

Brown adipose tissue in the treatment of obesity and diabetes: Are we hot enough?

Chong Yew Tan*, Ko Ishikawa, Samuel Virtue, Antonio Vidal-Puig

ABSTRACT

The identification of functional brown adipose tissue in human adults has intensified interest in exploiting thermogenic energy expenditure for the purpose of weight management. However, food intake and energy expenditure are tightly regulated and it is generally accepted that variation in one component results in compensatory changes in the other. In the context of weight loss, additional biological adaptations occur in an attempt to further limit weight loss. In the present review, we discuss the relationship between increasing energy expenditure and body weight in humans, including the effects of cold exposure. The data raise the possibility that some processes, particularly those involved in thermogenesis, induce less compensatory food intake for a given magnitude of additional energy expenditure, a state we term the 'thermogenic disconnect'. Although cold exposure increases thermogenesis and can putatively be exploited to induce weight loss, there are multiple adaptive responses to cold, of which many actually reduce energy expenditure. In order to optimally exploit either cold itself or agents that mimic cold for thermogenic energy expenditure, these non-thermogenic cold responses must be considered. Finally, the relative contribution of brown adipose tissue vs other thermogenic processes in humans remains to be defined. However, overall the data suggest that activation of cold-induced thermogenic processes are promising targets for interventions to treat obesity and its secondary metabolic complications. (*J Diabetes Invest*, doi: 10.1111/j.2040-1124.2011.00158.x, 2011)

KEY WORDS: Brown adipose tissue, Energy expenditure, Weight loss

INTRODUCTION

To date, therapeutic interventions aimed at reducing caloric intake have not been successful. Caloric restriction in combination with exercise is effective in the short term, but long-term weight maintenance is poor and weight regain, sometimes rising above pre weight loss levels, is common^{1–3}. Modestly effective pharmacological interventions, such as sibutramine and rimonabant, have both been withdrawn from the market owing to side effects. Only orlistat, a gastrointestinal lipase inhibitor, remains available; however, it is of limited efficacy and causes unpleasant side effects. An alternative strategy is to increase energy expenditure (EE) as a means to prevent weight gain and to induce weight loss.

Weight reduction through caloric restrictions results in a series of responses that minimize weight loss. Rosenbaum *et al.*^{4,5} demonstrated that weight loss through caloric restriction reduces total daily EE (TDEE) by 25% beyond that predicted by post-weight loss fat-free mass. This is accounted for, in part, by increased efficiency of skeletal muscle metabolism, altered hypothalamic–pituitary–thyroid axis, and lowered leptin levels. In addition, hunger and satiety scores after weight loss continue to favor caloric intake beyond daily needs. These changes are

sustained over a follow-up period of 1–2 years. Conversely, caloric excess promotes a largely opposite response to caloric reduction, although the response is short lived⁶. This ability to defend body energy stores as fat was termed the 'adipostat hypotheses'.

Based on the adipostat hypothesis, increasing EE to promote weight loss should be hindered by opposing responses, particularly an increase in food intake. Ideally, any strategy to increase EE should be achieved with only a small, or preferably no, reliance on concomitant caloric restriction, thus improving long-term compliance. More recently, Cannon and Nedergaard⁷ reviewed murine models examining the role of cold-induced thermogenesis (CIT) in weight control and concluded that increasing EE, particularly by utilizing brown adipose tissue (BAT), does not result in caloric intake that matches the elevated metabolic rate. In the present review, we will term the concept that, in the absence of caloric restriction, energy loss through thermogenesis is not completely matched by an increase in food intake as the 'thermogenic disconnect'.

Recently, BAT has gained renewed interest as a target for increasing EE as a means to treat obesity⁷. A putative therapeutic role for BAT was first postulated over three decades ago. Rothwell and Stock suggested that resistance to diet-induced obesity (DIO) in rats was a result of increased EE in response to a cafeteria diet⁸. This increase in EE was later shown to be mediated by BAT and termed 'diet-induced thermogenesis'⁹. Rothwell and Stock postulated that rats resistant to DIO were able to

Metabolic Research Laboratories, Addenbrooke's Hospital, Cambridge, UK

*Corresponding author. Chong Yew Tan Tel: +44-1223-768-629

Fax: +44-1223-330-598 E-mail address: cyt23@medschl.cam.ac.uk

Received 28 June 2011; revised 11 July 2011; accepted 12 July 2011

evoke an apparently wasteful increase in EE as a protective mechanism against an inadvertent increase in food intake.

The discovery that BAT is present in energetically significant amounts in adult humans makes this tissue a potential therapeutic target against obesity^{10–12}. In the present review, we provide a brief summary of the relationship between increasing EE and weight loss. Because there are no data regarding specific BAT EE in humans, we will review data regarding the effects of cold exposure, a state known to be the most potent physiological activator of BAT in rodents and humans, on EE and body weight. Finally, using the cold response as a model, we will review the potential issues involved in exploiting CIT.

INCREASING ENERGY EXPENDITURE FOR WEIGHT LOSS

Obesity is a result of chronic caloric imbalance. On a population level, this net imbalance is thought to be small, resulting in gradual but progressive weight gain amounting to approximately 1 kg/year in Western populations^{13–15}. A weight gain of 1 kg/year equates to only 105 kJ (approximately one teaspoon of sugar) per day. However as weight (both fat and fat-free mass) is gained, caloric intake would necessarily rise to maintain energy balance. A larger organism requires more energy, due to both an increase in cell numbers and an increase in mechanical work. Bouchard¹⁶ argue that when an individual gains weight from a body mass index (BMI) of 25 to 35 kg/m², his/her intake would have to increase by 2.7 MJ/day compared with his/her leaner self to maintain energy balance. That is, a net 105 kJ/day positive balance will result in a gain of 3.5 kg after 3.5 years. After factoring the effects of age (of 3.5 years), TDEE will rise by 105 kJ/day, effectively balancing the excess caloric intake. In order to explain the sustained rise in body weight, caloric intake would have to increase not only to match the added expenditure, but also to maintain the net positive balance. Therefore, the magnitude of caloric restriction required for an obese person to lose weight would be much larger than 105 kJ/day¹⁷.

Conversely, the reasons for increasing EE to promote weight loss go beyond simple thermodynamics. Individuals who have a lower than predicted weight-adjusted resting metabolic rate (RMR) are more likely to become obese^{18,19}. Obese individuals who have lost weight also have a lower than predicted TDEE compared with their pre-weight loss state that cannot be explained by the loss of fat-free mass alone. Therefore, increasing EE would be expected to counter these biological effects.

RELATIONSHIP BETWEEN THE MAGNITUDE OF ENERGY EXPENDED AND DEGREE OF WEIGHT LOSS

Energy Expenditure and Weight Loss Through Exercise

Obesity is thought to be driven, in part, by the modern sedentary life style, although this remains a matter for debate²⁰. Evidence for a decrease in physical activity was noted in epidemiological studies, cross-sectional studies, and from data demonstrating an increase in sedentary behavior, such as rising

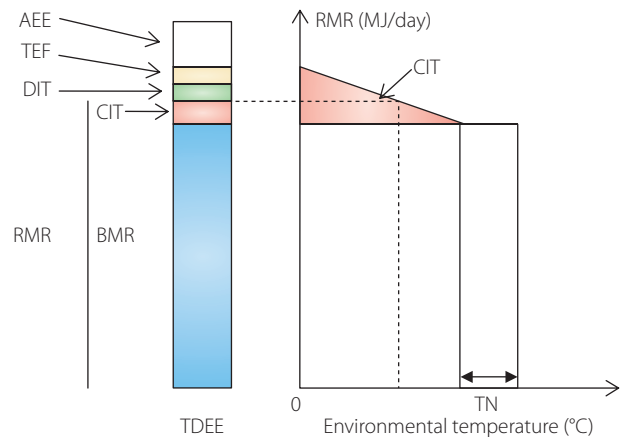


Figure 1 | Components contributing to total daily energy expenditure and the role of cold-induced thermogenesis (CIT). The bar graph on the left depicts the relative contribution of various components to total daily energy expenditure (TDEE), namely activity energy expenditure (AEE), thermal energy of feeding (TEF), diet-induced thermogenesis (DIT), CIT, and basal metabolic rate (BMR). The BMR is measured in a lightly clothed subject at rest in a thermoneutral environment. Below thermoneutrality, CIT is evoked to defend core body temperature. In this situation, the measured energy expenditure at rest is termed the resting metabolic rate (RMR). The graph on the right shows the magnitude of the RMR for a given environmental temperature. At thermoneutral temperatures (TN), RMR = BMR. At cooler temperature, CIT (shaded area) is evoked. The magnitude of CIT is determined by the severity of cold exposure (dotted line).

car sales and television watching. Therefore, promoting physical activity was postulated as an effective treatment for obesity.

To determine how much energy is required to induce weight loss through exercise, it is important to understand that measurements are influenced by the method of quantification (free living vs laboratory measurements). In addition, energy balance in the short term is highly variable, but remains remarkably well balanced, particularly in weight-stable lean individuals, over the mid to long term^{21,22}. This is of significant importance when extrapolating short-term energy balance data to long-term weight loss. The use of doubly labeled water to measure average daily EE (ADEE) coupled with calorimetry to assess resting metabolic rate (RMR) is generally accepted as the gold standard for determining free living activity EE (AEE). In this method, thermal energy of feeding and diet-induced thermogenesis are assumed to account for 10% of ADEE; hence, ADEE = 10% of ADEE + RMR + AEE (Figure 1).

In the context of weight loss, there appears to be a critical threshold above which increases in AEE are not matched by an increase in caloric intake²³. Using Physical Activity Level (PAL; whereby PAL = ADEE/RMR) as a marker of AEE, short- to mid-term measurements from the general population and among soldiers in training indicate that the average sustainable PAL score falls between 1.4 and 2.5²⁴. At a PAL score above 2.4, significant weight loss occurs. Although food intake

increases with increased PAL, it is thought that consumption and/or absorption of nutrients becomes limiting. This is supported by data from elite athletes, in whom PAL scores >2.4 without weight loss are sustainable only through the consumption of energy-dense supplements²⁵.

If we take an obese sedentary individual (PAL 1.4) with an RMR of 7.5 MJ/day, this individual will have an estimated ADEE of 10.5 MJ/day. To achieve sustained weight loss through exercise, an increase of ADEE to 18.1 MJ/day (achieving a PAL score of 2.4), equivalent to an increase of 7.6 MJ/day, is required. Four hours of moderate physical activity at four MET (where a MET, or metabolic equivalent, is the ratio of work metabolic rate to resting metabolic rate; by definition, one MET is equivalent to 1 kcal/kg/h or 4.185 kJ/kg/h) expends 4.0 MJ of energy. Energy expenditures below this is expected to induce some initial weight loss followed by a new and lower steady state weight.

Not surprisingly, exercise alone has achieved limited success in weight loss studies. Numerous studies comparing the effects of diet and exercise, diet alone, or exercise alone indicate that weight loss induced by moderate exercise alone is modest. A meta-analysis of 28 studies²⁶ suggested that exercise alone results in weight loss of 3 kg after 30 weeks intervention in men and 1.4 kg after 12 weeks intervention in women. A subsequent meta-analysis of weight loss trials following the publication of the 1998 National Institutes of Health Obesity Report²⁷, demonstrated that prescribed exercise intervention for a minimum of 1 year results in an average weight loss of approximately 2 kg at 6 months, followed by a gradual rise thereafter³.

In terms of energy expended, most exercise intervention studies achieve an increase of between 1.2 and 2.8 MJ/day²⁸. A 16-month study by Donnelly *et al.*¹⁵ showed that despite a sustained increase of 1.6 MJ/day throughout the study, the net result was a 5.2 kg weight loss that was achieved within the first 9 months of the study. In addition, female subjects did not lose weight despite a sustained increase of 875 kJ/day over 16 months. In that study, ADEE was measured with doubly labeled water and exercise was performed under strict supervision. Although significantly larger than the imbalance (approximately 105 kJ/day) thought to cause weight gain, this increase in EE still falls below the PAL score of 2.4 required to overcome compensatory increases in food intake.

In contrast, exercise has been shown to be effective in weight maintenance and this can be achieved at PAL scores well below 2.4. The precise amount of exercise required is still debatable, but it is expected to be higher than that recommended for promoting cardiovascular health. Energy expenditure of 8.4 MJ/week (1.2–1.3 MJ/day) may be sufficient for the purpose of weight maintenance²⁹. Although the magnitude expended is 10-fold >105 kJ/day, the food intake is perfectly matched with EE and so weight remains stable.

In summary, exercise is an effective means of increasing EE. Despite this, the expected weight loss is not observed (even when compliance is maximized). Exercise-induced EE expenditure is

fully compensated for by an increase in food intake up to a PAL score of 2.4. However, exercise can aid weight maintenance at a much lower PAL when combined with caloric control.

Energy Expenditure and Weight Loss in Hyperthyroidism

Individuals with hypothyroidism on average gain approximately 15% of their body weight^{30,31}. Not surprisingly, it was once thought that obesity may be a result of thyroid hypofunction. However, the majority of obese individuals have normal thyroid function^{32,33}. Although there is debate regarding the possibility of subtle thyroid dysfunction in obesity^{34–36}, it is unlikely that it can account for the obesity epidemic. Individuals with hyperthyroidism lose, on average, 15% of their pre-morbid weight. In cohorts of untreated hyperthyroid subjects, an increase in RMR of approximately 30–47% was observed compared with weight-matched controls^{37,38}. Considering a median delay of 4 months between the onset of symptoms and diagnosis³⁶, an 80-kg woman with a pre-morbid RMR of 6.7 MJ/day will expend an additional 1.9–3.1 MJ/day once she becomes hyperthyroid. After 4 months, this equates to 6.4–10 kg of fat mass loss, which is equivalent to weight loss of 8–12.5%.

Additional interest in the thyroid status in obese individuals comes from the observation that EE falls during caloric restriction and weight loss. This fall is greater than that predicted for weight and is accompanied by a fall in tri-iodothyronine (T₃) and a rise in reverse (r) T₃^{39–41}. This fall in EE would rapidly balance the caloric restriction, resulting in the stabilization of weight. As such, a number of studies have looked at the effects of supplementing T₃ during caloric restriction^{42–45}. Supplementation with T₃ was expected to allow sustained weight loss by preventing the hypometabolic effects of caloric restriction. In one study in which RMR was measured⁴⁵, supplementation, following initial caloric restriction, for 3 months with T₃ (20 µg, 8 hourly) resulted in an approximate 35% higher adjusted RMR compared with placebo controls. Given that the reported average subject weight of 100 kg and that control subjects dropped their RMR by 27%, the expected post-weight loss RMR would be 5.8 MJ/day in the control group. A 35% higher RMR (2.0 MJ/day) in the treated group would result in an additional 4.4 kg weight loss, which compares well with the reported 5.5 kg weight loss⁴⁵. Other shorter-term studies using higher doses of T₃ produced proportionally higher amounts of weight loss^{42,46}.

Hyperthyroidism is associated with an increase in food intake and yet, despite this, weight loss occurs. The weight loss in response to T₃ described above is mostly accounted for by increased RMR, implying that any additional calories consumed are either inefficiently absorbed and/or energy is expended through thermal energy of feeding and AEE. Crucially, and in contrast with what is observed with exercise (typically 1.2–2.8 MJ/day to lose approximately 2 kg after 6 months), increases in EE due to elevated thyroid hormones (1.9–3.1 MJ/day to lose 6.4–10 kg after 4 months) translate to weight loss and are not fully compensated for by food intake.

Despite decades of research, the relative contribution of various mechanisms responsible for thyroid hormone-mediated EE has not been completely worked out. Various processes have been postulated (for a review, see Silva⁴⁷). In general, increased fuel consumption via oxidative metabolism can only occur under states of increased ATP utilization or a reduced bioenergetic efficiency of ATP synthesis^{47,48}. In this regard, thyroid hormone is believed to increase ATP utilization via futile cycling of substrates (glycolysis–gluconeogenesis, lipolysis–lipogenesis, protein synthesis–proteolysis) and ions (Na⁺–K⁺ cycling, skeletal muscle Ca²⁺ cycling), with heat, rather than useful work, the ultimate energetic product. In addition, increased thyroid-mediated sympathetic nervous system (SNS) tone may contribute to higher non-voluntary skeletal muscle activity. In rodents, thyroid hormones have been found to be crucial for the activation of BAT and for cold acclimation^{49,50}. Cold-induced SNS activation of BAT increases the expression of deiodinase 2 (DIO2), which raises the local levels of T₃. Together, T₃ and β -adrenergic receptor stimulation promote the expression of uncoupling protein 1 (UCP1), resulting in mitochondrial uncoupling and diminished efficiency of ATP synthesis⁴⁹. Mice deficient in DIO2 are cold intolerant⁵⁰. Whether BAT uncoupling contributes significantly to EE in adult humans is unknown.

In summary, excess thyroid hormone is associated with an increase in EE. The magnitude of increase is similar to that achieved through exercise. Unlike exercise, much of the increase in EE is translatable to weight loss. Thyroid hormone regulates metabolism through a number of possible mechanisms, including an increase in ATP-utilizing processes and, in rodents, an increase in UCP1 expression in BAT. The incomplete compensation in terms of food intake in hyperthyroid states suggests the possibility of a thermogenic disconnect in humans. Why this should be applicable to hyperthyroid states is uncertain. It is possible that: (i) following hyperthyroid weight loss, the orexogenic signal of low leptin levels is dampened by hyperthyroid states; (ii) the EE through involuntary hypermetabolic processes may not be appropriately sensed; and (iii) the induction of low-grade EE, sustained throughout the day, may have a qualitatively different effect on appetite compared with bouts of EE from exercise.

Energy Expenditure and Weight Loss with Chemical Uncouplers

Studies using isolated mitochondria demonstrate that respiration is inhibited when ADP is depleted⁵¹. Depletion of ADP removes the substrate for ATP synthase and results in a rise of the inner mitochondrial membrane proton gradient to a point where the electron transport chain is inhibited. However, respiration can recommence in the absence of ADP if the mitochondrial membrane is disrupted and the proton gradient released, thus uncoupling oxidative metabolism from ATP synthesis. In BAT, the expression of UCP1 serves as a regulated uncoupler allowing an increase in biochemical heat production without reliance on ADP or build up of surplus ATP. The heat generated contributes to regulation of core body temperature.

In the early 1930s, dinitrophenol (DNP), a chemical uncoupler, was introduced as a means of increasing metabolism for the purpose of weight loss^{52–54}. It was found to be highly effective at increasing metabolic rate and resulted predominantly in a loss of fat with a conservation of muscle mass. However, patients exposed to DNP were later found to develop cataracts months after completing treatment^{53,55}, resulting in the eventual withdrawal of DNP from the market. In one early study, DNP was administered to 170 subjects without caloric restriction. Five subjects did not lose weight, but the remainder lost an average of 7.8 kg over an average period of 88 days. The average dose used in that study was 340 mg/day, resulting in a 33% rise in RMR⁵⁶. If we take an obese male with an RMR of 7.5 MJ/day, a 33% rise in RMR over 88 days (2.5 MJ/day) would result in a predicted weight loss of 5.2 kg.

The relationship between an increase in energy expended and net weight loss due to DNP is surprisingly similar to that in hyperthyroid states. The predicted amount of weight loss underestimates the observed weight loss, despite the absence of caloric restriction. No food intake data are available, but it would appear that intake does not increase sufficiently to compensate and may even fall. Like hyperthyroidism, chemical uncouplers induce a state of sustained low-grade hypermetabolism and use involuntary biochemical processes. The only difference is that the mode of action of DNP is through mitochondrial uncoupling, whereas hyperthyroidism use a number of biochemical processes. However, if BAT and regulated uncoupling are shown to be functionally relevant in humans, this may point to a common mechanism that may result in a state of thermogenic disconnect.

Energy Expenditure and Weight Loss with BAT Activation

There are no data regarding the magnitude of EE inducible in humans through BAT activation. Virtanen *et al.*¹¹ compared fluorodeoxyglucose uptake in five healthy individuals following 2 h exposure to approximately 18°C compared with no prior cold exposure. Using data from one subject, the authors estimated the supraclavicular BAT depot to have a mass of 63 g. Assuming that glucose accounts for 10% of substrate utilized, with the remainder from fat, the authors estimated that sustained activation of 63 g BAT will metabolize an equivalent of 4.1 kg fat over 1 year. That is, the estimated increase in EE due to BAT activation is approximately 418 kJ/day (equivalent to a 4.5% increase in TDEE for an average male with 9.2 MJ TDEE).

Although it would be wrong to assume that cold-induced increases in EE are equivalent to BAT thermogenesis, it is reasonable to assume that BAT thermogenesis will fall within the range observed in cold exposure studies, given that cold is the most potent physiological activator of BAT. Interestingly, the estimated increase in EE that could be attributed to BAT (as calculated by Virtanen *et al.*¹¹) falls within the range of increases observed in calorimetric studies under similar cold stimulation conditions. Increase in TDEE of 2% (22°C vs 16°C for 48 h)⁵⁷, 5% (22°C vs 16°C for 84 h)⁵⁸, 6% (24°C vs 19°C for 12 h)⁵⁹,

7% (28°C vs 22°C for 30 h)⁶⁰, 7–11% (22°C vs 15°C for 3 h)⁶¹, and 17.8% (27°C vs 19°C for 1.7 h)⁶² have been described. Provided that the estimates of Virtanen *et al.* are accurate, it would appear that cold-induced BAT activation could contribute significantly to adult human thermogenesis.

Whether these increases are sufficient to alter body weight remains uncertain. Compared with the values known to induce weight loss through exercise, hyperthyroidism, or DNP (as described above), mild cold-induced increases in EE would appear insufficient. Conversely, if cold exposure can induce a state of thermogenic disconnect, weight loss can potentially occur with this degree of increased EE. In rodents, prolonged cold exposure leads to an increase in BAT tissue mass and an increase in maximal thermogenic capacity⁴⁹. Whether CIT could be augmented in adult humans following repeated or prolonged cold exposure needs to be examined.

Indigenous populations living in desert environments are exposed to extreme fluctuations in temperature. During the night, temperatures can fall below 5°C. In addition, shelter and clothing are often minimal. As such, it can be considered a natural form of repeated cold exposure in humans. As opposed to data observed in rodents, studies of Aborigines living in the deserts of Australia⁶³ and Bushmen living in the Kalahari Desert of southern Africa⁶⁴ reveal that repeated nocturnal cold exposure does not increase EE; rather, core body temperature and RMR are observed to fall in the cold. Such a response would be of benefit for short-term energy conservation with the expectation that the prevailing thermal stress will be transient. In keeping with this, Leppaluoto *et al.* have published extensively^{65–75} on the hormonal changes that occur following repeated cold exposure. They observed that the endocrine (rise in thyroid hormone and adrenergic response) and metabolic (rise in free fatty acids) changes in a standardized cold stimulation tend to fall following repeated cold exposure. Such a response is termed hypothermic cold adaptation. The defended core body temperature is set lower (by approximately 0.5°C) and the core temperature that triggers shivering thermogenesis is lowered by a similar amount.

Despite hypothermic adaptation, measurements of EE following repeated cold exposure are much more encouraging. Bittel⁷⁶ exposed individuals to repeated cold water immersion and demonstrated the effects of hypothermic adaptation as described above. However, EE induced by acute cold stimulation did not diminish despite this. Furthermore, in some individuals, an additional 10% increase in EE was demonstrated upon a standardized cold exposure compared with the pre-acclimated state. This augmentation in EE despite cold adaptation is further supported by other studies⁷⁷.

The increase in EE is much more consistent following sustained cold exposure. A recent study looked at the effects of cold acclimatization following the winter months and compared this with the summer⁶¹. Individuals were exposed to a standardized mild cold exposure of 15°C for 3 h. In summer, the increase in RMR was 7% and in winter it was 11.5%, representing a 64% increase in metabolic capacity following winter acclimatization.

Data from longer-term cold exposure (>6 months) necessarily comes from colder environments at latitudes closer to the poles^{78,79}. Although these data will be less controlled, the consistency of the findings suggests that long-term cold exposure does increase thermogenic capacity in terms of responses to an acute cold stimulation compared with pre-acclimatization. More importantly, the magnitude of the increase appears higher than that inducible through repeated cold exposure. The effect of long-term cold exposure in soldiers stationed at different latitudes was published by Johnson and Kark⁷⁸. The authors collected estimates of food intake and demonstrated a progressive increase in total caloric requirements at higher latitudes and cooler environmental temperatures. Leonard *et al.*⁷⁹ published a meta-analysis of data from studies of Indigenous and non-Indigenous population living at high latitudes. Both Indigenous and non-Indigenous populations showed a higher RMR than that predicted for their age, height, weight, gender, and surface area. Non-Indigenous male subjects had a consistently higher RMR of between 10 and 19% compared with that predicted for fat-free mass. This supports the possibility that, in humans, cold-induced EE can be augmented following cold acclimatization.

Whether this increase in EE translates to weight change is of more relevance. To date, no controlled data regarding the effects of cold exposure on weight loss are available. Bergmann's rule states that at lower ambient temperature, a reduction in the surface area to volume ratio will reduce heat loss and be of survival benefit⁸⁰. Numerous studies have shown that this may be applicable to humans because there is a trend for increased body weight in populations living at higher latitudes^{81,82}. Other studies have looked at the effects on weight during extended polar expeditions. In general, these studies conclude that food intake^{83,84} and EE^{85–87} both increase in the cold, with significant fluctuations between the Antarctic/Arctic winter and summer months. More importantly, most individuals gain a small amount of weight over the period of their stay, which ranges from a few weeks to as much as 3 years⁸⁸. This would suggest that the higher EE from cold exposure is not associated with weight loss. However, these studies involve extreme climates and it may be possible that less extreme exposures applied in a regulated manner may have an impact on body weight regulation.

In summary, cold exposure induces an increase in EE. This increase is small relative to amounts known to achieve weight loss under conditions of exercise, DNP treatment, or hyperthyroidism. However, the increase in EE can be augmented with repeated or sustained cold exposure. Although long-term cold exposure is associated with increased EE, it is not associated with lower body weight.

RESPONSE TO COLD IN HUMANS AND POTENTIAL BARRIERS TO EXPLOITING CIT FOR WEIGHT MANAGEMENT

Homeotherms can maintain body temperature at a consistently stable level despite fluctuating environmental temperatures.

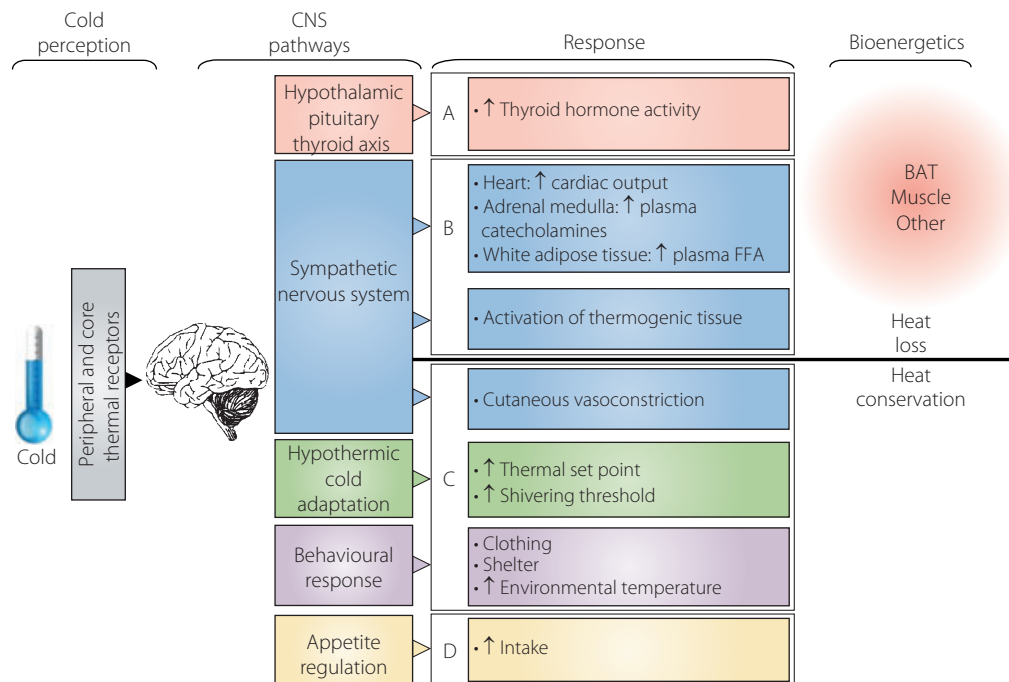


Figure 2 | Physiological responses to cold exposure. From left to right, cold stimuli are sensed by peripheral and central thermal receptors. Within the central nervous system (CNS), neurobehavioral and endocrine responses are activated to defend core body temperature, including the hypothalamic–pituitary–thyroid axis, the sympathetic nervous system, behavioral responses, hypothermic cold adaptation, and appetite regulation. In turn, these produce physiological responses that support thermogenesis (A), activate thermogenic effector tissues (B), conserve heat (C) and increase caloric intake (D). For maximal thermogenic induced weight loss, Processes A and B should be optimal, whereas Processes C and D should be minimized. BAT, brown adipose tissue; FFA, free fatty acids.

Thermoneutrality is defined as a narrow range of temperature whereby additional EE above obligatory thermogenesis is not required for thermal homeostasis (Figure 1). In a lightly clothed human, thermoneutrality lies between 23 and 26°C^{89,90}. Temperatures below thermoneutrality will elicit a series of mechanisms to defend core body temperature.

In humans, behavioral and physiological processes both contribute to thermal defense. In humans, behavioral responses predominate, such as seeking shelter; increasing clothing levels, and, perhaps most commonly in modern life, increasing our environmental temperature. Physiological responses can be separated into three major forms: (i) hypothermic (including habituation); (ii) insulative (vasoconstriction and piloerection); and (iii) metabolic (facultative thermogenesis or CIT). The latter is the only response that involves purposeful heat production. In addition, CIT can be further divided into non-shivering thermogenesis and shivering thermogenesis. Pertinent to weight management, activation of molecular processes involved in non-shivering thermogenesis represents the most promising (and comfortable) approach (Figure 2).

The relative contribution of each physiological response is dependent on interindividual differences^{23,91}, the severity of cold exposure⁶⁷, the rate of temperature change⁹², and prior acclimatization⁵⁸. The neurocircuitry regulating the balance of these three physiological responses is unclear. Much of the work has

been done on rodents and is beyond the scope of the present review (for recent reviews, see Whittle *et al.*⁹³ and Morrison *et al.*⁹⁴). Briefly, environmental temperature is sensed by specific cold receptors (belonging to a class of the transient receptor potential [TRP] family) located in the periphery. Together with central core temperature sensors, thermal signals are integrated within the central nervous system (CNS). Within the CNS, various pathways are thought to modulate food intake, the hypothalamic–pituitary–thyroid axis, and SNS tone to the adrenals, heart, skeletal muscle, cutaneous blood vessels, white, and brown adipose tissue. The resultant rise in core temperature forms a feedback that reduces the stimuli on core thermal receptors. If core or environmental temperatures rise above optimum, heat is sensed by separate warm receptors and their activation effectively terminates the cold response. Any further rise in core temperature will trigger vasodilatation and sweating. Any attempts at exploiting CIT for weight loss will require a specific cold signal to activate thermogenic processes, minimize insulative and hypothermic adaptations, and maximize heat loss (Figure 2).

The molecular mechanism underlying heat production during CIT in humans is not completely understood. In cold-adapted murine models, CIT is largely determined by BAT activity. Little is known about the role of BAT in humans, but the factors required for optimal BAT function in rodents should be applicable to humans. Following cold adaptation in mice, there is

hypertrophy and an increase in blood flow in BAT. Cardiac output is increased, with concomitant cardiac hypertrophy such that up to 60% of output is delivered to the BAT⁹⁵. The SNS tone to the BAT is increased, which regulates the expression of lipoprotein lipase and glucose transporters, thus increasing substrate uptake⁴⁹. Central to the ability of BAT to generate heat is the expression of UCP1. This requires both SNS activity and local increases in thyroid hormone activity. Finally, facilitated SNS tone to white adipose tissue increases the rate of lipolysis. The free fatty acids released are then delivered directly to the BAT or indirectly via hepatic production of triglyceride-rich lipoproteins.

Although the cold-induced changes within the BAT have yet to be documented in humans, many of the changes outside of the BAT have been described (Figure 2). Following acute mild cold exposure in humans, serum catecholamines, thyroid hormone, free fatty acids, and triglycerides are increased^{68,72,96,97}. Although heart rate falls, blood pressure is increased, indicating a rise in total peripheral resistance. This is most likely mediated by cutaneous vasoconstriction and is seen as a fall in skin temperature in the extremities. Heart rate variability analysis performed during cold exposure reveals that autonomic tone to the heart is altered with increases in both low-frequency (sympathetic and parasympathetic) and high-frequency (parasympathetic) components⁷². Altogether, optimal activation of BAT/CIT requires a complex series of physiological changes.

In summary, the effects of cold in humans are complex and the physiological processes are regulated by feedback loops centered around the defense of core body temperature. Non-physiological manipulation of thermogenic effector tissues (BAT or otherwise) in isolation may benefit from coordinated changes in the vasculature, hormonal profile, and circulating substrates. Alternatively, physiological activation of thermogenesis through cold exposure will require minimization of non-thermogenic responses and manipulation of core temperature negative feedback.

CONCLUSIONS

From a thermodynamic perspective, increasing EE to induce weight loss is a practical approach. When applied to biological systems, compensatory increases in energy intake often occur. The adipostat hypothesis postulates that fat storage is well defended, especially in the context of negative energy balance.

Exercise is an effective means of increasing EE. Despite this, energy expended during exercise does not result in the predicted amount of weight loss, even when compliance with exercise is maintained. Consistent with the adipostat hypothesis, EE induced by exercise is largely compensated for by an increase in food intake. Conversely, metabolic states, such as DNP treatment and hyperthyroidism, which result in conversion of energy to heat, do not appear to be fully compensated by increases in food intake. Although it is possible that factors other than EE, such as appetite suppression or reduced absorption, may be involved, the current data suggest that it is more than plausible

that activating thermogenesis can induce clinically relevant weight loss without caloric restriction, a state of thermogenic disconnect.

The effects of CIT share many common mechanisms with thyroid hormone heat production. In addition, the discovery of functional BAT in adult humans raises the possibility of exploitable mitochondrial uncoupling in humans. Targeting BAT mass and activity as a means of increasing EE has many potential benefits. First, unlike the use of DNP, uncoupling will only occur in one tissue type. Potentially, this would reduce any toxic effect on tissues not normally suited to high rates of uncoupling. Second, BAT contains the necessary vasculature to ensure an adequate supply of nutrients and the dissipation of excess heat. If humans are appropriately cold adapted, nutrient delivery and heat dissipation would be expected to meet BAT activity and thus maximize EE, heat loss, and ultimately weight loss. Finally, BAT activation may have other metabolic benefits independent of weight loss. In murine models, BAT has been shown to contribute significantly to whole-body glucose and lipid clearance. Although this may not directly alter insulin resistance at the molecular level, BAT could contribute to concomitant glycemic control^{98–102} and lipid lowering^{103,104} in obese individuals.

In the absence of an optimal method of specifically activating BAT in humans, studies using cold exposure without inducing shivering are a practical alternative. As such, a number of areas require further investigation, as outlined below.

1. Can CIT be augmented (through controlled cold exposure) to achieve a level sufficient to induce weight loss?
2. How much does food intake change in response to CIT? (That is, can we quantify the magnitude of the thermogenic disconnect?)
3. In terms of magnitude, what is the relative contribution of BAT-mediated compared with non-BAT-mediated processes to CIT in humans?
4. Can BAT (or other CIT processes) be activated without cold exposure?
5. If weight loss is not possible, will activation of thermogenic processes contribute to weight stabilization?
6. Will thermogenesis contribute to metabolic health independent of weight loss?

Some tools that may be required to investigate the questions listed above include: (i) a practical way of applying cold exposure outside the laboratory; (ii) a method to quantify BAT and non-BAT-mediated thermogenesis; and (iii) a method of distinguishing BAT activity from BAT mass. The answers to these questions and further investigations into the molecular basis of thermogenesis may reveal therapeutic targets that will be translatable into effective obesity and metabolic therapy.

ACKNOWLEDGEMENTS

CYT is funded by the Wellcome Trust, KI is funded by Manpei Suzuki Diabetes Foundation, SV is funded by the Biotechnology

and Biological Sciences Research Council (BBSRC). The authors declare no conflict of interest.

REFERENCES

1. Wu T, Gao X, Chen M, et al. Long-term effectiveness of diet-plus-exercise interventions vs. diet-only interventions for weight loss: a meta-analysis. *Obes Rev* 2009; 10: 313–323.
2. Barte JCM, Ter Bogt NCW, Bogers RP, et al. Maintenance of weight loss after lifestyle interventions for overweight and obesity, a systematic review. *Obes Rev* 2010; 11: 899–906.
3. Franz MJ, VanWormer JJ, Crain AL, et al. Weight-loss outcomes: a systematic review and meta-analysis of weight-loss clinical trials with a minimum 1-year follow-up. *J Am Diet Assoc* 2007; 107: 1755–1767.
4. Rosenbaum M, Hirsch J, Gallagher DA, et al. Long-term persistence of adaptive thermogenesis in subjects who have maintained a reduced body weight. *Am J Clin Nutr* 2008; 88: 906–912.
5. Rosenbaum M, Kissileff HR, Mayer LE, et al. Energy intake in weight-reduced humans. *Brain Res* 2010; 1350: 95–102.
6. Leibel RL, Rosenbaum M, Hirsch J. Changes in energy expenditure resulting from altered body weight. *N Engl J Med* 1995; 332: 621–628.
7. Cannon B, Nedergaard J. Thermogenesis challenges the adipostat hypothesis for body-weight control. *Proc Nutr Soc* 2009; 68: 401–407.
8. Rothwell NJ, Saville ME, Stock MJ. Effects of feeding a 'cafeteria' diet on energy balance and diet-induced thermogenesis in four strains of rat. *J Nutr* 1982; 112: 1515–1524.
9. Rothwell NJ, Stock MJ. Luxusconsumption, diet-induced thermogenesis and brown fat: the case in favour. *Clin Sci* 1983; 64: 19–23.
10. Cypess AM, Lehman S, Williams G, et al. Identification and importance of brown adipose tissue in adult humans. *N Engl J Med* 2009; 360: 1509–1517.
11. Virtanen KA, Lidell ME, Orava J, et al. Functional brown adipose tissue in healthy adults. *N Engl J Med* 2009; 360: 1518–1525.
12. van Marken Lichtenbelt WD, Vanhommerig JW, Smulders NM, et al. Cold-activated brown adipose tissue in healthy men. *N Engl J Med* 2009; 360: 1500–1508.
13. Rosenbaum M, Leibel RL, Hirsch J. Obesity. *N Engl J Med* 1997; 337: 396–407.
14. Stroebele N, Hill JO, Willich SN. Identifying the energy gap in the German population using results from representative national health surveys (1985–2002). *Public Health Nutr* 2011; 14: 44–48.
15. Donnelly JE, Hill JO, Jacobsen DJ, et al. Effects of a 16-month randomized controlled exercise trial on body weight and composition in young, overweight men and women: the Midwest Exercise Trial. *Arch Intern Med* 2003; 163: 1343–1350.
16. Bouchard C. The magnitude of the energy imbalance in obesity is generally underestimated. *Int J Obes* 2008; 32: 879–880.
17. Hill JO, Peters JC, Wyatt HR. Using the energy gap to address obesity: a commentary. *J Am Diet Assoc* 2009; 109: 1848–1853.
18. Ravussin E. Metabolic differences and the development of obesity. *Metabolism* 1995; 44(Suppl 3): 12–14.
19. Astrup A, Buemann B, Toubro S, et al. Low resting metabolic rate in subjects predisposed to obesity: a role for thyroid status. *Am J Clin Nutr* 1996; 63: 879–883.
20. Westerterp KR, Speakman JR. Physical activity energy expenditure has not declined since the 1980s and matches energy expenditures of wild mammals. *Int J Obes* 2008; 32: 1256–1263.
21. Edholm OG, Fletcher JG, Widdowson EM, et al. The energy expenditure and food intake of individual men. *Br J Nutr* 1955; 9: 286–300.
22. Melzer K, Kayser B, Saris WHM, et al. Effects of physical activity on food intake. *Clin Nutr* 2005; 24: 885–895.
23. van Marken Lichtenbelt WD, Schrauwen P, van De Kerckhove S, et al. Individual variation in body temperature and energy expenditure in response to mild cold. *Am J Physiol Endocrinol Metab* 2002; 282: E1077–E1083.
24. Black AE, Coward WA, Cole TJ, et al. Human energy expenditure in affluent societies: an analysis of 574 doubly-labelled water measurements. *Eur J Clin Nutr* 1996; 50: 72–92.
25. Sjodin AM, Andersson AB, Hogberg JM, et al. Energy balance in cross-country skiers: a study using doubly labeled water. *Med Sci Sports Exerc* 1994; 26: 720–724.
26. Garrow JS, Summerbell CD. Meta-analysis: effect of exercise, with or without dieting, on the body composition of overweight subjects. *Eur J Clin Nutr* 1995; 49: 1–10.
27. Pi-Sunyer FX, Becker DM, Bouchard C, et al. Clinical guidelines on the identification, evaluation, and treatment of overweight and obesity in adults: the evidence report. National Institutes of Health. *Obes Res* 1998; 6(Suppl 2): S51–S209.
28. Slentz CA, Duscha BD, Johnson JL, et al. Effects of the amount of exercise on body weight, body composition, and measures of central obesity: STRRIDE. A randomized controlled study. *Arch Intern Med* 2004; 164: 31–39.
29. Jakicic JM, Otto AD. Physical activity considerations for the treatment and prevention of obesity. *Am J Clin Nutr* 2005; 82(Suppl): S226–S229.
30. Krotkiewski M. Thyroid hormones and treatment of obesity. *Int J Obes Relat Metab Disord* 2000; 24(Suppl 2): S116–S119.
31. Hoogwerf BJ, Nuttall FQ. Long-term weight regulation in treated hyperthyroid and hypothyroid subjects. *Am J Med* 1984; 76: 963–970.
32. Douyon L, Scheingart DE. Effect of obesity and starvation on thyroid hormone, growth hormone, and cortisol secretion. *Endocrinol Metab Clin North Am* 2002; 31: 173–189.
33. Michalaki MA, Vagenakis AG, Leonardou AS, et al. Thyroid function in humans with morbid obesity. *Thyroid* 2006; 16: 73–78.
34. Biondi B. Thyroid and obesity: an intriguing relationship. *J Clin Endocrinol Metab* 2010; 95: 3614–3617.

35. Roti E, Minelli R, Salvi M. Thyroid hormone metabolism in obesity. *Int J Obes Relat Metab Disord* 2000; 24(Suppl 2): S113–S115.
36. Reinehr T. Obesity and thyroid function. *Mol Cell Endocrinol* 2010; 316: 165–171.
37. Jansson S, Lie-Karlsen K, Stenqvist O, et al. Oxygen consumption in patients with hyperthyroidism before and after treatment with beta-blockade versus thyrostatic treatment: a prospective randomized study. *Ann Surg* 2001; 233: 60–64.
38. Mitchell CS, Savage DB, Dufour S, et al. Resistance to thyroid hormone is associated with raised energy expenditure, muscle mitochondrial uncoupling, and hyperphagia. *J Clin Invest* 2010; 120: 1345–1354.
39. Kiortsis DN, Durack I, Turpin G. Effects of a low-calorie diet on resting metabolic rate and serum tri-iodothyronine levels in obese children. *Eur J Pediatr* 1999; 158: 446–450.
40. Buscemi S, Verga S, Maneri R, et al. Influences of obesity and weight loss on thyroid hormones. A 3–3.5-year follow-up study on obese subjects with surgical bilio-pancreatic by-pass. *J Endocrinol Invest* 1997; 20: 276–281.
41. Spaulding SW, Chopra IJ, Sherwin RS, et al. Effect of caloric restriction and dietary composition of serum T3 and reverse T3 in man. *J Clin Endocrinol Metab* 1976; 42: 197–200.
42. Koppeschaar HP, Meinders AE, Schwarz F. The effect of a low-calorie diet alone and in combination with triiodothyronine therapy on weight loss and hypophyseal thyroid function in obesity. *Int J Obes* 1983; 7: 123–131.
43. Rozen R, Abraham G, Falcou R, et al. Effects of a 'physiological' dose of triiodothyronine on obese subjects during a protein-sparing diet. *Int J Obes* 1986; 10: 303–312.
44. Kaptein SA, Gignac MA, Badley EM. Differences in the workforce experiences of women and men with arthritis disability: a population health perspective. *Arthritis Rheum* 2009; 61: 605–613.
45. Moore R, Grant AM, Howard AN, et al. Treatment of obesity with triiodothyronine and a very-low-calorie liquid formula diet. *Lancet* 1980; 1: 223–226.
46. Wilson JH, Lamberts SW. The effect of triiodothyronine on weight loss and nitrogen balance of obese patients on a very-low-calorie liquid-formula diet. *Int J Obes* 1981; 5: 279–282.
47. Silva JE. Thermogenic mechanisms and their hormonal regulation. *Physiol Rev* 2006; 86: 435–464.
48. Silva JE. The thermogenic effect of thyroid hormone and its clinical implications. *Ann Intern Med* 2003; 139: 205–213.
49. Cannon B, Nedergaard J. Brown adipose tissue: function and physiological significance. *Physiol Rev* 2004; 84: 277–359.
50. Christoffolete MA, Linardi CC, de Jesus L, et al. Mice with targeted disruption of the *Dio2* gene have cold-induced over-expression of the uncoupling protein 1 gene but fail to increase brown adipose tissue lipogenesis and adaptive thermogenesis. *Diabetes* 2004; 53: 577–584.
51. Balaban RS. Regulation of oxidative phosphorylation in the mammalian cell. *Am J Physiol* 1990; 258: C377–C389.
52. Dunlop DM. The use of 2,4-dinitrophenol as a metabolic stimulant. *Br Med J* 1934; 1: 524–527.
53. Rabinowitch IM. The danger from dinitrophenol. *Can Med Assoc J* 1934; 30: 564–565.
54. Tainter ML, Cutting WC, Stockton AB. Use of dinitrophenol in nutritional disorders: a critical survey of clinical results. *Am J Public Health Nations Health* 1934; 24: 1045–1053.
55. Boardman WW. Rapidly developing cataracts after dinitrophenol. *Cal West Med* 1935; 43: 118–119.
56. Tainter ML, Stockton AB, Cutting WC. Dinitrophenol in the treatment of obesity. *JAMA* 1935; 105: 332–337.
57. Wijers SL, Saris WH, van Marken Lichtenbelt WD. Cold-induced adaptive thermogenesis in lean and obese. *Obesity* 2010; 18: 1092–1099.
58. Wijers SL, Saris WH, van Marken Lichtenbelt WD. Individual thermogenic responses to mild cold and overfeeding are closely related. *J Clin Endocrinol Metab* 2007; 92: 4299–4305.
59. Celi FS, Brychta RJ, Linderman JD, et al. Minimal changes in environmental temperature result in a significant increase in energy expenditure and changes in the hormonal homeostasis in healthy adults. *Eur J Endocrinol* 2010; 163: 863–872.
60. Dauncey MJ. Influence of mild cold on 24 h energy expenditure, resting metabolism and diet-induced thermogenesis. *Br J Nutr* 1981; 45: 257–267.
61. van Ooijen AM, van Marken Lichtenbelt WD, van Steenhoven AA, et al. Seasonal changes in metabolic and temperature responses to cold air in humans. *Physiol Behav* 2004; 82: 545–553.
62. Yoneshiro T, Aita S, Matsushita M, et al. Brown adipose tissue, whole-body energy expenditure, and thermogenesis in healthy adult men. *Obesity* 2011; 19: 13–16.
63. Scholander PF, Hammel HT, Hart JS, et al. Cold adaptation in Australian Aborigines. *J Appl Physiol* 1958; 13: 211–218.
64. Wyndham CH, Morrison JF. Adjustment to cold of Bushmen in the Kalahari Desert. *J Appl Physiol* 1958; 13: 219–225.
65. Hassi J, Sikkilä K, Ruokonen A, et al. The pituitary–thyroid axis in healthy men living under subarctic climatological conditions. *J Endocrinol* 2001; 169: 195–203.
66. Korhonen I, Hassi J, Leppaluoto J. Serum concentrations of thyroid and adrenal hormones and TSH in men after repeated 1 h-stays in a cold room. *Int J Circumpolar Health* 2001; 60: 604–608.
67. Leppaluoto J, Korhonen I, Hassi J. Habituation of thermal sensations, skin temperatures, and norepinephrine in men exposed to cold air. *J Appl Physiol* 2001; 90: 1211–1218.
68. Leppaluoto J, Korhonen I, Huttunen P, et al. Serum levels of thyroid and adrenal hormones, testosterone, TSH, LH, GH and prolactin in men after a 2-h stay in a cold room. *Acta Physiol Scand* 1988; 132: 543–548.
69. Leppaluoto J, Paakkonen T, Korhonen I, et al. Pituitary and autonomic responses to cold exposures in man. *Acta Physiol Scand* 2005; 184: 255–264.

70. Leppaluoto J, Sikkilä K, Hassi J. Seasonal variation of serum TSH and thyroid hormones in males living in subarctic environmental conditions. *Int J Circumpolar Health* 1998; 57(Suppl 1): 383–385.
71. Leppaluoto J, Westerlund T, Huttunen P, et al. Effects of long-term whole-body cold exposures on plasma concentrations of ACTH, beta-endorphin, cortisol, catecholamines and cytokines in healthy females. *Scand J Clin Lab Invest* 2008; 68: 145–153.
72. Mäkinen TM, Mantysaari M, Paakkonen T, et al. Autonomic nervous function during whole-body cold exposure before and after cold acclimation. *Aviat Space Environ Med* 2008; 79: 875–882.
73. Mäkinen TM, Paakkonen T, Palinkas LA, et al. Seasonal changes in thermal responses of urban residents to cold exposure. *Comp Biochem Physiol A Mol Integr Physiol* 2004; 139: 229–238.
74. Paakkonen T, Leppaluoto J. Cold exposure and hormonal secretion: a review. *Int J Circumpolar Health* 2002; 61: 265–276.
75. Smolander J, Leppaluoto J, Westerlund T, et al. Effects of repeated whole-body cold exposures on serum concentrations of growth hormone, thyrotropin, prolactin and thyroid hormones in healthy women. *Cryobiology* 2009; 58: 275–278.
76. Bittel JH. Heat debt as an index for cold adaptation in men. *J Appl Physiol* 1987; 62: 1627–1634.
77. Bruck K, Baum E, Schwennicke HP. Cold-adaptive modifications in man induced by repeated short-term cold-exposures and during a 10-day and-night cold-exposure. *Pflügers Arch* 1976; 363: 125–133.
78. Johnson RE, Kark RM. Environment and food intake in man. *Science* 1947; 105: 378–379.
79. Leonard WR, Sorensen MV, Galloway VA, et al. Climatic influences on basal metabolic rates among circumpolar populations. *Am J Hum Biol* 2002; 14: 609–620.
80. Steegmann AT Jr. Human cold adaptation: an unfinished agenda. *Am J Hum Biol* 2007; 19: 218–227.
81. Ruff C. Variation in human body size and shape. *Annu Rev Anthropol* 2002; 31: 211–232.
82. Holliday TW, Hilton CE. Body proportions of circumpolar peoples as evidenced from skeletal data: Ipiutak and Tigara (Point Hope) versus Kodiak Island Inuit. *Am J Phys Anthropol* 2010; 142: 287–302.
83. Milan FA, Rodahl K. Caloric requirements of man in the Antarctic. *J Nutr* 1961; 75: 152–156.
84. Orr NW. Food requirements and weight changes of men on Antarctic expeditions. *Br J Nutr* 1965; 19: 79–91.
85. Budd GM, Warhaft N. Body temperature, shivering, blood pressure and heart rate during a standard cold stress in Australia and Antarctica. *J Physiol* 1966; 186: 216–232.
86. Budd GM, Warhaft N. Cardiovascular and metabolic responses to noradrenaline in man, before and after acclimatization to cold in Antarctica. *J Physiol* 1966; 186: 233–242.
87. Milan FA, Elsner RW, Rodahl K. Thermal and metabolic responses of men in the Antarctic to a standard cold stress. *J Appl Physiol* 1961; 16: 401–404.
88. Simpson A. The effect of Antarctic residence on energy dynamics and aerobic fitness. *Int J Circumpolar Health* 2010; 69: 220–235.
89. Hey E. Thermal neutrality. *Br Med Bull* 1975; 31: 69–74.
90. Hill JR. Reaction of the new-born animal to environmental temperature. *Br Med Bull* 1961; 17: 164–167.
91. Lopez M, Sessler DI, Walter K, et al. Rate and gender dependence of the sweating, vasoconstriction, and shivering thresholds in humans. *Anesthesiology* 1994; 80: 780–788.
92. Leblanc J. Factors affecting cold acclimation and thermogenesis in man. *Med Sci Sports Exerc* 1988; 20(Suppl): S193–S196.
93. Whittle AJ, Lopez M, Vidal-Puig A. Using brown adipose tissue to treat obesity: the central issue. *Trends Mol Med* 2011; 17: 405–411.
94. Morrison SF, Nakamura K, Madden CJ. Central control of thermogenesis in mammals. *Exp Physiol* 2008; 93: 773–797.
95. Foster DO, Frydman ML. Nonshivering thermogenesis in the rat. II. Measurements of blood flow with microspheres point to brown adipose tissue as the dominant site of the calorigenesis induced by noradrenaline. *Can J Physiol Pharmacol* 1978; 56: 110–122.
96. Hong JH, Kim HJ, Kim KJ, et al. Comparison of metabolic substrates between exercise and cold exposure in skaters. *J Physiol Anthropol* 2008; 27: 273–281.
97. Vallerand AL, Jacobs I. Influence of cold exposure on plasma triglyceride clearance in humans. *Metabolism* 1990; 39: 1211–1218.
98. Inokuma K, Ogura-Okamoto Y, Toda C, et al. Uncoupling protein 1 is necessary for norepinephrine-induced glucose utilization in brown adipose tissue. *Diabetes* 2005; 54: 1385–1391.
99. Shimizu Y, Nikami H, Saito M. Sympathetic activation of glucose utilization in brown adipose tissue in rats. *J Biochem* 1991; 110: 688–692.
100. Shibata H, Perusse F, Vallerand A, et al. Cold exposure reverses inhibitory effects of fasting on peripheral glucose uptake in rats. *Am J Physiol* 1989; 257: R96–R101.
101. Cawthorne MA. Does brown adipose tissue have a role to play in glucose homeostasis? *Proc Nutr Soc* 1989; 48: 207–214.
102. Greco-Perotto R, Zaninetti D, Assimacopoulos-Jeannet F, et al. Stimulatory effect of cold adaptation on glucose utilization by brown adipose tissue. Relationship with changes in the glucose transporter system. *J Biol Chem* 1987; 262: 7732–7736.
103. Bartelt A, Bruns OT, Reimer R, et al. Brown adipose tissue activity controls triglyceride clearance. *Nat Med* 2011; 17: 200–205.
104. Laplante M, Festuccia WT, Soucy G, et al. Tissue-specific postprandial clearance is the major determinant of PPAR-gamma-induced triglyceride lowering in the rat. *Am J Physiol* 2009; 296: R57–R66.