



# Extracting Extra-Telomeric Phenotypes from Telomerase Mouse Models

Young Hoon Sung,\* Muhammad Ali,\* and Han-Woong Lee

Department of Biochemistry, College of Life Science and Biotechnology, Laboratory Animal Research Center, Yonsei University, Seoul, Korea.

Received: October 4, 2013

Co-corresponding authors:

Dr. Young Hoon Sung and Dr. Han-Woong Lee,  
Department of Biochemistry,  
College of Life Science, and Biotechnology,  
Laboratory Animal Research Center,  
Yonsei University, 50 Yonsei-ro,  
Seodaemun-gu, Seoul 120-752, Korea.  
Tel: 82-2-2123-7642, Fax: 82-2-2123-8682  
E-mail: [sungyh@yonsei.ac.kr](mailto:sungyh@yonsei.ac.kr); [hwl@yonsei.ac.kr](mailto:hwl@yonsei.ac.kr)

\*Young Hoon Sung and Muhammad Ali  
contributed equally to this work.

The authors have no financial conflicts of  
interest.

Telomerase reverse transcriptase (TERT) is the protein component of telomerase and combined with an RNA molecule, telomerase RNA component, forms the telomerase enzyme responsible for telomere elongation. Telomerase is essential for maintaining telomere length from replicative attrition and thus contributes to the preservation of genome integrity. Although diverse mouse models have been developed and studied to prove the physiological roles of telomerase as a telomere-elongating enzyme, recent studies have revealed non-canonical TERT activities beyond telomeres. To gain insights into the physiological impact of extra-telomeric roles, this review revisits the strategies and phenotypes of telomerase mouse models in terms of the extra-telomeric functions of telomerase.

**Key Words:** Telomerase reverse transcriptase, extra-telomeric function of TERT, transgenic, knockout mouse, genetically engineered mouse, stem cells, senescence, anti-apoptosis, metabolic fitness, cancer

## INTRODUCTION

During DNA replication, the linear ends of chromosomes are eroded at each cell division due to the end replication problem.<sup>1</sup> Telomeres, the very ends of linear chromosomes, are predominantly composed of tandem repeats of short sequences; in vertebrates, the repeats consist of the TTAGGG hexanucleotide.<sup>2</sup> Telomere lengths are also remarkably heterogeneous among individuals and vary according to the origin, age, and proliferative history of cells.<sup>3,4</sup> Telomere length variations among individuals of the same age are, therefore, thought to be related to variations in ageing and longevity.<sup>5</sup> As a ribonucleoprotein complex that is composed of telomerase reverse transcriptase (TERT) and telomerase RNA component (TERC),<sup>6</sup> telomerase is responsible for elongation of the telomeres, and thus maintains genome stability.<sup>7,8</sup> The enzymatic activity of telomerase is not detected in normal somatic cells, but is detected in embryonic and highly proliferative adult tissues.<sup>9</sup> Furthermore, telomerase is re-activated in most cancers,<sup>10</sup> thus suggesting the possibility that telomerase is a potential therapeutic target in cancers.

Recently, a number of reports have shown extra-telomeric functions of TERT. The following observations pose questions regarding the rationality of telomere length-dependent singular role of telomerase:

### © Copyright:

Yonsei University College of Medicine 2014

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

1) Although tissue stem and progenitor cells have sufficient telomere reserves, they highly express TERT. Notably, laboratory mice have significantly longer telomeres than humans (40-60 kb vs. 5-15 kb),<sup>11-14</sup> however, no apparent role for long telomeres has been found in the survival of mice.

2) TERT overexpression promotes tumor development without further telomere elongation.<sup>15</sup>

3) The reconstitution ability of hematopoietic stem cells (HSCs) is essentially linked with TERT, although there is no evidence that HSC activities are fully dependent on the telomere-elongation function of TERT.<sup>16,17</sup>

4) Transgenic mice overexpressing murine TERT show significant resistance to ischemic brain injury and N-methyl-D-aspartic acid (NMDA) receptor-mediated excitotoxicity without any detectable change in telomere length.<sup>18</sup> Interestingly, ischemic injury induces TERT expression in the wild-type brain.<sup>18</sup>

5) Suppression of TERT expression decreases cell growth rate and induces apoptosis prior to measurable telomere shortening,<sup>19</sup> and the expression of specific TERT mutants lacking telomerase activity prevents apoptotic cell death.<sup>20</sup>

6) Ectopic expression of TERT in the hair follicle stem cells of mouse epidermis activates stem cell capacities.<sup>21</sup> The phenotype appears to be independent of telomerase activity.

7) Cancers without functional telomerase (10-15% of all cases) maintain their telomere lengths by adopting an alternative lengthening of telomeres (ALT) pathway.<sup>22</sup>

Furthermore, indirect evidence also indicates extra-telomeric functions of telomerase: TERT may have additional functions because the reverse transcriptase (RT) domain of TERT is only 15 kDa, which is less than 10% of the total molecular weight. Thus, it is quite possible that other regions may mediate distinct activities other than telomerase activity.<sup>23</sup> In support of this hypothesis, alternatively spliced forms of TERT devoid of the RT domain have been identified in humans.<sup>24</sup> Although TERT is known as a nuclear protein, it is exported from the nucleus and delays replicative senescence in endothelial cells dependent on reactive oxygen species (ROS).<sup>25</sup>

Several lines of evidence obtained using mouse models have strengthened the idea that TERT contributes to preventing ageing and cell death, as well as promoting carcinogenesis, cell signaling, and transcriptional regulation, which are unrelated to its role in telomere lengthening. Here, we review the studies encompassing the extra-telomeric functions of telomerase conducted using transgenic or knockout mouse models.

## SURVIVAL-PROMOTING FUNCTIONS OF TELOMERASE

Apoptosis induced by critically short telomeres has been extensively documented. Dysfunctional telomeres increase apoptosis in highly proliferative tissues including intestine,<sup>26,27</sup> male germ cells,<sup>26,28</sup> and splenocytes (B cells) from immunized fifth and sixth generation mice after mitogen treatment.<sup>29</sup> These dysfunctional telomeres can be generated in early generations, and can increase apoptosis in these tissues as well. Protection of Telomeres 1 (POT1) is a single-stranded telomere binding protein that is essential for proper maintenance of telomere length. When *Pot1* is deficient, abnormal apoptosis is induced in proliferative tissues as well as cells derived from *Terc*<sup>-/-</sup> mice, including male germ cells, hematopoietic cells, and intestinal cells.<sup>30</sup> Critically short telomeres also affect highly proliferative developmental processes. *Terc*<sup>-/-</sup> embryos at embryonic day 10.5 (E10.5) with dysfunctional telomeres frequently exhibit neural tube closure defects, suggesting that this is one of the most sensitive developmental processes to telomere loss and chromosomal instability.<sup>31</sup> Although cardiac tissues are not highly proliferative, the balance between cell growth and cell death is critical for maintaining normal heart function. Consistently, dysfunctional telomeres lead to abnormal apoptosis in cardiomyocytes, resulting in cardiac dilatation and heart failure in the late generation of *Terc*<sup>-/-</sup> mice.<sup>32</sup> Additionally, cardiomyocyte survival is promoted in transgenic mice overexpressing wild-type TERT, but not expressing mutant TERT.<sup>33</sup> *Tert* deficiency also induces apoptotic phenotypes; *Tert*<sup>-/-</sup> and *Terc*<sup>-/-</sup> mice show frequent apoptosis in intestinal crypt cells<sup>34-36</sup> and male germ cells<sup>37</sup> respectively in their late generations. Furthermore, the deleterious effects can be rescued by turning on telomerase activity in the late generation of homozygous *ER-Tert* knock-in mice by treating with tamoxifen.<sup>37</sup> These results clearly demonstrate that telomerase deficiency elicits telomere erosion, resulting in abnormal apoptotic phenotypes *in vivo*.

Telomerase-deficient mouse models have provided opportunities for unraveling the mechanisms which induce these apoptotic phenotypes in late generations. Rajaraman, et al.<sup>36</sup> showed using *Tert*<sup>-/-</sup> mice that apoptosis is dependent on S phase, and thus is primarily triggered by newly uncapped (or critically short) telomeres. It is not triggered by chromosome fusion-bridge breakage because mitotic blockade did not alter the apoptotic pattern.<sup>36</sup> It is also de-

pendent on p53 that is activated by genotoxic stresses, including critically short telomeres.<sup>36,38,39</sup> In fact, growth arrest and/or apoptosis in late generations of *Terc*<sup>-/-</sup> mice are dependent on proper p53 activation.<sup>32,40</sup> However, p53-mediated regulation of the phenotypic manifestations in telomerase knockout mice is complicated by its negative effect on *TERT* gene expression.<sup>41</sup> Rahman, et al.<sup>20</sup> evaluated the effect of human TERT (hTERT) overexpression on p53-dependent apoptosis. In HCT116 colon carcinoma cells carrying endogenous p53, genotoxic stress-induced apoptosis that is p53-dependent is suppressed by constitutive hTERT expression. Indeed, a telomerase-inactive hTERT mutant equally antagonizes p53-induced apoptosis.<sup>20</sup> Similarly, ectopic mouse TERT expression in mouse embryonic stem cells that exhibit high levels of telomerase activity and maintain sufficiently long telomeres confers resistance to p53-dependent apoptosis.<sup>42</sup> In addition to telomere-associated functions, these results indicate that TERT exerts antiapoptotic activity beyond telomeres.

In addition to the phenotypes induced by critically short telomeres, emerging evidence has indicated the existence of extra-telomeric functions of telomerase. The first clue for an extra-telomeric role of TERT was obtained by revealing the neuroprotective effect of TERT on neuronal cell death induced by the neurotoxic protein amyloid  $\beta$ -peptide, a protein believed to promote neuronal degeneration in Alzheimer's disease.<sup>43</sup> Using transgenic mice ubiquitously overexpressing TERT, we also have provided evidence that TERT prevents NMDA neurotoxicity through the transfer of cytosolic free Ca<sup>2+</sup> into the mitochondria, thereby playing a protective role in ameliorating ischemic neuronal cell death.<sup>18</sup> Because TERT is induced in postmitotic neurons by ischemic brain injury and its overexpression confers resistance against NMDA neurotoxicity, these protective phenotypes are considered to be independent of telomerase activity. Similarly, first generation (G1) *Tert*-deficient mouse embryonic fibroblasts (MEFs) displayed increased sensitivity to staurosporine (STS), whereas *Tert* transgenic MEFs were more resistant to STS-induced apoptosis than wild-type.<sup>44</sup> Consistent phenotypes were also observed upon NMDA treatment of *Tert*-deficient and *Tert* transgenic mice, respectively.<sup>44</sup> Although extensive studies were conducted, it remains unclear whether the protective function is dependent on telomerase activity.<sup>44</sup> In fact, *Terc* deficiency does not alter the sensitivity of *Tert* transgenic MEFs to STS treatment, and NMDA-induced excitotoxic cell death of primary neurons was suppressed by TERT, but not by *Terc* deficiency, *in*

*vitro* and *in vivo*.<sup>44</sup> Furthermore, although telomerase activity is evidently suppressed in transgenic mice overexpressing hTERT, hTERT transgenic MEFs still show resistance to STS-induced apoptosis.<sup>44</sup> Based on these lines of evidence, telomerase activity must not be essential for the protective function of TERT. Therefore, independent of its roles in telomere maintenance, diverse telomerase mouse models have demonstrated that TERT-mediated antiapoptotic functions may contribute to tumorigenesis.

## ONCOGENIC ROLES OF TELOMERASE IN TUMORIGENESIS

Telomerase knockout or transgenic mouse models have been extensively employed to elucidate the *in vivo* roles of telomerase and dysfunctional telomeres in tumorigenesis.

Telomeres are dedicated to the maintenance of linear chromosomes and thus prevent chromosomal abnormalities. In cultivated cells from late generations of *Terc*<sup>-/-</sup> mice, critically short or dysfunctional telomeres induce aneuploidy and chromosomal abnormalities, including end-to-end fusions.<sup>45</sup> These phenomena are prevalent in cancer, and spontaneous tumors are more frequently induced in late generation *Terc*<sup>-/-</sup> mice,<sup>46</sup> indicating that dysfunctional telomeres are genotoxic and possess mutagenic effects in mice. Additionally, it is plausible that *p53* deficiency significantly attenuates genotoxic stresses triggered by telomere dysfunction. In fact, *p53* deficiency contributes to the neoplastic transformation of cells with critically short telomeres from late generation *Terc*<sup>-/-</sup> mice<sup>40</sup> and promotes non-reciprocal translocations and epithelial cancers.<sup>47</sup> In contrast, these critically short telomeres also suppress tumor formation in cancer-prone *Ink4a/Arf*-deficient mice that still possess intact DNA damage responses.<sup>48</sup> The phenotypes obtained from studies using *Terc*<sup>-/-</sup> mice support the tumor suppressive role of intact telomeres in maintaining genomic integrity, and prove the intimate genetic interaction between telomere regulation and p53-governed genomic surveillance.

*Tert* deficiency also results in overtly similar phenotypes to *Terc* deficiency in tumorigenesis, but the phenotypic manifestations are not completely identical, thus revealing the extra-telomeric role of TERT in tumorigenesis. The seminal observation was obtained from *in vitro* experiments employing human cell lines. Immortalized human cells are frequently transformed by introducing an oncogene such as ras; however, oncogenic ras cannot fully transform immor-

talized human cells that are *TERT*-deficient ALT cells.<sup>49</sup> Interestingly, hTERT overexpression confers fully malignant traits to cells expressing oncogenic ras.<sup>49</sup> A hemagglutinin (HA) epitope-tagged hTERT (hTERT-HA) that is defective in maintaining telomeres *in vivo* also exhibits comparable effects on cellular transformation.<sup>49</sup> Similarly, hTERT overexpression in human mammary epithelial cells with epigenetically silenced *p16<sup>INK4a</sup>* resulted in increased resistance to growth arrest mediated by transforming growth factor  $\beta$  (TGF- $\beta$ ).<sup>50</sup> Because resistance to TGF- $\beta$ -induced growth inhibition is independent of telomere length,<sup>50</sup> TERT possesses telomere-independent roles that cooperate with *p16<sup>INK4a</sup>* inactivation to promote tumor development. These results clearly demonstrate an oncogenic role of TERT beyond telomeres.

Extensive studies adopting diverse models have recently revealed the extra-telomeric roles of oncogenic TERT at an organismal level (Table 1). For example, telomere dysfunction in late generation *Terc*<sup>-/-</sup> mice enhances the initiation of hepatocellular carcinogenesis, but suppresses progression into fully malignant carcinomas.<sup>51</sup> In contrast, enhanced tumor initiation does not occur in late generations of *Tert*<sup>-/-</sup> mice,<sup>35</sup> indicating a possible oncogenic effect of TERT, other than telomeres, in tumorigenesis. Strong induction of TERT expression in hepatic neoplasms may also support its procarcinogenic effect on hepatic tumorigenesis.<sup>35</sup> Consistently, transgenic overexpression of *Tert* promotes the development of spontaneous cancers in ageing mice.<sup>52</sup> When TERT overexpression is targeted to basal keratinocytes using the bovine keratin 5 promoter, these transgenic mice show normal telomere length in their stratified epithelia even with high levels of telomerase activity.<sup>15</sup> Interestingly, these mice are more susceptible to experimental skin carcinogenesis employing 7,12-dimethylbenz[a]anthracene and 12-*o*-tetradecanoylphorbol 13-acetate than wild-type mice.<sup>15</sup> In addition, TERT overexpression actively promotes proliferation in epidermal tissues without telomere elongation.<sup>15</sup> These results from mouse models suggest extra-telomeric roles of TERT, particularly in promoting tumor progression.

Telomerase mouse models have been also extensively used to validate telomerase as an important target for anti-cancer therapies. Since telomere dysfunction increases the chemo-sensitivity of *Terc*-deficient transformed MEFs, the combination of chemotherapy and telomerase inhibition may be an effective anticancer approach.<sup>53</sup> Recently, Ding, et al.<sup>34</sup> showed that telomerase reactivation by conditional rescue of *Tert* expression in mice with dysfunctional telo-

meres resulted in bone metastases of prostate tumors. Although the authors did not discuss the extra-telomeric roles of TERT in their study, this report is reminiscent of ALT cell transformation by hTERT overexpression.<sup>49</sup> From this standpoint, anti-telomeric drugs are considered as an effective strategy for curing cancers. However, anti-telomerase therapy certainly provokes ALT and mitochondrial adaptive mechanisms in cancer,<sup>54</sup> and with respect to the extra-telomeric functions of telomerase, anti-telomeric drugs may not be the best drug candidates.<sup>55</sup>

Taken together, telomerase exerts pleiotropic effects in cancer both dependent on and independent from its roles in telomeres. As described in Table 1, there are complex genetic interactions of telomerase with diverse genes. In conjunction with the currently emerging mechanisms of extra-telomeric roles, telomerase mouse models will expedite the invention of anti-telomerase strategies for cancer treatment.

## REGULATION OF STEM CELLS BY TELOMERASE

Stem cells support tissue homeostasis and regeneration after certain types of damage. Because stem cells possess self-renewal potential and indefinitely propagate, high levels of telomerase activity should be essential for telomere maintenance. Therefore, extensive studies have been conducted to identify the patho-physiological consequences of telomerase deficiency or overexpression in stem cell function using diverse telomerase mouse models. However, telomere dysfunction is likely to affect stem cell functions in a context-dependent manner. Indeed, late-generation *Terc*<sup>-/-</sup> HSCs with short telomeres exhibit reduced proliferation capacity, but still possess long-term repopulating ability.<sup>56</sup> Interestingly, when serially transplanted into recipient mice, the telomeres are considerably shortened even in wild-type HSCs, which is accelerated by approximately 2-fold in both *Terc*<sup>-/-</sup> and *Tert*<sup>-/-</sup> mice.<sup>16</sup> Consistently, these telomerase-deficient HSCs exhibit considerably reduced replicative capacity compared to wild-type HSCs.<sup>16</sup> However, although the telomere length of HSCs is constantly maintained by TERT overexpression in the transgenic mice, the long-term transplantation capacity of HSCs is not enhanced.<sup>57</sup> Furthermore, *Tert* deficiency exacerbates senescence and the sensitivity of *ataxia-telangiectasia mutated* deficient murine HSCs against ROS-induced apoptosis, which does not accompany telomere shortening or dysfunction.<sup>17</sup> These results

**Table 1.** Phenotypes of Telomerase Mouse Models

Mouse models	Affected tissue/organ	Dependence on telomere length?	Phenotypes	Refs	Functions
$\alpha$ MHC* <i>-hTERT-Tg</i>	Heart	Yes	Promotes cardiac myocyte survival	Oh, et al. <sup>33</sup>	Survival
CAG <sup>†</sup> <i>-Tert-Tg</i>	Whole body	No	Protects neuronal cells against ischemic cell death	Kang, et al. <sup>18</sup>	
CAG <sup>†</sup> <i>-hTERT-Tg</i>	Whole body	No	Protects motor neurons from sciatic nerve axotomy induced apoptosis	Lee, et al. <sup>44</sup>	
<i>Tert</i> <sup>-/-</sup>	Whole body	No	Mitochondrial TERT may contribute to the anti-ageing and anti-apoptosis	Haendeler, et al. <sup>69</sup>	
<i>Terc</i> <sup>-/-</sup>	Whole body	Yes	Defect in the closing of neural tube; decreased overall fitness and well-being; progressive loss of organismal viability	Herrera, et al. <sup>31</sup> Rudolph, et al. <sup>46</sup> Herrera, et al. <sup>70</sup>	Tumorigenesis
<i>Terc</i> <sup>-/-</sup>	Whole body	Yes	Frequent chromosomal abnormalities and tumor formation	Blasco, et al. <sup>45</sup>	
<i>Terc</i> <sup>-/-</sup>	Whole body	Yes	Decreases cell proliferation and impaired function of reproductive organs.	Lee, et al. <sup>71</sup>	
<i>Terc</i> <sup>-/-</sup> <i>p53</i> <sup>+/- or -/-</sup>	Whole body	Yes	Results in genetic catastrophe; prone to development of epithelial cancers	Chin, et al. <sup>40</sup> Artandi, et al. <sup>47</sup>	
K5 <sup>‡</sup> <i>-Tert-Tg</i>	Skin	No	Increased cell proliferation and wound-healing	González-Suárez, et al. <sup>15</sup>	Stem cell behavior
Lck <sup>§</sup> <i>-Tert-Tg</i>	Thymocytes	No	High incidence of spontaneous T-cell lymphoma	Canela, et al. <sup>72</sup>	
CAG-rtTA-i- <i>Tert</i> <sup>  </sup> <i>Terc</i> <sup>+/- or +/- or -/-</sup>	Whole body	No	Induced proliferation of resting epithelial stem cells	Sarin, et al. <sup>13</sup>	Gene regulation/ cell signaling
LSL <i>Tert</i> <sup>¶</sup> <i>p53</i> <sup>L/L</sup> <i>Pten</i> <sup>L/L</sup> <i>PB-Cre</i>	Prostate tissues	Yes	Telomerase reactivation confers tumor cells to metastatic potential	Ding, et al. <sup>34</sup>	
K5- <i>Tert-Tg</i> <i>Terc</i> <sup>+/+ or +/-</sup>	Skin	No	Increased stem cell mobility	Flores, et al. <sup>21</sup>	
<i>Tert</i> <sup>-/-</sup> , <i>Atm</i> <sup>-/-</sup>	Whole body	No	Decelerated ageing through protection of HSCs	Nitta, et al. <sup>17</sup>	Gene regulation/ cell signaling
iK5- <i>Tert</i> <sup>¶</sup> -Tg <sup>**</sup> , iK5- <i>Tert-Tg</i>	Skin	No	Promotes epithelial proliferation by controlling transcription of genes	Choi, et al. <sup>60</sup>	
<i>Tert</i> <sup>-/-</sup>	Whole body	No	Transcriptionally induce Wnt/ $\beta$ -catenin signaling pathway	Park, et al. <sup>61</sup>	
<i>Tert</i> <sup>-/-</sup> , <i>Terc</i> <sup>+/- or -/-</sup>	Whole body	No	Induction of NF- $\kappa$ B-dependent genes	Ghosh, et al. <sup>65</sup>	

TERT, telomerase reverse transcriptase; HSC, hematopoietic stem cell.

\*Mouse  $\alpha$ -myosin heavy chain (MHC) promoter.

†Human cytomegalovirus immediate-early enhancer linked to the chicken  $\beta$ -actin promoter (CAG).

‡Bovine keratin 5 promoter (K5).

§Thymus-specific light-chain kinase (Lck) promoter.

||Actin-rtTA<sup>†</sup>;tetop-TERT<sup>†</sup> (termed as doxycycline-inducible *Tert* or *i-Tert*).

¶Lox-Stop-Lox cassette.

\*\*Doxycycline-inducible expression of mutant TERT without telomerase activity under the regulation of K5-rtTA promoter.

suggest that telomerase may regulate the long-term replicative capacity of HSCs independently of telomere length.

Epidermal stem cells are also regulated by telomerase both dependent and independent of telomeres. In late generation *Terc*<sup>-/-</sup> mice, epidermal stem cell functions are significantly suppressed by critically short telomeres.<sup>21,58</sup> However, epidermal overexpression of TERT under the control

of the K5 promoter does not alter telomere length, but promotes stem cell mobilization, hair growth, and stem cell proliferation *in vitro*.<sup>18</sup> Similarly, transgenic mice conditionally overexpressing TERT show robust hair growth via proliferation of quiescent, multipotent stem cells in the hair follicles.<sup>13</sup> These phenotypes are also reproduced in a *Terc*-deficient genetic background without telomere dysfunction,<sup>13</sup>

thereby indicating the extra-telomeric activity of TERT.

In addition to the critical roles of p53 in mediating phenotypic manifestations against critically short telomeres,<sup>59</sup> clues for the molecular mechanisms governing the extra-telomeric roles of telomerase have been obtained by identifying the positive effect of TERT on gene expression. Choi, et al.<sup>60</sup> found that TERT triggers a rapid change in gene expression in the skin and hair follicles. This gene expression pattern significantly overlaps those controlling natural hair follicle cycling in wild-type mice. TERT affects the developmental program mediated by Myc and Wnt, which is intimately associated with stem cell function and cancer.<sup>60</sup> Furthermore, TERT binds BRG1 (also called SMARCA4), a SWI/SNF-related chromatin remodeling protein, and directly modulates Wnt/ $\beta$ -catenin signaling as a cofactor in the  $\beta$ -catenin transcriptional complex.<sup>61</sup> Therefore, independently of telomeres, TERT can act as a transcriptional regulator that is directly involved in stem cell functions, including in mouse epidermal tissues.

## CONCLUSIONS

For more than a decade, diverse telomerase mouse models have provided us with precious opportunities for evaluating the patho-physiological significance of telomerase in genetically defined environments and at an organismal level. With an emphasis on defective telomeres, these mouse models have considerably contributed to understanding a broad spectrum of phenomena associated with cancer and ageing. Furthermore, growing evidence has indicated that defective telomerase functions are involved in distinct diseases other than human cancers including dyskeratosis congenita, atherosclerosis, and renal diseases.<sup>62-64</sup> The list of disease-associated mutations has been expanding. To genetically define the pathological aspects and thus to establish animal models of these mutations, novel mouse models should be still generated and analyzed.

Despite the evident roles in telomeres, currently emerging extra-telomeric functions of telomerase are completely changing the scope of this enzyme. Notably, the direct roles of TERT in transcriptional regulation (e.g. Wnt/ $\beta$ -catenin and nuclear factor- $\kappa$ B or NF $\kappa$ B) provide good rationale for several phenotypes that cannot be explained by telomere dysfunction, and their physiological significance has been also confirmed using telomerase mouse models.<sup>62,64,65</sup> As might be expected, these lines of evidence make us consider that di-

verse observations supporting extra-telomeric roles of telomerase should be scrutinized and validated *in vivo* by generating novel mouse models. For example, in addition to the effect of short telomeres on mitochondria,<sup>39</sup> mitochondrial targeting of telomerase upon certain stressful conditions<sup>66</sup> and the recently identified RNA-dependent RNA polymerase activity of TERT,<sup>67,68</sup> indicates that telomerase has direct roles in mitochondria. Furthermore, considering the important roles of telomerase in cellular homeostasis, telomerase may be a critical factor for regulating the subcellular organelle homeostasis. Undoubtedly, we believe that these extra-telomeric functions of telomerase should be intimately associated with life span regulation, and that some regions of TERT, other than the RT domain, will be required for mediating protein-protein interactions with known functions in controlling the life span of an organism. In this context, we cannot rule out speculations for divergent mechanisms of telomerase function regulating survival, tumor progression, development/differentiation, and stress responses. These extra-telomeric functions have inevitably complicated the phenotypic manifestations elicited by dysfunctional telomeres and vice versa; thus, to separate these distinct functions of telomerase, more sophisticated genetic strategies should be developed in mice.

## ACKNOWLEDGEMENTS

This work was supported by National Research Foundation of Korea (NRF) grants funded by Ministry of Education, Science and Technology (MEST) of the Korean government (2009-0081177, 2010-0020878); a Korea Healthcare Technology R&D Project from the Ministry for Health & Welfare Affairs (A085136). M.A. was also supported by Higher Education Commission (HEC) of Pakistan government.

## REFERENCES

1. Bischoff C, Graakjaer J, Petersen HC, Hjelmborg Jv, Vaupel JW, Bohr V, et al. The heritability of telomere length among the elderly and oldest-old. *Twin Res Hum Genet* 2005;8:433-9.
2. Kipling D, Cooke HJ. Hypervariable ultra-long telomeres in mice. *Nature* 1990;347:400-2.
3. Aubert G, Lansdorp PM. Telomeres and aging. *Physiol Rev* 2008;88:557-79.
4. Lansdorp PM. Telomere length and proliferation potential of hematopoietic stem cells. *J Cell Sci* 1995;108(Pt 1):1-6.
5. Heidinger BJ, Blount JD, Boner W, Griffiths K, Metcalfe NB,

- Monaghan P. Telomere length in early life predicts lifespan. *Proc Natl Acad Sci U S A* 2012;109:1743-8.
6. Cohen SB, Graham ME, Lovrecz GO, Bache N, Robinson PJ, Reddel RR. Protein composition of catalytically active human telomerase from immortal cells. *Science* 2007;315:1850-3.
  7. Bianchi A, Shore D. Early replication of short telomeres in budding yeast. *Cell* 2007;128:1051-62.
  8. Laterreur N, Eschbach SH, Lafontaine DA, Wellinger RJ. A new telomerase RNA element that is critical for telomere elongation. *Nucleic Acids Res* 2013;41:7713-24.
  9. Wright WE, Piatyszek MA, Rainey WE, Byrd W, Shay JW. Telomerase activity in human germline and embryonic tissues and cells. *Dev Genet* 1996;18:173-9.
  10. Kim NW, Piatyszek MA, Prowse KR, Harley CB, West MD, Ho PL, et al. Specific association of human telomerase activity with immortal cells and cancer. *Science* 1994;266:2011-5.
  11. Allshire RC, Dempster M, Hastie ND. Human telomeres contain at least three types of G-rich repeat distributed non-randomly. *Nucleic Acids Res* 1989;17:4611-27.
  12. Prowse KR, Greider CW. Developmental and tissue-specific regulation of mouse telomerase and telomere length. *Proc Natl Acad Sci U S A* 1995;92:4818-22.
  13. Sarin KY, Cheung P, Gilson D, Lee E, Tennen RI, Wang E, et al. Conditional telomerase induction causes proliferation of hair follicle stem cells. *Nature* 2005;436:1048-52.
  14. Starling JA, Maule J, Hastie ND, Allshire RC. Extensive telomere repeat arrays in mouse are hypervariable. *Nucleic Acids Res* 1990;18:6881-8.
  15. González-Suárez E, Samper E, Ramírez A, Flores JM, Martín-Caballero J, Jorcano JL, et al. Increased epidermal tumors and increased skin wound healing in transgenic mice overexpressing the catalytic subunit of telomerase, mTERT, in basal keratinocytes. *EMBO J* 2001;20:2619-30.
  16. Allsopp RC, Morin GB, DePinho R, Harley CB, Weissman IL. Telomerase is required to slow telomere shortening and extend replicative lifespan of HSCs during serial transplantation. *Blood* 2003;102:517-20.
  17. Nitta E, Yamashita M, Hosokawa K, Xian M, Takubo K, Arai F, et al. Telomerase reverse transcriptase protects ATM-deficient hematopoietic stem cells from ROS-induced apoptosis through a telomere-independent mechanism. *Blood* 2011;117:4169-80.
  18. Kang HJ, Choi YS, Hong SB, Kim KW, Woo RS, Won SJ, et al. Ectopic expression of the catalytic subunit of telomerase protects against brain injury resulting from ischemia and NMDA-induced neurotoxicity. *J Neurosci* 2004;24:1280-7.
  19. Folini M, Brambilla C, Villa R, Gandellini P, Vignati S, Paduano F, et al. Antisense oligonucleotide-mediated inhibition of hTERT, but not hTERC, induces rapid cell growth decline and apoptosis in the absence of telomere shortening in human prostate cancer cells. *Eur J Cancer* 2005;41:624-34.
  20. Rahman R, Latonen L, Wiman KG. hTERT antagonizes p53-induced apoptosis independently of telomerase activity. *Oncogene* 2005;24:1320-7.
  21. Flores I, Cayuela ML, Blasco MA. Effects of telomerase and telomere length on epidermal stem cell behavior. *Science* 2005;309:1253-6.
  22. Heaphy CM, Subhawong AP, Hong SM, Goggins MG, Montgomery EA, Gabrielson E, et al. Prevalence of the alternative lengthening of telomeres telomere maintenance mechanism in human cancer subtypes. *Am J Pathol* 2011;179:1608-15.
  23. Sýkorová E, Fajkus J. Structure-function relationships in telomerase genes. *Biol Cell* 2009;101:375-92.
  24. Ulaner GA, Hu JF, Vu TH, Giudice LC, Hoffman AR. Telomerase activity in human development is regulated by human telomerase reverse transcriptase (hTERT) transcription and by alternate splicing of hTERT transcripts. *Cancer Res* 1998;58:4168-72.
  25. Haendeler J, Hoffmann J, Diehl JF, Vasa M, Spyridopoulos I, Zeicher AM, et al. Antioxidants inhibit nuclear export of telomerase reverse transcriptase and delay replicative senescence of endothelial cells. *Circ Res* 2004;94:768-75.
  26. Khoo CM, Carrasco DR, Bosenberg MW, Paik JH, Depinho RA. Ink4a/Arf tumor suppressor does not modulate the degenerative conditions or tumor spectrum of the telomerase-deficient mouse. *Proc Natl Acad Sci U S A* 2007;104:3931-6.
  27. Siegl-Cachedenier I, Muñoz P, Flores JM, Klatt P, Blasco MA. Deficient mismatch repair improves organismal fitness and survival of mice with dysfunctional telomeres. *Genes Dev* 2007;21:2234-47.
  28. Liu L, Franco S, Spyropoulos B, Moens PB, Blasco MA, Keefe DL. Irregular telomeres impair meiotic synapsis and recombination in mice. *Proc Natl Acad Sci U S A* 2004;101:6496-501.
  29. Herrera E, Martínez-A C, Blasco MA. Impaired germinal center reaction in mice with short telomeres. *EMBO J* 2000;19:472-81.
  30. He H, Wang Y, Guo X, Ramchandani S, Ma J, Shen MF, et al. Pot1b deletion and telomerase haploinsufficiency in mice initiate an ATR-dependent DNA damage response and elicit phenotypes resembling dyskeratosis congenita. *Mol Cell Biol* 2009;29:229-40.
  31. Herrera E, Samper E, Blasco MA. Telomere shortening in mTR-/- embryos is associated with failure to close the neural tube. *EMBO J* 1999;18:1172-81.
  32. Leri A, Franco S, Zacheo A, Barlucchi L, Chimenti S, Limana F, et al. Ablation of telomerase and telomere loss leads to cardiac dilatation and heart failure associated with p53 upregulation. *EMBO J* 2003;22:131-9.
  33. Oh H, Taffet GE, Youker KA, Entman ML, Overbeek PA, Michael LH, et al. Telomerase reverse transcriptase promotes cardiac muscle cell proliferation, hypertrophy, and survival. *Proc Natl Acad Sci U S A* 2001;98:10308-13.
  34. Ding Z, Wu CJ, Jaskeliouff M, Ivanova E, Kost-Alimova M, Prottopov A, et al. Telomerase reactivation following telomere dysfunction yields murine prostate tumors with bone metastases. *Cell* 2012;148:896-907.
  35. Farazi PA, Glickman J, Horner J, Depinho RA. Cooperative interactions of p53 mutation, telomere dysfunction, and chronic liver damage in hepatocellular carcinoma progression. *Cancer Res* 2006;66:4766-73.
  36. Rajaraman S, Choi J, Cheung P, Beaudry V, Moore H, Artandi SE. Telomere uncapping in progenitor cells with critical telomere shortening is coupled to S-phase progression in vivo. *Proc Natl Acad Sci U S A* 2007;104:17747-52.
  37. Jaskeliouff M, Muller FL, Paik JH, Thomas E, Jiang S, Adams AC, et al. Telomerase reactivation reverses tissue degeneration in aged telomerase-deficient mice. *Nature* 2011;469:102-6.
  38. Cosme-Blanco W, Shen MF, Lazar AJ, Pathak S, Lozano G, Murtani AS, et al. Telomere dysfunction suppresses spontaneous tumorigenesis in vivo by initiating p53-dependent cellular senescence. *EMBO Rep* 2007;8:497-503.
  39. Sahin E, Depinho RA. Linking functional decline of telomeres, mitochondria and stem cells during ageing. *Nature* 2010;464:520-8.
  40. Chin L, Artandi SE, Shen Q, Tam A, Lee SL, Gottlieb GJ, et al.

- p53 deficiency rescues the adverse effects of telomere loss and cooperates with telomere dysfunction to accelerate carcinogenesis. *Cell* 1999;97:527-38.
41. Xu D, Wang Q, Gruber A, Björkholm M, Chen Z, Zaid A, et al. Downregulation of telomerase reverse transcriptase mRNA expression by wild type p53 in human tumor cells. *Oncogene* 2000;19:5123-33.
  42. Lee MK, Hande MP, Sabapathy K. Ectopic mTERT expression in mouse embryonic stem cells does not affect differentiation but confers resistance to differentiation- and stress-induced p53-dependent apoptosis. *J Cell Sci* 2005;118(Pt 4):819-29.
  43. Zhu H, Fu W, Mattson MP. The catalytic subunit of telomerase protects neurons against amyloid beta-peptide-induced apoptosis. *J Neurochem* 2000;75:117-24.
  44. Lee J, Sung YH, Cheong C, Choi YS, Jeon HK, Sun W, et al. TERT promotes cellular and organismal survival independently of telomerase activity. *Oncogene* 2008;27:3754-60.
  45. Blasco MA, Lee HW, Hande MP, Samper E, Lansdorp PM, DePinho RA, et al. Telomere shortening and tumor formation by mouse cells lacking telomerase RNA. *Cell* 1997;91:25-34.
  46. Rudolph KL, Chang S, Lee HW, Blasco M, Gottlieb GJ, Greider C, et al. Longevity, stress response, and cancer in aging telomerase-deficient mice. *Cell* 1999;96:701-12.
  47. Artandi SE, Chang S, Lee SL, Alson S, Gottlieb GJ, Chin L, et al. Telomere dysfunction promotes non-reciprocal translocations and epithelial cancers in mice. *Nature* 2000;406:641-5.
  48. Greenberg RA, Chin L, Femino A, Lee KH, Gottlieb GJ, Singer RH, et al. Short dysfunctional telomeres impair tumorigenesis in the INK4a(delta2/3) cancer-prone mouse. *Cell* 1999;97:515-25.
  49. Stewart SA, Hahn WC, O'Connor BF, Banner EN, Lundberg AS, Modha P, et al. Telomerase contributes to tumorigenesis by a telomere length-independent mechanism. *Proc Natl Acad Sci U S A* 2002;99:12606-11.
  50. Stampfer MR, Garbe J, Levine G, Lichtsteiner S, Vasserot AP, Yaswen P. Expression of the telomerase catalytic subunit, hTERT, induces resistance to transforming growth factor beta growth inhibition in p16INK4A(-) human mammary epithelial cells. *Proc Natl Acad Sci U S A* 2001;98:4498-503.
  51. Farazi PA, Glickman J, Jiang S, Yu A, Rudolph KL, DePinho RA. Differential impact of telomere dysfunction on initiation and progression of hepatocellular carcinoma. *Cancer Res* 2003;63:5021-7.
  52. Artandi SE, Alson S, Tietze MK, Sharpless NE, Ye S, Greenberg RA, et al. Constitutive telomerase expression promotes mammary carcinomas in aging mice. *Proc Natl Acad Sci U S A* 2002;99:8191-6.
  53. Lee KH, Rudolph KL, Ju YJ, Greenberg RA, Cannizzaro L, Chin L, et al. Telomere dysfunction alters the chemotherapeutic profile of transformed cells. *Proc Natl Acad Sci U S A* 2001;98:3381-6.
  54. Hu X, Li Y, Li C, Fu Y, Cai F, Chen Q, et al. Combination of fucosanthin and conjugated linoleic acid attenuates body weight gain and improves lipid metabolism in high-fat diet-induced obese rats. *Arch Biochem Biophys* 2012;519:59-65.
  55. Roh JI, Sung YH, Lee HW. Clinical implications of antitelomeric drugs with respect to the nontelomeric functions of telomerase in cancer. *Onco Targets Ther* 2013;6:1161-6.
  56. Samper E, Fernández P, Eguía R, Martín-Rivera L, Bernad A, Blasco MA, et al. Long-term repopulating ability of telomerase-deficient murine hematopoietic stem cells. *Blood* 2002;99:2767-75.
  57. Allsopp RC, Morin GB, Horner JW, DePinho R, Harley CB, Weissman IL. Effect of TERT over-expression on the long-term transplantation capacity of hematopoietic stem cells. *Nat Med* 2003;9:369-71.
  58. Siegl-Cachedenier I, Flores I, Klatt P, Blasco MA. Telomerase reverses epidermal hair follicle stem cell defects and loss of long-term survival associated with critically short telomeres. *J Cell Biol* 2007;179:277-90.
  59. Flores I, Blasco MA. A p53-dependent response limits epidermal stem cell functionality and organismal size in mice with short telomeres. *PLoS One* 2009;4:e4934.
  60. Choi J, Southworth LK, Sarin KY, Venteicher AS, Ma W, Chang W, et al. TERT promotes epithelial proliferation through transcriptional control of a Myc- and Wnt-related developmental program. *PLoS Genet* 2008;4:e10.
  61. Park JI, Venteicher AS, Hong JY, Choi J, Jun S, Shkreli M, et al. Telomerase modulates Wnt signalling by association with target gene chromatin. *Nature* 2009;460:66-72.
  62. Gizard F, Heywood EB, Findeisen HM, Zhao Y, Jones KL, Cudejko C, et al. Telomerase activation in atherosclerosis and induction of telomerase reverse transcriptase expression by inflammatory stimuli in macrophages. *Arterioscler Thromb Vasc Biol* 2011;31:245-52.
  63. Mitchell JR, Wood E, Collins K. A telomerase component is defective in the human disease dyskeratosis congenita. *Nature* 1999;402:551-5.
  64. Shkreli M, Sarin KY, Pech MF, Papeta N, Chang W, Brockman SA, et al. Reversible cell-cycle entry in adult kidney podocytes through regulated control of telomerase and Wnt signaling. *Nat Med* 2011;18:111-9.
  65. Ghosh A, Saginc G, Leow SC, Khattar E, Shin EM, Yan TD, et al. Telomerase directly regulates NF- $\kappa$ B-dependent transcription. *Nat Cell Biol* 2012;14:1270-81.
  66. Ahmed S, Passos JF, Birket MJ, Beckmann T, Brings S, Peters H, et al. Telomerase does not counteract telomere shortening but protects mitochondrial function under oxidative stress. *J Cell Sci* 2008;121(Pt 7):1046-53.
  67. Maida Y, Masutomi K. RNA-dependent RNA polymerases in RNA silencing. *Biol Chem* 2011;392:299-304.
  68. Maida Y, Yasukawa M, Furuuchi M, Lassmann T, Possemato R, Okamoto N, et al. An RNA-dependent RNA polymerase formed by TERT and the RMRP RNA. *Nature* 2009;461:230-5.
  69. Haendeler J, Dröse S, Büchner N, Jakob S, Altschmied J, Goy C, et al. Mitochondrial telomerase reverse transcriptase binds to and protects mitochondrial DNA and function from damage. *Arterioscler Thromb Vasc Biol* 2009;29:929-35.
  70. Herrera E, Samper E, Martín-Caballero J, Flores JM, Lee HW, Blasco MA. Disease states associated with telomerase deficiency appear earlier in mice with short telomeres. *EMBO J* 1999;18:2950-60.
  71. Lee HW, Blasco MA, Gottlieb GJ, Horner JW 2nd, Greider CW, DePinho RA. Essential role of mouse telomerase in highly proliferative organs. *Nature* 1998;392:569-74.
  72. Canela A, Martín-Caballero J, Flores JM, Blasco MA. Constitutive expression of tert in thymocytes leads to increased incidence and dissemination of T-cell lymphoma in Lck-Tert mice. *Mol Cell Biol* 2004;24:4275-93.