

Research Article

RCAN1 Inhibits BACE2 Turnover by Attenuating Proteasome-Mediated BACE2 Degradation

Kaixin Qiu,^{1,2,3} Shuai Wang^{1,2,3}, Xin Wang,^{2,3} Fengting Wang,^{2,3} and Yili Wu^{2,3}

¹Cheeloo College of Medicine, Shandong University, Wenhua West Road No. 44, Lixia District, Jinan, Shandong, China 250012

²Shandong Collaborative Innovation Center for Diagnosis, Treatment and Behavioral Interventions, Institute of Mental Health, Jining Medical University, Hehua Road No. 133, Taibaihu New District, Jining, Shandong, China 272067

³Shandong Key Laboratory of Behavioral Medicine, School of Mental Health, Jining Medical University, Hehua Road No. 133, Taibaihu New District, Jining, Shandong, China 272067

Correspondence should be addressed to Yili Wu; yili_wu2004@yahoo.ca

Received 4 January 2020; Accepted 22 May 2020; Published 5 June 2020

Academic Editor: Anna Chiarini

Copyright © 2020 Kaixin Qiu et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Amyloid- β protein ($A\beta$) is the main component of neuritic plaques, the pathological hallmark of Alzheimer's disease (AD). β -site APP cleaving enzyme 1 (BACE1) is a major β -secretase contributing to $A\beta$ generation. β -site APP cleaving enzyme 2 (BACE2), the homolog of BACE1, is not only a θ -secretase but also a conditional β -secretase. Previous studies showed that regulator of calcineurin 1 (RCAN1) is markedly increased by AD and promotes BACE1 expression. However, the role of RCAN1 in BACE2 regulation remains elusive. Here, we showed that RCAN1 increases BACE2 protein levels. Moreover, RCAN1 inhibits the turnover of BACE2 protein. Furthermore, RCAN1 attenuates proteasome-mediated BACE2 degradation, but not lysosome-mediated BACE2 degradation. Taken together, our work indicates that RCAN1 inhibits BACE2 turnover by attenuating proteasome-mediated BACE2 degradation. It advances our understanding of BACE2 regulation and provides a potential mechanism of BACE2 dysregulation in AD.

1. Introduction

Alzheimer's disease (AD) is the most common neurodegenerative disease in the elderly. Neuritic plaques are the major pathological hallmark of AD, while amyloid- β protein ($A\beta$) is the main component of neuritic plaques [1]. $A\beta$ is derived from amyloid- β precursor protein (APP) by sequential cleavage of β - and γ -secretase [2–4]. β -site APP cleaving enzyme 1 (BACE1) is the major β -secretase contributing to $A\beta$ generation [2]. Sun et al. reported that β -site APP cleaving enzyme 2 (BACE2), the homolog of BACE1 cleaves APP at the Phe20 site to yield a C-terminal fragment (CTF) with 80 amino acids (CTF θ or C80), which is further cleaved by γ -secretase to release a truncated $A\beta$ [3, 5, 6]. Thus, BACE2 is considered as a θ -secretase. Consistently, overexpression of BACE2 in primary neurons derived from Swedish mutant APP transgenic mice significantly reduces $A\beta$ generation [6]. Moreover, BACE2 prevents neuronal apoptosis by

cleaving a potassium channel at the surface of the neuronal plasma membrane [7].

However, growing evidence indicates that BACE2 may be implicated in the pathogenesis of AD. First, our recent study demonstrated that the binding of clusterin to the juxtamembrane helix (JH) of APP triggers BACE2-mediated β -cleavage of APP, indicating that BACE2 has comparable β -secretase activity to that of BACE1. Both BACE2 and clusterin are elevated in aged mouse brains, indicating that increased clusterin in the elderly may facilitate the β -secretase activity of BACE2, contributing to the pathogenesis of AD [8]. In addition, increased BACE2 expression and activity are detected in the neurons of AD brains [9]. Genetic data shows that BACE2 is associated with AD risk [10]. For example, the BACE2 haplotype associates with AD, while SNPs in BACE2 (e.g., rs2252576, rs2837990, and rs7281733) predispose to early onset of AD in Down syndrome. Recently, the association between SNPs in BACE2 and AD was reported in APOE

ε4 noncarriers, which may be mediated by altered BACE2 expression-mediated Aβ generation and clearance [11].

Regulator of calcineurin 1 (RCAN1), also called Down syndrome critical region 1 (DSCR1), is highly expressed in the human brain and heart [12]. By alternative splicing and alternative promoter usage, two major transcripts are generated, transcript 1 and transcript 4, *RCAN1.1* and *RCAN1.4* [13–15]. Transcript 1 encodes two isoforms by alternative usage of two translational start sites, the long isoform of RCAN1.1 (RCAN1.1L) and the short isoform of RCAN1.1 (RCAN1.1S), which consist of 252 and 197 amino acids, respectively [16]. RCAN1.1L is the major isoform of RCAN1.1, which is upregulated in AD. However, RCAN1.1S is hard to be detected. Transcript 4 encodes RCAN1.4 with 197 amino acids. RCAN1.1L (hereinafter referred to as RCAN1) is highly expressed in the brains and is upregulated in AD brains. Increased RCAN1 plays a pivotal role in AD pathogenesis [15] including neuronal loss [17, 18], tau hyperphosphorylation [19, 20], and synaptic dysfunction [21, 22]. Previous study showed that RCAN1 significantly increases BACE1 expression, while BACE1 and BACE2 share an approximate 75% similarity of amino acids. However, the role of RCAN1 in BACE2 regulation remains elusive. In this study, we reported that RCAN1 increases BACE2 protein levels. Moreover, RCAN1 inhibits the turnover of BACE2 protein. Furthermore, RCAN1 attenuates proteasome-mediated BACE2 degradation, but not lysosome-mediated BACE2 degradation. Taken together, our work indicates that RCAN1 inhibits BACE2 turnover by attenuating proteasome-mediated BACE2 degradation, leading to the upregulation of BACE2. It advances our understanding of BACE2 regulation and provides a potential mechanism of BACE2 dysregulation in AD.

2. Materials and Methods

2.1. Cell Culture and Transfection. Human embryonic kidney HEK293 cells and HRNLM cells obtained from Dr. Weihong Song's lab were cultured in high-glucose DMEM containing 10% fetal bovine serum and 1% penicillin-streptomycin. HRNLM cells are derived from HEK293 cells, which stably overexpress RCAN1 with a C-terminal myc tag. All cells were maintained at 37°C with 5% CO₂ in an incubator as described previously [16, 23, 24]. pBACE2-mycHis refers to pZ-BACE2mycHis in this study, which is constructed previously [25]. Transient transfection was performed by using the polyetherimide (PEI) method as described previously [26, 27]. Briefly, HEK293 and HRNLM cells were seeded 24h prior to transfection. The regular culture medium was replaced with high-glucose DMEM without serum 1h prior to transfection. 6h after transfection, the medium was replaced with regular culture medium.

2.2. Pharmacological Treatments. HEK293 cells and HRNLM cells were transiently transfected with pBACE2-mycHis. 24h after transfection, the cells were equally seeded into 6cm culture dishes. 48h after transfection, the cells were treated with different drugs, respectively. To measure the half-life of BACE2, 100 μg/mL cycloheximide (CHX) was used to

treat cells as described previously [16, 23, 24]. After treatment, the cells were harvested at 0, 2, 4, 8, and 12h time points, respectively. The lysosomal inhibitor NH₄Cl was applied to determine the involvement of the lysosome pathway in BACE2 degradation, while the proteasomal inhibitor N-carbobenzoxy-L-leucyl-L-leucyl-L-leucinal (MG-132) was applied to determine the involvement of proteasome pathway in BACE2 degradation, respectively [16, 23, 28–30]. MG-132 and NH₄Cl were purchased from Sigma.

2.3. Western Blotting. Cells were lysed with RIPA-Doc buffer (Tris-HCl, 50 mM; NaCl, 150 mM; Triton X-100, 1%; deoxycholate, 1%; and SDS, 0.1%; supplemented with 1/100 protease inhibitors). Cell lysates were separated by 10% Tris-glycine SDS-PAGE gels and transferred to PVDF membranes. The membranes were blocked in 5% nonfat milk for 1h, then incubated overnight at 4°C with anti-myc (9E10), anti-GFP, anti-RCAN1, and anti-β-actin (AC-15) antibodies, respectively [23]. The membranes were washed in TBST with 0.1% Tween-20 and incubated with HRP-labeled goat anti-mouse or HRP-labeled goat anti-rabbit antibodies at room temperature for 1h. Anti-myc antibody 9E10 was obtained from Abcam. β-Actin antibody AC-15, HRP-labeled goat anti-rabbit antibody, and HRP-labeled goat anti-mouse antibody were obtained from ZSGB-BIO. Anti-GFP antibody was purchased from Proteintech Group. Anti-RCAN1 antibody was purchased from Sigma. The image was obtained by using FluorChem R imaging system.

2.4. Statistical Analysis. The protein expression was quantified by using ImageJ. Student's *t* test or two-way ANOVA was used for data analysis with three or more independent experiments. *P* < 0.05 was considered as a significant difference.

3. Results

3.1. RCAN1 Increases BACE2 Expression. To explore the effect of RCAN1 on BACE2 regulation, HEK293 cells and HRNLM cells (i.e., HEK293 cells stably overexpressing myc-tagged RCAN1) were cotransfected with plasmids pEGFP and pBACE2-mycHis at the ratio of 1:5. Exogenous GFP was used as a control for transfection efficiency in both cell lines. We found that the level of BACE2 protein was significantly higher in HRNLM cells than in HEK293 cells, while the levels of GFP were similar in HEK293 cells and HRNLM cells (Figure 1(a)). After normalization to the level of GFP, BACE2 was significantly increased to 4.75 ± 0.60 -fold in HRNLM cells comparing with that in HEK293 cells (Figure 1(b)). It indicated that RCAN1 significantly upregulated BACE2 expression.

3.2. RCAN1 Inhibits BACE2 Turnover. Protein degradation plays an important role in protein homeostasis. To determine whether RCAN1 affects BACE2 turnover rate contributing to its upregulation, the degradation rate of BACE2 was examined in HEK293 and HRNLM cells. HEK293 cells and HRNLM cells were transfected with the plasmid pBACE2-mycHis. 24 hours after transfection, cells were divided equally into five dishes. 48 hours after transfection, the cells were treated with 100 μg/mL cycloheximide (CHX), a potent

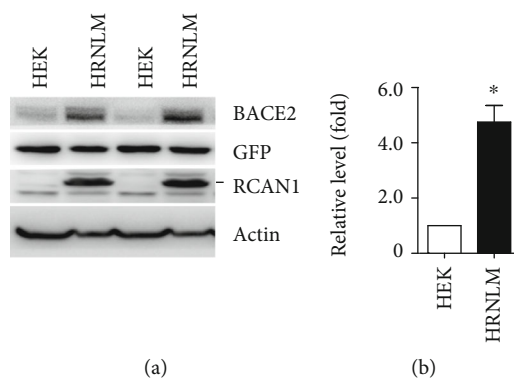


FIGURE 1: RCAN1 increases BACE2 expression. (a) HEK293 cells and HRNLM cells were cotransfected with plasmids pEGFP and pBACE2-mycHis. Cell lysates were resolved by 10% SDS-PAGE. BACE2 expression was detected by using 9E10 antibody. GFP was detected by GFP antibody. Anti-RCAN1 antibody was used to detect RCAN1. β -Actin was detected by AC-15 antibody. (b) Quantification of BACE2 levels. GFP levels were used to normalize the transfection efficiency. Values are mean \pm SEM; $n \geq 3$, * $P < 0.05$ by Student's t test.

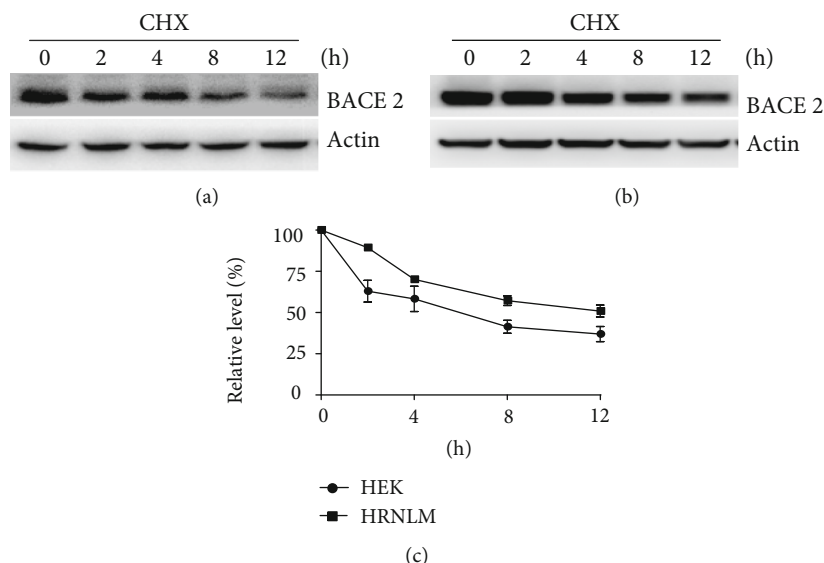


FIGURE 2: RCAN1 inhibits BACE2 turnover. (a) HEK293 cells and (b) HRNLM cells were transfected with pBACE2-mycHis. 48 hours after transfection, cells were treated with 100 μ g/mL CHX for 0, 2, 4, 8, or 12 h. Cell lysates were resolved by 10% SDS-PAGE. 9E10 antibody was used to detect myc-tagged BACE2 protein. β -Actin served as an internal control. (c) Quantification of BACE2 levels. Values represent mean \pm SEM.

inhibitor of protein synthesis, to block BACE2 synthesis [16, 23, 29, 31]. The cells were harvested at 0, 2, 4, 8, and 12h after CHX treatment, and western blot analysis was performed to determine the level of remaining BACE2 protein relative to the BACE2 level at 0h time point. BACE2 level was decreased to $63 \pm 6.6\%$, $58 \pm 7.6\%$, $45 \pm 6.7\%$, and $37 \pm 4.6\%$ at 2, 4, 8, and 12h time points in HEK293 cells (Figures 2(a) and 2(c)). It indicated that the half-life of BACE2 was approximately 6 h in HEK293 cells. In HRNLM cells, BACE2 level was decreased to $89 \pm 1.7\%$, $70 \pm 1.6\%$, $55 \pm 4.1\%$, and $51 \pm 3.6\%$ at 2, 4, 8, and 12h time points, respectively, indicating that the half-life of BACE2 was approximately 12 h in HRNLM cells (Figures 2(b) and 2(c)). Our data indicated that RCAN1 inhibited BACE2 turnover by prolonging its half-life.

3.3. RCAN1 Has No Effect on Lysosome-Mediated BACE2 Degradation. As the previous study showed that BACE2 degradation is mediated by the lysosome pathway, the effect of RCAN1 on lysosome-mediated BACE2 degradation was examined. HEK293 cells and HRNLM cells were transfected with pBACE2-mycHis, respectively. 24 hours after transfection, cells were divided equally into four dishes. 48 hours after transfection, the lysosomal inhibitor NH_4Cl was applied to the two cell lines. The cells were treated with 0, 10, 25, or 50 mM NH_4Cl for 24h. Western blot analysis showed that NH_4Cl treatment significantly increased BACE2 levels to 1.64 ± 0.23 -, 4.18 ± 0.33 -, and 5.38 ± 0.27 -fold, respectively, compared with those of the control in HEK293 cells, $P < 0.05$ (Figures 3(a) and 3(c)). In HRNLM cells, the level of BACE2 protein was increased to 1.52 ± 0.19 -, 4.25 ± 0.22 -,

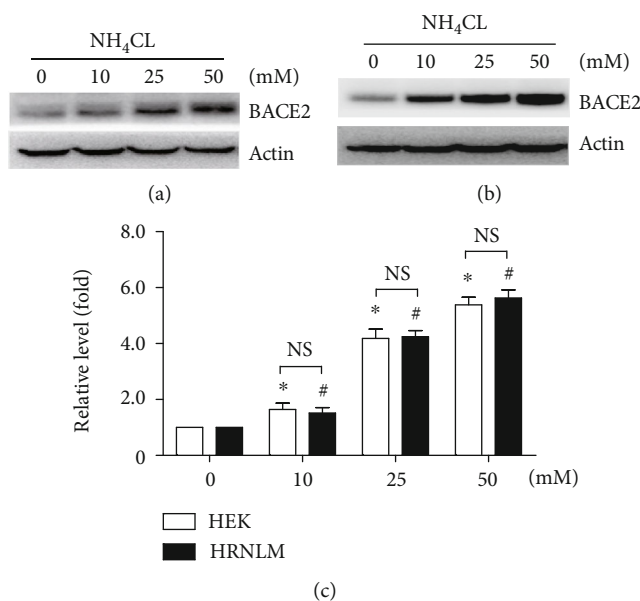


FIGURE 3: RCAN1 has no effect on lysosome-mediated BACE2 degradation. (a) HEK293 cells and (b) HRNLM cells were transfected with pBACE2-mycHis. 48 hours after transfection, cells were treated with 0, 10, 25, or 50 mM lysosomal inhibitor NH₄Cl for 24 h. Cell lysates were resolved by 10% SDS-PAGE. 9E10 antibody was used to detect myc-tagged BACE2 protein. β -Actin served as an internal control. (c) Quantification of BACE2 levels. Values represent mean \pm SEM; $n \geq 3$, * $P < 0.05$ by two-way ANOVA.

and 5.63 ± 0.28 -fold, respectively, $P < 0.05$, when the cells were treated with 10, 25, or 50 mM NH₄Cl (Figures 3(b) and 3(c)). However, no significant difference of relative BACE2 changes was detected between HEK293 cells and HRNLM cells when the same dose of NH₄Cl was applied (Figures 3(a)–3(c)). It indicated that the increase of RCAN1 has no effect on lysosome-mediated BACE2 degradation.

3.4. RCAN1 Attenuates Proteasome-Mediated BACE2 Degradation. As the previous study showed that BACE2 is also degraded via the proteasome pathway, the effect of RCAN1 on proteasome-mediated BACE2 degradation was further investigated [32]. HEK293 cells and HRNLM cells were transfected with pBACE2-mycHis, respectively. 24 hours after transfection, cells were divided equally into four dishes. 48 hours after transfection, the proteasomal inhibitor MG-132 was applied to two cell lines. The cells were treated with 0, 10, 15, and 20 μ M MG-132 for 12 h, respectively. MG-132 treatment significantly increased BACE2 levels to 4.52 ± 0.29 -, 4.30 ± 0.22 -, and 4.42 ± 0.23 -fold, respectively, compared with those of the control in HEK293 cells, $P < 0.05$ (Figures 4(a) and 4(c)). In HRNLM cells, the level of BACE2 protein was increased to 1.97 ± 0.23 -, 1.97 ± 0.12 -, and 2.15 ± 0.15 -fold, respectively, $P < 0.05$, when the cells were treated with 10, 15, and 20 μ M MG-132 (Figures 4(b) and 4(c)). Importantly, the increase of BACE2 in HEK293 cells was much higher than that in HRNLM cells when same dose of MG-132 was applied, 4.52 ± 0.29 - vs. 1.97 ± 0.23 -fold at the dose of 10 μ M, 4.30 ± 0.22 - vs. 1.97 ± 0.12 -fold at the dose of 15 μ M, and 4.42 ± 0.23 - vs. 2.15 ± 0.15 -fold at the dose of 20 μ M, $P < 0.05$ (Figures 4(a)–4(c)). It indicated that the increase of RCAN1 attenuates proteasome-mediated BACE2 degradation.

4. Discussion

The dysregulation of BACE2 is observed in AD. However, the underlying mechanisms remain elusive. Our data showed that RCAN1 increases BACE2, while RCAN1 upregulation in AD is mediated by different mechanisms. It indicates that multiple stresses and risk factors of AD may implicate in the increase of BACE2 in AD, which is mediated by RCAN1 upregulation. For example, the stress hormone glucocorticoids are increased in AD, which are stimulated to be released by various stressful risk factors of AD [33]. The glucocorticoid significantly increases RCAN1 transcription mediated by a glucocorticoid response element in the RCAN1 promoter region [24, 34, 35], indicating that stress hormone may contribute to the BACE2 upregulation. In addition, *ApoE* allele4, a known risk factor of AD, markedly increases RCAN1 expression, indicating that it may also promote BACE2 expression [36]. NF- κ B promotes RCAN1 transcription, indicating that inflammation may play a key role in the upregulation of RCAN1 and subsequent BACE2 upregulation in AD [37]. The putative transcription factor HIF-1 binding site within the RCAN1 promoter region may mediate the alteration of RCAN1 expression under hypoxia, ischemia, and stroke conditions, promoting AD development. It is also supported by the fact that hypoxia, ischemia, and stroke are also risk factors of AD. Moreover, chronic hyperglycemia, the characteristic of diabetes, increases RCAN1 expression in pancreatic β cells, suggesting that chronic hyperglycemia may also contribute to RCAN1 elevation in the brain [38]. As both stroke and diabetes are age-associated diseases, their role in RCAN1 regulation may also be affected by the aging process [39]. The above evidence suggests that stroke and diabetes may promote the upregulation of BACE2

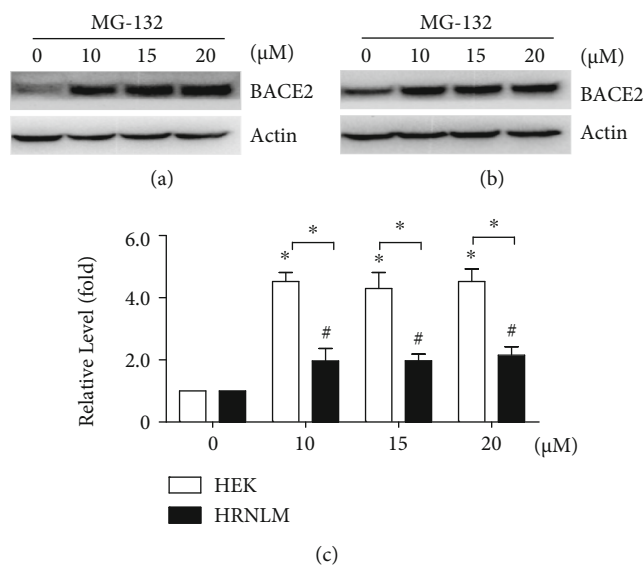


FIGURE 4: RCAN1 attenuates proteasome-mediated BACE2 degradation. (a) HEK293 cells and (b) HRNLM cells were transfected with pBACE2-mycHis. 48 hours after transfection, cells were treated with 0, 10, 15, or 20 μM proteasomal inhibitor MG-132 for 12 h. Cell lysates were resolved by 10% SDS-PAGE. 9E10 antibody was used to detect myc-tagged BACE2 protein. β -Actin served as an internal control. (c) Quantification of BACE2 levels. Values represent mean \pm SEM; $n \geq 3$, * $P < 0.05$ by two-way ANOVA.

mediated by increased RCAN1 expression. In fact, inhibition of BACE2 counteracts IAPP-induced β cell loss and functional defects [40, 41]. Furthermore, calcineurin and GSK3 β activities are significantly altered in AD brains, which are involved in regulating RCAN1 degradation rate by modulating its phosphorylation status [42–48]. It indicates that the dysregulation of calcineurin and GSK3 β may be implicated in the increase of BACE2 in AD, which is mediated by RCAN1 upregulation, although further investigation is needed.

Our data showed that RCAN1-induced increase of BACE2 expression is caused by attenuating proteasome-mediated BACE2 degradation. Although the mechanism of RCAN1 attenuating proteasome-mediated BACE2 degradation is not clear, the following mechanisms may be implicated in RCAN1 attenuating the proteasome-mediated BACE2 degradation. Previous studies showed that RCAN1.1S could stabilize oligosaccharyltransferase (OST) by interacting with the OST component ribophorin I (RPN I) [49]. It is possible that RCAN1 may stabilize BACE2 by directly interacting with BACE2. In addition, RCAN1.1S promotes the N-glycosylation of BACE1 and BACE2, a type of protein posttranslational modification, which is associated with their upregulation [49]. It is possible that RCAN1 may also facilitate BACE2 N-glycosylation contributing to the increase of BACE2 expression. Moreover, RCAN1 not only regulates the activity of various transcriptional factors, e.g., NFAT, but also shows RNA binding activity, which may be implicated in the increase of BACE2 by regulating the expression of OST components or ubiquitin-proteasome system- (UPS-) associated molecules.

5. Conclusions

In conclusion, our work indicates that RCAN1 inhibits BACE2 turnover by attenuating proteasome-mediated BACE2 degradation, leading to the upregulation of BACE2.

It advances our understanding of BACE2 regulation and provides a potential mechanism of BACE2 dysregulation in AD. Moreover, it provides a novel insight that RCAN1 dysregulation promotes the pathogenesis of AD through BACE2 7in addition to other known mechanisms. It suggests that RCAN1 and BACE2 may be potential targets for AD treatment.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare no conflict of interest.

Authors' Contributions

Kaixin Qiu and Shuai Wang contributed equally to this work.

Acknowledgments

This research was funded by the National Natural Science Foundation of China (NSFC), Grant 81771147 to YW, and the Natural Science Foundation of Shandong Province, Grant ZR2019PC038 to SW.

References

- [1] G. G. Glenner and C. W. Wong, "Alzheimer's disease and Down's syndrome: sharing of a unique cerebrovascular amyloid fibril protein," *Biochemical and Biophysical Research Communications*, vol. 122, no. 3, pp. 1131–1135, 1984.
- [2] K. Bromley-Brits and W. Song, "The Role of TMP21 in Trafficking and Amyloid- β Precursor Protein (APP) Processing

- in Alzheimer's Disease," *Current Alzheimer Research*, vol. 9, no. 4, pp. 411–424, 2012.
- [3] K. Qiu, X. Zhang, S. Wang et al., "TMP21 in Alzheimer's disease: molecular mechanisms and a potential target," *Frontiers in Cellular Neuroscience*, vol. 13, 2019.
 - [4] X. Wang, X. Zhou, G. Li, Y. Zhang, Y. Wu, and W. Song, "Modifications and trafficking of APP in the pathogenesis of Alzheimer's disease," *Frontiers in Molecular Neuroscience*, vol. 10, p. 294, 2017.
 - [5] F. Acquati, M. Accarino, C. Nucci et al., "The gene encoding DRAP (BACE2), a glycosylated transmembrane protein of the aspartic protease family, maps to the down critical region," *FEBS Letters*, vol. 468, no. 1, pp. 59–64, 2000.
 - [6] X. Sun, G. He, and W. Song, "BACE2, as a novel APP theta-secretase, is not responsible for the pathogenesis of Alzheimer's disease in Down syndrome," *FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology*, vol. 20, no. 9, pp. 1369–1376, 2006.
 - [7] F. Liu, Y. Zhang, Z. Liang et al., "Cleavage of potassium channel Kv2.1 by BACE2 reduces neuronal apoptosis," *Molecular Psychiatry*, vol. 23, no. 7, pp. 1542–1554, 2018.
 - [8] Z. Wang, Q. Xu, F. Cai, X. Liu, Y. Wu, and W. Song, "BACE2, a conditional β -secretase, contributes to Alzheimer's disease pathogenesis," *JCI insight*, vol. 4, no. 1, 2019.
 - [9] C. J. Holler, R. L. Webb, A. L. Laux et al., "BACE2 expression increases in human neurodegenerative disease," *The American Journal of Pathology*, vol. 180, no. 1, pp. 337–350, 2012.
 - [10] L. Mylykangas, F. Wavrant-de Vrièze, T. Polvikoski et al., "Chromosome 21 BACE2 haplotype associates with Alzheimer's disease: A two-stage study," *Journal of the Neurological Sciences*, vol. 236, no. 1-2, pp. 17–24, 2005.
 - [11] M. Huentelman, M. De Both, W. Jepsen et al., "Common BACE2 polymorphisms are associated with altered risk for Alzheimer's disease and CSF amyloid biomarkers in APOE ϵ 4 non-carriers," *Scientific Reports*, vol. 9, no. 1, p. 9640, 2019.
 - [12] J. J. Fuentes, M. A. Pritchard, A. M. Planas, A. Bosch, I. Ferrer, and X. Estivill, "A new human gene from the Down syndrome critical region encodes a proline-rich protein highly expressed in fetal brain and heart," *Human Molecular Genetics*, vol. 4, no. 10, pp. 1935–1944, 1995.
 - [13] K. J. A. Davies, G. Ermak, B. A. Rothermel et al., "Renaming the DSCR1 / Adapt78 gene family as RCAN: regulators of calcineurin," *FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology*, vol. 21, no. 12, pp. 3023–3028, 2007.
 - [14] J. J. Fuentes, M. A. Pritchard, and X. Estivill, "Genomic organization, alternative splicing, and expression patterns of the DSCR1 (Down syndrome candidate region 1) gene," *Genomics*, vol. 44, no. 3, pp. 358–361, 1997.
 - [15] X. Sun, Y. Wu, B. Herculano, and W. Song, "RCAN1 overexpression exacerbates calcium overloading-induced neuronal apoptosis," *PLoS One*, vol. 9, no. 4, article e95471, 2014.
 - [16] Y. Wu and W. Song, "Regulation of RCAN1 translation and its role in oxidative stress-induced apoptosis," *FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology*, vol. 27, no. 1, pp. 208–221, 2012.
 - [17] M. Dierssen, G. Arque, J. McDonald et al., "Behavioral characterization of a mouse model overexpressing DSCR1/RCAN1," *PLoS One*, vol. 6, no. 2, article e17010, 2011.
 - [18] K. R. Martin, A. Corlett, D. Dubach et al., "Over-expression of RCAN1 causes Down syndrome-like hippocampal deficits that alter learning and memory," *Human Molecular Genetics*, vol. 21, no. 13, pp. 3025–3041, 2012.
 - [19] G. Ermak, M. A. Pritchard, S. Dronjak, B. Niu, and K. J. A. Davies, "Do RCAN1 proteins link chronic stress with neurodegeneration?," *FASEB Journal : Official Publication of the Federation of American Societies for Experimental Biology*, vol. 25, no. 10, pp. 3306–3311, 2011.
 - [20] D. Poppek, S. Keck, G. Ermak et al., "Phosphorylation inhibits turnover of the tau protein by the proteasome: influence of RCAN1 and oxidative stress," *Biochemical Journal*, vol. 400, no. 3, pp. 511–520, 2006.
 - [21] K. T. Chang and K. T. Min, "Upregulation of three Drosophila homologs of human chromosome 21 genes alters synaptic function: implications for Down syndrome," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 106, no. 40, pp. 17117–17122, 2009.
 - [22] T. Sun, X. S. Wu, J. Xu et al., "The role of calcium/calmodulin-activated calcineurin in rapid and slow endocytosis at central synapses," *The Journal of neuroscience : the official journal of the Society for Neuroscience*, vol. 30, no. 35, pp. 11838–11847, 2010.
 - [23] X. Liu, Z. Wang, Y. Wu, J. Wang, and W. Song, "BACE2 degradation mediated by the macroautophagy-lysosome pathway," *The European Journal of Neuroscience*, vol. 37, no. 12, pp. 1970–1977, 2013.
 - [24] X. Sun, Y. Wu, B. Chen et al., "Regulator of calcineurin 1 (RCAN1) facilitates neuronal apoptosis through caspase-3 activation," *The Journal of Biological Chemistry*, vol. 286, no. 11, pp. 9049–9062, 2011.
 - [25] X. Sun, Y. Wang, H. Qing et al., "Distinct transcriptional regulation and function of the human BACE2 and BACE1 genes," *The FASEB Journal*, vol. 19, no. 7, pp. 739–749, 2005.
 - [26] G. Backliwal, M. Hildinger, S. Chenuet, M. Dejesus, and F. M. Wurm, "Coexpression of acidic fibroblast growth factor enhances specific productivity and antibody titers in transiently transfected HEK293 cells," *New Biotechnology*, vol. 25, no. 2-3, pp. 162–166, 2008.
 - [27] G. Backliwal, M. Hildinger, S. Chenuet, S. Wulhfard, M. De Jesus, and F. M. Wurm, "Rational vector design and multi-pathway modulation of HEK 293E cells yield recombinant antibody titers exceeding 1 g/l by transient transfection under serum-free conditions," *Nucleic Acids Research*, vol. 36, no. 15, article e96, 2008.
 - [28] C. Li, X. Wang, X. Li et al., "Proteasome inhibition activates autophagy-lysosome pathway associated with TFEB dephosphorylation and nuclear translocation," *Frontiers in Cell and Developmental Biology*, vol. 7, 2019.
 - [29] S. Liu, K. Bromley-Brits, K. Xia, J. Mittelholtz, R. Wang, and W. Song, "TMP21 degradation is mediated by the ubiquitin-proteasome pathway," *The European Journal of Neuroscience*, vol. 28, no. 10, pp. 1980–1988, 2008.
 - [30] H. Qing, W. Zhou, M. A. Christensen, X. Sun, Y. Tong, and W. Song, "Degradation of BACE by the ubiquitin-proteasome pathway," *The FASEB Journal*, vol. 18, no. 13, pp. 1571–1573, 2004.
 - [31] A. J. Whiffen, "The production, assay, and antibiotic activity of actidione, an antibiotic from *Streptomyces griseus*," *Journal of Bacteriology*, vol. 56, no. 3, pp. 283–291, 1948.

- [32] K. Qiu, W. Liang, S. Wang et al., "BACE2 degradation is mediated by both the proteasome and lysosome pathways," *BMC molecular and cell biology*, vol. 21, no. 1, p. 13, 2020.
- [33] F. Masugi, T. Ogihara, K. Sakaguchi et al., "High plasma levels of cortisol in patients with senile dementia of the Alzheimer's type," *Methods and Findings in Experimental and Clinical Pharmacology*, vol. 11, no. 11, pp. 707–710, 1989.
- [34] N. L. Yoshida, T. Miyashita, U. Mami et al., "Analysis of gene expression patterns during glucocorticoid-induced apoptosis using oligonucleotide arrays," *Biochemical and Biophysical Research Communications*, vol. 293, no. 4, pp. 1254–1261, 2002.
- [35] M. U. L. Shen, T. Oshida, J. Miyauchi, M. Yamada, and T. Miyashita, "Identification of novel direct transcriptional targets of glucocorticoid receptor," *Leukemia*, vol. 18, no. 11, pp. 1850–1856, 2004.
- [36] M. C. Badia, A. Lloret, E. Giraldo et al., "Lymphocytes from young healthy persons carrying the ApoE4 allele overexpress stress-related proteins involved in the pathophysiology of Alzheimer's disease," *Journal of Alzheimer's Disease*, vol. 33, no. 1, pp. 77–83, 2012.
- [37] L. Zheng, H. Liu, P. Wang, W. Song, and X. Sun, "Regulator of calcineurin 1 gene transcription is regulated by nuclear factor-kappaB," *Current Alzheimer Research*, vol. 11, no. 2, pp. 156–164, 2014.
- [38] H. Peiris, R. Raghupathi, C. F. Jessup et al., "Increased expression of the glucose-responsive gene, RCAN1, causes hypoinsulinemia, β -cell dysfunction, and diabetes," *Endocrinology*, vol. 153, no. 11, pp. 5212–5221, 2012.
- [39] Y. Tian, S. Wang, F. Jiao, Q. Kong, C. Liu, and Y. Wu, "Telomere length: a potential biomarker for the risk and prognosis of stroke," *Frontiers in Neurology*, vol. 10, p. 624, 2019.
- [40] G. Alcarraz-Vizan, P. Casini, L. Cadavez et al., "Inhibition of BACE2 counteracts hIAPP-induced insulin secretory defects in pancreatic β -cells," *The FASEB Journal*, vol. 29, no. 1, pp. 95–104, 2014.
- [41] G. Finzi, F. Franzi, C. Placidi et al., "BACE2 is Stored in Secretory Granules of Mouse and Rat Pancreatic β Cells," *Ultrastructural Pathology*, vol. 32, no. 6, pp. 246–251, 2009.
- [42] S. Abbasi, J. D. Lee, B. Su et al., "Protein kinase-mediated regulation of calcineurin through the phosphorylation of modulatory calcineurin-interacting protein 1," *The Journal of Biological Chemistry*, vol. 281, no. 12, pp. 7717–7726, 2006.
- [43] J. J. Fuentes, L. Genesca, T. J. Kingsbury et al., "DSCR1, overexpressed in Down syndrome, is an inhibitor of calcineurin-mediated signaling pathways," *Human Molecular Genetics*, vol. 9, no. 11, pp. 1681–1690, 2000.
- [44] E. J. Lee, S. R. Seo, J. W. Um, J. Park, Y. Oh, and K. C. Chung, "NF-kappaB-inducing kinase phosphorylates and blocks the degradation of Down syndrome candidate region 1," *The Journal of Biological Chemistry*, vol. 283, no. 6, pp. 3392–3400, 2008.
- [45] Q. Liu, J. C. Busby, and J. D. Molkentin, "Interaction between TAK1-TAB1-TAB2 and RCAN1-calcineurin defines a signaling nodal control point," *Nature Cell Biology*, vol. 11, no. 2, pp. 154–161, 2009.
- [46] M. A. Mines, E. Beurel, and R. S. Jope, "Regulation of Cell Survival Mechanisms in Alzheimer's Disease by Glycogen Synthase Kinase-3," *International Journal of Alzheimer's Disease*, vol. 2011, pp. 1–11, 2011.
- [47] J. J. Pei, E. Braak, H. Braak et al., "Distribution of active glycogen synthase kinase 3beta (GSK-3beta) in brains staged for Alzheimer disease neurofibrillary changes," *Journal of Neuro-pathology and Experimental Neurology*, vol. 58, no. 9, pp. 1010–1019, 1999.
- [48] L. C. Reese and G. Taghialatela, "A role for calcineurin in Alzheimer's disease," *Current Neuropharmacology*, vol. 9, no. 4, pp. 685–692, 2011.
- [49] T. Wang, H. Liu, Y. Wang, C. Liu, and X. Sun, "RCAN1 increases A β generation by promoting N-glycosylation via oligosaccharyltransferase," *Current Alzheimer Research*, vol. 11, no. 4, pp. 332–339, 2014.