



# The relationship between serum hepatitis B virus DNA level and liver histology in patients with chronic HBV infection

Changjiang Liu<sub>0</sub><sup>1</sup>, Li Wang<sup>2</sup>, Huizhong Xie<sub>0</sub><sup>1</sup>\*, Liyuan Zhang<sup>3</sup>, Bingshu Wang<sup>2</sup>, Chun Luo<sup>2</sup>, Suiqun Wang<sup>2</sup>, Mingliang Tang<sup>1</sup>, Zhongbiao Fu<sup>1</sup>, Hailan Ruan<sup>1</sup>, Zhengjin Liu<sup>1</sup>, Ling Wei<sup>1</sup>, Wenyi Yi<sup>1</sup>, Yunqian Xie<sup>1</sup>

- 1 Department of Gastroenterology, The Second Affiliated Hospital of Hainan Medical University, Haikou, Hainan Province, China, 2 Department of Pathology, The Second Affiliated Hospital of Hainan Medical University, Haikou, Hainan Province, China, 3 Department of Infectious Diseases, The Second Affiliated Hospital of Hainan Medical University, Haikou, Hainan Province, China
- \* hnxiehuizhong@163.com.



# **Background**

There is no consensus regarding the relationship between HBV DNA and liver fibrosis, and the relationship between HBV DNA and the degree of liver cirrhosis has not been reported in patients with chronic HBV infection.

#### **Methods**

From January 2011 to December 2016, liver biopsies were performed on 396 patients with chronic hepatitis B and cirrhosis. Assessments of liver fibrosis and cirrhosis were based on the Laennec staging system.

#### Results

Serum levels of HBV DNA were correlated with fibrosis and cirrhosis (KW = 73.946, P<0.001). Serum HBV DNA level was correlated with mild fibrosis, moderate to severe fibrosis and cirrhosis (P = 0.009, P<0.001, and P<0.001, respectively). The HBeAg-positive group and HBeAg-negative group showed significant differences in HBV DNA levels, and the rates of mild fibrosis, severe fibrosis and cirrhosis were significantly different between these two groups (F = 17.585, P<0.001 and F = 6.017, P = 0.003, respectively). The replication status of the serum HBV DNA affected fibrosis formation as well as cirrhosis ( $\chi^2$  = 53.76, P<0.001). In the HBeAg-positive group, the sensitivity, specificity and AUC values of HBV DNA as a predictor for mild fibrosis and cirrhosis were 64.3%, 78.94% and 0.818, respectively, and 81.0%, 69.2%, and 0.871, respectively. In the HBeAg-negative group, the sensitivity, specificity and AUC values of HBV DNA for liver sclerosis prediction were 48%, 76.8% and 0.697, respectively.



# OPEN ACCESS

Citation: Liu C, Wang L, Xie H, Zhang L, Wang B, Luo C, et al. (2018) The relationship between serum hepatitis B virus DNA level and liver histology in patients with chronic HBV infection. PLoS ONE 13(11): e0206060. https://doi.org/10.1371/journal.pone.0206060

**Editor:** Pavel Strnad, Medizinische Fakultat der RWTH Aachen, GERMANY

Received: May 11, 2018

Accepted: October 6, 2018

Published: November 7, 2018

Copyright: © 2018 Liu et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files

**Funding:** The authors received no specific funding for this work.

**Competing interests:** The authors have declared that no competing interests exist.



#### **Conclusions**

Different HBV DNA levels had different effects on the formation of fibrosis and sclerosis in liver tissues. HBV DNA levels can predict mild fibrosis and cirrhosis in liver tissue, which is enhanced in HBeAg-positive patients.

#### Introduction

Hepatitis B virus (HBV) infection can manifest as different clinical types, including infection with Hepatitis B surface antigen (HBsAg) positivity and non-clinical symptoms, chronic Hepatitis B, cirrhosis and liver cancer [1]. During disease progression, HBV infection can be characterized by different clinical types, fibrosis and pathological changes in the liver tissue. Elucidation of the pathological changes of liver tissue in these patients is important in determining patient treatment plans and assessing their prognosis. At present, the observation of pathological changes in the liver tissue is the gold standard for evaluating fibrosis and cirrhosis of liver tissues in HBV-related liver diseases [2]. There are many other ways to assess the degree of liver fibrosis, such as FibroScan; however, this instrument has not been popularized in China, especially in economically underdeveloped areas, because of its high cost. Serological determination of HBV DNA level is simple and can avoid the trauma of biopsy and the limitation of tissue size. This method is useful to evaluate and predict the degree of liver fibrosis and cirrhosis based on the different levels of HBV DNA in clinical practice [3]. The aims of this prospective study were to confirm the correlation between HBV DNA level and liver histology in 396 patients with HBV-related liver disease and provide a theoretical basis for clinical treatment and disease course assessment.

#### Materials and methods

#### Study population

From January 2011 to December 2016, a total of 610 patients with HBV-related liver diseases (including chronic hepatitis B and cirrhosis) who underwent liver biopsy were included in the study. The patients were all hospitalized in the Department of Gastroenterology, Department of Infectious Diseases or Department of Hepatobiliary Surgery. Demographic characteristics and laboratory data including age, sex, alanine aminotransferase (ALT) level, platelet count (PLT), HBeAg status and HBV DNA level were collected at the time of liver biopsy. All patients were HBsAg-positive for at least 6 months before enrolling in the study. Overall, 197 patients were excluded based on exclusion criteria, 17 patients were excluded due to unqualified liver biopsy samples and indeterminate pathological results. The flow diagram of the study population is shown in Fig 1. A final total of 396 HBV-related patients were enrolled in this study, with 287 males and 109 females. Their mean age was 49.62±13.56 years, and the age ranged from 20 to 80 years. The patients were diagnosed according to the EASL 2017 standard [4]. All participants provided written informed consent, and ethical approval for these studies was obtained from the Human Research Ethics Committee of the Second Affiliated Hospital of Hainan Medical University, China.

#### Serological HBV DNA assay

Serum ALT was detected with a Hitachi 7600 automatic biochemistry analyzer.



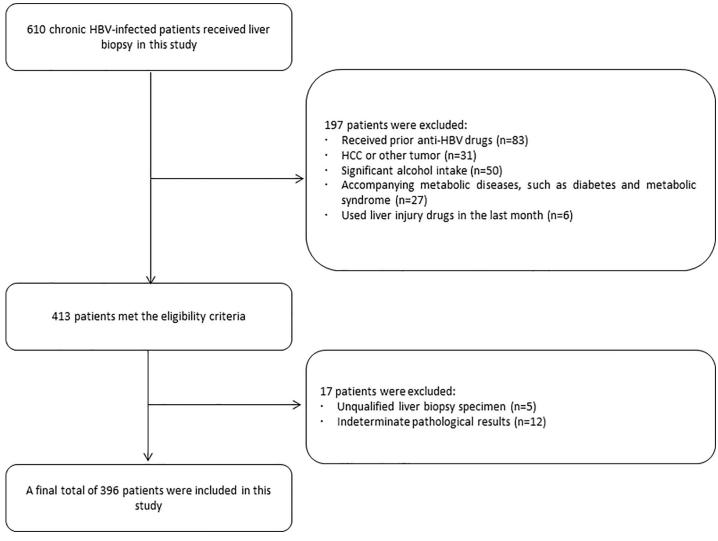


Fig 1. Flow diagram of the study population selection.

https://doi.org/10.1371/journal.pone.0206060.g001

PLT was detected with a Sysmex-1800 automated hematology analyzer with five classification levels. Quantification of serum HBV DNA was performed by real-time fluorescence quantitative polymerase chain reaction (PCR). The instrument was an FO-33A PCR amplification instrument from Hangzhou Bori Technology Co., Ltd. The kits were purchased from Shanghai Renaissance Medical Science and Technology Development Corporation. The specimen collection, preservation and detection process were carried out according to the instructions of the reagents. The detection range was  $2.24-8.24 \log IU/ml (1x10^2-1x10^9 copies/ml)$ . Based on the replication degree of HBV DNA, samples were divided into three groups. HBV DNA <2.24 log IU/ml was the no replication group,  $2.24 \le HBV DNA <4.24 \log IU/ml$  was the low replication group, and HBV DNA $\ge 4.24 \log IU/ml$  was the high replication group [5].

## Liver biopsy

A Madison SA-6000C color Doppler ultrasound instrument was used. The puncture device was a convex vibration probe, with a lateral direction needle on an externally fixed puncture



frame. The biopsy gun was a 16G, 20-cm-long disposable Bud puncture needle with a range of 22 mm. The patients were positioned on the left side, and the correct puncture entry point and path were selected after ultrasound localization. After skin disinfection and local anesthesia, the probe angle was fixed when the screen showed a clear target. A total of three liver tissues were obtained. All 396 liver biopsies were performed in the same B ultrasonic room of the hospital by the same ultrasound doctor. The needle was withdrawn rapidly, and a bandage was used for compression hemostasis. The liver tissues were sent to the pathology department for pathological observation.

## Histopathological assessment

The liver biopsy specimens were stained with hematoxylin-eosin (HE). The assessment of liver fibrosis and cirrhosis was performed at low magnification by three senior pathologists unaware of the clinical and virological results. According to the Laennec staging system (grades L0-L4C), liver fibrosis and cirrhosis were divided into 7 grades [6–9]. If there were different opinions on the stage of the liver biopsy, the patient was excluded.

## Statistical analysis

Statistical analyses were performed using SPSS software version 20.0 (SPSS Inc., Chicago, IL, United States). Due to the non-normal distribution of the HBV-DNA levels, these results are expressed logarithmically. The normally distributed variables are shown as the mean and standard deviation, and the groups were compared by analysis of variance. The rank sum test was used for the comparison between groups with non-normally distributed data. Numerical data were analyzed with the  $\chi^2$  test. The correlation between the Laennec staging system and HBV DNA levels was analyzed with Spearman's correlation. The influence of clinical indicators on liver fibrosis and cirrhosis was analyzed by multivariate logistic regression. The differences in HBV DNA levels in liver tissues of different pathological grades were compared with single factor variance analysis. The predictive value of HBV DNA level for liver fibrosis and cirrhosis was analyzed by ROC curve analysis. A P value of less than 0.05 was considered statistically significant.

#### **Results**

#### HBV DNA quantification and liver fibrosis and cirrhosis

The 396 liver tissue samples were divided into 7 groups based on the diagnostic criteria of the Laennec staging system. Age, sex, HBV DNA, PLT and ALT levels were compared among the seven groups. The results showed that there was no significant difference in sex among the groups, but the differences in other clinical indicators among the seven groups were statistically significant (P<0.001). The results are shown in Table 1.

#### Pathological diagnosis of liver fibrosis and cirrhosis

Among the 396 liver tissue samples, there were 21 cases of L0, 40 of L1, 75 of L2, 89 of L3, 94 of L4A, 44 of L4B and 33 of L4C, as shown in Fig 2.

# The correlation between the serum HBV DNA level and liver Laennec staging

There were 61 cases of mild fibrosis (L0—L1), and the mean HBV DNA level was 5.71±1.98 log IU/ml. The moderate to severe fibrosis (L0—L1) group had 164 cases, and the mean HBV DNA level was 4.83±1.76 log IU/ml. There were 171 cases in the liver cirrhosis (L4A - L4C)



Table 1	Correlation analysis	batwaan Laannac stagin	g and clinical indicators.
Table I.	Correlation analysis	between Laennec stagin	g and clinical indicators.

Laennec staging	age	n	Sex (male/female)	HBV DNA (log IU/ml)	ALT (U/L)	PLT (10 <sup>9</sup> /L)
L0	38.86±13.25	21	14/7	5.53±1.97	149.38±112.69	182.91±72.35
L1	40.83±13.12	40	26/14	5.80±1.99	171.05±161.09	160.33±47.19
L2	49.19±14.62	75	46/29	5.08±1.92	150.67±111.63	150.75±58.20
L3	48.13±13.66	89	63/26	4.62±1.59	132.03±106.65	133.24±64.52
L4A	52.81±11.86	94	77/17	3.69±1.55	64.95±49.55	118.94±43.74
L4B	54.36±9.52	44	34/10	3.61±1.49	63.27±44.08	117.02±55.69
L4C	56.7±10.86	33	27/6	3.18±1.26	62.97±44.52	97.33±35.52
$F/\chi^2$	47.38		12.42	73.95	79.75	67.75
P value	< 0.001		0.06	< 0.001	< 0.001	< 0.001

Laennec staging: L0 is normal, L1-L3 indicate liver fibrosis, L4A -L4C indicate cirrhosis.

https://doi.org/10.1371/journal.pone.0206060.t001

group, and the HBV DNA level was  $3.57\pm1.49 \log$  IU/ml. The results showed that the pathological fibrosis of liver tissues increased with the decrease in HBV DNA level. There was a significant difference in the HBV DNA levels among the three groups (P = 0.009, P<0.001, P<0.001). The results are shown in Fig 3.

# The correlation between the serum HBV DNA level and liver Laennec staging of the HBeAg-positive group

The 396 patients were divided into two groups: a HBeAg-positive group and HBeAg-negative group; 151 (38.13%) cases were positive, and 245 (61.87%) cases were negative. There was a significant difference between the HBV DNA levels and the liver tissues with mild fibrosis, moderate to severe fibrosis and cirrhosis in the HBeAg-positive group (F = 17.585, P < 0.001), and the HBeAg-negative group had the same results (F = 6.017, P = 0.003). The results are shown in Table 2.

#### HBV DNA levels and liver fibrosis and cirrhosis in different groups

Cases were divided into three groups according to serum HBV DNA level. The results showed a significant difference in the stage of liver fibrosis and cirrhosis among the three groups ( $\chi^2 = 53.76$ , P<0.001). The results are shown in Table 3.

#### Logistic analysis of liver biopsy stages and clinical indexes

Liver biopsy samples were divided into three groups according to the severity of liver fibrosis cirrhosis: a mild fibrosis group (L0-L1), severe fibrosis group (L2-L3) and liver cirrhosis group (L4A-L4C). Multiple factors including sex, age, HBeAg state, HBV DNA quantitative logarithm, PLT and ALT level were analyzed by ordered multi-classification logistic regression. The results showed that HBV DNA was an independent predictor of the degree of liver fibrosis and cirrhosis (OR 0.778, 95% CI 0.697-0.869, P<0.001).

# Prediction of liver fibrosis and cirrhosis by HBV DNA cut-off values in patients with different HBeAg states

The ROC curve analysis showed that the sensitivity and specificity of the prediction of mild liver fibrosis when the HBV DNA cut-off value was >6.68 log IU/ml were 64.3% and 78.94%, respectively, and the AUC value was 0.818 in the HBeAg-positive group. When the HBV DNA cut-off value was <4.94 log IU/ml, the sensitivity and specificity of the prediction of liver



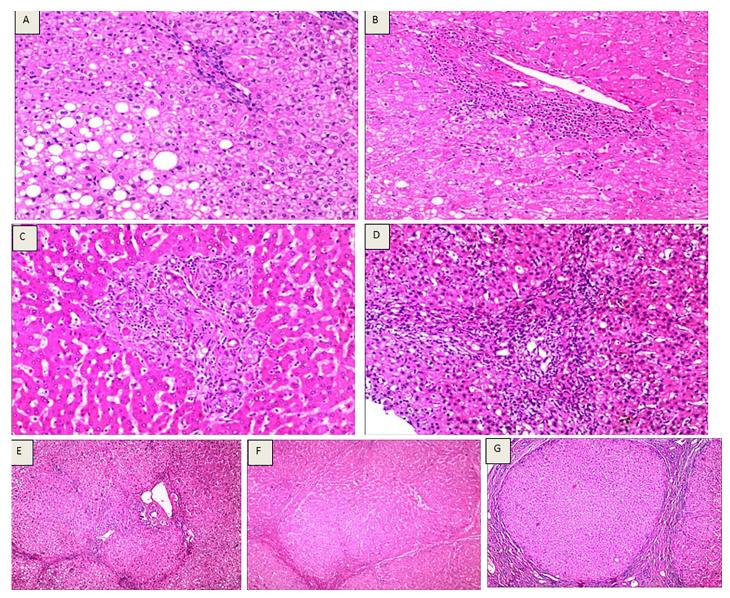


Fig 2. Assessments of liver fibrosis and cirrhosis according to the Laennec staging system. (A) Laennec staging 0, HE staining (10×10), hepatocytic loosening, steatosis, focal necrosis, mild lymphocytic infiltration in the portal area, and no fibrosis. (B) Laennec staging 1, HE staining (4×10), loose liver cell cytoplasm, fatty degeneration, focal point necrosis, mild fiber hyperplasia and enlargement, no fiber spacing, lymphocyte infiltration within the portal area. (C) Laennec staging 2, HE staining (4×10), hepatocyte eosinophilic degeneration, eosinophilic body formation, focal necrosis, enlargement of fibrous tissue in the portal area, formation of fibrous septa, infiltration of lymphocytes, and hyperplasia of small bile ducts. (D) Laennec staging 3, HE staining (4×10), liver cell cytoplasm of osteoporosis, eosinophilic degeneration, cholestasis, focal necrosis and severe periportal piecemeal necrosis, hyperplasia of fibrous tissue expansion, formation of fibrous septa, segmentation of hepatic lobule, periportal and septa lymphocytes. (E) Laennec staging 4A, HE staining (4×10). Hepatic lobule structure disappearance, hepatocyte loosening, steatosis, focal necrosis, fibrous tissue hyperplasia, fibrous septum, formation of false lobules, infiltration of lymphocytes in the portal area and intercellular septum. (F) Laennec staging 4B, HE staining (4×10), hepatic lobule structure disappearance, liver cell vacuolization, focal necrosis and moderate periportal piecemeal necrosis, hyperplasia of fibrous tissue formation of fibrous septa, pseudolobule formation, changes in part of the fiber interval width, portal area and fibrous septum lymphocytic infiltration. (G) Laennec staging 4C, HE staining (4 x 10), hepatic lobule structure disappearance, liver cell vacuolization, focal necrosis, portal fibrosis formation of fibrous septa, fibrous septum lymphocytic infiltration).

https://doi.org/10.1371/journal.pone.0206060.g002

cirrhosis were 81.0% and 69.2%, respectively, and the AUC value was 0.871 in the same group. However, in the HBeAg-negative group, an HBV DNA cut-off value of  $<2.68 \log$  IU/ml could predict liver cirrhosis with a sensitivity and specificity of 48.0% and 76.8%, respectively, and the AUC value was 0.697. The results are shown in Fig 4.



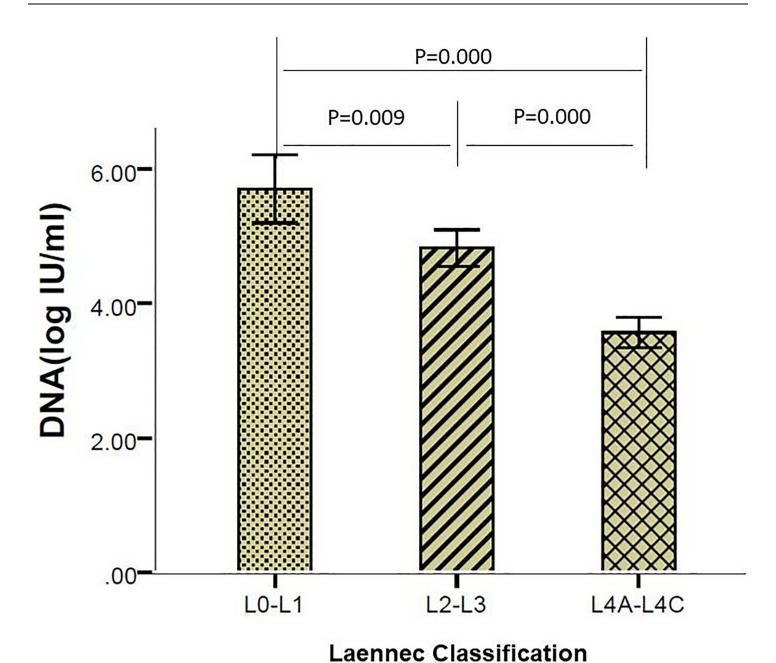


Fig 3. Comparison of the three Laennec grade groups and HBV DNA levels.

https://doi.org/10.1371/journal.pone.0206060.g003

Table 2. Comparison of the Laennec staging and HBeAg-positive HBV DNA in pathological liver tissues.

group	I	IBeAg positive	HBeAg negative			
	n (%)	HBV DNA (log IU/ml)	n (%)	HBV DNA (log IU/ml)		
L0-1	42(27.81)	6.50±1.63	19(7.76)	3.95±1.50		
L2-3	88(58.28)	5.32±1.72	76(31.02)	4.27±1.63		
L4A-L4C	21(13.91)	3.88±1.64	150(61.22)	3.53±1.47		
F		17.585		6.017		
P value		< 0.001		0.003		

https://doi.org/10.1371/journal.pone.0206060.t002



Table 3. Comparison of groups with different HBV DNA levels and liver fibrosis and cirrhosis.

HBV DNA groups	LO	L1	L2	L3	L4A	L4B	L4C	N
no replication	2	4	8	17	34	17	19	101
low replication	4	7	20	16	24	9	5	85
high replication	15	29	47	56	36	8	9	210
Total	21	40	75	89	94	44	33	396

 $\chi^2$  = 53.76, P<0.001

https://doi.org/10.1371/journal.pone.0206060.t003

# **Discussion**

The effect of HBV DNA level on liver fibrosis and cirrhosis is a research hotspot  $[\underline{10}-\underline{13}]$ . However, the results and conclusions of previous studies have not been consistent  $[\underline{14},\underline{15}]$ . The samples included in the studies were mostly patients with chronic hepatitis B  $[\underline{16},\underline{17}]$ . Few

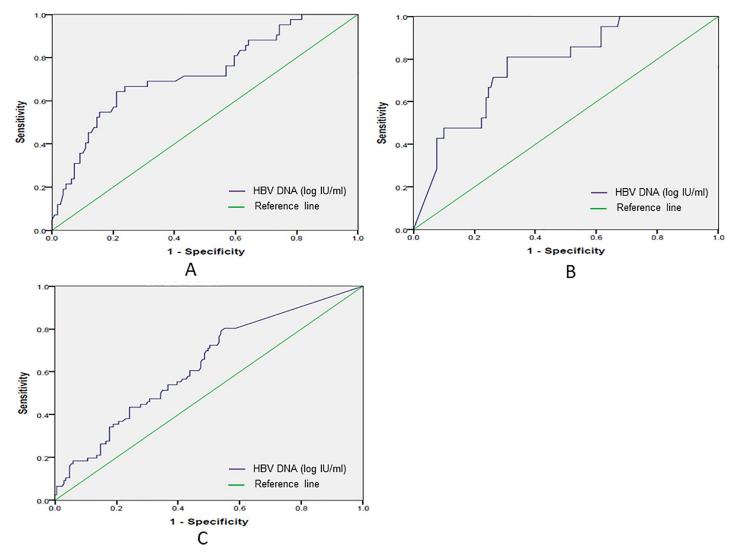


Fig 4. Prediction of liver fibrosis and cirrhosis by HBV DNA cut-off values. (A): L0-1 in HBeAg-positive patients. (B): L4A-4C in HBeAg-positive patients. (C): L4A-4C in HBeAg-negative patients).

https://doi.org/10.1371/journal.pone.0206060.g004



studies have investigated liver cirrhosis samples. Most publications used the Ishak, METAVIR and Batts systems [18–20] to classify the degree of fibrosis in the liver tissue. There are relatively few studies on the relationship between HBV DNA levels and different stages of liver cirrhosis. The purpose of our research was to explore the correlation between these variables and provide a theoretical basis for treatment strategies and evaluation of prognosis of patients with hepatitis B and different HBV DNA levels who develop liver fibrosis or cirrhosis.

In this study, we used the Laennec histological staging system to divide the liver tissues with fibrosis or cirrhosis into 7 grades according to the thickness and quantities of hepatic interlobular septum: grades L0, L1, L2, L3, L4A, L4B and L4C. The results showed that the Laennec staging system, to some extent, could reflect the degree of liver cirrhosis, the clinical staging and the liver function and predict the incidence of complications in patients with cirrhosis [8,21,22].

The results of our research showed that the HBV DNA level decreased gradually with the severity of Laennec staging of the liver tissue. According to the degree of liver fibrosis and cirrhosis, we divided the liver samples into the mild fibrosis group (L0-L1), severe fibrosis group (L2-L3) and liver cirrhosis group (L4A-L4C). The differences in HBV DNA level among the three groups were significant. HBV DNA was an independent predictor of liver fibrosis staging. The result was consistent with the findings of Diktas H [16] and Surat P [23].

According to the Laennec staging system, the degree of liver tissue cirrhosis was divided into the L4A, L4B and L4C groups. We compared the HBV DNA levels among these three groups. The results showed that there was no significant difference between the L4A and L4B groups (Z = -0.260, P = 0.795), but the differences between L4A and L4C and between L4B and L4C were statistically significant (Z = -2.013, P = 0.041; Z = -1.961, P = 0.049). The HBV DNA level gradually decreased with the severity of liver tissue cirrhosis.

The results of Kim JD [5] suggested that patients in the late stage of liver cirrhosis (decompensation period) could still have significant inflammation of the portal area even if their serum alanine transaminase was normal or serum viral load was very low (<2000 copies/ml). The Oliveira VO study confirmed that low-load HBV DNA did not indicate that liver injury ceased; instead, the liver cell injury continued [24]. HBV DNA can still be detected in liver tissues of patients with liver cirrhosis without HBV DNA detection in the serum [25]. The studies of Chen JD and Fung J suggested that the pathological processes in liver tissues of inactive HBV carriers could show progression, and some may even develop into end-stage liver disease or hepatocellular carcinoma [26,27]. The continuous suppression or removal of the virus could reduce the progression of inflammation and fibrosis in the liver, preventing fibrosis or cirrhosis of the liver [28,29]. Our results showed that low levels of HBV DNA and the liver pathological changes were not parallel. Thus, the HBV DNA level may not reflect liver disease progression. For hepatitis B cirrhosis patients, anti-viral inclusion based only on the HBV DNA level or clinical diagnosis should be further studied.

In this study, 396 HBV-related liver disease patients were divided into the HBeAg-positive group and HBeAg-negative group. The HBV DNA levels showed significant differences among the mild fibrosis, severe fibrosis and cirrhosis cases in the HBeAg-positive group (F = 17.585, P < 0.001), and the HBV DNA level decreased significantly with the severity in pathological changes of liver tissue. These findings were consistent with the studies of Xie Q [30] and Gao Y [31], who reported that extensive and repeated immune-mediated responses led to decreased viral load and liver damage. Our study suggested that the effects of HBV DNA level on the formation of liver fibrosis and cirrhosis might be enhanced in HBeAg-positive patients.

Many studies have reported that liver injury in patients with chronic hepatitis B was associated with HBV-mediated immune response rather than direct injury to hepatocytes [32,33].



The occurrence and development of liver fibrosis and cirrhosis might be caused by liver interstitial vascular chronic ischemia [34,35]. Chronic liver injury leads to endothelial cell injury and microvascular thrombosis, and then, cell proliferation and fibrosis gradually appear [36]. The degree of liver tissue injury may not have increased with the serum HBV DNA level, or there may be no parallel relationship between the two [1,37,38].

Our study results showed that the HBV DNA level decreased with the degree of liver fibrosis and cirrhosis. The replication of HBV DNA, a marker of HBV replication, depended on the liver cells, which may explain these findings. After infection by HBV, the body produces specific cellular and humoral immune responses that cause hepatocyte injury. With persistence of HBV infection, the cumulative immune damage increases gradually, and the extent of liver cell damage is also aggravated, causing extensive necrosis of the hepatocytes. Finally, hepatocytes are extensively necrotic, leading to a decrease in HBV, as replication is dependent on hepatocytes. Therefore, the more extensive the liver cell damage, the lower the HBV DNA load. We considered whether this process could similar to "seed growth dependent on the soil"; this hypothesis requires further study and discussion. Some scholars believe that the course of most patients with chronic hepatitis B is long. During this period, liver cell degeneration and necrosis occur repeatedly, and excess collagen deposition continues. All of these situations could lead to liver fibrosis or even cirrhosis. The study confirmed that the liver tissue injury and formation of liver fibrosis and cirrhosis had a significant correlation with the long-term infection of HBV DNA [39]. High replication of HBV DNA was an independent risk factor for the development of cirrhosis in chronic hepatitis B patients [40]. The continuous replication of HBV DNA in the liver cells resulted in the continuous aggravation of inflammatory necrosis and liver fibrosis. In patients infected by HBV, the degree of liver tissue inflammatory activity and fibrosis were significantly increased after repeated liver injury. Long-term HBV infection promoted the formation and severity of liver fibrosis and cirrhosis.

The etiology of liver disease in this study was only HBV, as other causes were excluded. The results of the study independently showed the influence of HBV DNA on liver fibrosis and sclerosis and their relationships. In this study, patients were divided into the HBeAg-positive group and the HBeAg-negative group according to HBeAg status so that we could evaluate liver fibrosis and sclerosis of patients infected by HBV at different stages.

Our study also has certain limitations. First, we lacked data on the HBV genotypes due to the limitations of our hospital's testing methods. Second, this study consists entirely of Chinese individuals in Hainan Province, who may have unique characteristics. Our results need to be confirmed by a multicenter study. The third limitation is that all patients included had not received regular antiviral treatment. With the current worldwide level of knowledge of hepatitis B, patients with chronic hepatitis B and cirrhosis who do not receive antiviral treatment are relatively rare, so the study is not repeatable.

This study results showed that the Laennec staging system could be used to classify the degree of liver fibrosis and cirrhosis according to the thickness and quantity of the interval. We investigated the relationship between the HBV DNA level and liver fibrosis and cirrhosis and provided one of the evidences supporting or discouraging antiviral therapy in patients at different stages. The relationship between HBV DNA level and liver fibrosis may suggest causality, and the results could provide support for antiviral therapy for patients at different stages. The study results suggested that the HBV DNA level decreased with the aggravation of fibrosis and cirrhosis of liver tissues. HBV DNA level was a good index for evaluating the degree of fibrosis and cirrhosis of patients with HBV-related liver diseases, especially HBeAg-positive patients.



# **Supporting information**

S1 Fig. Flow diagram of the study population selection.

(TIF)

S2 Fig. Assessments of liver fibrosis and cirrhosis according to the Laennec staging system. (TIF)

S3 Fig. Comparison of the three Laennec grade groups and HBV DNA levels. (TIF)

S4 Fig. Prediction of liver fibrosis and cirrhosis by HBV DNA cut-off values. (TIF)

S1 Dataset. Relevant data underlying the findings described in manuscript. (XLSX)

# **Acknowledgments**

We thank American Journal Experts (AJE) for English language editing.

#### **Author Contributions**

Data curation: Liyuan Zhang.

Formal analysis: Li Wang.

**Resources:** Mingliang Tang, Zhongbiao Fu, Hailan Ruan, Zhengjin Liu, Ling Wei, Wenyi Yi, Yunqian Xie.

Validation: Bingshu Wang, Chun Luo, Suiqun Wang.

Writing – original draft: Changjiang Liu.
Writing – review & editing: Huizhong Xie.

#### References

- Sarin SK, Kumar M, Lau GK, Abbas Z, Chan HL, Chen CJ, et al. Asian-Pacific clinical practice guidelines on the management of hepatitis B: a 2015 update. Hepatology International. 2016; 10:1–98.
- Wang J, Yan X, Yang Y, Chang H, Jia B, Zhao XA, et al. A novel predictive model using routinely clinical parameters to predict liver fibrosis in patients with chronic hepatitis B. Oncotarget. 2017; 8:59257– 59267. https://doi.org/10.18632/oncotarget.19501 PMID: 28938634
- Sanai FM, Helmy A, Bzeizi KI, Babatin MA, Al-Qahtani A, Al-Ashgar HA, et al. Discriminant value of serum HBV DNA levels as predictors of liver fibrosis in chronic hepatitis B. J Viral Hepat. 2011; 18:217– 225.
- European Association for the Study of the Liver. EASL 2017 clinical practice guidelines on the management of hepatitis B virus infection. J Hepatol. 2017; 67:370–398. <a href="https://doi.org/10.1016/j.jhep.2017.03.">https://doi.org/10.1016/j.jhep.2017.03.</a>
   PMID: 28427875
- Kim JD, Choi JY, Bae SH, Yoon SK, Yang JM, Han NI, et al. Hepatitis B virus load in serum does not reflect histologic activity in patients with decompensated cirrhosis. Clin Gastroenterol Hepatol. 2010; 8:60–65. https://doi.org/10.1016/j.cgh.2009.09.026 PMID: 19804841
- Sanai FM, Farah T, Albeladi K, Batwa F, Dahlan Y, Babatin MA, et al. Diminished accuracy of biomarkers of fibrosis in low replicative chronic hepatitis B. BMC Gastroenterol. 2017; 17:101. https://doi.org/10.1186/s12876-017-0658-x PMID: 28841822
- Kim MY, Cho MY, Baik SK, Park HJ, Jeon HK, Im CK, et al. Histological subclassification of cirrhosis using the Laennec fibrosis scoring system correlates with clinical stage and grade of portal hypertension. J Hepatol. 2011; 55:1004–1009. https://doi.org/10.1016/j.jhep.2011.02.012 PMID: 21354227



- Rastogi A, Maiwall R, Bihari C, Ahuja A, Kumar A, Singh T, et al. Cirrhosis histology and Laennec staging system correlate with high portal pressure. Histopathology. 2013; 62:731–741. https://doi.org/10.1111/his.12070 PMID: 23470026
- Wanless IR, Sweeney G, Dhillon AP, Guido M, Piga A, Galanello R, et al. Lack of progressive hepatic fibrosis during long-term therapy with deferiprone in subjects with transfusion-dependent beta-thalassemia. Blood. 2002; 100:1566–1569. https://doi.org/10.1182/blood-2002-01-0306 PMID: 12176871
- Alam MM, Mahtab MA, Akbar SM, Kamal M, Rahman S. Hepatic necroinflammation and severe liver fibrosis in patients with chronic hepatitis B with undetectable HBV DNA and persistently normal alanine aminotransferase. Bangladesh Med Res Counc Bull. 2014; 40:92–96. PMID: 26402972
- 11. Abdo AA, Bzeizi KI, Babatin MA, AlSohaibani F, AlMana H, Alsaad KO, et al. Predictors of significant fibrosis in chronic hepatitis B patients with low viremia. J Clin Gastroenterol. 2014; 48: 50–56.
- Pan AN, Xu WW, Luo YL, Yu HH, Hu YB, Sun QF, et al. A novel system for predicting liver histopathology in patients with chronic hepatitis B. Medicine (Baltimore). 2017; 96:e6465.
- Alavian SM, Imanieh MH, Imanieh MH. Predictive Factors in the Incidence of Cirrhosis in Chronic Hepatitis B Virus Infections. Hepat Mon. 2016; 16:e34790. <a href="https://doi.org/10.5812/hepatmon.34790">https://doi.org/10.5812/hepatmon.34790</a> PMID: 27330536
- 14. Harkisoen S, Arends JE, van den Hoek JA, Whelan J, van Erpecum KJ, Boland GJ, et al. Historic and current hepatitis B viral DNA and quantitative HBsAg level are not associated with cirrhosis in non-Asian women with chronic hepatitis B. Int J Infect Dis. 2014; 29:133–138. https://doi.org/10.1016/j.ijid. 2014.08.005 PMID: 25449247
- 15. Ke WM, Xie SB, Li XJ, Zhang SQ, Lai J, Ye YN, et al. There were no differences in serum HBV DNA level between HBeAg-positive and HBeAg-negative chronic hepatitis B with same liver histological necroinflammation grade but differences among grades 1, 2, 3 and 4 apportioned by the same hepatic parenchyma cell volume. J Viral Hepat. 2011; 18:637–645. https://doi.org/10.1111/j.1365-2893.2011. 01444.x PMID: 21794025
- Diktas H, Karacaer Z, Özturk II, Cicek H. Comparison of relationship between histopathological, serological and biochemical parameters in patients with chronic hepatitis B infection. Postgraduate Medical Journal. 2016; 92:693–696. https://doi.org/10.1136/postgradmedj-2016-134069 PMID: 27170311
- Vardar R, Gunsar F, Sertoz R, Ozacar T, Nart D, Barbet FY, et al. The relationship between HBV-DNA level and histology in patients with naive chronic HBV infection. Hepatogastroenterology. 2010; 57:908–912. PMID: 21033250
- Batts KP, Ludwig J. Chronic hepatitis. An update on terminology and reporting. Am J Surg Pathol. 1995; 19:1409–1417. PMID: 7503362
- Ishak K, Baptista A, Bianchi L, Callea F, De Groote J, Gudat F, et al. Histological grading and staging of chronic hepatitis. J Hepatol. 1995; 22:696–699. PMID: 7560864
- Bedossa P, Poynard T. An algorithm for the grading of activity in chronic hepatitis C. The METAVIR Cooperative Study Group. Hepatology. 1996; 24:289–293. <a href="https://doi.org/10.1002/hep.510240201">https://doi.org/10.1002/hep.510240201</a> PMID: 8690394
- Wang W, Li J, Pan R, A S, Liao C. Association of the Laennec staging system with degree of cirrhosis, clinical stage and liver function. Hepatol Int. 2015; 9:621–626. <a href="https://doi.org/10.1007/s12072-015-9648-7">https://doi.org/10.1007/s12072-015-9648-7</a> PMID: 26208820
- 22. Zhang EL, Zhang ZY, Wang SP, Xiao ZY, Gu J, Xiong M, et al. Predicting the severity of liver cirrhosis through clinical parameters. J Surg Res. 2016; 204:274–281. https://doi.org/10.1016/j.jss.2016.04.036 PMID: 27565061
- Praneenararat S, Chamroonkul N, Sripongpun P, Kanngurn S, Jarumanokul R, Piratvisuth T. HBV DNA level could predict significant liver fibrosis in HBeAg negative chronic hepatitis B patients with biopsy indication. BMC Gastroenterol. 2014; 14:218. https://doi.org/10.1186/s12876-014-0218-6 PMID: 25523185
- 24. Oliveira VO, Oliveira JP, França EV, Brito HL, Nascimento TV, Franca A. Advanced liver injury in patients with chronic hepatitis B and viral load below 2,000 IU/mL. Rev Inst Med Trop Sao Paulo. 2016; 58:65, https://doi.org/10.1590/S1678-9946201658065 PMID: 27680170
- 25. Lee CZ, Huang GT, Yang PM, Sheu JC, Lai MY, Chen DS. Correlation of HBV DNA levels in serum and liver of chronic hepatitis B patients with cirrhosis. Liver. 2002; 22:130–135. PMID: 12028407
- Chen JD, Yang HI, Iloeje UH, You SL, Lu SN, Wang LY, et al. Carriers of inactive hepatitis B virus are still at risk for hepatocellular carcinoma and liver-related death. Gastroenterology. 2010; 138:1747– 1754. https://doi.org/10.1053/j.gastro.2010.01.042 PMID: 20114048
- Fung J, Yuen MF, Yuen JC, Wong DK, Lai CL. Low serum HBV DNA levels and development of hepatocellular carcinoma in patients with chronic hepatitis B: a case-control study. Aliment Pharmacol Ther. 2007; 26:377–382. https://doi.org/10.1111/j.1365-2036.2007.03390.x PMID: 17635372



- 28. Brown A, Goodman Z. Hepatitis B-associated fibrosis and fibrosis/cirrhosis regression with nucleoside and nucleotide analogs. Expert Rev Gastroenterol Hepatol. 2012; 6:187–198. https://doi.org/10.1586/egh.12.4 PMID: 22375524
- Chung GE, Lee JH, Kim YJ. Does antiviral therapy reduce complications of cirrhosis? World J Gastroenterol. 2014; 20:7306–7311. https://doi.org/10.3748/wjg.v20.i23.7306 PMID: 24966601
- Xie Q, Hu X, Zhang Y, Jiang X, Li X, Li J. Decreasing hepatitis B viral load is associated with a risk of significant liver fibrosis in hepatitis B e antigen positive chronic hepatitis B. J Med Virol. 2014; 86:1828–1837. https://doi.org/10.1002/jmv.24000 PMID: 25145769
- Gao Y, Zou G, Ye J, Pan G, Rao J, Li F, et al. Relationship between hepatitis B surface antigen, HBV DNA quantity and liver fibrosis severity. Zhonghua Gan Zang Bing Za Zhi. 2015; 23:254–257. https://doi.org/10.3760/cma.j.issn.1007-3418.2015.04.005 PMID: 26133815
- Wu JF, Chang MH. Natural history of chronic hepatitis B virus infection from infancy to adult life—the mechanism of inflammation triggering and long-term impacts. J Biomed Sci. 2015; 22:92. https://doi. org/10.1186/s12929-015-0199-y PMID: 26487087
- **33.** Dandri M, Locarnini S. New insight in the pathobiology of hepatitis B virus infection. Gut. 2012; 61 Suppl 1:i6–17.
- Mancuso A. Cirrhosis development probably arises from chronic micro-vascular ischemia. Med Hypotheses. 2014; 82:244. https://doi.org/10.1016/j.mehy.2013.11.044 PMID: 24389105
- Mancuso A. The ischemic liver cirrhosis theory and its clinical implications. Med Hypotheses. 2016;
   94:4–6. https://doi.org/10.1016/j.mehy.2016.06.008 PMID: 27515188
- Mancuso A. Letter: why treatments of hepatitis C virus are less effective in cirrhosis. Aliment Pharmacol Ther. 2015; 41:707. https://doi.org/10.1111/apt.13118 PMID: 25736155
- Gish RG, Given BD, Lai CL, Locarnini SA, Lau JY, Lewis DL, et al. Chronic hepatitis B: Virology, natural history, current management and a glimpse at future opportunities. Antiviral Res. 2015; 121:47–58. https://doi.org/10.1016/j.antiviral.2015.06.008 PMID: 26092643
- Tseng TC, Kao JH. Treating Immune-tolerant Hepatitis B. J Viral Hepat. 2015; 22:77–84. https://doi. org/10.1111/jvh.12370 PMID: 25424771
- Mani H, Kleiner DE. Liver biopsy findings in chronic hepatitis B. Hepatology. 2009; 49:S61–71. https://doi.org/10.1002/hep.22930 PMID: 19399798
- Iloeje UH, Yang HI, Su J, Jen CL, You SL, Chen CJ, et al. Predicting cirrhosis risk based on the level of circulating hepatitis B viral load. Gastroenterology. 2006; 130:678–686. https://doi.org/10.1053/j.gastro. 2005.11.016 PMID: 16530509