Protective effects of AMP-activated protein kinase in the cardiovascular system

Qiang Xu, Liang-Yi Si*

Department of Geriatrics, Southwest Hospital, Third Military Medical University, Chongqing, China

Received: April 22, 2010; Accepted: August 25, 2010

- Introduction
- AMPK structure and function
- Role of AMPK in the heart
- Role of AMPK in endothelial cells
- Role of AMPK in vascular smooth muscle cells

- Atherosclerosis
 Hypertension
 Ischaemic stroke
- Conclusion and perspectives

· Role of AMPK in vascular angiogenesis

Role of AMPK in cardiovascular diseases

Abstract

Cardiovascular diseases remain the leading cause of mortality worldwide. Recent studies of AMP-activated protein kinase (AMPK), a highly conserved sensor of cellular energy status, suggest that there might be therapeutic value in targeting the AMPK signaling pathway. AMPK is found in most mammalian tissues, including those of the cardiovascular system. As cardiovascular diseases are typically associated with blood flow occlusion and blood occlusion may induce rapid energy deficit, AMPK activation may occur during the early phase upon nutrient deprivation in cardiovascular organs. Therefore, investigation of AMPK in cardiovascular organs may help us to understand the pathophysiology of defence mechanisms in these organs. Recent studies have provided proof of concept for the idea that AMPK is protective in heart as well as in vascular endothelial and smooth muscle cells. Moreover, dysfunction of the AMPK signalling pathway is involved in the genesis and development of various cardiovascular diseases, including atherosclerosis, hypertension and stroke. The roles of AMPK in the cardiovascular system, as they are currently understood, will be presented in this review. The interaction between AMPK and other cardiovascular signalling pathways such as nitric oxide signalling is also discussed.

Keywords: AMPK • cardiovascular system • therapeutic target • energy stress • protection

Introduction

AMP-activated protein kinase (AMPK) is a phylogenetically conserved serine/threonine protein kinase [1]; AMPK is a member of the SNF1/AMPK protein kinase family and is found in all eucaryotes [2, 3]. It is a critical metabolic controller and stress sensor that is activated by nutrient deprivation, hypoxia/ischaemia, starvation, exercise and some therapeutic agents [1]. The name AMPK was first suggested in 1988 when Munday *et al.* showed that the enzyme formerly known as acetyl-CoA carboxylase (ACC) kinase-3 is the primary kinase responsible for regulation of the ACC V_{max} [4]. Later, the name AMPK was used in 1989 by Carling *et al.* [5], who demonstrated that AMPK is the major 3-hydroxy-3-methyl-glutaryl-CoA (HMG-CoA) reductase kinase in rat liver. Activated AMPK not only stimulates a number of energy-producing metabolic pathways, but also inhibits energy-consuming pathways to

*Correspondence to: Liang-Yi SI, M.D.,

Department of Geriatrics, Southwest Hospital,

Third Military Medical University 29 Gaotanyan Main Street, Shapingba District,

doi:10.1111/j.1582-4934.2010.01179.x

reserve energy for cell survival. Thus, AMPK is a signal switch that monitors and regulates systemic and cellular energy status [6].

As the name suggests, AMPK activity increases in response to increases in the intracellular level of adenosine monophosphate (AMP), which generally occurs as a product of hydrolysis of adenosine triphosphate (ATP) in metabolic processes. In response to a reduction in energy charge (decrease in the ATP/AMP ratio), AMPK shuts down anabolic pathways such as fatty acid, triglyceride and cholesterol synthesis, as well as transcriptional processes that consume ATP, and switches on catabolic pathways that generate ATP, including fatty acid oxidation, glycolysis and protein decomposition [7]. Increases in the AMP/ATP ratio activate AMPK by a number of mechanisms, including direct allosteric activation, and render it more easily phosphorylated by an AMP-dependent

Chongqing 400038, PR China. Fax: +86-23-6875-4151 Tel.: +86-23-6875-4151 E-mail: siliangyi@yahoo.com



Fig. 1 Proposed model illustrating protein factors upstream and downstream of AMPK (refer to text for additional detail). Upper arrows show kinase-mediated activation of AMPK via Ca²⁺/calmodulin-dependent protein kinase β (CaMKK β), LKB1 and TGF β -activated kinase (TAK1). Lower arrows indicate targets of AMPK, including endothelial nitric oxide synthase (eNOS), acetyl-CoA carboxylase (ACC), glucose transporter type 4 (GLUT4) and 6-phosphofructo-1-kinase (PFK1).

AMPK kinase (AMPKK) on several sites of the α -subunit, including at Thr¹⁷² [8, 9]. Thus, the ubiquitous expression of AMPK and its involvement in the uptake, mobilization and utilization of glucose, fatty acids and protein, stress the general importance of this pathway. Apart from these diverse functions in metabolism, AMPK is, in fact, a key regulator involved in many biological functions of the cardiovascular system. In this review, we specifically focus on the roles of AMPK in cardiovascular tissues and data suggesting that dysfunction in AMPK signalling contributes to the genesis and progression of cardiovascular diseases.

AMPK structure and function

AMPK is a heterotrimeric complex consisting of a catalytic α subunit and regulatory β and γ subunits [1]. The α and β subunits each exist as two isoforms: $\alpha 1$ and $\alpha 2$ and $\beta 1$ and $\beta 2$. The γ subunit has three known isoforms: $\gamma 1$, $\gamma 2$ and $\gamma 3$ [10, 11]. Although the C-terminal region of the α subunit is required for association with the β/γ subunits in both $\alpha 1$ and $\alpha 2$ isoforms, the subunits have distinct expression patterns. For example, in peripheral metabolic tissues, AMPK complexes containing the $\alpha 1$ catalytic subunit are mainly expressed in adipose tissue [12-14], whereas those containing the $\alpha 2$ catalytic subunit pre-dominate in skeletal and heart muscle [15–17]. In central nervous system tissues, the α 1 catalytic subunit is almost absent [18]. The serine/threonine kinase activity of AMPK is located at the α subunit and is characterized by the presence of a threonine residue (Thr172) in a loop that must be phosphorylated for activation [19–21]. The AMPK α catalytic subunit can also be phosphorylated at other sites: Thr258 and Ser485 for $\alpha 1$ and Ser491 for α 2. However, the upstream kinase and the biological significance of these phosphorylation sites are currently unclear [22]. The ß subunits are modified by both myristoylation and multi-site phosphorylation, including Ser24/25, Ser96, Ser101, Ser108 and Ser182. These modifications appear to be required for the activation of AMPK and its subcellular localization [22, 23]. The y subunits are responsible for the binding of AMP. Mutations at the AMP/ATP binding sites (CBS or Bateman domains) lead to reduction of AMPK activity and cause glycogen accumulation [24, 25].

The phosphorylation of the sites referred to above is regulated by three upstream AMPK kinases (AMPKKs): LKB1, Ca²⁺/calmodulindependent protein kinase B (CaMKKB) and transforming growth factor β activated kinase 1 (TAK1) shown schematically in Fig. 1. LKB1 is an upstream AMPKK that regulates AMPK and AMP-related kinases [20] and is believed to be the most potent positive regulator of AMPK [26, 27]. A cytosolic protein complex containing LKB1, putative kinase STRAD and the MO25 scaffold protein can interact with, and directly phosphorylate, AMPK [20]. When the AMP/ATP ratio increases, the phosphorylation of AMPK by LKB1 is enhanced: the LKB1 pathway is thought to be the major route by which AMPK is activated in response to energy demand and to maintain metabolic homeostasis. In addition, an increasing number of studies indicate that LKB1 is the principal provider of AMPKK activity, especially in peripheral tissues such as muscle, adipose tissue and liver. The second AMPKK, CaMKKB, is particularly relevant to situations where Ca²⁺ signalling pathways are activated. Several studies have shown that Ca²⁺ signalling activates AMPK by CaMKK_B-dependent phosphorylation [28-30]. Some investigators have suggested that the CaMKK_B–AMPK axis plays an important role in cerebral Ca²⁺ signal transduction under physiological conditions as well as in ischaemic states (e.g., ischaemia-induced calcium influx through NMDA receptor activation) [31, 32]. Recently, TAK1, the third putative upstream AMPKK, (also referred to as MAPKK kinase-7 or MAP3K7) was identified by Momcilovic et al. [33, 34]. Subsequently, TAK1 has been shown to activate AMPK-dependent cytoprotective autophagy in tumour necrosis factor (TNF)-related apoptosis-inducing ligandtreated epithelial cells [35]. However, the precise role of TAK1 as an AMPKK requires further investigation.

Role of AMPK in the heart

The role of AMPK in the heart is currently under intensive study. The heart can be viewed simplistically as a circulating pump. In performing its function, the heart consumes considerable energy as a result of oxidation of various substrates, including fatty acids and glucose [36, 37]. Therefore, reduction of oxygen availability, as would occur during an ischaemic episode, has been postulated to trigger AMPK activation, which then acts as an emergency signal to restore cell energy homeostasis in the cardiomyocytes [38, 39]. Consistent with this, numerous reports have indicated that AMPK is very important in regulating substrate metabolism and in protecting against ischaemic reperfusion injury in the heart [24, 40, 41]. Further, two of the AMPKKs described earlier are found in the heart, LKB1 and CaMKKB [28, 42]. Importantly, it has been shown that LKB1 responds to ischaemic stress in heart tissue by phosphorylating the AMPK α subunit at Thr172 [43]. In a recent study, Sakamoto et al. demonstrated that under the ischaemic status LKB1 selectively phosphorylates the α 2 but not the α 1 subunit, suggesting differential regulation and distinct physiological roles of various catalytic complexes and their upstream kinases [16].

AMPK protects ischaemic cardiomvocvtes, although several mechanisms. First, it increases glucose uptake by stimulating the translocation of glucose transporter type 4 (GLUT4, Fig. 1) to the sarcolemmal membrane. Nishino et al. found that ischaemic preconditioning activates AMPK and up-regulates GLUT4 expression in a manner dependent on protein kinase C (PKC), suggesting that these upregulation events may contribute to attenuation of myocardial stunning [44]. The nitric oxide pathway also contributes to AMPK stimulation of glucose uptake and GLUT4 translocation in heart muscle [45]. Recently. Horie et al. demonstrated that oxidative stress induces GLUT4 translocation by activation of phosphoinositide 3-kinase (PI3K) and Akt and by dual AMPKK (CaMKKB and LKB1) activation in cardiac myocytes [46]. Secondly, AMPK indirectly stimulates 6-phosphofructo-1-kinase (PFK1, Fig. 1) activity by phosphorylating and activating 6-phosphofructo-2kinase (PFK2), the enzyme that synthesizes fructose 2,6-bisphosphate, to generate more energy. Third, AMPK protects cardiocytes by inhibiting apoptosis through several mechanisms, thus promoting cell survival. In 2005. Shibata et al. showed that the AMPK pathway is essential for the apoptosis-inhibiting effects of adiponectin in heart [47]. The translocation of Bax, a pro-apoptotic protein, to mitochondria is thought to be an early step in apoptosis induced by ischaemia and this process appears to occur in response to activation of p38 MAPK downstream of AMPK in heart [48]. Recently, AMPK was further defined as a powerful cardiac protector against cardiomyocyte apoptosis triggered by TNF- α through phosphorylation of Bad; subsequent suppression of the interaction between Bad and Bcl-xL limits cytochrome c release and caspase-3 activation [49]. Because ischaemic injury concurrently elevates TNF- α production, cytochrome *c* release, the caspase cascade and AMPK activation, these newly identified anti-apoptosis functions of AMPK may represent important pathogenic activities of AMPK pathway. Finally, AMPK is a critical controller in the autophagy process in ischaemic heart. Autophagy has been identified as a survival mechanism in ischaemic myocardium that reserves energy and substances necessary for cell survival [50-53]. In 2007, Liang et al. first demonstrated that the LKB1–AMPK pathway dictates entry into autophagy or apoptosis under ischaemic conditions by regulating p27(kip1) phosphorylation in heart [54]. Of relevance, it was found that ischaemia stimulates autophagy through an AMPK-dependent mechanism, and this autophagy is cardioprotective [55]. Collectively, these studies provide compelling evidence for the beneficial effects of AMPK on cardiac function.

Animal models have been applied to address the question whether AMPK plays a positive or negative role during processes associated with ischaemia/reperfusion [56]. The α 2 isoform is the pre-dominant catalytic isoform in the heart. The phosphorylation of the main substrate of AMPK, acetyl-CoA carboxylase (ACC, Fig. 1), is clearly decreased in both normoxic and ischaemic conditions in AMPK α 2 knockout mice [57, 58]. These studies also indicated that AMPK a2 is necessary for maintaining myocardial energy homeostasis during ischaemia. This phenotype was further confirmed in AMPK a2-dominant negative (DN) mouse hearts [59, 60]. Moreover, genetic manipulation of the AMPK pathway indicated additional roles for this enzyme in heart. In AMPK α 2-DN mouse hearts, AMPK critically contributes to p38 MAPK activation during ischaemia by promoting auto-phosphorylation of p38 MAPK through interaction with the scaffold protein transforming growth factor β activated protein kinase 1 binding protein 1 [61]. In a similar fashion, it was found that AMPK mediates pre-conditioning in cardiac cells by regulating the activity and recruitment of sarcolemmal K (ATP) channels without being a part of the signalling pathway that regulates mitochondrial membrane potential [62]. Furthermore, using a knock-down approach. Ahmad et al. demonstrated that the AMPK $\alpha 2$, rather than the $\alpha 1$ subunit, is the primary mediator of the effects of the 5'-AMP-activated protein kinase subunit γ^2 gene mutation, such as cardiomyopathy with cardiac hypertrophy, pre-excitation and glycogen deposition [63].

Role of AMPK in endothelial cells

The endothelium is a single layer of cells covering the inner surface of all blood vessels, including conduit arteries, resistance and capacitance vessels and capillaries. Endothelial cells are involved in many aspects of vascular biology, including control of blood pressure, thrombosis, fibrinolysis, atherosclerosis and inflammation. The pre-dominant catalytic isoform of AMPK expressed in vascular endothelial cells is $\alpha 1$ rather than $\alpha 2$ [64–66]. Endothelium-dependent vasodilatation is a vital mechanism of systemic blood flow regulation that occurs during periods of increased metabolic demand such as glucose deprivation and hypoxia. As a metabolic sensor, vascular AMPK is believed to be involved in the metabolic regulation of blood flow. It is now well established that a major function of endothelial cells is nitric oxide synthesis, which is regulated by the activity of endothelial nitric oxide synthase (eNOS, Fig. 1) [67-68]. Chen et al. (1999) first suggested that eNOS might represent a target for AMPK in endothelium [69]. In 2008, Hu et al. demonstrated that AMPK could phosphorylate eNOS at Ser1179, enhancing activity of this enzyme [70]. Recently, a second phosphorylation site on eNOS,

Ser633, was identified; the authors confirmed that AMPK phosphorylation of eNOS Ser633 is a functional signalling event contributing to nitric oxide bioavailability in endothelial cells [71]. Thus, AMPK directly affects endothelial cells function, although regulation of the phosphorylation of eNOS.

Consistent with these reports, we demonstrated in cultured human umbilical vein endothelial cells (HUVECs) that resveratrol, a natural extract, enhances phosphorylation of eNOS at Ser1177 and increases nitric oxide production via the AMPK pathway. Further, the enhanced eNOS activity and nitric oxide production is totally blocked by the specific AMPK inhibitor, Compound C [72]. Moreover, the AMPK pathway is required for the resveratrolinduced relaxation of aorta isolated from mice fed a high glucose diet [72]. In addition to these effects on eNOS, AMPK also appears to play an important role in the level of oxidative stress in endothelial cells. Zou et al. suggested that the increased generation of reactive oxygen species (ROS) promotes AMPK activation by the LKB1 complex in cultured endothelial cells [73]. Further, experiments by Ido et al. [74] showed that the AMPK activator, 5aminoimidazole-4-carboxamide ribonucleoside (AICAR), increases AMPK activity and completely prevents the marked increase in HUVEC apoptosis that results from oxidative stress.

The AMPK pathway may represent a 'common pathway' involved in the protective effects of many exogenous drugs or endogenous factors on endothelial cells. For example, it has been reported that a class of drugs that target peroxisome proliferatoractivated receptor α , used to modify blood lipids levels, stimulate eNOS phosphorylation and nitric oxide production through AMPK activation [75, 76]. In addition, metformin, a commonly used antidiabetes drug, reportedly exerts its therapeutic effects by activating the AMPK signalling pathway in endothelial cells [77–79]. The mechanisms underlying this protective effect include prevention of inflammatory cytokine production [77] and increased angiogenesis [79]. Rosiglitazone, an anti-diabetic drug in the thiazolidinedione class of drugs, not only protects endothelial cells against alucose-induced oxidative stress in an AMPK-dependent manner [80] but also stimulates nitric oxide synthesis in human aortic endothelial cells via AMPK [81]. In addition, some important hormones, such as adiponectin [82-84], ghrelin [85] and leptin [86], exert their biological functions though the AMPK signalling pathway. Collectively, these studies support the notion that the AMPK pathway is a physiologically relevant and potent protection mechanism in endothelial cells.

Role of AMPK in vascular smooth muscle cells

The most important function of vascular smooth muscle cells (VSMC) is vasoconstriction in response to many physiological and pathological stimuli. Moreover, proliferation and migration of VSMCs play critical roles in vascular inflammation and development of atherosclerosis. Vascular smooth muscle AMPK differs in

its expression profile compared to striated and cardiac muscle cells, with both $\alpha 1$ and $\alpha 2$ catalytic subunits being evident [64, 65, 87]. Further, the relative proportion of the α 1 and α 2 subunits varies between vessels [64, 65, 87]. Studies from Rubin et al. demonstrated that endothelium-denuded porcine carotid artery segments exhibit an increase in AMPK activity within 1 minute of being metabolically challenged with 2-deoxyglucose (2DG) and 2-deoxyglucose plus N₂ (N₂-2DG) [65]. Further, AMPK activation by N2-2DG was associated with a rapid and pronounced reduction in phosphorylation of Akt and ERK 1/2. Moreover, it has been reported that AMPK can phosphorylate and desensitize smooth muscle myosin light chain kinase to attenuate vasoconstriction, implying that AMPK contributes to the reduced ATP turnover in the tonic phase of smooth muscle contraction [88]. Using AMPK α 1 and α 2 knockout mice, Goirand *et al.* demonstrated that activation of AMPK α 1, but not α 2, leads to relaxation of aorta in mice in an endothelium- and eNOS-independent manner [87]. Consistent with these reports. AMPK was also found to be a key regulator of the contractile response of pulmonary arteries to acute hypoxia [89, 90]. Therefore, AMPK is a player in the complex signalling pathways that regulate vascular smooth muscle tone.

Apart from the acute regulation of vascular tone, AMPK is also involved in the proliferation of VSMC. AMPK expression blocks the growth of vascular smooth muscle induced by angiotensin-II [91]. In a rat femoral artery wire injury model, continuous activation of AMPK by AICAR injection significantly inhibits neointima formation [91]. Igata *et al.* showed that AMPK increased the expression of the cyclin-dependent kinase inhibitor p21cip via inhibition of phosphorylation of the retinoblastoma gene product, leading to inhibition of the proliferation of vascular smooth muscle [92]. Moreover, Liang *et al.* demonstrated that berberine, a herbal extract, inhibits platelet-derived growth factor-induced growth and migration in VSMCs, partly through the AMPK pathway [93]. Interestingly, AMPK also contributes to the activation of NAD(P)H:quinone oxidoreductase 1, which can prevent arterial restenosis by suppressing VSMC proliferation [94].

Role of AMPK in vascular angiogenesis

Angiogenesis is a physiological process involving the growth of new blood vessels from pre-existing vessels. It represents a normal and pivotal biological process in growth and development and is also important in wound healing and tumour growth. A critical role for AMPK in angiogenesis was first identified in cancer cells under nutrient deprivation. The introduction of AMPK antisense RNA expression vectors into pancreatic cancer cell lines, PANC-1 and AsPC-1, diminished their tolerance to glucose deprivation; in addition, the stable transfection of AMPK antisense RNA into PANC-1 cells inhibited tumour growth in nude mice [95]. Results from the studies of Nagata *et al.* demonstrated that AMPK signalling was a potent regulator of vascular angiogenesis and was specifically required for endothelial cell migration and differentiation under conditions of hypoxia [96]. These observations suggest that endothelial AMPK signalling may be a critical determinant of blood vessel recruitment to tissues exposed to ischaemic stress. Subsequently, the involvement of AMPK signalling in the proangiogenesis actions of adiponectin was observed [97]. Moreover, Yun et al. found that glucose deprivation increased mRNA stability of vascular endothelial growth factor (VEGF), a key hormone regulating angiogenesis, through activation of AMPK [98]. In another report, Orchi et al. provided the first direct evidence that AMPK activation by AICAR treatment increased VEGF expression and accelerated angiogenic repair of ischaemic hindlimbs in mice [99]. The mechanisms by which AMPK promotes vascular angiogenesis are being gradually elucidated. Levine et al. suggested that an AMPK-Rac1-Akt-eNOS axis in endothelial cells plays a central role in the vascular angiogenesis process [100]. Although several other studies support this hypothesis [101, 102], data from the studies of Zwetsloot et al. [103] suggest that AMPK is not necessarv for the angiogenic response to exercise, indicating that the role of AMPK may be more complex than it currently appears.

Role of AMPK in cardiovascular diseases

Atherosclerosis

Atherosclerosis is a chronic disease affecting medium and large arteries. In the past several decades, there has been a marked increase in our understanding of the pathogenesis of this vascular disorder [104]. Previously, atherosclerosis was presumed to be primarily a plumbing problem. Now it is deemed a chronic inflammatory disease in the walls of arteries, which originates from the interaction between cells of the arterial wall, lipoproteins and inflammatory cells, leading to the development of complex lesions or plagues. AMPK not only regulates proliferation, migration, apoptosis and autophagy of vascular endothelial and smooth muscle cells but also affects biological functions of macrophages, which are critical activators of atherosclerosis. Also relevant to the development of atherosclerosis is the fact that AMPK participates in whole body glucose, lipid and protein homeostasis, thus affecting serum lipid and glucose levels and insulin resistance. Therefore, it is an attractive hypothesis that AMPK-mediated signalling is intimately involved in the pathophysiological process of atherosclerosis.

The entry of vascular cells, especially VSMCs, into the cell cycle plays an important role in the development and progression of atherosclerosis. Studies have reported that treatment of human aortic smooth muscle cells (HASMC), with the AMPK activator AICAR results in phosphorylation of AMPK and ACC while significantly inhibiting HASMC proliferation induced by either platelet-derived growth factor-BB or fetal calf serum [92]. Cell cycle analysis showed that AMPK activation increases the number of cells in G0/G1-phase and reduces numbers in S- and G2/M-phase, sug-

gesting that AMPK activation can cause cell cycle arrest [105]. Consistent with this report, AICAR inhibits angiotensin II-stimulated VSMC thymidine incorporation, and administration of AICAR prevents neointimal formation in the rat balloon injury model. This suggests that AMPK might serve as a therapeutic target in treatment of atherosclerosis. However, there is currently no direct evidence conclusively showing AMPK to be protective in atherosclerosis. In 2006, Zang et al. showed that polyphenols increase AMPK and ACC phosphorylation and inhibit the hepatic lipid accumulation, hyperlipidaemia and accelerated aortic lesion formation seen in type 1 diabetic LDL receptor-deficient mice. These data imply that activation of AMPK may represent a potential protective mechanism against atherosclerosis [106]. Studies from Devaraj et al. revealed that adiponectin decreased C-reactive protein synthesis and secretion by endothelial cells, which could be mimicked by the AMPK activator AICAR and reversed by inhibition of AMPK [107]. Moreover, AMPK activation was found to contribute to the activity of adiponectin, which inhibits insulin-like growth factor-1 induced cell migration in VSMCs [108]. AMPK regulates the antioxidant status of vascular endothelial cells [109], and reduction of AMPK increases endoplasmic reticulum stress and atherosclerosis in vivo [110]. In the latter report, Dong et al. also demonstrated that AMPK $\alpha 2$ is the main physiological suppressor of endoplasmic reticulum stress in endothelial cells [110]. Notably, AMPK suppresses oxidized low-density lipoprotein-induced macrophage proliferation, which is a key event underlying the development of atherosclerosis [111]. Collectively, these studies suggest that AMPK protects the vascular wall from atherosclerosis and may represent a new therapeutic target in the treatment of atherosclerosis.

Hypertension

Vessel remodelling and/or vessel hypertrophy are responsible for the alterations in vascular structure observed in hypertension. Vessel remodelling occurs through reorganization of existing intracellular and extracellular components around the vascular lumen. In contrast, hyperplasia involves VSMC proliferation, migration, hypertrophy and polyploidy. Thus, AMPK may be involved in the genesis and development of hypertension. Studies from Kurdi et al. (2004) comparing left ventricle gene expression between severely hypertensive rats (transgenic rats that express the renin gene in the salivary gland) and normotensive control rats by cDNA macroarray revealed that AMPK is overexpressed in hypertensive rats [112]. Hattori et al. reported that metformin attenuates cytokine-induced expression of proinflammatory and adhesion molecule genes by inhibiting NF-kB activation via AMPK activation [77], which is pivotal in development of hypertension [113]. Moreover, impaired adiponectin-AMPK signalling has been reported in insulin-sensitive tissues of hypertensive rats by RodrÌguez et al. [114]. Recently, it was further reported that deletion of AMPK α 2 enhances the increase of phosphate-4E binding protein-1 seen in transverse aortic constriction and augments the T transverse aortic constriction-induced increase of phosphate-Akt, indicating that the AMPK signalling pathway exerts a cardiac

protective effect against pressure-overload-induced ventricular hypertrophy and dysfunction [115]. Collectively, the results from these studies support the notion that AMPK is protective against hypertension. However, it should be noted that activation of AMPK was found to enhance angiotensin II-induced proliferation in cardiac fibroblasts [116], implying AMPK might exert different effects in different cells in hypertension. Therefore, further investigation is needed to reveal the precise role of AMPK in hypertension, including the relevance of AMPK activation to the end-organ damage induced by hypertension.

Ischaemic stroke

Whether AMPK is beneficial or detrimental to the ischaemic brain is in dispute. Evidence from several independent groups indicates that AMPK is neuroprotective in ischaemic stroke [117-119], whereas there are also reports showing activation of AMPK is harmful in ischaemic stroke [120-121]. The brain, a high energyconsuming organ with very low energy reserves, relies heavily on a continuous flow of blood. Thus, AMPK in brain is likely to be activated when the ischaemic stroke occurs; a suggestion which is supported by a number of studies [117-121]. During severe energy depletion, neurons attempt to reserve energy. Apoptosis and autophagy both occur in neuronal cell outcomes after ischaemic stroke [122]. Moreover, different from other organs, AMPK a1 is expressed at low levels in brain; the AMPK a2 catalytic subunit accounts for more than 90% of AMPK in cerebral tissue [18]. In 2001, Culmsee et al. reported that AMPK activator AICAR protected hippocampal neurons against death induced by glucose deprivation, chemical hypoxia and exposure to glutamate and amyloid B peptide [119]. Dasgupta et al. found that resveratrol protects neurons through stimulation of AMPK activity [118]. Also, AMPK is associated with GABA(B) receptors, serving to enhance neuronal survival after ischaemia [32, 117]. It is worth noting that the detrimental effects of AMPK in stroke have been reported by only one group [120, 121]. Future studies using more selective AMPK activators or inhibitors, selective genetic manipulation in animal models and isolated genetic knockout neuronal cells will provide more information on this role of AMPK in stroke.

Conclusion and perspectives

Although the AMPK pathway is traditionally thought of as an intracellular fuel switch and controller of metabolism, the significance of AMPK in the cardiovascular system has recently been recognized [123]. A large body of evidence indicates that approaches aimed at increasing AMPK activity in the cardiovascular system may be therapeutic. A-769662, a thienopyridone, has been identified as a potent AMPK activator which directly stimulates partially purified rat liver AMPK [124]. Treatment of *ob/ob* mice with this agent lowered plasma glucose by 40%, reduced body-weight gain and significantly decreased both plasma and liver triglyceride levels. Evaluation of small molecular weight AMPK activators will shed new light on the treatment of cardiovascular disease.

Conflicts of interest

The authors confirm that there are no conflicts of interest.

References

- Kahn BB, Alquier T, Carling D, et al. AMP-activated protein kinase: ancient energy gauge provides clues to modern understanding of metabolism. *Cell Metab.* 2005: 1: 15–25.
- Kemp BE, Stapleton D, Campbell DJ, et al. AMP-activated protein kinase, super metabolic regulator. *Biochem Soc Trans.* 2003; 31: 162–8.
- Winder WW, Hardie DG. AMP-activated protein kinase, a metabolic master switch: possible roles in type 2 diabetes. *Am J Physiol.* 1999; 277: E1–10.
- Munday MR, Campbell DG, Carling D, et al. Identification by amino acid sequencing of three major regulatory phosphorylation sites on rat acetyl-CoA carboxylase. Eur J Biochem. 1988; 175: 331–8.
- 5. **Carling D, Clarke PR, Zammit VA**, *et al.* Purification and characterization of the

AMP-activated protein kinase. Copurification of acetyl-CoA carboxylase kinase and 3hydroxy-3-methylglutaryl-CoA reductase kinase activities. *Eur J Biochem*. 1989; 186: 129–36.

- Sanders MJ, Grondin PO, Hegarty BD, et al. Investigating the mechanism for AMP activation of the AMP-activated protein kinase cascade. *Biochem J.* 2007; 403: 139–48.
- Steinberg GR, Kemp BE. AMPK in Health and Disease. *Physiol Rev.* 2009; 89: 1025–78.
- Sakamoto K, Goransson O, Hardie DG, et al. Activity of LKB1 and AMPK-related kinases in skeletal muscle: effects of contraction, phenformin, and AICAR. Am J Physiol Endocrinol Metab. 2004; 287: E310–7.
- 9. Woods A, Johnstone SR, Dickerson K, et al. LKB1 is the upstream kinase in the

AMP-activated protein kinase cascade. *Curr Biol.* 2003; 13: 2004–8.

- Kemp BE, Mitchelhill KI, Stapleton D, et al. Dealing with energy demand: the AMP-activated protein kinase. Trends Biochem Sci. 1999; 24: 22–5.
- 11. **Hardie DG, Carling D**. The AMP-activated protein kinase-fuel gauge of the mammalian cell? *Eur J Biochem.* 1997; 246: 259–73.
- Kelly M, Keller C, Avilucea PR, et al. AMPK activity is diminished in tissues of IL-6 knockout mice: the effect of exercise. Biochem Biophys Res Commun. 2004; 320: 449–54.
- Ruderman NB, Park H, Kaushik VK, et al. AMPK as a metabolic switch in rat muscle, liver and adipose tissue after exercise. Acta Physiol Scand. 2003; 178: 435–42.

- Rutter GA, Da Silva Xavier G, Leclerc I. Roles of 5'-AMP-activated protein kinase (AMPK) in mammalian glucose homoeostasis. *Biochem J.* 2003; 375: 1–16.
- Sakamoto K, McCarthy A, Smith D, et al. Deficiency of LKB1 in skeletal muscle prevents AMPK activation and glucose uptake during contraction. *EMBO J.* 2005; 24: 1810–20.
- Sakamoto K, Zarrinpashneh E, Budas GR, et al. Deficiency of LKB1 in heart prevents ischemia-mediated activation of AMPKalpha2 but not AMPKalpha1. Am J Physiol Endocrinol Metab. 2006; 290: E780–8.
- Thomson DM, Porter BB, Tall JH, et al. Skeletal muscle and heart LKB1 deficiency causes decreased voluntary running and reduced muscle mitochondrial marker enzyme expression in mice. Am J Physiol Endocrinol Metab. 2007; 292: E196–202.
- Turnley AM, Stapleton D, Mann RJ, et al. Cellular distribution and developmental expression of AMP-activated protein kinase isoforms in mouse central nervous system. J Neurochem. 1999; 72: 1707–16.
- Hawley SA, Davison M, Woods A, et al. Characterization of the AMP-activated protein kinase kinase from rat liver and identification of threonine 172 as the major site at which it phosphorylates AMP-activated protein kinase. J Biol Chem. 1996; 271: 27879–87.
- Lizcano JM, Goransson O, Toth R, et al. LKB1 is a master kinase that activates 13 kinases of the AMPK subfamily, including MARK/PAR-1. EMBO J. 2004; 23: 833–43.
- Shaw RJ, Kosmatka M, Bardeesy N, et al. The tumor suppressor LKB1 kinase directly activates AMP-activated kinase and regulates apoptosis in response to energy stress. Proc Natl Acad Sci U S A. 2004; 101: 3329–35.
- Woods A, Vertommen D, Neumann D, et al. Identification of phosphorylation sites in AMP-activated protein kinase (AMPK) for upstream AMPK kinases and study of their roles by site-directed mutagenesis. J Biol Chem. 2003; 278: 28434–42.
- Warden SM, Richardson C, O'Donnell J Jr., et al. Post-translational modifications of the beta-1 subunit of AMP-activated protein kinase affect enzyme activity and cellular localization. *Biochem J.* 2001; 354: 275–83.
- Carling D. The AMP-activated protein kinase cascade-a unifying system for energy control. *Trends Biochem Sci.* 2004; 29: 18–24.

- 25. **Hardie DG.** The AMP-activated protein kinase pathway-new players upstream and downstream. *J Cell Sci.* 2004; 117: 5479–87.
- Hardie DG. New roles for the LKB1->AMPK pathway. *Curr Opin Cell Biol.* 2005; 17: 167–73.
- Jansen M, Ten Klooster JP, Offerhaus GJ, et al. LKB1 and AMPK family signaling: the intimate link between cell polarity and energy metabolism. *Physiol Rev.* 2009; 89: 777–98.
- Hawley SA, Pan DA, Mustard KJ, et al. Calmodulin-dependent protein kinase kinase-beta is an alternative upstream kinase for AMP-activated protein kinase. *Cell Metab.* 2005; 2: 9–19.
- Hong SP, Momcilovic M, Carlson M. Function of mammalian LKB1 and Ca²⁺/calmodulin-dependent protein kinase kinase alpha as Snf1-activating kinases in yeast. J Biol Chem. 2005; 280: 21804–9.
- Hurley RL, Anderson KA, Franzone JM, et al. The Ca²⁺/calmodulin-dependent protein kinase kinases are AMP-activated protein kinase kinases. J Biol Chem. 2005; 280: 29060–6.
- Birnbaum MJ. Activating AMP-activated protein kinase without AMP. *Mol Cell.* 2005; 19: 289–90.
- Hardie DG, Frenguelli BG. A neural protection racket: AMPK and the GABA(B) receptor. *Neuron*. 2007; 53: 159–62.
- Momcilovic M, Hong SP, Carlson M. Mammalian TAK1 activates Snf1 protein kinase in yeast and phosphorylates AMPactivated protein kinase *in vitro. J Biol Chem.* 2006; 281: 25336–43.
- Xie M, Zhang D, Dyck JR, et al. A pivotal role for endogenous TGF-beta-activated kinase-1 in the LKB1/AMP-activated protein kinase energy-sensor pathway. Proc Natl Acad Sci U S A. 2006; 103: 17378–83.
- Herrero-Martin G, Hoyer-Hansen M, Garcia-Garcia C, et al. TAK1 activates AMPK-dependent cytoprotective autophagy in TRAIL-treated epithelial cells. EMBO J. 2009; 28: 677–85.
- Lloyd SG, Wang P, Zeng H, et al. Impact of low-flow ischemia on substrate oxidation and glycolysis in the isolated perfused rat heart. Am J Physiol Heart Circ Physiol. 2004; 287: H351–62.
- Rösen P, Adrian M, Feuerstein J, et al. Glycolysis and glucose oxidation in the rat heart under nonrecirculating perfusion conditions. *Basic Res Cardiol.* 1984; 79: 307–12.
- 38. **Dyck JR, Lopaschuk GD.** AMPK alterations in cardiac physiology and pathology:

enemy or ally? *J Physiol.* 2006; 574: 95–112.

- Hue L, Beauloye C, Marsin AS, et al. Insulin and ischemia stimulate glycolysis by acting on the same targets through different and opposing signaling pathways. J Mol Cell Cardiol. 2002; 34: 1091–7.
- Young LH, Li J, Baron SJ, et al. AMP-activated protein kinase: a key stress signaling pathway in the heart. *Trends Cardiovasc Med.* 2005; 15: 110–8.
- Arad M, Seidman CE, Seidman JG. AMPactivated protein kinase in the heart: role during health and disease. *Circ Res.* 2007; 100: 474–88.
- Hawley SA, Boudeau J, Reid JL, et al. Complexes between the LKB1 tumor suppressor, STRAD alpha/beta and MO25 alpha/beta are upstream kinases in the AMP-activated protein kinase cascade. J Biol. 2003; 2.
- Altarejos JY, Taniguchi M, Clanachan AS, et al. Myocardial ischemia differentially regulates LKB1 and an alternate 5'-AMPactivated protein kinase kinase. J Biol Chem. 2005; 280: 183–90.
- Nishino Y, Miura T, Miki T, et al. Ischemic preconditioning activates AMPK in a PKC-dependent manner and induces GLUT4 up-regulation in the late phase of cardioprotection. *Cardiovasc Res.* 2004; 61: 610–9.
- 45. Li J, Hu X, Selvakumar P, et al. Role of the nitric oxide pathway in AMPK-mediated glucose uptake and GLUT4 translocation in heart muscle. Am J Physiol Endocrinol Metab. 2004; 287: E834–41.
- Horie T, Ono K, Nagao K, et al. Oxidative stress induces GLUT4 translocation by activation of PI3-K/Akt and dual AMPK kinase in cardiac myocytes. J Cell Physiol. 2008; 215: 733–42.
- Shibata R, Sato K, Pimentel DR, et al. Adiponectin protects against myocardial ischemia-reperfusion injury through AMPK- and COX-2-dependent mechanisms. Nat Med. 2005; 11: 1096–103.
- Capano M, Crompton M. Bax translocates to mitochondria of heart cells during simulated ischaemia: involvement of AMPactivated and p38 mitogen-activated protein kinases. *Biochem J.* 2006; 395: 57–64.
- Kewalramani G, Puthanveetil P, Wang F, et al. AMP-activated protein kinase confers protection against TNF-{alpha}induced cardiac cell death. Cardiovasc Res. 2009; 84: 42–53.
- 50. Yan L, Vatner DE, Kim SJ, et al. Autophagy in chronically ischemic

myocardium. *Proc Natl Acad Sci U S A.* 2005; 102: 13807–12.

- 51. **Rothermel BA, Hill JA.** Myocyte autophagy in heart disease: friend or foe? *Autophagy.* 2007; 3: 632–4.
- Nakai A, Yamaguchi O, Takeda T, et al. The role of autophagy in cardiomyocytes in the basal state and in response to hemodynamic stress. Nat Med. 2007; 13: 619–24.
- Hamacher-Brady A, Brady NR, Logue SE, et al. Response to myocardial ischemia/ reperfusion injury involves Bnip3 and autophagy. *Cell Death Differ.* 2007; 14: 146–57.
- Liang J, Shao SH, Xu ZX, et al. The energy sensing LKB1-AMPK pathway regulates p27(kip1) phosphorylation mediating the decision to enter autophagy or apoptosis. *Nat Cell Biol.* 2007; 9: 218–24.
- Matsui Y, Takagi H, Qu X, et al. Distinct roles of autophagy in the heart during ischemia and reperfusion: roles of AMPactivated protein kinase and Beclin 1 in mediating autophagy. Circ Res. 2007; 100: 914–22.
- Viollet B, Athea Y, Mounier R, et al. AMPK: Lessons from transgenic and knockout animals. Front Biosci. 2009; 14: 19–44.
- Zarrinpashneh E, Carjaval K, Beauloye C, et al. Role of the alpha2-isoform of AMPactivated protein kinase in the metabolic response of the heart to no-flow ischemia. Am J Physiol Heart Circ Physiol. 2006; 291: H2875–83.
- Carvajal K, Zarrinpashneh E, Szarszoi O, et al. Dual cardiac contractile effects of the alpha2-AMPK deletion in low-flow ischemia and reperfusion. Am J Physiol Heart Circ Physiol. 2007; 292: H3136–47.
- Russell RR, 3rd, Li J, Coven DL, et al. AMP-activated protein kinase mediates ischemic glucose uptake and prevents postischemic cardiac dysfunction, apoptosis, and injury. J Clin Invest. 2004; 114: 495–503.
- Xing Y, Musi N, Fujii N, et al. Glucose metabolism and energy homeostasis in mouse hearts overexpressing dominant negative alpha2 subunit of AMP-activated protein kinase. J Biol Chem. 2003; 278: 28372–7.
- Li J, Miller EJ, Ninomiya-Tsuji J, et al. AMP-activated protein kinase activates p38 mitogen-activated protein kinase by increasing recruitment of p38 MAPK to TAB1 in the ischemic heart. Circ Res. 2005; 97: 872–9.
- 62. **Sukhodub A, Jovanovic S, Du Q, et al.** AMP-activated protein kinase mediates

preconditioning in cardiomyocytes by regulating activity and trafficking of sarcolemmal ATP-sensitive K(+) channels. *J Cell Physiol.* 2007; 210: 224–36.

- Ahmad F, Arad M, Musi N, et al. Increased alpha2 subunit-associated AMPK activity and PRKAG2 cardiomyopathy. *Circulation*. 2005; 112: 3140–8.
- Evans AM, Mustard KJ, Wyatt CN, et al. Does AMP-activated protein kinase couple inhibition of mitochondrial oxidative phosphorylation by hypoxia to calcium signaling in O₂-sensing cells? J Biol Chem. 2005; 280: 41504–11.
- Rubin LJ, Magliola L, Feng X, et al. Metabolic activation of AMP kinase in vascular smooth muscle. J Appl Physiol. 2005; 98: 296–306.
- 66. Davis BJ, Xie Z, Viollet B, et al. Activation of the AMP-activated kinase by antidiabetes drug metformin stimulates nitric oxide synthesis in vivo by promoting the association of heat shock protein 90 and endothelial nitric oxide synthase. Diabetes. 2006; 55: 496–505.
- Dudzinski DM, Michel T. Life history of eNOS: partners and pathways. *Cardiovasc Res.* 2007; 75: 247–60.
- Förstermann U, Münzel T. Endothelial nitric oxide synthase in vascular disease: from marvel to menace. *Circulation*. 2006; 113: 1708–14.
- Chen ZP, Mitchelhill KI, Michell BJ, et al. AMP-activated protein kinase phosphorylation of endothelial NO synthase. FEBS Lett. 1999: 443: 285–9.
- Hu Z, Chen J, Wei Q, et al. Bidirectional actions of hydrogen peroxide on endothelial nitric-oxide synthase phosphorylation and function: co-commitment and interplay of Akt and AMPK. J Biol Chem. 2008; 283: 25256–63.
- Chen Z, Peng I-C, Sun W, et al. AMP-activated protein kinase functionally phosphorylates endothelial nitric oxide synthase Ser633. Circ Res. 2009; 104: 496–505.
- Xu Q, Hao X, Yang Q, et al. Resveratrol prevents hyperglycemia-induced endothelial dysfunction via activation of adenosine monophosphate-activated protein kinase. Biochem Biophys Res Commun. 2009; 388: 389–94.
- Zou M-H, Kirkpatrick SS, Davis BJ, et al. Activation of the AMP-activated protein kinase by the anti-diabetic drug metformin *in vivo*. Role of mitochondrial reactive nitrogen species. J Biol Chem. 2004; 279: 43940–51.
- 74. **Ido Y, Carling D, Ruderman N**. Hyperglycemia-induced apoptosis in

human umbilical vein endothelial cells: inhibition by the AMP-activated protein kinase activation. *Diabetes*. 2002; 51: 159–67.

- Murakami H, Murakami R, Kambe F, et al. Fenofibrate activates AMPK and increases eNOS phosphorylation in HUVEC. Biochem Biophys Res Commun. 2006; 341: 973–8.
- Okayasu T, Tomizawa A, Suzuki K, et al. PPARalpha activators upregulate eNOS activity and inhibit cytokine-induced NFkappaB activation through AMP-activated protein kinase activation. *Life Sci.* 2008; 82: 884–91.
- Hattori Y, Suzuki K, Hattori S, et al. Metformin inhibits cytokine-induced nuclear factor kappaB activation via AMPactivated protein kinase activation in vascular endothelial cells. *Hypertension*. 2006; 47: 1183–8.
- Xie Z, Dong Y, Scholz R, et al. Phosphorylation of LKB1 at serine 428 by protein kinase C-zeta is required for metformin-enhanced activation of the AMPactivated protein kinase in endothelial cells. *Circulation*. 2008; 117: 952–62.
- Xie Z, Dong Y, Zhang J, et al. Identification of the serine 307 of LKB1 as a novel phosphorylation site essential for its nucleocytoplasmic transport and endothelial cell angiogenesis. *Mol Cell Biol.* 2009; 29: 3582–96.
- Ceolotto G, Gallo A, Papparella I, et al. Rosiglitazone reduces glucose-induced oxidative stress mediated by NAD(P)H oxidase via AMPK-dependent mechanism. Arterioscler Thromb Vasc Biol. 2007; 27: 2627–33.
- Boyle JG, Logan PJ, Ewart M-A, et al. Rosiglitazone stimulates nitric oxide synthesis in human aortic endothelial cells via AMP-activated protein kinase. J Biol Chem. 2008; 283: 11210–7.
- Deng G, Long Y, Yu Y-R, et al. Adiponectin directly improves endothelial dysfunction in obese rats through the AMPK-eNOS Pathway. Int J Obes (Lond). 2010; 34: 165–71.
- Tomizawa A, Hattori Y, Kasai K. Induction of gene expression in response to globular adiponectin in vascular endothelial cells. *Life Sci.* 2009; 85: 457–61.
- 84. Chandrasekar B, Boylston WH, Venkatachalam K, et al. Adiponectin blocks interleukin-18-mediated endothelial cell death via APPL1-dependent AMPactivated protein kinase (AMPK) activation and IKK/NF-kappaB/PTEN suppression. *J Biol Chem.* 2008; 283: 24889–98.

- Xu X, Jhun BS, Ha CH, et al. Molecular mechanisms of ghrelin-mediated endothelial nitric oxide synthase activation. *Endocrinology*. 2008; 149: 4183–92.
- Procopio C, Andreozzi F, Laratta E, et al. Leptin-stimulated endothelial nitric-oxide synthase via an adenosine 5'-monophosphate-activated protein kinase/Akt signaling pathway is attenuated by interaction with C-reactive protein. Endocrinology. 2009: 150: 3584–93.
- Goirand F, Solar M, Athea Y, et al. Activation of AMP kinase alpha1 subunit induces aortic vasorelaxation in mice. *J Physiol.* 2007; 581: 1163–71.
- Horman S, Morel N, Vertommen D, et al. AMP-activated protein kinase phosphorylates and desensitizes smooth muscle myosin light chain kinase. J Biol Chem. 2008; 283: 18505–12.
- Robertson TP, Mustard KJW, Lewis TH, et al. AMP-activated protein kinase and hypoxic pulmonary vasoconstriction. Eur J Pharmacol. 2008; 595: 39–43.
- Barman SA, Zhu S, White RE. Hypoxia modulates cyclic AMP activation of BkCa channels in rat pulmonary arterial smooth muscle. *Lung.* 2005; 183: 353–61.
- Nagata D, Takeda R, Sata M, et al. AMP-activated protein kinase inhibits angiotensin II-stimulated vascular smooth muscle cell proliferation. *Circulation*. 2004; 110: 444–51.
- Igata M, Motoshima H, Tsuruzoe K, et al. Adenosine monophosphate-activated protein kinase suppresses vascular smooth muscle cell proliferation through the inhibition of cell cycle progression. *Circ Res.* 2005; 97: 837–44.
- Liang K-W, Yin S-C, Ting C-T, et al. Berberine inhibits platelet-derived growth factor-induced growth and migration partly through an AMPK-dependent pathway in vascular smooth muscle cells. Eur J Pharmacol. 2008; 590: 343–54.
- Kim S-Y, Jeoung NH, Oh CJ, et al. Activation of NAD(P)H:quinone oxidoreductase 1 prevents arterial restenosis by suppressing vascular smooth muscle cell proliferation. *Circ Res.* 2009; 104: 842–50.
- Kato K, Ogura T, Kishimoto A, et al. Critical roles of AMP-activated protein kinase in constitutive tolerance of cancer cells to nutrient deprivation and tumor formation. Oncogene. 2002; 21: 6082–90.
- Nagata D, Mogi M, Walsh K. AMPactivated protein kinase (AMPK) signaling in endothelial cells is essential for angiogenesis in response to hypoxic stress. *J Biol Chem.* 2003; 278: 31000–6.

- Ouchi N, Kobayashi H, Kihara S, et al. Adiponectin stimulates angiogenesis by promoting cross-talk between AMP-activated protein kinase and Akt signaling in endothelial cells. J Biol Chem. 2004; 279: 1304–9.
- Yun H, Lee M, Kim S-S, et al. Glucose deprivation increases mRNA stability of vascular endothelial growth factor through activation of AMP-activated protein kinase in DU145 prostate carcinoma. J Biol Chem. 2005: 280: 9963–72.
- Ouchi N, Shibata R, Walsh K. AMP-activated protein kinase signaling stimulates VEGF expression and angiogenesis in skeletal muscle. *Circ Res.* 2005; 96: 838–46.
- 100. Levine YC, Li GK, Michel T. Agonist-modulated regulation of AMP-activated protein kinase (AMPK) in endothelial cells. Evidence for an AMPK → Rac1 → Akt → endothelial nitric-oxide synthase pathway. *J Biol Chem.* 2007; 282: 20351–64.
- Webler AC, Michaelis UR, Popp R, et al. Epoxyeicosatrienoic acids are part of the VEGF-activated signaling cascade leading to angiogenesis. *Am J Physiol Cell Physiol.* 2008; 295: C1292–301.
- 102. Phoenix KN, Vumbaca F, Claffey KP. Therapeutic metformin/AMPK activation promotes the angiogenic phenotype in the ERalpha negative MDA-MB-435 breast cancer model. *Breast Cancer Res Tr.* 2009; 113: 101–11.
- 103. Zwetsloot KA, Westerkamp LM, Holmes BF, et al. AMPK regulates basal skeletal muscle capillarization and VEGF expression, but is not necessary for the angiogenic response to exercise. J Physiol. 2008; 586: 6021–35.
- Libby P, Theroux P. Pathophysiology of coronary artery disease. *Circulation*. 2005; 111: 3481–8.
- 105. Igata M, Motoshima H, Tsuruzoe K, et al. Adenosine monophosphate-activated protein kinase suppresses vascular smooth muscle cell proliferation through the inhibition of cell cycle progression. *Circ Res.* 2005; 97: 837–44.
- 106. Zang M, Xu S, Maitland-Toolan KA, et al. Polyphenols stimulate AMP-activated protein kinase, lower lipids, and inhibit accelerated atherosclerosis in diabetic LDL receptor-deficient mice. *Diabetes.* 2006; 55: 2180–91.
- 107. Devaraj S, Torok N, Dasu MR, et al. Adiponectin decreases C-reactive protein synthesis and secretion from endothelial cells: evidence for an adipose tissuevascular loop. Arterioscler Thromb Vasc Biol. 2008; 28: 1368–74.

- 108. Motobayashi Y, Izawa-Ishizawa Y, Ishizawa K, et al. Adiponectin inhibits insulin-like growth factor-1-induced cell migration by the suppression of extracellular signal-regulated kinase 1/2 activation, but not Akt in vascular smooth muscle cells. Hypertens Res. 2009; 32: 188–93.
- Colombo SL, Moncada S. AMPKalpha1 regulates the antioxidant status of vascular endothelial cells. *Biochem J.* 2009; 421: 163–9.
- 110. Dong Y, Zhang M, Liang B, et al. Reduction of AMP-activated protein kinase {alpha}2 increases endoplasmic reticulum stress and atherosclerosis in vivo. Circulation. 2010; 121: 792–803.
- 111. Ishii N, Matsumura T, Kinoshita H, et al. Activation of AMP-activated protein kinase suppresses oxidized low-density lipoprotein-induced macrophage proliferation. *J Biol Chem.* 2009; 284: 34561–9.
- 112. Kurdi M, Cerutti C, Randon J, et al. Macroarray analysis in the hypertrophic left ventricle of renin-dependent hypertensive rats: identification of target genes for renin. J Renin Angiotensin Aldosterone Syst. 2004; 5: 72–8.
- 113. Zhou M-S, Schulman IH, Raij L. Vascular inflammation, insulin resistance, and endothelial dysfunction in salt-sensitive hypertension: role of nuclear factor kappa B activation. J Hypertens. 2010; 28: 527–35.
- 114. Rodríguez A, Catalán V, Becerril S, et al. Impaired adiponectin-AMPK signalling in insulin-sensitive tissues of hypertensive rats. *Life Sci.* 2008; 83: 540–9.
- 115. Zhang P, Hu X, Xu X, et al. AMP activated protein kinase-alpha2 deficiency exacerbates pressure-overload-induced left ventricular hypertrophy and dysfunction in mice. Hypertension. 2008; 52: 918–24.
- 116. Hattori Y, Akimoto K, Nishikimi T, et al. Activation of AMP-activated protein kinase enhances angiotensin II-induced proliferation in cardiac fibroblasts. *Hypertension*. 2006; 47: 265–70.
- 117. Kuramoto N, Wilkins ME, Fairfax BP, et al. Phospho-dependent functional modulation of GABA(B) receptors by the metabolic sensor AMP-dependent protein kinase. *Neuron.* 2007; 53: 233–47.
- 118. **Dasgupta B, Milbrandt J.** Resveratrol stimulates AMP kinase activity in neurons. *Proc Natl Acad Sci USA.* 2007; 104: 7217–22.
- 119. Culmsee C, Monnig J, Kemp BE, et al. AMP-activated protein kinase is highly expressed in neurons in the developing rat brain and promotes neuronal survival

following glucose deprivation. *J Mol Neurosci.* 2001; 17: 45–58.

- McCullough LD, Zeng Z, Li H, et al. Pharmacological inhibition of AMP-activated protein kinase provides neuroprotection in stroke. J Biol Chem. 2005; 280: 20493–502.
- 121. Li J, Zeng Z, Viollet B, et al. Neuroprotective effects of adenosine

monophosphate-activated protein kinase inhibition and gene deletion in stroke. *Stroke.* 2007; 38: 2992–9.

- Li J, McCullough LD. Effects of AMPactivated protein kinase in cerebral ischemia. *J Cerebr Blood F Met.* 2010; 30: 480–92.
- 123. Nagata D, Hirata Y. The role of AMP-activated protein kinase in the

cardiovascular system. *Hypertens Res.* 2010; 33: 22–8.

124. Cool B, Zinker B, Chiou W, et al. Identification and characterization of a small molecule AMPK activator that treats key components of type 2 diabetes and the metabolic syndrome. *Cell Metab.* 2006; 3: 403–16.