

# ET-1 Promotes Epithelial–Mesenchymal Transition in Oral Squamous Cell Carcinoma Cells via the microRNA-489-3p /TWIST Axis

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**Objective:** Oral squamous cell carcinoma (OSCC) constitutes almost 90% of head and neck malignancies and has a poor prognosis. To improve the efficacy of OSCC therapy, it is of great significance to explore other therapy for OSCC. Endothelin-1 (ET-1), a potent vasoconstrictor peptide, is implicated in cancer pathogenesis. Moreover, ET-1 promotes epithelial-mesenchymal transition (EMT) during the development of human cancers. We further to found that ET-1 exposure induced EMT in human squamous cell carcinoma cell lines SCC4 and SAS, by enhancing the expression of EMT biomarkers N-cadherin and vimentin and reducing E-cadherin expression via upregulation of the transcription factor TWIST.

**Materials and Methods:** Cell motility was examined by migration, invasion and wound-healing assays. Quantitative real time polymerase chain reaction (q-PCR), and promoter assays confirmed the inhibitory effects of ET-1 on miRNAs expression in oral cancer cells. We demonstrate an intravenous injection model of lung metastasis followed by an advanced method for quantifying metastatic tumor using image analysis software.

**Results:** In addition, ET-1/ETAR reduced levels of microRNA-489-3p (miR-489-3p), a transcriptional repressor of TWIST. We have identified a novel bypass mechanism through which ET-1/ETAR are involved in TWIST signaling and downregulate miR-489-3p expression, enabling OSCC cells to acquire the EMT phenotype. Notably, ET-1 knockdown dramatically decreased levels of EMT markers and cell migration potential.

**Conclusion:** The role of ET-1 in OSCC progression is supported by our findings from an in vivo murine model of OSCC. ET-1 may therefore represent a novel molecular therapeutic target in OSCC metastasis.

**Keywords:** oral squamous cell carcinoma, endothelin-1, TWIST, microRNA-489-3p, epithelial–mesenchymal transition

## Introduction

Oral squamous cell carcinoma (OSCC) is a common cause of mortality worldwide, with oral squamous cell carcinoma (OSCC) being the predominant form, accounting for over 90% of all oral malignancies.<sup>1</sup> However, despite treatment advances including multi-agent chemotherapy, radiotherapy, and targeted therapy, the overall 5-year survival rate for OSCC remains disappointingly low (<50%), because of its aggressively invasive and metastatic nature, such that most patients with metastatic disease die within 1 year. Thus, metastasis is a major obstacle that must be overcome for the successful treatment of OSCC. Exploring the molecular basis of metastasis may help to improve the early detection, prevention, intervention, and prognostic evaluation of OSCC patients.<sup>2,3</sup>

EMT plays a major role in local recurrence and lymph node metastasis and is associated with a low survival rate in patients with oral squamous cell carcinoma (OSCC).<sup>4</sup> During EMT, epithelial cells gradually lose epithelial structural molecules, polarity, and adhesion capacity, and acquire mesenchymal traits, motility, and invasiveness.<sup>1,15</sup> Activation of the EMT results in decreased expression of epithelial markers (E-cadherin and  $\beta$ -catenin) and increased expression of adhesion and mesenchymal proteins (including vimentin, N-cadherin and fibronectin). Twist1, a basic helix-loop-helix transcription factor that induces tumor initiation, cell proliferation, development of resistance to anti-cancer drugs, and inhibition of apoptosis. However, the most well characterized role of Twist1 is in cell invasion, epithelial-to-mesenchymal transition (EMT), and tumor metastasis.

Previously studies, shown Serum big ET-1 or salivary ET-1 both as a biomarker of OSCC, by correlating it with the clinical staging and the histopathological grading.<sup>5,6</sup> The secreted peptide endothelin-1 (ET-1) is a widely expressed member of the endothelin family of proteins. Overexpression of the ET axis (ie, the combination of ET-1 and its receptors ET-A and ET-B) is implicated in the pathobiology of numerous different types of tumors.<sup>7</sup> Evidence supports the importance of an association between ET-1 expression and tumor progression and metastasis<sup>8</sup> and it is recognized that the ET-1/ET-A receptor (ET<sub>A</sub>R) autocrine pathway induces an invasive phenotype that drives EMT in ovarian cancer cells.<sup>9,10</sup> Thus, it may be worth targeting ET-1-induced EMT-dependent phenotypes as a way of preventing or treating tumor angiogenesis and metastasis.

Endogenous microRNAs (miRNAs) are small, evolutionarily conserved non-coding ribonucleotide acids that are crucial players in many biological processes involving normal cell development and cellular homeostasis, as well as cancer-relevant processes.<sup>11</sup> By binding to complementary sequences in the 3'-untranslated regions (3'-UTRs) of their target mRNAs, miRNAs interfere with mRNA degradation and translation.<sup>12,13</sup> Moreover, miRNAs modulate the metastatic process in many tumors.<sup>14,15</sup> Recently, using miRNA microarray analysis, we identified a group of miRNAs differentially expressed in mesenchymal-like versus epithelial-like cancer cells.<sup>16,17</sup> Remarkably, miR-489-3p was downregulated in cancer cells that had undergone EMT compared with those that had a typical epithelial phenotype, suggesting that miR-489-3p may regulate the EMT process.<sup>18</sup>

Among patients with oral squamous cell carcinoma (OSCC), high ET-1 or Twist1 expression has been reported in OSCC patients with a poorly differentiated tumor accompanied by metastasis. Twist1, in turn, activates Bmi1, both of which are essential for promoting EMT and tumor-initiating capacity.<sup>19–21</sup> However, there have been no studies on the importance of the expression of the ET-1/Twist1 axis, and this axis signalling pathways in OSCC is still poorly understood. Therefore, we examined whether miR-489-3p/Twist1 plays a role in ET-1-mediated metastasis in human OSCC. We found that ET-1 promotes tumor metastasis and EMT markers expression by down regulating miR-489-3p expression via the TWIST signalling pathway.

## Materials and Methods

### Cell Culture

Two kinds of human tongue squamous carcinoma derived cells were used, including SAS cell line (a gift from Ph.D. Chih-Hsin Tang, 2018 from China Medical University, Taichung, Taiwan), SCC-4 cell line (CRL-1624; American Type Culture Collection (ATCC)).<sup>22</sup> The SAS cell line has been authenticated by short tandem repeat (STR) DNA typing (Mission Biotech, Taipei, Taiwan). Cells were maintained in F12/Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS), 1% penicillin, 15 mM HEPES, 1 mM sodium pyruvate and 400 mg/mL hydrocortisone. Cells were maintained at 37°C in an incubator with 5% CO<sub>2</sub>. The medium was changed every 3 days. Subcultivation was carried out at 70–80% of cell confluence.

### Migration and Invasion Assay

The migration assay was performed using Transwell inserts (Costar, NY, USA; 8-mm pore size) in 24-well dishes. For invasion assay, filters were precoated with 30  $\mu$ L Matrigel basement membrane matrix (BD Biosciences, Bedford, MA, USA) for 30 min. The following procedures were the same for both migration and invasion assays. After the treatment with ET-1 (0, 10, 30 and 100 nM) for 24 h, cells were harvested and seeded to Transwell at  $1 \times 10^4$  cells/well in serum-free medium and then incubated for 24 h at 37 °C in 5% CO<sub>2</sub>. Cells were then fixed in 3.7% formaldehyde for 5 min and stained with 0.05% crystal violet in PBS for 15 min. Cells on the upper side of the filters were removed with cotton-tipped swabs, and the

filters were washed with PBS. Cells on the underside of the filters were examined and counted under a microscope. Each experiment was performed in triplicate and repeated at least three times.<sup>23</sup>

## Quantitative Real-Time PCR

Total cDNA (100 ng) was mixed with TaqMan<sup>®</sup> primers and probes as well as PCR Master Mix. The StepOnePlus<sup>™</sup> system was used in the quantitative RT-PCR assays. For the detection of miRNAs, reverse transcription was performed using Mir-X<sup>™</sup> miRNA First-Strand Synthesis and the SYBR<sup>®</sup> RT-PCR kit. qPCR analysis was carried out according to an established protocol. All mRNA sequences have been provided in the supplementary information ([Supplementary data Table S1](#) and [S2](#)).

## Plasmid Construction and Luciferase Activity Assay

Wild-type (wt) Twist-3'-UTR was constructed into the pGL2-Control vector. The predicted Twist binding site for miRNA was identified by the Targetscan (<http://www.targetscan.org/>). Mutant plasmids that attenuate the interaction between Twist-3 3'UTR and miRNA were generated using a QuikChange Site-Directed Mutagenesis kit (Stratagene, Cedar Creek, TX, USA). These plasmids were transfected into cells using Lipofectamine 2000. Following transfection, these cells were incubated with the indicated agents. Cell extracts were prepared and used for measuring the luciferase and  $\beta$ -galactosidase activities.<sup>24</sup>

## In vivo Tumor Xenograft Study

SCC4 cells that constitutively expressed pLenti CMV V5-Luc were co-transfected with pCMV plasmids alone or harboring human SCC-4/Luc control shRNA, or SCC4/Luc-ET-1 shRNA, respectively. These cells ( $2 \times 10^6$ ) that were resuspended in serum-free DMEN/ $\alpha$ -MEM were intravenously injected into the lateral tail vein of severe combined immunodeficiency (SCID) mice. Lung metastasis was monitored using an in vivo imaging system (Xenogen IVIS imaging system). After six weeks, the mice were humanely sacrificed and the tumor tissues were removed and photographed. The mice study was handled in accordance with the Animal Care and Use Guidelines of the Tunghai University (Taichung, Taiwan) under a protocol approved by the Institutional Animal Care and Use.

## Immunohistochemistry (IHC)

In this study, most human OSCC tissue array was purchased from Biomax (Rockville, MD; 6 cases for normal cartilage, 24 cases for grade I OSCC, 9 cases for grade II OSCC, and 15 cases for grade III OSCC). And some human tissues were purchased from Taipei medical hospital and the Ethics Committee of Taipei medical Hospital approved the study protocol, which followed institutional guidelines (N201804035). OSCC sample was treated with 3% hydrogen peroxide, then cover with 3% BSA. The tissue samples were incubated with primary antibodies against ET-1 and Twist. IHC analysis was carried out to determine the expression of lymphangiogenic marker according to the standard protocol.<sup>25</sup>

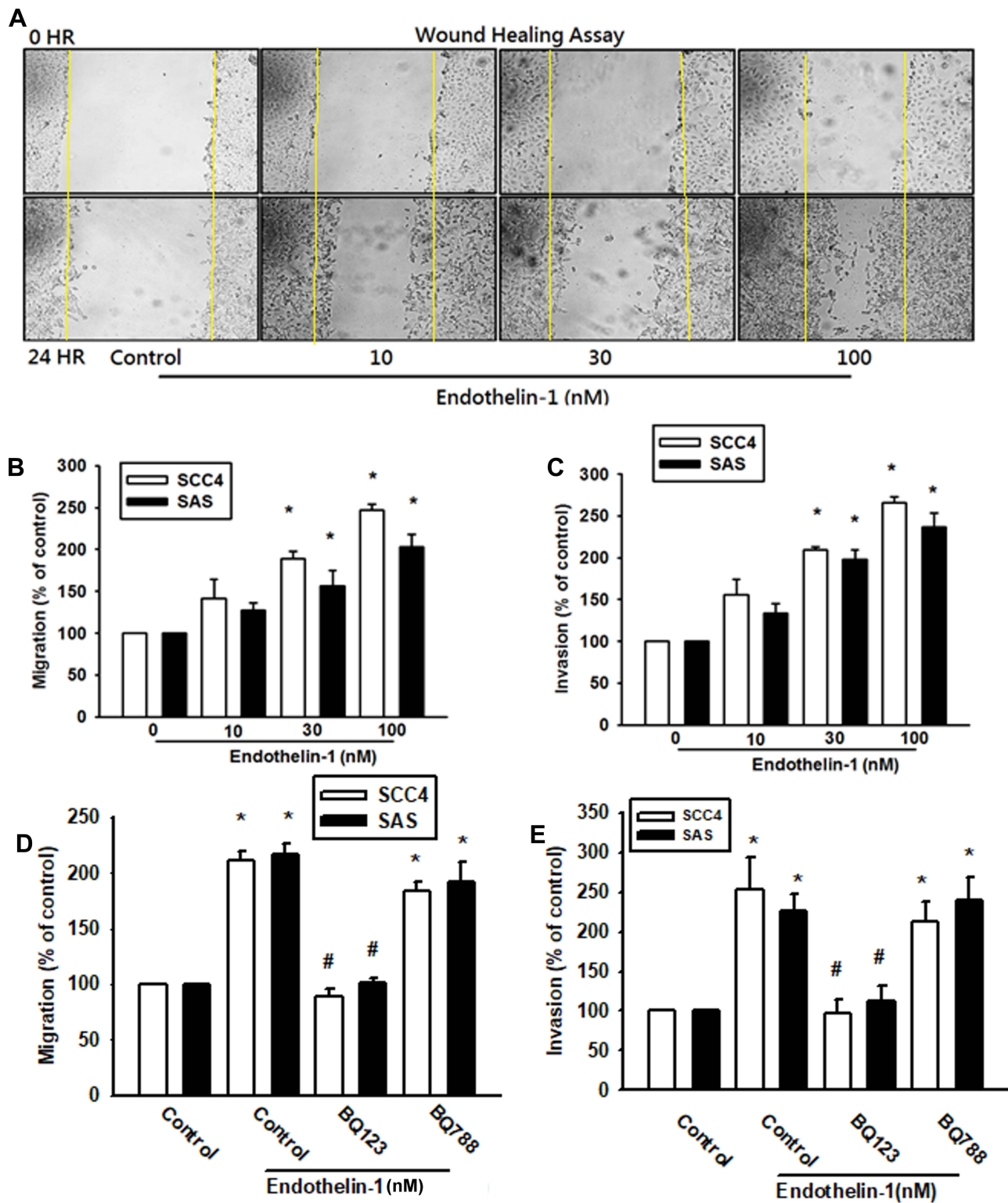
## Statistics

All quantified results were calculated using GraphPad Prism 5.0 (GraphPad Software, Inc; San Diego, CA, USA) or SigmaPlot 10.0 software (Systat Software, Inc; San Jose, CA, USA) and are presented as the mean  $\pm$  standard error of the mean (SEM). Pearson's correlation analyses are presented in clinical data analysis. The Student's *t*-test was used to compare the means of two experimental groups. We performed a one-way ANOVA followed by Bonferroni's post hoc comparison tests on results from statistical comparisons involving more than 2 groups. In all cases,  $p < 0.05$  was considered statistically significant.

## Results

### ET-1 Induced OSCC Cell Migration and Invasion via the ET-A Receptor

Cancer cells gain migratory and invasive properties through active EMT functioning, which occurs during wound healing and in the initiation of cancer metastasis.<sup>26,27</sup> To investigate whether ET-1 was associated with migratory activity in OSCC cells, we examined the migratory ability of the human laryngeal squamous cell carcinoma SCC4 cell line. As shown in [Figure 1A](#), ET-1 enhanced cell migration as demonstrated by the wound healing assay in a concentration-dependent manner. Under the same conditions, ET-1 also enhanced cell migration and invasion of the human SCC4 & SAS OSCC cell lines ([Figure 1B](#) and [C](#)). As ET-1 acts via two distinct subtypes of G protein-coupled receptors, subtype A (ET<sub>A</sub>R) and subtype B (ET<sub>B</sub>R), we hypothesized that ET receptors may be involved in ET-1-induced EMT in



**Figure 1** ET-1 induced OSCC cell migration through ETAR. Cells were treatment by ET-1 (10–100 nM) and cell wound's healing, migration and invasion was monitored (A–C) (n=4–6). Cells were pre-treated with BQ123 or BQ788 for 30 mins, then treated with ET-1 100nM. Cell migration was measured by the Transwell assay and the invasion assay (n=6–8) (D, E). Results are expressed as the mean  $\pm$  S.E.M. \* $p$ <0.05 compared with controls. # $p$ <0.05 compared with ET-1 treatment group.

OSCC. Following pretreatment of OSCC cells with the ET<sub>A</sub>R antagonist BQ123 and the ET<sub>B</sub>R antagonist BQ788, only BQ123 abolished ET-1-induced cell

migration and invasion, indicating that ET-1 acts through the ET<sub>A</sub>R to enhance the EMT process (Figure 1D and E).



## The ETR Signaling Pathway is Involved in ET-1-Induced Expression of EMT Markers

We hypothesized that ET-1 may promote cell migration by increasing the expression of specific EMT marker proteins. Using quantitative polymerase chain reaction (qPCR) analysis, we found that ET-1 increased levels of N-cadherin and vimentin expression, but suppressed E-cadherin expression in SCC4 and SAS cells (Figure 2B-D). Similar results were obtained with proteins levels by Western blot assay of SCC4 cells (Figure 2A). Pretreatment of SCC4 and SAS cells with the ETR antagonist BQ123 abolished ET-1-induced cell migration, invasion and levels of N-cadherin and vimentin expression, but increased E-cadherin levels (Figure 2E-G).

## Transcription Factor TWIST is Required for ET-1-Mediated Expression of EMT Markers in OSCC Cells and Subsequently Elicits Cell Migration

Previous studies have indicated that TWIST is a key transcriptional factor EMT.<sup>28,29</sup> We therefore hypothesized that TWIST may be involved in ET-1-mediated expression of EMT markers in human OSCC cells. Stimulation of SCC4 cells with ET-1 promoted TWIST mRNA and proteins expression (Figure 3A and F). Pretreatment of OSCC cells with the ETR antagonist BQ123 abolished ET-1-induced increases in TWIST mRNA expression (Figure 3B). After they were transfected with TWIST siRNA to suppress Twist1 protein levels (Supplementary data Figure S1). Stimulated with ET-1 the Twist1 proteins levels (Figure 3F), Transwell assay results revealed that ET-1 significantly increased cell migration and invasion, which were dramatically attenuated in the presence of TWIST siRNA (Figure 3C and D). Transfection of cells with specific siRNAs attenuated ET-1-induced expression of N-cadherin and vimentin, but suppressed E-cadherin mRNA levels (Figure 3E). These findings indicate that TWIST transactivation plays a critical role in ET-1-induced EMT marker expression and migration.

## MiR-489-3p is an Important Factor in ET-1-Induced Cell Migration and TWIST Expression

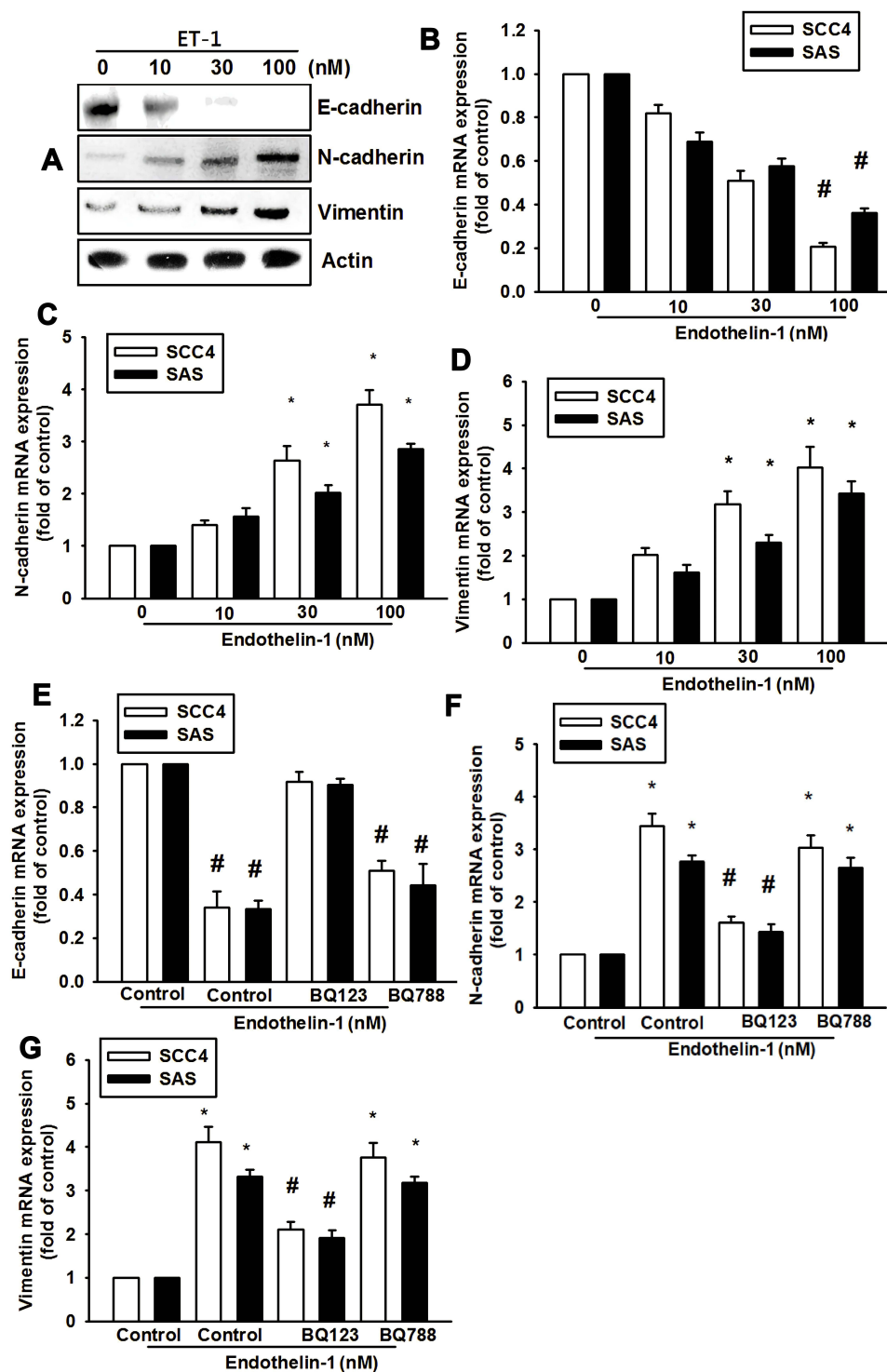
MiRNAs are important regulators of cancer progression and metastasis. Our results indicate that ET-1 promotes cell migration by upregulating TWIST expression. We

therefore undertook a bioinformatics analysis using open-source database software (miRDB and TargetScan) to screen for miRNAs that regulate TWIST expression. We found that the 3'-UTR region of TWIST mRNA harbors potential binding sites for candidate miRNAs and that the most significant downregulation of miR-489-3p occurs after ET-1 stimulation (Figure 4A). Using RT-qPCR, we observed that ET-1 directly reduced miR-489-3p expression in a concentration-dependent manner of SCC4 cells (Figure 4B). To confirm the involvement of miR-489-3p in ET-1-mediated cell migration and invasion, a miR-489-3p mimic was transfected into OSCC cells. Following ET-1 treatment, we found that the miR-489-3p mimic abolished ET-1-induced cell migration and invasion (Figure 4C and D). To further examine whether EMT-related proteins are mediated by ET-1 via the reduction of miR-489-3p expression, we pre-treated cells with miR-489-3p mimic. This abolished ET-1-induced expression of E-cadherin, N-cadherin, vimentin and TWIST EMT-related proteins (Figure 4E and F).

To demonstrate whether miR-489-3p regulates the 3'-UTRs of TWIST, we constructed luciferase reporter vectors harboring the wild-type 3'-UTR of TWIST mRNA (WT-TWIST-2-3'-UTR) and a vector containing mismatches in the predicted miR-489-3p binding site (MUT-TWIST-3'-UTR; Figure 4H). In addition, treatment with ET-1 induced luciferase activity with the WT-TWIST-3'-UTR plasmid, but not with the MUT-TWIST-3'-UTR, indicating that miR-489-3p directly suppresses by ET-1. Furthermore, ET-1 can inhibit the expression of miR489-3p binding WT-TWIST 3'UTR further increasing TWIST mRNA expression. On the contrary, this phenomenon has not been seen in MT-TWIST 3'UTR. This shows that miR489-3p is specific to TWSIT 3'UTR and will be regulated by ET-1 (Figure 4I). These data indicate that ET-1 promotes cell migration via suppresses miR-489-3p and up-regulated Twist-1mRNA level.

## Knockdown of ET-1 Inhibits Metastasis of OSCC Cells in the Lungs of an Animal Model

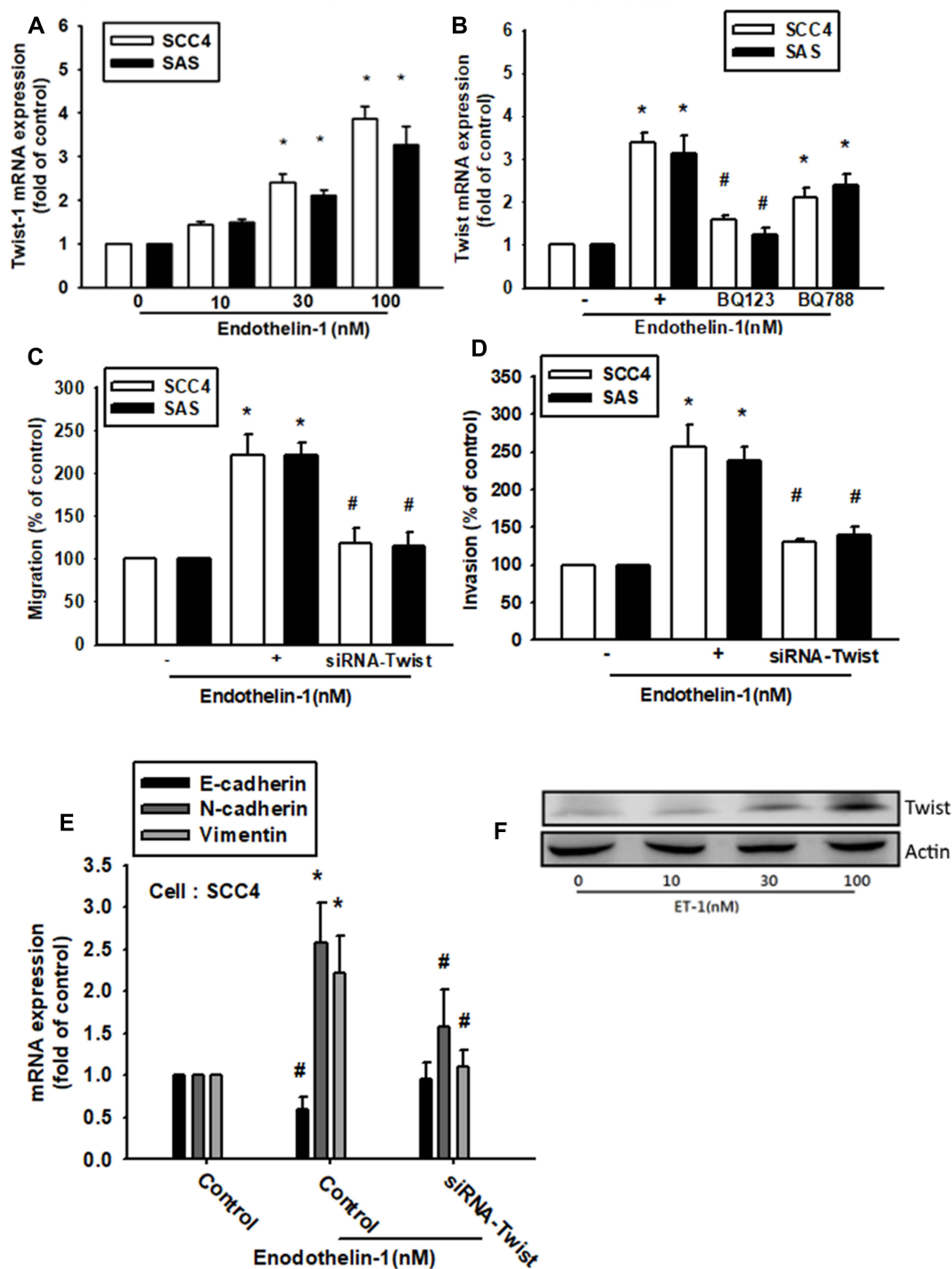
To verify the involvement of ET-1 expression in the migration of OSCC cells, ET-1 expression was knocked down via lentivirus-mediated delivery of ET-1 shRNA. Knockdown efficiency of ET-1 was determined by Western blot and qPCR (Supplementary data Figure S2 and S3). As shown in Figure 5A, silencing of ET-1



**Figure 2** ET-1 promotes EMT through ETAR activation in OSCC cells. ET-1 induces epithelial-to-mesenchymal transition (EMT) functioning via N-cadherin and Vimentin upregulation and E-cadherin downregulation. Cells were incubated with different concentrations of ET-1 (0–100 nM) for 24 h and EMT marker expression was evaluated by qPCR and Western blot assay (A–D) (n=6). Cells were pre-treated by BQ123 or BQ788 in 30mins, and then treated with ET-1 100nM. mRNA expression levels were measured by qPCR assay (E–G) (n=6). Results are expressed as the mean  $\pm$  S.E.M. \*p<0.05 compared with controls. #p<0.05 compared with ET-1 treatment group.

expression reduced ET-1-shRNA SCC4 cell migratory activity and enhanced miRNA489-3p expression (Figure 5B). As shown in Figure 5C and D, ET-1 shRNA

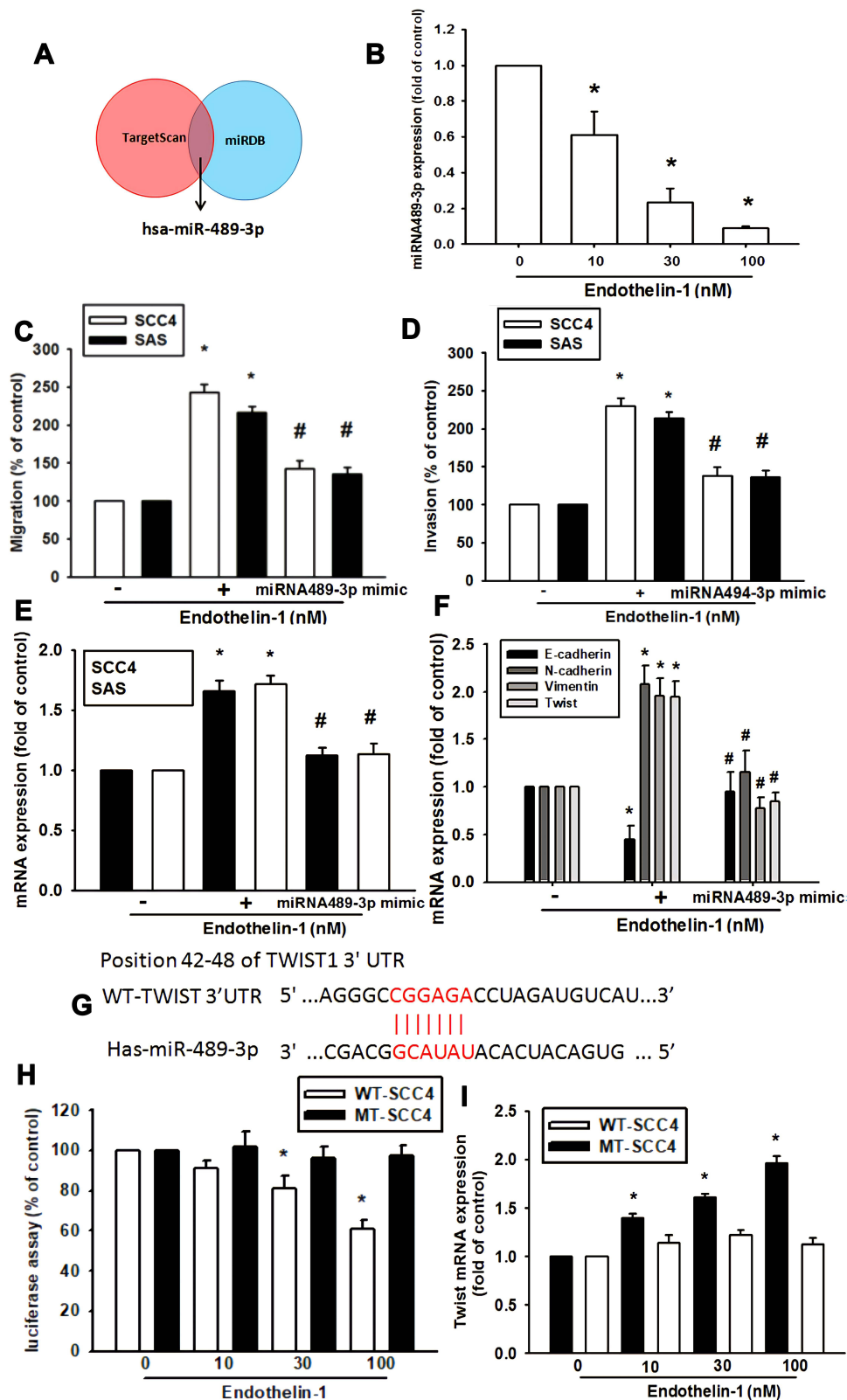
repressed OSCC cells growth via a remarkable reduction of bioluminescence activity. Furthermore, we development of lung metastases in mouse models of OSCC. According



**Figure 3** ET-1 enhances migration and invasion through TWIST activation. Cells were incubated with ET-1 (10–100 nM) for 24 h. TWIST mRNA expression levels were measured by qPCR assay (A). Cells were incubated with BQ123 or BQ788 for 30 min, and then treated with ET-1 100nM for 24 h. TWIST protein expression levels were measured by qPCR assay (B). Cells were transfected with TWIST siRNA for 24 h, followed by stimulation with ET-1 100nM for 24 h (n=4–6). Cell migration was examined by Transwell migration assay and the invasion assay (C, D). EMT-related marker protein levels were examined by qPCR assay (E). Results are expressed as the mean ± S.E.M. \*p<0.05 compared with controls. #p<0.05 compared with the ET-1-treated group.

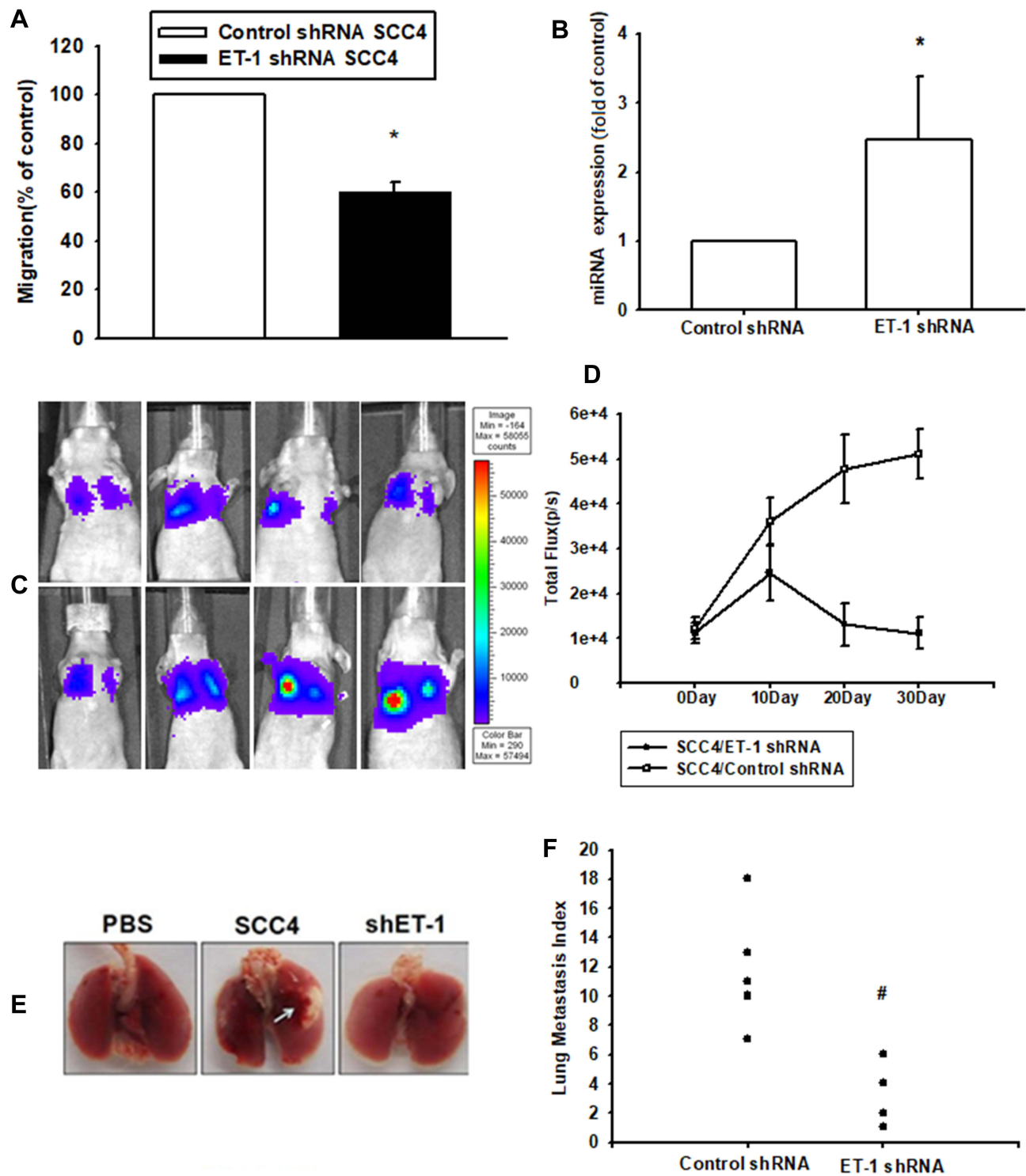
to recent studies, whether it is orthotopic or tail vein metastasis, is suitable for the construction of animal models of oral cancer metastasis. The orthotopic model

presented spontaneous lung metastases in 50% of the animals and the tail vein model, a lung metastasis rate of 60% was observed.<sup>30,31</sup> ET-1 shRNA decreased the



**Figure 4** ET-1 promotes OSCC cell migration and invasion, and EMT marker expression by downregulating miR-489-3p expression. Open-source software used to predict and identify target miRNAs (A). Cells were treated with ET-1 and miR-489-3p expression was examined by q-PCR assay (B). Transfected with a control miRNA or miR-494-3p mimic in OSCC cells. Cell migration (C), invasion (D) and EMT related marker expression (E, F) were examined (n=4-6). Schematic 3'-UTR representation of human TWIST1 containing the miR-489-3p binding site (G). Cells were transfected with a wild-type or mutant TWIST 3'UTR luciferase plasmid, then treated with ET-1, and the relative luciferase activity was determination (H). Twist mRNA expression was measured by qPCR assay (I). Results are expressed as the mean ± S.E.M. \*p<0.05 compared with controls. #p<0.05 compared with the ET-1-treated group.





**Figure 5** Knockdown of ET-1 suppressed lung metastasis and reversed microRNA-489-3p levels in an animal model of OSCC. SCC4 cells stably expressing control- or ET-1-shRNA were established. Cell migration in the SCC4/control-shRNA and SCC4/ET-1-shRNA cells was examined by Transwell assay (A). MiR-489-3p expression in the SCC4/control-shRNA and SCC4/ET-1-shRNA cells was detected by RT-qPCR assay (B). SCC4/Luc or SCC4/shET-1-Luc cells were injected into the lateral tail veins of mice, and the lung metastasis was determined by bioluminescence imaging at the indicated time intervals (C). Quantification of in vivo bioluminescence imaging images (photons/s of lung region) (D). After mice were sacrificed and then lung tissues were excised and photographed (E and F). The results are expressed as the mean ± SEM. \*p<0.05 compared with the JJ012/control-shRNA group. #p<0.05 compared with ET-1 treatment group.

number of lung metastatic nodules of OSCC xenografts (Figure 5E and F). Therefore, our results suggest that ET-1 is involved in OSCC metastasis.

## ET-1 and TWIST Expression Levels Correlate Positively with the Degree of Malignancy in OSCC

We observed strong positive correlations between increased concentrations of ET-1 and TWIST expression with tumor progression in clinical samples of OSCC lung metastases (Figure 6A–C). Pearson's correlation analysis confirmed a significant positive correlation between ET-1 and TWIST expression (Figure 6D). These findings suggest that ET-1 is associated with TWIST expression and tumor metastasis in OSCC.

## Discussion

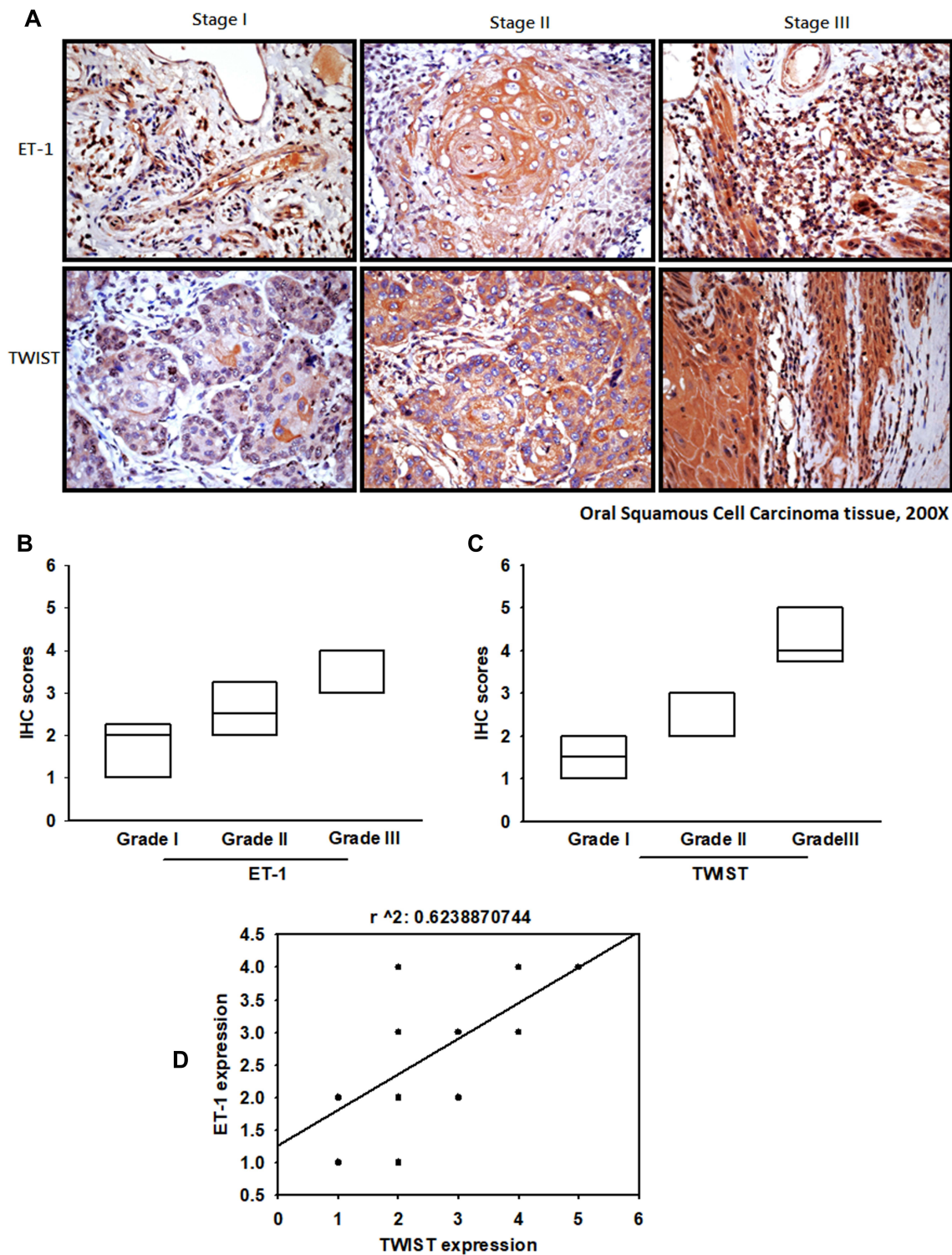
Despite the introduction of targeted treatment protocols combining EGFR monoclonal antibodies<sup>32</sup> and radiotherapy combined with anticancer agents,<sup>33</sup> in the hopes of improving therapeutic outcomes for OSCC, 5-year survival rates are poor and marked by toxicity.<sup>34</sup> Moreover, the rapid division of cells in the mucosal lining makes them highly susceptible to damage from radiotherapy, resulting in serious oral cavity problems and deleterious quality of life outcomes.<sup>35,36</sup>

ET has previously been reported to be directly related to cell metastasis. Recently studies reported that the endothelin receptors ET<sub>A</sub> and ET<sub>B</sub> are highly expressed in head and neck cancer. And they were overexpressed in tumor cells of tongue cancer samples by immunohistochemistry. Previously studies mentioned the survival rates were significantly lower among the patients who were strongly positive for ET-1 and the ETAR-positive patients compared to negative patients. There was also a significant difference between ET/ETAR expression and the degree of histological differentiation and mode of invasion, and the survival rate of the positive cases was significantly lower than that of the negative cases.<sup>37</sup> These reports suggested that ET/ETAR assessments are important for assessing the malignancy of cancer cells and predicting the prognoses of OSCC patients. These results are consistent with our findings. Moreover, our research further indicated that endothelin signalling may, in part, play important roles in miRNA 489–3p/Twist1 in OSCCs through the ET-1/ETAR pathway.

ET-1 plays an important role in several cancers, promoting tumor cell migration and metastasis.<sup>38</sup> Previous study indicated that ET-1 facilitates oncogenesis in human chondrosarcoma by induced angiogenesis or metastasis via increasing VEGF-A, matrix-metalloproteinase (MMP) family, and cyclooxygenase (COX)-2.<sup>39–42</sup> Clinical studies have indicated that the presence of ET-1 immunoreactivity in OSCC specimens enhances the aggressive behavior of poorly-differentiated OSCC, especially metastasis.<sup>43–45</sup> In contrast, while salivary ET-1 levels have been detected in patients with OSCC, the levels do not appear to reflect tumor progression.<sup>46–48</sup> Thus, the biological function and significance of ET-1 expression in malignancy has been controversial. In this study, we hypothesized that ET-1 may drive metastasis of OSCC. Direct administration of exogenous ET-1 promoted cell migration and invasion activity in OSCC cells. Conversely, ET-1-regulated TWIST expression and cell motility were abolished by ET-1 shRNA. These data suggest that ET-1 increases TWIST expression and subsequently promotes cell migration in human OSCC.

Indeed, most research about EMT are related with Snail & Slug pathways but the major purpose of our research is previously reported mentioned that twist1 is a molecular marker for a poor prognosis in oral cancer and represents a potential therapeutic target.<sup>49</sup> This result was corroborated by the clinical observation that Twist1 up-regulation predicted the occurrence of lymph node and lung metastases as well as poor patient survival.<sup>50</sup> In support of Twist1 as a driver of OSCC progression, the up-regulation of Twist1 was observed in cells isolated from patients with metastatic OSCC. Seemly, TWIST1 is a critical role in clinical metastasis; therefore, we further verify whether ET-1 is the main cause of TWIST1 activation.

As shown in previous studies, ET receptors are present on the surface of tumor cells and are responsible for ET-1-mediated cell motility.<sup>51</sup> We found in this study that pretreatment of OSCC with an ET<sub>A</sub>R inhibitor blocked ET-1-induced cell migration and reduced levels of TWIST, N-cadherin and vimentin, but increased E-cadherin expression. The ET-1/ET<sub>A</sub>R axis interaction therefore mediated EMT activity and promoted cell migration in OSCC. MiRNAs control gene expression by binding to complementary sequences in the 3'-UTRs of target mRNAs. Deregulated miRNA expression has been noted in human cancers and may affect multiple steps during



**Figure 6** OSCC specimens indicate the significant correlations with ET-1, TWIST and tumor progression. OSCC specimens were stained with ET-1 and TWIST antibodies for IHC analysis (A). IHC stain intensities quantify expression levels (B, C). IHC staining scores were paired from the same specimens and the correlation between ET-1 and TWIST expression was shown by linear regression in OSCC specimens (D).

metastasis. In particular, the following miRNAs can regulate metastatic ability in OSCC: miR-146a, miR-191, RNA-1297, miR-155, miR-221 and miR-31-5p.<sup>52-55</sup> Our study indicates that miR-489-3p is downregulated in

response to ET-1; miR-489-3p reportedly suppresses tumor formation in prostate cancer, making it an attractive candidate biomarker for predicting responses to cancer treatment.<sup>56-58</sup>



In this study, transfection of cells with a miR-489-3p mimic reduced ET-1-induced cell migration, suggesting that miR-489-3p can function as a tumor suppressor. Our tumor metastasis animal model provides strong support that ET-1 is associated with higher incidence of metastasis. Here, we found that decreased ET-1 levels significantly downregulated OSCC cell migration and upregulated miRNA-489-3p expression. These changes affect metastatic ability in OSCC. In the present study, staining of clinical tissue specimens showed that levels of ET-1 and TWIST expression correlate with tumor development, which is consistent with previous clinical studies. These in vivo findings suggest that ET-1 regulates TWIST and OSCC metastasis through ET-1/ET<sub>A</sub>R/ miRNA-489-3p mechanisms.

## Conclusions

In conclusion, existing therapeutic options for OSCC are characterized by poor survival outcomes and significant toxicity. Novel therapeutic targets are needed. Our study findings provide novel insights into the role of ET-1 in OSCC metastasis through ETRs/miR489-3p/Twist pathways ([Supplementary data Figure S4](#)) and indicate that ET-1 may be a novel therapeutic target.

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## Author Contributions

All authors contributed to data analysis, drafting or revising the article, have agreed on the journal to which the article will be submitted, gave final approval of the version to be published, and agree to be accountable for all aspects of the work. Huey-En Tzeng and Chih-Hsin Tang contributed equally to this work and are co-first authors. Min-Huan Wu and Yun Yen contributed equally to this work and are co-corresponding authors.

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## Disclosure

The authors report no conflicts of interest for this work.

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