RESEARCH ARTICLE



Novel lures and COI sequences reveal cryptic new species of *Bactrocera* fruit flies in the Solomon Islands (Diptera, Tephritidae, Dacini)

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Abstract

Results from a snap-shot survey of Dacine fruit flies carried out on three of the Solomon Islands in April 2018 are reported. Using traps baited with the male lures cue-lure, methyl eugenol, and zingerone, 30 of the 48 species previously known to occur in the Solomon Islands were collected. Six species are newly described here: *Bactrocera allodistincta* **sp. nov.**, *B. geminosimulata* **sp. nov.**, *B. kolombangarae* **sp. nov.**, *B. quasienochra* **sp. nov.**, *B. tsatsiai* **sp. nov.**, and *B. vargasi* **sp. nov.**, all authored by Leblanc & Doorenweerd. An illustrated key to the 54 species now known to be present in the country is provided.

Keywords

Dacus, Oceania, pest species, taxonomy, Zeugodacus

Introduction

Dacine fruit flies (Diptera: Tephritidae: Dacini), a species-rich Old World tropical group, is composed of 947 currently known species, including 83 crop pests (White and Elson-Harris 1992; Vargas et al. 2015; Doorenweerd et al. 2018). Diversity is particularly high in Australasia, with 332 species described and an imminent publication

of 65 new species from Papua New Guinea (R.A.I. Drew, pers. comm.). Many more new species are being discovered, especially cryptic species, with ever improving molecular diagnostic tools and the emergence of new generation male lures (De Meyer et al. 2015; Manrakhan et al. 2017; Royer et al. 2018, 2019; Doorenweerd et al. 2020).

The earliest Dacine fruit fly record in the Solomon Islands was the description of *Bactrocera longicornis* Macquart, in 1835. By 1939, eleven species were known (Malloch 1939), growing to 26 five decades later (Drew 1989). Extensive survey efforts through trapping and host fruit surveys during the Regional Fruit Fly Projects in the Pacific (Allwood and Drew 1997; Allwood 2000; Lidner and McLeod 2008) nearly doubled the number of species to 48 (Drew and Romig 2001). Two decades later, we carried out a snap-shot survey on three islands (Guadalcanal, Kolombangara, Gizo), with the inclusion of zingerone lure, to collect fresh material and develop molecular diagnostic tools to help further characterize the species found in the Solomon Islands. In just a couple of weeks, we discovered six new species, including cryptic species that would not have been detectable without molecular characterization. We herein describe these new species and provide a key to the 54 species now present in the Solomon Islands.

Materials and methods

Collecting and curation

We maintained 79 sets of three traps separately baited with male lures (cue-lure, methyl eugenol and zingerone) in the Solomon Islands in April 2018. We used commercially available cue-lure and methyl eugenol plugs (Scentry Biologicals, Billings, Montana). Zingerone (= vanillylacetone) lure was prepared by dipping dental cotton wicks in zingerone powder (Sigma-Aldrich) melted over a hot plate and allowed to solidify in the wicks. Small vertical bucket traps (Leblanc et al. 2015: fig. 1) were made of 5-oz urine sample cups (Stockwell Scientific, Scottsdale, Arizona) with two 20 mm wide lateral circular openings on opposite sides, 12 mm below the top, with a hole drilled in the lid center, through which a 30-cm-long, 15-gauge, aluminum tie wire was inserted, and bent into a hook below the lid. The male lure unit and a 10 × 10 mm piece of dichlorvos (DVVP) strip (Vaportape II; Hercon Environmental, Emingsville, PA) were attached to the hook below the lid. A 10-cm-wide black square plastic food plate (Waddington North America) was placed on top of the trap to prevent flooding by frequent rain. A solution of 25% propylene glycol (Better World Manufacturing, Fresno, CA) was used in the trap to preserve captured flies, until they were transferred to 95% ethanol during trap servicing. The 79 sets of traps in agricultural areas and endemic forest on the islands of Guadalcanal and Kolombangara, and agricultural areas on Gizo Island (Fig. 1) were maintained for 12, four, and six days, respectively. Forest trapping sites were ca. 50 meters apart along transects that followed trails. Sampled flies were stored in 95% ethanol in a -20 °C freezer to preserve DNA. All flies were identified to species using available keys (Drew 1989; Drew and Romig 2001). We pulled one or two legs from specimens

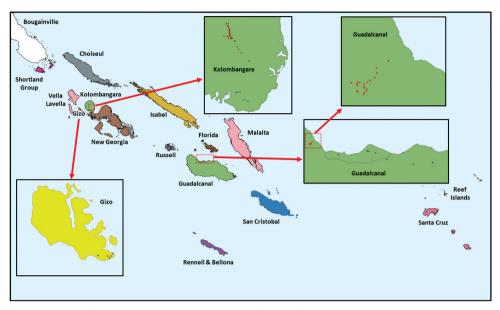


Figure 1. Map of the Solomon Islands, with groups of islands for distribution records, and trapping locations on Guadalcanal, Gizo and Kolombangara islands in the 2018 survey. Red circles are sites located in forest and blue triangles are sites in agricultural environments.

that were selected for DNA extraction (for further details on DNA extraction methods see Doorenweerd et al. 2020). All holotypes and all, or a subset of, the paratypes were double-mounted to be stored as dry specimens in collections for permanent future reference. Before drying flies for double-mounting (White and Elson-Harris 1992), we pinned them through the scutum with a minuten pin and soaked them in diethyl-ether for 3–12 hours to fix and preserve their natural coloration. We photographed specimens using a Nikon D7100 camera attached to an Olympus SZX10 microscope and used Helicon Focus pro v6.7.1 to merge pictures taken at a range of focal planes. To measure specimens (all available or up to 10 specimens measured per species), we used an ocular grid mounted on an Olympus SZ30 dissecting microscope.

Morphological terms and taxonomic assignment

Morphological terminology used in the descriptions follows White et al. (1999) and assignment of species to genera follows Doorenweerd et al. (2018). We treat *Zeugodacus* as a distinct genus from *Bactrocera* and *Dacus* (Krosch et al. 2012; Virgilio et al. 2015; Dupuis et al. 2017; San Jose et al. 2018). Subgenus assignment for each species follows reclassifications recently published by Hancock and Drew (Hancock 2015; Drew and Hancock 2016; Hancock and Drew 2015, 2018a, b). The host plant records included in the key follow the compilation published by Leblanc et al. (2012). For accurate taxonomic application of host plant records from the literature we used the World Flora Online (WFO 2021).

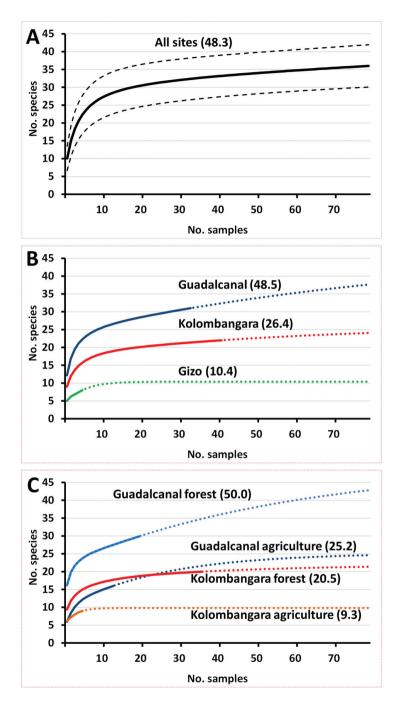


Figure 2. Species accumulation curves based on the 2018 survey of Solomon Islands **A** for all sampled sites with 95% confidence interval range **B** for the three individually sampled islands, and **C** comparing forest and agricultural sites separately on Guadalcanal and Kolombangara. Estimated species numbers for each curve (in brackets) based on the Chao 2 estimator.

COI sequence analysis

Representatives of all species newly described here were also included in the cytochrome c oxidase I (COI) study of Doorenweerd et al. (2020), under tentative species names. For that study, 1493 base pairs of the COI gene were sequenced and comparatively analyzed in a dataset that included 163 species of *Bactrocera*. We include here the maximum like-lihood gene tree from that study and the summary *Bactrocera* species statistics as supplementary material (Suppl. material 1: Fig. S1, Suppl. material 2: Table S1). For the methods for DNA extraction, sequencing and analyses we refer to Doorenweerd et al. (2020). Collecting information as well as COI sequences are available on BOLD (Ratnasingham and Hebert 2007) dataset (DOI: http://dx.doi.org/10.5883/DS-DACCOI), as well as NCBI GenBank (accessions MZ196488–MZ196507). Each specimen for which DNA was extracted was assigned a unique code in the format "UHIM.ms000000", physically labelled as such, and this number forms the 'Sample ID' in BOLD.

Estimating biodiversity

We used EstimateS software (Colwell 2019) to generate species accumulation curves and estimate species diversity, using the incidence-based Chao 2 algorithm. We generated accumulation curves, with 100 randomizations without replacement for confidence intervals for all sites collectively, separately for each island, and comparing agricultural and forest sites in Guadalcanal and Kolombangara.

Abbreviations

Bernice Pauahi Bishop Museum, Honolulu, Hawaii, United States;
Biosecurity Solomon Islands, Honiara, Solomon Islands;
William F. Barr Entomological Museum, Moscow, Idaho, United States;
University of Hawaii Insect Museum, Honolulu, Hawaii, United States;
National Museum of Natural History, Smithsonian Institution, Washing-
ton DC, United States.

Results and taxonomy

Bactrocera (*Bactrocera*) *allodistincta* Leblanc & Doorenweerd, sp. nov. http://zoobank.org/6D929FD2-D802-42D1-B15D-B14C78CF4442 Fig. 3A–E

Type material. *Holotype.* SOLOMON ISLANDS • ♂; Guadalcanal, forest; -9.4067, 159.8647; 167 m; 4–16 Apr. 2018; L. Leblanc, F. Tsatsia leg.; cue-lure baited trap FFS0015. Deposited in UHIM. *Paratypes.* 11 males. SOLOMON ISLANDS • 1 ♂; Guadalcanal forest; -9.4041, 159.8628; 153 m; 4–16 Apr. 2018; L. Leblanc, F. Tsatsia leg.;

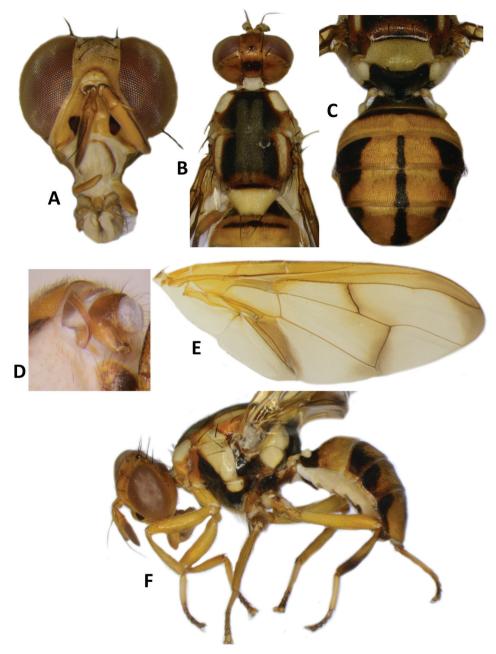


Figure 3. *Bactrocera allodistincta* sp. nov. **A** head **B** head and scutum **C** abdomen **D** male genitalia **E** wing **F** lateral view.

cue-lure baited trap FFS0011 • 1 3; same locality and date as for preceding; -9.4067, 159.8647; 167 m; trap FFS0015 • 1 3; same locality and date as for preceding; -9.4072, 159.8664; 153 m; trap FFS0016 • 2 3; same locality and date as for preced-

ing; -9.4064, 159.8671; 145 m; trap FFS0018; molecular voucher UHIM.ms08766 • 2 3; same locality and date as for preceding; -9.4059, 159.8672; 133 m; trap FFS0019 • 1 3; same locality and date as for preceding; -9.4055, 159.8665; 145 m; trap FFS0020 • 1 3; same locality and date as for preceding; -9.4040, 159.8652; 125 m; trap FFS0023 • 1 3; same locality and date as for preceding; -9.4026, 159.8695; 57 m; trap FFS0027 • 1 3; same locality and date as for preceding; -9.4000, 159.8700; 57 m; trap FFS0029. Seven of the paratypes are deposited at UHIM, three at WFBM, and one at USNM.

Differential diagnosis. *Bactrocera allodistincta* differs from *B. pseudodistincta* (Drew) (Fig. 4) in the presence of orange-brown lateral and posterior markings on the predominantly black scutum, abdominal tergites III–V with a narrower medial black stripe, the lateral black markings on tergite IV narrowed posteriorly, and the rather diffuse fuscous crossband on the wing. It differs from *B. distincta* (Malloch) in that the costal band is diffuse orange-brown and the crossband is sinuous, with a bend along vein M (Fig. 3E), whereas the entire costal band, including in the basicostal and costal cells, is dark fuscous and the crossband is broad and straight in *B. distincta* (Fig. 5E).

Molecular diagnosis. We obtained a single COI sequence (UHIM.ms08766) which matches closest to *Bactrocera pedestris* (Bezzi) [misidentified as *B. gombokensis* Drew & Hancock, 1994 in Doorenweerd et al. 2020], at 10.25% pairwise distance. *Bactrocera pseudodistincta* (Drew) [N = 2] is also represented in the dataset and does not appear as a close match, but *B. distincta* is not represented.

Description of adult. Male. *Head* (Fig. 3A). Height 1.56 ± 0.12 (SD) (1.37-1.67) mm. Frons of even width, 0.80 ± 0.03 (0.73-0.83) mm long and 1.40 ± 1.05 (1.33-1.50) times as long as broad; fulvous, sometimes fuscous around orbital seta and anteromedial hump; latter covered by short red-brown microtrichia; three pairs of dark fuscous frontal setae present; lunule fulvous. Ocellar triangle black. Vertex fulvous with two pairs of dark fuscous vertical setae. Face fulvous with a pair of moderately sized oval black spots in antennal furrows; length 0.49 ± 0.05 (0.43-0.60) mm. Gena fulvous, with a fuscous subocular spot and a dark fuscous seta. Occiput fulvous with a dark fuscous to black dorsomedial marking; a row of 4–6 dark fuscous postocular setae present behind eye. Antenna with scape and pedicel fulvous and first flagellomere fulvous with pale fuscous on lateral surface of flagellum; a strong red-brown dorsal seta on pedicel; arista fulvous basally and black distally; length of segments: 0.22 ± 0.04 (0.17-0.27) mm; 0.27 ± 0.03 (0.23-0.33) mm; 0.71 ± 0.04 (0.67-0.73) mm.

Thorax (Fig. 3B). Scutum black except orange-brown ventral to and narrowly medial to lateral postsutural vitta, around notopleural suture, along lateral margin between postpronotal lobe and notopleuron, medial to postpronotal lobe, and along posterior margin of scutum. Scutum with two broad parallel medial stripes of dense silvery microtrichia along entire scutum length. Pleural areas black except orange-brown anterior margin of anepisternum and proeipsternum. Yellow markings: postpronotal lobe; notopleuron; moderately broad paired parallel-sided lateral postsutural vitta reaching intra-alar seta posteriorly; broad anepisternal stripe with anterior margin straight, reaching to anterior notopleural seta dorsally; a large transverse spot on katepisternum

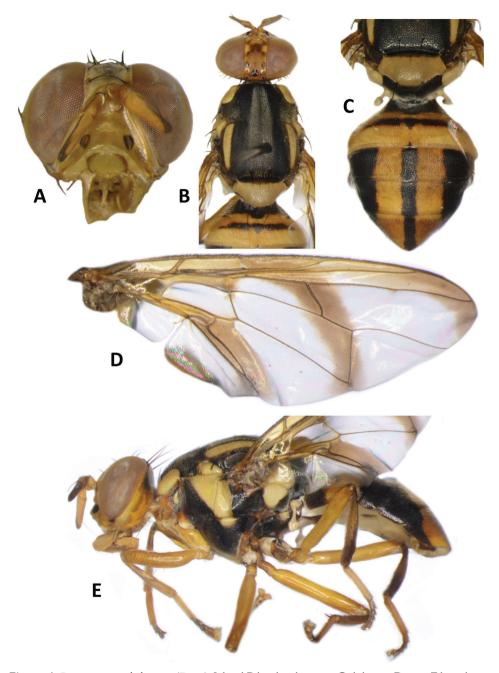


Figure 4. Bactrocera pseudodistincta (Drew) A head B head and scutum C abdomen D wing E lateral view.

below the anepisternal stripe; anterior ⁴/₅ of anatergite and ³/₄ of katatergite (posteriorly black). Mediotergite black. Scutellum yellow except for very narrow black basal band. Setae: 1 pair scutellar; 1 pair prescutellar acrostichal; 1 pair intra-alar; 1 pair postalar;

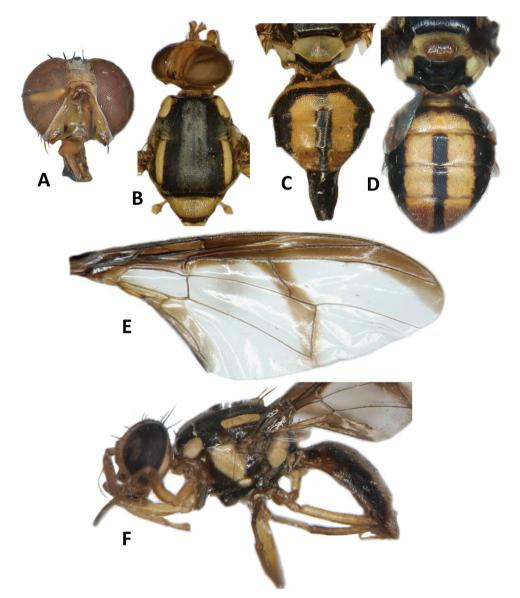


Figure 5. *Bactrocera distincta* (Malloch) **A** head **B** head and scutum **C** female abdomen **D** male abdomen **E** wing **F** lateral view.

1 pair postsutural supra-alar; 1 pair anepisternal; 2 pairs notopleural; 2 pairs scapular; all setae well developed and dark fuscous.

Legs (Fig. 3F). All legs entirely fulvous with apical ²/₅ of hind tibia fuscous. Fore femur with a row of long pale dorsal setae. Mid-tibia with apical black spur.

Wing (Fig. 3E). Length 5.6 \pm 0.2 (5.3–5.9) mm; basal costal and costal cells fuscous with microtrichia in posterodistal corner of costal cell; broad fuscous costal

band confluent with R_{4+5} , remaining broad at apex and ending at apex of medial vein; a diffuse orange-brown crossband along crossvein r-m, continuing along M and dmcu to reach posterior wing margin, and a broad fuscous anal streak over cell bcu and basal margin of cu₁; remainder of wing light fuscous; dense aggregation of microtrichia around A₁ + CuA₂; supernumerary lobe weakly developed.

Abdomen (Fig. 3C, D). Oval with tergites not fused; pecten present on tergite III; posterior lobe of surstylus short; abdominal sternite V with a deep concavity on posterior margin. Base of syntergite I+II wider than long. Syntergite I+II orange-brown with base black and a narrow sub-basal transverse medial black band. Tergites III–V orange-brown with moderately broad medial black stripe reaching apex, and large lateral black markings on tergite III and anterolateral corners of tergites IV and V. Ceromata on tergite V indistinct from abdomen orange-brown color. Sternite I dark fuscous, sternite II fulvous, and sternites III–V fulvous tending fuscous medially.

Female. Unknown

Male attractant. Cue-lure.

Etymology. The specific name is a noun in apposition, derived from the Greek *allos* (another) and the species resembles *B. distincta* (Malloch). Previously, *B. pseudo-distincta* (Drew) had been described as a species with similar appearance to *B. distincta*. All three are present in Oceania.

Notes. *Bactrocera allodistincta* was included as *B*. spnSol01 in Doorenweerd et al. (2020).

Bactrocera (Bactrocera) geminosimulata Leblanc & Doorenweerd, sp. nov. http://zoobank.org/49835D53-30EE-47F9-9F64-DE320C2E046A Fig. 6A–E, 9E–G

Type material. *Holotype.* SOLOMON ISLANDS • \mathcal{J} ; Guadalcanal, forest; -9.4045, 159.8665; 120 m; 4–16 Apr. 2018; L. Leblanc, F. Tsatsia leg.; cue-lure baited trap FFS0022; molecular voucher UHIM.ms09156". Deposited in UHIM. *Paratypes.* 13 males. SOLOMON ISLANDS • 4 \mathcal{J} ; Guadalcanal, forest; -9.4072, 159.8664; 153 m; 4–16 Apr. 2018; L. Leblanc, F. Tsatsia leg.; cue-lure baited trap FFS0016; molecular voucher UHIM.ms08673 • 2 \mathcal{J} ; same locality and date as for preceding; -9.4064, 159.8664; 153 m; trap FFS0017 • 2 \mathcal{J} ; same locality and date as for preceding; -9.4064, 159.8665; 139 m; trap FFS0018 • 1 \mathcal{J} ; same locality and date as for preceding; or preceding; -9.4045, 159.8665; 139 m; trap FFS0022 • 2 \mathcal{J} ; same locality and date as for preceding; -9.4038, 159.8646; 103 m; trap FFS0024; molecular voucher UHIM. ms09155) • 2 \mathcal{J} ; same locality and date as for preceding; -9.4026, 159.8695; 57 m; trap FFS0027; molecular vouchers UHIM.ms09153, UHIM.ms09154. Nine of the paratypes are deposited at UHIM, three at WFBM, and one at USNM.

Differential diagnosis. Bactrocera geminosimulata is identical in all points to the sympatric *B. simulata* (Malloch), only distinguished by a subtle difference in wing infuscation in the presence of a light fuscous tinge as a broad, somewhat triangular

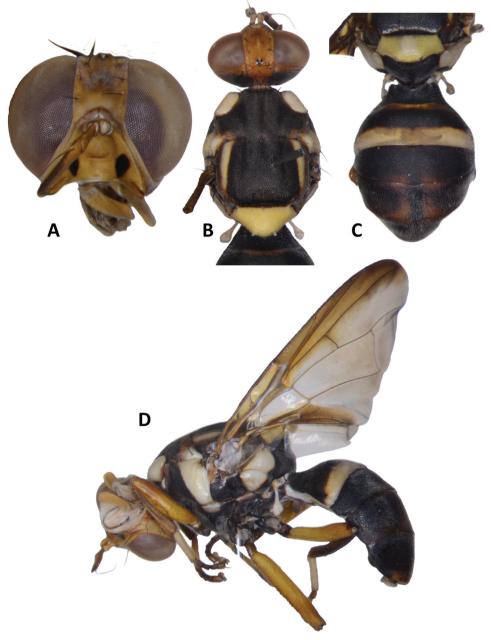


Figure 6. Bactrocera geminosimulata sp. nov. A head B head and scutum C abdomen D lateral view and wing.

area covering much of the middle of the wing, including the areas bordering r-m and dm-cu (Fig. 9E–G); the latter is absent in *B. simulata* (Fig. 9A–D). The new species can be distinguished from *B. bryoniae* (Tryon) by the lighter fuscous tinge of the costal band, a narrower anal streak and the largely to entirely black abdomen, whereas the

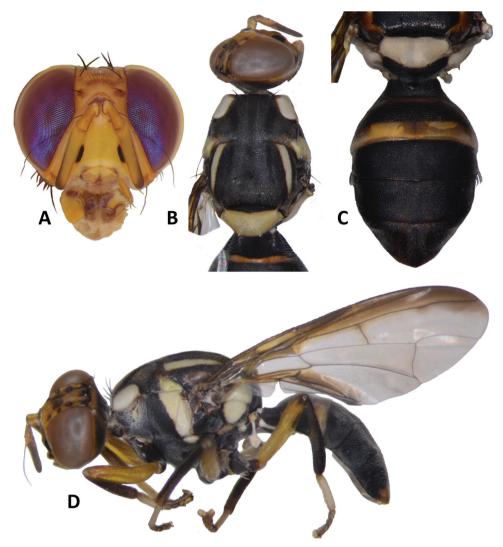
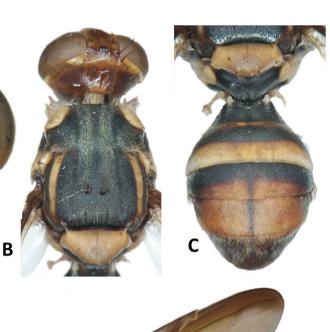


Figure 7. Bactrocera simulata (Malloch) A head B head and scutum C abdomen D lateral view and wing.

abdomen in *B. bryoniae* is orange-brown with a narrow black 'T'-shaped pattern (Fig. 8). *Bactrocera bryoniae* is widespread in Australia and New Guinea but is absent from the Solomon Islands.

Molecular diagnosis. The COI sequences of *B. geminosimulata* [N = 4] are similar to those of *B. bryoniae* [N = 5], but with a minimum of 1.47% pairwise distance. The reference COI dataset only includes *B. bryoniae* from Australia. The COI sequences suggest no close relationship with *B. simulata*, and can be used to reliably distinguish *B. geminosimulata* from *B. simulata*.

Description of adult. Male. *Head* (Fig. 6A). Height 2.02 ± 0.18 (SD) (1.77–2.17) mm. Frons, of even width, 0.98 ± 0.11 (0.83–1.07) mm long and 1.33 ± 0.08





Α

Figure 8. Bactrocera bryoniae (Tryon) A head B head and scutum C abdomen D lateral view and wing.

(1.24–1.43) times as long as broad; generally fulvous; anteromedial hump covered by short red-brown microtrichia; three pairs of black frontal setae present; lunule yellow. Ocellar triangle black. Vertex fulvous with two pairs of black vertical setae. Face fulvous with a pair of large circular black spots in antennal furrows; length 0.62 \pm 0.07 (0.53–0.67) mm. Gena fulvous, with or without a faint dark fuscous subocular spot; red-brown seta present. Occiput dark fuscous and narrowly fulvous along eye margin; a row of 6–8 black postocular setae present behind eye. Antenna with scape and pedicel

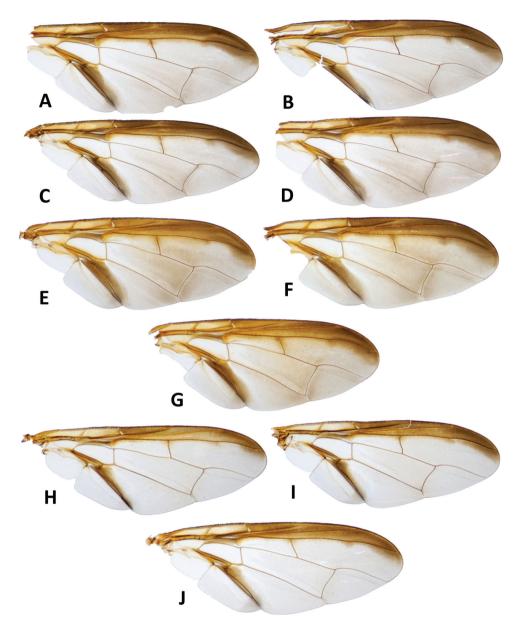


Figure 9. Wings of *Bactrocera simulata* (Malloch) A molecular voucher ms09146 B ms09147 C ms09148
D ms09151, *Bactrocera geminosimulata* sp. nov. E ms09153 F ms09154 G ms09155 *Bactrocera bryoniae* (Tryon) H ms01515 I ms01516 J ms07717.

fulvous and flagellum fulvous with light fuscous lateral surface; a strong red-brown dorsal seta on pedicel; arista fulvous basally and black distally; length of segments: 0.30 \pm 0.03 (0.27–0.33) mm; 0.40 \pm 0.05 (0.33–0.43) mm; 0.95 \pm 0.07 (0.89–1.03) mm.

Thorax (Fig. 6B). Scutum black with small orange-brown markings anterior and posterior to lateral postsutural vitta. Pleural areas black. Yellow markings: postpronotal

lobe; notopleuron; moderately broad paired lateral postsutural vitta, tapering posteriorly and ending before intra-alar seta posteriorly; moderately broad anepisternal stripe with anterior margin straight, ending before anterior notopleural seta dorsally; a large transverse spot on katepisternum below the anepisternal stripe; anterior ¾ of anatergite and katatergite (posteriorly black). Mediotergite black. Scutellum yellow with narrow black basal band. Setae: 1 pair scutellar; 1 pair prescutellar acrostichal; 1 pair intra-alar; 1 pair postalar; 1 pair postsutural supra-alar; 1 pair anepisternal; 2 pairs notopleural; 2 pairs scapular; all setae well developed and black.

Legs (Fig. 6E). Coxae and trochanters black. Remainder of legs fulvous with hind tibia tending fuscous to dark fuscous. Fore femur with a row of long dark dorsal setae. Mid-tibia with an apical black spur.

Wing (Fig. 9E–G). Length 6.4 \pm 0.4 (5.9–6.9) mm; basal costal and costal cells fulvous with microtrichia in posterodistal corner of costal cell; broad dark fuscous costal band confluent with R₄₊₅, ending between R₄₊₅ and medial vein; light fuscous tinge as a broad, somewhat triangular area covering much of the middle of the wing, including the areas bordering r-m and dm-cu (absent in *B. simulata*); broad dark fuscous anal streak; dense aggregation of microtrichia around A₁ + CuA₂; supernumerary lobe moderately developed.

Abdomen (Fig. 6C, D). Oval with tergites not fused; pecten present on tergite III; posterior lobe of surstylus short; abdominal sternite V with a deep concavity on posterior margin. Base of syntergite I+II wider than long. Syntergite I+II black except for yellow along posterior half of and narrowly orange-brown along anterior margin of tergite II. Tergites III–V entirely black or with two broad longitudinal orange-brown areas running from center of tergite IV to posterior margin of tergite V, each side of a broad medial longitudinal dull black stripe. Ceromata on tergite V black. Abdominal sternites black.

Female. Unknown

Male attractant. Cue-lure.

Etymology. The specific name is a noun in apposition, derived from the Latin noun *geminus* (twins) and the epithet of the sympatric and morphologically nearly identical *B. simulata* (Malloch).

Notes. *Bactrocera geminosimulata* was included as *B.* spSol12 in Doorenweerd et al. (2020).

Bactrocera (Parazeugodacus) kolombangarae Leblanc & Doorenweerd, sp. nov. http://zoobank.org/DEED6917-CAC4-4647-BB4F-4454B7F7AE3C Fig. 10A–E

Type material. *Holotype.* SOLOMON ISLANDS • ♂; Kolombangara, forest; -8.0252, 157.1159; 455 m; 9–13 Apr. 2018; L. Leblanc, F. Tsatsia leg.; zingerone baited trap FFS0059. Deposited in UHIM. *Paratypes.* 18 males. SOLOMON ISLANDS • 1 ♂; Guadalcanal forest; -9.4048, 159.8645; 144 m; 4–16 Apr. 2018; L. Leblanc, F. Tsatsia leg.; zingerone baited trap FFS0013 • 1 ♂; Kolombangara, forest; -8.0680, 157.1434;

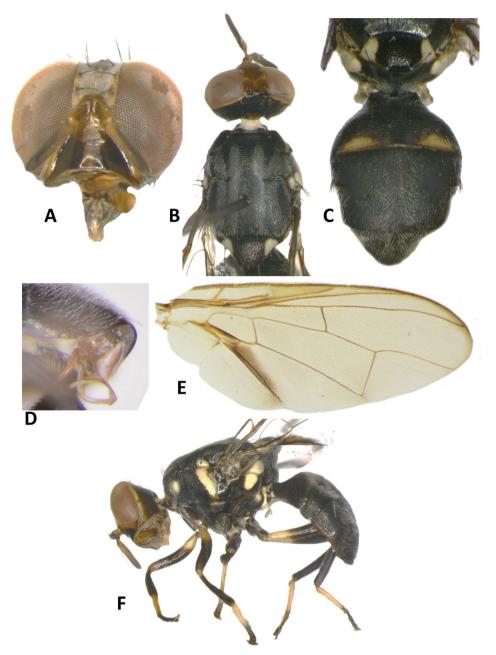


Figure 10. *Bactrocera kolombangarae* sp. nov. **A** head **B** head and scutum **C** abdomen **D** male genitalia **E** wing **F** lateral view.

156 m; 9–13 Apr. 2018; L. Leblanc, F. Tsatsia leg.; zingerone baited trap FFS0044 • 1 \Im ; same locality and date as for preceding; -8.0563, 157.1320; 232 m; trap FFS0046 • 2 \Im ; same locality and date as for preceding; -8.0512, 157.1287; 263 m; trap FFS0047;

molecular vouchers UHIM.ms08663, UHIM.ms08664 • 1 3; same locality and date as for preceding; -8.0479, 157.1262; 267 m; trap FFS0048 • 1 3; same locality and date as for preceding; -8.0364, 157.1186; 331 m; trap FFS0050 • 1 3; same locality and date as for preceding; -8.0297, 157.1166; 403 m; trap FFS0055 • 1 3; same locality and date as for preceding; -8.0273, 157.1160; 433 m; trap FFS0057 • 1 3; same locality and date as for preceding; -8.0260, 157.1156; 446 m; trap FFS0058 • 3 3; same locality and date as for preceding; -8.0238, 157.1157; 464 m; trap FFS0060 • 1 3; same locality and date as for preceding; -8.015, 157.1143; 523 m; trap FFS0068 • 1 3; same locality and date as for preceding; -8.0331, 157.1081; 325 m; trap FFS0071 • 1 3; same locality and date as for preceding; -8.0331, 157.1129; 245 m; trap FFS0073 • 2 3; same locality and date as for preceding; -8.0328, 157.1164; 356 m; trap FFS0075. Nine of the paratypes are deposited at UHIM, five at WFBM, three at USNM, and one at BSI.

Differential diagnosis. Bactrocera kolombangarae appears similar to *B. morula* (Fig. 11), but has two pairs of setae on the scutellum, a narrow anepisternal stripe, and the costal band very narrow and faint beyond the apex of R_{2+3} (Fig. 10). It is also similar to *B. (Parazeugodacus) abbreviata* (Hardy), a species from Southeast Asia. Unlike *B. kolombangarae*, *B. abbreviata* has yellow femora, very short lateral postsutural vitta, and orange-brown medially on abdomen tergites III–V.

Molecular diagnosis. We obtained two COI sequences (UHIM.ms08663, 4) that group with other members of subgenus *Parazeugodacus* in the maximum likelihood tree (Suppl. material 1: Fig. S1). The sequences are closest to *Bactrocera pendleburyi* (Perkins) [N = 11] but at a distance of 3.62%, *B. abbreviata* [N = 29] and *B. morula* [N = 3] are separated with larger distances.

Description of adult. Male. *Head* (Fig. 10A). Height 1.46 ± 0.11 (SD) (1.30-1.70) mm. Frons, of even width, 0.71 ± 0.06 (0.63-0.83) mm long and 1.57 ± 0.08 (1.46-1.77) times as long as broad; dark fulvous and frequently fuscous around orbital setae and on anteromedial hump; latter covered by short red-brown microtrichia; three pairs of black frontal setae present; lunule fulvous. Ocellar triangle black. Vertex dark fulvous with two pairs of black vertical setae. Face varying from mostly black, to lower $\frac{3}{5}$ entirely black with or without traces of dark fulvous medially, and upper $\frac{2}{5}$ dark fulvous, to a pair of very large spots in antennal furrows; length 0.47 ± 0.04 (0.43-0.53) mm. Gena fulvous, with fuscous subocular spot and a black seta. Occiput black and narrowly fulvous along eye margin; a row of 4-6 black postocular setae present behind eye. Antenna with scape dark fulvous, and pedicel and flagellum dark fuscous tending dark fulvous on inner surface; a strong red-brown dorsal seta on pedicel; arista fulvous basally and black apically; length of segments: 0.19 ± 0.02 (0.17-0.20) mm; 0.26 ± 0.03 (0.23-0.30) mm; 0.71 ± 0.06 (0.63-0.83) mm.

Thorax (Fig. 10B). Scutum entirely black with four parallel longitudinal rows of dense silvery microtrichia along entire length and two outer rows starting before no-topleural sutures. Pleural areas black. Yellow markings: notopleuron; sometimes faint marking on posterior margin of postpronotal lobe; narrow anepisternal stripe with anterior margin straight, reaching to mid distance between anterior and posterior

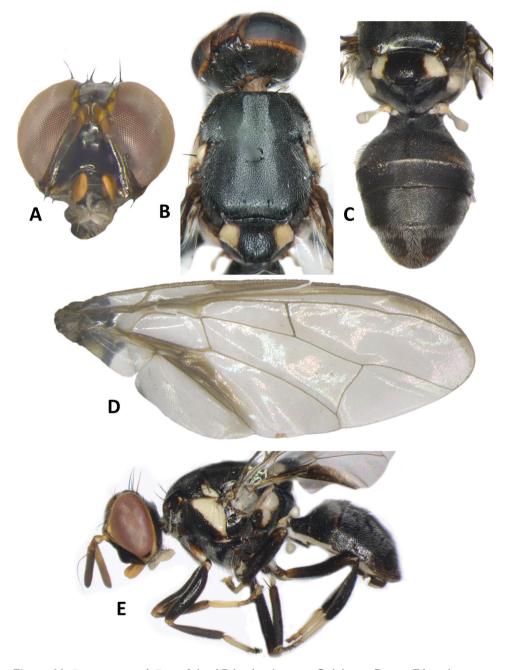


Figure 11. Bactrocera morula Drew A head B head and scutum C abdomen D wing E lateral view.

notopleural setae dorsally; a very small spot on katepisternum below the anepisternal stripe; anterior ¼ of anatergite and anterior half of katatergite (posteriorly black). Mediotergite black. Scutellum black and narrowly yellow anterolaterally. Setae: 2 pairs scutellar; 1 pair prescutellar acrostichal; 1 pair intra-alar; 1 pair postalar; 1 pair postsutural supra-alar; 1 pair anepisternal; 2 pairs notopleural; 2 pairs scapular; all setae well developed and black.

Legs (Fig. 10F). Legs black with yellow at basal 2/5 of fore and hind femora and basal 1/6 of mid femur, and yellow fore basitarsus and mid and hind tarsi. Fore femur with a row of long pale dorsal setae. Mid-tibia with an apical black spur.

Wing (Fig. 10E). Length 4.9 ± 0.3 (4.5–5.6) mm; basal costal and costal cells hyaline with microtrichia in posterodistal corner of costal cell; narrow faint fuscous costal band confluent with R_{2+3} , remaining narrow and ending shortly past the apex of R_{2+3} ; and moderately broad anal streak; remainder of wing hyaline; dense aggregation of microtrichia around A_1 + CuA₃; supernumerary lobe weakly developed.

Abdomen (Fig. 10C, D). Oval with tergites not fused; pecten present on tergite III; posterior lobe of surstylus short; abdominal sternite V with a shallow concavity on posterior margin. Base of syntergite I+II wider than long. Tergites entirely black except for elongate creamy yellow short sublateral bands along posterior margin of tergite II. Ceromata on tergite V black. Abdominal sternites dark except for yellow sternite II.

Female. Unknown

Male attractant. Zingerone.

Etymology. This species epithet is a noun in genitive case, derived from the locality where the majority of the specimens were collected; Kolombangara Island.

Notes. This species belongs to the subgenus *Parazeugodacus* as defined by Hancock and Drew (2015), based on morphological characters (shallow posterior concavity on male sternite V, posterior lobe of surstylus short, postpronotal seta absent, postsutural supra-alar, prescutellar acrostichal and two pairs of scutellar setae present, costal band very narrow and nearly indistinct). Its COI sequences also suggest closest affinity with other members of *Parazeugodacus* (Suppl. material 1: Fig. S1). *Bactrocera kolombangarae* was included as *B.* spnSol06 in Doorenweerd et al. (2020).

Bactrocera (*Bactrocera*) *quasienochra* Leblanc & Doorenweerd, sp. nov. http://zoobank.org/3A13A2D0-6F79-4338-B501-887EEA24C356 Fig. 12A–E

Type material. *Holotype.* SOLOMON ISLANDS • ♂; Guadalcanal, forest; -9.4064, 159.8671; 145 m; 4–16 Apr. 2018; L. Leblanc, F. Tsatsia leg.; cue-lure baited trap FFS0018; molecular voucher UHIM.ms08789. Deposited in UHIM.

Differential diagnosis. *Bactrocera quasienochra* (Fig. 12) is similar to *B. enochra* (Drew) (Fig. 13). It differs by the absence of broad black lateral markings on abdomen tergites III–V, and the narrower lateral postsutural vitta, ending before intra-alar seta.

Molecular diagnosis. We sequenced the holotype for COI, and its sequence is closest to an undescribed species from Malaysia (*B.* spMalaysia11 in Doorenweerd et al. (2020)) at 11.19% pairwise distance. The *B. quasienochra* sequence has an even greater distance to those of *B. enochra* [N = 6].

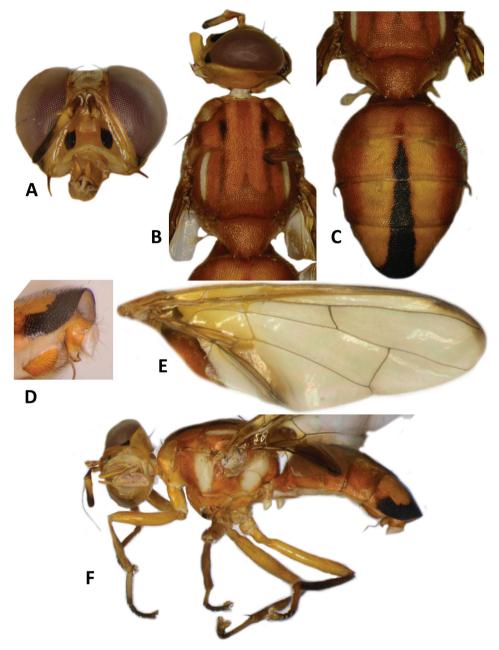


Figure 12. *Bactrocera quasienochra* sp. nov. **A** head **B** head and scutum **C** abdomen **D** male genitalia **E** wing **F** lateral view.

Description of adult. Male. *Head* (Fig. 12A). Height 1.83 mm. Frons, of even width, 0.93 mm long and 1.56 times as long as broad; dark fulvous and narrowly fulvous anterolaterally; anteromedial hump covered by short red-brown microtrichia; three pairs of dark fuscous frontal setae present; lunule fulvous. Ocellar triangle black.

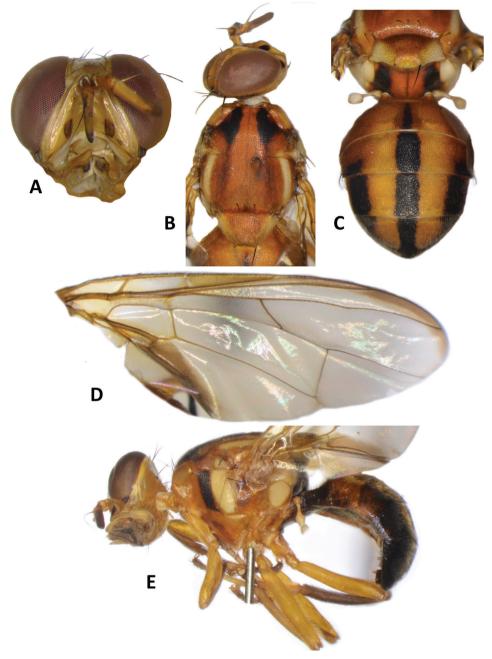


Figure 13. Bactrocera enochra (Drew) A head B head and scutum C abdomen D wing E lateral view.

Vertex fulvous with two pairs of dark fuscous vertical setae. Face fulvous with a pair of large oval black spots in antennal furrows; length 0.53 mm. Gena fulvous, with large dark fuscous subocular spot and a red-brown seta. Occiput fulvous and dark fulvous behind vertex; row of postocular setae weakly developed, with ca. four nearly indistinct

setae. Antenna with scape and pedicel dark fulvous and flagellum fulvous with lateral surface and inner apical half dark fuscous; a strong fulvous dorsal seta on pedicel; arista fulvous basally and black distally; length of segments: 0.27 mm; 0.30 mm; 0.87 mm.

Thorax (Fig. 12B). Scutum orange-brown with two short sublateral dark fuscous markings anterior to notopleural suture, and continued posteriorly as parallel lines formed by black microtrichia. Pleural areas orange-brown. Notopleuron light fulvous. Yellow markings: posterior half of postpronotal lobe (anteriorly orange-brown); narrow paired parallel-sided lateral postsutural vitta, slightly tapered posteriorly and ending before intra-alar seta; moderately broad anepisternal stripe with anterior margin straight, reaching to mid distance between anterior and posterior notopleural setae dorsally; anterior ²/₃ anatergite and katatergite (posteriorly orange-brown). Mediotergite orange-brown. Scutellum orange-brown, and yellow on anterolateral surface and ventrally. Setae: 1 pair scutellar; prescutellar acrostichal absent; 1 pair intra-alar; 1 pair postalar; 1 pair postsutural supra-alar; 1 pair anepisternal; 2 pairs notopleural; 1 pair scapular (lateral position); all setae well developed and fuscous.

Legs (Fig. 12F). Legs entirely fulvous with hind tibia tending fuscous on dorsal surface. Fore femur with a row of long fulvous dorsal setae. Mid-tibia with an apical black spur.

Wing (Fig. 12E). Length 6.7 mm; basal costal and costal cells fuscous with microtrichia in posterodistal corner of costal cell; narrow fuscous costal band confluent with R_{2+3} , not expanded at apex, and ending mid distance between apex of R_{4+5} and medial vein, and broad fuscous anal streak; remainder of wing hyaline; dense aggregation of microtrichia around A_1 + CuA₂; supernumerary lobe weakly developed.

Abdomen (Fig. 12C, D). Elongate-oval with tergites not fused; pecten present on tergite III; posterior lobe of surstylus short; abdominal sternite V with a deep concavity on posterior margin. Base of syntergite I+II wider than long. All tergites orange-brown with a medial longitudinal black stripe gradually broadened from base of tergite III and extended apically along the entire lateral margins of tergite V except their bases. Ceromata on tergite V indistinct from abdomen orange-brown color. Abdominal sternites fulvous.

Female. Unknown

Male attractant. Cue-lure.

Etymology. The species name is a noun in apposition, derived from the Latin adverb *quasi* (just as if) used in conjunction with the epithet of the species it closely resembles; *B. enochra*.

Notes. *Bactrocera quasienochra* was included as *B*. spnSol03 in Doorenweerd et al. (2020).

Bactrocera (*Bactrocera*) *tsatsiai* Leblanc & Doorenweerd, sp. nov. http://zoobank.org/8B4AC740-8648-44ED-87C8-84056641FEC4 Fig. 14A–I, 15

Type material. *Holotype.* SOLOMON ISLANDS • ∂; Guadalcanal, forest; -9.4053, 159.8664; 139 m; 4–16 Apr. 2018; L. Leblanc, F. Tsatsia leg.; zingerone baited trap

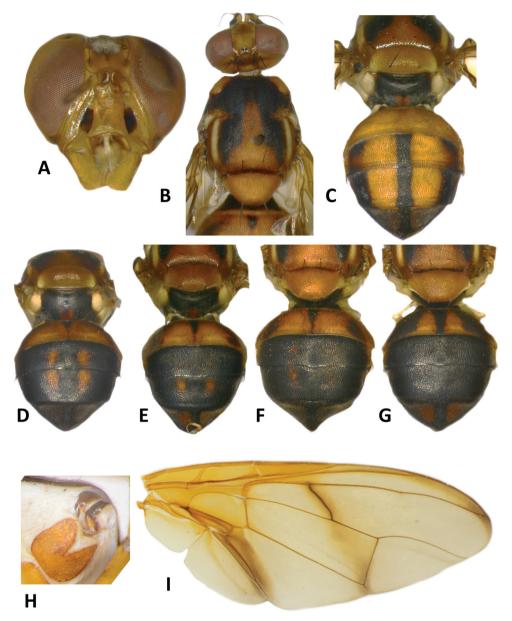


Figure 14. *Bactrocera tsatsiai* sp. nov. A head B head and scutum C-G abdomen variants H male genitalia I wing.

FFS0021. Deposited in UHIM. *Paratypes.* 28 males. SOLOMON ISLANDS • 1 \bigcirc Guadalcanal, forest; -9.4041, 159.8628; 153 m; 4–16 Apr. 2018; L. Leblanc, F. Tsatsia leg.; zingerone baited trap FFS0011 • 2 \bigcirc ; same locality and date as for preceding; -9.4064, 159.8644; 167 m; trap FFS014 • 1 \bigcirc ; same locality and date as for preceding; -9.4067, 159.8647; 167 m; trap FFS015 • 1 \bigcirc ; same locality and date as for preceding; -9.4067,

-9.4069, 159.8664; 153 m; trap FFS0017 • 1 &; same locality and date as for preceding: -9.4059, 159.8672; 133 m; trap FFS0019 • 2 d; same locality and date as for preceding; -9.4035, 159.8681; 85 m; trap FFSo026; molecular voucher UHIM.ms08671 • 1 (; Kolombangara, forest; -8.0312, 157.1160; 348 m; 9–13 Apr. 2018; L. Leblanc, F. Tsatsia leg.; zingerone baited trap FFS0053 \bullet 3 $\textcircled{\circ}$; same locality and date as for preceding; -8.0297, 157.1166; 403 m; trap FFS0055 • 1 Å; same locality and date as for preceding; -8.0283, 157.1159; 426 m; trap FFSo056 • 3 (); same locality and date as for preceding; -8.0218, 157.1150; 491 m; trap FFSo062 • 2 3; same locality and date as for preceding; -8.0200, 157.1143; 508 m; trap FFS0063 • 2 3; same locality and date as for preceding; -8.0190, 157.1133; 520 m; trap FFSo064 • 1 Å; same locality and date as for preceding; -8.0181, 157.1129; 518 m; trap FFS0065 • 1 %; same locality and date as for preceding; -8.0181, 157.1134; 526 m; trap FFS0066 • 1 d; same locality and date as for preceding; -8.0157, 157.1118; 506 m; trap FFSo067 • 1 3; same locality and date as for preceding; -8.0150, 157.1143; 523 m; trap FFSo068 • 1 Å; same locality and date as for preceding; -8.0327, 157.1159; 333 m; trap FFS0070 • 2 Å; same locality and date as for preceding; -8.0356, 157.1193; 352 m; trap FFSo077 • 1 Å; same locality and date as for preceding; -8.0357, 157.1200; 352 m; trap FFS0078. Fifteen of the paratypes are deposited at UHIM, seven at WFBM, four at USNM, and two at BSI.

Differential diagnosis. The broad orange-brown medial marking on the scutum uniquely defines *Bactrocera tsatsiai* within the genus, where all other species have either a yellow mark or no mark.

Molecular diagnosis. We obtained two COI sequences that are most similar to *Bactrocera hantanae* Tsuruta & White but at 10.79% pairwise distance.

Description of adult. Male. *Head* (Fig. 14A). Height 2.00 ± 0.09 (SD) (1.87–2.13) mm. Frons, of even width, 0.99 ± 0.04 (0.93-1.07) mm long and 1.56 ± 0.06 (1.47-1.63) times as long as broad; fulvous with red-brown microtrichia on anteromedial hump; three pairs of black frontal setae present; lunule yellow. Ocellar triangle black. Vertex fuscous with two pairs of black vertical setae. Face fulvous with a pair of large oval black spots in antennal furrows; length 0.60 ± 0.04 (0.53-0.67) mm. Gena fulvous, with small dark fuscous subocular spot and a black seta. Occiput fulvous; a row of 6–9 black postocular setae present behind eye. Antenna with scape and pedicel fulvous and flagellum fuscous with fulvous on inner surface; a strong black dorsal seta on pedicel; arista fulvous basally and black distally; length of segments: 0.25 ± 0.03 (0.20-0.30) mm; 0.32 ± 0.03 (0.27-0.37) mm; 0.87 ± 0.05 (0.80-0.93) mm.

Thorax (Fig. 14B). Scutum dark fuscous with orange-brown ventral to and narrowly anterior to lateral postsutural vitta, narrowly englobing notopleural suture, between postpronotal lobe and notopleuron, and as a medial band starting before notopleural suture and enlarged posteriorly to cover entire posterior margin region of scutum. Pleural areas black except orange-brown anepisternum and proepisternum. Yellow markings: postpronotal lobe (or may be anteriorly to entirely orange-brown), notopleuron; moderately broad paired parallel-sided lateral postsutural vitta ending at intra-alar seta posteriorly; moderately broad anepisternal stripe with anterior margin



Figure 15. Bactrocera tsatsiai sp. nov. lateral view.

slightly convex, reaching to mid distance between anterior and posterior notopleural setae dorsally; a small transverse spot on katepisternum below the anepisternal stripe; anterior ³/₄ of anatergite and katatergite (posteriorly black). Mediotergite black. Scutellum orange-brown, and yellow ventrally and narrowly on dorsolateral surface. Setae: 1 pair scutellar; 1 pair prescutellar acrostichal; 1 pair intra-alar; 1 pair postalar; 1 pair postsutural supra-alar; 1 pair anepisternal; 2 pairs notopleural; 2 pairs scapular; all setae well developed and black.

Legs (Fig. 15). All legs entirely fulvous with hind femur and fore tarsomeres II–IV fuscous. Fore femur with a row of long pale dorsal setae. Mid-tibia with an apical black spur.

Wing (Fig. 14I). Length 7.1 \pm 0.3 (6.6–7.5) mm; basal costal and costal cells fuscous with microtrichia in posterodistal corner of costal cell; light fuscous costal band confluent with R₂₊₃, not expanded at apex and ending mid distance between apex of R₄₊₅ and medial vein, a diffuse broad fuscous cross band along r-m crossvein, continuing in straight line through discal medial (dm) cell and reaching wing margin at level of CuA₁, and a broad fuscous anal streak; remainder of wing hyaline; dense aggregation of microtrichia around A₁ + CuA₂; supernumerary lobe moderately developed. **Abdomen** (Fig. 14C–H). Oval with tergites not fused; pecten present on tergite III; posterior lobe of surstylus short; abdominal sternite V with a deep concavity on posterior margin. Base of syntergite I+II wider than long. Syntergite I+II with tergite I black and tergite II orange-brown with or without a small basal black triangular and two small sublateral black markings. Tergites III–V orange-brown with broad medial longitudinal black stripe reaching apex of tergite V and extended apically along entire lateral margins of tergite V, and two broad sublateral stripes covering tergite III (may be interrupted on that tergite) and continuing on tergite IV and along lateral margins on tergite V. Dark marking variable and may cover almost all of tergites III–V (Fig. 14C–G). Ceromata on tergite V dark fuscous. Abdominal sternites fulvous.

Female. Unknown

Male attractant. Zingerone.

Etymology. The epithet *tsatsiai* is a noun in genitive case, referring to the personal name Francis Tsatsia, a long-time colleague, friend, co-author of the present publication, and currently the director of Biosecurity Solomon Islands.

Notes. Bactrocera tsatsiai was included as B. spnSol05 in Doorenweerd et al. (2020).

Bactrocera (Bactrocera) vargasi Leblanc & Doorenweerd, sp. nov.

http://zoobank.org/BC8E46E7-1917-412C-AF67-7A487BDEFAFE Fig. 16A–F

Type material. *Holotype.* SOLOMON ISLANDS • ♂; Kolombangara, forest; -8.0563, 157.1320; 232 m; 9-13 Apr. 2018; L. Leblanc, F. Tsatsia leg.; zingerone baited trap FFS0046. Deposited in UHIM. Paratypes. 42 males. SOLOMON ISLANDS • 2 3; Guadalcanal, forest; 4-16-iv-2018; L. Leblanc, F. Tsatsia leg.; -9.4041, 159.8628; 153 m; zingerone trap FFS0011 • 2 $\vec{\circ}$; same locality and date as for preceding; -9.4045, 159.8644; 142 m; trap FFS0012 • 2 3; same locality and date as for preceding; -9.4048, 159.8645; 144 m; trap FFS0013 • 2 3; same locality and date as for preceding; -9.4064, 159.8644; 167 m; trap FFSo14 • 3 Å; same locality and date as for preceding; -9.4067, 159.8647; 167 m; trap FFSo015; molecular vouchers UHIM. ms08665, UHIM.ms08666, UHIM.ms08667 • 2 👌; same locality and date as for preceding; -9.4069, 159.8664; 153 m; trap FFSo017 • 4 ♂; same locality and date as for preceding; -9.4064, 159.8671; 145 m; trap FFSo018 • 2 3; same locality and date as for preceding; -9.4059, 159.8672; 133 m; trap FFSo019 • 2 3; same locality and date as for preceding; -9.4055, 159.8665; 145 m; trap FFSo020 • 1 Å; same locality and date as for preceding; -9.4053, 159.8664; 139 m; trap FFS0021 • 3 🖑; same locality and date as for preceding; -9.4040, 159.8652; 125 m; trap FFS0023 • 3 d; same locality and date as for preceding; -9.4038, 159.8646; 103 m; trap FFS0024 • 1 3; same locality and date as for preceding; -9.4039, 159.8673; 103 m; trap FFS0025 • 2 Å; same locality and date as for preceding; -9.4035, 159.8681; 85 m; trap FFSo026 • 2 Å; same locality and date as for preceding; -9.4026, 159.8695; 57 m; trap FFS0027 • 1 ♂; same locality and date as for preceding; -9.400, 159.8700; 50 m; trap FFSo029 • 2 ; Kolombangara, forest; -8.0563, 157.1320; 232 m; 9–13 Apr. 2018; L. Leblanc,

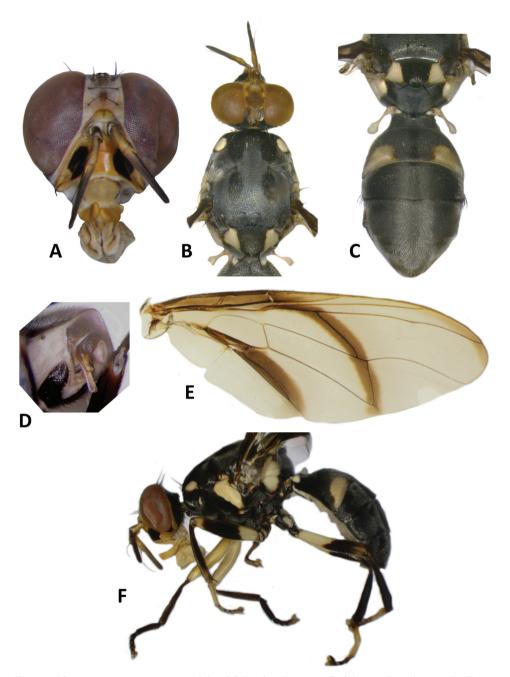


Figure 16. *Bactrocera vargasi* sp. nov. A head B head and scutum C abdomen D male genitalia E wing F lateral view.

F. Tsatsia leg.; zingerone baited trap FFS0046 • 2 3; same locality and date as for preceding; -8.0479, 157.1262; 267 m; trap FFS0048 • 1 3; same locality and date as for preceding; -8.0306, 157.1168; 389 m; trap FFS0054 • 1 3; same locality and date as

for preceding; -8.0252, 157.1159; 455 m; trap FFS0059 • 1 3; same locality and date as for preceding; -8.0328, 157.1164; 356 m; trap FFS0075 • 1 3; same locality and date as for preceding; -8.0395, 157.1237; 308 m; trap FFS0079. 29 of the paratypes are deposited at UHIM, seven at WFBM, four at USNM, and two at BSI.

Differential diagnosis. The overall appearance and specifically the wing of *B. var*gasi (Fig. 16E) is very similar to that of *B. frauenfeldi* (Schiner) (Fig. 17) [Solomon Island populations], *B. trilineola* Drew and *B. parafrauenfeldi* Drew [all three are members of the morphological *B. frauenfeldi* complex], but *B. vargasi* differs from *B. triline*ola and *B. parafrauenfeldi* in having a nearly entirely black abdomen (Fig. 16), and can be separated from *B. frauenfeldi* in lacking lateral postsutural yellow vitta.

Molecular diagnosis. We sequenced three specimens which have COI sequences closest to *B. quasiinfulata* Drew & Romig at 7.24% minimum pairwise distance. The maximum intraspecific distance is 1.2%. Sequences of the morphologically similar *B. frauenfeldi* and *B. trilineola* were also included in the reference dataset but are highly dissimilar to *B. vargasi* with >8% pairwise distance. *Bactrocera parafrauenfeldi* was not included in the reference set but is presumed to be closely related to *B. trilineola* (Drew 1989).

Description of adult. Male. *Head* (Fig. 16A). Height 1.61 ± 0.22 (SD) (1.33-1.93) mm. Frons, of even width, 0.83 ± 0.08 (0.67-0.93) mm long and 1.63 ± 0.11 (1.43-1.85) times as long as broad; fulvous and narrowly yellow along eye margin; anteromedial hump covered by short red-brown microtrichia; three pairs of black frontal setae present; lunule yellow. Ocellar triangle black. Vertex black with yellow spot behind ocellar triangle and two pairs of black vertical setae. Face fulvous with a pair of very large circular black spots in antennal furrows; length 0.51 ± 0.06 (0.43-0.63) mm. Gena fulvous, with small dark fuscous subocular spot and a red-brown seta. Occiput black and narrowly fulvous along eye margin; a row of 6-9 black postocular setae present behind eye. Antenna with scape and pedicel fulvous and flagellum dark fuscous tending dark fulvous on inner surface; a strong red-brown dorsal seta on pedicel; arista fulvous basally and black distally; length of segments: 0.31 ± 0.03 (0.27-0.33) mm; 0.36 ± 0.03 (0.33-0.40) mm; 0.88 ± 0.09 (0.73-1.00) mm.

Thorax (Fig. 16B). Scutum entirely black with dense silvery microtrichia on all scutum except two broad parallel longitudinal shining black areas interrupted at level of notopleural suture. Pleural areas black. Yellow markings: posterior half of postpronotal lobe (anteriorly fuscous); notopleuron; moderately broad anepisternal stripe with anterior margin convex, reaching to mid distance between anterior and posterior notopleural setae dorsally; a very small transverse spot on katepisternum below the anepisternal stripe; anterior $\frac{3}{5}$ of anatergite and katatergite (posteriorly black). Mediotergite black. Scutellum broadly black medially and yellow laterally. Setae: 1 pair scutellar; 1 pair prescutellar acrostichal; 1 pair intra-alar; 1 pair postalar; 1 pair postsutural supra-alar; 1 pair anepisternal; 2 pairs notopleural; 2 pairs scapular; and setae well developed and black.

Legs (Fig. 16F). Legs black with yellow fore femur, basal ²/₅ of mid and hind femur, and mid and hind tarsi. Fore femur with a row of long pale dorsal setae. Mid-tibia with an apical black spur.

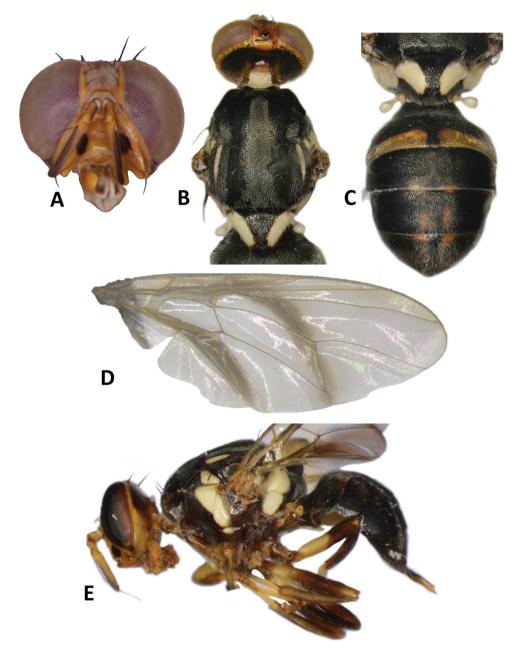


Figure 17. *Bactrocera frauenfeldi* (Schiner) **A** head **B** head and scutum **C** abdomen, male **D** wing **E** lateral view, female.

Wing (Fig. 16E). Length 6.2 ± 0.6 (5.3–6.9) mm; basal costal and costal cells dark fuscous with microtrichia covering both cells; faint narrow fuscous costal band confluent with R_{2+3} , remaining narrow to end shortly past the apex of R_{2+3} ; dark fuscous

straight band across r-m and dm-cu veins and reaching wing margin; broad dark fuscous anal streak; remainder of wing hyaline; dense aggregation of microtrichia around $A_1 + CuA_2$; supernumerary lobe weakly developed.

Abdomen (Fig. 16C). Oval with tergites not fused; pecten present on tergite III; posterior lobe of surstylus short; abdominal sternum V with a deep concavity on posterior margin. Base of syntergite I+II wider than long. Tergites entirely black with yellow lateral bands along posterior margin of tergite II. Ceromata on tergite V black. Abdominal sternites black.

Female. Unknown.

Etymology. We proudly name this species to honor the famous fruit fly ecologist Roger I. Vargas (1947–2018) (Stark et al. 2018). The species name *vargasi* is a noun in genitive case. Roger and LL collaborated extensively on projects during years spent in the South Pacific Islands. Roger brought LL to Hawaii in 2003 to continue working on fruit flies, and he secured funding and provided guidance that allowed LL to obtain a PhD title in 2010.

Male attractant. Zingerone.

Notes. Bactrocera vargasi was included as B. spnSol07 in Doorenweerd et al. (2020).

Key to Dacine fruit fly species of Solomon Islands

This is a modified version of the key published by Drew and Romig (2001). We include for each species subgenus assignment and information on male lure attraction and host fruit (after Leblanc et al. 2012), whenever known.

- 1 Elongate large wasp-like fly; antenna longer than height of face; abdomen elongate and petiolate (base of syntergite I+II longer than wide), with a pronounced hump on tergite V in lateral view (unique to that species) (Fig. 18A–D) (cue-lure) (pest of cucurbit fruits) *Dacus (Mellesis) solomonensis* (Malloch)
- 2 Wing without complete costal band, with large faint light fuscous spot covering apex, and a swelling (bulla) in CuA₂ cell; medial postsutural vitta large and triangular (Fig. 18 E–H) (bred from *Terminalia catappa* and *Gnetum gnemon*)
- males of *B*. (*Bulladacus*) *penefurva* Drew
 Wing with complete costal band, although sometimes noticeably paler beyond apex of R₁, with marking (when present) not as large spot at apex, and bulla absent
- (except in males of *B. pacificae*); medial postsutural vitta present or absent.......**3**

4	Scutum with medial postsutural vitta (Figs 18I–20A)5
_	Scutum without medial postsutural vitta (Figs 20D–24D)11
5	Prescutellar acrostichal seta absent; postsutural supra-alar seta present or absent 6
_	Prescutellar acrostichal and postsutural supra-alar seta present
6	Postsutural supra-alar seta absent; abdomen fulvous with broad dark fuscous lat-
-	eral stripes on tergites III–V (Fig. 18I–K) (zingerone)
	B. (Tetradacus) pagdeni (Malloch)
_	Postsutural supra-alar seta present; abdomen with black spot on tergite V (Fig.
	18M) or with narrow medial and lateral stripes (Fig. 19B)
7	Postpronotal lobe and notopleuron joined by lateral yellow band; wing with nar-
,	row pale infuscation along crossveins r-m and dm-cu; abdominal tergites entirely
	red-brown except for black spot in center of tergite V (Fig. 18L–N) (cue-lure)
	<i>Z. (Zeugodacus) univittatus (Drew)</i>
_	Postpronotal lobe and notopleuron not joined by yellow band; wing with fuscous
	tint throughout, broad fuscous costal band to R_{4+5} , narrow infuscation along r-m
	crossvein and broad infuscation along dm-cu crossvein; abdominal tergites III–
	V with narrow medial and lateral longitudinal black stripes (Fig. 19A–C) (cue-
	lure)
8	Scutum glossy black; Z-shaped fuscous pattern across wing (Fig. 19D–F) (cue-
0	lure)
_	Scutum basically red-brown; wing with infuscation on one or both crossveins,
0	9 Sentere and have a still date mathing and a sentence and in a star
9	Scutum red-brown with small dark markings and a very narrow medial postsu-
	tural vitta; scutellum with one or two pairs of setae (Fig. 19G–I) (cue-lure, zinge-
	rone) (pest of cucurbit flowers and fruits)
_	Scutum entirely red-brown or red-brown with large dark markings and a broader
	medial postsutural vitta (Figs 19J, 20A); scutellum with two pairs of setae10
10	Wing with infuscation on dm-cu crossvein only; scutum entirely red-brown; ab-
	dominal tergites III–V red-brown without a distinct dark 'T'-shaped pattern (Fig.
	19J–L) Z. (Javadacus) fuscipennulus (Drew & Romig)
_	Wing with infuscation on both crossveins; scutum red-brown with large black
	markings; abdominal tergites III–V red-brown with a black 'T'-shaped pattern
	(Fig. 20A–C) (cue-lure)Z. (Javadacus) abdoangustus (Drew)
11	Infuscation on wing on one crossvein only (Fig. 20F, J, N)12
—	Infuscation on wing more extensive, as a very broad pattern across most of mem-
	brane, a recurved band, or one or more transverse bands14
12	Lateral postsutural vitta very short and narrowing posteriorly to end well before
	intra-alar seta (Fig. 20D–F) (methyl eugenol)
_	Lateral postsutural vitta broad, parallel sided (or with only a slight narrowing
	posteriorly) and ending at intra-alar seta13

13	An episternal stripe reaching to postpronotal lobe dorsally; abdominal tergites III – V red-brown with a black 'T'-shaped and broad lateral black margins (Fig. 20G–I)
	(cue-lure) (bred from <i>Alpinia purpurata</i>)
-	Anepisternal stripe reaching to anterior notopleural seta dorsally; abdominal ter-
	gites mostly black (Fig. 24K–N) (methyl eugenol)
14	Scutellum with a black triangular dorsal marking or with an apical dark spot .15
-	Scutellum yellow or orange-brown, except for a narrow dark basal band20
15	Costal band pale and indistinct beyond apex of R_1 ; a narrow transverse fuscous
	band across wing16
—	Costal band distinct for entire length; infuscation across wing as a single band or a recurved band
16	Lateral postsutural vitta present; abdomen orange-brown with a broad medial and
10	
	two broad lateral black bands along tergites III–V (Fig. 17A–E) (cue-lure, zinge-
	rone) (polyphagous fruit pest bred from 28 host species in Solomon Islands)
-	Lateral postsutural vitta absent; abdomen entirely black except yellow lateral
	bands along posterior margin of tergite II (Fig. 16A–F) (zingerone)
. –	<i>B. (Bactrocera) vargasi</i> sp. nov.
17	Infuscation across wing as a recurved band (Fig. 21C, F)
_	Infuscation across wing as a single band (Figs 21I, 22C)
18	Single U-shaped band across wing (Fig. 21A–C) (methyl eugenol)
10	
10	
_	Broad recurved band across center of wing and a narrow transverse band across
_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart
- 19	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel-
_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto-
_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow;
_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral
_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)
_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>hollingsworthi</i> Drew & Romig
_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)
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_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>hollingsworthi</i> Drew & Romig Scutellum yellow with at most an apical dark spot; postpronotal lobe yellow;
_	<i>B.</i> (<i>Bactrocera</i>) reclinata Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) longicornis Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)
_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)
_	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)
_ 19 _	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>hollingsworthi</i> Drew & Romig Scutellum yellow with at most an apical dark spot; postpronotal lobe yellow; postpronotal lobe and notopleuron joined by a narrow yellow band; lateral post- sutural vitta well developed; abdomen tergites III–V orange-brown with a black medial longitudinal stripe and well defined black lateral markings (Fig. 22A–C) (cue-lure)
_ 19 _	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)
_ 19 _	<i>B.</i> (<i>Bactrocera</i>) <i>reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B.</i> (<i>Bactrocera</i>) <i>longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)
_ 19 _ 20 _	<i>B. (Bactrocera) reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B. (Bactrocera) longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)
_ 19 _ 20 _	<i>B. (Bactrocera) reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B. (Bactrocera) longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)
- 19 - 20 - 21	<i>B. (Bactrocera) reclinata</i> Drew Broad recurved band across center of wing and a narrow transverse band across apex (Fig. 21D–F) (cue-lure) <i>B. (Bactrocera) longicornis</i> Macquart Scutellum with a broad medial longitudinal black stripe; postpronotal lobe yel- low except for anterior third dark fuscous to black; postpronotal lobe and noto- pleuron not joined by a yellow band; lateral postsutural vitta short and narrow; abdomen tergites III–V orange-brown with small irregularly shaped sublateral markings and a narrow medial longitudinal stripe (Fig. 21G–I) (cue-lure)

23	Wing membrane almost entirely fuscous; abdomen tergite II largely fulvous, con- trasting with the black tergites III–V (Fig. 22J–L) (methyl eugenol)
	B. (Bactrocera) pepisalae (Froggatt)
_	Wing membrane colorless with distinct fuscous markings; abdomen entirely
	black, or with at most an orange-brown band along posterior margin of tergite II
24	r-m crossvein strongly oblique; broad dark fuscous band across wing from cos-
	tal band to hind margin, enclosing both crossveins; legs entirely fulvous (Fig.
	23A-C) (methyl eugenol)B. (Bactrocera) obliquivenosa Drew & Romig
_	r-m crossvein not oblique; transverse fuscous band across wing broad and cover-
	ing more than outer half of discal medial cell; legs fulvous with apical half of mid
	and hind femur black (Fig. 23D–F) (methyl eugenol)
	B. (Bactrocera) biarcuata (Walker)
25	Z-shaped fuscous pattern across wing
_	Single fuscous band of variable shape across wing27
26	Lateral postsutural vitta short and tapering posteriorly; wing markings dark fus-
	cous; lateral and medial longitudinal black stripes on abdominal tergites III-V
	sometimes joined across base of tergite III (Fig. 23G-I) (cue-lure)
	B. (Bactrocera) nigrescentis (Drew)
_	Lateral postsutural vitta broad, parallel sided and reaching to intra-alar seta; wing
	markings pale fuscous; lateral and medial longitudinal black stripes on abdominal
	tergites III-V not joined (Fig. 23J-L) (cue-lure) (bred from Pycnarrhena ozantha
	in Vanuatu)
27	Large species (body length 11 mm or more); transverse fuscous band across wing
	broad and covering more than outer half of discal medial cell (Fig. 24A-C) (me-
	thyl eugenol)
_	Moderately sized species (body length 9 mm or less); transverse fuscous band
	across wing of medium width, covering outer third of discal medial cell28
28	Wing crossband dark fuscous and broad; costal band confluent with $\mathrm{R}_{\!\!\!\!_{4+5}}$ and
	greatly expanded at apex of wing (Fig. 24D-F) (cue-lure) (bred from Burckela
	sp.)B. (Bactrocera) decumana (Drew)
-	Wing crossband light fuscous; costal band not greatly expanded at apex of wing. 29
29	Scutum dark fuscous to black with a broad orange-brown medial stripe, starting
	before notopleural suture and enlarged posteriorly to cover entire posterior mar-
	gin region of scutum; scutellum largely orange-brown, and yellow ventrally and
	narrowly on dorsolateral surface; anepisternal stripe moderately broad, reaching
	to mid distance between anterior and posterior notopleural setae dorsally (Figs
	14A–I, 15) (zingerone) B. (Bactrocera) tsatsiai sp. nov.
-	Scutum predominantly to entirely black, at most narrowly orange-brown laterally
	and posteriorly; scutellum yellow; anepisternal stripe broad, almost reaching to
	anterior notopleural seta dorsally
30	Scutum entirely black, except for yellow postpronotal lobe, notopleuron and lateral postsutural vitta; abdominal tergites III–V with moderately broad medial

	longitudinal black stripe and lateral black markings on tergite IV not narrowed posteriorly; fuscous crossband on wing clearly defined (Fig. 4A–E) (cue-lure) B. (Bactrocera) pseudodistincta (Drew)
_	Scutum predominantly black with orange-brown laterally and posteriorly; ab- dominal tergites III–V with narrower medial longitudinal black stripe and lateral black markings on tergite IV narrowed posteriorly; fuscous crossband on wing
31	diffuse (Fig. 3A–F) (cue-lure)
_	Lateral postsutural vitta absent
32	Scutellum entirely yellow; abdominal tergites orange-brown with a narrow me-
	dian longitudinal stripe on tergites III–V (Fig. 24G–I) (cue-lure) (bred from Cer-
	bera spp and Antiaris toxicaria in Vanuatu)B. (Bactrocera) minuta (Drew)
—	Scutellum yellow and broadly black medially; abdominal tergites black
33	Scutellum with two pairs of setae; anepisternal stripe narrow and reaching to mid
	distance between anterior and posterior notopleural setae dorsally (Fig. 10A–F)
	(zingerone)
_	rior notopleural seta dorsally (Fig. 11A–E) (cue-lure)
	<i>B. (Bactrocera) morula</i> Drew
34	Costal band confluent with or overlapping R_{4+5}
_	Costal band not reaching to R ₄₊₅ 43
35	Scutum and abdominal tergites mostly red-brown
_	Scutum black or dark fuscous with a pair of longitudinal black bands; abdominal
26	tergites mostly black or orange-brown or red-brown with dark markings
36	Anepisternal stripe reaching to anterior notopleural seta; pecten present on male abdominal tergite III (Fig. 24J–L) (methyl eugenol) (bred from <i>Nauclea</i> sp.)
_	anterior notopleural seta; pecten absent from male abdominal tergite III (Fig.
	25A–C) (bred from <i>Spondias dulcis</i>)
37	Abdominal tergites III–V red-brown with a black 'T'-shaped pattern and narrow
	lateral dark margins; pecten absent from male abdominal tergite III (Fig. 25D–F)
	(bred from <i>Calophyllum</i> spp)B. (Calodacus) calophylli (Perkins & May)
—	Abdominal tergites III-V mostly black or orange-brown with broad medial and
	longitudinal black stripes; pecten present on male abdominal tergite III
38	Abdominal tergites III–V orange-brown with broad medial and lateral longitudi-
	nal black stripes that are not joined (Fig. 25G–I) (methyl eugenol) B. (Bactrocera) froggatti (Bezzi)
_	Abdominal tergites mostly black
39	Costal band overlapping R_{4+5} for entire length; abdominal tergites mostly black,
-	with some orange-brown centrally on tergites IV and V
_	Costal band confluent with R, ; abdominal tergites entirely black

 $- \quad \ \ \text{Costal band confluent with } \mathsf{R}_{_{4+5}\!;} \text{ abdominal tergites entirely black}.....41$

Wing (Fig. 9E–G) with a light fuscous tinge as a broad, somewhat triangular area covering much of the middle of the wing, including the areas bordering r-m and
dm-cu (Figs 6A–E, 9E–G) (cue-lure)B. (Bactrocera) geminosimulata sp. nov.
Wing (Fig. 9A–D) without a light fuscous tinge in the area described above (Figs 7A–D, 9A–D) (cue-lure) (bred from <i>Coccinia grandis</i>)
<i>B. (Bactrocera) simulata (Malloch)</i>
Microtrichia covering all of basal costal and costal cells in wing (Fig. 25J–L) (di-
hydroeugenol, isoeugenol) (bred from Allophylus cobbe (formerly Pometia pin-
nata))B. (Bactrocera) quadrisetosa (Bezzi)
Microtrichia restricted to posterodistal corner of costal cell in wing
Legs mostly black; scutellum with a broad black basal band; an episternal stripe $(T_{1}^{2}, 2(A, C))$
narrow, just wider than notopleuron (Fig. 26A–C) (cue-lure)
Legs mostly fulvous; scutellum with a narrow black basal band; anepisternal stripe
reaching to anterior notopleural seta (Fig. 26D–F) (cue-lure)
Scutellum yellow with dark markings or orange-brown and narrowly yellow later-
ally
Scutellum entirely yellow or entirely orange-brown
Scutellum with a dark apical spot (Fig. 26G–I) (methyl eugenol) (bred from
Moraceae in Australia and <i>Pimelodendron amboicinum</i> in Papua New Guinea)
Scutellum with a black or brown longitudinal marking over dorsal surface
Scutum and abdomen predominantly black (Fig. 26J–L) (methyl eugenol)
B. (Bactrocera) picea (Drew)
Scutum and abdomen predominantly red-brown
Postpronotal lobe fuscous or orange-brown anteriorly and yellow posteriorly; ab-
dominal tergites III–V with a broad medial longitudinal black stripe and with or
without narrow sublateral longitudinal black bands over tergites III–V which are all joined across posterior margin of tergite V by a narrow transverse black band 47
Postpronotal lobe entirely yellow; abdominal tergites III–V with either a narrow
medial longitudinal black band on all tergites or a broad medial band on tergite V
only
Abdomen with broad sublateral black stripes on tergites III-V, in addition to
medial stripe (Fig. 13A–E) (cue-lure) (bred from <i>Medusanthera laxiflora</i> in Papua
New Guinea)
Abdomen with sublateral black stripes absent on tergites III–V (Fig. 12A–F) (cue-lure)
Anepisternal stripe reaching anterior notopleural seta; bulla present in male wing (Fig.
27A–D) (bred from <i>Gnetum gnemon</i>) B. (Bulladacus) pacificae Drew & Romig
Anepisternal stripe reaching midway between anterior margin of notopleuron
and anterior notopleural seta; no bulla in male wing (Fig. 27E–G) (cue-lure)

49	Scutum basically red-brown
_	Scutum predominantly black
50	Lateral postsutural vitta very short and tapering to a point posteriorly; a circular
	black spot present on tergite V (Fig. 27H–J)
_	Lateral postsutural vitta long and reaching to intra-alar seta; abdominal tergites
	uniformly pale colored or with patterns of dark markings
51	Costal cells colorless; abdominal tergites III-V with broad lateral longitudinal
	fuscous stripes (Fig. 27K–M) B. (Bactrocera) aithogaster Drew
_	Costal cells with pale fuscous coloration; abdominal tergites III-V uniformly
	orange-brown or with a black 'T'-shaped pattern (Fig. 28A–C) (cue-lure, zinge-
	rone) (bred from <i>Inocarpus fagifer</i>) B. (Bactrocera) moluccensis (Perkins)
52	Postpronotal lobe dark fuscous (Fig. 28D–F) (cue-lure)
	B. (Bactrocera) furvescens Drew
_	Postpronotal lobe yellow
53	Abdominal tergites entirely black (Fig. 28G–I) (cue-lure)
20	<i>B. (Bactrocera) aterrima (Drew)</i>
_	Abdominal tergites orange-brown with or without dark color patterns
54	Abdominal tergites either entirely orange-brown or with very narrow black lines
<i>J</i> 1	anterolaterally on tergite III and occasionally with a narrow medial black stripe
	over tergites III–V (Fig. 26G–I) (methyl eugenol) (bred from Moraceae in Aus-
	tralia and <i>Pimelodendron amboicinum</i> in Papua New Guinea)
_	Abdominal tergites orange-brown with distinct dark markings laterally and medi-
	ally
55	Medial postsutural vitta present (Fig. 18E–H) (bred from <i>Terminalia catappa</i> and
))	Gnetum gnemon)
_	Medial postsutural vitta absent (Fig. 28J–L) (methyl eugenol)

Discussion

The snap-shot survey yielded 16,843 Dacine flies, belonging to 30 known and six new species, described herein, increasing the number of species known from the Solomon Islands from 48 to 54 (Table 1). Twenty-eight species were represented by at least ten specimens and the five most collected species were *Bactrocera frauenfeldi* (Schiner) (43.0% of all specimens), *B. froggatti* (Bezzi) (13.4%), *B. umbrosa* (Fabricius) (9.6%), *B. morula* Drew (7.1%), and *B. pagdeni* (Malloch) (7.0%). Our sampling effort was very fruitful, yielding 29 of the 37 species previously collected by trapping over eight years, plus we found six new species, and 18 new island records (Table 1). We collected 31 of 48, 22 of 31, and 8 of 14 species found on Guadalcanal, Kolombangara, and

Gizo, respectively. The species accumulation curves (Fig. 2) demonstrate the highest species diversity to be in the forests of Guadalcanal, with twice as many species as in agricultural sites (Fig. 2C). Despite the deployment of 36 sets of traps in the rich protected forests of Kolombangara (688 km²), the number of collected and projected species was still half as many as on Guadalcanal (5,302 km²) (Fig. 2B, C), consistent with previously published accounts (Drew and Romig 2001; Hollingsworth et al. 2003). The difference is related to island size, with number of fruit fly species clearly correlated ($r^2 = 60.9\%$) to island size in the Solomon Islands (Suppl. material 3: Fig. S2).

In addition to collecting three new species, the use of zingerone lure revealed that Bactrocera pagdeni, formerly known only by its female holotype (Drew 1974), one specimen at the Bishop Museum collections (BPBM), and a few males recently captured in zingerone traps (Hancock and Drew 2018b), is actually a common and widespread species, with 1,174 specimens collected during our survey (Table 1). Likewise, the recent discovery of the attraction of Bactrocera quadrisetosa (Bezzi) to dihydroeugenol and isoeugenol lures in Vanuatu (Leblanc, unpublished) will likely reveal that this species is also common and widespread in the Solomon Islands. Clearly, many new species are awaiting discovery with the increasing availability of new generation lures (Manrakhan et al. 2017; Royer et al. 2018, 2019). Several rare species that require further attention in future surveys include: B. aithogaster Drew (known by only two specimens), B. bancroftii (Tryon) (one specimen of this Australian species from Guadalcanal), B. furvescens Drew, a Papua New Guinea species of which a single specimen was collected in 1971 in Honiara, and B. unipunctata (Malloch) known from a single specimen collected on Florida Island (Malloch 1939). The species from Mount Austen (Guadalcanal), identified as *B. musae* (Tryon) (Drew and Romig 2001), is likely a nonpest species member of the B. musae complex (Drew et al. 2011). Bactrocera musae is a major pest of banana, and no fruit fly infestations have been observed on banana in the Solomon Islands, even in recent years (FT, pers. obs.).

The COI sequences we obtained for the new species typically have large minimum pairwise distances to their nearest congeners, up to 12%, whereas the average minimum distance between species for *Bactrocera* is 6.09% (Suppl. material 2: Table S1; Doorenweerd et al. 2020). This is likely due to a lack of species from Papua New Guinea represented in the reference dataset, where there is a large, mostly unstudied, diversity of *Bactrocera* (White and Evenhuis 1999; Drew pers. comm.). As a consequence, the currently available reference data suggests that COI reliably distinguishes all newly described species, but further sampling of species in New Guinea may reduce the pairwise distance resolution (Doorenweerd et al. 2020). There is one potentially new species (*B.* spnSol08; molecular voucher UHIM.ms08767), for which we have one specimen, that we leave undescribed. Although its COI sequence is highly divergent, closest to *B. hantanae* Tsuruta & White at 9.89%, there is only a single specimen and its morphology has no apparent differences with that of *B. dorsalis* (Hendel). Future sampling will hopefully bring in a larger series of this potentially new species to enable further examination of the morphological characters.

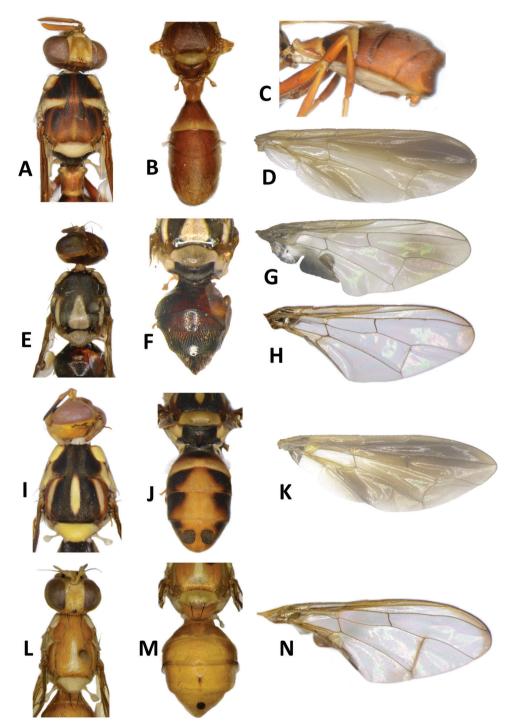


Figure 18. Dacus solomonensis Malloch A head and scutum B, C abdomen D wing. Bactrocera penefurva
Drew E head and scutum F abdomen G male wing H female wing. Bactrocera pagdeni (Malloch) I head
and scutum J abdomen K wing. Zeugodacus univittatus (Drew) L head and scutum M abdomen N wing.

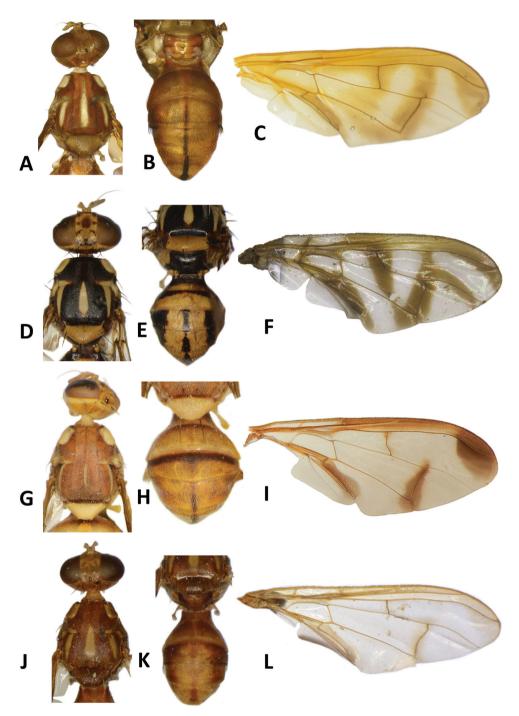


Figure 19. Zeugodacus hamaceki Drew & Romig A head and scutum B abdomen C wing. Zeugodacus amoenus (Drew) D head and scutum E abdomen F wing. Zeugodacus cucurbitae (Coquillett) G head and scutum H abdomen I wing. Zeugodacus fuscipennulus (Drew & Romig) G head and scutum H abdomen I wing.

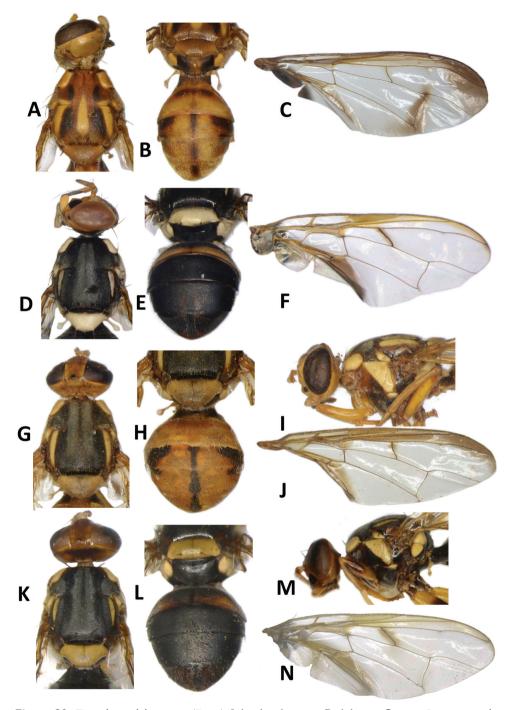


Figure 20. Zeugodacus abdoangustus (Drew) A head and scutum B abdomen C wing. Bactrocera melanogaster Drew D head and scutum E abdomen F wing. Bactrocera phaea (Drew) G head and scutum H abdomen I lateral view J wing. Bactrocera neonigrita Drew K head and scutum L abdomen M lateral view N wing.

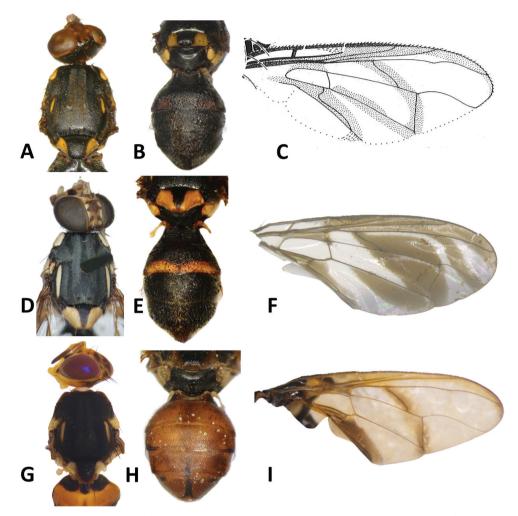


Figure 21. Bactrocera reclinata Drew A head and scutum B abdomen C wing (reproduced from Drew 1989). Bactrocera longicornis Macquart D head and scutum E abdomen F wing. Bactrocera hollingsworthi Drew & Romig G head and scutum H abdomen I wing.

In addition to the data from this survey, we summarized trapping data in the Solomon Islands generated during the Regional Fruit Fly Projects in the Pacific, as a further indication of the relative abundance and to update the distribution of each species (Table 1). Over 1.8 million flies were collected from 180 sites maintained throughout the archipelago between 1994 and 2001 (Vagalo et al. 1997; Drew and Romig 2001; Hollingsworth et al. 2003; Leblanc et al. 2012). A few specimens of then undescribed *B. geminosimulata* and *B. quasienochra* may have been included among these records.

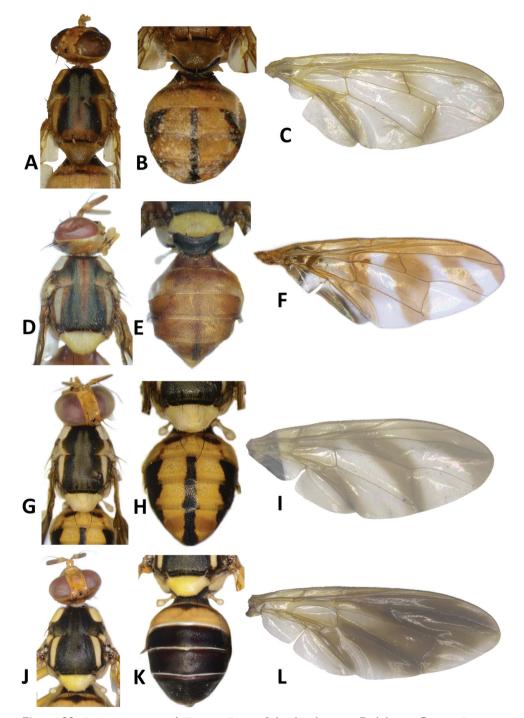


Figure 22. *Bactrocera unitaeniola* Drew & Romig **A** head and scutum **B** abdomen **C** wing. *Bactrocera umbrosa* (Fabricius) **D** head and scutum **E** abdomen **F** wing. *Bactrocera unifasciata* (Malloch) **G** head and scutum **H** abdomen **I** wing. *Bactrocera pepisalae* (Froggatt) **J** head and scutum **K** abdomen **L** wing.

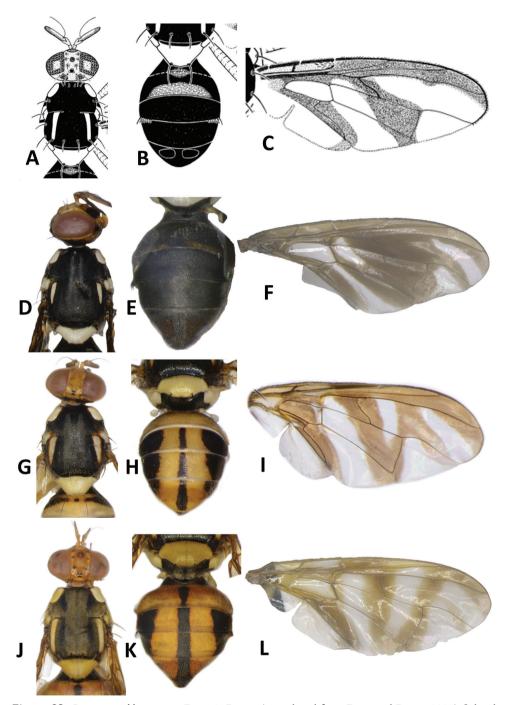


Figure 23. Bactrocera obliquivenosa Drew & Romig (reproduced from Drew and Romig 2001) A head and scutum B abdomen C wing. Bactrocera biarcuata (Walker) D head and scutum E abdomen F wing. Bactrocera nigrescentis (Drew) G head and scutum H abdomen I wing. Bactrocera redunca (Drew) J head and scutum K abdomen L wing.

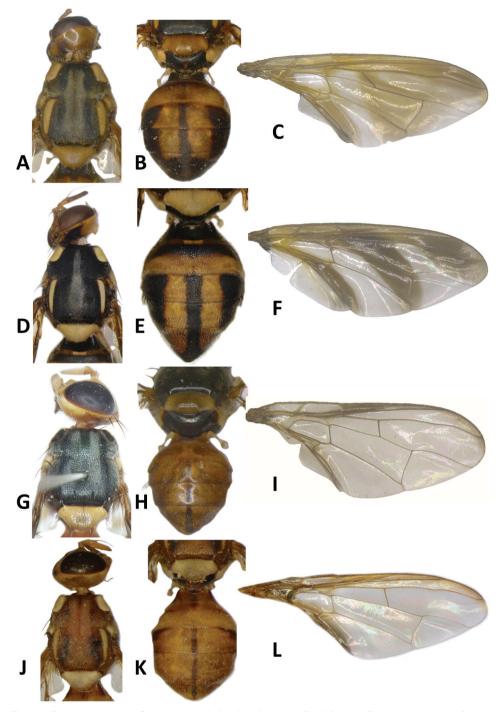


Figure 24. Bactrocera confluens (Drew) A head and scutum B abdomen C wing. Bactrocera decumana (Drew) D head and scutum E abdomen F wing. Bactrocera minuta (Drew) G head and scutum H abdomen I wing. Bactrocera naucleae (Drew & Romig) J head and scutum K abdomen L wing.

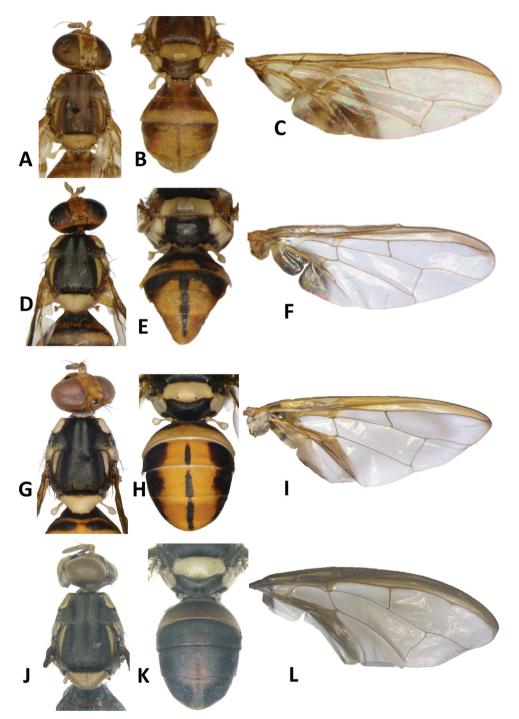


Figure 25. Bactrocera hastigerina (Hardy) A head and scutum B abdomen C wing. Bactrocera calophylli (Perkins & May) D head and scutum E abdomen F wing. Bactrocera froggatti (Bezzi) G head and scutum H abdomen I wing. Bactrocera quadrisetosa (Bezzi) J head and scutum K abdomen L wing.

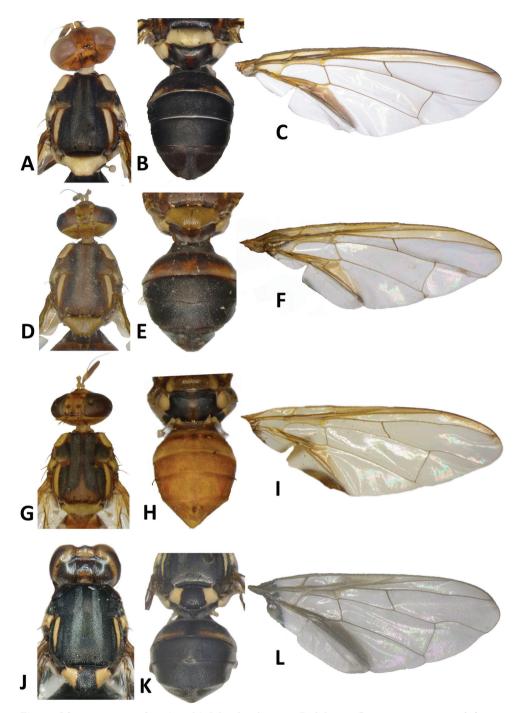


Figure 26. Bactrocera epicharis (Hardy) A head and scutum B abdomen C wing. Bactrocera atrabifasciata
Drew & Romig D head and scutum E abdomen F wing. Bactrocera bancroftii (Tryon) (specimen from Australia) G head and scutum H abdomen I wing. Bactrocera picea (Drew) J head and scutum K abdomen L wing.

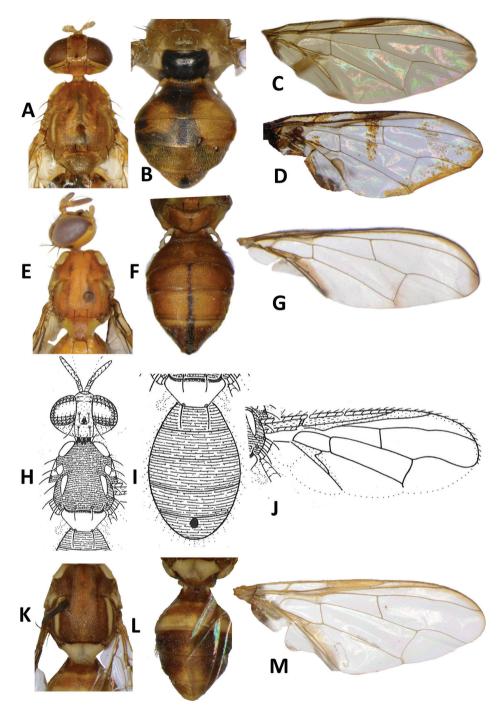


Figure 27. Bactrocera pacificae Drew & Romig A head and scutum B abdomen C female wing D male wing. Bactrocera buinensis Drew E head and scutum F abdomen G wing. Bactrocera unipunctata (Malloch) (reproduced from Drew 1989) H head and scutum I abdomen J wing. Bactrocera aithogaster Drew K scutum L abdomen M wing.

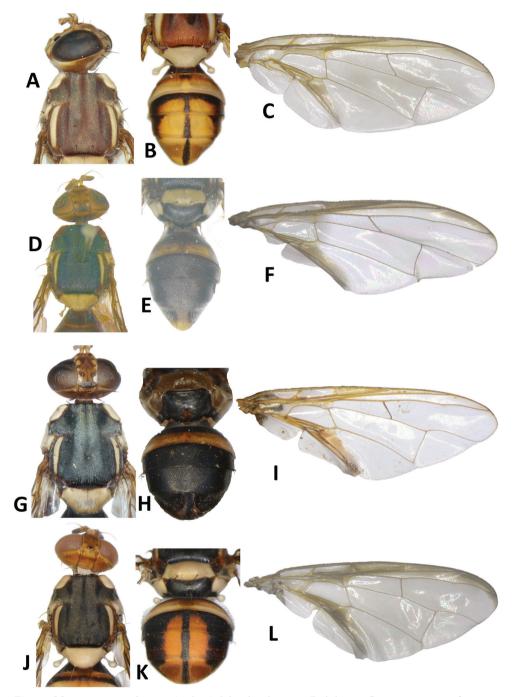


Figure 28. Bactrocera moluccensis (Perkins) A head and scutum B abdomen C wing. Bactrocera furvescens Drew D head and scutum E abdomen F wing. Bactrocera aterrima (Drew) G head and scutum H abdomen I wing. Bactrocera parafroggatti Drew & Romig J head and scutum K abdomen L wing.

Table 1. Checklist of Dacine fruit flies of Solomon Islands, including number of specimens collected during the Regional Fruit Fly Projects (1994–2001) and the 2018 survey. References to earliest record for each group on islands are: A: Macquart 1835, B: Froggatt 1910, C: Bezzi 1919, D: Malloch 1939, E: Hardy 1954, F: Drew 1972, G: Drew 1974, H: Eta 1985, I: Drew 1989, J: Waterhouse 1993; K: Hollingsworth et. al. 1997, L: Drew and Romig 2001, new: previously unpublished or new record, W: widespread species.

Species	Lure	# trapped Solomon Islands (1994–2001)	# trapped Gizo (2018)	# trapped Kolombangara (2018)	# trapped Guadalcanal (2018)	Shortland Group	Choiseul	Vella Lavella	Gizo	Kolombangara	New Georgia	Isabel	Russell	Florida (Ngella & Savo)	Guadalcanal	Malaita	San Cristobal	Rennell & Bellona	Santa Cruz	Reef Islands
BACTROCERA B. aithogaster	No known lure				1										I					
Drew, 1989	INO KHOWH JUIC																			
B. <i>allodistincta</i> Leblanc & Doorenweerd	Cue-lure				12										new					
<i>B. aterrima</i> (Drew, 1972)	Cue-lure	10			1	Ι	L			L		L			new					
<i>B. atrabifasciata</i> Drew & Romig, 2001	Cue-lure	14	1		1	L			L			L		L	new					
B. bancroftii (Tryon, 1927)	Methyl eugenol	118													L					
<i>B. biarcuata</i> (Walker, 1865)	Methyl eugenol	299		28	3	Ι	L		L	L	L	L	L	L	L		L			
<i>B. buinensis</i> Drew, 1989	Cue-lure	16		163			L			L					new		new			
<i>B. calophylli</i> (Perkins & May, 1949)	No known lure	bred from fruit													L					
B. confluens (Drew, 1971)	Methyl eugenol	412			1		new			new				L	Ι	new				
<i>B. decumana</i> (Drew, 1972)	Cue-lure	1,226		208		Ι	L	L	L	L		L	L	L	L	L	L			
<i>B. enochra</i> (Drew, 1972)	Cue-lure	33	1	19	1	F			new	L		L			L					
<i>B. epicharis</i> (Hardy, 1970)	Cue-lure	168		119	48	Ι	new			L		new		L	L	L				
<i>B. frauenfeldi</i> (Schiner, 1868)	Cue-lure, zingerone	1,271,832	1686	921	4636	W	W	W	W	W	W	W	D	W	W	W	W	W	W	W
<i>B. froggatti</i> (Bezzi, 1928)	Methyl eugenol	33,514	36	983	1236	Ι	L	L	G	L	L	L	С	D	D	L	L	L		
<i>B. furvescens</i> Drew, 1989	Cue-lure														Ι					
<i>B. geminosimulata</i> Leblanc & Doorenweerd	Cue-lure				14										new					
<i>B. hastigerina</i> (Hardy, 1954)	No known lure	bred from fruit													L					
<i>B. hollingsworthi</i> Drew & Romig, 2001	Cue-lure	5		1						new									L	
B. <i>kolombangarae</i> Leblanc & Doorenweerd	Zingerone			18	1					new					new					
<i>B. longicornis</i> Macquart, 1835	Cue-lure											А								

Species	Lure		8)	8	ıal									(o)						
		# trapped Solomon Islands (1994–2001)	# trapped Gizo (2018)	# trapped Kolombangara (2018)	# trapped Guadalcanal (2018)	Shortland Group	Choiseul	Vella Lavella	Gizo	Kolombangara	New Georgia	Isabel	Russell	Florida (Ngella & Savo)	Guadalcanal	Malaita	San Cristobal	Rennell & Bellona	Santa Cruz	Reef Islands
<i>B. melanogaster</i> Drew, 1989	Methyl eugenol	820		2	10	new	Ι	new	new	Ι	Ι	Ι	Ι	Ι	Ι					
<i>B. minuta</i> (Drew, 1971)	Cue-lure	45																	L	
B. moluccensis (Perkins, 1939)	Cue-lure	71,499	9	5	435	L	L	L	L	L	L	L	L	L	L	L	L	L	L	
<i>B. morula</i> Drew, 1989	Cue-lure	861			1202				L						Ι		L			
<i>B. naucleae</i> Drew & Romig, 2001	Methyl eugenol	130					L					L			L					
<i>B. neonigrita</i> Drew, 1989	Methyl eugenol					Ι														
B. nigrescentis (Drew, 1971)	Cue-lure	279	1	32	331		L	L	new	new	new	L		L	L			L	L	
<i>B. obliquivenosa</i> Drew & Romig, 2001	Methyl eugenol	1								L										
<i>B. pacificae</i> Drew & Romig, 2001	No known lure	bred from fruit													L				L	
B. pagdeni (Malloch, 1939)	Zingerone			718	456					new				D	new					
<i>B. parafroggatti</i> Drew & Romig, 2001	Methyl eugenol	1,645			85		L			new	L	L		L	L		L	L		
<i>B. penefurva</i> Drew, 1989	No known lure	bred from fruit													Ι					
<i>B. pepisalae</i> (Froggatt, 1910)	Methyl eugenol	7,746			35	new	L	E		L	L	L	В	D	G	L	G			
<i>B. phaea</i> (Drew, 1971)	Cue-lure	97										L		L	L					
<i>B. picea</i> (Drew, 1972)	Methyl eugenol	726		227	19	Ι	L	L	L	L	L	L	Ι	L	L	new				
B. pseudodistincta (Drew, 1971)	Cue-lure	433		5			L			L		L		L	L	L	L	L	L	
B. quadrisetosa (Bezzi, 1928)	Dihydroeugenol, isoeugenol	bred from fruit									L			D	Ι				L	
B. <i>quasienochra</i> Leblanc &	Cue-lure				1										new					
Doorenweerd <i>B. reclinata</i> Drew,	Methyl eugenol	1													L					
1989 <i>B. redunca</i> (Drew,	Cue-lure	7,031			524	I	L			L		L	L	L	L	L	new		L	
1971) <i>B. simulata</i> (Malloch, 1939)	Cue-lure	32,810		16	350	I	L	L	L	L	I	L	L	D	I	L	L	L	L	
B. <i>tsatsiae</i> Leblanc & Doorenweerd	Zingerone			20	9					new					new					
<i>B. umbrosa</i> (Fabricius, 1805)	Methyl eugenol	362,783	157	170	1296	new	W	W	W	W	W	W	W	D	W	W	W	W	D	W
B. unifasciata (Malloch, 1939)	Cue-lure	5		1	18					new		L			D					
<i>B. unipunctata</i> (Malloch, 1939)	No known lure													D						
<i>B. unitaeniola</i> Drew & Romig, 2001	Cue-lure	13					L					L			L					

Species	Lure	# trapped Solomon Islands (1994–2001)	# trapped Gizo (2018)	# trapped Kolombangara (2018)		Shortland Group	Choiseul	Vella Lavella	Gizo	Kolombangara	New Georgia	Isabel	Russell	Florida (Ngella & Savo)	Guadalcanal	Malaita	San Cristobal	Rennell & Bellona	Santa Cruz	Reef Islands
B. <i>vargasi</i> Leblanc & Doorenweerd	Zingerone			9	34					new					new					
DACUS																				
D. solomonensis Malloch, 1939	Cue-lure	23,085			60		L			new	L	L	L	L	D	L	L			
ZEUGODACUS																				
Z. abdoangustus (Drew, 1972)	Cue-lure	38			11							L			L	L				
Z. amoenus (Drew, 1972)	Cue-lure	3										L								
<i>Z. cucurbitae</i> (Coquillett, 1899)	Cue-lure, zingerone	43,294	7	305	44	Н	J	J	J	J	J	J	K		К	K				
Z. fuscipennulus (Drew & Romig, 2001)	Cue-lure	101		1						L	new	new			L		new			
Z. hamaceki (Drew & Romig, 2001)	Cue-lure	115			5					L		L			L					
Z. univittatus (Drew, 1972)	Cue-lure	118			95					new		L			L		L			

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Supplementary material I

Figure S1. COI Phylogeny

Authors: Camiel Doorenweerd

Data type: phylogenetic

- Explanation note: Maximum likelihood tree based on COI sequence data, modified from Doorenweerd et al. (2020) to have species / species complexes 'collapsed' into triangles where the horizontal width indicates the maximum pairwise distance of that clade. Newly described species are indicated in green.
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Link: https://doi.org/10.3897/zookeys.1057.68375.suppl1

Supplementary material 2

Table S1. Pairwise molecular distance

Authors: Camiel Doorenweerd

Data type: phylogenetic

- Explanation note: COI Pairwise distance statistics for the *Bactrocera* included in Doorenweerd et al. (2020), including the species newly described here.
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Link: https://doi.org/10.3897/zookeys.1057.68375.suppl2

Supplementary material 3

Figure S2. Number of dacine fruit fly species in relation to island size in the Solomon Islands

Authors: Luc Leblanc, Francis Tsatsia, Camiel Doorenweerd

Data type: pdf. file

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