

Heat stress and β -adrenergic agonists alter the adipose transcriptome and fatty acid mobilization in ruminant livestock¹

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Transl. Anim. Sci. 2020.4:S141–S144
doi: 10.1093/tas/txaa122

INTRODUCTION

Growth and feed efficiency of cattle are improved by supplementation with the beta-adrenergic agonists (β AA), ractopamine hydrochloride (RH; β_1 AA) or zilpaterol hydrochloride (ZH; β_2 AA) (Elam et al., 2009). β AA supplementation alters adipose deposition by inhibiting fatty acid biosynthesis and promoting lipolysis of stored triacylglycerols into free fatty acids (FFAs) (Johnson et al., 2014). However, β_2 adrenoceptors (β AR) desensitize with chronic activation (Re et al., 1997); supplementation is thus limited to the last 20 to 40 d of feeding.

The annual economic impact of heat stress (HS) has been estimated to exceed \$2.4 billion (St-Pierre et al., 2003). Heat-stressed livestock have reduced growth rates, dry matter intake, and average daily gain (Mitlöhner et al., 2001; St-Pierre et al., 2003). In response to acute stress,

signaling pathways for lipolysis of circulating and stored triglycerides are activated, while chronic stress increases lipogenesis and adipogenesis (Campbell et al., 2009; Peckett et al., 2011). In cattle, HS also increases the responsiveness of adipocytes to lipolytic signals, increasing lipolysis (Faylon et al., 2015).

The objective of this study was to understand how HS and β AA independently and interactively affect adipose tissue. Prior work identified minimal impact of RH on metabolic properties (Barnes et al., 2019) and on the transcriptome of skeletal muscle (Kubik et al., 2018). We therefore hypothesized that RH may be primarily affecting adipose; specifically, that lipolytic activity is increased due to heat and β AA in an additive fashion. We tested this hypothesis in RH-supplemented lambs and ZH-supplemented cattle exposed to HS for 30 and 21 d, respectively.

MATERIALS AND METHODS

This study was approved by the Institutional Animal Care and Use Committee at the University of Nebraska-Lincoln, which is accredited by AAALAC International.

For study 1, wether lambs were fed a high-energy feedlot diet for 30 d under thermal neutral (TN; Temperature Humidity Index [THI] = 65; $n = 14$) or HS (THI = 80; $n = 12$) conditions and supplemented with ractopamine HCl (RH; 60 mg/head/d)

¹This project is based on research that was partially supported by the Nebraska Agricultural Experiment Station with funding from the Hatch Multistate Research capacity funding program (accession number 1011055) from the USDA National Institute of Food and Agriculture. A portion of this work was completed utilizing the Holland Computing Center of the University of Nebraska, which receives support from the Nebraska Research Initiative.

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Received April 29, 2020.

Accepted July 8, 2020.

or none (NS) in a 2×2 factorial. Thermal neutral lambs were pair-fed to the average intake of HS lambs. At harvest, subcutaneous fat was flash frozen. Poly-A⁺ selected libraries were sequenced from isolated RNA using 150 bp, paired-end reads to a minimum depth of 20 million reads/sample (Michigan State University). After quality control, transcripts as annotated in Oar_rambouillet_v1.0 were quantified (STAR; Dobin et al., 2013). Differential expression (DE) analyses were performed in DESeq2 (Love et al., 2014) with a significance threshold False Discovery Rate (FRD) of 0.05. Exploration of data was performed with DAVID (Huang et al., 2009a, 2009b). To study the main effects of environment and supplement, loci with a significant interaction were removed and the remaining loci reevaluated. Pathway analysis (Qiagen) was conducted on all loci with raw $P < 0.05$ for the main effects.

For study 2, Red Angus-based steers were fed a high-energy diet for 21 d under TN ($n = 12$; pair-fed to HS average) or HS (THI = 83; $n = 12$) conditions and supplemented with zilpaterol hydrochloride (ZH; 8.38 mg/kg DM/d) or none (NS) in a 2×2 factorial. At harvest, visceral adipose was flash frozen to determine fatty acid mobilization in a method adapted from Raclot and Groscolas (1993). Briefly, modified Krebs Ringer buffer (MKRB) was made (9 g of KRB [Sigma], 900 mL ddH₂O, 15 mM NaHCO₃, 2.5 mM CaCl₂ dihydrate, and 4% fatty acid free BSA, pH adjusted to 7.4 and sterilized). Visceral fat was minced, strained (200 μ M), washed (37 °C MKRB), and minced again. Adipose (400 ± 10 mg) was added to 5 mL MKRB containing 0 or 1 μ M epinephrine. Samples were incubated in a shaking water bath (2 h, 37 °C), and the media filtered (2.4 cm glass microfiber filter) and stored at -80 °C. Free fatty acids were quantified by colorimetric detection (Sigma Aldrich Free Fatty Acid Quantification Kit), read at 570 nm (BioTek EPOCH). Concentrations were determined using a 0 to 4 nmol/ μ L standard curve for palmitic acid and analyzed using the PROC MIXED procedure (SAS Institute Inc., Cary, NC). $P < 0.05$ were considered significant.

RESULTS

Seventy-one loci were DE ($P_{\text{adj}} < 0.05$) due to a temperature by supplement interaction. Differential expression loci with the greatest logFC include G-protein-coupled receptor, *GPRC5A*, and others involved in G-protein receptor signaling (e.g., *SAAI*, *NPW*, and *NGEF*; Table 1). When loci DE due to interaction were removed, *RBM3* and *ATXN7L1* were DE ($P_{\text{adj}} < 0.1$) due to temperature (Table 2).

Table 1. Loci DE due to an interaction of environment and supplement in adipose of wether lambs

Gene ID	<i>P</i> -value	Adjusted <i>P</i> -value
<i>GPRC5A</i>	1.10E-07	<0.001
<i>FOSB</i>	1.43E-04	0.033
<i>FOS</i>	1.92E-06	0.002
<i>SAA2</i>	1.09E-04	0.023
<i>SERPINF2</i>	6.63E-08	<0.001
<i>SAAI</i>	3.31E-05	0.012
<i>NPW</i>	2.33E-04	0.048
<i>FMOD</i>	4.08E-05	0.014
<i>CIQTNF3</i>	2.55E-06	0.002
<i>SFRP4</i>	8.97E-07	0.001
<i>TPSB2</i>	2.09E-05	0.008
<i>LTC4S</i>	449E-09	<0.001
<i>NGEF</i>	9.53E-05	0.025
<i>IL34</i>	3.23E-05	0.012

Table 2. Loci DE in wether lamb adipose due to HS

Gene ID	Log ₂ fold change	<i>P</i> -value	Adjusted <i>P</i> -value
<i>RBM3</i>	-0.888	2.53E-07	0.006
<i>ATXN7L1</i>	0.550	1.06E-05	0.10

No loci were DE due to supplement. Pathway analysis predicted the “Adipogenesis Pathway” to be altered ($P < 0.001$) but without clear directionality of dysregulation. The top regulator effect networks included the biological functions: concentration of fatty acid, molecule transport, and cell migration (Table 3).

There was no interaction between environment and supplement for ex vivo FFA mobilization from steer adipose. Free fatty acid concentration did not differ among groups at 0 μ M (Fig. 1). At 1 μ M epinephrine, FFA was greater ($P < 0.05$) in TN than HS. All treatment groups responded to epinephrine, with TN/ZH having the highest concentration (Fig. 1).

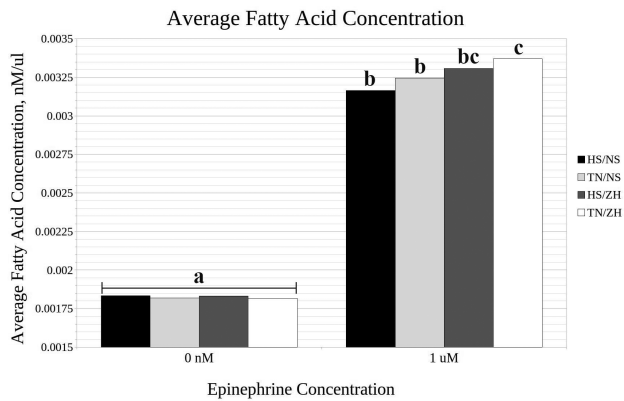
DISCUSSION

We identified interacting and independent effects of HS and β AA supplementation on adipogenesis and lipolysis in sheep, as indicated by gene expression. We also found that fatty acid mobilization was impaired by HS but enhanced by β AA supplementation in cattle. Zilpaterol hydrochloride supplementation in heat-stressed cattle increased the FFA-mobilization response to epinephrine even after 21 d, but the response was less due to HS.

Transcriptome analyses in wethers did not clearly predict whole-pathway changes due to the interaction of HS and β AA. Several loci implicated

Table 3. Top regulator effect networks in the adipose transcriptome due to HS

Regulators	Disease and functions	Consistency score
CD28, CD3 group, LCN2, RETN	Concentration of fatty acid, transport of molecule	7.217
CD3 group	Concentration of fatty acid, migration of cells	6.425

**Figure 1.** Mean fatty acid concentration for each treatment group. Superscripts denote a significance ($P < 0.05$).

however are components of G-protein receptor pathways, through which β A signal. Others such as *FOS* and *FOSB* are associated with inflammation (Wagner and Efreil, 2005). Additional data from this study demonstrated an inflammatory response due to HS that was moderated by RH (Swanson et al., 2020). Heat stress alone altered expression of two loci in adipose. *RBM3* encodes for a protein induced in response to hypoxia and cold shock (Wellmann et al., 2010); it was downregulated in HS lambs. *ATXN7L1* is associated with neurological disorders (Carlson et al., 2009), but Komolka et al. (2016) reported it is downregulated in the longissimus dorsi of cattle with greater intramuscular fat. Increased *ATXN7L1* expression in HS lambs could therefore contribute to the observed decrease in fat (Swanson et al., 2020). Pathway analyses predicted adipogenesis to be dysregulated due to HS. This was not surprising, considering that there are presumably mechanisms whereby HS affects adipose tissue not apparent from RNA analyses alone. Two regulator effect networks identified affected fatty acid concentrations and movement, supporting our hypothesis that HS impairs fat homeostasis. Conversely, we did not identify transcriptome changes in subcutaneous adipose attributable solely to RH. It is worth noting that samples were collected after 30 d on RH, and thus it is possible that β AR had been desensitized. The chronic effect β A on adipose can also be difficult to observe from a single time-point, as β A enhance

lipid mobilization but increase fatty acid synthesis (Yang and McElligott, 1989).

In cattle, HS and ZH supplementation each modified fatty acid mobilization of adipose due to epinephrine independently, although the two factors did not have interacting effects. Zilpaterol hydrochloride preferentially binds β_2 AR, and thus the heightened effect could be due to epinephrine acting on β_1 AR with greater frequency than it would in the absence of ZH. It is more likely, however, that exposure to both epinephrine and ZH had an additive effect. Decreased FFA mobilization due to HS was possibly an effect of chronic exposure. In study 1, HS caused increased circulating epinephrine (Swanson et al., 2020). If steers in study 2 responded likewise, it is reasonable to speculate that their β AR became desensitized and less responsive to epinephrine. Acute HS increases adipocyte responses to lipolytic signals (Faylon et al., 2015), but we postulate that chronic HS decreased responsiveness due to downregulated β AR.

IMPLICATIONS

Loci in adipose have altered expression due to the combined impact of HS and β A supplementation. Specific loci associated with inflammation were observed, warranting additional investigation in order to fully elucidate their role in adipose HS. We also provide evidence that increased lipolysis is a mechanism by which ZH can reduce carcass fat and promote feed efficiency. Conversely, HS impaired fatty acid mobilization, presumably via β AR desensitization resulting from chronic stimulation. Finally, no interacting effects of HS and β A supplementation on mechanisms that would impact animal wellbeing were apparent. Building a better understanding of the mechanisms by which animals respond to HS and β A supplementation will aid in generating improved management practices to improve sustainability of livestock production.

Conflict of interest statement. None declared.

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