

## 

## Complete Genome Sequences of the Probiotic Lactic Acid Bacteria Lactobacillus helveticus D75 and D76

V. A. Toropov,<sup>a</sup> T. Y. Vakhitov,<sup>a</sup> O. N. Shalaeva,<sup>a</sup> E. K. Roshchina,<sup>a</sup> S. I. Sitkin<sup>a</sup>

<sup>a</sup>State Research Institute of Highly Pure Biopreparations of the FMBA of the Russian Federation, St. Petersburg, Russia

**ABSTRACT** Lactobacillus helveticus D75 and D76 were isolated from the intestinal tract of a healthy child. Both strains possess symbiotic, probiotic, and antagonistic activities. We have sequenced and annotated the whole genomes of *L. helveticus* D75 and D76 and have conducted a preliminary genome comparative analysis.

Lactobacillus helveticus D75 and D76 have pronounced fermentative and probiotic activities (1). It was important to have information that was as complete as possible about the genomes of *L. helveticus* strains D75 and D76 to understand the mechanisms of regulation of their probiotic activity and syntrophic interactions.

*L. helveticus* D75 and D76 were grown on de Man-Rogosa-Sharpe (MRS) medium (2) at 37°C up to the mid-exponential-growth phase. The modified method of cell lysis of Gram-positive bacteria and the conventional method of DNA extraction with organic solvents were used to obtain the chromosomal DNA (3). The cell lysis consisted of two phases. Initially, a Tris-EDTA buffer containing mutanolysin (final concentration, 300 U/ ml) and lysozyme (final concentration, 2 mg/ml) was added to the cell pellet, and the resulting mixture was incubated at 37°C for 1 h. Then, a solution containing sodium dodecyl sulfate (final concentration, 1.5%) and proteinase K (final concentration, 1 mg/ ml) was added, and the mixture was incubated at 50°C for 1 h.

Both genomic DNAs were sequenced using the PacBio RS II platform (Macrogen, Inc., Republic of Korea) (4–7). Genome libraries consisting of 150,292 and 166,471 reads with  $N_{50}$  values of 22,778 bp and 10,700 bp were obtained for *L. helveticus* D75 and D76, respectively. The HGAP algorithm in the SMRT Analysis pipeline version 2.3.0 was used to assemble the genomes of *L. helveticus* D75 and D76 from PacBio RS raw reads (8). The lengths of the whole genomes obtained were 2,053,066 bp (with 422× read multiplicity) and 2,058,319 bp (with 375× read multiplicity) for the *L. helveticus* D75 and D76 strains, respectively.

The genome annotations of *L. helveticus* D75 and D76 were done using the Prokaryotic Genome Annotation Pipeline (PGAP) algorithm of the National Center for Biotechnological Information (NCBI) (9). The annotated genome of *L. helveticus* D75 contained 2,092 coding sequences (CDS), with 1,693 protein-coding genes, 64 tRNA genes, and 15 rRNA genes. The total number of pseudogenes was 317. *L. helveticus* D76 includes 2,068 CDS, with 1,986 protein-coding genes, 64 tRNA genes, and 15 rRNA genes. The total number of pseudogenes was 265. The BLASTn algorithm was applied for preliminary genome comparative analysis of *L. helveticus* D75 and D76 with the genomes of *L. helveticus* DPC4571 and *L. helveticus* R0052, which were annotated and deposited in GenBank (10, 11). The *L. helveticus* D75 and D76 strains had 99.18% identity (90.65% coverage) and 97.73% identity (80.45% coverage) to the genomes of the DPC4571 and R0052 strains, respectively.

Calculating the average nucleotide identity (ANI) (12–14) showed that the genome sequences of *L. helveticus* D75 and D76 were 99.22% identical (with 76.3% coverage of the genome) to the genomes of *L. helveticus* species. Therefore, D75 and D76 strains,

Received 9 January 2018 Accepted 11 January 2018 Published 15 March 2018

Citation Toropov VA, Vakhitov TY, Shalaeva ON, Roshchina EK, Sitkin SI. 2018. Complete genome sequences of the probiotic lactic acid bacteria *Lactobacillus helveticus* D75 and D76. Genome Announc 6:e01552-17. https://doi .org/10.1128/genomeA.01552-17.

**Copyright** © 2018 Toropov et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to T. Y. Vakhitov, vakhitov@hpb-spb.com.

previously attributed to *L. acidophilus* species (based both on phenotype and 16S rRNA genetic analysis), were reclassified as *L. helveticus* D75 and D76.

Accession number(s). These whole-genome shotgun projects have been deposited in DDBJ/ENA/GenBank under the accession no. CP020029 (*L. helveticus* D75) and CP016827 (*L. helveticus* D76). The versions described in this paper are the first versions.

## ACKNOWLEDGMENT

The study was financially supported exclusively by the State Research Institute of Highly Pure Biopreparations of the FMBA of the Russian Federation.

## REFERENCES

- Vakhitov TY, Verbitskaya NB, Dobrolezh OV, Polevaya EV, Kobatov AI. 2013. The effect of probiotic and pathogenic bacteria metabolites on antagonistic activity of *Lactobacillus acidophilus* D No. 75. Sci J KubSAU 92:339–357.
- De Man JC, Rogosa M, Sharpe ME. 1960. A medium for the cultivation of lactobacilli. J Appl Bacteriol 23:130–135. https://doi.org/10.1111/j.1365 -2672.1960.tb00188.x.
- Bollet C, Gevaudan MJ, de Lamballerie X, Zandotti C, de Micco P. 1991. A simple method for the isolation of chromosomal DNA from Gram positive or acid-fast bacteria. Nucleic Acids Res 19:1995. https://doi.org/ 10.1093/nar/19.8.1955.
- Khan IU, Yadav JS. 2004. Development of a single-tube, cell lysis-based, genus-specific PCR method for rapid identification of mycobacteria: optimization of cell lysis, PCR primers and conditions, and restriction pattern analysis. J Clin Microbiol 42:453–457. https://doi.org/10.1128/ JCM.42.1.453-457.2004.
- Van Soolingen D, Hermans PW, de Haas PE, Soll DR, van Embden JD. 1991. Occurrence and stability of insertion sequences in *Mycobacterium tuberculosis* complex strains: evaluation of an insertion sequencedependent DNA polymorphism as a tool in the epidemiology of tuberculosis. J Clin Microbiol 29:2578–2586.
- Rhoads A, Au KF. 2015. PacBio sequencing and its applications. Genomics Proteomics Bioinformatics 13:278–289. https://doi.org/10.1016/j.gpb .2015.08.002.
- Zhu L, Zhong J, Jia X, Liu G, Kang Y, Dong M, Zhang X, Li Q, Yue L, Li C, Fu J, Xiao J, Yan J, Zhang B, Lei M, Chen S, Lv L, Zhu B, Huang H, Chen F. 2016. Precision methylome characterization of *Mycobacterium tuberculosis* complex (MTBC) using PacBio single-molecule real-time (SMRT)

technology. Nucleic Acids Res 44:730-743. https://doi.org/10.1093/nar/gkv1498.

- Chin CS, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. https://doi.org/10.1038/ nmeth.2474.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI prokaryotic genome annotation pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.
- Callanan M, Kaleta P, O'Callaghan J, O'Sullivan O, Jordan K, McAuliffe O, Sangrador-Vegas A, Slattery L, Fitzgerald GF, Beresford T, Ross RP. 2008. Genome sequence of *Lactobacillus helveticus*, an organism distinguished by selective gene loss and insertion sequence element expansion. J Bacteriol 190:727–735. https://doi.org/10.1128/JB.01295-07.
- Tompkins TA, Barreau G, Broadbent JR. 2012. Complete genome sequence of *Lactobacillus helveticus* R0052, a commercial probiotic strain. J Bacteriol 194:6349. https://doi.org/10.1128/JB.01638-12.
- Arahal DR. 2014. Whole-genome analyses: average nucleotide identity. Methods Microbiol 41:103–122.
- Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, Tiedje JM. 2007. DNA-DNA hybridization values and their relationship to whole-genome sequence similarities. Int J Syst Evol Microbiol 57:81–91. https://doi.org/10.1099/ijs.0.64483-0.
- Konstantinidis KT, Tiedje JM. 2005. Genomic insights that advance the species definition for prokaryotes. Proc Natl Acad Sci U S A 102:2567–2572. https://doi.org/10.1073/pnas.0409727102.