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Incidence and Distribution of UroSEEK Gene Panel in a Multi-institutional Cohort of Bladder Urothelial Carcinoma

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Abstract

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Disclosure/Conflict of Interest

Nickolas Papadopoulos, Ken W Kinzler, Bert Vogelstein: Founder of Personal Genome Diagnostics and PapGene and advise Sysmex-Inostics. Kinzler and Vogelstein also advise Eisai. Vogelstein is also an advisor to Camden Partners. These companies and others have licensed technologies from Johns Hopkins that are related to the work described in this paper. These licenses are associated with equity or royalty payments to Papadopoulos, Kinzler, Netto, and Vogelstein. Additional patent applications on the work described in this paper may be filed by Johns Hopkins University. The terms of these arrangements are managed by the university in accordance with its conflict of interest policies. The other authors declare that no competing interests exist.

Non-invasive approaches for early detection of bladder cancer are actively being investigated. We recently developed a urine based molecular assay for the detection and surveillance of bladder neoplasms (UroSEEK). UroSEEK is designed to detect alterations in 11 genes that includes most common genetic alterations in bladder cancer. In this study we analyzed 527 cases including 373 non-invasive and 154 invasive urothelial carcinomas of bladder from trans-urethral resections or cystectomies performed at 4 institutions (1991–2016). Two different mutational analysis assays of a representative tumor area were performed: first, a singleplex PCR assay for evaluation of the *TERT* promoter region (TERTSeqS) and second, a multiplex PCR assay using primers designed to amplify regions of interest of 10 (*FGFR3*, *PIK3CA*, *TP53*, *HRAS*, *KRAS*, *ERBB2*, *CDKN2A*, *MET*, *MLL*, and *VHL*) genes (UroSeqS). Overall 92% of all bladder tumors were positive for at least one genetic alteration in the UroSEEK panel. We found *TERT* promoter mutations in 77% of low-grade non-invasive papillary carcinomas with relatively lower incidence of 65% in high-grade non-invasive papillary carcinomas and carcinomas in situ; $p=0.017$. Seventy-two percent of pT1 and 63% of muscle-invasive bladder tumors harbored *TERT* promoter mutations with g.1295228C>T alteration being the most common in all groups. *FGFR3* and *PIK3CA* mutations were more frequent in low-grade non-invasive papillary carcinomas compared to high-grade non-invasive papillary carcinomas and carcinomas in situ ($p<0.0001$), while the opposite was true for *TP53* ($p<0.0001$). Significantly higher rates of *TP53* and *CDKN2A* mutation rates ($p=0.005$ and 0.035, respectively) were encountered in muscle-invasive bladder tumors compared to those of pT1 stage. The overwhelming majority of all investigated tumors showed at least one mutation among UroSEEK assay genes confirming the comprehensive coverage of the panel and supporting its potential utility as a non-invasive urine based assay.

Keywords

bladder cancer; non-invasive assay; urine; molecular diagnostics; FFPE

Introduction

Bladder cancer is the fourth most common cancer in men and the most common malignancy of the urinary tract in both male and female. In 2019, 80,470 new cases are being estimated in the USA leading to 17,670 deaths [1].

Invasive urothelial carcinoma evolves through two distinct pathways: low-and high-grade non-invasive papillary urothelial carcinoma and “flat” carcinoma in situ [2]. Due to high recurrence rates and likelihood of progression to muscle-invasive bladder cancer, periodical follow-up with cystoscopy and cytology for patients diagnosed with non-invasive tumors is required [3–6]. These procedures entail an estimated three billion dollar burden to the health care system every year [7]. Alternative approaches for surveillance are therefore needed.

Early detection of bladder cancer remains a challenge in clinical practice. Hematuria, with or without lower urinary tract symptoms, is the most common presenting symptom. While asymptomatic microscopic hematuria is prevalent (up to 31%) [8, 9], only a small fraction of patients will ultimately be diagnosed with bladder cancer (3–5%) [10]. Therefore, risk stratification with a non-invasive method to avoid unneeded cystoscopy is of great utility.

Desquamated urothelial cells in urine have long been used as a valuable source for non-invasive detection of bladder cancer. Beside cystoscopy, urine cytology remains the gold standard for bladder cancer detection. However, its overall low sensitivity (11–76%) especially in low-grade tumors tampers its utility. Several FDA approved urine-based non-invasive assays are currently available (e.g. UroVysion, ImmunoCyt/uCyt+™, NMP22®, and BTA®) for both early detection and surveillance of bladder cancer. Sensitivity and specificity for these assays range from 56% to 78% and 74% to 88%, respectively [11–18]. We recently developed a non-invasive bladder cancer assay with promising performance characteristics in both early detection and surveillance setting. When combined with cytology, a sensitivity of 95% and a specificity of 93% were reached in early detection cohort [19]. The assay, termed “UroSEEK”, consists of three components (TERTSeqS, UroSeqS and Fast SeqS). It covers molecular alterations that are frequently encountered in bladder cancer in addition to aneuploidy (FastSeqS). The alterations include *TERT* promoter mutations (TERTSeqS) that occur in up to 80% of bladder cancers [20, 21] and 10 additional genes: *FGFR3*, *PIK3CA*, *HRAS*, *KRAS*, *TP53*, *CDKN2A*, *ERBB2*, *MLL*, *MET*, and *VHL* (UroSeqS) [22–24].

Evidently, a non-invasive mutation based approach can only be effective if its genes are closely matched to the tumors it aims to detect. Therefore, in the current study we sought to identify the distribution of the UroSEEK gene panel in archival tumor tissues from a multi-institutional international cohort of bladder urothelial carcinoma. Relationship with tumor grade and stage was also assessed.

Material and Methods

Patient samples and clinical data

The study was approved by the Institutional Board Review of participating institutions. Required material transfer agreements were obtained. The 527 formalin-fixed, paraffin-embedded bladder urothelial carcinoma specimens were collected between 1991 and 2016 from four international academic institutions (Johns Hopkins Hospital, Baltimore, MD, United States; A.C. Camargo Cancer Center, Sao Paulo, Brazil; Osaka University Hospital, Osaka, Japan; and Hacettepe University Hospital, Ankara, Turkey). The mutational findings of the UroSEEK gene panel in a subset (102 tumors) were described in our previously reported study describing the non-invasive multigene assay (UroSEEK) [19]. All histologic sections from transurethral resection of bladder tumor and cystectomy specimens were reviewed by a genitourinary pathologist to confirm the diagnosis and select a representative tumor area. Corresponding formalin-fixed, paraffin-embedded blocks were cored for DNA purification as previously described [20]. Clinicopathologic data was obtained from electronic medical record. Only cases with a minimum follow-up time of 3 month were included in outcome analysis. Disease recurrence was defined as the development of histologically documented tumor occurrence. Progression was defined as the occurrence of histologically documented upgrade or upstage of disease.

Mutation analysis

Mutation and data analysis was performed as previously described [19, 20, 25, 26]. In brief, purified DNA was submitted for SafeSeqS analysis, a sequencing error-reduction technique capable of discriminating mutations from artifactual sequencing variants introduced during the sequencing process [27, 28]. Two different mutational analysis assays were performed: first, a singleplex PCR assay for evaluation of *TERT* promoter region (TERTSeqS) and a second multiplex PCR assay [20] using primers designed to amplify regions of interest of 10 (*FGFR3*, *PIK3CA*, *TP53*, *HRAS*, *KRAS*, *ERBB2*, *CDKN2A*, *MET*, *MLL*, and *VHL*) genes (UroSeqS). Primers are listed in Supplementary Table S1 [19].

In order to evaluate the statistical significance of observed mutations, DNA from white blood cells of 188 unrelated healthy individuals was also assessed. A variant was scored as a mutation only if the mutant allele frequency was much higher than that observed in normal white blood cells. As previously described [19], the classification of a sample's DNA status was based on two complementary criteria applied to each mutation: 1) the difference between the average mutant allele frequency in the sample of interest and the corresponding maximum mutant allele frequency observed for that same mutation in a set of controls; and 2) the Stouffer's Z-score obtained by comparing the mutant allele frequency in the sample of interest to a distribution of normal controls.

Statistical analysis

Statistical analysis was performed using R version 3.5.1 (2018-07-02) from the R Foundation for Statistical Computing (Vienna, Austria). For hypothesis testing, statistical significance was established at $p < 0.05$ for 2 tails of distribution. The relationship between mutation status and pathological and outcome variables was analyzed by Chi-square test with Yates' continuity correction.

Results

Clinicopathologic features

Five hundred twenty-seven tumors from 484 patients were included in the study. One-hundred and thirteen patients were female and 371 were male. The median age was 68 years (range 28–96). Three hundred twenty-nine patients were Caucasian, 68 were Asian, 46 were African-American with the race in the remaining 41 patients being undetermined or from other category. The tumor included 188 low-grade non-invasive papillary carcinomas, 129 high-grade non-invasive papillary carcinomas, 56 carcinomas in situ, and 154 high grade invasive urothelial carcinoma including 111 pT1 and 43 pT2 (muscle-invasive) tumors.

Mutation Analysis

The frequency of mutations of all genes of the UroSEEK gene panel across histopathological categories is summarized in Figure 1. The mutation variants of analyzed genes are listed in Supplementary Table S2. Overall 93% of non-invasive and 92% of invasive tumors were positive for at least one mutation in genes included in UroSEEK assay.

TERT promoter mutations were identified in 70% of all cases with the most common alteration being g.1295228C>T, followed by g.1295250C>T. A previously unreported variant (g.1295223G>T; in 1 case) was detected. A higher incidence of *TERT* promoter mutation was identified in low-grade non-invasive papillary carcinomas compared to high-grade non-invasive papillary carcinomas and carcinomas in situ (77% vs 65%; $p=0.017$; see Table 1). Although higher frequency of *TERT* promoter mutations occurred in pT1 cases (72%) compared to muscle-invasive bladder cancer (63%), the difference was not statistically significant.

Among the 10 genes included in the UroSeqS assay, *FGFR3* and *PIK3CA* mutations occurred significantly more often in low-grade non-invasive papillary carcinoma tumors compared to high-grade non-invasive papillary carcinomas and carcinomas in situ ($p<0.0001$), while the reverse was true for *TP53* ($p<0.0001$; see Table 1). In invasive bladder cancer *CDKN2A* and *TP53* mutations were more commonly observed in muscle-invasive bladder cancer compared to pT1 tumors ($P=0.035$ and 0.005 , respectively; see Table 2). Regarding mutation variants, p.S249C was the most frequent *FGFR3* mutation (67%), while p.E545K and p.H66P were the most common *PIK3CA* and *CDKN2A* mutations, (49% and 79%, respectively; see Supplementary Table S2).

Two hundred and nineteen tumors demonstrated co-occurrence of mutations in both assay components with 59% of tumors harboring *TERT* promoter mutation also showing a mutation in at least one UroSeqS gene ($p=0.001$). Finally, occasional cases demonstrated multiple variants of *TERT* promoter (13 tumors) and *TP53* (14 tumors) mutations and one case displayed two *FGFR3* variant mutations.

Mutational analysis in patients with sequential tumors

Tissues from sequential tumors were available for mutational analysis in 36 patients. As illustrated in Figure 2, *TERT* promoter mutations were absent in all samples in 7 of the 36 patients. In the remaining 29 patients, the same *TERT* promoter mutation was persistently present across tumors in 22 and variably found in 7 patients. Only 2 of 15 patients with observed *FGFR3* mutations had the same mutation present across tumors.

Association of UroSEEK assay with outcome

The distribution of the three hundred-three cases that met the minimum follow-up requirement for outcome analysis was as follows: 124 low-grade non-invasive papillary carcinomas, 78 high-grade non-invasive papillary carcinomas, 24 carcinomas in situ, 58 pT1 and 19 muscle-invasive bladder tumors.

Association of tumor UroSEEK, TERTSeqS and UroSeqS findings with recurrence and progression are summarized in Table 3. As shown, 94% of tumors with subsequent recurrence had mutation in at least one of the 11 genes included in UroSEEK. A comparable rate of 95% was found in tumors without recurrence ($p=0.784$). Only 21 patients developed progression during follow-up. Tumors with progression were less likely to harbor a mutation in one or more UroSEEK assay genes (81% vs. 96%; $p=0.016$).

Discussion

In our current multi-center study we investigated the comprehensive coverage of the UroSEEK gene assay in archival bladder cancer tissues. The TERTSeqS component detected mutations in the *TERT* promoter region in 70% of all tumors. Mutations in UroSeqS gene panel were found in 63% of all cases. Combining these two components led to a capture rate for alteration of the UroSEEK assay in 92% of all tumors. More specifically, in the subset of low-grade non-invasive papillary carcinoma cases, we found at least one gene alteration in 98% of the cases with 77% being positive for TERTSeqS and 70% positive for UroSeqS. This is especially important given the low sensitivity of routine cytology for the diagnosis of low grade non-invasive tumors. As evidenced in our recent study, urine cytology was negative in all 49 low-grade non-invasive papillary carcinoma cases, while urine UroSEEK was positive in 2/3 of these cases [19].

The current observation of 54% of *TP53* mutation in muscle-invasive bladder cancer is in line with findings of recent studies such as The Cancer Genome Atlas [23] and Kim et al. [29] where *TP53* was altered in 48% and 57% of cases, respectively. We observed a lower frequency for *PIK3CA*, *ERBB2* and *MLL* in muscle-invasive bladder cancer compared to these two studies.

Our study represents one of the largest assessment of a multigene assay in low-grade non-invasive papillary carcinoma tumors to date where 188 cases were analyzed. In this group, *FGFR3* mutations were the most frequent (46%) among the 10 genes included in UroSeqS followed by *PIK3CA* mutation in 19%. These rates are lower than those obtained by Hurst et al. [30] in their study of whole exome and targeted sequencing of 82 Ta tumors (79% and 54% for *FGFR3* and *PIK3CA* mutations, respectively). This could be due to our hotspot focused mutational analysis approach given that Hurst et al. have identified many non-hotspot mutations in their analysis.

TERT promoter mutations are one of the most frequent alteration in bladder cancer and its variants [25, 26, 31–33]. In the current analysis, we found *TERT* promoter mutations in 70% of all cases with the highest rate observed in low-grade non-invasive papillary carcinomas (77%). The here found distribution rates of *TERT* promoter mutations in non-invasive lesions is in line with our originally reported rates in conventional urothelial carcinoma in Kinde et al. where the low-grade non-invasive papillary carcinoma group had the highest rate of *TERT* promoter mutations of 86% [20]. In contrast, Pietzak et al. found *TERT* promoter mutations to be more frequent in their high-grade non-invasive papillary carcinoma cases compared to low-grade non-invasive papillary carcinomas (88% and 61 %, respectively) [34].

Our analysis of sequential tumors in 36 patients unveiled an identical *TERT* promoter mutation across tumors in 22 of the 29 patients harboring *TERT* promoter mutations. In surveillance setting, UroSEEK is envisioned to be use for follow-up in patients whose index tumor is positive for at least one alteration in the panel. This consistency of *TERT* promoter alteration across tumors, if proven in a larger cohort of sequential tumors, suggests that assessment of the index tumor might be sufficient and could obviate the need for repeat

sequential tumor testing. The same approach would not apply to cases where the index tumor is only positive for one of the ten genes in UroSeqS where we found inconsistency in mutation detection across tumors from the same patient. This might in part be due to tumor molecular heterogeneity that was not captured by our adopted technique.

Given the nature of the cohort in the original study describing the non-invasive multigene assay (UroSEEK) [19], analysis of association with outcome could only be performed in a subset of patients. This is due to the fact that in a proportion of cases tumors that are contemporaneous with analyzed urine samples were selected. Strengths of the study include the large number of cases in the multi-institutional cohort (527 tumors) and the advantageous error proof nature of the SafeSeqS technique.

In conclusion, the overwhelming majority of all investigated tumors showed at least one mutation among genes included in the recently reported UroSEEK assay, across tumor grade and stage. This confirms the comprehensive coverage of UroSEEK panel and support its potential utility as a non-invasive urine based assay. Our findings are especially reassuring in the subset of low-grade non-invasive papillary carcinoma where routine cytology lacks sensitivity.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

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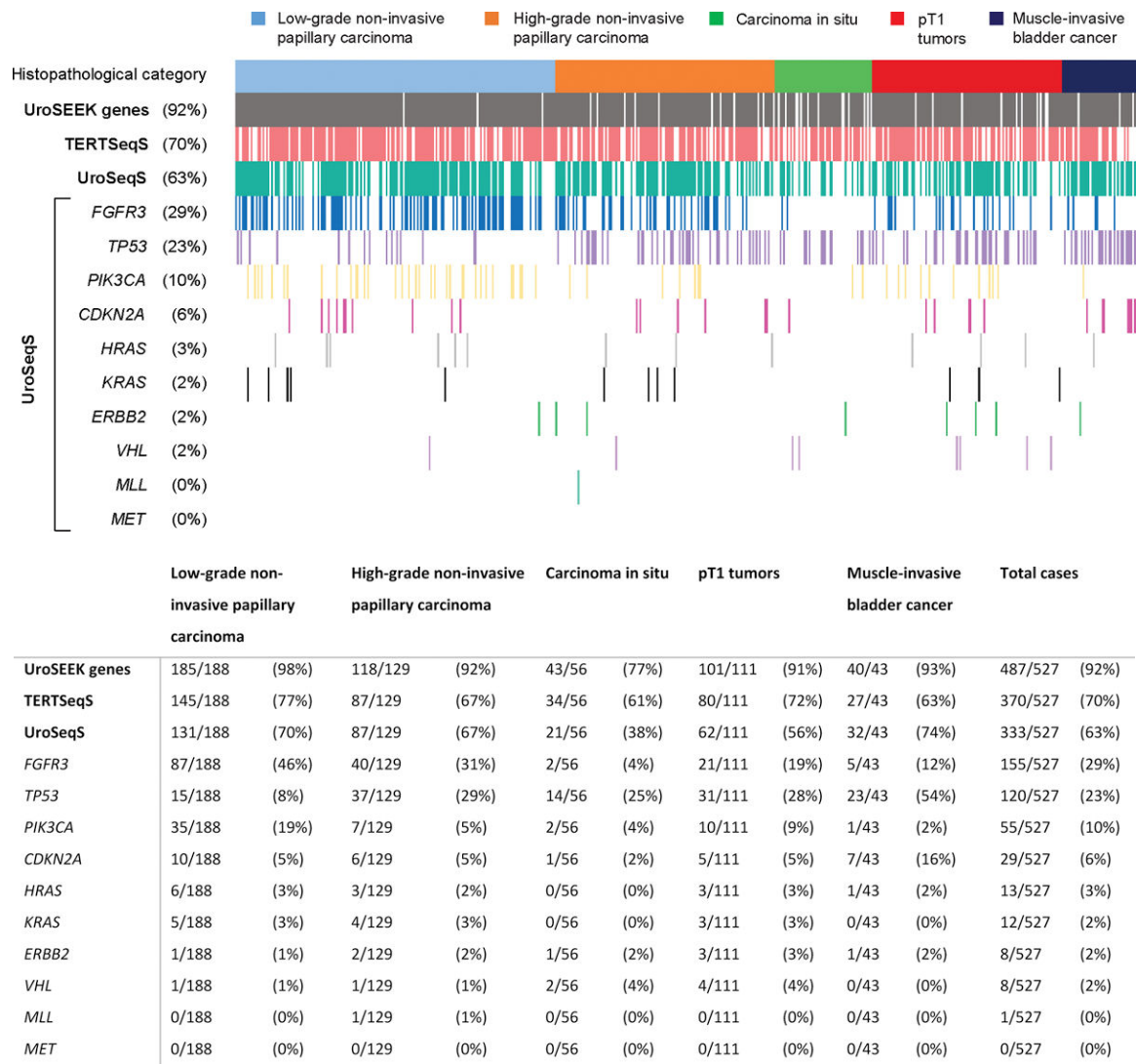


Figure 1. Distribution of mutations in UroSEEK gene panel and its TERTSeqS and UroSeqS components across 527 bladder carcinomas. A) Oncoplot graphic representation of data. Each column represents one tumor. The top line colored boxes indicate the histopathological category. Rows indicate affected gene(s) and their rate of mutations. Detailed listing of the absolute numbers of mutations is shown in B).

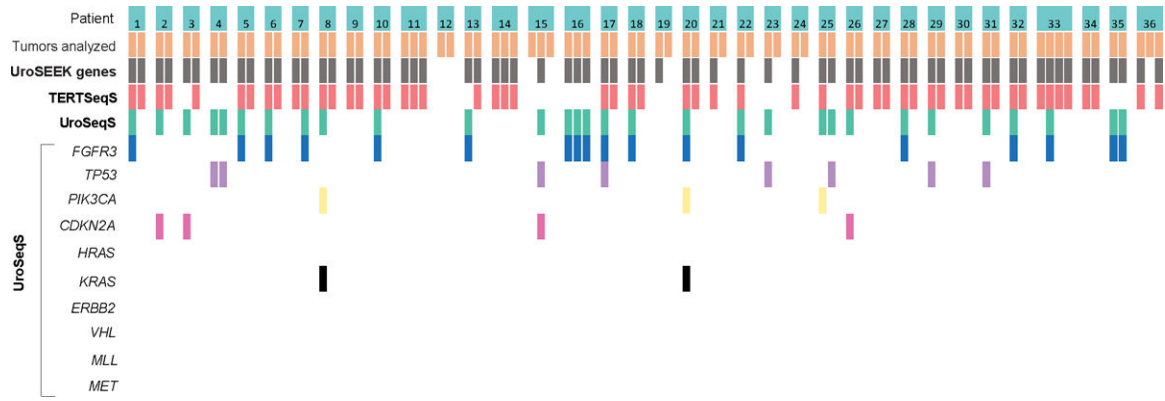


Figure 2. Oncoplot representation of mutations in UroSEEK gene panel and its TERTSeqS and UroSeqS components in 36 patients with sequential tumors. Each broad column represents one patient. The individual tumor per patient (orange box) and their mutated gene(s) are indicated in each row.

Table 1.

Distribution of mutations of UroSEEK gene panel in non-invasive bladder cancer

	Low-grade non-invasive papillary carcinoma		High-grade non-invasive papillary carcinoma + Carcinoma in situ		p-value
UroSEEK genes	185/188	(98%)	161/185	(87%)	<0.0001
TERTSeqS	145/188	(77%)	121/185	(65%)	0.017
UroSeqS	131/188	(70%)	108/185	(58%)	0.030
<i>FGFR3</i>	87/188	(46%)	42/185	(23%)	<0.0001
<i>TP53</i>	15/188	(8%)	51/185	(28%)	<0.0001
<i>PIK3CA</i>	35/188	(19%)	9/185	(5%)	<0.0001
<i>CDKN2A</i>	10/188	(5%)	7/185	(4%)	0.644
<i>HRAS</i>	6/188	(3%)	3/185	(2%)	0.515
<i>KRAS</i>	5/188	(3%)	4/185	(2%)	1.0
<i>ERBB2</i>	1/188	(1%)	3/185	(2%)	0.604
<i>VHL</i>	1/188	(1%)	3/185	(2%)	0.604
<i>MLL</i>	0/188	(0%)	1/185	(1%)	0.994
<i>MET</i>	0/188	(0%)	0/185	(0.0%)	.

Table 2.

Distribution of mutations of UroSEEK gene panel in invasive bladder cancer

	PT1 TUMORS		Muscle-invasive bladder cancer		p-value
UroSEEK genes	101/111	(91%)	40/43	(93%)	0.933
TERTSeqS	80/111	(72%)	27/43	(63%)	0.354
UroSeqS	62/111	(56%)	32/43	(74%)	0.053
<i>FGFR3</i>	21/111	(19%)	5/43	(12%)	0.399
<i>TP53</i>	31/111	(28%)	23/43	(54%)	0.005
<i>PIK3CA</i>	10/111	(9%)	1/43	(2%)	0.273
<i>CDKN2A</i>	5/111	(5%)	7/43	(16%)	0.035
<i>HRAS</i>	3/111	(3%)	1/43	(2%)	1.0
<i>KRAS</i>	3/111	(3%)	0/43	(0%)	0.661
<i>ERBB2</i>	3/111	(3%)	1/43	(2%)	1.0
<i>VHL</i>	4/111	(4%)	0/43	(0%)	0.486
<i>MLL</i>	0/111	(0%)	0/43	(0%)	.
<i>MET</i>	0/111	(0%)	0/43	(0%)	.

Table 3.

Association of UroSEEK gene panel with outcome

	Recurrence		p-value	Progression		p-value
	Yes N=132	No N=171		Yes N=21	No N=282	
Mutation in UroSEEK genes	124/132 (94%)	163/171 (95%)	0.784	17/21 (81%)	270/282 (96%)	0.016
Mutation in TERTSeqS	101/132 (77%)	115/171 (67%)	0.101	12/21 (57%)	204/282 (72%)	0.217
Mutation in UroSeqS	87/132 (66%)	120/171 (70%)	0.505	12/21 (57%)	195/282 (69%)	0.369

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