

RAD-QTL Mapping Reveals Both Genome-Level Parallelism and Different Genetic Architecture Underlying the Evolution of Body Shape in Lake Whitefish (*Coregonus clupeaformis*) Species Pairs

Martin Laporte,^{*,1} Sean M. Rogers,[†] Anne-Marie Dion-Côté,^{*} Eric Normandeau,^{*}

Pierre-Alexandre Gagnaire,[‡] Anne C. Dalziel,^{*} Jobran Chebib,[†] and Louis Bernatchez^{*}

^{*}Institut de Biologie Intégrative et des Systèmes (IBIS), Département de Biologie, Université Laval, Pavillon Charles-Eugène-Marchand, Québec, G1V 0A6, Canada, [†]Department of Biological Sciences, University of Calgary, 2500 University Drive N.W., Calgary, Alberta, T2N 1N4, Canada, and [‡]Institut des Sciences de l'Évolution – Montpellier (ISEM - CNRS UMR5554), SMEL, 2 rue des Chantiers, 34200 Sète, France

ABSTRACT Parallel changes in body shape may evolve in response to similar environmental conditions, but whether such parallel phenotypic changes share a common genetic basis is still debated. The goal of this study was to assess whether parallel phenotypic changes could be explained by genetic parallelism, multiple genetic routes, or both. We first provide evidence for parallelism in fish shape by using geometric morphometrics among 300 fish representing five species pairs of Lake Whitefish. Using a genetic map comprising 3438 restriction site–associated DNA sequencing single-nucleotide polymorphisms, we then identified quantitative trait loci underlying body shape traits in a backcross family reared in the laboratory. A total of 138 body shape quantitative trait loci were identified in this cross, thus revealing a highly polygenic architecture of body shape in Lake Whitefish. Third, we tested for evidence of genetic parallelism among independent wild populations using both a single-locus method (outlier analysis) and a polygenic approach (analysis of covariation among markers). The single-locus approach provided limited evidence for genetic parallelism. However, the polygenic analysis revealed genetic parallelism for three of the five lakes, which differed from the two other lakes. These results provide evidence for both genetic parallelism and multiple genetic routes underlying parallel phenotypic evolution in fish shape among populations occupying similar ecological niches.

KEYWORDS

adaptive radiation
parallel evolution
fish body shape
geometric morphometrics
genotyping-by-sequencing

Understanding the genetic basis of adaptation of complex phenotypic traits is a major goal in ecologic and evolutionary genetics. In particular, the mechanisms linking genotype to phenotype can be complex (e.g., pleiotropy, polygeny, epistasis) and result in genetic architectures that may either constrain or promote the effects of selection on beneficial

alleles in a new environment (Gompel and Prud'homme 2009; Elmer and Meyer 2011; Losos 2011; Conte *et al.* 2012; Rogers *et al.* 2013). Therefore, elucidating whether parallel adaptation to similar environments relies on similar genetic changes is crucial to better understand how populations can adapt to different environments.

Parallel evolution refers to the evolution of ecologically and phenotypically similar traits in independently derived populations or species (Endler 1986; Losos 2011; Conte *et al.* 2012; Merilä 2014). Parallel evolution has best been documented in microbial experimental evolution studies (Rainey and Travisano 1998; Gerstein *et al.* 2012; Heron and Doeblí 2013). However, it is a relatively common feature observed in fishes inhabiting lakes in north temperate regions (Mcphail 1984; Bernatchez and Dodson 1990; Skúlason and Smith 1995; Taylor 1999; Østbye *et al.* 2006; Laporte *et al.* 2011; Elmer *et al.* 2014). Evidence for phenotypic parallelism is commonly used to infer the possible role of adaptive mechanisms being involved, because the probability that such phenotypic transitions occurred

Copyright © 2015 Laporte *et al.*

doi: 10.1534/g3.115.019067

Manuscript received December 8, 2014; accepted for publication May 11, 2015; published Early Online May 21, 2015.

This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Supporting information is available online at www.g3journal.org/lookup/suppl/doi:10.1534/g3.115.019067/-/DC1

¹Corresponding author: Institut de Biologie Intégrative et des Systèmes (IBIS), Département de Biologie, Université Laval, 1030 avenue de la Médecine, Québec, G1V 0A6, Québec, Canada. E-mail: uni.mlaporte@gmail.com

multiple times only through stochastic processes (e.g., genetic drift) typically is considered to be low (Schluter 2000; Nosil 2012).

Body shape is a complex phenotypic trait composed of a suite of underlying characters that often are genetically based but also may vary by direct environmental induction (Proulx and Magnan 2004; Albert *et al.* 2008; Klingenberg 2010; Rogers *et al.* 2012; Rogers and Jaminiczky 2014). In fishes, shape differentiation can influence a variety of performance traits such as prolonged or burst swimming, feeding efficiency, and mating displays (Webb 1982, 1984; Langerhans and Reznick 2010; Walker 2010). Moreover, the strength of selection acting on whole-animal performance may vary among habitats. For examples, a slender, fusiform body shape reduces drag and may be selected in fish that swim long distances, a stout caudal peduncle allows for faster, more powerful burst swimming that can be beneficial for fish that ambush their prey or those that sprint to escape predators, and larger eyes and a terminal mouth increase feeding efficiency in fish that feed on small prey and may evolve via natural selection in planktivorous fish (Webb 1982, 1984; Langerhans and Reznick 2010; Walker 2010; Willacker *et al.* 2010). Identifying the genetic mechanisms by which fish shape evolves would contribute to improve our understanding of adaptation for a complex, ecologically important phenotypic trait.

The Lake Whitefish (*Coregonus clupeaformis*) species complex exhibits adaptive divergence in association with shape contributing to species-pairs referred to as “normal” and “dwarf.” The “normal” species occupies a benthic niche whereas the derived “dwarf” species is specialized for a limnetic niche (Bernatchez and Dodson 1990; Lu and Bernatchez 1999; Landry *et al.* 2007; Landry and Bernatchez 2010). The dwarf species has evolved independently and repeatedly from the normal ones in several lakes in the St. John River Drainage (Québec, Canada, and Maine, USA) after the last glacial maximum [~12,000 years ago or about 3000 generations (Bernatchez and Dodson 1990; Pigeon *et al.* 1997; Lu *et al.* 2001; Bernatchez *et al.* 2010)]. The species differ in adult body size and several ecologic and physiologic traits associated with occupying distinct trophic niches (Bernatchez 2004; Bernatchez *et al.* 2010; Evans and Bernatchez 2012; Evans *et al.* 2012; Evans *et al.* 2013). Indeed, these differences match ecologic predictions for habitat use (Willacker *et al.* 2010; Harrod *et al.* 2010; Ozerov *et al.* 2015). Shape differentiation resulting in specialization to benthic and limnetic niches is expected to have evolved in these species pairs, but whether similar genetic changes underlie the evolution of body shape in different replicate lakes is unknown.

The first objective of this study was to test for shape differentiation and parallelism at the phenotypic level between sympatric Whitefish species that evolved independently in five geographically isolated lakes using geometric morphometrics. The second objective was to identify quantitative trait loci (QTL) underlying these same shape traits in a backcross family. We then examined the putative functions of genes associated with QTL toward proposing elucidating the potential molecular mechanisms underlying fish shape differentiation. As third objective we tested for evidence of genetic parallelism among independent wild populations by using both a single-locus method (outlier analysis) and a polygenic approach (analysis of covariation among markers) at shape QTL.

MATERIALS AND METHODS

Sampling, experimental crosses, and genetic mapping

A total of 300 wild Whitefish were sampled from five lakes in the Saint John river basin harboring sympatric Whitefish species pairs using gill nets between June and July 2010 (Table 1). Between 50 and 67 specimens

■ **Table 1 Geographic sampling coordinates, dates, and sample sizes of dwarf and normal Whitefish photographed in the five lakes surveyed in this study**

Lakes	Geographic Coordinates	Date	Normal	Dwarf	Total
Cliff	46°24'20 N 69°15'60 W	June 2010	25	35	60
East	47°10'42 N 69°32'52 W	July 2010	35	33	68
Indian	46°15'24 N 69°16'49 W	June 2010	24	31	55
Témiscouasta	47°39'58 N 68°49'22 W	July 2010	34	33	67
Webster	46° 09'18 N 69° 05'17 W	June 2010	25	25	50
Total	—	—	143	157	300

were collected from each lake (Table 1). These were used to determine whether body shape differentiation was present between dwarf and normal Whitefish of a given species pairs and if parallelism at the phenotypic level was present among lakes.

A laboratory-raised backcross (BC) family derived from the *F1 hybrid x dwarf* cross previously used for QTL mapping (Rogers and Bernatchez 2007) was used to map shape QTL in this study. A total of 198 progeny were produced, 102 of which were subsequently available for constructing a second generation linkage map containing 3438 single-nucleotide polymorphisms (SNP) genotyped by restriction site-associated DNA (RAD) sequencing and assigned to 40 different linkage groups (LGs) with an average resolution of 0.89 cM between mapped markers (mapping details provided in Gagnaire *et al.* 2013a).

Measuring fish shape and landmark positioning

Immediately after euthanization, digital photographs of the left side of the 300 wild and 102 BC fish were taken with a Nikon Coolpix P7700 camera to avoid shape deformation that can be associated with preservation. Fish were placed on a Styrofoam board with fins extended and fixed with needles. Fifteen landmarks were digitized on each image using tpsDig v2.16 (Rohlf 2010) to quantify shape differences between individuals (Figure 1). Landmarks were chosen for an optimal coverage of the body (Zelditch *et al.* 2004), reflecting characteristics expected to be under differential selection between limnetic and benthic species (Webb 1982, 1984; Willacker *et al.* 2010). To preserve information on shape differences among fish and to remove information unrelated to shape (*i.e.*, scale, position, and orientation), a partial generalized procrustes analysis superimposition was first applied (Rohlf and Slice 1990; Dryden and Mardia 1998). This resulted in 15 × (abscissa) and 15 y (ordinate) coordinates providing 30 phenotypic traits for further analyses. All protocols were in accordance with the Canadian Council for Animal Care.

Testing for fish shape differentiation and parallelism in the wild

We first estimated fish shape from the superimposed coordinates projected on a $k \times p - 4$ dimensional principal component analysis (PCA) (k = number of coordinates; P = number of landmarks; the four dimensions being lost during the Procrustes superimposition; see Claude 2008) on the 300 wild Whitefish. We used a broken-stick distribution to determine the number of informative principal components (PCs) (Legendre and Legendre 1998). Euclidian distance and angles between the five lines connecting species pairs and the first axis

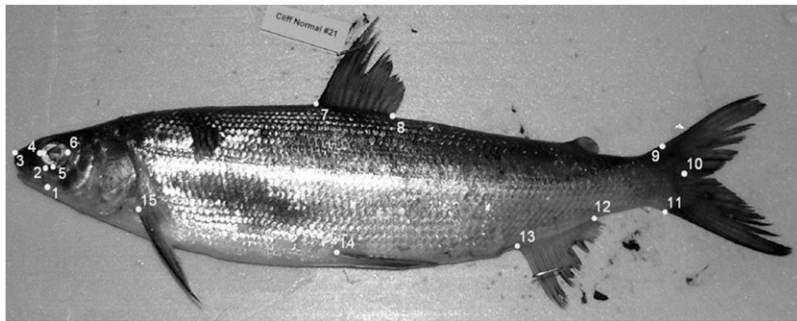


Figure 1 Position of 15 landmarks (30 x, y coordinates) used to study body shape in Lake Whitefish (1: lower extent of distal maxilla; 2: upper extent of distal maxilla; 3: tip of snout; 4: anterior extent of orbital; 5: ventral extent of orbital; 6: anterior extent of orbital; 7: anterior insertion of dorsal fin; 8: posterior insertion of dorsal fin; 9: dorsal insertion of caudal fin; 10: base of caudal fin; 11: ventral insertion of caudal fin; 12: posterior insertion of anal fin; 13: anterior insertion of anal fin; 14: anterior insertion of pelvic fin; 15: anterior insertion of pectoral fin).

of a between-group PCA were computed using the *groupPCA* function in R/Morph (Schlager 2014). Euclidian distance and angle comparison serve respectively to quantify the fish shape differentiation and the extent of parallelism among species pairs. To visualize the effects of species identity on fish shape for each lake, the juxtaposition of the mean shape of the fish between species from the same lake was conducted.

In addition, a multivariate analysis of variance (MANOVA), from PC scores of the informative axes, was then run to test for the effect of “species” (Dwarf or Normal), “lake,” “sex” (male or female), and their interactions on fish shape. Identification of sex was performed by visual inspection of the gonads and/or by a sex determination locus for Whitefish (locus 82008; Gagnaire *et al.* 2013a) in cases in which the sex was not determined anatomically. To test for significant shape differentiation among species pairs, we ran five independent MANOVAs for each lake with the explanatory variables “species,” “sex,” and their interaction. Shape differentiation was also assessed by verifying the significance of the Euclidian distance with 10,000 permutations using the *groupPCA* function in R/Morph (Schlager 2014).

The effect of sex on fish shape in laboratory-raised conditions

Identification of sex for BC fish also was performed by visual inspection of the gonads (Rogers and Bernatchez 2007). A MANOVA was then run on fish shape (response variable) with the explained variable sex to test for an effect of sex. Differences in fish shape were visualized with the juxtaposition of the mean fish shape of sexes. All statistical analyses were performed with R software v2.15.1 (Claude 2008; R Core Team 2014).

Shape QTL analyses

Data files, including phenotypes and genotypes of laboratory-raised Whitefish, were created with the four-way cross format and imported with the *read.cross* function in R/qtl (allowing for four segregating alleles; Broman *et al.* 2003). Genotype probabilities were estimated with the *calc.geneprob* function and a step size of 1 cM for interval mapping. We used the 30 shape coordinates directly instead of shape principal components for QTL mapping, because although the principal components are orthogonal, they do not correspond to independent dimensions of genetic variation (Albert *et al.* 2008; Rogers *et al.* 2012; Berner 2012). We scanned the genetic map for QTL using the Haley-Knott regression method (Haley and Knott 1992) in R/qtl with the *scanone* function (Broman *et al.* 2003). For each of the 30 shape coordinates, a significance threshold was determined for each LG (at 1% significance level) and genome-wide (at 5% significance level) using 10,000 permutations. Finally, the percent variance explained (PVE) of each QTL was computed with the *fitqtl* function and we used the 1.5 LOD support to delimit the confidence interval of each QTL.

Annotation of body shape QTL

As QTL were genotyped by RAD sequencing, we were able to map these sequenced regions onto a draft Lake Whitefish genome (Pavey *et al.* unpublished data) to gain insight into the functions of some of the genes associated with shape differentiation QTL. This was done using the *blastn* program with the SNP RAD sequences (63 bp) associated with QTL against the assembled contigs of the draft Lake Whitefish genome. Although this genome is not yet published, its quality is sufficient to ensure reliable annotations of genes in subsequent steps (100× coverage, total scaffolds size = 1.3 Gb, N scaffolds = 464 428, scaffold N50 = 3802 bp). The sequence of scaffolds used in this study is presented in Supporting Information, Table S3. When multiple hits were found, only the contig with the greatest e-value was retained. If multiple hits had equal e-values, all contigs were retained for subsequent steps. We then annotated genome scaffolds using the function *blastx* against the Swiss-Prot database (last accessed March 21, 2015), keeping again only the best hit. If multiple hits with equal e-values were found, these were kept only when the annotations were similar (*e.g.*, same protein in different species). We used this two-step procedure to minimize false positives and multiple hits of our SNPs against public databases due to the short length of RAD sequences.

Testing for selection on shape QTL

A total of 200 wild Whitefish (20 for each of the 10 species/lake groups) were genotyped in a previous study by RAD sequencing for the 3438 mapped markers from the same 5 lakes sampled in this study. Details are provided in Gagnaire *et al.* (2013b). We used these genetic data to test for selection on shape QTL. To determine whether a shape QTL was potentially under divergent selection (single-locus approach), the extent of genetic differentiation between each of the five sympatric species pairs was compared with neutral predictions obtained using coalescent simulations in Arlequin v3.5.1 software (*i.e.*, five nonhierarchical Fdist analyses; Excoffier *et al.* 2009; Excoffier and Lischer 2010). The test was conducted independently for each lake. Corrections for multiple comparisons were performed by controlling the false discovery rate at 5% (Benjamini and Hochberg 1995) with the function *p.adjust* in R v3.0.3 (R Core Team 2014). The number and identity of outlier QTL or associated SNPs was then compared across lakes to evaluate the extent to which parallelism in fish shape is associated with repeated divergent selection at body shape QTL.

Under quantitative genetic theory, rapid adaptation of complex traits is considered highly polygenic (Pritchard *et al.* 2010; Messer and Petrov 2013). It is expected that simultaneous selection of variants at many loci (*i.e.*, polygenic adaptation) will result in subtle variation in allelic frequencies on several covarying loci, yielding a combined effect greater than the effect of individual loci on the phenotype (McKay and Latta 2002; Pritchard *et al.* 2010; Le Corre and Kremer 2012; Bourret *et al.* 2014; Pavey *et al.* 2015). Such subtle changes in allelic frequency

are not expected to be detected by genome scan methods that are based on the classical hitch-hiking model (Maynard Smith and Haigh 1974; Pritchard *et al.* 2010; Messer and Petrov 2013; Kemper *et al.* 2014). Therefore, we also tested for the occurrence of a group of covarying markers (which we refer to as a “polygenic approach”) among the 138 body shape QTL that differentiated the two species among the five lakes (see the section *Results*). To search for a group of covarying markers across population pairs, we used the randomForest function implemented in the Random Forest R package (Liam and Wiener 2002). The Random Forest algorithm is a tree-based ensemble machine learning tool more suited to detect evidence of polygenic adaptation since it search for correlation and interactions among loci (Goldstein *et al.* 2011; Boulesteix *et al.* 2012). The efficiency of Random Forest approach in finding a group of covarying markers that differentiate complex traits has been shown in several medicine and agriculture studies (Shi *et al.* 2005; Cordell 2009; Tang *et al.* 2009; Xu *et al.* 2011; Poland *et al.* 2012; Mokry *et al.* 2013; Jarquín *et al.* 2014) but still infrequent in evolutionary molecular [but see Briec *et al.* 2015; Pavay *et al.* 2015]. As recommended by Strobl *et al.* (2009) and Chen and Ishwaran (2012), a total of 100 forests (runs) sets with different seed numbers were computed to ensure randomness of the test. Each run had a total of 10,000 trees. The “importance of markers” (*i.e.*, an indicator of how a marker in interaction with other markers will successfully classify an individual) was used to select a set of covarying markers which we subsequently refer to as “important markers” (*sensu* Chen and Ishwaran 2012), for comparison with the single-locus approach. We added the 99% confidence interval of the 100 runs to this threshold in the selection of important markers to decrease the probability of type I error. The proportion of trees classifying an individual in the dwarf species group and the classification error rate of the Random Forest analysis were used as criteria to determine the presence of similar genetic changes among population pairs. It is expected that the proportion of such trees (which we subsequently refer to as “proportion of votes”; *sensu* Chen and Ishwaran 2012) will be high for dwarf individuals and low for normal individuals. Classification error rate should also be low if similar genetic changes are present among species pairs.

Further evidence of genetic parallelism was searched by calculating mean F_{ST} estimates of the important markers between normal and dwarf species in each lake. It has been documented that dwarf Whitefish evolved postglacially [12,000 years before present (ypb)] from the normal ones after a secondary contact in the St. John River basin (Bernatchez *et al.* 2010). If genes that altered fish shape have differentiated under a recent polygenic adaptation, it will thus be expected to observe low F_{ST} for the important markers. Moreover, if true genetic parallelism occurs among species pairs, we predicted that a Random Forest analysis should correctly assign Whitefish of a given species pair even if these fish are not included in the identification of important loci. Consequently, we i) computed a Random Forest analysis without a given species pair, ii) test the ability of the important markers obtained in the assignment of the excluded species pair individuals using the software GeneClass2 (Piry *et al.* 2004), and iii) redo this procedure for all species pairs. All computations were performed in R v3.0.3 (R Core Team 2014).

RESULTS

Fish shape parallelism in the wild

Broken-stick distribution indicated that only the two first PCs were informative. The first axis represents 50.6% of the variation and show variation among lakes (Figure 2). With the exception of Témiscouata Lake, dwarf Whitefish species also showed lower mean score on this

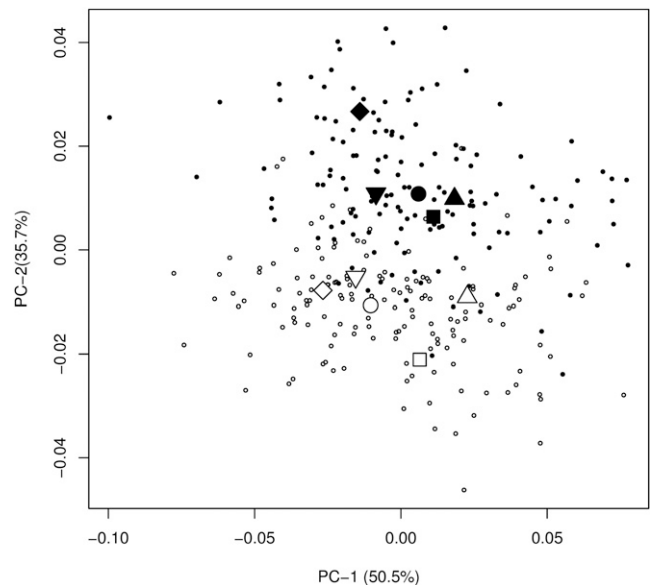


Figure 2 Fish shape variation among individuals along PC1 and PC2 in all five lakes. Each dot represents an individual (black color = normal species; white color = dwarf species). The means fish shape for each species lakes combination were also indicated (circle = Cliff; square = East; diamond = Indian; upper triangle = Témiscouata and lower triangle = Webster).

axis than normal Whitefish (Figure 2). The second axis represents 35.7% of the variation and showed a clear distinction between all dwarf and normal Whitefish (Figure 2). Euclidian distance showed an increasing gradient in the following order: Webster (0.020; P -value = 0.032), Témiscouata (0.021; P -value < 0.001), Cliff (0.028; P -value < 0.001), East (0.032; P -value < 0.001), and Indian (0.038; P -value < 0.001). This gradient roughly correspond to the one observed with the juxtaposition of mean shape (Figure 3). In addition, all between-group PCA angles point in a similar direction (9–55°). Interestingly, the lower angle observed (Cliff: 9°) is linked to a ventral thinning in dwarf in comparison to normal Whitefish (Figure 3), the greater angle (Témiscouata: 55°) is linked to a dorsal thinning in dwarf (Figure 3) and the intermediary angles (Indian: 18°, Webster: 22°, and East: 25°) are linked to both ventral and dorsal thinning (Figure 3). Dwarf Whitefish also have bigger eyes and longer tails than the normal fish in all of the five lakes (Figure 3).

The effect of both “species” (dwarf vs. normal) and “lake” on fish shape were highly significant ($P < 0.001$) (Table 2). The interaction “species × lake” and “lake × sex” also were significant and a trend of a possible effect of “sex” was observed ($P < 0.1$) (Table 2). As observed with Euclidian distance, phenotypic differentiation between dwarf and normal Whitefish was significant according to the five within-lake MANOVAs (Table 2). Effect of “sex” also was marginally significant in Témiscouata Lake. The effect of “sex” on shape in laboratory-raised fish was also significant (df: 1, 71; Pillai = 0.57; approx F: 3.55; P -value < 0.001). In comparison with males, females have smaller eyes, a deeper body, a shorter tail, and a more anterior insertion of the pectoral fin (Figure S1). Given this and also the fact that previous transcriptomics studies on Whitefish revealed a pronounced sex effect on expression QTL (Derôme *et al.* 2008; Whiteley *et al.* 2008), the identification of shape QTL was performed by considering sex as a cofactor.

Whitefish shape QTL: laboratory-reared Whitefish

QTL analysis detected 138 QTL that were significant at the LG level, including 26 that also were significant at the genome-wide level

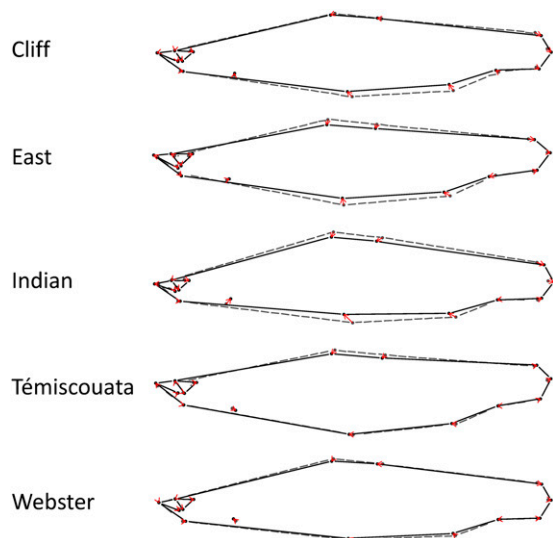


Figure 3 Mean shape comparisons between normal (white dot; dashed line) and dwarf Whitefish (black dot; full line) for Cliff, East, Indian, Témiscouata, and Webster lakes. Red arrows show how shape changes from normal to dwarf Whitefish.

(Table S1). There was an average of 3.40 (SD \pm 2.57) QTL per LG and 5.67 (SD \pm 1.86) QTL per shape coordinate (Table S1). Only three of the 40 LGs did not contain any shape QTL (LG20, 22, and 30) and all shape coordinates had a minimum of 3 QTL (Table S1). The average distance between a QTL and a RAD marker was 0.17 (SD \pm 0.43) cM (Table S1).

Annotation of shape QTL: laboratory-raised Whitefish

Of all 138 SNPs linked to QTL, 113 (81.9%) mapped against the draft assembly of the Lake Whitefish genome. Among these, 35 SNPs could be annotated unambiguously (Table S2). Three SNPs are of particular interest, as they are potentially under selection and involved in development: i) the collagen alpha-1 (XXVII) chain B (RAD marker 3771), ii) the microtubule-actin cross-linking factor 1 (RAD marker 110970), and iii) the nesprin 1 (RAD marker 123874).

Selection and parallelism in shape QTL: wild Whitefish

A total of 19 of the 138 SNPs that were associated with body shape QTL were significant outliers between dwarf and normal species for at least one lake after corrections for multiple tests with the single-locus Fdist analysis, suggesting that their level of divergence between dwarf and normal Whitefish may have been influenced by divergent selection. These were distributed over many linkage groups (Table 3). An average of four outlier loci were found in each lake (four loci in Cliff, seven in East, five in Indian, two in Témiscouata, and two in Webster, Table 3). Only one locus showed parallelism for two lakes (Indian and Webster); LG 12, position 38.2 (RAD marker 69063; Table 3). Two loci separated by only 1.2 cM showed potential divergent selection between dwarf and normal Whitefish from two lakes (Cliff and Indian); LG 21, position 81 and 82.2 (RAD marker 107544 and 37687; Table 3).

The polygenic Random Forest algorithm produced on the five lakes identified 33 important markers. The proportion of votes differentiated individuals of both species in Cliff, Indian, and Webster lakes, but not in East and Témiscouata lakes (Figure 4A). This finding suggests that these three lakes shared a common genetic architecture underlying morphologic differentiation between dwarf and normal

Table 2 Effect of both “species” (dwarf vs. normal) and “lake” on fish shape

Effect of	DF	Approx F	P-value
All lakes^a			
Species	1, 215	161.25	<0.001*
Lake	4, 432	11.97	<0.001*
Sex	1, 215	2.61	0.076**
Species \times lake	4, 432	2.75	0.006*
Species \times sex	1, 215	0.37	0.691
Lake \times sex	4, 432	2.14	0.031*
Species \times lake \times sex	4, 432	0.99	0.444
Within lake^b			
Cliff			
Species	1, 29	48.10	<0.001*
Sex	1, 29	0.67	0.519
Species \times sex	1, 29	0.79	0.465
East			
Species	1, 55	40.13	<0.001*
Sex	1, 55	1.83	0.171
Species \times sex	1, 55	0.28	0.756
Indian			
Species	1, 35	90.05	<0.001*
Sex	1, 35	2.26	0.119
Species \times sex	1, 35	1.86	0.171
Témiscouata			
Species	1, 48	17.56	<0.001*
Sex	1, 48	3.15	0.051*
Species \times sex	1, 48	1.13	0.332
Webster			
Species	1, 44	27.41	<0.001*
Sex	1, 44	1.33	0.275
Species \times sex	1, 44	1.18	0.317

MANOVA, multivariate analysis of variance.

^a Effect of species identity (Dwarf vs. Normal), lake, sex, and their interaction on the two first axes of a PCA on 15 landmarks representing fish shape tested with a (MANOVA).

^b Five independent MANOVAs (one for each lake) testing for the effect of species identity, sex and their interaction on the same response variables

* Significant result (P-value < 0.05).

** Marginally significant result (P-value < 0.1)

Whitefish (*i.e.*, similar genotypes on the 138 shape QTL allowing one to differentiate the two species) that is not shared with the other two lakes. The overall Random Forest classification error rate was 24.3% and was concentrated mainly in East and Témiscouata lakes according to the proportion of votes (Figure 4A). Given this, we ran the Random Forest algorithm two more times; i) on the species pairs from Cliff, Indian, and Webster lakes, to test whether the classification error rate will decrease when considering these three lakes only and ii) on the species pairs from East and Témiscouata to test whether genetic parallelism specific to these two species pairs is present. We found a classification error rate of only 5.9% when considering Cliff, Indian and Webster lakes and 38 important markers (Figure 4B). The mean F_{ST} for these 38 important markers was low, averring 0.08. Moreover, the mean F_{ST} between dwarf and normal Whitefish from Cliff, Indian, and Webster lakes was significantly greater for RAD markers under potential divergent selection ($n = 10$; four from Cliff, four from Indian, one from Webster, and one shared between Indian and Webster) than for the 38 important markers among the three lakes ($F_{ST} = 0.32$ vs. 0.08; $t = -4.94$, P-value < 0.001). Only three markers were shared between the two approaches. Yet, there was a trend for the mean PVE for individual marker to be greater for the important markers (PVE = 4.85%) than for the outliers potentially under divergent selection

■ **Table 3** Description of SNP markers potentially under divergent selection in wild populations of Lake Whitefish as revealed by Fdist analysis

SNP	Associated QTL		F_{ST} of Loci Under Potential Divergent Selection in Lake				
	LG	Position, cM	Cliff (0.22)	East (0.03)	Indian (0.11)	Témiscouata (0.01)	Webster (0.05)
46086	1	20.0	–	–	0.49	–	–
78628	4	80.3	–	–	–	0.13	–
147541	4	92.3	–	0.22	–	–	–
107600	6	16.0	–	0.27	–	–	–
110970	10	37.0	–	0.24	–	–	–
132516	12	27.1	–	0.26	–	–	–
69063	12	38.2	–	–	0.67	–	0.37
33001	16	60.4	0.96	–	–	–	–
123874	17	51.5	–	–	–	–	0.31
107544	21	81.0	–	–	0.67	–	–
37687	21	82.2	0.39	–	–	–	–
1790	25	29.1	–	0.17	–	–	–
35278	26	36.7	–	–	0.44	–	–
88462	28	38.3	–	0.27	–	–	–
101670	31	10.8	0.77	–	–	–	–
3771	31	14.9	–	–	–	0.27	–
74955	34	36.2	0.78	–	–	–	–
45298	36	19.0	–	0.28	–	–	–
71661	39	21.9	–	–	0.68	–	–

The LG as defined in Gagnaire *et al.* (2013a) and position of the body shape–associated QTL are indicated. In addition, F_{ST} values between dwarf and normal Whitefish in the lake in which potential selection was detected are indicated. The mean F_{ST} value for divergence between dwarf and normal Whitefish for 3438 SNPs is indicated in parentheses under the lake names. In bold, a marker (69063) that shows potential genetic parallelism between Indian and Webster species pairs and two others (107544 and 37687) physically ‘close’ (1.2 cM) on the genetic map that could represent another case of genetic parallelism between Cliff and Indian species pairs. SNP, single-nucleotide polymorphism; QTL, quantitative trait loci; LG, linkage group.

(PVE = 3.07%), although this difference was not statistically significant ($t = 1.58$, P -value = 0.131). In addition, important markers obtained from Indian and Webster species pairs allowed assigning correctly 100% of the individuals from Cliff lake. Similarly, analysis from Cliff and Webster species pairs assigned correctly 95% of the individuals from Indian lake and analysis from Cliff and Indian species pairs correctly assigned 85% of the individuals from Webster lake. Thus, an average of 93.3% of individuals were correctly assigned when not including a given species pair for identifying important markers. In contrast, genetic parallelism appears to be absent between East and Témiscouata lakes with a classification error rate of 40.5% (Figure 4C, number of important markers = 4, see Table S1).

DISCUSSION

We found evidence for parallel body shape differentiation among dwarf and normal Lake Whitefish in five different lakes. In all cases dwarf fish have larger eyes, more slender bodies, and longer tails compared with the benthic normal species. These differences are consistent with expectations regarding adaptation to the limnetic niche (Langerhans and Reznick 2010; Willacker *et al.* 2010). We also identified QTL underlying variation in body shape, thus revealing a genetic basis for shape variation, although this does not exclude by any mean an environmental effect on shape variation as well (in this section, to follow) as this would be the case for most phenotypic traits in any species. Moreover, the polygenic approach used to detect a group of covarying markers that differentiated the two species showed evidence of genetic parallelism in three of the five lakes (Cliff, Indian, and Webster) we studied. A low mean $F_{ST} = 0.08$ of these covarying markers suggests a recent polygenic adaptation and high percentages of correct assignment, when a species pairs was not included to identify important markers, also support the occurrence of genetic parallelism underlying phenotypic parallelism among these three

lakes. In contrast, there was little evidence for genetic parallelism with the single-locus approach that detects outlier under potential divergent selection, as only two genomic regions associated with a body shape QTL showed parallelism between at least two of the five lakes (LG 12, Position 38.2: Indian and Webster and LG 21, Position 81.0 to 82.2: Cliff and Indian). Interestingly, no genetic parallelism was observed between East and Témiscouata Lakes, suggesting a possibility of three different genetic paths that have led to the evolution of dwarf-normal Whitefish species pairs in this system (1: Cliff, Indian and Webster; 2: East and 3: Témiscouata).

Two non-mutually exclusive hypotheses may explain the observation of variation in the extent of parallelism between the phenotype and the genotype: i) a polygenic architecture in which different groups of loci contribute to phenotypic differentiation among lakes and ii) environmental influences on the phenotype which differ among lakes leading to phenotypic plasticity. The high number of shape-related QTL found in the BC family support the polygenic architecture hypothesis since all fish were reared in identical conditions. In addition, quantitative genetic theory predicts that polygenic methods may be better at exploring the genetic basis of complex traits such as body shape (Le Corre and Kremer 2012). For instance, as observed in Bourret *et al.* (2014) in Atlantic salmon, the polygenic analysis revealed genetic parallelism associated with differential mortality whereas single-locus outlier method did not, and recent studies using Random Forest algorithm show high power to detect polygenic selection (Briec *et al.* 2015; Pavay *et al.* 2015). Body shape differentiation in Lake Whitefish may therefore be explained by a complex polygenic architecture and the combined effects of relatively modest allelic frequency changes, as reflected by the low mean F_{ST} observed at the markers identified by the Random Forest analysis. As mentioned previously, the results of these analyses suggest that phenotypic differentiation observed in this study may have evolved through three

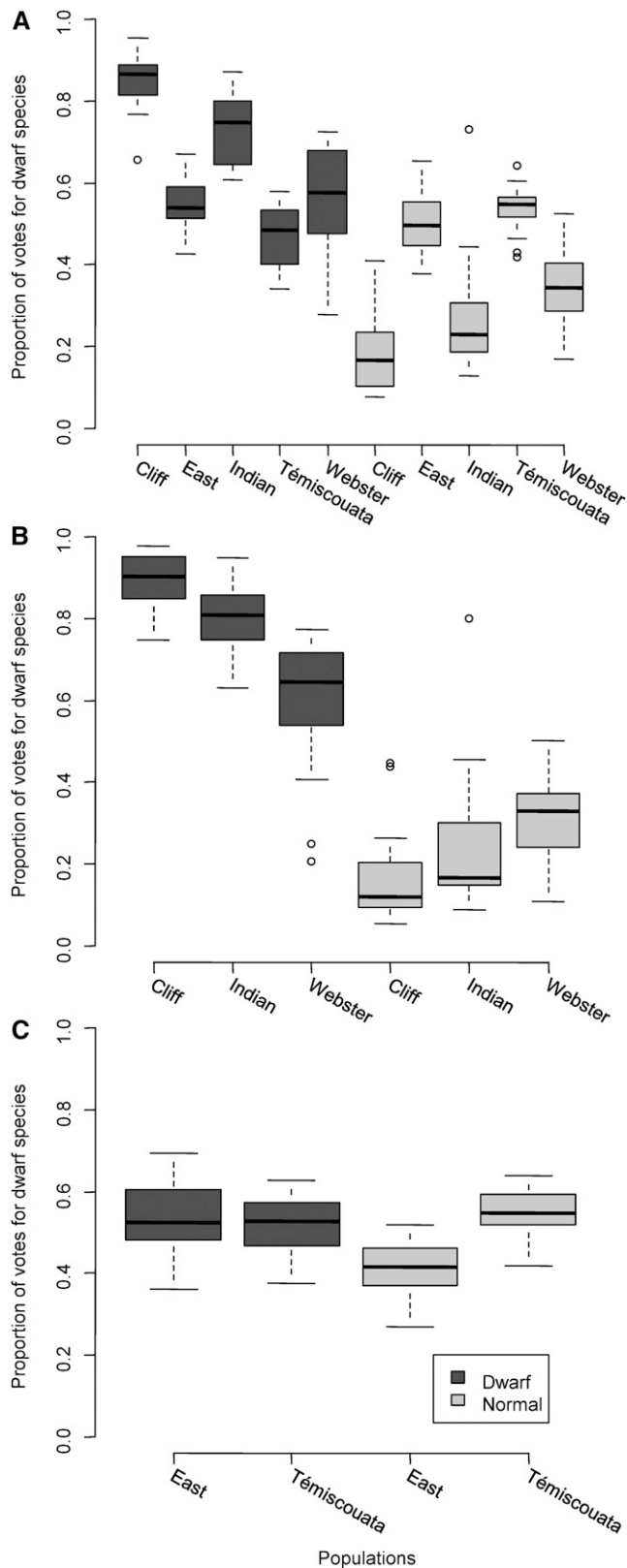


Figure 4 Boxplot of the proportion of “votes” for Whitefish from dwarf populations (dark gray) and normal populations (light gray) (x-axis) for the dwarf species group (y-axis) obtained via 1,000,000 trees produced with random forest analyses on 138 shape-related QTL. A “vote” refers the classification of an individual by a single tree. (A) All 10 populations from

different patterns of subtle allelic frequency changes in i) Témiscouata, ii) East, and iii) Cliff/Indian/Webster lakes. Historic contingency offers an explanation for the presence of these three potential genetic pathways found for body shape differentiation as Cliff, Indian, and Webster lakes are closer geographically. In addition, their environments may have been more similar when independent secondary contacts occurred between the Acadian and Atlantic glacial lineages in those lakes (Bernatchez and Dodson 1990). Evidence also suggests that the sympatric pairs from East Lake may have evolved from a single glacial lineage, unlike the other lakes studied (Pigeon *et al.* 1997; Rico *et al.* 2013). Moreover, gene flow between Témiscouata species pairs is the highest documented among the five lakes (Gagnaire *et al.* 2013b; Rico *et al.* 2013) and could result in a loss of genetic differentiation at some loci that are related to body shape differentiation. In addition to a polygenic architecture, phenotypic plasticity evolves following adaptive divergence in new environments and could therefore contribute to an increase in phenotypic parallelism among lakes, in addition to the polygenic basis for body shape that we have identified here (Aubin-Horth and Renn 2009; Angers *et al.* 2010; Pfennig *et al.* 2010, Morris *et al.* 2014).

We identified a total of 138 loci (including 26 at genome-wide significance) that affected at least one body shape coordinate. On average, each phenotypic trait is associated with more than five QTL (between 3 and 11). These data indicate a highly polygenic architecture for body shape in Lake Whitefish which agrees i) with our previous findings of a polygenic basis underlying the expression of other complex ecologically important traits in Lake Whitefish such as growth rate, age at maturity, and behavior (Rogers and Bernatchez 2007; Filteau *et al.* 2013; Gagnaire *et al.* 2013a). This is also congruent with other studies revealing polygenic architecture of shape and other morphometrics (Albert *et al.* 2008; Cooper *et al.* 2011; Wang *et al.* 2011; Hecht *et al.* 2012; Rogers *et al.* 2012; Franchini *et al.* 2014). Also, it must be considered that QTL detected in this study resulted from crossing a normal Whitefish from Aylmer Lake not involved in wild populations considered in this study. This is because it was not possible to obtain wild sexually mature normal fish from the studied populations. Therefore, some QTL found in this study could be cross specific and not segregate in other populations. However, we are confident that this bias is not major in the detection of parallelism among our samples. Namely, previous QTL studies involving other phenotypic traits in this system (*e.g.*, growth) using this same cross and comparing with natural populations revealed the occurrence of parallelism for the same segregating QTL (Rogers and Bernatchez 2007). If anything, this render our interpretations of parallelism more conservative given that we are detecting parallelism despite the possible confounding effect of variation in genetic architecture among populations. To our defense also, very few studies have actually built genetic maps for each individual wild populations being investigated in similar types of studies as performed here.

Functional morphology provides clear *a priori* expectations for the effects of changes in body shape. A slender, fusiform body shape and caudal peduncle diminish drag and can reduce the energetic costs of prolonged swimming in fishes (Webb 1982, 1984; Langerhans and Reznick 2010; Walker 2010; Willacker *et al.* 2010). Thus, we predicted

five lakes—classification error rate of 24.3%; (B) six of these populations from three lakes (Cliff, Indian, and Webster)—classification error rate of 5.9% and (C) the four remaining populations from two lakes (East and Témiscouata)—classification error rate of 40.5%.

that the dwarf Whitefish species would evolve a more slender shape. This is because the dwarf Whitefish forages on zooplankton in open water, which shows greater variation in abundance and distribution in lakes in comparison to benthic prey (Del Giorgio and Gasol 1995). Moreover, the Lake Trout (*Salvelinus namaycush*), common in all lakes surveyed, is a main predator of Lake Whitefish (Scott and Crossman 1973; Chouinard *et al.* 1996), and is more likely to chase the dwarf Whitefish for longer distances in the greater luminosity of the limnetic environment (Vogel and Beauchamp 1999). Previous studies in controlled experimental conditions also showed that dwarf Whitefish exhibit genetically-based differences in swimming behavior in accordance with expectations based on their ecology (Rogers *et al.* 2002; Rogers and Bernatchez 2007). However, the dwarf Whitefish caudal peduncle depth was not more slender than in the normal species. Since a stout peduncle better powers “fast-start burst swims” (Webb 1982, 1984; Langerhans and Reznick 2010; Walker 2010), it is possible that dwarf Whitefish cannot maximize drag reduction because this may come at the cost of burst swimming performance. Therefore, a slender body but a similar peduncle depth in comparison with the normal species could represent a trade-off to minimize the costs of foraging and predation risk and maintain adequate burst and prolonged swimming capacities. This hypothesis will need to be tested by measuring swimming performance in dwarf and normal Whitefish in future experiments.

Among the annotated SNPs associated with body shape QTL, three are of particular interest because they are found in genes with well-known functions in vertebrate development and are potentially under divergent selection: MACF1, Col27a1b, and Nesprin-1. The microtubule-actin cross-linking factor 1 (MACF1; RAD marker 110970) is highly expressed in mouse neurons and skeletal muscle during embryonic development (Barsi *et al.* 2005; Chen 2006; Koo *et al.* 2007) and regulates animal-vegetal polarity in the zebrafish oocyte (Gupta *et al.* 2010). It has a role in the activation of the canonical Wnt signaling pathway (Wnt/ β -catenin pathway), which is involved in embryo patterning and the determination of cell fate through transcriptional activation of target genes, among other functions (Chen 2006). The collagen alpha-1(XXVII) chain B (Col27a1b) (RAD marker 3771) is expressed in the notochord and cartilage of zebrafish embryo. The knockout of this protein results in curvature of the notochord and scolioses in zebrafish embryos. It is thus involved in notochord morphogenesis and axial skeletogenesis (Christiansen *et al.* 2009). Finally, Nesprin-1 (RAD marker 123874) is involved in muscle cell differentiation, nuclear positioning and anchorage (Zhang *et al.* 2001, 2010). Nesprin-1 knockout in mice results in a decrease in survival rates, growth, and exercise capacity and increased variability in body weight (Zhang *et al.* 2010). All of these traits are potentially linked with the phenotypic differentiation observed between Lake Whitefish wild species pairs. As mentioned previously, these genes were all under potential divergent selection in at least one lake (MACF1: East; Col27a1b: Témiscouata; Nesprin-1: Webster) and they are also mistranscribed in malformed hybrid backcrossed Whitefish embryos relative to pure parental forms (Dion-Côté *et al.* 2014). The role of these candidate genes during development suggests that shape could be genetically determined early during the embryonic stage in Lake Whitefish. This finding is supported by the presence of a pronounced shape differentiation between young juveniles of both species raised in common garden (M. Laporte, unpublished data). Of the 35 annotated QTL, 11 corresponded to transposable elements. This is noteworthy as transposable elements are recognized as “powerful facilitators of evolution and phenotypic diversity” (Oliver and Greene 2009) through their transcriptional impact on gene networks (Feschotte 2008). Hence, our results raise the possibility that transposable elements could affect

the expression of key genes that are responsible for fish shape determination in the Lake Whitefish. Incidentally, a recent study in Lake Whitefish revealed a reactivation of the expression of transposable elements in backcrossed hybrids which is associated with phenotypic malformation of hybrids during their early development (Dion-Côté *et al.* 2014). Although the effects of these alleles on fish shape in Lake Whitefish embryos are unknown, these proteins and transposable elements are candidates for future studies aiming to decipher the molecular basis of shape differentiation in the Lake Whitefish system.

In conclusion, we found a pronounced pattern of phenotypic parallelism in body shape between dwarf and normal Whitefish in five different species pairs, which fitted the *a priori* predictions based on the known ecology of dwarf and normal Lake Whitefish. This brings support to the role for natural selection in driving body shape differences between these forms. Body shape was also shown to be under polygenic control and genotypic parallelism appears to be present in three of the five lakes as revealed by a polygenic analytical approach. In comparison, the single-locus approach identified very limited cases of genetic parallelism in body shape. We also identified candidate genes to be further investigated and that could contribute to fish shape differentiation based on functional annotation. Overall, our results support the view that both multiple genetic routes and genetic parallelism can occur during the evolution of fish shape to produce phenotypically similar adaptive changes in body shape when facing similar environmental challenges.

ACKNOWLEDGMENTS

We are grateful to K. Broman, G. Côté, M. Evans, A. Kusler, and S. A. Pavey for their invaluable logistic support and/or comments on earlier versions of the manuscript. We are also grateful to the staff of Maine Department of Inland Fisheries & Wildlife for their help in sampling. We are also grateful to T. Slotte and two anonymous referees for their constructive comments on a previous version of this manuscript. This research was supported by a Discovery research grant from the Natural Sciences and Engineering Research Council of Canada (NSERC) led by L.B., who holds the Canadian Research Chair in genomics and conservation of aquatic resources. M.L. was supported by a Postdoctoral fellowship from the Fonds de Recherche Nature et Technologies du Québec (FQRNT), SMR is supported by an Alberta Innovates Technology Futures New Faculty Award and an NSERC Discovery Grant Award. A.-M.D.-C. and A.C.D. received, respectively, a NSERC postgraduate and postdoctoral scholarship and A.-M.D.-C. also received a stipend from Québec-Océan.

LITERATURE CITED

- Albert, A. Y. K., A. Sawaya, T. H. Vines, A. K. Knecht, C. T. Miller *et al.*, 2008 The genetic of adaptive shape shift in stickleback: pleiotropy and effect size. *Evolution* 62: 76–85.
- Angers, B., E. Castonguay, and R. Massicotte, 2010 Environmentally induced phenotypes and DNA methylation: how to deal with unpredictable conditions until the next generation and after. *Mol. Ecol.* 19: 1283–1295.
- Aubin-Horth, N., and S. C. P. Renn, 2009 Genomic reaction norms: using integrative biology to understand molecular mechanisms of phenotypic plasticity. *Mol. Ecol.* 18: 3763–3780.
- Barsi, J. C., R. Rajendra, J. I. Wu, and K. Artzt, 2005 Mind bomb1 is a ubiquitin ligase essential for mouse embryonic development and Notch signaling. *Mech. Dev.* 122: 1106–1117.
- Benjamini, Y., and Y. Hochberg, 1995 Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J. R. Stat. Soc., B* 57: 289–300.
- Bernatchez, L., 2004 Ecological theory of adaptive radiation: an empirical assessment from coregonine fishes (Salmoniformes). *Evolution Illuminated:*

- Salmon and Their Relatives*, edited by Hendry, A. P., and S. C. Stearns. Oxford University Press, Oxford, UK.
- Bernatchez, L., and J. J. Dodson, 1990 Allopatric origin of sympatric populations of Lake Whitefish (*Coregonus clupeaformis*) as revealed by mitochondrial DNA restriction analysis. *Evolution* 44: 1263–1271.
- Bernatchez, L., S. Renaut, A. R. Whiteley, D. Campbell, N. Derôme *et al.*, 2010 On the origin of species: insights from the ecological genomics of Whitefish. *Philos. Trans. R. Soc. Lond.* 365: 1783–1800.
- Berner, D., 2012 How much can the orientation of G's eigenvectors tell us about genetic constraints? *Ecol. Evol.* 2: 1834–1842.
- Boulesteix, A.-L., S. Janitzka, J. Kruppa, and I. R. König, 2012 Overview of random forest methodology and practical guidance with emphasis on computational biology and bioinformatics. *WIREs Data Min. Knowl. Discov.* 2: 493–507.
- Bourret, V., M. Dionne, and L. Bernatchez, 2014 Detecting genotypic changes associated with selective mortality at sea in Atlantic salmon: polygenic multilocus analysis surpasses genome scan. *Mol. Ecol.* 23: 4444–4457.
- Briec, M. S. O., K. Ono, D. Drinan, and K. A. Naish, 2015 Integration of Random Forest with population-based outlier analyses to provides insight on the genomic basis and evolution of run timing in Chinook salmon (*Oncorhynchus tshawytscha*). *Mol. Ecol.* (in press).
- Broman, K. W., H. Wu, S. Sen, and G. A. Churchill, 2003 R/qtl: QTL mapping in experimental crosses. *Bioinform.* 19: 889–890.
- Chen, H. J., 2006 The role of microtubule actin cross-linking factor 1 (MACF1) in the Wnt signaling pathway. *Genes Dev.* 20: 1933–1945.
- Chen, X., and H. Ishwaran, 2012 Random forests for genomic data analysis. *Genomics* 99: 323–329.
- Chouinard, A., D. Pigeon, and L. Bernatchez, 1996 Lack of specialization in trophic morphology between genetically differentiated dwarf and normal forms of Lake Whitefish (*Coregonus clupeaformis*, Mitchill) in Lac de l'Est, Québec. *Can. J. Zool.* 74: 1989–1998.
- Christiansen, H. E., M. R. Lang, J. M. Pace, and D. M. Parichy, 2009 Critical early roles for col27a1a and col27a1b in zebrafish notochord morphogenesis, vertebral mineralization and post-embryonic axial growth. *PLoS One* 4: e8481.
- Claude, J., 2008 *Morphometrics with R*. Springer Science + Business Media, L.L.C., New York, NY.
- Conte, G. L., M. E. Arnegard, C. L. Peichel, and D. Schluter, 2012 The probability of genetic parallelism and convergence in natural populations. *Proc. Biol. Sci.* 279: 5039–5047.
- Cooper, W. J., J. Wernle, K. Mann, and R. C. Albertson, 2011 Functional and genetic integration in the skulls of Lake Malawi cichlids. *Evol. Biol.* 38: 316–334.
- Cordell, H. J., 2009 Detecting gene–gene interactions that underlie human diseases. *Nat. Rev. Genet.* 10: 392–404.
- Del Giorgio, P. A., and J. M. Gasol, 1995 Biomass distribution in freshwater plankton communities. *Am. Nat.* 146: 135–152.
- Derôme, N., B. Bougas, S. M. Rogers, A. Whiteley, A. Labbe *et al.*, 2008 Pervasive sex-linked effects on transcription regulation as revealed by eQTL mapping in Lake Whitefish species pairs (*Coregonus* sp, Salmonidae). *Genetics* 179: 1903–1917.
- Dion-Côté, A.-M., S. Renaut, E. Normandeau, and L. Bernatchez, 2014 RNA-seq reveals transcriptomic shock involving transposable elements reactivation in hybrids of young Lake Whitefish species. *Mol. Biol. Evol.* 31: 1188–1199.
- Dryden, I. L., and K. V. Mardia, 1998 *Statistical Shape Analysis*. Wiley, New York, NY.
- Elmer, K. R., and A. Meyer, 2011 Adaptation in the age of ecological genomics: insights from parallelism and convergence. *Trends Ecol. Evol.* 26: 298–306.
- Elmer, K. R., S. Fan, H. Kusche, M. L. Spreitzer, A. F. Kautt *et al.*, 2014 Parallel evolution of Nicaraguan crater lake cichlid fishes via non-parallel routes. *Nat. Commun.* 5: 5168.
- Endler, J. A., 1986 *Natural Selection in the Wild*. Princeton University Press, Princeton, NJ.
- Evans, M. L., and L. Bernatchez, 2012 Oxydative phosphorylation gene transcription in Whitefish species pairs reveals patterns of parallel and non-parallel physiological divergence. *J. Evol. Biol.* 25: 1823–1834.
- Evans, M. L., K. Præbel, S. Peruzzi, and L. Bernatchez, 2012 Parallelism in the oxygen transport system of the Lake Whitefish: the role of physiological adaptive divergence in ecological speciation. *Mol. Ecol.* 21: 4038–4050.
- Evans, M. L., L. J. Chapman, I. Mitrofanov, and L. Bernatchez, 2013 Variable extent of parallelism in respiratory, circulatory, and neurological traits across Lake Whitefish species pairs. *Ecol. Evol.* 3: 546–557.
- Excoffier, L., 2010 Arlequin suite ver. 3.5: A new series of programs to perform population genetics analyses under Linux and Windows. *Mol. Ecol. Res.* 10: 564–567.
- Excoffier, L., T. Hofer, and M. Foll, 2009 Detecting loci under selection in a hierarchically structured population. *Heredity* 103: 285–298.
- Feschotte, C., 2008 Transposable elements and the evolution of regulatory networks. *Nat. Rev. Genet.* 9: 397–405.
- Filteau, M., S. A. Pavey, J. St-Cy, and L. Bernatchez, 2013 Gene co-expression networks reveal key drivers of phenotypic divergence in Lake Whitefish. *Mol. Biol. Evol.* 30: 1384–1396.
- Franchini, P., C. Fruciano, M. L. Spreitzer, J. C. Jones, K. R. Elmer *et al.*, 2014 Genomic architecture of ecologically divergent body shape in a pair of sympatric crater lake cichlid fishes. *Mol. Ecol.* 23: 1828–1845.
- Gagnaire, P. -A., E. Normandeau, S. A. Pavey, and L. Bernatchez, 2013a Mapping phenotypic, expression and transmission ratio distortion QTL using RAD markers in the Lake Whitefish (*Coregonus clupeaformis*). *Mol. Ecol.* 22: 3036–3048.
- Gagnaire, P. -A., S. A. Pavey, E. Normandeau, and L. Bernatchez, 2013b The genetic architecture of reproductive isolation during speciation-with-gene-flow in Lake Whitefish species pairs assessed by RAD-sequencing. *Evolution* 67: 2483–2497.
- Gerstein, A. C., D. S. Lo, and S. P. Otto, 2012 Parallel genetic changes and non-parallel gene-environment interactions characterize the evolution of drug resistance in yeast. *Genetics* 192: 241–252.
- Goldstein, B. A., E. C. Polley, and F. B. S. Briggs, 2011 Random forests for genetic association studies. *Stat. Appl. Genet. Mol. Biol.* 10: 32.
- Gompel, N., and B. Prud'homme, 2009 The causes of repeated genetic evolution. *Dev. Biol.* 332: 36–47.
- Gupta, T., F. L. Marlow, D. Ferriola, K. Mackiewicz, J. Daprich *et al.*, 2010 Microtubule actin crosslinking factor 1 regulates the Balbiani body and animal-vegetal polarity of the zebrafish oocyte. *PLoS Genet.* 6: e1001073.
- Haley, C. S., and S. A. Knott, 1992 A simple regression method for mapping quantitative trait loci in line crosses using flanking markers. *Heredity* 69: 315–324.
- Harrod, C., J. Mallela, and K. K. Kahilainen, 2010 Phenotype-environment correlations in a putative whitefish adaptive radiation. *J. Anim. Ecol.* 79: 1057–1068.
- Hecht, B. C., F. P. Thrower, M.C. Hale, M.R. Miller, and K.M. Nichols, 2012 Genetic architecture of migration-related traits in rainbow and stellhead trout, *Oncorhynchus mykiss*. *G3 (Bethesda)* 2: 1113–1127.
- Heron, M. D., and M. Doebli, 2013 Parallel evolutionary dynamics of adaptive diversification in *Escherichia coli*. *PLoS Biol.* 11: e1001490.
- Jarquín, D., K. Kocak, L. Posadas, K. Hyma, J. Jedlicka *et al.*, 2014 Genotyping by sequencing for genomic prediction in a soybean breeding population. *BMC Genomics* 15: 740.
- Kemper, K. E., S. J. Saxton, S. Bolormaa, B. J. Hayes, and M. E. Goddard, 2014 Selection for complex traits leaves little or no classic signatures of selection. *Genomics* 15: 246.
- Klingenberg, C. P., 2010 Evolution and development of shape: integrating quantitative approach. *Nat. Rev. Genet.* 11: 623–635.
- Koo, B. K., M. J. Yoon, K. J. Yoon, S. K. Im, Y. Y. Kim *et al.*, 2007 An obligatory role of mind bomb-1 in notch signaling of mammalian development. *PLoS One* 2: e1221.
- Landry, L., and L. Bernatchez, 2010 Role of epibenthic resource opportunities in the parallel evolution of Lake Whitefish species pairs (*Coregonus* sp.). *J. Evol. Biol.* 23: 2602–2613.

- Landry, L., W. F. Vincent, and L. Bernatchez, 2007 Parallelism between limnological features and phenotypic evolution of Lake Whitefish dwarf ecotypes. *J. Evol. Biol.* 20: 971–984.
- Langerhans, R. B., and D. N. Reznick, 2010 Ecology and evolution of swimming performance in fishes: predicting evolution with biomechanics, pp. 200–248 in *Fish Locomotion: an Etho-Ecological Perspective*, edited by Domenici, P., and B. G. Kapoor. Science Publishers, Enfield, NH.
- Laporte, M., P. Magnan, and B. Angers, 2011 Genetic differentiation between the blue and the yellow phenotypes of walleye (*Sander vitreus*): an example of parallel evolution. *Ecoscience* 18: 124–129.
- Le Corre, V., and A. Kremer, 2012 The genetic differentiation at quantitative trait loci under local adaptation. *Mol. Ecol.* 21: 1548–1566.
- Legendre, P., and L. Legendre, 1998 *Numerical Ecology*. Second English Ed. Elsevier, Amsterdam.
- Liam, A., and M. Wiener, 2002 Classification and regression by random-Forest. *R News*. 2: 18–22.
- Losos, J. B., 2011 Convergence, adaptation, and constraint. *Evolution* 65: 1827–1840.
- Lu, G., and L. Bernatchez, 1999 Correlated trophic specialization and genetic divergence in sympatric Lake Whitefish ecotypes (*Coregonus clupeaformis*): support for the ecological speciation hypothesis. *Evolution* 53: 1491–1505.
- Lu, G., D. J. Basley, and L. Bernatchez, 2001 Contrasting patterns of mitochondrial DNA and microsatellite introgressive hybridization between lineages of Lake Whitefish (*Coregonus clupeaformis*): relevance for speciation. *Mol. Ecol.* 10: 965–985.
- Maynard Smith, J., and J. Haigh, 1974 The hitch-hiking effect of a favourable gene. *Genet. Res.* 23: 23–35.
- McKay, J. K., and R. G. Latta, 2002 Adaptive population divergence: markers, QTL and traits. *Trends Ecol. Evol.* 17: 285–291.
- McPhail, J. D., 1984 Ecology and evolution of sympatric sticklebacks (*Gasterosteus*): Morphological and genetic evidence for a species pair in Enos Lake, British Columbia. *Can. J. Zool.* 62: 1402–1408.
- Merilä, J., 2014 Lakes and ponds as model systems to study parallel evolution. *J. Limnol.* 73: 33–45.
- Messer, P. W., and D. A. Petrov, 2013 Population genomics of rapid adaptation by soft selective sweeps. *Trends Ecol. Evol.* 28: 659–669.
- Mokry, F. B., R. H. Higa, M. A. Mudadu, A. O. Lima, S. L. C. Meirelles *et al.*, 2013 Genome-wide association study for backfat thickness in Canchim beef cattle using Random Forest approach. *BMC Genet.* 14: 47.
- Morris, M. R. J., R. Richard, E. H. Leder, R. D. H. Barrett, N. Aubin-Horth *et al.*, 2014 Gene expression plasticity facilitates and evolves in response to colonization of freshwater lakes in threespine stickleback. *Mol. Ecol.* 23: 3226–3240.
- Nosil, P., 2012 *Ecological Speciation*. Oxford University Press, Oxford.
- Oliver, K. R., and W. K. Greene, 2009 Transposable elements: powerful facilitators of evolution. *BioEssays* 31: 703–714.
- Østbye, K., P. A. Amundsen, L. Bernatchez, A. Klemetsen, R. Knudsen *et al.*, 2006 Parallel evolution of ecomorphological traits in the European Whitefish *Coregonus lavaretus* (L.) species complex during postglacial times. *Mol. Ecol.* 15: 3983–4001.
- Ozerov, M. Y., M. Himberg, T. Aykanat, D. S. Sendek, H. Hägerstrand *et al.*, 2015 Generation of a neutral FST baseline for testing local adaptation on gill raker number within and between European whitefish ecotypes in the Baltic Sea basin. *J. Evol. Biol.* 28: 1170–1183.
- Pavey, S. A., J. Gaudin, E. Normandeau, M. Dionne, M. Canstonguay *et al.*, 2015 RAD-sequencing highlights polygenic discrimination of habitat ecotypes in the panmictic American Eel. *Curr. Biol.* DOI: 10.1016/j.cub.2015.04.062.
- Pfennig, D. W., M. A. Wund, E. C. Snell-Rood, T. Cruickshank, C. D. Schlichting *et al.*, 2010 Phenotypic plasticity's impacts on diversification and speciation. *Trends Ecol. Evol.* 25: 459–467.
- Pigeon, D. A., A. Chouinard, and L. Bernatchez, 1997 Multiple modes of speciation involved in the parallel evolution of sympatric morphotypes of Lake Whitefish (*Coregonus clupeaformis*). *Evolution* 51: 196–205.
- Piry, S., A. Alapetite, J.-M. Cornuet, D. Paetkau, and A. Estoup, 2004 GENECLASS2: A software for genetic assignment and first-generation migrant detection. *J. Hered.* 95: 536–539.
- Poland, J., J. Endelman, J. Dawson, J. Rutkoski, S. Wu *et al.*, 2012 Genomic selection in wheat breeding using genotyping-by-sequencing. *Plant Genome*. 5: 103.
- Pritchard, J. K., J. K. Pickrell, and G. Coop, 2010 The genetics of human adaptation: Hard sweeps, soft sweeps, and polygenic adaptation. *Curr. Biol.* 20: 208–215.
- Proulx, R., and P. Magnan, 2004 Contribution of phenotypic plasticity and heredity to the trophic polymorphism of lacustrine brook charr (*Salvelinus fontinalis* M.). *Evol. Ecol. Res.* 6: 503–522.
- R Core Team., 2014 R version 3.0.3. The R Foundation for Statistical Computing. Available at: <http://cran.r-project.org/bin/windows/base/old/3.0.3/>. Accessed May 26, 2015.
- Rainey, P. B., and M. Travisano, 1998 Adaptive radiation in a heterogeneous environment. *Nature* 32: 69–72.
- Rico, R., E. Normandeau, A.-M. Dion-Côté, M. I. Rico, G. Côté *et al.*, 2013 Combining next-generation sequencing and online databases for microsatellite development in non-model organisms. *Sci. Rep.* 3: 3376.
- Rogers, M. R., V. Gagnon, and L. Bernatchez, 2002 Genetically based phenotype-environment association for swimming behavior in Lake Whitefish ecotypes (*Coregonus clupeaformis* Mitchell). *Evolution* 56: 2322–2329.
- Rogers, S. M., and L. Bernatchez, 2007 The genetic architecture of ecological speciation and the association with signatures of selection in natural Lake Whitefish (*Coregonus* sp. Salmonidae) species pairs. *Mol. Biol. Evol.* 24: 1423–1438.
- Rogers, S. M., P. Tamkee, B. Summers, S. Balabhadra, M. Marks *et al.*, 2012 Genetic signature of adaptive peak shift in threespine stickleback. *Evolution* 68: 2439–2450.
- Rogers, S. M., J. A. Mee, and E. Bowles, 2013 The consequences of genomic architecture on ecological speciation in postglacial fishes. *Cur. Zool.* 59: 53–71.
- Rogers, S. M., and H. Jamniczky, 2014 Perspective: The shape of things to come in the study of the origin of species. *Mol. Ecol.* 23: 1650–1652.
- Rohlf, F. J., and D. Slice, 1990 Extensions of the Procrustes method for the optimal superimposition of landmarks. *Syst. Zool.* 39: 40–59.
- Rohlf, F. J., 2010 *tpsDig, Digitize Landmarks and Outlines, version 2.16*. Department of Ecology and Evolution, State University of New York at Stony Brook, Stony Brook, NY.
- Schlager, S., 2014 Morpho: Calculations and visualisations related to geometric morphometrics. The R Foundation for Statistical Computing. Available at: <http://cran.r-project.org/web/packages/Morpho/Morpho.pdf>. Accessed May 26, 2015.
- Schluter, D., 2000 *The Ecology of Adaptive Radiation*, Oxford University Press, Oxford.
- Scott, W. B., and E. J. Crossman, 1973 *Freshwater Fishes of Canada. Bulletin 184*. Fisheries Research Board of Canada, Ottawa, Ontario.
- Shi, T., D. Seligson, A. S. Beldegrun, A. Palotie, and S. Horvath, 2005 Tumor classification by tissue microarray profiling: random forest clustering applied to renal cell carcinoma. *Mod. Pathol.* 18: 547–557.
- Skúlason, S., and T. B. Smith, 1995 Resource polymorphisms in vertebrates. *Trends Ecol. Evol.* 10: 366–370.
- Strobl, C., J. Malley, and G. Tutz, 2009 An introduction to recursive partitioning: rational, applications, and characteristics and regression trees, bagging, and random forests. *Psych. Met.* 14: 323–348.
- Tang, R., J. P. Sinnwell, L. Jia, D. N. Ryder, M. de Aridrade *et al.*, 2009 Identification of genes and haplotypes that predict rheumatoid arthritis using random forests. *BMC Proc.* 3: S68.
- Taylor, E. B., 1999 Species pairs of north temperate freshwater fishes: evolution, taxonomy, and conservation. *Rev. Fish Biol. Fish.* 9: 299–334.
- Vogel, J. L., and D. A. Beauchamp, 1999 Effect of light, prey size, and turbidity on reaction distances of lake trout (*Salvelinus namaycush*) to salmonid prey. *Can. J. Fish. Aquat. Sci.* 56: 1293–1297.
- Walker, J. A., 2010 An integrative model of evolutionary covariance: a symposium on body shape in fishes. *Integr. Comp. Biol.* 50: 1051–1056.
- Wang, C. M., L. C. Lo, Z. Y. Zhu, H. Y. Pang, H. M. Liu *et al.*, 2011 Mapping QTL for an adaptive trait: the length of caudal fin in *Lates calcarifer*. *Mar. Biotechnol.* (NY) 13: 73–82.

- Webb, P. W., 1982 Locomotor patterns in the evolution of actinopterygian fishes. *Am. Zool.* 22: 329–342.
- Webb, P. W., 1984 Form and function in fish swimming. *Sci. Am.* 251: 58–68.
- Whiteley, A. R., N. Derôme, S. M. Rogers, J. St-Cyr, A. Labbe, and L. Bernatchez, 2008 The phenomics of brain transcriptomes regulating adaptive divergence in lake whitefish species pairs. *Genetics.* 180: 147–164.
- Willacker, J. J., F. A. Von Hippel, P. R. Wilton, and K. M. Walton, 2010 Classification of threespine stickleback along the benthic–limnetic axis. *Biol. J. Linn. Soc. Lond.* 101: 595–608.
- Xu, M., K. G. Tantisira, A. Wu, A. A. Litonjua, J. H. Chu *et al.*, 2011 Genome wide association study to predict severe asthma exacerbations in children using random forests classifiers. *BMC Med. Genet.* 12: 90.
- Zelditch, M. L., D. L. Swiderki, H. D. Sheets, and W. L. Fink, 2004 *Geometric Morphometrics for Biologists.* Elsevier, San Diego, CA.
- Zhang, Q., J. N. Skepper, F. Yang, J. D. Davies, L. Hegyi *et al.*, 2001 Nesprins: a novel family of spectrin-repeat-containing proteins that localize to the nuclear membrane in multiple tissues. *J. Cell Sci.* 114: 4485–4498.
- Zhang, J., A. Felder, Y. Liu, L. T. Guo, S. Lange *et al.*, 2010 Nesprin 1 is critical for nuclear positioning and anchorage. *Hum. Mol. Genet.* 19: 329–341.

Communicating editor: T. Slotte