

Article

Aminoglycoside 6'-N-acetyltransferase Type Ib [AAC(6')-Ib]-Mediated Aminoglycoside Resistance: Phenotypic Conversion to Susceptibility by Silver Ions

Craig M. Reeves, Jesus Magallon, Kenneth Rocha, Tung Tran, Kimberly Phan, Peter Vu, Yang Yi, Crista L. Oakley-Havens, José Cedano, Verónica Jimenez , Maria S. Ramirez  and Marcelo E. Tolmasky * 

Center for Applied Biotechnology Studies, Department of Biological Science, California State University Fullerton, Fullerton, CA 92831, USA; wizard5424@csu.fullerton.edu (C.M.R.); jesusmagallon91@csu.fullerton.edu (J.M.); kenneth.rocha@csu.fullerton.edu (K.R.); tungtran6186@yahoo.com (T.T.); kkphan@csu.fullerton.edu (K.P.); vupeter8@csu.fullerton.edu (P.V.); yiyang6655@csu.fullerton.edu (Y.Y.); cristalee0810@csu.fullerton.edu (C.L.O.-H.); Jcedano@csu.fullerton.edu (J.C.); vjimenezortiz@Exchange.FULLERTON.EDU (V.J.); msramirez@fullerton.edu (M.S.R.)

* Correspondence: mtolmasky@fullerton.edu; Tel.: +1-657-278-5263

Abstract: Clinical resistance to amikacin and other aminoglycosides is usually due to the enzymatic acetylation of the antimicrobial molecule. A ubiquitous resistance enzyme among Gram-negatives is the aminoglycoside 6'-N-acetyltransferase type Ib [AAC(6')-Ib], which catalyzes acetylation using acetyl-CoA as a donor substrate. Therapies that combine the antibiotic and an inhibitor of the inactivation reaction could be an alternative to treat infections caused by resistant bacteria. We previously observed that metal ions such as Zn²⁺ or Cu²⁺ in complex with ionophores interfere with the AAC(6')-Ib-mediated inactivation of aminoglycosides and reduced resistance to susceptibility levels. Ag¹⁺ recently attracted attention as a potentiator of aminoglycosides' action by mechanisms still in discussion. We found that silver acetate is also a robust inhibitor of the enzymatic acetylation mediated by AAC(6')-Ib in vitro. This action seems to be independent of other mechanisms, like increased production of reactive oxygen species and enhanced membrane permeability, proposed to explain the potentiation of the antibiotic effect by silver ions. The addition of this compound to *aac(6')-Ib* harboring *Acinetobacter baumannii* and *Escherichia coli* cultures resulted in a dramatic reduction of the resistance levels. Time-kill assays showed that the combination of silver acetate and amikacin was bactericidal and exhibited low cytotoxicity to HEK293 cells.

Keywords: ESKAPE; *Acinetobacter*; aminoglycosides; amikacin; acetyltransferase; silver; adjuvant



Citation: Reeves, C.M.; Magallon, J.; Rocha, K.; Tran, T.; Phan, K.; Vu, P.; Yi, Y.; Oakley-Havens, C.L.; Cedano, J.; Jimenez, V.; et al. Aminoglycoside 6'-N-acetyltransferase Type Ib [AAC(6')-Ib]-Mediated Aminoglycoside Resistance: Phenotypic Conversion to Susceptibility by Silver Ions. *Antibiotics* **2021**, *10*, 29. <https://doi.org/10.3390/antibiotics10010029>

Received: 8 December 2020

Accepted: 29 December 2020

Published: 31 December 2020

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Silver has been used for the treatment of human diseases since ancient times [1,2]. In particular, until the advent of antibiotics in the mid-20th century, silver may have been the most used medicine to treat infections [2]. Despite taking a backstage role after the advent of antibiotics, silver has been continuously used in different forms [1,2]. Furthermore, due to the multidrug resistance crisis, the utilization of silver as a component of combination therapies or other novel formulations has regained interest [3–7].

Multidrug resistant Gram-negatives are one of the most serious threats to human health [8,9]. In particular, *Acinetobacter baumannii* is responsible for a large fraction of multiresistant hospital outbreaks [10,11]. Infections caused by this bacterium present multiple clinical manifestations, high mortality, and refraction to treatment [12–14]. These characteristics positioned *A. baumannii* within the U.S. Centers for Disease Control's list of threats to human health [15,16]. Amikacin and other aminoglycosides are important components of the armamentarium against *A. baumannii* and other bacterial infections [17,18]. Furthermore, based on recent docking experiments, amikacin could be useful in the treatment of viral infections including COVID-19 [19]. Unfortunately, a substantial percentage

of *A. baumannii* clinical isolates have acquired resistance to these antibiotics. A common mechanism of resistance to aminoglycosides is the enzymatic transfer of an acetyl group from acetyl-CoA to the 6' amine group of the antibiotic molecule. The aminoglycoside 6'-N-acetyltransferase type Ib [AAC(6')-Ib], an enzyme coded for by the *aac(6')-Ib* gene found in integrons, transposons, plasmids, and chromosomes of Gram-negative bacteria, is responsible for most amikacin-resistant strains [20,21]. A strategy to regain aminoglycosides' effectiveness and reduce the risks of infections caused by multidrug resistant Gram-negatives is to develop inhibitors of the enzymatic reaction that could be administered in combination with the antibiotic [20,22,23]. This path proved to be successful in the case of β -lactamase-mediated resistance to β -lactams [22,24]. The recent finding that metal ions such as Zn^{2+} and Cu^{2+} , in complex with ionophores, inhibit the acetylation of aminoglycosides mediated by AAC(6')-Ib and reverse amikacin resistance in laboratory assays increased the expectations that viable formulations that can treat infections caused by resistant *A. baumannii* will be designed in the near future [25–28]. In this article, we describe the inhibition of AAC(6')-Ib-mediated amikacin-resistance by Ag^{1+} in *A. baumannii* and *E. coli*.

2. Results

2.1. Effect of Ag^{1+} on AAC(6')-Ib-Mediated Acetylation of Amikacin

Ag^{1+} drastically interfered with the acetylation of amikacin, kanamycin, and tobramycin catalyzed by AAC(6')-Ib. Figure 1A shows that while the addition of sodium acetate did not produce any changes in the acetylation levels, silver acetate completely obliterated the incorporation of an acetyl group to the aminoglycoside molecule. The strength of inhibition was assessed by determining the 50% inhibitory concentration values (IC₅₀) using kanamycin, tobramycin, or amikacin as substrates. The values found were 5.1, 3.5, and 3.1 μ M, respectively (Figure 1B).

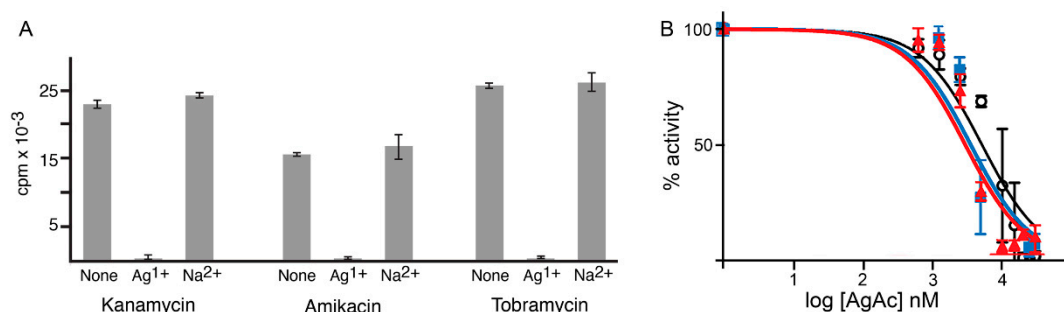


Figure 1. Effect of Ag^{1+} on AAC(6')-Ib activity. (A) acetylation activity of AAC(6')-Ib using kanamycin, tobramycin, or amikacin as substrates in the presence of Ag^{1+} . Silver acetate (AgAc) and sodium acetate were added at 100 μ M and the activity was compared to that observed in its absence. (B) The percentage of acetylation activity by AAC(6')-Ib was calculated for reaction mixtures containing different concentrations of silver acetate.

2.2. Silver Acetate Interferes with AAC(6')-Ib-Mediated Resistance to Amikacin

To determine if the silver acetate-mediated inhibition of AAC(6')-Ib activity observed in vitro has a significant impact on bacterial resistance, we assessed the effect on the growth of *aac(6')-Ib*-harboring *A. baumannii* and *E. coli* strains in amikacin containing media. Figure 2 shows that silver acetate dramatically reduced the growth of all these otherwise amikacin-resistant strains. Cultures in the presence of silver acetate were not affected at all, while partial inhibition of growth was observed in the presence of amikacin. These results showed that the presence of Ag^{1+} interferes with resistance to amikacin mediated by AAC(6')-Ib.

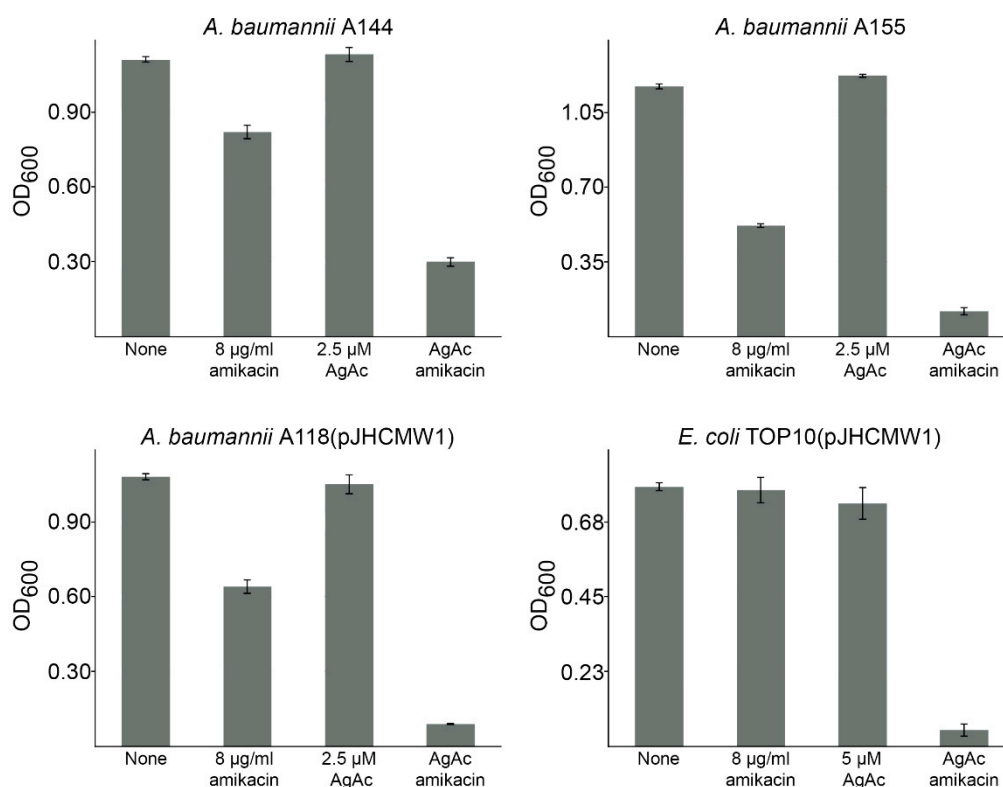


Figure 2. Effect of Ag^{1+} on AAC(6′)-Ib-mediated resistance to amikacin. *A. baumannii* A155, A144, and A118(pJHCMW1), and *E. coli* TOP10(pJHCMW1) were cultured in 100 μL Mueller–Hinton broth in microtiter plates at 37 °C, with the additions indicated in the figure and the OD₆₀₀ was determined after 20 h. The concentrations when both compounds were added to the cultures are those used when they were added as single addition. *p* values of the cultures containing silver acetate (AgAc), and amikacin were calculated with respect to the results obtained in cultures containing only amikacin. All four *p* values were statistically significant (<0.05).

2.3. Bactericidal Effect

We carried out time-kill assays to confirm that the severe reduction of growth observed when all four strains were cultured in the presence of the combination of silver acetate and amikacin was due to a bactericidal effect. Figure 3 shows that in tests using *A. baumannii* A144, A155, A118(pJHCMW1), and *E. coli* TOP10(pJHCMW1), the addition of silver acetate and amikacin had a robust bactericidal effect. As expected, these strains showed healthy growth when one of the components of the mix was omitted. These results confirmed that amikacin in the presence of Ag^{1+} ions regained full bactericidal power when resistance was mediated by the AAC(6′)-Ib enzyme.

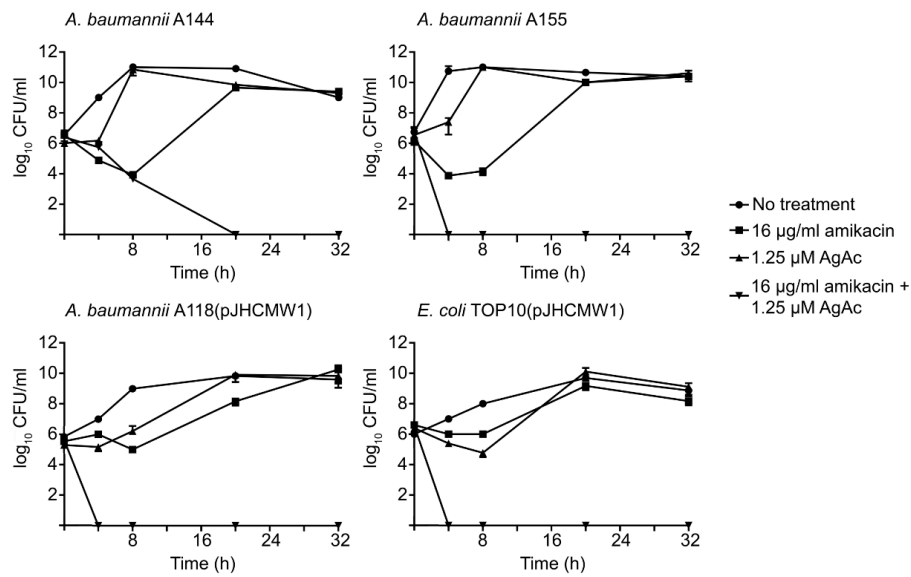


Figure 3. Time-kill assay curves for amikacin in the presence of silver acetate. *A. baumannii* A155, A144, and A118(pJHCMW1), and *E. coli* TOP10(pJHCMW1) cultures in Mueller-Hinton broth were incubated until they contained the indicated CFU/mL. Then, the different compounds were added, and the cultures were incubated at 37 °C. The CFU/mL values were measured at different intervals. AgAc, silver acetate.

2.4. Cytotoxicity

The cytotoxicity of the mix of silver acetate and amikacin, as well as that of the individual components, were tested on HEK293 cells as described in the Materials and Methods section. Addition of silver acetate or amikacin alone or in combination at the concentrations required to overcome resistance did not cause significant mortality in treated cells with respect to the control (Figure 4).

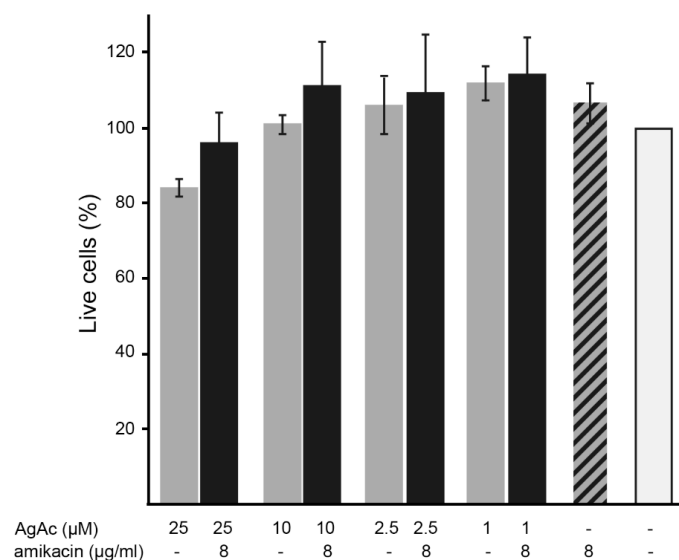


Figure 4. Cytotoxicity of silver acetate (AgAc) and amikacin. Cytotoxicity of silver acetate alone and in combination with amikacin on HEK293 cells was assayed using a LIVE/DEAD kit. The percentage of surviving cells was calculated relative to cells untreated (white bar). Cells incubated with 70% methanol were used as a control of maximum toxicity (striped bar). The experiments were conducted in triplicate and the values are mean \pm SD.

3. Discussion

The antibiotic resistance crisis has triggered the interest in finding creative alternatives to extend the life of antibiotics currently in use. Since one of the most ubiquitous acquired mechanisms of resistance to aminoglycosides is their enzymatic modification, compounds that interfere with this process would fulfill the purpose stated above. Numerous compounds have been proposed as candidates to inhibit enzymatic modification of aminoglycosides [20,22,23,29–31]. In the case of the AAC(6′)-Ib, the most common aminoglycoside resistance enzyme among AAC(6′)-I-carrying Gram-negatives [21,32], some small molecules have been identified that act as enzymatic inhibitors [20,23,29,33–35]. In addition to these advances, recent research revealed that Zn²⁺ and other metal ions inhibit the acetylation of aminoglycosides mediated by AAC(6′)-Ib in vitro [25–28,36,37]. The mechanism by which these metal ions interfere with enzymatic acetylation of aminoglycosides mediated by AAC(6′)-Ib is not yet known. A hypothesis has been put forward proposing the formation of a coordination complex that protects the substrate aminoglycoside from modification [26,38]. This manuscript shows that silver ions are also potent inhibitors of the enzymatic inactivation of aminoglycosides. However, while to overcome resistance to aminoglycosides in the presence of Zn²⁺ or Cu²⁺ ions at low concentrations, they had to be complexed to ionophores like pyrithione or clioquinol [25–28], the addition of low μM concentrations of silver acetate was sufficient. Our experiments showed that concentrations lower than 10 μM of silver acetate completely abolish resistance to amikacin in *aac(6′)-Ib*-carrying *A. baumannii* and *E. coli* strains. Furthermore, time-kill assays confirmed that the effect of the combination amikacin/silver acetate was bactericidal.

The potentiating effect of Ag¹⁺ ions on aminoglycoside antibiotics has been recently reported [4,7]. One of these reports proposed that an increase in reactive oxygen species production is the primary mechanism by which silver ions act as adjuvants [4]. On the other hand, another series of experiments pointed to an enhanced uptake level of antibiotics induced by silver ions as the molecular mechanism behind the observed activity [7]. Furthermore, the authors found no correlation between the silver activity and levels of reactive oxygen species [7]. While in these studies all experiments were carried on growing bacterial cells, our assays included the demonstration of interferences with acetylation in in vitro enzymatic reactions. Our results do not discard the possibilities proposed by the earlier work [4,7] but positively add another mechanism of enhancing resistance to aminoglycosides by inhibiting the enzymatic modification catalyzed by AAC(6′)-Ib. The results reported by other researchers [4,7] along with those described in this article led us to conclude that silver could act in vivo at multiple levels resulting in a synergistic effect that obliterates the resistance. The increase in membrane permeability produced by Ag¹⁺ [4,7] may be why an ionophore was not necessary to keep the active concentrations of the adjuvant at the μM levels. The mechanism by which silver increases permeability might be a combination of disturbances, including direct alteration of membrane proteins or an effect of the metal on ribosomes that interferes with proper translation and results in aborted and misfolded membrane proteins [1,4,7].

Silver has been used to treat infections since ancient times; the discovery that it can also act as potentiator of antimicrobials, in particular aminoglycosides, by numerous mechanisms that include the inhibition of aminoglycoside modifying enzymes, which can be very active and exist in high concentrations in the cytoplasm of resistant strains, makes combinations between Ag¹⁺ and aminoglycosides excellent candidates to treat multidrug resistant infections. The excitement about the possibility of using these mixes as therapeutic agents for a variety of infections is enhanced by the low toxicity the mix exhibited in our tests.

4. Materials and Methods

4.1. Bacterial Strains and Plasmids

Plasmids and strains used in this work are described in Table 1. All three *A. baumannii* strains used in this study are clinical isolates. Two of them, A144 and A155, naturally carry

aac(6′)-Ib. *A. baumannii* and *E. coli* TOP10 were transformed with the plasmid pJHCMW1, which harbors *aac(6′)-Ib*.

Table 1. Bacterial strains and plasmids used in this study.

Bacterial Strain	Relevant Characteristics, Genotype or Phenotype	Source or Reference
<i>E. coli</i>		
TOP10(pJHCMW1)	TOP10 (F ⁻ <i>mcrA</i> Δ(<i>mrr-hsdRMS-mcrBC</i>) Φ80 <i>lacZ</i> Δ <i>M15</i> Δ <i>lacX74</i> <i>recA1</i> <i>araD139</i> Δ(<i>ara-leu</i>)7697 <i>galU galK rpsL</i> (Str ^R) <i>endA1 nupG</i>). Transformed with pJHCMW1	[39]
<i>A. baumannii</i>		
A144	Human clinical isolate. Naturally carries <i>aac(6′)-Ib</i>	[40]
A155	Human clinical isolate. Naturally carries <i>aac(6′)-Ib</i>	[41]
A118(pJHCMW1)	Human clinical isolate transformed with pJHCMW1	[27]
Plasmids		
pJHCMW1	A 17 copies/cell plasmid harboring <i>aac(6′)-Ib</i>	[42,43]

4.2. General Procedures

Bacteria were cultured in Lennox L broth (1% tryptone, 0.5% yeast extract, 0.5% NaCl), and 2% agar was added in the case of solid medium. Inhibition of *A. baumannii* strains growth was determined in Mueller-Hinton broth at 37 °C with shaking in microtiter plates. The optical density at 600 nm (OD₆₀₀) of the cultures containing the specified additions was determined after 20 h incubation at 37 °C using the BioTek Synergy 5 microplate reader as described before [27]. Amikacin sulfate concentration is expressed in the usual µg/mL units; these numbers multiplied by 1.27 give the µM concentration. For time-kill assays, the cells were cultured in Mueller-Hinton broth until they reached 10⁶ CFU/mL before adding the indicated compounds. The cultures were continued at 37 °C with shaking, and the number of cells was determined by taking aliquots after 0, 4, 8, 20, and 32 h [28]. *E. coli* TOP10 and *A. baumannii* A118 were transformed with pJHCMW1 DNA [44] as described by Cohen et al. [45] and Ramirez et al. [46], respectively. Plasmid DNA preparations and DNA gel extractions were performed with the QIAprep Spin miniprep kit and QIAquick gel extraction kit, respectively (QIAGEN). Purification of AAC(6′)-Ib was carried out as before [29].

4.3. Acetyltransferase Assays

Acetyltransferase activity was assessed using two methodologies. One of them, the phosphocellulose paper binding assay [47], was carried out as previously described [48,49]. Briefly, 120 µg of protein from a soluble extract obtained from sonically disrupted *E. coli* TOP10(pJHCMW1) cells were added to a reaction mixture containing 200 mM Tris HCl pH 7.6 buffer, 0.25 mM MgCl₂, 330 µM antibiotic, 100 µM sodium acetate or silver acetate, and 0.05 µCi of [acetyl-1-¹⁴C]-acetyl-coenzyme A (specific activity 60 µCi/µmol) in a final volume of 30 µL. This mixture was incubated at 37 °C for 15 min and 20 µL were spotted on phosphocellulose paper strips. The unreacted radioactive substrate was washed once by submersion in 80 °C water followed by two washes with room temperature water. After drying, the radioactivity corresponding to the acetylated antibiotic was determined. The other method, utilized to calculate IC₅₀ values, consisted of monitoring the increase

in absorbance at 412 nm when the released CoA-SH from the substrate acetyl-CoA reacts with Ellman's reagent [5,5'-dithiobis(2-nitrobenzoic acid, DTNB)] [50]. The reaction mixture (150 μ M acetyl CoA, 0.2 mM DTNB, 20 mM Tris-HCl pH 7.5 buffer, 18 μ M antibiotic, and the required concentrations of silver acetate) was incubated at room temperature for 10 min. At this moment purified AAC(6')-Ib was added and acetylation was followed using a BioTek Synergy 2 plate reader monitoring absorbance at 412 nm. The initial velocities were calculated using the Gen 5 software, version 2.01.13. Inhibition was assessed by comparison of initial velocities of acetylation reactions in the presence or absence of silver acetate.

4.4. Cytotoxicity Assays

The cytotoxicity of the formulation silver acetate/amikacin was assessed on HEK293 cells [51] as described previously [29]. Briefly, 10^3 cells per well were inoculated and cultured overnight using flat-bottom 96-well, black microtiter plates. After this period, the compounds to be tested were added to cell-containing wells at the concentrations indicated, and incubation was continued for 24 h. At this point, the cells were washed with sterile D-PBS, resuspended in the LIVE/DEAD reagent (2 μ M ethidium homodimer 1 and 1 μ M calcein-AM) (Molecular Probes), and incubated for 30 min at 37 °C before determining the fluorescence levels at 645 nm (dead cells) and 530 nm (live cells). The percentage of dead cells was calculated relative to the untreated control cells. The maximum toxicity control was calculated treating the cells with 70% methanol for 10 min. Experiments were conducted in triplicate. The results were expressed as mean \pm SD of three independent experiments.

5. Conclusions

An enhancing effect of Ag¹⁺ ions on aminoglycosides had been recently reported. The causes proposed by different research groups were an increase in the production of reactive oxygen species and an enhanced uptake level of antibiotics induced by silver ions. The results showed here indicate that besides these mechanisms, Ag¹⁺ inhibits the acetylation of aminoglycosides catalyzed by AAC(6')-Ib. Addition of silver acetate at concentrations lower than 10 μ M sufficed to dramatically reduce the resistance to amikacin in *A. baumannii* and *E. coli* strains carrying the *aac(6')-Ib* gene. This observation, together with the low toxicity, make combinations of amikacin and silver acetate excellent candidates to treat infections caused by multidrug resistant Gram-negatives.

Author Contributions: Conceptualization, M.E.T., C.M.R., and M.S.R.; formal analysis, C.M.R., V.J., M.S.R., and M.E.T.; funding acquisition, M.E.T.; methodology, C.M.R., J.M., K.R., T.T., K.P., P.V., Y.Y., V.J., C.L.O.-H., J.C., M.S.R., and M.E.T.; resources, M.S.R. and M.E.T.; writing—original draft preparation, M.E.T.; writing—review and editing, M.S.R. and M.E.T. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by Public Health Service Grants 2R15AI047115 (MET) and SC3GM125556 (MSR) from the National Institute of Health. T.T. was partially supported by grant LA Basin Minority Health and Health Disparities Research Training Program (MHRT) T37MD001368 from the National Institute on Minority Health and Health Disparities, National Institute of Health.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Data is contained within the article.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Barras, F.; Aussel, L.; Ezraty, B. Silver and antibiotic, new facts to an old Story. *Antibiotics* **2018**, *7*, 79. [[CrossRef](#)] [[PubMed](#)]
2. Mijndonckx, K.; Leys, N.; Mahillon, J.; Silver, S.; Van Houdt, R. Antimicrobial silver: Uses, toxicity and potential for resistance. *Biomaterials* **2013**, *26*, 609–621. [[CrossRef](#)] [[PubMed](#)]

3. Montelongo-Peralta, L.Z.; Leon-Buitimea, A.; Palma-Nicolas, J.P.; Gonzalez-Christen, J.; Morones-Ramirez, J.R. Antibacterial activity of combinatorial treatments composed of transition-metal/antibiotics against *Mycobacterium tuberculosis*. *Sci. Rep.* **2019**, *9*, 5471. [[CrossRef](#)] [[PubMed](#)]
4. Morones-Ramirez, J.R.; Winkler, J.A.; Spina, C.S.; Collins, J.J. Silver enhances antibiotic activity against gram-negative bacteria. *Sci. Transl. Med.* **2013**, *5*, 190ra181. [[CrossRef](#)] [[PubMed](#)]
5. Garza-Cervantes, J.A.; Chavez-Reyes, A.; Castillo, E.C.; Garcia-Rivas, G.; Antonio Ortega-Rivera, O.; Salinas, E.; Ortiz-Martinez, M.; Gomez-Flores, S.L.; Pena-Martinez, J.A.; Pepi-Molina, A.; et al. Synergistic antimicrobial effects of silver/transition-metal combinatorial treatments. *Sci. Rep.* **2017**, *7*, 903. [[CrossRef](#)]
6. Garza-Cervantes, J.A.; Mendiola-Garza, G.; de Melo, E.M.; Dugmore, T.I.J.; Matharu, A.S.; Morones-Ramirez, J.R. Antimicrobial activity of a silver-microfibrillated cellulose biocomposite against susceptible and resistant bacteria. *Sci. Rep.* **2020**, *10*, 7281. [[CrossRef](#)]
7. Herisse, M.; Duverger, Y.; Martin-Verstraete, I.; Barras, F.; Ezraty, B. Silver potentiates aminoglycoside toxicity by enhancing their uptake. *Mol. Microbiol.* **2017**, *105*, 115–126. [[CrossRef](#)]
8. Adler, A.; Friedman, N.D.; Marchaim, D. Multidrug-resistant Gram-negative bacilli: Infection control implications. *Infect. Dis. Clin. N. Am.* **2016**, *30*, 967–997. [[CrossRef](#)]
9. Boucher, H.W.; Talbot, G.H.; Bradley, J.S.; Edwards, J.E.; Gilbert, D.; Rice, L.B.; Scheld, M.; Spellberg, B.; Bartlett, J. Bad bugs, no drugs: No ESKAPE! An update from the Infectious Diseases Society of America. *Clin. Infect. Dis.* **2009**, *48*, 1–12. [[CrossRef](#)]
10. Hartstein, A.I.; Rashad, A.L.; Liebler, J.M.; Actis, L.A.; Freeman, J.; Rourke, J.W., Jr.; Stibolt, T.B.; Tolmasky, M.E.; Ellis, G.R.; Crosa, J.H. Multiple intensive care unit outbreak of *Acinetobacter calcoaceticus* subspecies *anitratus* respiratory infection and colonization associated with contaminated, reusable ventilator circuits and resuscitation bags. *Am. J. Med.* **1988**, *85*, 624–631. [[CrossRef](#)]
11. McConnell, M.J.; Actis, L.; Pachon, J. *Acinetobacter baumannii*: Human infections, factors contributing to pathogenesis and animal models. *FEMS Microbiol. Rev.* **2013**, *37*, 130–155. [[CrossRef](#)] [[PubMed](#)]
12. Ramirez, M.S.; Bonomo, R.A.; Tolmasky, M.E. Carbapenemases: Transforming *Acinetobacter baumannii* into a yet more dangerous menace. *Biomolecules* **2020**, *10*, 720. [[CrossRef](#)] [[PubMed](#)]
13. Falagas, M.E.; Rafailidis, P.I. Attributable mortality of *Acinetobacter baumannii*: No longer a controversial issue. *Crit. Care* **2007**, *11*, 134. [[CrossRef](#)] [[PubMed](#)]
14. Dijkshoorn, L.; Nemec, A.; Seifert, H. An increasing threat in hospitals: Multidrug-resistant *Acinetobacter baumannii*. *Nat. Rev. Microbiol.* **2007**, *5*, 939–951. [[CrossRef](#)]
15. CDC. *Antibiotic Resistance Threats in the United States*; Centers for Disease Control: Atlanta, GA, USA, 2019.
16. Tacconelli, E.; Carrara, E.; Savoldi, A.; Harbarth, S.; Mendelson, M.; Monnet, D.L.; Pulcini, C.; Kahlmeter, G.; Kluytmans, J.; Carmeli, Y.; et al. Discovery, research, and development of new antibiotics: The WHO priority list of antibiotic-resistant bacteria and tuberculosis. *Lancet Infect. Dis.* **2018**, *18*, 318–327. [[CrossRef](#)]
17. Murray, C.K.; Hospenthal, D.R. Treatment of multidrug resistant *Acinetobacter*. *Curr. Opin. Infect. Dis.* **2005**, *18*, 502–506. [[CrossRef](#)]
18. Ramirez, M.S.; Tolmasky, M.E. Amikacin: Uses, resistance, and prospects for inhibition. *Molecules* **2017**, *22*, 2267. [[CrossRef](#)]
19. Kumar, P.; Bhardwaj, T.; Kumar, A.; Gehi, B.R.; Kapuganti, S.K.; Garg, N.; Nath, G.; Giri, R. Reprofilling of approved drugs against SARS-CoV-2 main protease: An in-silico study. *J. Biomol. Struct. Dyn.* **2020**, 1–15. [[CrossRef](#)]
20. Ramirez, M.S.; Tolmasky, M.E. Aminoglycoside modifying enzymes. *Drug Resist. Updates* **2010**, *13*, 151–171. [[CrossRef](#)]
21. Ramirez, M.S.; Nikolaidis, N.; Tolmasky, M.E. Rise and dissemination of aminoglycoside resistance: The *aac(6′)-Ib* paradigm. *Front. Microbiol.* **2013**, *4*, 121. [[CrossRef](#)]
22. Tolmasky, M.E. Strategies to prolong the useful life of existing antibiotics and help overcoming the antibiotic resistance crisis. In *Frontiers in Clinical Drug Research-Anti Infectives*; Rhaman, A., Ed.; Bentham Books: Sharjah, UAE, 2017; Volume 1, pp. 1–27.
23. Labby, K.J.; Garneau-Tsodikova, S. Strategies to overcome the action of aminoglycoside-modifying enzymes for treating resistant bacterial infections. *Future Med. Chem.* **2013**, *5*, 1285–1309. [[CrossRef](#)]
24. Bush, K.; Bradford, P.A. Interplay between beta-lactamases and new beta-lactamase inhibitors. *Nat. Rev. Microbiol.* **2019**, *17*, 295–306. [[CrossRef](#)]
25. Chiem, K.; Fuentes, B.A.; Lin, D.L.; Tran, T.; Jackson, A.; Ramirez, M.S.; Tolmasky, M.E. Inhibition of aminoglycoside 6′-N-acetyltransferase type Ib-mediated amikacin resistance in *Klebsiella pneumoniae* by zinc and copper pyrithione. *Antimicrob. Agents Chemother* **2015**, *59*, 5851–5853. [[CrossRef](#)]
26. Chiem, K.; Hue, F.; Magallon, J.; Tolmasky, M.E. Inhibition of aminoglycoside 6′-N-acetyltransferase type Ib-mediated amikacin resistance by zinc complexed with clioquinol, an ionophore active against tumors and neurodegenerative diseases. *Int. J. Antimicrob. Agents* **2018**, *51*, 271–273. [[CrossRef](#)]
27. Lin, D.L.; Tran, T.; Alam, J.Y.; Herron, S.R.; Ramirez, M.S.; Tolmasky, M.E. Inhibition of aminoglycoside 6′-N-acetyltransferase type Ib by zinc: Reversal of amikacin resistance in *Acinetobacter baumannii* and *Escherichia coli* by a zinc ionophore. *Antimicrob. Agents Chemother* **2014**, *58*, 4238–4241. [[CrossRef](#)] [[PubMed](#)]
28. Magallon, J.; Chiem, K.; Tran, T.; Ramirez, M.S.; Jimenez, V.; Tolmasky, M.E. Restoration of susceptibility to amikacin by 8-hydroxyquinoline analogs complexed to zinc. *PLoS ONE* **2019**, *14*, e0217602. [[CrossRef](#)] [[PubMed](#)]
29. Tran, T.; Chiem, K.; Jani, S.; Arivett, B.A.; Lin, D.L.; Lad, R.; Jimenez, V.; Farone, M.B.; Debevec, G.; Santos, R.; et al. Identification of a small molecule inhibitor of the aminoglycoside 6′-N-acetyltransferase type Ib [AAC(6′)-Ib] using mixture-based combinatorial libraries. *Int. J. Antimicrob. Agents* **2018**, *51*, 752–761. [[CrossRef](#)] [[PubMed](#)]

30. Guan, J.; Vong, K.; Wee, K.; Fakhoury, J.; Dullaghan, E.; Auclair, K. Cellular studies of an aminoglycoside potentiator reveal a new inhibitor of aminoglycoside resistance. *Chembiochem* **2018**, *19*, 2107–2113. [[CrossRef](#)]
31. Vong, K.; Tam, I.S.; Yan, X.; Auclair, K. Inhibitors of aminoglycoside resistance activated in cells. *ACS Chem. Biol.* **2012**, *7*, 470–475. [[CrossRef](#)]
32. Haldorsen, B.C.; Simonsen, G.S.; Sundsfjord, A.; Samuelsen, O.; Norwegian Study Group on Aminoglycoside Resistance. Increased prevalence of aminoglycoside resistance in clinical isolates of *Escherichia coli* and *Klebsiella* spp. in Norway is associated with the acquisition of AAC(3)-II and AAC(6′)-Ib. *Diagn. Microbiol. Infect. Dis.* **2014**, *78*, 66–69. [[CrossRef](#)]
33. Chiem, K.; Jani, S.; Fuentes, B.; Lin, D.L.; Rasche, M.E.; Tolmasky, M.E. Identification of an inhibitor of the aminoglycoside 6′-N-acetyltransferase type Ib [AAC(6′)-Ib] by glide molecular docking. *Medchemcomm* **2016**, *7*, 184–189. [[CrossRef](#)] [[PubMed](#)]
34. Lin, D.L.; Tran, T.; Adams, C.; Alam, J.Y.; Herron, S.R.; Tolmasky, M.E. Inhibitors of the aminoglycoside 6′-N-acetyltransferase type Ib [AAC(6′)-Ib] identified by in silico molecular docking. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 5694–5698. [[CrossRef](#)] [[PubMed](#)]
35. Lombes, T.; Begis, G.; Maurice, F.; Turcaud, S.; Lecourt, T.; Dardel, F.; Micouin, L. NMR-guided fragment-based approach for the design of AAC(6′)-Ib ligands. *Chembiochem* **2008**, *9*, 1368–1371. [[CrossRef](#)] [[PubMed](#)]
36. Bohlmann, L.; De Oliveira, D.M.P.; El-Deeb, I.M.; Brazel, E.B.; Harbison-Price, N.; Ong, C.Y.; Rivera-Hernandez, T.; Ferguson, S.A.; Cork, A.J.; Phan, M.D.; et al. Chemical synergy between ionophore PBT2 and zinc reverses antibiotic resistance. *mBio* **2018**, *9*, e02391-18. [[CrossRef](#)] [[PubMed](#)]
37. Li, Y.; Green, K.D.; Johnson, B.R.; Garneau-Tsodikova, S. Inhibition of aminoglycoside acetyltransferase resistance enzymes by metal salts. *Antimicrob. Agents Chemother* **2015**, *59*, 4148–4156. [[CrossRef](#)] [[PubMed](#)]
38. Kozłowski, H.; Kowalik-Jankowska, T.; Jezowska-Bojczuk, M. Chemical and biological aspects of Cu²⁺ interactions with peptides and aminoglycosides. *Coordin Chem Rev.* **2005**, *249*, 2323–2334. [[CrossRef](#)]
39. Ramirez, M.S.; Parenteau, T.R.; Centron, D.; Tolmasky, M.E. Functional characterization of Tn1331 gene cassettes. *J. Antimicrob. Chemother* **2008**, *62*, 669–673. [[CrossRef](#)]
40. Vilacoba, E.; Deraspe, M.; Traglia, G.M.; Roy, P.H.; Ramirez, M.S.; Centron, D. Draft genome sequence of an international clonal lineage 1 *Acinetobacter baumannii* strain from Argentina. *Genome Announc.* **2014**, *2*, e01190-14. [[CrossRef](#)]
41. Arivett, B.A.; Fiester, S.E.; Ream, D.C.; Centron, D.; Ramirez, M.S.; Tolmasky, M.E.; Actis, L.A. Draft genome of the multidrug-resistant *Acinetobacter baumannii* strain A155 clinical isolate. *Genome Announc.* **2015**, *3*, e00212-15. [[CrossRef](#)]
42. Reyes-Lamothe, R.; Tran, T.; Meas, D.; Lee, L.; Li, A.M.; Sherratt, D.J.; Tolmasky, M.E. High-copy bacterial plasmids diffuse in the nucleoid-free space, replicate stochastically and are randomly partitioned at cell division. *Nucleic Acids Res.* **2014**, *42*, 1042–1051. [[CrossRef](#)]
43. Sarno, R.; McGillivray, G.; Sherratt, D.J.; Actis, L.A.; Tolmasky, M.E. Complete nucleotide sequence of *Klebsiella pneumoniae* multiresistance plasmid pJHCMW1. *Antimicrob. Agents Chemother* **2002**, *46*, 3422–3427. [[CrossRef](#)] [[PubMed](#)]
44. Tolmasky, M.E.; Crosa, J.H. Tn1331, a novel multiresistance transposon encoding resistance to amikacin and ampicillin in *Klebsiella pneumoniae*. *Antimicrob. Agents Chemother* **1987**, *31*, 1955–1960. [[CrossRef](#)] [[PubMed](#)]
45. Cohen, S.N.; Chang, A.C.; Hsu, L. Nonchromosomal antibiotic resistance in bacteria: Genetic transformation of *Escherichia coli* by R-factor DNA. *Proc. Natl. Acad. Sci. USA* **1972**, *69*, 2110–2114. [[CrossRef](#)] [[PubMed](#)]
46. Ramirez, M.S.; Don, M.; Merckier, A.K.; Bistue, A.J.; Zorreguieta, A.; Centron, D.; Tolmasky, M.E. Naturally competent *Acinetobacter baumannii* clinical isolate as a convenient model for genetic studies. *J. Clin. Microbiol.* **2010**, *48*, 1488–1490. [[CrossRef](#)]
47. Haas, M.J.; Dowding, J.E. Aminoglycoside-modifying enzymes. *Methods Enzymol.* **1975**, *43*, 611–628.
48. Tolmasky, M.E.; Roberts, M.; Woloj, M.; Crosa, J.H. Molecular cloning of amikacin resistance determinants from a *Klebsiella pneumoniae* plasmid. *Antimicrob. Agents Chemother* **1986**, *30*, 315–320. [[CrossRef](#)]
49. Woloj, M.; Tolmasky, M.E.; Roberts, M.C.; Crosa, J.H. Plasmid-encoded amikacin resistance in multiresistant strains of *Klebsiella pneumoniae* isolated from neonates with meningitis. *Antimicrob. Agents Chemother* **1986**, *29*, 315–319. [[CrossRef](#)]
50. Ellman, G.L.; Courtney, K.D.; Andres, V., Jr.; Feather-Stone, R.M. A new and rapid colorimetric determination of acetylcholinesterase activity. *Biochem. Pharmacol.* **1961**, *7*, 88–95. [[CrossRef](#)]
51. Graham, F.L.; Smiley, J.; Russell, W.C.; Nairn, R. Characteristics of a human cell line transformed by DNA from human adenovirus type 5. *J. Gen. Virol.* **1977**, *36*, 59–74. [[CrossRef](#)]