



Complete Genome Sequence of *Enterococcus faecium* FS86, Used for Propagation of Bacteriophages with Therapeutic Potential

Rustam M. Buzikov,^a Emma G. Pilgrimova,^a  Andrey M. Shadrin^a

^aLaboratory of Bacteriophage Biology, G. K. Skryabin Institute of Biochemistry and Physiology of Microorganisms, Pushchino Scientific Center for Biological Research of the Russian Academy of Sciences, Federal Research Center, Pushchino, Moscow Oblast, Russia

ABSTRACT The *Enterococcus faecium* FS86 genome consists of a 2,685,395-bp chromosome and a 9,751-bp plasmid. The plasmid harbors mobilization-related genes. The pathogenicity factor genotype is *cylA* negative, *aggA* negative, *gelE* negative, *sprE* negative, *esp* negative, *eep* positive, and *efaA* positive. *E. faecium* FS86 belongs to multilocus sequence type 296, together with the probiotic strain *E. faecium* SF68.

Enterococcus faecium is a common member of the human gut microflora (1) that is often used as a probiotic (2). However, several strains of enterococci are known to be associated with urinary tract infections, bacteremia, endocarditis, and other infectious diseases (1). Bacteriophages can be used to treat infections caused by these antibiotic-resistant *E. faecium* strains (3). In this paper, we report the whole-genome sequence of the *E. faecium* FS86 strain, which was used for propagation of certain bacteriophages with therapeutic potential.

E. faecium FS86 was isolated from a capsule of Bifiform (Ferrosan) by plating the material onto KF *Streptococcus* agar (Difco). The procedure was repeated three times, and a single colony from the third plate was grown in a test tube with KF *Streptococcus* broth (Difco) to obtain a stock culture, which was maintained at -80°C . Total DNA was isolated by the standard phenol-chloroform DNA extraction method (4). The MiSeq library was prepared with the KAPA HyperPlus kit. A total of 29,676,276 Illumina MiSeq paired-end reads with an average length of 101 nucleotides (nt) were obtained. The Oxford Nanopore library was prepared using SQK-LSK109 and EXP-NBD104 kits; 19,280 Oxford Nanopore reads, with an average length of 2,374 nt (maximum read length, 77,526 nt), were used for the assembly. Genome assembly was performed using the Genome Assembly Service at the PATRIC v3.6.5 website (<http://patricbrc.org>) (5), with both Illumina paired-end reads and Nanopore reads, using the Unicycler v0.4.8 assembly strategy; read trimming was performed with the Trim Galore tool (6) implemented in PATRIC. The resulting assembly graph was visualized in Bandage v0.8.1 (7). All genome assembly software was used with default parameters, except that the “trim reads before assembly” parameter was changed to “true” for the Trim Galore tool. Two circular contigs (lengths of 2,685,395 bp and 9,751 bp, and average coverages of $751.5\times$ and $3,006\times$, respectively) were obtained, suggesting that the genome is represented by a chromosome and a plasmid with a copy number of ~ 4 per chromosome. The GC contents are 38.4% and 32.6% for the chromosome and the plasmid, respectively.

The *E. faecium* FS86 chromosome was annotated by the NCBI Prokaryotic Genome Annotation Pipeline v4.11 (8). In total, 2,594 putative genes were identified, including 2,503 coding sequences (CDSs), 6 clusters of the 16S, 23S, and 5S rRNA genes, 69 tRNA genes, 1 transfer-messenger RNA gene, and 3 noncoding RNA genes. The pFS86 plasmid was annotated by RAST v2 (9), and 14 genes were identified. The plasmid

Citation Buzikov RM, Pilgrimova EG, Shadrin AM. 2020. Complete genome sequence of *Enterococcus faecium* FS86, used for propagation of bacteriophages with therapeutic potential. *Microbiol Resour Announc* 9:e00776-20. <https://doi.org/10.1128/MRA.00776-20>.

Editor Kenneth M. Stedman, Portland State University

Copyright © 2020 Buzikov et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Andrey M. Shadrin, andrey2010s@gmail.com.

Received 17 July 2020

Accepted 3 September 2020

Published 24 September 2020

harbors mobilization (CDS1 to CDS3) and replication (CDS6 and CDS7) gene modules similar to those of the pHY (10) and pCIZ2 (11) plasmids harboring bacteriocin genes. No genes encoding antibiotic-modifying enzymes were detected on the plasmid, but the FS86 chromosome encodes aminoglycoside *N*-acetyltransferase AAC(6')-II, providing resistance to aminoglycosides.

The FS86 strain belongs to sequence type 296 (ST296) from clonal cluster 98 (CC98) according to the scheme developed by Homan et al. (12, 13). Interestingly, the *E. faecium* SF68 strain, which has been widely used as a probiotic in the past 4 decades (2), also belongs to ST296.

Data availability. This whole-genome project has been deposited in GenBank under accession number [CP053704](#) for the chromosome and accession number [MT501398.1](#) for the plasmid. Raw reads are available in the Sequence Read Archive under BioProject accession number [PRJNA633747](#).

ACKNOWLEDGMENTS

This work was project 075-03-2019-525/2 (regulation AAAA-A19-119120390010-1) of State Assignment/Ministry of Science and Higher Education of the Russian Federation.

We thank Gleb Speshilov, BioSpark, for his advice on sequencing technology.

REFERENCES

1. Fiore E, Van Tyne D, Gilmore MS. 2019. Pathogenicity of enterococci. *Microbiol Spectrum* 7:GPP3-0053-2018. <https://doi.org/10.1128/microbiolspec.GPP3-0053-2018>.
2. Wunderlich PF, Braun L, Fumagalli I, D'Apuzzo V, Heim F, Karly M, Lodi R, Politta G, Vonbank F, Zeltner L. 1989. Double-blind report on the efficacy of lactic acid-producing enterococcus SF68 in the prevention of antibiotic-associated diarrhoea and in the treatment of acute diarrhoea. *J Int Med Res* 17:333–338. <https://doi.org/10.1177/030006058901700405>.
3. Shlezinger M, Copenhagen-Glazer S, Gelman D, Beyth N, Hazan R. 2019. Eradication of vancomycin-resistant enterococci by combining phage and vancomycin. *Viruses* 11:954. <https://doi.org/10.3390/v11100954>.
4. Sambrook J, Fritsch EF, Maniatis T. 1989. *Molecular cloning: a laboratory manual*. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.
5. Wattam AR, Davis JJ, Assaf R, Boisvert S, Brettin T, Bun C, Conrad N, Dietrich EM, Disz T, Gabbard JL, Gerdes S, Henry CS, Kenyon RW, Machi D, Mao C, Nordberg EK, Olsen GJ, Murphy-Olson DE, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Vonstein V, Warren A, Xia F, Yoo H, Stevens RL. 2017. Improvements to PATRIC, the all-bacterial bioinformatics database and analysis resource center. *Nucleic Acids Res* 45:D535–D542. <https://doi.org/10.1093/nar/gkw1017>.
6. Krueger F. 2015. Trim Galore: a wrapper tool around Cutadapt and FastQC to consistently apply quality and adapter trimming to FastQ files. https://www.bioinformatics.babraham.ac.uk/projects/trim_galore.
7. Wick RR, Schultz MB, Zobel J, Holt KE. 2015. Bandage: interactive visualisation of de novo genome assemblies. *Bioinformatics* 31:3350–3352. <https://doi.org/10.1093/bioinformatics/btv383>.
8. Tatusova T, DiCuccio M, Badredin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44:6614–6624. <https://doi.org/10.1093/nar/gkw569>.
9. Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST Server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
10. Yamashita H, Tomita H, Inoue T, Ike Y. 2011. Genetic organization and mode of action of a novel bacteriocin, bacteriocin 51: determinant of VanA-type vancomycin-resistant *Enterococcus faecium*. *Antimicrob Agents Chemother* 55:4352–4360. <https://doi.org/10.1128/AAC.01274-10>.
11. Criado R, Diep DB, Aakra Á, Gutiérrez J, Nes IF, Hernández PE, Cintas LM. 2006. Complete sequence of the enterocin Q-encoding plasmid pCIZ2 from the multiple bacteriocin producer *Enterococcus faecium* L50 and genetic characterization of enterocin Q production and immunity. *Appl Environ Microbiol* 72:6653–6666. <https://doi.org/10.1128/AEM.00859-06>.
12. Homan WL, Tribe D, Poznanski S, Li M, Hogg G, Spalburg E, van Embden JDA, Willems RJ. 2002. Multilocus sequence typing scheme for *Enterococcus faecium*. *J Clin Microbiol* 40:1963–1971. <https://doi.org/10.1128/jcm.40.6.1963-1971.2002>.
13. Jolley KA, Bray JE, Maiden MC. 2018. Open-access bacterial population genomics: BIGSdb software, the PubMLST.org website and their applications. *Wellcome Open Res* 3:124. <https://doi.org/10.12688/wellcomeopenres.14826.1>.