



Neuroimmunoendocrine Link Between Chronic Kidney Disease and Olfactory Deficits

Rebeca Corona¹, Benito Ordaz¹, Ludivina Robles-Osorio², Ernesto Sabath² and Teresa Morales^{1*}

¹ Instituto de Neurobiología, Universidad Nacional Autónoma de México, Querétaro, Mexico, ² Facultad de Nutrición, Universidad Autónoma de Querétaro, Querétaro, Mexico

Chronic kidney disease (CKD) is a multifactorial pathology that progressively leads to the deterioration of metabolic functions and results from deficient glomerular filtration and electrolyte imbalance. Its economic impact on public health is challenging. Mexico has a high prevalence of CKD that is strongly associated with some of the most common metabolic disorders like diabetes and hypertension. The gradual loss of kidney functions provokes an inflammatory state and endocrine alterations affecting several systems. High serum levels of prolactin have been associated with CKD progression, inflammation, and olfactory function. Also, the nutritional status is altered due to impaired renal function. The decrease in calorie and protein intake is often accompanied by malnutrition, which can be severe at advanced stages of the disease. Nutrition and olfactory functioning are closely interconnected, and CKD patients often complain of olfactory deficits, which ultimately can lead to deficient food intake. CKD patients present a wide range of deficits in olfaction like odor discrimination, identification, and detection threshold. The chronic inflammatory status in CKD damages the olfactory epithelium leading to deficiencies in the chemical detection of odor molecules. Additionally, the decline in cognitive functioning impairs the capacity of odor differentiation. It is not clear whether peritoneal dialysis and hemodialysis improve the olfactory deficits, but renal transplants have a strong positive effect. In the present review, we discuss whether the olfactory deficiencies caused by CKD are the result of the induced inflammatory state, the hyperprolactinemia, or a combination of both.

OPEN ACCESS

Edited by:

Beatriz Gomez-Gonzalez, Universidad Autonoma Metropolitana, Iztapalapa, Mexico

Reviewed by:

Santos Ángel Depine, Simón Bolívar University, Colombia Pierre-Yves Martin, Hôpitaux Universitaires de Genève, Switzerland

*Correspondence:

Teresa Morales marter@unam.mx

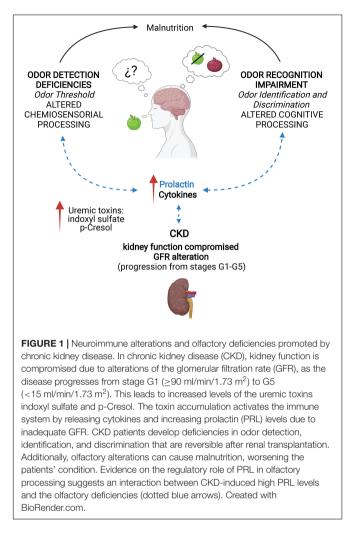
Received: 24 August 2021 Accepted: 07 January 2022 Published: 31 January 2022

Citation:

Corona R, Ordaz B, Robles-Osorio L, Sabath E and Morales T (2022) Neuroimmunoendocrine Link Between Chronic Kidney Disease and Olfactory Deficits. Front. Integr. Neurosci. 16:763986. doi: 10.3389/fnint.2022.763986 Keywords: chronic kidney disease, olfactory deficit, neuroinflammation, prolactin, olfactory epithelium, olfactory bulb

INTRODUCTION

The kidneys are in charge of filtering and eliminating the metabolic products and toxins from the blood, maintaining the control of the extracellular fluid, and the electrolyte and acid-base balance (Dhondup and Qian, 2017). Chronic kidney disease (CKD) is a major public health problem affecting approximately 9–12% of the population worldwide (Ji-Cheng and Lu-Xia, 2019; Cockwell and Fisher, 2020). In Mexico, diabetes and hypertension, two chronic metabolic diseases, are associated with CKD; studies on the latter's prevalence and incidence are scarce (Gutierrez-Padilla et al., 2010; Aguilar and Madero, 2019).



CHRONIC KIDNEY DISEASE

CKD is defined as structural and functional abnormalities in the kidney for more than 3 months, mainly caused by deficient glomerular function (Figure 1; No Author, 2013; Levey et al., 2020). The glomerular filtration rate (GFR) is used as an overall index of kidney function. CKD is classified to facilitate the diagnosis and treatment of patients with kidney damage and is based on an estimated GFR for three consecutive months. The classification is divided into five stages (G1-G5): normal to mildly decreased GFR (G1–G2; \geq 90 and 60–89 ml/min/1.73 m², respectively), moderately to severely decreased GFR (G3-G4; 30-59 and 15-29 ml/min/1.73 m², respectively) and kidney failure (G5; $< 15 \text{ ml/min}/1.73 \text{ m}^2$) (Levey et al., 2020). When GFR is < 15 ml/min/1.73 m² (G5), patients are diagnosed with end-stage kidney disease (ESKD) and usually require a renal replacement therapy that could be dialysis or kidney transplantation (Krajewska et al., 2020).

CKD leads to uremia, a condition induced by the accumulation of uremic toxins in the body (Cigarran Guldris et al., 2017). Classically, the uremic toxins are classified based on their physicochemical characteristics into small water-soluble

compounds, protein-bound, and middle molecules (Cobo et al., 2018; Vanholder et al., 2018). Uremic toxin retention negatively interacts with a variety of biological systems, and the progression of CKD to ESKD is strongly associated with the accumulation of uremic metabolites in the blood, with concentrations that reach 10–100-fold in CKD patients compared to the healthy population (Bobot et al., 2020).

The origin of uremic toxins in CKD is manifold. In CKD, dysbiosis in the intestinal microflora with increased pathogenic flora occurs (Cigarran Guldris et al., 2017). Uremic toxins are generated through protein fermentation by colonic microbiota (Rysz et al., 2021). Proteins degraded in the colon are broken down by intestinal bacteria to metabolites, such as phenols and indoles (Cigarran Guldris et al., 2017), and subsequently eliminated in the feces. However, some of them are absorbed and eliminated by the kidney (Cigarran Guldris et al., 2017).

In CKD, the uremic toxins generated by the intestinal microflora are protein-bound indoles and phenols (Rysz et al., 2021). Indole is a metabolite of the dietary amino acid L-tryptophan after fermentation by intestinal bacteria, which is rapidly absorbed by intestinal epithelial cells and subsequently sulfated to indoxyl sulfate (IS) in the liver (Cigarran Guldris et al., 2017). IS directly induces apoptotic and necrotic cell death of tubular cells, increases oxidative stress and the production of transforming growth factor $\beta 1$ (TGF- $\beta 1$), decreases the intracellular glutathione, and promotes renal fibrosis with the consequent decline in the kidney function (Figure 1; Cheng et al., 2020). The most studied phenolic uremic toxin is p-Cresol, which is generated by the intestinal microbial breakdown of tyrosine/phenylalanine and causes endothelial dysfunction and vascular calcification among other effects (Opdebeeck et al., 2019). In proximal tubular cells, it increases NADPH activity, mRNA levels of inflammatory cytokines, and the secretion of TGF-β1 (Watanabe et al., 2013). P-Cresol is associated with renal fibrosis and progression of CKD (Figure 1; Watanabe et al., 2013; Opdebeeck et al., 2019).

Oxidative stress and chronic inflammation have a central role in the pathophysiological process of uremia, causing secondary complications in various systems, among them the central nervous system (CNS) (Watanabe et al., 2014; Tsuruya and Yoshida, 2018). More specifically, patients commonly present neurological disorders such as restless leg syndrome (Safarpour et al., 2020), dementia, stroke (Bugnicourt et al., 2013), and uremic encephalopathy (Olano et al., 2021). In animal models, chronic exposure to IS promotes its accumulation in the brain stem, reducing local monoamine levels and leading to reduced locomotor and exploratory activity (Karbowska et al., 2020). IS promotes the expression of oxidative stress markers and inflammation in glial cells compromising their functioning and leading to neurodegeneration (Adesso et al., 2017).

CKD is characterized by a low-grade systemic inflammatory status that plays a key role in the progression of the disease and the increased morbidity and mortality (Mihai et al., 2018), which are also affected by malnutrition and chronic inflammation (Tinti et al., 2021). As illustrated in **Figure 1**, CKD is associated with the dysregulation of synthesis, release, and degradation of soluble molecules of the immune system, the disruption of cytokines and inflammatory mediators, and decreased adrenal clearance accounting for high levels of circulating cytokines (Rossaint et al., 2016). Inflammatory processes are highly influenced by sex hormones, with males being more susceptible to exacerbation of CKD (Kang et al., 2014; Franco-Acevedo et al., 2021).

Endocrine System and Chronic Kidney Disease

Sex and gender disparities exists in CKD: the worldwide data analysis of the 2016 Global Burden of Disease, Injuries, and Risk Factors Study revealed that the CKD prevalence is higher in women than in men (Bikbov et al., 2018). However, the CKD progression and risk of death is lower in women, while men tend to escalate more rapidly to ESKD (Brar and Markell, 2019). In this vein, as pointed out in a recent review by Machluf et al. (2020), sex differences and their relationship to CKD incidence are complex, and various factors, such as age, ethnicity, lifestyle habits, and access to health care, should be considered (Mills et al., 2015; Bikbov et al., 2018; Carrero et al., 2018; Machluf et al., 2020).

The deficient and progressive GFR leads to renal damage and CKD with some sex differences explaining part of the sex disparities (Brar and Markell, 2019). For instance, men have larger and heavier kidneys compared to women (Miletić et al., 1998; Neugarten et al., 2002; Sabolic et al., 2007; Franco-Acevedo et al., 2021; Lima-Posada and Bobadilla, 2021). In rodents, males develop glomerular hypertrophy, tubular fibrosis, proteinuria, and enhanced oxidative stress after kidney ischemiareperfusion injury, while females do not present CKD-related deterioration, suggesting that testosterone and estrogen may contribute to the susceptibility and progression of the disease (Lima-Posada et al., 2017). Furthermore, gonadectomized males presented reduced inflammation with decreased expression of pro-inflammatory cytokines, and testosterone replacement exacerbated the inflammatory response. However, in females, ovariectomy aggravated the inflammation but treatment with estrogen did not completely revert the effect (Kang et al., 2014).

It is not surprising that sex hormones influence kidney function since androgen, estrogen, and progesterone receptors exist in this organ (Davidoff et al., 1980; Takeda et al., 1990; Bhat et al., 1993; Han et al., 2001; Sabolic et al., 2007). In humans, sex hormones are also produced within the kidney, raising the possibility of autocrine and paracrine regulation of renal functions (Sharpe, 1998; Quinkler et al., 2003). Locally produced sexual hormones activate their receptors even in the absence of circulating hormones (Martucci and Fishmann, 1993; Dalla Valle et al., 2004). Furthermore, several studies on CDK show its association with elevated concentrations of prolactin (PRL) (**Figure 1**; Franco-Acevedo et al., 2021).

Prolactin and Chronic Kidney Disease

PRL is a peptide hormone synthesized in the anterior pituitary gland and released into the systemic circulation to carry out more than 300 biological actions: it is associated with the reproductive system, including the classical lactogenic action in the mammary gland, osmoregulation, immune response, brain function, and behavior (Ben-Jonathan et al., 1996). The PRL receptor is a member of the class I cytokine-hematopoietin

receptor superfamily, widely expressed in different systems (Ma et al., 2005). Within the CNS, PRL actions include the control of its secretion (Freeman et al., 2000), neurogenesis in the olfactory bulb (Shingo et al., 2003), and neuroprotection (Morales, 2011), among others. The secretion of PRL from the pituitary is regulated by the inhibitory action of dopamine, released from the tuberoinfundibular (TIDA) neurons of the arcuate nucleus (Freeman et al., 2000). Dopamine tonically inhibits the secretion of PRL (Demaria et al., 2000) through the short feedback loop between lactotrophs and TIDA neurons, which is largely responsible for PRL homeostasis (Grattan et al., 2001).

The PRL regulatory loop might be disrupted in CKD since elevated concentrations of PRL or hyperprolactinemia are present in \sim 30% of the patients in the early stages and 60–80% in the advanced stages of CKD (Lo et al., 2017; Dourado et al., 2020). To date, there is no evidence on whether hyperprolactinemia is caused by deficient metabolic depuration or the central dysregulation of PRL secretion (Carrero et al., 2012; Gungor et al., 2013), but deleterious effects of hyperprolactinemia are known to include endocrine, metabolic, and immune effects (Borba et al., 2018).

As for the systemic accumulation of PRL and its inadequate clearance related to deficient renal filtration, there is some evidence to consider. This hormone participates in osmoregulation, principally in salt and water metabolism (Loretz and Bern, 1982). Expression of the PRL gene and the binding of the hormone have been identified in the Bowman's capsule and proximal tubule cells of the kidney (Sakai et al., 1999). A large percentage of PRL in the renal artery is metabolized and eliminated by the kidneys (Emmanouel et al., 1981). PRL reduces sodium, potassium, and water excretion (Stier et al., 1984), while unilateral nephrectomy and water deprivation significantly increase serum PRL levels (Horrobin et al., 1971; Loretz and Bern, 1982; Franco-Acevedo et al., 2021).

The PRL receptor is also expressed in immune system cells, including lymphocytes, macrophages, and thymic epithelial cells (Bouchard et al., 1999), where the ligand acts as an immunomodulatory cytokine (Leite De Moraes et al., 1995). Hyperprolactinemia is associated with autoimmune diseases such as lupus and multiple sclerosis (Borba et al., 2018) and possibly contributes to the inflammatory mechanisms associated with CKD (Tinti et al., 2021).

A recent study addressing the consequences of elevated PRL in CKD proposed that the diagnosis of hyperprolactinemia in this population is difficult and dialysis therapies, such as conventional hemodialysis and peritoneal dialysis, do not normalize PRL levels (Lo et al., 2017). However, renal transplantation and the subsequent improvement in glomerular filtration result in the normalization of serum PRL levels (Bry-Gauillard et al., 1999; Saha et al., 2002). Studies using the dopaminergic agonists cabergoline and bromocriptine in CKD are scarce. However, when prescribed for clinical conditions such as galactorrhea or hypogonadism, their use is safe (Degli Esposti et al., 1985; Salmela et al., 2006).

Dopaminergic neurons' functionality and survival are sensitive to cytokines in chronic inflammation conditions (Felger and Miller, 2012). TIDA neurons have estrogen receptors (Mitchell et al., 2003; Ribeiro et al., 2015) and chronic exposure to estradiol affects them, leading to hyperprolactinemia. This initiates an inflammatory cascade in the arcuate nucleus: increased cytokine and nitric oxide production lead to reduced TIDA neuron functioning, which culminates in lower dopamine synthesis and high levels of PRL that increase the risk for mammary and pituitary tumor development (Gilbreath et al., 2019). This process highlights the importance of inflammatory regulators in pathological alterations of the PRL system, by "highjacking" the TIDA circuit and ultimately promoting hyperprolactinemia.

Interestingly, experimental evidence suggests that a hyperprolactinemic state might alter the olfactory function (Corona et al., 2021). Adequate olfactory information processing relies on the precise function of the olfactory bulb (OB), which involves continuous plastic changes that tune the circuit (Shingo et al., 2003; Corona and Levy, 2015). PRL regulates OB neurogenesis, necessary for healthy olfactory functions and behavioral responses (Shingo et al., 2003; Corona and Levy, 2015). Recently, we demonstrated that juvenile hyperprolactinemia alters OB mitral cell activation in female mice exposed to social odors, suggesting that PRL participates in the maturation and response of the OB circuits (Corona et al., 2021). Interestingly, OB mitral cells express PRL receptors (Freemark et al., 1996; Canavan et al., 2011; de Moura et al., 2015), implying that hyperprolactinemia possibly regulates OB functioning and, as a result, the olfactory capacity in CKD.

OLFACTORY FUNCTION IN DISEASE

Olfactory Impairments in Disease

Human olfaction involves complex processes that detect, discriminate, and code thousands of odors (McGann, 2017). Olfactory impairments reflect deficient functionality in the periphery with variable odor thresholds or compromised central processing of the olfactory information resulting in diminished odor discrimination and identification (Frasnelli et al., 2002; Raff et al., 2008; Robles-Osorio et al., 2020; Iacono et al., 2021; Yusuf et al., 2021a).

Within the nasal cavity resides the epithelium, where odor molecules bind to the olfactory receptors (Ihara et al., 2013). This information is transduced into a neural signal and sent through the axons of the olfactory sensory neurons (OSN) to synapse the projection mitral or tufted cells, in the glomeruli of the OBs (Martin-Lopez et al., 2012). This first step is essential for olfaction. The OSNs have a lifelong regenerative capacity necessary for adequate olfaction (Lazarini and Lledo, 2011). Reduced tissue maintenance by poor basal stem cell proliferation within the epithelium can generate hyposmia (a significant reduction in olfactory abilities), anosmia (complete loss of olfaction), or olfactory threshold deficiencies due to the lack of neural signals in the OB (Goncalves and Goldstein, 2016; McGann, 2017).

There is a growing interest in olfactory disorders owing to their role as leading indicators in numerous pathologies. Olfactory disorders are associated with many causes like nasal inflammation, upper olfactory tract abnormalities, neurological pathologies, and aging (Huttenbrink et al., 2013; Schriever et al., 2014; Kar et al., 2015). Moreover, olfaction strongly influences human behavior: a wide range of environmental odors primes our safety (e.g., avoiding fire, gas leakage, or rotten food) but also influences eating behavior (e.g., associating memories and emotions with food) and social interactions (McGann, 2017). Initially, olfactory impairments remain unnoticed, forcing the individuals into situations where they must deal with daily issues like feeding, safety, and social situations, which, in many cases, lead to anxiety and depression (Sivam et al., 2016; Kondo et al., 2020). Olfactory deficits promote food aversion and, consequently, malnutrition (Frasnelli et al., 2002).

Olfactory Function and Inflammation

Inflammatory conditions of the nasal cavity reduce the proliferation of the basal stem cells, while the newly differentiated OSNs do not survive easily (Chen et al., 2019). Inflammation induces OB atrophy by interfering with the input from the OSN axons, reducing the apical dendrites of mitral and tufted cells, and shrinking the glomerular layer thickness (Chen et al., 2019). Also, OB inflammation is accompanied by glial activation and high expression of the cytokines interleukin 1B and tumor necrosis factor α (Hasegawa-Ishii et al., 2020). The basal stem cells directly regulate the inflammation, the basal cells release cytokines and chemokines to shut down the stem cell proliferation, generating olfactory dysfunctions (Chen et al., 2019).

Damage to the nasal epithelium is reversible when the inflammation is reduced or controlled (Hasegawa-Ishii et al., 2020). This recovery probably explains the discrepancies observed in odor threshold deficiencies, which improve after dialysis treatment (Landis et al., 2011). The complete recovery of odor discrimination and identification after renal transplantation is due to the decreased generalized inflammation that allows for sufficient neuronal information processing (Vreman et al., 1980; Griep et al., 1997; Frasnelli et al., 2002; Landis et al., 2011; Koseoglu et al., 2017; Iacono et al., 2021; Yusuf et al., 2021a). Interestingly, odor stimulation contributes to the recovery of OB atrophy (Hasegawa-Ishii et al., 2020). Olfactory training could serve as a novel strategy and additional treatment in CKD since it has been demonstrated to ameliorate olfactory deficits in aging and various pathologies (Livermore and Hummel, 2004; Hummel et al., 2009; Haehner et al., 2013; Schriever et al., 2014).

Olfactory Impairments in Chronic Kidney Disease

CKD and particularly ESKD patients present significantly decreased olfactory function (**Figure 1**; Frasnelli et al., 2002; Landis et al., 2011; Attems et al., 2015; Kondo et al., 2020; Robles-Osorio et al., 2020; Iacono et al., 2021; Yusuf et al., 2021a,b). Additionally, CKD patients follow a controlled diet with reduced calorie uptake, and \sim 70% of them suffer from loss of taste (Frasnelli et al., 2002), which can lead to insufficient or unbalanced feeding behaviors. Malnutrition, one of the major causes of morbidity and mortality in CKD (**Figure 1**;

Anderson et al., 2016), begins when renal functions decline, promoting a gradually reduced protein and total calorie intake, leading to cachexia, and increasing the risk of death (Nigwekar et al., 2017; Yusuf et al., 2021b). Although the straightforward link between impaired olfactory function and the nutritional state is still debatable, poor odor perception is associated with elevated serum urea and a high protein catabolic rate (Griep et al., 1997). The pathological changes underlying the malnutritioninflammatory state may also contribute to impaired olfactory function in ESKD patients (Frasnelli et al., 2002; Raff et al., 2008; Koseoglu et al., 2017; Robles-Osorio et al., 2020; Iacono et al., 2021; Yusuf et al., 2021a).

Up to date, several studies have highlighted the importance of understanding the link between CKD and olfactory sense loss due to its relevance in nutrition and quality of life of the patients (Robles-Osorio et al., 2020; Yusuf et al., 2021a). In recent studies with CKD and ESKD patients, \sim 70% presented olfactory dysfunctions, with 72% (Yusuf et al., 2021a) to 83% (Koseoglu et al., 2017) of CKD patients presenting some degree of hyposmia and, in extreme cases, 5% presenting anosmia (Yusuf et al., 2021a).

Renal failure pathologies affect the function of the CNS. In CKD, there is a generalized disequilibrium of biochemical parameters that provokes systemic inflammation (Adesso et al., 2017; Cobo et al., 2018). Odor discrimination and identification are compromised in CKD due to deficient neuronal functioning (Frasnelli et al., 2002). The lack of efficient glomerulus filtration generates endogenous intoxication that damages axons and dendrites, consequently leading to neurodegeneration (Bobot et al., 2020). Furthermore, the impaired blood-brain barrier by the neuropathic uremia promotes changes in the hydration levels within the brain and the cerebral blood flow, altering the synthesis of neurotransmitters (Wiggins et al., 1984; Griep et al., 1997; Burn and Bates, 1998; Frasnelli et al., 2002; Raff et al., 2008; Goncalves and Goldstein, 2016; Nigwekar et al., 2017; Robles-Osorio et al., 2020). It has been consistently shown that CKD and -even more prominently- ESKD patients present deficits in odor discrimination and identification but the odor threshold impairments are less evident or inconsistent across different studies (Schiffman et al., 1978; Griep et al., 1997; Frasnelli et al., 2002; Raff et al., 2008; Landis et al., 2011; Nigwekar et al., 2017; Robles-Osorio et al., 2020; Iacono et al., 2021; Yusuf et al., 2021a).

DISCUSSION

CKD is characterized by a persistent deficiency in GFR that leads to increased levels of uremic toxins, such as indoxyl sulfate and p-Cresol. The accumulation of uremic toxins promotes the activation of the immune system through the release of cytokines and the lack of clearance of PRL due to inadequate glomerular filtration. CKD patients develop deficiencies in odor detection because of alterations in the olfactory epithelium that modify the odor threshold and are regulated by inflammation. Odor identification and discrimination impairments frequently occur in CKD due to chronic intoxication and inflammation. These olfactory impairments are reversible after renal transplantation. The olfactory alterations promoted by CKD can cause malnutrition, exacerbating the patients' condition. Evidence on the regulatory role of PRL in olfactory processing suggests an interaction between CKD-induced high PRL levels and the olfactory deficiencies (**Figure 1**).

In CKD, the deficient elimination of uremic toxins increases the levels of inflammatory cytokines, causing oxidative stress and consequently cellular damage to a variety of systems. CKD is associated with various hormonal alterations. Hyperprolactinemia is a frequent endocrine abnormality in this population as it has been reported in more than 30% of CKD patients (Lo et al., 2017; Serret-Montaya et al., 2020; Franco-Acevedo et al., 2021). PRL levels are directly associated with endothelial dysfunction and increased risk of cardiovascular events and mortality in CKD (Carrero et al., 2012). Although the mechanisms of hyperprolactinemia in CKD are yet to be unraveled, it could be a result of PRL accumulation due to deficient renal clearance. Among the causes of hyperprolactinemia in CKD, the following stand out: (a) the established negative feedback loop in TIDA neurons is "highjacked," promoting PRL production, (b) estrogens increase the inflammatory response, reducing dopamine production, therefore, enhancing PRL secretion, and (c) renal dysfunction directly alters the PRL metabolic clearance (Sievertsen et al., 1980).

The inflammation-oxidative stress "duo" causes cognitive impairments in CKD patients (Tsuruya and Yoshida, 2018), as the chronic and systemic inflammatory state deteriorates the odor detection at the level of the olfactory epithelium, leading to odor threshold deficiencies which can be reverted after controlling the inflammation (Hasegawa-Ishii et al., 2020). CKD-induced neuropathy compromises odor discrimination and identification, affecting the quality of life and leading to malnutrition and metabolic problems (Frasnelli et al., 2002). In some cases, these alterations are improved after dialysis treatment, while renal transplantation leads to recovery of the olfaction (Robles-Osorio et al., 2020; Yusuf et al., 2021a).

Understanding the link between CKD and the loss of the sense of smell is relevant to the nutritional status and the quality of life of patients. Research in animal models and humans should be directed toward the understanding of the relationship between CKD-induced hyperprolactinemia and the loss or alteration of olfaction. New insights into the timing and progression of the disease and the role of inflammation on neuroendocrine systems, such as PRL, could expand our knowledge and open an avenue toward the improvement of the quality of life in CKD patients. Additionally, the development of a prevention strategy with screening olfactory tests in the early stages of CKD can complement the therapeutical approaches together with the introduction of physical activity and dietary programs.

AUTHOR CONTRIBUTIONS

RC, BO, LR-O, ES, and TM wrote and edited the review article. RC and BO created the figure. All authors contributed to the article and approved the submitted version.

FUNDING

Cited research supported by UNAM-DGAPA-PAPIIT (IA202218, IA200820) and CONACyT (A1S8948).

REFERENCES

- Adesso, S., Magnus, T., Cuzzocrea, S., Campolo, M., Rissiek, B., Paciello, O., et al. (2017). Indoxyl sulfate affects glial function increasing oxidative stress and neuroinflammation in chronic kidney disease: interaction between astrocytes and microglia. *Front. Pharmacol.* 8:370. doi: 10.3389/fphar.2017.00370
- Aguilar, D. J., and Madero, M. (2019). Other Potential CKD hotspots in the world: the cases of mexico and the united states. *Semin. Nephrol.* 39, 300–307. doi: 10.1016/j.semnephrol.2019.02.008
- Anderson, C. A., Nguyen, H. A., and Rifkin, D. E. (2016). Nutrition interventions in chronic kidney disease. *Med. Clin. North Am.* 100, 1265–1283.
- Attems, J., Walker, L., and Jellinger, K. A. (2015). Olfaction and aging: a minireview. Gerontology 61, 485–490. doi: 10.1159/000381619
- Ben-Jonathan, N., Mershon, J. L., Allen, D. L., and Steinmetz, R. W. (1996). Extrapituitary prolactin: distribution, regulation, functions, and clinical aspects. *Endoc. Rev.* 17, 639–669. doi: 10.1210/edrv-17-6-639
- Bhat, H. K., Hacker, H. J., Bannasch, P., Thompson, E. A., and Liehr, J. G. (1993). Localization of estrogen receptors in interstitial cells of hamster kidney and in estradiol-induced renal tumors as evidence of the mesenchymal origin of this neoplasm. *Cancer Res.* 53, 5447–5451.
- Bikbov, B., Perico, N., and Remuzzi, G. (2018). Disparities in chronic kidney disease prevalence among males and females in 195 countries: analysis of the global burden of disease 2016 study. *Nephron* 139, 313–318. doi: 10.1159/000489897
- Bobot, M., Thomas, L., Moyon, A., Fernandez, S., McKay, N., Balasse, L., et al. (2020). Uremic toxic blood-brain barrier disruption mediated by ahr activation leads to cognitive impairment during experimental renal dysfunction. *J. Am. Soc. Nephrol.* 31, 1509–1521. doi: 10.1681/ASN.2019070728
- Borba, V. V., Zandman-Goddard, G., and Shoenfeld, Y. (2018). Prolactin and Autoimmunity. *Front. Immunol.* 9:73.
- Bouchard, B., Ormandy, C. J., Di Santo, J. P., and Kelly, P. A. (1999). Immune system development and function in prolactin receptor-deficient mice. *J. Immunol.* 163, 576–582.
- Brar, A., and Markell, M. (2019). Impact of gender and gender disparities in patients with kidney disease. *Curr. Opin. Nephrol. Hyperten.* 28, 178–182. doi: 10.1097/MNH.000000000000482
- Bry-Gauillard, H., Touraine, P., Mamzer-Bruneel, M. F., Simoes-Vaz, A., Kuttenn, F., and Legendre, C. (1999). Complete regression of a major hyperprolactinaemia after renal transplantation. *Nephrol. Dial. Transpl.* 14, 466–468. doi: 10.1093/ndt/14.2.466
- Bugnicourt, J. M., Godefroy, O., Chillon, J. M., Choukroun, G., and Massy, Z. A. (2013). Cognitive disorders and dmentia in CKD: the neglected kidney-brainaxis. J. Am. Soc. Nephrol. 24, 353–363. doi: 10.1681/ASN.2012050536
- Burn, D. J., and Bates, D. (1998). Neurology and the kidney. J. Neurol. Neurosurg. Psychiatry 65, 810–821. doi: 10.1136/jnnp.65.6.810
- Canavan, S. V., Mayes, L. C., and Treloar, H. B. (2011). Changes in maternal gene expression in olfactory circuits in the immediate postpartum period. *Front. psychiatry* 2:40. doi: 10.3389/fpsyt.2011.00040
- Carrero, J. J., Hecking, M., Chesnaye, N. C., and Jager, K. J. (2018). Sex and gender disparities in epidemiology and outcomes of chronic kidney disease. *Nat. Rev. Nephrol.* 14, 151–164. doi: 10.1038/nrneph.2017.181
- Carrero, J. J., Kyriazis, J., Sonmez, A., Tzanakis, I., Qureshi, A. R., Stenvinkel, P., et al. (2012). Prolactin levels, endothelial dysfunction, and the risk of cardiovascular events and mortality in patients with CKD. *Clin. J. Am. Soc. Nephrol.* 7, 207–215. doi: 10.2215/CJN.06840711
- Chen, M., Reed, R. R., and Lane, A. P. (2019). Chronic inflammation directs an olfactory stem cell functional switch from neuroregeneration to immune defense. *Cell Stem cell* 25:e505. doi: 10.1016/j.stem.2019.08.011
- Cheng, T. H., Ma, M. C., Liao, M. T., Zheng, C. M., Lu, K. C., Liao, C. H., et al. (2020). Indoxyl sulfate, a tubular toxin, contributes to the development of chronic kidney disease. *Toxins* 12:684. doi: 10.3390/toxins12110684

ACKNOWLEDGMENTS

We thank Ilektra Anagnostou, for editing the English version of this manuscript.

- Cigarran Guldris, S., GonzalezParra, E., and CasesAmenos, A. (2017). Gut microbiota in chronic kidney disease. *Nefrologia : Publicacion Oficial De La Sociedad Espanola Nefrologia* 37, 9–19.
- Cobo, G., Lindholm, B., and Stenvinkel, P. (2018). Chronic inflammation in endstage renal disease and dialysis. *Nephrol. Dial Transpl.* 33, iii35–iii40. doi: 10. 1093/ndt/gfy175
- Cockwell, P., and Fisher, L. A. (2020). The globan burden of chronic kidney disease. *Lancet* 395, 662–664.
- Corona, R., Jayakumar, P., Carbajo Mata, M. A., Del Valle-Diaz, M. F., Luna-Garcia, L. A., and Morales, T. (2021). Sexually dimorphic effects of prolactin treatment on the onset of puberty and olfactory function in mice. *Gener. Compar. Endocrinol.* 301:113652. doi: 10.1016/j.ygcen.2020.11 3652
- Corona, R., and Levy, F. (2015). Chemical olfactory signals and parenthood in mammals. *Hormon. Behav.* 68, 77–90. doi: 10.1016/j.yhbeh.2014.06.018
- Dalla Valle, L., Toffolo, V., Vianello, S., Belvedere, P., and Colombo, L. (2004). Expression of cytochrome P450c17 and other steroid-converting enzymes in the rat kidney throughout the life-span. J. Ster. Biochem. Mol. Biol. 91, 49–58. doi: 10.1016/j.jsbmb.2004.01.008
- Davidoff, M., Caffier, H., and Schiebler, T. H. (1980). Steroid hormone binding receptors in the rat kidney. *Histochemistry* 69, 39–48. doi: 10.1007/BF00508365
- de Moura, A. C., Lazzari, V. M., Becker, R. O., Gil, M. S., Ruthschilling, C. A., Agnes, G., et al. (2015). Gene expression in the CNS of lactating rats with different patterns of maternal behavior. *Neurosci. Res.* 99, 8–15. doi: 10.1016/ j.neures.2015.05.003
- Degli Esposti, E., Sturani, A., Santoro, A., Zuccala, A., Chiarini, C., and Zucchelli, P. (1985). Effect of bromocriptine treatment on prolactin, noradrenaline and blood pressure in hypertensive haemodialysis patients. *Clin. Sci.* 69, 51–56. doi: 10.1042/cs0690051
- Demaria, J. E., Nagy, G. M., Lerant, A. A., Fekete, M. I., Levenson, C. W., and Freeman, M. E. (2000). Dopamine transporters participate in the physiological regulation of prolactin. *Endocrinology* 141, 366–374. doi: 10.1210/endo.141.1. 7281
- Dhondup, T., and Qian, Q. (2017). Acid-Base and electrolyte disorders in patients with and without chronic kidney disease: an update. *Kidney Dis.* 3, 136–148. doi: 10.1159/000479968
- Dourado, M., Cavalcanti, F., Vilar, L., and Cantilino, A. (2020). Relationship between prolactin, chronic kidney disease, and cardiovascular risk. *Int. J. Endocrinol.* 2020, 1–6 doi: 10.1155/2020/9524839
- Emmanouel, D. S., Fang, V. S., and Katz, A. I. (1981). Prolactin metabolism in the rat: role of the kidney in degradation of the hormone. *Am. J. Physiol.* 240, F437–F445. doi: 10.1152/ajprenal.1981.240.5.F437
- Felger, J. C., and Miller, A. H. (2012). Cytokine effects on the basal ganglia and dopamine function: the subcortical source of inflammatory malaise. *Front. Neuroendocrinol.* 33:315–327. doi: 10.1016/j.yfrne.2012.09.003
- Franco-Acevedo, A., Echavarría, R., and Melo, Z. (2021). Sex differences in renal function: participation of gonadal hormones and prolactin. *Endocrines* 2, 185– 202. doi: 10.1016/j.envint.2017.10.006
- Frasnelli, J. A., Temmel, A. F., Quint, C., Oberbauer, R., and Hummel, T. (2002). Olfactory function in chronic renal failure. Am. J. Rhinol. 16, 275–279. doi: 10.1177/194589240201600511
- Freeman, M. E., Kanyicska, B., Lerant, A., and Nagy, G. (2000). Prolactin: structure, function, and regulation of secretion. *Physiol. Rev.* 80, 1523–1631. doi: 10.1152/ physrev.2000.80.4.1523
- Freemark, M., Driscoll, P., Andrews, J., Kelly, P. A., and Royster, M. (1996). Ontogenesis of prolactin receptor gene expression in the rat olfactory system: potential roles for lactogenic hormones in olfactory development. *Endocrinology* 137, 934–942. doi: 10.1210/endo.137.3.8603606
- Gilbreath, E. T., Jaganathan, L., Subramanian, M., Balasubramanian, P., Linning, K. D., MohanKumar, S. M. J., et al. (2019). Chronic estrogen affects TIDA

neurons through IL-1beta and NO: effects of aging. J. Endocrinol. 240, 157–167. doi: 10.1530/JOE-18-0274

- Goncalves, S., and Goldstein, B. J. (2016). Pathophysiology of olfactory disorders and potential treatment strategies. *Curr. Otorhinolaryngol. Rep.* 4, 115–121. doi: 10.1007/s40136-016-0113-5
- Grattan, D. R., Xu, J., McLachlan, M. J., Kokay, I. C., Bunn, S. J., Hovey, R. C., et al. (2001). Feedback regulation of PRL secretion is mediated by the transcription factor, signal transducer, and activator of transcription 5b. *Endocrinology* 142, 3935–3940. doi: 10.1210/endo.142.9.8385
- Griep, M. I., Van der Niepen, P., Sennesael, J. J., Mets, T. F., Massart, D. L., and Verbeelen, D. L. (1997). Odour perception in chronic renal disease. *Nephrol. Dial. Transpl.* 12, 2093–2098.
- Gungor, O., Kircelli, F., Voroneanu, L., Covic, A., and Ok, E. (2013). Hormones and arterial stiffness in patients with chronic kidney disease. J. Atheroscl. Thromb. 20, 698–707. doi: 10.5551/jat.18580
- Gutierrez-Padilla, J. A., Mendoza-García, M., Plascencia-Pérez, S., Renoirte-López, K., García-García, G., Lloyd, A., et al. (2010). Screening for CKD and cardiovascular disease risk factors using mobile clinics in Jalisco, México. Am. J. Kidney Dis. 55, 474–484. doi: 10.1053/j.ajkd.2009.0 7.023
- Haehner, A., Tosch, C., Wolz, M., Klingelhoefer, L., Fauser, M., Storch, A., et al. (2013). Olfactory training in patients with Parkinson's disease. *PloS One* 8:e61680. doi: 10.1371/journal.pone.0061680
- Han, H. J., Park, S. H., Park, H. J., Lee, J. H., Lee, B.-C., and Hwang, W. S. (2001). Effects of Sex Hormones on Na+/Glucose cotransporter of renal proximal tubular cells following oxidant injury. *Kidney Blood Press Res.* 24, 159–165. doi: 10.1159/000054223
- Hasegawa-Ishii, S., Imamura, F., Nagayama, S., Murata, M., and Shimada, A. (2020). Differential effects of nasal inflammation and odor deprivation on layerspecific degeneration of the mouse olfactory bulb. *eNeuro* 7:2020. doi: 10.1523/ ENEURO.0403-19.2020
- Horrobin, D. F., Lloyd, I. J., Lipton, A., Burstyn, P. G., Durkin, N., and Muiruri, K. L. (1971). Actions of prolactin on human renal function. *Lancet* 2, 352–354. doi: 10.1016/s0140-6736(71)90065-1
- Hummel, T., Rissom, K., Reden, J., Hahner, A., Weidenbecher, M., and Huttenbrink, K. B. (2009). Effects of olfactory training in patients with olfactory loss. *Laryngoscope* 119, 496–499. doi: 10.1002/lary. 20101
- Huttenbrink, K. B., Hummel, T., Berg, D., Gasser, T., and Hahner, A. (2013). Olfactory dysfunction: common in later life and early warning of neurodegenerative disease. *Deutsch. Arzteblatt Int.* 110:e1. doi: 10.3238/arztebl. 2013.0001
- Iacono, V., Lombardi, G., Ottaviano, G., Gambaro, G., and Zaza, G. (2021). Impact of renal replacement therapies on olfactory ability: results of a cross-sectional case control study. J. Nephrol. 2021:983. doi: 10.1007/s40620-021-00983-6
- Ihara, S., Yoshikawa, K., and Touhara, K. (2013). Chemosensory signals and their receptors in the olfactory neural system. *Neuroscience* 254, 45–60. doi: 10.1016/ j.neuroscience.2013.08.063
- Ji-Cheng, L., and Lu-Xia, Z. (2019). Prevalence and disease burden of chronic kidney disease. Adv. Exp. Med. Biol. 1165, 3–15. doi: 10.1007/978-981-13-88 71-2_1
- Kang, K. P., Lee, J. E., Lee, A. S., Jung, Y. J., Kim, D., Lee, S., et al. (2014). Effect of gender differences on the regulation of renal ischemia-reperfusion-induced inflammation in mice. *Mol. Med. Rep.* 9, 2061–2068. doi: 10.3892/mmr.2014. 2089
- Kar, T., Yildirim, Y., Altundag, A., Sonmez, M., Kaya, A., Colakoglu, K., et al. (2015). The relationship between age-related macular degeneration and olfactory function. *Neuro-Degener. Dis.* 15, 219–224. doi: 10.1159/00038 1216
- Karbowska, M., Hermanowicz, J. M., Tankiewicz-Kwedlo, A., Kalaska, B., Kaminski, T. W., Nosek, K., et al. (2020). Neurobehavioral effects of uremic toxin-indoxyl sulfate in the rat model. *Sci. Rep.* 10:9483. doi: 10.1038/s41598-020-66421-y
- Kondo, K., Kikuta, S., Ueha, R., Suzukawa, K., and Yamasoba, T. (2020). Agerelated olfactory dysfunction: epidemiology, pathophysiology, and clinical management. *Front. Aging Neurosci.* 12:208. doi: 10.3389/fnagi.2020.0 0208

- Koseoglu, S., Derin, S., Huddam, B., and Sahan, M. (2017). The effect of nondiabetic chronic renal failure on olfactory function. *Eur. Ann. Otorhinolaryngol. Head Neck Dis.* 134, 161–164. doi: 10.1016/j.anorl.2016.04.022
- Krajewska, J., Krajewski, W., and Zatonski, T. (2020). Otorhinolaryngological dysfuntions induced by chronic kidney disease in pre- and post-transplant stages. *Eur. Arch. Oto-Rhino-Laryngol.* 277, 1575–1591. doi: 10.1007/s00405-020-05925-9
- Landis, B. N., Marangon, N., Saudan, P., Hugentobler, M., Giger, R., Martin, P. Y., et al. (2011). Olfactory function improves following hemodialysis. *Kidney Int*, 80, 886–893. doi: 10.1038/ki.2011.189
- Lazarini, F., and Lledo, P. M. (2011). Is adult neurogenesis essential for olfaction? *Trends Neurosci.* 34, 20–30. doi: 10.1016/j.tins.2010.09.006
- Leite De Moraes, M. C., Touraine, P., Gagnerault, M. C., Savino, W., Kelly, P. A., and Dardenne, M. (1995). Prolactin receptors and the immune system. *Ann. Endocrinol.* 56, 567–570.
- Levey, A. S., Eckardt, K. U., Dorman, N. M., Christiansen, S. L., Hoorn, E. J., Ingelfinger, J. R., et al. (2020). Nomenclature for kidney function and disease: report of a kidney disease: improving global outcomes (KDIGO) Consensus Conference. *Kidney Int.* 97, 1117–1129. doi: 10.1016/j.kint.2020.0 2.010
- Lima-Posada, I., and Bobadilla, N. A. (2021). Understanding the opposite effects of sex hormones in mediating renal injury. *Nephrology* 26, 217–226. doi: 10.1111/ nep.13806
- Lima-Posada, I., Portas-Cortes, C., Perez-Villalva, R., Fontana, F., Rodriguez-Romo, R., Prieto, R., et al. (2017). Gender differences in the acute kidney injury to chronic kidney disease transition. *Sci. Rep.* 7:12270. doi: 10.1038/s41598-017-09630-2
- Livermore, A., and Hummel, T. (2004). The influence of training on chemosensory event-related potentials and interactions between the olfactory and trigeminal systems. *Chem. Senses* 29, 41–51. doi: 10.1093/chemse/bjh013
- Lo, J. C., Beck, G. J., Kaysen, G. A., Chan, C. T., Kliger, A. S., Rocco, M. V., et al. (2017). Hyperprolactinemia in end-stage renal disease and effects of frequent hemodialysis. *Hemodial. Int.* 21, 190–196. doi: 10.1111/hdi.12489
- Loretz, C. A., and Bern, H. A. (1982). Prolactin and osmoregulation in vertebrates. An update. *Neuroendocrinology* 35, 292–304. doi: 10.1159/00012 3397
- Ma, F. Y., Anderson, G. M., Gunn, T. D., Goffin, V., Grattan, D. R., and Bunn, S. J. (2005). Prolactin specifically activates signal transducer and activator of transcription 5b in neuroendocrine dopaminergic neurons. *Endocrinology* 146, 5112–5119. doi: 10.1210/en.2005-0770
- Machluf, Y., Chaiter, Y., and Tal, O. (2020). Gender medicine: Lessons from COVID-19 and other medical conditions for designing health policy. *World J. Clin. Cases* 8, 3645–3668. doi: 10.12998/wjcc.v8.i17.3645
- Martin-Lopez, E., Corona, R., and Lopez-Mascaraque, L. (2012). Postnatal characterization of cells in the accessory olfactory bulb of wild type and reeler mice. *Front. Neuroanat.* 6:15. doi: 10.3389/fnana.2012.00015
- Martucci, C. P., and Fishmann, J. (1993). P450 enzyme of estrogen metabolism. *Pharmacol. Ther.* 57, 237–257.
- McGann, J. P. (2017). Poor human olfaction is a 19th-century myth. *Science* 356:7263. doi: 10.1126/science.aam7263
- Mihai, S., Codrici, E., Popescu, I. D., Enciu, A. M., Albulescu, L., Necula, L. G., et al. (2018). Inflammation-related mechanisms in chronic kidney disease prediction, progression, and outcome. *J. Immunol. Res.* 2018:2180373. doi: 10.1155/2018/ 2180373
- Miletić, D., Fuckar, Z., Sustić, A., Mozetic, V., Stimac, D., and Zauhar, G. (1998). Sonographic measurement of absolute and relative renal length in adults. *J. Clin. Ultrasound* 26, 185–189. doi: 10.1002/(sici)1097-0096(199805)26:4<185::aidjcu1>3.0.co;2-9
- Mills, K. T., Xu, Y., Zhang, W., Bundy, J. D., Chen, C. S., Kelly, T. N., et al. (2015). A systematic analysis of worldwide population-based data on the global burden of chronic kidney disease in 2010. *Kidney Int.* 88, 950–957. doi: 10.1038/ki.201 5.230
- Mitchell, V., Loyens, A., Spergel, D. J., Flactif, M., Poulain, P., Tramu, G., et al. (2003). A confocal microscopic study of gonadotropin-releasing hormone (GnRH) neuron inputs to dopaminergic neurons containing estrogen receptor alpha in the arcuate nucleus of GnRH-green fluorescent protein transgenic mice. *Neuroendocrinology* 77, 198–207. doi: 10.1159/000069511

- Morales, T. (2011). Recent findings on neuroprotection against excitotoxicity in the hippocampus of female rats. J. Neuroendocrinol. 23, 994–1001. doi: 10.1111/j. 1365-2826.2011.02141.x
- Neugarten, J., Kasiske, B., Silbiger, S. R., and Nyengaard, J. R. (2002). Effects of sex on renal structure. Nephron 90, 139–144. doi: 10.1159/000049033
- Nigwekar, S. U., Weiser, J. M., Kalim, S., Xu, D., Wibecan, J. L., Dougherty, S. M., et al. (2017). Characterization and correction of olfactory deficits in kidney disease. J. Am. Soc. Nephrol. 28, 3395–3403. doi: 10.1681/ASN.2016121308
- No Author. (2013). Chapter 1: Definition and classification of CKD. *Kidney Int. Suppl.* 3, 19–62. doi: 10.1038/kisup.2012.64
- Olano, C. G., Akram, S. M., and Bhatt, H. (2021). *Uremic Encephalopathy*. Treasure Island, FL: StatPearls.
- Opdebeeck, B., Maudsley, S., Azmi, A., De Mare, A., De Leger, W., Meijers, B., et al. (2019). Indoxyl Sulfate and p-Cresyl sulfate promote vascular calcification and associate with glucose intolerance. *J. Am. Soc. Nephrol.* 30, 751–766. doi: 10.1681/ASN.2018060609
- Quinkler, M., Bumke-Vogt, C., Meyer, B., Bahr, V., Oelkers, W., and Diederich, S. (2003). The human kidney is a progesterone-metabolizing and androgenproducing organ. J Clin. Endocrinol. Metabol. 88, 2803–2809. doi: 10.1210/jc. 2002-021970
- Raff, A. C., Lieu, S., Melamed, M. L., Quan, Z., Ponda, M., Meyer, T. W., et al. (2008). Relationship of impaired olfactory function in ESRD to malnutrition and retained uremic molecules. *Am. J. Kidney Dis.* 52, 102–110. doi: 10.1053/j. aikd.2008.02.301
- Ribeiro, A. B., Leite, C. M., Kalil, B., Franci, C. R., Anselmo-Franci, J. A., and Szawka, R. E. (2015). Kisspeptin regulates tuberoinfundibular dopaminergic neurones and prolactin secretion in an oestradiol-dependent manner in male and female rats. J. Neuroendocrinol. 27, 88–99. doi: 10.1111/jne.12242
- Robles-Osorio, M. L., Corona, R., Morales, T., and Sabath, E. (2020). Chronic kidney disease and the olfactory system. *Nefrologia* 40, 120–125. doi: 10.1016/j. nefro.2019.04.009
- Rossaint, J., Oehmichen, J., Van Aken, H., Reuter, S., Pavenstadt, H. J., Meersch, M., et al. (2016). FGF23 signaling impairs neutrophil recruitment and host defense during CKD. J. Clin. Invest. 126, 962–974. doi: 10.1172/JCI83470
- Rysz, J., Franczyk, B., Lawinski, J., Olszewski, R., Cialkowska-Rysz, A., and Gluba-Brzozka, A. (2021). The Impact of CKD on Uremic Toxins and Gut Microbiota. *Toxins* 13:252. doi: 10.3390/toxins13040252
- Sabolic, I., Asif, A. R., Budach, W. E., Wanke, C., Bahn, A., and Burckhardt, G. (2007). Gender differences in kidney function. *Pflug. Archiv.* 455, 397–429. doi: 10.1016/b978-012440905-7/50305-4
- Safarpour, Y., Vaziri, N. D., and Jabbari, B. (2020). Movement disorders in chronic kidney disease - a descriptive review. J. Stroke Cerebrovascul. Dis. 30:105408. doi: 10.1016/j.jstrokecerebrovasdis.2020.105408
- Saha, M. T., Saha, H. H., Niskanen, L. K., Salmela, K. T., and Pasternack, A. I. (2002). Time course of serum prolactin and sex hormones following successful renal transplantation. *Nephron* 92, 735–737. doi: 10.1159/000064079
- Sakai, Y., Hiraoka, Y., Ogawa, M., Takeuchi, Y., and Aiso, S. (1999). The prolactin gene is expressed in the mouse kidney. *Kidney Int.* 55, 833–840. doi: 10.1046/j. 1523-1755.1999.055003833.x
- Salmela, O., Li, L., Mamputu, J. C., Beauchamp, F., Maingrette, F., and Renier, G. (2006). The influences of hyperprolactinemia and obesity on cardiovascular risk markers: effects of cabergoline therapy. *Clin. Endocrinol.* 64, 366–370. doi: 10.1111/j.1365-2265.2006.02469.x
- Schiffman, S. S., Nash, M. L., and Dackis, C. (1978). Reduced olfactory discrimination in patients on chronic hemodialysis. *Physiol. Behav.* 21, 239–242. doi: 10.1016/0031-9384(78)90046-x
- Schriever, V. A., Lehmann, S., Prange, J., and Hummel, T. (2014). Preventing olfactory deterioration: olfactory training may be of help in older people. J. Am. Geriat. Soc. 62, 384–386. doi: 10.1111/jgs.12669
- Serret-Montaya, J., Zurita-Cruz, J. N., Villasis-Keever, M. A., Aguilar-Kitsu, A., Zepeda-Martinez, C. C., Cruz-Anleu, I., et al. (2020). Hyperprolactinemia as a prognostic factor for menstrual disorders in female adolescents with advanced chronic kidney disease. *Pediatric Nephrol.* 35, 1041–1049. doi: 10.1007/s00467-020-04494-7
- Sharpe, R. M. (1998). The roles of oestrogen in the male. Trends Endocrinol. Metabol. 9, 371–377.

- Shingo, T., Gregg, C., Enwere, E., Fujikawa, H., Hassam, R., Geary, C., et al. (2003). Pregnancy-stimulated neurogenesis in the adult female forebrain mediated by prolactin. *Science* 299, 117–120. doi: 10.1126/science.1076647
- Sievertsen, G. D., Lim, V. S., Nakawatase, C., and Frohman, L. A. (1980). Metabolic clearance and secretion rates of human prolactin in normal subjects and in patients with chronic renal failure. J. Clin. Endocrinol. Metabol. 50, 846–852. doi: 10.1210/jcem-50-5-846
- Sivam, A., Wroblewski, K. E., Alkorta-Aranburu, G., Barnes, L. L., Wilson, R. S., Bennett, D. A., et al. (2016). Olfactory dysfunction in older adults is associated with feelings of depression and loneliness. *Chem. Senses* 41, 293–299. doi: 10. 1093/chemse/bjv088
- Stier, C. T. Jr., Cowden, E. A., Friesen, H. G., and Allison, M. E. (1984). Prolactin and the rat kidney: a clearance and micropuncture study. *Endocrinology* 115, 362–367. doi: 10.1210/endo-115-1-362
- Takeda, H., Chodak, G., Mutchnik, S., Nakamoto, T., and Chang, C. (1990). Immunohistochemical localization of androgen receptors with mono- and polyclonal antibodies to androgen receptor. J. Endocrinol. 126, 17–25. doi: 10.1677/joe.0.1260017
- Tinti, F., Lai, S., Noce, A., Rotondi, S., Marrone, G., Mazzaferro, S., et al. (2021). Chronic kidney disease as a systemic inflammatory syndrome: update on mechanisms involved and potential treatment. *Life* 11:419. doi: 10.3390/ life11050419
- Tsuruya, K., and Yoshida, H. (2018). Brain athrophy and cognitive impairment in chronic kidney disease. *Contrib. Nephrol.* 196, 27–36.
- Vanholder, R., Pletinck, A., Schepers, E., and Glorieux, G. (2018). Biochemical and clinical impact of organic uremic retention solutes: a comprehensive update. *Toxins* 10:33. doi: 10.3390/toxins10010033
- Vreman, H. J., Venter, C., Leegwater, J., Oliver, C., and Weiner, M. W. (1980). Taste, smell and zinc metabolism in patients with chronic renal failure. *Nephron* 26, 163–170. doi: 10.1159/000181974
- Watanabe, H., Miyamoto, Y., Honda, D., Tanaka, H., Wu, Q., Endo, M., et al. (2013). p-Cresyl sulfate causes renal tubular cell damage by inducing oxidative stress by activation of NADPH oxidase. *Kidney Int.* 83, 582–592. doi: 10.1038/ ki.2012.448
- Watanabe, K., Watanabe, T., and Nakayama, M. (2014). Cerebro-renal interactions: impact of uremic toxins on cognitive function. *Neurotoxicology* 44, 184–193. doi: 10.1016/j.neuro.2014.06.014
- Wiggins, R. C., Fuller, G., and Enna, S. J. (1984). Undernutrition and the development of brain neurotransmitter systems. *Life Sci.* 35, 2085–2094. doi: 10.1016/0024-3205(84)90507-1
- Yusuf, T., Raji, Y. R., Daniel, A., Bamidele, O. T., Fasunla, A. J., and Lasisi, O. A. (2021a). Effect of chronic kidney disease on olfactory function: a case-control study. *Ear Nose Throat J.* 2021:145561321996628. doi: 10.1177/ 0145561321996628
- Yusuf, T., Raji, Y. R., Lasisi, T. J., Daniel, A., Bamidele, O. T., Fasunla, A. J., et al. (2021b). Predictors of taste dysfunction and its severity among patients with chronic kidney disease. *Ear Nose Throat J.* 2021:1455613211019708. doi: 10.1177/01455613211019708

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's Note: All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Corona, Ordaz, Robles-Osorio, Sabath and Morales. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.