2019, *380*, 1103–1115. [CrossRef]
- 68. Motzer, R.J.; Rini, B.I.; McDermott, D.F.; Frontera, O.A.; Hammers, H.J.; Carducci, M.A.; Salman, P.; Escudier, B.; Beuselinck, B.; Amin, A.; et al. Nivolumab plus ipilimumab versus sunitinib in first-line treatment for advanced renal cell carcinoma: Extended follow-up of efficacy and safety results from a randomised, controlled, phase 3 trial. *Lancet Oncol.* **2019**, *20*, 1370–1385. [CrossRef]
- 69. Rini, B.I.; Powles, T.; Atkins, M.B.; Escudier, B.; McDermott, D.F.; Suarez, C.; Bracarda, S.; Stadler, W.M.; Donskov, F.; Lee, J.L.; et al. Atezolizumab plus bevacizumab versus sunitinib in patients with previously untreated metastatic renal cell carcinoma (IMmotion151): A multicentre, open-label, phase 3, randomised controlled trial. *Lancet* **2019**, *393*, 2404–2415. [CrossRef]
- Motzer, R.J.; Escudier, B.; McDermott, D.F.; George, S.; Hammers, H.J.; Srinivas, S.; Tykodi, S.S.; Sosman, J.A.; Procopio, G.; Plimack, E.R.; et al. Nivolumab versus Everolimus in Advanced Renal-Cell Carcinoma. *N. Engl. J. Med.* 2015, 373, 1803–1813. [CrossRef]
- McFalls, E.O.; Ward, H.B.; Moritz, T.E.; Goldman, S.; Krupski, W.C.; Littooy, F.; Pierpont, G.; Santilli, S.; Rapp, J.; Hattler, B.; et al. Coronary-artery revascularization before elective major vascular surgery. *N. Engl. J. Med.* 2004, 351, 2795–2804. [CrossRef] [PubMed]
- 72. De Bono, J.S.; Oudard, S.; Ozguroglu, M.; Hansen, S.; Machiels, J.P.; Kocak, I.; Gravis, G.; Bodrogi, I.; Mackenzie, M.J.; Shen, L.; et al. Prednisone plus cabazitaxel or mitoxantrone for metastatic castration-resistant prostate cancer progressing after docetaxel treatment: A randomised open-label trial. *Lancet* 2010, 376, 1147–1154. [CrossRef]

- 73. Tawbi, H.A.; Burgess, M.; Bolejack, V.; Van Tine, B.A.; Schuetze, S.M.; Hu, J.; D'Angelo, S.; Attia, S.; Riedel, R.F.; Priebat, D.A.; et al. Pembrolizumab in advanced soft-tissue sarcoma and bone sarcoma (SARC028): A multicentre, two-cohort, single-arm, open-label, phase 2 trial. *Lancet Oncol.* 2017, 18, 1493–1501. [CrossRef]
- 74. D'Angelo, S.P.; Mahoney, M.R.; Van Tine, B.A.; Atkins, J.; Milhem, M.M.; Jahagirdar, B.N.; Antonescu, C.R.; Horvath, E.; Tap, W.D.; Schwartz, G.K.; et al. Nivolumab with or without ipilimumab treatment for metastatic sarcoma (Alliance A091401): Two open-label, non-comparative, randomised, phase 2 trials. *Lancet Oncol.* 2018, 19, 416–426. [CrossRef]
- 75. Kelly, C.M.; Antonescu, C.R.; Bowler, T.; Munhoz, R.; Chi, P.; Dickson, M.A.; Gounder, M.M.; Keohan, M.L.; Movva, S.; Dholakia, R.; et al. Objective Response Rate Among Patients with Locally Advanced or Metastatic Sarcoma Treated With Talimogene Laherparepvec in Combination with Pembrolizumab: A Phase 2 Clinical Trial. *JAMA Oncol.* 2020, *6*, 402–408. [CrossRef] [PubMed]
- Wilky, B.A.; Trucco, M.M.; Subhawong, T.K.; Florou, V.; Park, W.; Kwon, D.; Wieder, E.D.; Kolonias, D.; Rosenberg, A.E.; Kerr, D.A.; et al. Axitinib plus pembrolizumab in patients with advanced sarcomas including alveolar soft-part sarcoma: A single-centre, single-arm, phase 2 trial. *Lancet Oncol.* 2019, 20, 837–848. [CrossRef]
- 77. Green, D.A.; Rink, M.; Xylinas, E.; Matin, S.F.; Stenzl, A.; Roupret, M.; Karakiewicz, P.I.; Scherr, D.S.; Shariat, S.F. Urothelial carcinoma of the bladder and the upper tract: Disparate twins. *J. Urol.* 2013, 189, 1214–1221. [CrossRef] [PubMed]
- Yates, D.R.; Catto, J.W. Distinct patterns and behaviour of urothelial carcinoma with respect to anatomical location: How molecular biomarkers can augment clinico-pathological predictors in upper urinary tract tumours. *World J. Urol.* 2013, 31, 21–29. [CrossRef]
- 79. Miyake, M.; Tatsumi, Y.; Matsumoto, H.; Nagao, K.; Matsuyama, H.; Inamoto, T.; Azuma, H.; Yasumoto, H.; Shiina, H.; Fujimoto, K. Nishinihon Uro-Oncology Collaborative Group. Outcomes of subsequent non-muscle-invasive bladder cancer treated with intravesical Bacillus Calmette-Guérin after radical nephroureterectomy for upper urinary tract urothelial carcinoma. *BJU Int.* 2018, 121, 764–773. [CrossRef]
- Morizawa, Y.; Miyake, M.; Shimada, K.; Hori, S.; Tatsumi, Y.; Nakai, Y.; Tanaka, N.; Fujii, T.; Fujimoto, K. Colony-stimulating factors detected in tumor cells and voided urine are potential prognostic markers for patients with muscle-invasive bladder cancer undergoing radical cystectomy. *Res. Rep. Urol.* 2018, 10, 103–111. [CrossRef]
- Hori, S.; Miyake, M.; Onishi, S.; Morizawa, Y.; Nakai, Y.; Tatsumi, Y.; Onishi, K.; Iida, K.; Gotoh, D.; Itami, Y.; et al. Evaluation of pro- and anti-tumor effects induced by three colony-stimulating factors, G-CSF, GM-CSF and M-CSF, in bladder cancer cells: Is G-CSF a friend of bladder cancer cells? *Int. J. Oncol.* 2019, 54, 2237–2249. [CrossRef]
- Hori, S.; Miyake, M.; Onishi, S.; Morizawa, Y.; Nakai, Y.; Tatsumi, Y.; Onishi, K.; Iida, K.; Gotoh, D.; Itami, Y.; et al. Supplementary granulocyte macrophage colony-stimulating factor to chemotherapy and programmed death-ligand 1 blockade decreases local recurrence after surgery in bladder cancer. *Cancer Sci.* 2019, *110*, 3315–3327. [CrossRef]
- 83. Sun, Y.; Xu, S. Tumor-Associated CD204-Positive Macrophage Is a Prognostic Marker in Clinical Stage I Lung Adenocarcinoma. *Biomed. Res. Int.* 2018, 2018, 8459193. [CrossRef]
- Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J. Clin.* 2018, 68, 394–424. [CrossRef]
- Motzer, R.J.; Hutson, T.E.; Tomczak, P.; Michaelson, M.D.; Bukowski, R.M.; Rixe, O.; Oudard, S.; Negrier, S.; Szczylik, C.; Kim, S.T.; et al. Sunitinib versus interferon alfa in metastatic renal-cell carcinoma. *N. Engl. J. Med.* 2007, 356, 115–124. [CrossRef]
- Rini, B.I.; Escudier, B.; Tomczak, P.; Kaprin, A.; Szczylik, C.; Hutson, T.E.; Michaelson, M.D.; Gorbunova, V.A.; Gore, M.E.; Rusakov, I.G.; et al. Comparative effectiveness of axitinib versus sorafenib in advanced renal cell carcinoma (AXIS): A randomised phase 3 trial. *Lancet* 2011, *378*, 1931–1939. [CrossRef]
- 87. Lam, J.S.; Leppert, J.T.; Belldegrun, A.S.; Figlin, R.A. Novel approaches in the therapy of metastatic renal cell carcinoma. *World J. Urol.* **2005**, *23*, 202–212. [CrossRef]
- 88. Barry, M.J.; Simmons, L.H. Prevention of Prostate Cancer Morbidity and Mortality: Primary Prevention and Early Detection. *Med. Clin. N. Am.* **2017**, *101*, 787–806. [CrossRef]

- Dy, G.W.; George, J.L.; Forouzanfer, M.H.; Naghavi, M.; Fitzmaurice, C. Global Burden of Urologic Cancers, 1990–2013. Eur. Urol. 2017, 71, 437–446. [CrossRef]
- De Bono, J.S.; Logothetis, C.J.; Molina, A.; Fizazi, K.; North, S.; Chu, L.; Chi, K.N.; Jones, R.J.; Goodman, O.B., Jr.; Saad, F.; et al. Abiraterone and increased survival in metastatic prostate cancer. *N. Engl. J. Med.* 2011, 364, 1995–2005. [CrossRef]
- 91. Beer, T.M.; Armstrong, A.J.; Rathkopf, D.E.; Loriot, Y.; Sternberg, C.N.; Higano, C.S.; Iversen, P.; Bhattacharya, S.; Carles, J.; Chowdhury, S.; et al. Enzalutamide in metastatic prostate cancer before chemotherapy. *N. Engl. J. Med.* **2014**, *371*, 424–433. [CrossRef] [PubMed]
- 92. Kantoff, P.W.; Higano, C.S.; Shore, N.D.; Berger, E.R.; Small, E.J.; Penson, D.F.; Redfern, C.H.; Ferrari, A.C.; Dreicer, R.; Sims, R.B.; et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N. Engl. J. Med.* 2010, 363, 411–422. [CrossRef]
- 93. Parker, C.; Nilsson, S.; Heinrich, D.; Helle, S.I.; O'Sullivan, J.M.; Fosså, S.D.; Chodacki, A.; Wiechno, P.; Logue, J.; Seke, M.; et al. Alpha emitter radium-223 and survival in metastatic prostate cancer. *N. Engl. J. Med.* 2013, 369, 213–223. [CrossRef]
- 94. Antonarakis, E.S.; Piulats, J.M.; Gross-Goupil, M.; Goh, J.; Ojamaa, K.; Hoimes, C.J.; Vaishampayan, U.; Berger, R.; Sezer, A.; Alanko, T.; et al. Pembrolizumab for Treatment-Refractory Metastatic Castration-Resistant Prostate Cancer: Multicohort, Open-Label Phase II KEYNOTE-199 Study. J. Clin. Oncol. 2020, 38, 395–405. [CrossRef]
- 95. Graff, J.N.; Beer, T.M.; Alumkal, J.J.; Slottke, R.E.; Redmond, W.L.; Thomas, G.V.; Thompson, R.F.; Wood, M.A.; Koguchi, Y.; Chen, Y.; et al. A phase II single-arm study of pembrolizumab with enzalutamide in men with metastatic castration-resistant prostate cancer progressing on enzalutamide alone. *J. Immunother. Cancer* **2020**, *8*, e000642. [CrossRef]
- 96. Hansen, A.R.; Massard, C.; Ott, P.A.; Haas, N.B.; Lopez, J.S.; Ejadi, S.; Wallmark, J.M.; Keam, B.; Delord, J.P.; Aggarwal, R.; et al. Pembrolizumab for advanced prostate adenocarcinoma: Findings of the KEYNOTE-028 study. Ann. Oncol. 2018, 29, 1807–1813. [CrossRef]
- 97. Kwon, E.D.; Drake, C.G.; Scher, H.I.; Fizazi, K.; Bossi, A.; van den Eertwegh, A.J.; Krainer, M.; Houede, N.; Santos, R.; Mahammedi, H.; et al. Ipilimumab versus placebo after radiotherapy in patients with metastatic castration-resistant prostate cancer that had progressed after docetaxel chemotherapy (CA184-043): A multicentre, randomised, double-blind, phase 3 trial. *Lancet Oncol.* 2014, *15*, 700–712. [CrossRef]
- Ebelt, K.; Babaryka, G.; Figel, A.M.; Pohla, H.; Buchner, A.; Stief, C.G.; Eisenmenger, W.; Kirchner, T.; Schendel, D.J.; Noessner, E. Dominance of CD4⁺ lymphocytic infiltrates with disturbed effector cell characteristics in the tumor microenvironment of prostate carcinoma. *Prostate* 2008, *68*, 1–10. [CrossRef]
- Yunger, S.; Bar El, A.; Zeltzer, L.A.; Fridman, E.; Raviv, G.; Laufer, M.; Schachter, J.; Markel, G.; Itzhaki, O.; Besser, M.J. Tumor-infiltrating lymphocytes from human prostate tumors reveal anti-tumor reactivity and potential for adoptive cell therapy. *Oncoimmunology* 2019, *8*, e1672494. [CrossRef]
- 100. Martin, A.M.; Nirschl, T.R.; Nirschl, C.J.; Francica, B.J.; Kochel, C.M.; van Bokhoven, A.; Meeker, A.K.; Lucia, M.S.; Anders, R.A.; DeMarzo, A.M.; et al. Paucity of PD-L1 expression in prostate cancer: Innate and adaptive immune resistance. *Prostate Cancer Prostatic Dis.* 2015, *18*, 325–332. [CrossRef]
- 101. Mercader, M.; Bodner, B.K.; Moser, M.T.; Kwon, P.S.; Park, E.S.; Manecke, R.G.; Ellis, T.M.; Wojcik, E.M.; Yang, D.; Flanigan, R.C.; et al. T cell infiltration of the prostate induced by androgen withdrawal in patients with prostate cancer. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 14565–14570. [CrossRef]
- 102. Haffner, M.C.; Guner, G.; Taheri, D.; Netto, G.J.; Palsgrove, D.N.; Zheng, Q.; Guedes, L.B.; Kim, K.; Tsai, H.; Esopi, D.M.; et al. Comprehensive Evaluation of Programmed Death-Ligand 1 Expression in Primary and Metastatic Prostate Cancer. *Am. J. Pathol.* **2018**, *188*, 1478–1485. [CrossRef]
- Burningham, Z.; Hashibe, M.; Spector, L.; Schiffman, J.D. The epidemiology of sarcoma. *Clin. Sarcoma Res.* 2012, 2, 14. [CrossRef] [PubMed]
- 104. Messiou, C.; Moskovic, E.; Vanel, D.; Morosi, C.; Benchimol, R.; Strauss, D.; Miah, A.; Douis, H.; van Houdt, W.; Bonvalot, S. Primary retroperitoneal soft tissue sarcoma: Imaging appearances, pitfalls and diagnostic algorithm. *Eur. J. Surg. Oncol.* 2017, 43, 1191–1198. [CrossRef]
- 105. Gronchi, A.; Strauss, D.C.; Miceli, R.; Bonvalot, S.; Swallow, C.J.; Hohenberger, P.; Van Coevorden, F.; Rutkowski, P.; Callegaro, D.; Hayes, A.J.; et al. Variability in patterns of recurrence after resection of primary retroperitoneal sarcoma (RPS): A report on 1007 patients from the multiinstitutional collaborative RPS Working Group. Ann. Surg. 2016, 263, 1002–1009. [CrossRef]

- 106. D'Ambrosio, L.; Touati, N.; Blay, J.Y.; Grignani, G.; Flippot, R.; Czarnecka, A.M.; Piperno-Neumann, S.; Martin-Broto, J.; Sanfilippo, R.; Katz, D.; et al. Doxorubicin plus dacarbazine, doxorubicin plus ifosfamide, or doxorubicin alone as a first-line treatment for advanced leiomyosarcoma: A propensity score matching analysis from the European Organization for Research and Treatment of Cancer Soft Tissue and Bone Sarcoma Group. *Cancer* 2020, *126*, 2637–2647. [CrossRef] [PubMed]
- 107. Mitsis, D.; Francescutti, V.; Skitzki, J. Current immunotherapies for sarcoma: Clinical trials and rationale. *Sarcoma* **2016**, *2016*, 9757219. [CrossRef] [PubMed]
- 108. Angelo, S.P.D. Manipulating the immune system with checkpoint inhibitors for patients with metastatic sarcoma. *Am. Soc. Clin. Oncol. Educ. Book* **2016**, *36*, e558–e564. [CrossRef]
- 109. Hendry, S.; Salgado, R.; Gevaert, T.; Russell, P.A.; John, T.; Thapa, B.; Christie, M.; Van De Vijver, K.; Estrada, M.V.; Gonzalez-Ericsson, P.I.; et al. Assessing Tumor-Infiltrating Lymphocytes in Solid Tumors: A Practical Review for Pathologists and Proposal for a Standardized Method from the International Immuno-Oncology Biomarkers Working Group: Part 2: TILs in Melanoma, Gastrointestinal Tract Carcinomas, Non-Small Cell Lung Carcinoma and Mesothelioma, Endometrial and Ovarian Carcinomas, Squamous Cell Carcinoma of the Head and Neck, Genitourinary Carcinomas, and Primary Brain Tumors. *Adv. Anat. Pathol.* 2017, 24, 311–335. [CrossRef]
- 110. Galsky, M.D.; Arija, J.Á.A.; Bamias, A.; Davis, I.D.; De Santis, M.; Kikuchi, E.; Garcia-Del-Muro, X.; De Giorgi, U.; Mencinger, M.; Izumi, K.; et al. Atezolizumab with or without chemotherapy in metastatic urothelial cancer (IMvigor130): A multicentre, randomised, placebo-controlled phase 3 trial. *Lancet* 2020, 395, 1547–1557. [CrossRef]
- 111. Sharma, P.; Shen, Y.; Wen, S.; Yamada, S.; Jungbluth, A.A.; Gnjatic, S.; Bajorin, D.F.; Reuter, V.E.; Herr, H.; Old, L.J.; et al. CD8 tumor-infiltrating lymphocytes are predictive of survival in muscle-invasive urothelial carcinoma. *Proc. Natl. Acad. Sci. USA* 2007, 104, 3967–3972. [CrossRef] [PubMed]
- Kolbeck, P.C.; Kaveggia, F.F.; Johansson, S.L.; Gune, M.; Taylor, R. The relationships among tumor-infiltrating lymphocytes, histopathologic findings, and long-term clinical follow-up in renal cell carcinoma. *Mod. Pathol.* 1992, 5, 420–425. [PubMed]
- 113. Bromwich, E.J.; McArdle, P.A.; Canna, K.; McMillan, D.; McNicol, A.; Brown, M.; Aitchisonet, M. The relationship between T-lymphocyte infiltration, stage, tumour grade and survival in patients undergoing curative surgery for renal cell cancer. *Br. J. Cancer.* **2003**, *89*, 1906–1908. [CrossRef] [PubMed]
- 114. Remark, R.; Alifano, M.; Cremer, I.; Lupo, A.; Dieu-Nosjean, M.C.; Riquet, M.; Crozet, L.; Ouakrim, H.; Goc, J.; Cazes, A.; et al. Characteristics and Clinical Impacts of the Immune Environments in Colorectal and Renal Cell Carcinoma Lung Metastases: Influence of Tumor Origin. *Clin. Cancer Res.* 2013, 19, 4079–4091. [CrossRef] [PubMed]
- 115. Nakano, O.; Sato, M.; Naito, Y.; Suzuki, K.; Orikasa, S.; Aizawa, M.; Suzuki, Y.; Shintaku, I.; Nagura, H.; Ohtani, H. Proliferative Activity of Intratumoral CD8+ T-Lymphocytes As a Prognostic Factor in Human Renal Cell Carcinoma. *Cancer Res.* 2001, *61*, 5132–5136. [PubMed]
- 116. Hotta, K.; Sho, M.; Fujimoto, K.; Shimada, K.; Yamato, I.; Anai, S.; Konishi, N.; Hirao, Y.; Nonomura, K.; Nakajima, Y. Prognostic significance of CD45RO+ memory T cells in renal cell carcinoma. *Br. J. Cancer.* 2011, 105, 1191–1196. [CrossRef]
- 117. Irani, J.; Goujon, J.M.; Ragni, E.; Peyrat, L.; Hubert, J.; Saint, F.; Mottet, N. High-grade inflammation in prostate cancer as a prognostic factor for biochemical recurrence after radical prostatectomy. *Urology* **1999**, *54*, 467–472. [CrossRef]
- 118. Karja, V.; Aaltomaa, S.; Lipponen, P.K.; Isotalo, T.; Talja, M.; Mokka, R. Tumour-infiltrating Lymphocytes: A Prognostic Factor of PSA-free Survival in Patients with Local Prostate Carcinoma Treated by Radical Prostatectomy. *Anticancer Res.* 2005, 25, 4435–4438.
- Zeigler-Johnson, C.; Morales, K.H.; Lal, P.; Feldman, M. The Relationship between Obesity, Prostate Tumor Infiltrating Lymphocytes and Macrophages, and Biochemical Failure. *PLoS ONE* 2016, *11*, e0159109. [CrossRef]
- 120. Richardsen, E.; Uglehaus, R.D.; Due, J.; Busch, C.; Busund, L.T. The prognostic impact of M-CSF, CSF-1 receptor, CD68 and CD3 in prostatic carcinoma. *Histopathology* **2008**, *53*, 30–38. [CrossRef]

121. McArdle, P.A.; Canna, K.; McMillan, D.C.; McNicol, A.M.; Campbell, R.; Underwood, M.A. The relationship between T-lymphocyte subset infiltration and survival in patients with prostate cancer. *Br. J. Cancer* 2004, *91*, 541–543. [CrossRef] [PubMed]

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).