

Gold nanoparticles promote osteogenic differentiation in human adipose-derived mesenchymal stem cells through the Wnt/ β -catenin signaling pathway

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Abstract: Gold nanoparticles (AuNPs) are attractive materials for use in biomedicine due to their physical properties. Increasing evidence suggests that several nanoparticles induce the differentiation of human mesenchymal stem cells into osteoblasts and adipocytes. In this study, we hypothesized that chitosan-conjugated AuNPs promote the osteogenic differentiation of human adipose-derived mesenchymal stem cells. For the evaluation of osteogenic differentiation, alizarin red staining, an alamarBlue[®] assay, and a quantitative real-time polymerase chain reaction analysis were performed. In order to examine specific signaling pathways, immunofluorescence and a western blotting assay were performed. Our results demonstrate that chitosan-conjugated AuNPs increase the deposition of calcium content and the expression of marker genes related to osteogenic differentiation in human adipose-derived mesenchymal stem cells at nontoxic concentrations. These results indicate that chitosan-conjugated AuNPs promote osteogenesis through the Wnt/ β -catenin signaling pathway. Therefore, chitosan-conjugated AuNPs can be used as a reagent for promoting bone formation.

Keywords: chitosan-conjugated gold nanoparticle, mineralization, nonphosphorylated beta-catenin

Introduction

Transplant engineering of stem cells is a potential therapeutic method for the regeneration of damaged tissue, such as with osteoporosis, which is becoming an important issue and health problem.¹⁻⁶ Human mesenchymal stem cells (hMSCs) derived from various tissues, including bone marrow, adipose tissue, and umbilical cord blood, possess multilineage differentiating potentials.⁷ Interests in the therapeutic potential of stem cells isolated from adipose tissue, called adipose-derived stem cells (ADSCs), has grown due to a less expensive acquisition and greater available quantities than bone marrow.⁸⁻¹¹ However, mesenchymal stem cells (MSCs) derived from various sources have limited proliferation activity and differentiation potentials.^{12,13} Therefore, it appears that developing new technologies to stimulate the differentiation of MSCs is required. Recently, gold nanoparticles (AuNPs) have been suggested as useful medical vehicles for regenerative tools due to their nontoxic effects, as compared to other types of nanoparticles (NPs).^{14,15} Various forms of AuNPs are known to be involved in the differentiation of hMSCs into various types of cells. For example, gold-coated iron oxide (gold/Fe₃O₄) and graphene stimulate the neural differentiation of rat olfactory bulb neural stem cells, and graphene oxide-encapsulated AuNPs are attractive

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detection materials for neural stem cell differentiation.^{16,17} In addition, it has been proposed that AuNPs regulate the myocardial differentiation of MSCs.^{18,19} Many reports suggest that AuNPs promote the differentiation of hMSCs into osteoblasts.^{20,21} Citrate-reduced AuNPs stimulate the osteogenic differentiation of bone marrow-derived MSCs through the mitogen-activated protein kinase signaling pathway, and the osteogenic differentiation of fibroblasts was enhanced by an AuNPs–hydrogel complex through the bone morphogenic protein signaling pathway.^{22,23} Influence of various nanomaterials on stem cell differentiation was studied;^{16–24} however, research regarding the effects of chitosan-conjugated AuNPs on the differentiation of hMSCs has not been reported.

In the present study, we investigated the effect of chitosan-conjugated AuNPs on the differentiation of human adipose-derived mesenchymal stem cells (hADMSCs) as a switch to determine cell fate into osteoblasts and the relating signaling pathway in cell differentiation.

Materials and methods

Culture of hADMSCs

The hADMSCs were purchased from ATCC (PCS-500-011) and cultured in low-glucose Dulbecco's Modified Eagle's Medium (GIBCO cell culture, Thermo Fisher Scientific, Waltham, MA, USA) with 10% fetal bovine serum and 1% antibiotics (Sigma-Aldrich, St Louis, MO, USA) incubating at 37°C in 5% CO₂. The culture medium was replaced every 3 days during the experiments.

Characterization of chitosan-conjugated AuNPs

In this study, we used the same chitosan-conjugated AuNPs which were used in our previous study.²⁵ In summary, positively charged AuNPs were prepared using the chitosan reduction method. The average diameter of chitosan-conjugated AuNPs was ~17 nm, and the zeta potentials were ~42 mV. The hydrodynamic diameters of chitosan-conjugated AuNPs were ~40 nm from the dynamic light scattering measurements.

Transmission electron microscope

The uptake of chitosan-conjugated AuNPs was examined using a transmission electron microscope (TEM). The hADMSCs were plated into six-well culture plates at 3×10³ cells/cm². After cell confluency, chitosan-conjugated AuNPs were added to the growth medium and incubated at 37°C in 5% CO₂ for 24 hours. Fixation, embedding, ultrathin sections,

and analysis using a TEM were operated in the Integrative Research Support Center in School of Medicine, the Catholic University of Korea.

Cell viability assay

The cell viability of hADMSCs was assessed using a Cell Counting Kit-8 (CCK-8; Dojindo Molecular Technologies, Inc. Kumamoto, Japan). Cells were seeded in 96-well culture plates at 3×10³ cells/cm². After cell confluency, chitosan-conjugated AuNPs were added to the culture medium. Wells containing the culture medium and NPs without cells were used as blanks. After 72 hours of incubation, 10 μL of the CCK-8 solution was added to the medium and incubated for 4 hours at 37°C. The absorbance of the optical density (OD) at 450 nm was measured using a microplate reader (TECAN, Männedorf, Switzerland). The cell viability (%) was calculated according to the formula: $(OD_{\text{sample}} - OD_{\text{blank}}) / (OD_{\text{control}} - OD_{\text{blank}}) \times 100$.

Proliferation of hADMSCs

The proliferation of the hADMSCs was determined using an alamarBlue[®] assay. Briefly, hADMSCs were seeded in each well of 96-well plates at a density of 3×10³ cells/cm² and incubated for 72 hours. After cell confluency, an osteogenic-inducing medium (OM) containing 100 nM dexamethasone (Calbiochem, EMD Millipore, Billerica, MA, USA), 50 μM L-ascorbic acid 2-phosphate sesquimagnesium salt hydrate (Sigma-Aldrich), and 10 mM beta-glycerophosphate disodium salt hydrate (Sigma-Aldrich) was added to each well with different concentrations of chitosan-conjugated AuNPs and incubated for 10, 14, and 21 days. Wells containing OM and NPs without cells were used as blanks. At each time point, 20 μL alamarBlue[®] solution (Invitrogen Corporation, Carlsbad, CA, USA) was added to the medium and incubated for 4 hours at 37°C. The absorbances at 570 nm and 620 nm were measured using a microplate reader (TECAN).

Alizarin Red S for mineralization

The osteogenic differentiation rates of hADMSCs in the presence of chitosan-conjugated AuNPs were determined by an Alizarin Red S (ARS) staining assay. The hADMSCs were seeded in a six-well culture plates at a cell density of 3×10³ cells/cm² and cultured for 10, 14, and 21 days in OM and chitosan-conjugated AuNPs. Briefly, the cells were fixed with 80% ethanol for 1 hour at room temperature. After being washed with distilled water, they were stained with 60 mM ARS (pH 4.2) for 15 minutes at room temperature. A quantitation of ARS staining was performed by elution with 10% (w/v) cetylpyridinium chloride (Sigma-Aldrich)

for 20 minutes at room temperature, and the absorbance was measured at 570 nm. The mineralization ratio was normalized to the proliferation ratio.

Oil Red O for lipid accumulation

The adipogenic differentiation rates of hADMSCs in the presence of chitosan-conjugated AuNPs were determined by an Oil Red O (ORO) staining assay. After osteogenic induction, the cells were washed with distilled water and they were stained with 0.6% (w/v) ORO solution (60% isopropanol) for 15 minutes at room temperature. For quantitation, the cells were washed with distilled water to remove unbound dye, and 2 mL isopropyl alcohol was added to the wells of the culture plates. After 20 minutes, the absorbance was measured by a microplate reader at 510 nm. The lipid accumulation ratio was normalized to the proliferation ratio.

Real-time RT-PCR analysis

The total RNA was isolated from the differentiated cells, which were treated with chitosan-conjugated AuNPs for 10, 14, and 21 days using a Hybrid-R prep kit (GeneAll Biotechnology Co., Ltd., Seoul, South Korea) according to the manufacturer's protocol. Reverse transcription (RT) was performed using M-MLV reverse transcriptase (Invitrogen). The relative mRNA expression of the osteogenic differentiation marker genes was normalized to the *GAPDH* gene and expressed as a fold change relative to the growth medium group. The polymerase chain reaction (PCR) conditions were an initial step at 95°C for 30 seconds and 40 denaturation cycles of 95°C for 5 seconds and annealing at 60°C for 45 seconds. Steps at 95°C for 15 seconds, 60°C for 1 minute, and 95°C for 15 seconds was added to minimize nonspecific products. The results were analyzed by comparing the $2^{-[\Delta\Delta Ct]}$ values of the mRNA of cells treated with chitosan-conjugated AuNPs to those of the cells in OM. Table 1 represents the list of primers used for real-time RT-PCR.

Western blotting analysis

A western blotting assay was applied to study the protein expressions of hADMSCs related to the β -catenin signaling pathway. Proteins were extracted and subjected to 10% sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) for the detection of active-form β -catenin, then transferred to PVDF membranes. The membranes were blocked with 5% skim milk in tris-buffered saline with Tween 20 for 1 hour at room temperature. Membranes were reacted with primary antibodies overnight at 4°C. Primary antibodies were as follows: anti-nonphosphorylated β -catenin (1:500 [EMD Millipore, Billerica, MA, USA]) and β -actin (1:1000 [Santa Cruz Biotechnology Inc., Dallas, TX, USA]), and the β -actin protein was used as the control.

Immunofluorescence for translocation of nonphosphorylated β -catenin into the nucleus

The hADMSCs were plated to confocal dishes (Nunc™ Lab-Tek chamber slides, Thermo Fisher Scientific) at a density of 3×10^3 cells/cm² and cultured for 7, 10, 14, and 21 days in an OM treated with chitosan-conjugated AuNPs. The cells were fixed with 4% paraformaldehyde for 15 minutes. Fixed cells were incubated with an active-form β -catenin (EMD Millipore) antibody for 48 hours at 4°C, then incubated with a secondary antibody, which was conjugated with Alexa Fluor® 555 for 1 hour at room temperature. Cells were stained with 2-(4-aminophenyl)-6-indolecarbamidine dihydrochloride for nucleus staining. Confocal images were acquired with a LSM 710 confocal laser scanning microscope (Carl Zeiss, Hamburg, Germany) operated in the National Center for Inter-University Research Facilities (Seoul National University, Korea).

Statistical analysis

Data was generally presented as the mean \pm standard error, and the statistical differences between the experimental

Table 1 Primers for quantitative real-time RT-PCR analysis

Gene (accession number)	Sequences	Product size (bp)
Alkaline phosphatase (NM_000478.4)	F 5'-CCTCCTCGGAAGACACTCTG-3' R 5'-GCAGTGAAGGGCTTCTTGTC-3'	139
Bone sialoprotein (NM_004967.3)	F 5'-AAAGTGAGAACGGGGAACCT-3' R 5'-GATGCAAAGCCAGAATGGAT-3'	161
Osteocalcin (NM_001199662.1)	F 5'-GACTGTGACGAGTTGGCTGA-3' R 5'-CTGGAGAGGAGCAGAACTGG-3'	119
GAPDH (NM_002046.4)	F 5'-CTCTGCTCCTCTGTTGAC-3' R 5'-ACGACCAAATCCGTTGACTC-3'	112

Abbreviations: F, forward; R, reverse; RT-PCR, reverse transcription polymerase chain reaction.

groups were analyzed by Student's *t*-test using statistical software Origin 8.0. $P < 0.05$ was considered statistically significant in all cases.

Results and discussion

Uptake of chitosan-conjugated AuNPs in hADMSCs

Recent studies suggested that the anionic AuNPs are endocytosed into cells, while half the amounts of cationic AuNPs escape the endocytotic pathway.²⁶ Cationic AuNPs may directly diffuse into cytosol by generating disruptions on the cell membranes.^{27,28} In addition, uptake mechanisms of NPs were reportedly dependent on the cell types.^{29,30} In an earlier study, chitosan-conjugated AuNPs demonstrated endocytotic uptake in lung cancer cells.²⁵ In order to identify uptake of chitosan-conjugated AuNPs in hADMSCs, internalized chitosan-conjugated AuNPs were measured using a TEM. The concentration of chitosan-conjugated AuNPs added for TEM was 1 ppm. Most of the internalized chitosan-conjugated AuNPs were detected in cytosol as shown in Figure 1. However, in this TEM images, chitosan-conjugated AuNPs internalized into hADMSCs were not inside endosomal vesicles. According to our results, chitosan-conjugated AuNPs entered into hADMSCs through direct diffusion or escaping the endocytotic pathway.

Effect of chitosan-conjugated AuNPs on hADMSC viability and proliferation

Over the past few years, the interactions of AuNPs with cell have been researched to confirm their uptake, distribution, and cytotoxicity. AuNPs with various sizes and shapes were taken in a variety of cell types. Recent evidence indicated that AuNPs decreased cell viability and differentiation of bone

marrow MSCs through triggered necrosis by excessive reactive oxygen species (ROS) generation.³¹ In particular, excessive ROS cause damage to MSCs, whereas low levels of ROS enhance osteogenesis of MSCs.³² To determine the viability of hADMSCs treated with chitosan-conjugated AuNPs, cell viability assays were performed for 72 hours in a growth medium that did not contain osteogenic-inducing substances (Figure 2A). As shown in Figure 2A, chitosan-conjugated AuNPs had no effect on the cell viability of hADMSCs at a concentration of up to 10 ppm. The enhancement of the osteogenic differentiation was followed by increased cell proliferation.^{7,33} To determine the proliferation of the hADMSCs in osteogenic-inducing conditions, a proliferation assay was examined at 10, 14, and 21 days in an OM-containing chitosan-conjugated AuNPs (Figure 2B). This result indicates that the value of OD was significantly increased in all experimental groups in a time-dependent manner. However, there is no statistical significance in the difference between the OM-only group and the OM-containing chitosan-conjugated AuNPs group. The degree of uptake and subcellular localization in cell systems showed different cytotoxicity.³⁴ In particular, gold nanostars located in the nucleus are more toxic compared to the gold nanostars located in the cytosol.³⁵ Therefore, it is possible that AuNPs located in membrane are lacking in cytotoxicity in hADMSCs.

Chitosan-conjugated AuNPs promote the osteogenic differentiation of hADMSCs

Pristine AuNPs without any appropriate modifications are generally unstable because of high surface energy. So a stabilizer should be added to apply AuNPs into cells.³⁶ In the present study, chitosan was used in the reduction of AuNPs as a stabilizer. Several researches suggest that

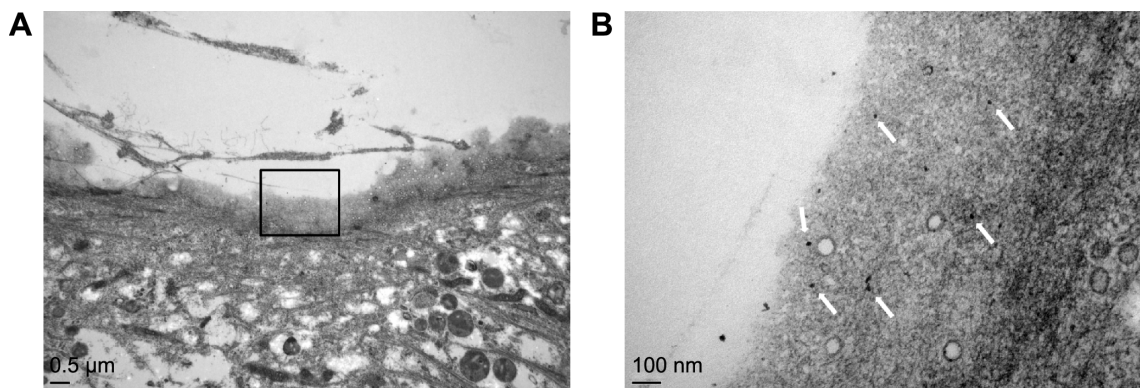


Figure 1 TEM images of hADMSCs exposed to chitosan-conjugated AuNPs.

Notes: hADMSCs were exposed to chitosan-conjugated AuNPs for 24 hours and then fixed for TEM. **(A)** Chitosan-conjugated AuNPs were internalized in cell membrane as magnified in the **(B)** image. White arrows represent chitosan-conjugated AuNPs in cell membrane. The scale bars in **(A)** and **(B)** are 0.5 μm and 100 nm.

Abbreviations: AuNPs, gold nanoparticles; hADMSCs, human adipose-derived mesenchymal stem cells; TEM, transmission electron microscope.

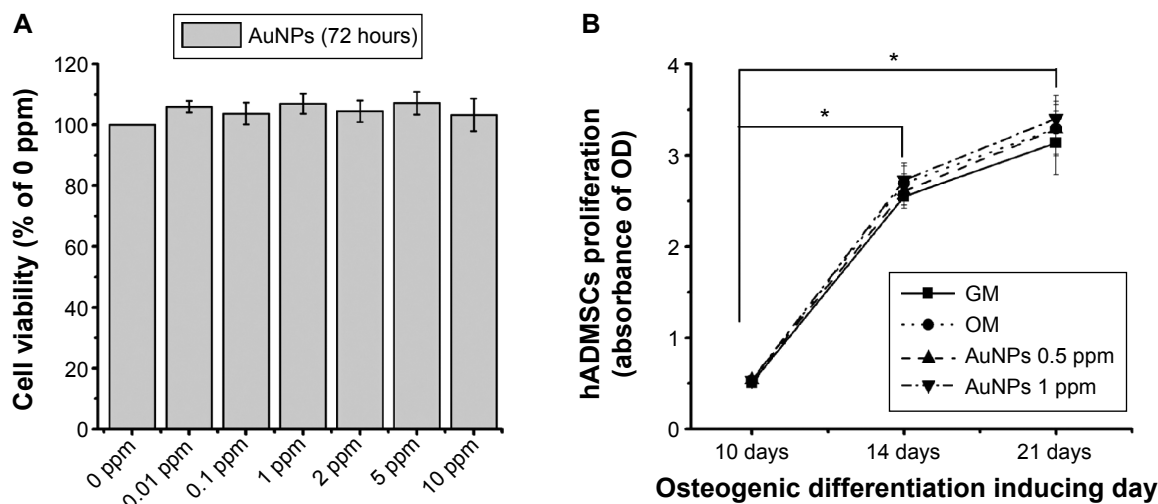


Figure 2 Effects of chitosan-conjugated AuNPs.

Notes: (A) On the cell viability of hADMSCs for 72 hours in a growth medium (GM) and (B) on the cell proliferation of hADMSCs for 10, 14, and 21 days after the induction of osteogenic differentiation. Results are mean \pm SE of the triplicate experiments. * $P < 0.05$.

Abbreviations: AuNPs, gold nanoparticles; hADMSCs, human adipose-derived mesenchymal stem cells; OM, osteogenic-inducing medium; SE, standard error; OD, optical density.

chitosan enhanced mineralization of bone marrow-derived hMSCs by upregulating genes associated with mineralization and calcium-binding proteins.^{37,38} In particular, multiwall carbon nanotube scaffolds modified with chitosan enhanced proliferation and mineralization of MG-63 cells more than chitosan scaffolds.³⁹ In order to confirm the effect of chitosan-conjugated AuNPs on the osteogenic differentiation of hADMSCs in osteogenic-inducing conditions, osteogenic differentiation was determined by measuring the mineralized calcium deposition. The result shows that the mineralized calcium deposition increased more in the hADMSCs containing OM with chitosan-conjugated AuNPs group than in the OM-only group in a time-dependent manner (Figure 3A). In 21 days, the mineralized calcium deposition increased in the containing AuNPs groups up to 1.4-fold at a concentration of 0.5 ppm and 1.35-fold at 1 ppm, as compared to the OM-only group (Figure 3B). Therefore, chitosan-conjugated AuNPs stimulated the differentiation of hADMSCs by osteogenesis.

Expression of osteogenic differentiation specific genes in hADMSCs

A real-time RT-PCR analysis was used to measure the expressions of specific genes to reveal the involvement of the osteogenic differentiation of hADMSCs in OM-containing chitosan-conjugated AuNPs. In the osteogenic differentiation of hMSCs, bone formation is a complex biological process involving several tightly regulated gene expression patterns of bone-related proteins, and specific marker genes, such as osteocalcin (OSC), osteopontin, and bone sialoprotein (BSP),

were expressed according to the progress of differentiation.⁴⁰ Alkaline phosphatase (ALP) is a key regulator of the early stage differentiation of bone marrow stromal cells, and gradually decreased as proceeding through differentiation into osteoblasts.⁴¹ The result shows that the ALP mRNA expression level increased more in the cells containing chitosan-conjugated AuNPs group as compared to the OM-only group at 10 days (Figure 4A). The ALP mRNA expression level in the OM-only group showed a maximal value at 14 days, and it decreased at 21 days. In the OM-containing chitosan-conjugated AuNPs group, the ALP mRNA expression levels increased at 10 days when compared to the OM-only group, and the levels were sustained at 14 days and 21 days. Lian and Stein suggest that the ALP mRNA expression changed during the developmental maturation stage of the osteoblast phenotype.⁴² The peak levels of the ALP mRNA represent the matrix maturation period, and the cellular levels of the ALP mRNA in the heavily mineralized stage decreased.⁴² We observed that the ALP mRNA expression was early induced by the chitosan-conjugated AuNPs group, as compared to the OM-only group. The BSP gene is up-regulated by promoting bone formation.⁴³ The BSP mRNA expression levels in the OM-containing chitosan-conjugated AuNPs group at 0.5 ppm and 1 ppm were higher than in the OM-only group at 14 days and 21 days (Figure 4B). In the OM-only group, the BSP mRNA level decreased in a time-dependent manner. However, the BSP mRNA level in the OM-containing chitosan-conjugated AuNPs group increased up to 14 days and then decreased at 21 days, as compared to the OM-only group. This indicates the BSP mRNA expression was stimulated

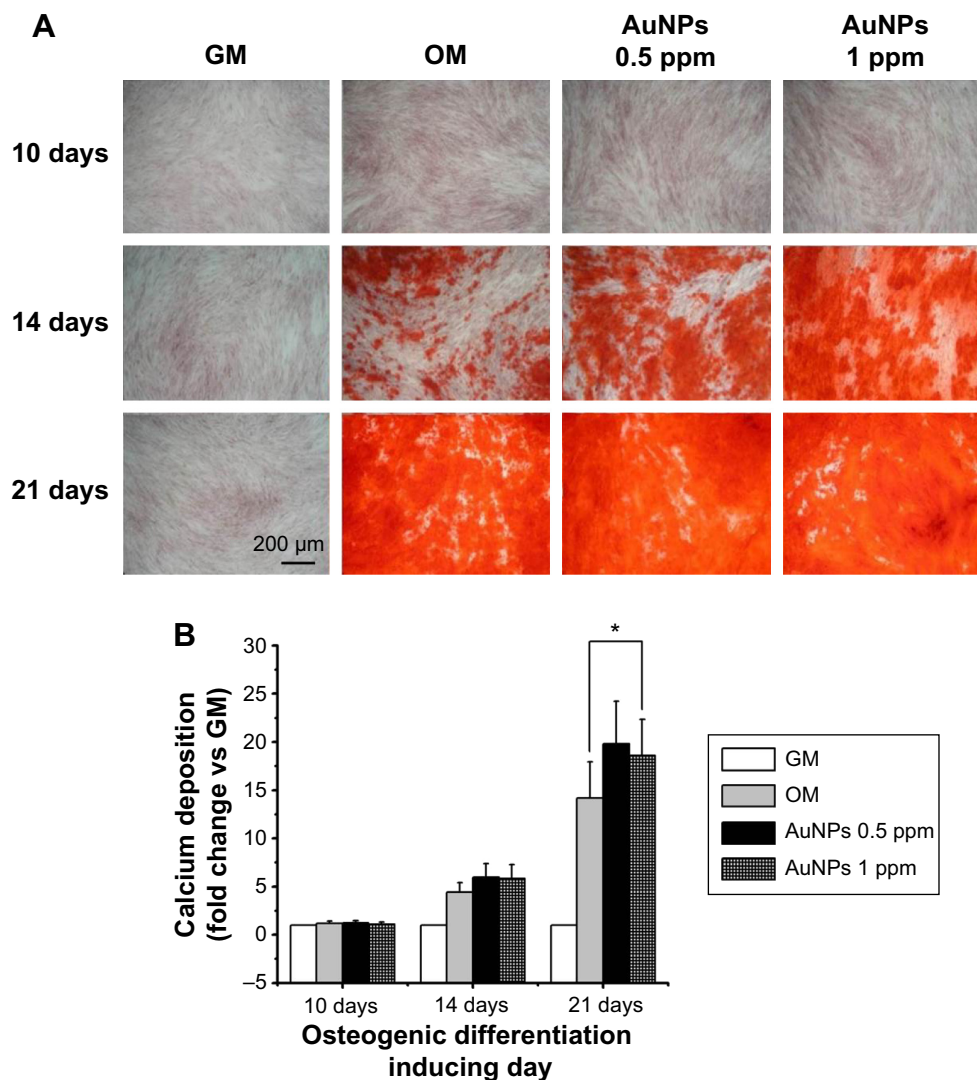


Figure 3 Effect of chitosan-conjugated AuNPs on the mineralization of hADMSCs.

Notes: (A) Stained calcium deposition by Alizarin Red S. (B) Mineralization was quantitated through the elution of Alizarin Red S from stained mineral deposits. Results are mean \pm SE of the triplicate experiments, * $P < 0.05$.

Abbreviations: AuNPs, gold nanoparticles; GM, growth medium; hADMSCs, human adipose-derived mesenchymal stem cells; OM, osteogenic-inducing medium; SE, standard error.

by the chitosan-conjugated AuNPs. The OSC gene is a key marker for the late maturation of osteoblasts and the expression of OSC is concomitantly up-regulated by more advanced osteoblasts.^{40,44} The OSC mRNA expression levels increased in cells induced by osteogenic differentiation, and the OSC mRNA expression levels in the OM-containing chitosan-conjugated AuNPs group were significantly higher than in the OM-only group at 10 days and 14 days (Figure 4C). Thus, the cell fate of the hADMSCs was more induced by the chitosan-conjugated AuNPs group into osteoblasts than by the OM-only group.

According to the integrated real-time PCR results, the expressions of the osteogenic marker genes were more activated by the chitosan-conjugated AuNPs group, and the osteogenic differentiation of hADMSCs treated with

chitosan-conjugated AuNPs was more promoted than in the OM-only group.

Chitosan-conjugated AuNPs do not affect the adipogenic differentiation in osteogenic-inducing hADMSCs

Osteogenesis and adipogenesis are reciprocal events sharing some transcription factors in the specific lineage fate of MSCs.⁴⁵ The stimulation of osteogenic differentiation by AuNPs inhibits the production of lipid droplets, which is a phenotype marker of the maturation of adipogenic differentiation.²² The effect of chitosan-conjugated AuNPs on the adipogenic differentiation of hADMSCs in osteogenic-inducing conditions was determined by measuring lipid droplets stained by ORO dye. In our study, the accumulation

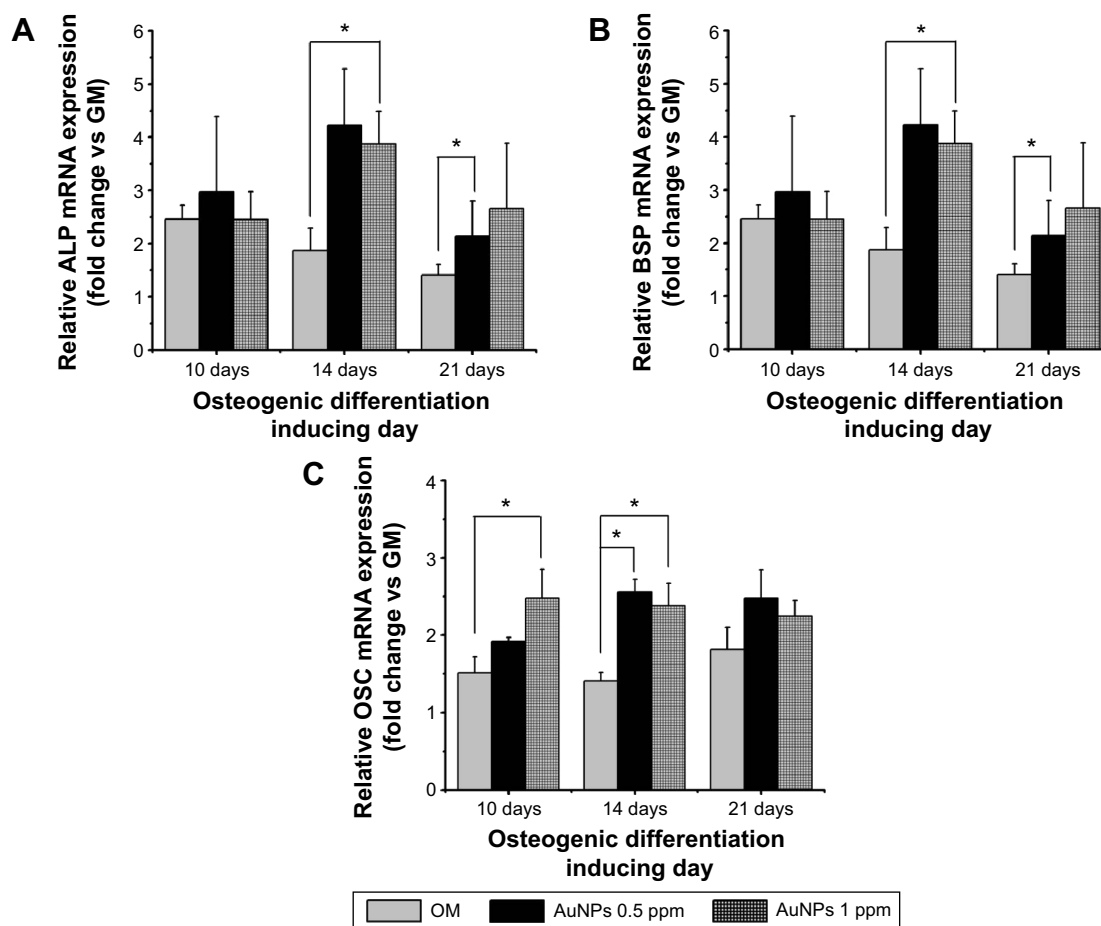


Figure 4 A real-time RT-PCR analysis of the osteogenic marker gene expression in hADMSCs treated with chitosan-conjugated AuNPs for 10, 14, and 21 days.

Notes: (A) Alkaline phosphatase (ALP); (B) bone sialoprotein (BSP); (C) osteocalcin (OSC). Results are mean \pm SE of the triplicate experiments. * $P < 0.05$.

Abbreviations: AuNPs, gold nanoparticles; GM, growth medium; hADMSCs, human adipose-derived mesenchymal stem cells; OM, osteogenic-inducing medium; RT-PCR, reverse transcription polymerase chain reaction; SE, standard error.

of lipid droplets was slightly increased in the OM-only group and the OM-containing chitosan-conjugated AuNPs group (Figure 5). However, the degree of accumulation was weak when compared to the mineralization, and it was similar between the OM-only group and the OM-containing chitosan-conjugated AuNPs group. Therefore, the adipogenic differentiation of hADMSCs was not induced by chitosan-conjugated AuNPs in OM conditions.

Chitosan-conjugated AuNPs enhance the osteogenic differentiation of hADMSCs through the Wnt/ β -catenin signaling pathway

Several studies demonstrated that the Wnt/ β -catenin signaling pathway was initially involved in osteoblast differentiation.^{46–48} Wnt/ β -catenin signaling inhibits adipogenic differentiation and alters the cell fate from adipocyte to osteoblast in hADMSCs.⁴⁹ To confirm the signaling

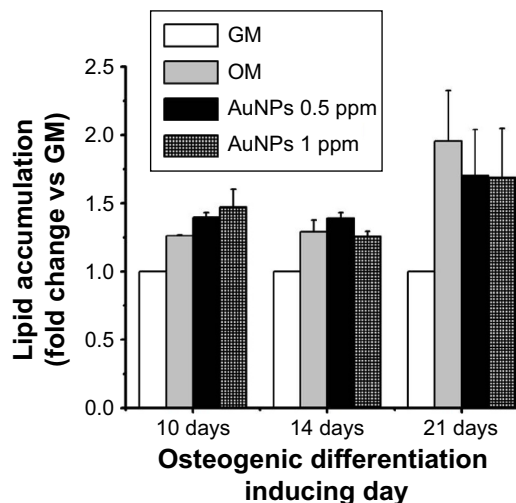


Figure 5 Effect of chitosan-conjugated AuNPs on adipogenic differentiation.

Notes: Lipid accumulation of hADMSCs treated with chitosan-conjugated AuNPs after 10, 14, and 21 days of culturing. Results are mean \pm SE of the triplicate experiments.

Abbreviations: AuNPs, gold nanoparticles; GM, growth medium; hADMSCs, human adipose-derived mesenchymal stem cells; OM, osteogenic-inducing medium; SE, standard error.

pathway related to osteogenic differentiation in hADMSCs, we examined nonphosphorylated β -catenin in protein levels. Nonphosphorylated β -catenin protein levels increased in the OM-containing chitosan-conjugated AuNPs group at 7 days, as compared to the OM-only group (Figure 6A). The nonphosphorylated β -catenin in the cytoplasm accumulated and then translocated into the nucleus while activating the Wnt/ β -catenin signaling pathway.⁵⁰⁻⁵² The translocation

of nonphosphorylated β -catenin into the nuclei of the OM group was determined by using an immunofluorescent antibody (Figure 6B). The translocation of nonphosphorylated β -catenin into nuclei of the OM-containing chitosan-conjugated AuNPs group was greater than in the OM-only group at 7 days and 10 days. After 10 days, the degree of translocated nonphosphorylated β -catenin proteins was similar between the OM-only group and the OM-containing

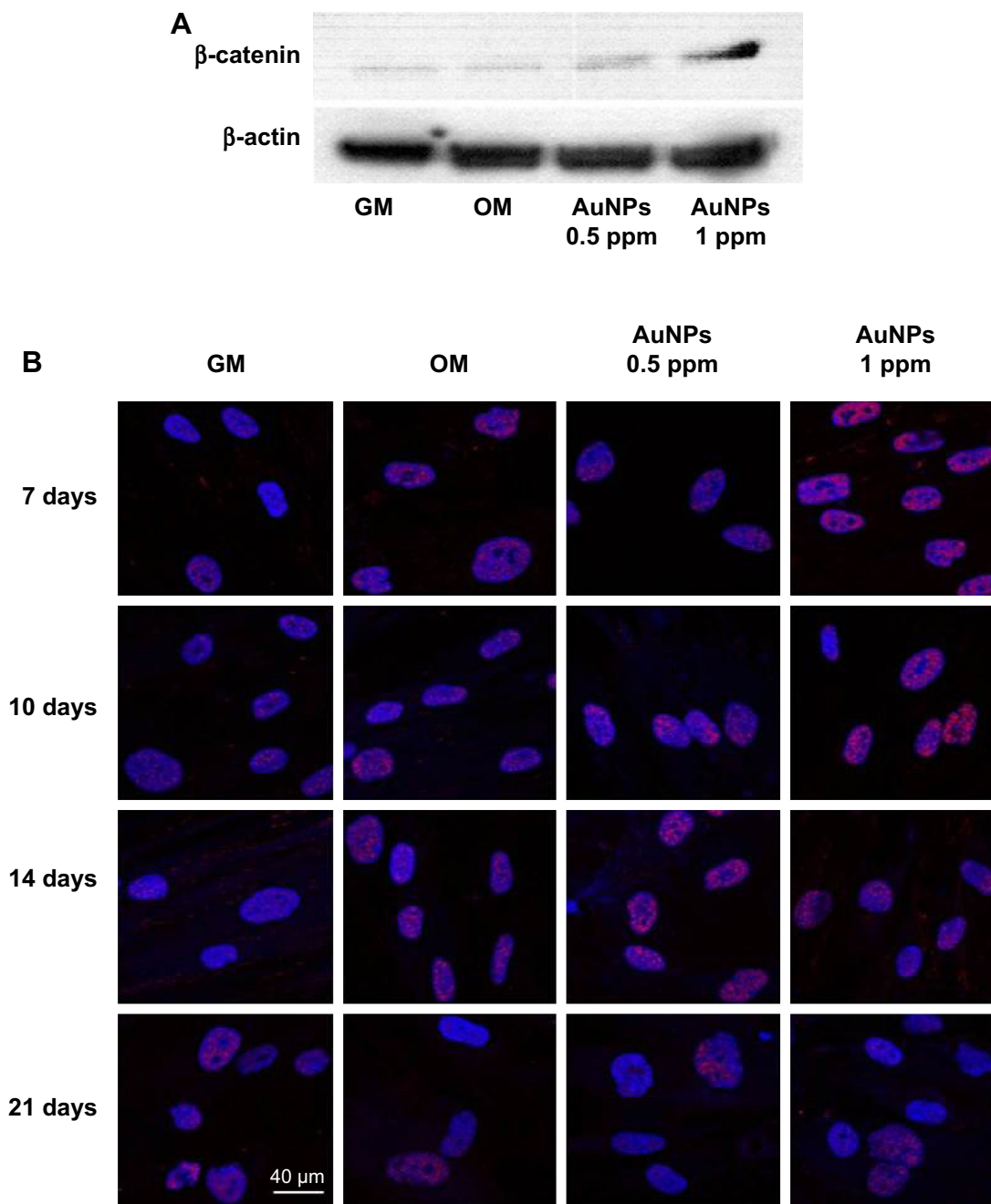


Figure 6 Activation of the osteogenic signaling pathway by chitosan-conjugated AuNPs.

Notes: (A) A western blot analysis of the nonphosphorylated β -catenin protein expression after treatment with chitosan-conjugated AuNPs for 7 days. (B) Translocation of nonphosphorylated β -catenin into the nuclei of hADMSCs treated with chitosan-conjugated AuNPs for 7, 10, 14, and 21 days.

Abbreviations: AuNPs, gold nanoparticles; GM, growth medium; hADMSCs, human adipose-derived mesenchymal stem cells; OM, osteogenic-inducing medium.

chitosan-conjugated AuNPs group. Some studies suggested that various type of NPs act in hMSCs by different action mechanism. Adipogenic and osteogenic differentiation of hMSCs added with silver NPs was inhibited by the Ag⁺ ion, which suppresses the expression of marker genes, and SH-SY5Y cells treated with silver NPs differentiate into neuronal cells through ROS activating the ERK-AKT pathway.^{53,54} Also, AuNPs enhance the osteogenic differentiation of MSCs through the activation of the mitogen-activated protein kinase and bone morphogenic protein signaling pathways.^{22,23} However, the mechanism of the action of the NPs in hADMSCs has not been clear. AuNPs induced mechanical stimulation in MSCs.²² Also, several researches suggested that mechanical stimulation enhance osteogenesis and inhibit adipogenesis through activation of Wnt/ β -catenin signaling.^{55–58}

According to our results, mechanical stimulation by uptake of chitosan-conjugated AuNPs in hADMSCs promote the differentiation of hADMSCs into osteoblasts through nonphosphorylated β -catenin accumulation, as it enhances the switching of the hADMSC cell fate from adipocyte to osteoblast.

Conclusion

This study showed that chitosan-conjugated AuNPs at nontoxic concentrations enhanced the osteogenic differentiation of hADMSCs. The uptake of chitosan-conjugated AuNPs in hADMSCs induced mechanical stimulation that regulated mRNA expression and mineralization by activating the Wnt/ β -catenin signaling pathway. Therefore, chitosan-conjugated AuNPs have been demonstrated as stimulators of the osteogenic differentiation of MSCs, and can be used as vehicles for promoting bone tissue regeneration.

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Disclosure

The authors report no conflicts of interest in this work.

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