

Conundrum of vitamin D on glucose and fuel homeostasis

Maria Mercedes Chang Villacreses, Rudrudee Karnchanasorn, Panadeekarn Panjawatanan, Horng-Yih Ou, Ken C Chiu

ORCID number: Maria Mercedes Chang Villacreses 0000-0001-9713-1450; Rudrudee Karnchanasorn 0000-0001-7134-1097; Panadeekarn Panjawatanan 0000-0003-3665-7904; Horng-Yih Ou 0000-0002-3350-6548; Ken C Chiu 0000-0002-2226-1039.

Author contributions: Chang Villacreses MM, Chiu KC, Karnchanasorn R, and Ou HY developed the central theme and concepts of this manuscript; Panjawatanan P collected the data and participated in data analyses with Chang Villacreses MM, Chiu KC, Karnchanasorn R, and Ou HY; Chang Villacreses MM and Chiu KC prepared the first draft of manuscript; Chang Villacreses MM, Chiu KC, Karnchanasorn R, and Ou HY took part in critical review and revision of manuscript.

Conflict-of-interest statement: Authors declare no conflict-of-interest.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works

Maria Mercedes Chang Villacreses, Panadeekarn Panjawatanan, Ken C Chiu, Department of Clinical Diabetes, Endocrinology, and Metabolism, City of Hope National Medical Center, Duarte, CA 91010, United States

Maria Mercedes Chang Villacreses, Ken C Chiu, Division of Endocrinology, Metabolism and Nutrition, Department of Internal Medicine, Harbor-UCLA Medical Center, Torrance, CA 90509, United States

Rudrudee Karnchanasorn, Division of Endocrinology, Department of Medicine, University of Kansas Medical Center, Kansas City, KS 66160, United States

Panadeekarn Panjawatanan, Department of Internal Medicine, Bassett Medical Center, Cooperstown, NY 13326, United States

Horng-Yih Ou, Department of Internal Medicine, National Cheng Kung University Hospital, National Cheng Kung University, Tainan 700, Taiwan

Corresponding author: Ken C Chiu, FACE, FACP, MD, Professor, Department of Clinical Diabetes, Endocrinology, and Metabolism, City of Hope National Medical Center, 1500 East Duarte Road, Duarte, CA 91010, United States. kenchiumd@gmail.com

Abstract

As an endocrine hormone, vitamin D plays an important role in bone health and calcium homeostasis. Over the past two decades, the non-calcemic effects of vitamin D were extensively examined. Although the effect of vitamin D on beta cell function were known for some time, the effect of vitamin D on glucose and fuel homeostasis has attracted new interest among researchers. Yet, to date, studies remain inconclusive and controversial, in part, due to a lack of understanding of the threshold effects of vitamin D. In this review, a critical examination of interventional trials of vitamin D in prevention of diabetes is provided. Like use of vitamin D for bone loss, the benefits of vitamin D supplementation in diabetes prevention were observed in vitamin D-deficient subjects with serum 25-hydroxyvitamin D < 50 nmol/L (20 ng/mL). The beneficial effect from vitamin D supplementation was not apparent in subjects with serum 25-hydroxyvitamin D > 75 nmol/L (30 ng/mL). Furthermore, no benefit was noted in subjects that achieved serum 25-hydroxyvitamin D > 100 nmol/L (40 ng/mL). Further studies are required to confirm these observations.

Key Words: Vitamin D; Glucose metabolism; Diabetes mellitus; Insulin sensitivity; Beta

on different terms, provided the original work is properly cited and the use is non-commercial. See: <http://creativecommons.org/licenses/by-nc/4.0/>

Manuscript source: Invited manuscript

Specialty type: Endocrinology and metabolism

Country/Territory of origin: United States

Peer-review report's scientific quality classification

Grade A (Excellent): 0

Grade B (Very good): 0

Grade C (Good): C

Grade D (Fair): 0

Grade E (Poor): 0

Received: January 26, 2021

Peer-review started: January 26, 2021

First decision: May 3, 2021

Revised: May 10, 2021

Accepted: August 5, 2021

Article in press: August 5, 2021

Published online: September 15, 2021

P-Reviewer: Yang L

S-Editor: Liu M

L-Editor: A

P-Editor: Guo X



cell function

©The Author(s) 2021. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Vitamin D deficiency is a well-recognized health issue and contributes to bone loss and calcium dysregulation. Evidence suggests that excess vitamin D is not in and of itself of therapeutic benefit. Available clinical data suggests that vitamin D supplementation appears to limit the development of diabetes in vitamin D deficient subjects. However, no benefit was observed in non-vitamin D deficient subjects. Furthermore, overreplacement of vitamin D is of no beneficial effect and could possibly be harmful.

Citation: Chang Villacreses MM, Karnchanasorn R, Panjawatanan P, Ou HY, Chiu KC. Conundrum of vitamin D on glucose and fuel homeostasis. *World J Diabetes* 2021; 12(9): 1363-1385

URL: <https://www.wjgnet.com/1948-9358/full/v12/i9/1363.htm>

DOI: <https://dx.doi.org/10.4239/wjd.v12.i9.1363>

INTRODUCTION

The potential role of vitamin D deficiency induced by migration of human beings has been suggested to be involved in human evolution and various modern health conditions[1]. The history prospective of vitamin D evaluation will enhance our understanding of the development in this field. The role of dietary deficiency in the pathogenesis of rickets was established by Platt[2] in 1919. Although it was thought to be caused by vitamin A deficiency initially, McCollum *et al*[3] identified a vitamin deficiency other than vitamin A that caused rickets in 1922. Since vitamin A, B, and C were already identified, the new molecule was named as vitamin D[4].

Beginning with its discovery in 1922, scientific publications focusing upon vitamin D numbered no more than some 10 per year but this increased to 35 per year by 1945 (Figure 1). As knowledge of the structure, molecular biology and function of vitamin D increased[5,6], there was a concurrent increase in vitamin D-specific publications. With the observations of the non-calcemic effects of vitamin D[7], vitamin D-focused publications peaked at 5152 in 2017. Recently, the role of vitamin D deficiency in relation to coronavirus disease 2019 (COVID-19) infection attracted attention[8].

Vitamin D on bone health

The role of vitamin D on calcium and bone metabolism was well-summarized[9]. There is no doubt about the association between rickets and vitamin D deficiency and the reversal and prevention of rickets with vitamin D supplementation. However, controversy still surrounds the efficacy of vitamin D supplementation upon bone mineral density and fracture prevention. Multiple studies failed to demonstrate any benefit from vitamin D supplementation[10-12] and a systematic review and meta-analysis also failed to confirm any beneficial effect on bone density or fracture prevention from vitamin D supplement[13]. Nevertheless, placebo-control randomized clinical trials revealed a threshold effect of vitamin D[14,15] with no benefit observed on the subjects with baseline 25-hydroxyvitamin D level ≥ 75 nmol/L (30 ng/mL). Furthermore, possible detrimental effects on bone mineral density were observed in subjects who received a higher dose of vitamin D (250 μ g or 10000 IU daily) with a mean 25-hydroxyvitamin D of 200 nmol/L or 80 ng/mL[12]. While not conclusive, these data suggest that the optimal effects of vitamin D are found at a 25-hydroxyvitamin D level of 75 nmol/L (30 ng/mL).

Vitamin D as a hormone

Vitamins are defined as micronutrients that cannot be self-synthesized and that necessary for the proper function of key enzymatic processes. Consequently, vitamins must be obtained through the diet. Vitamin D is synthesized from cholesterol to 7-dehydrocholesterol, also known as pro-vitamin D₃, in the skin through the action of ultraviolet radiation[16]. In addition, the liver forms 25-hydroxyvitamin D₃, also

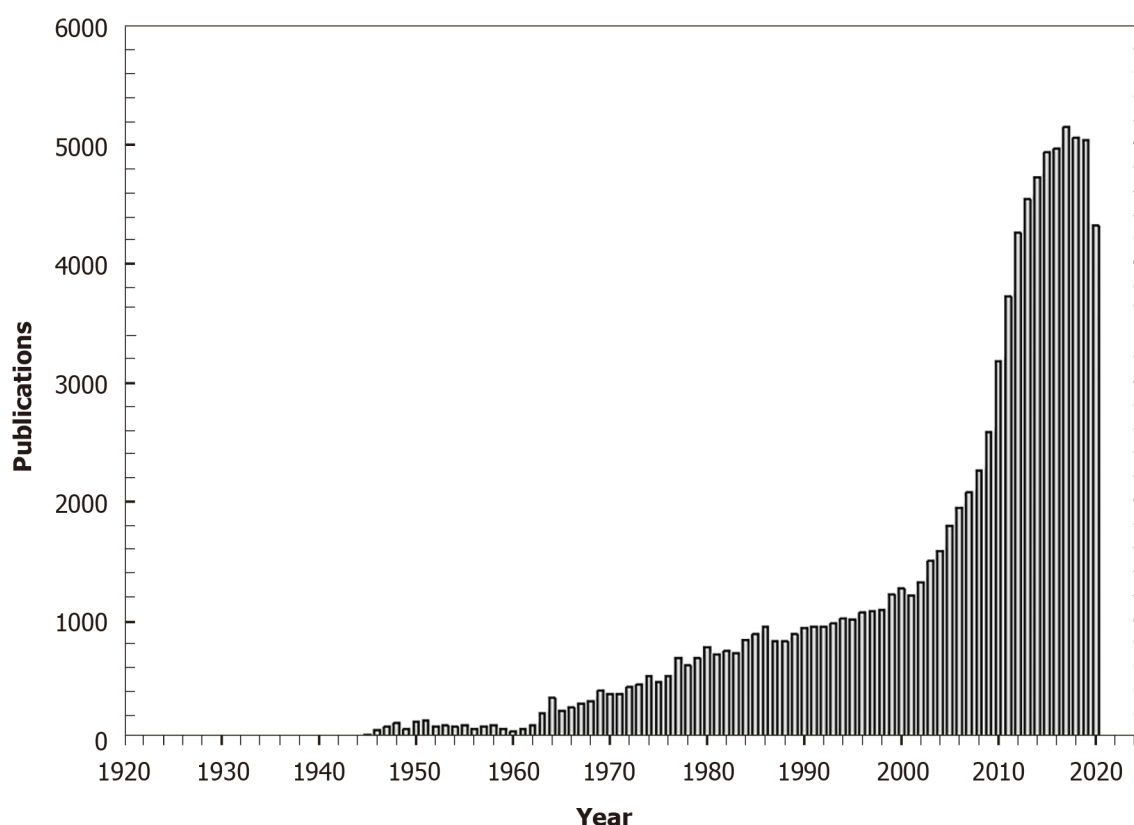


Figure 1 Vitamin D publications from 1922 to 2020. Data were obtained from PubMed (<https://pubmed.ncbi.nlm.nih.gov/>) accessed on October 20, 2020.

known as pre-vitamin D₃. To become an active compound, further hydroxylation in the kidney is required to form 1,25-dihydroxyvitamin D₃, which is a biologically active vitamin D. Then, 1,25-dihydroxyvitamin D is released into circulation to exert its effects on the target cells and promote calcium and bone homeostasis. Thus, vitamin D is a hormone and, like the pituitary-thyroid axis, has a complex natural history in the body (Table 1).

The half-life of thyroid hormone depends upon thyroid status[17]. The half-life for levothyroxine (T4) is 6-7 d in euthyroid subjects, 9-10 d in subjects with hypothyroidism, and 3-4 d in subjects with hyperthyroidism. The half-life of liothyronine (T3) is 18-24 h in euthyroid subjects, 12-16 h in hyperthyroid subjects, and 26-32 h in hypothyroid subjects. The half-life of vitamin D averages 15 h but depends upon of the type of vitamin D (cholecalciferol or vitamin D₃ vs ergocalciferol or vitamin D₂) and vitamin D binding protein concentration[18]. The half-life of 1,25-dihydroxyvitamin D is 10-20 h[19], while there is no information regarding the half-life of 1,25-dihydroxyvitamin D₃ vs D₂. Since 1,25-dihydroxyvitamin D is released into the blood and exerts its effects upon osteocytes to promote mineralization and on the gastrointestinal epithelium to increase calcium and phosphorus absorption, it is appropriate to classify vitamin D as a hormone.

EVIDENCE OF NON-CALCEMIC EFFECTS

In addition to the target organs, both the vitamin D receptor and 1 α -hydroxylase (CYP27B1) are expressed in various other tissues[20], suggesting additional functions of vitamin D beyond bone metabolism and calcium homeostasis. Interestingly, the vitamin D receptor is expressed in the pancreatic islets[21], liver[22], muscle[23], and adipose tissue[24]. 1 α -hydroxylase (CYP27B1) is expressed in pancreatic islets[25], liver[26], muscle[27], and adipose tissue[28]. Thus, it is possible that vitamin D could take part in glucose and fuel homeostasis.

In contrast to calcemic effects of vitamin D which is primary mediated by circulating 1,25-dihydroxyvitamin D produced in the kidney, the non-calcemic effects of vitamin D are mediated by circulating 25-hydroxyvitamin D through a paracrine or autocrine function[29]. Within the target cells or its vicinity, circulatory 25-hydroxyvitamin D

Table 1 Vitamin D as a hormone: Comparison of the pituitary-thyroid and parathyroid hormone-vitamin D axes

	Pituitary-thyroid axis	Parathyroid-vitamin D axis
Organ(s)	Thyroid glands	Skin/liver/kidney
Source compound	Iodine, tyrosine	Cholecalciferol (cholesterol), ergocalciferol
Prehormone	Levothyroxine, $T_{1/2} = 6-7$ d	25-hydroxyvitamin D ₂ /D ₃ , $T_{1/2} = 13-17$ d
Active hormone	Triiodothyronine, $T_{1/2} = 14-24$ h	1,25-dihydroxyvitamin D ₂ /D ₃ , $T_{1/2} = 10-20$ h
Transportation	Thyroxine binding globulin	Vitamin D binding protein
Receptor	Thyroid hormone receptor	Vitamin D receptor
Stimulating factor	Thyroid stimulating hormone	Parathyroid hormone
Effect	Energy homeostasis	Calcium homeostasis

enters cells and is converted to 1,25-dihydroxyvitamin D by the locally existing 1 α -hydroxylase (CYP27B1). Hence, 25-hydroxyvitamin D is the key circulatory element for the non-calcemic effects of vitamin D whereas 1,25-dihydroxyvitamin D promotes the calcemic effects.

EFFECTS UPON CELL DIFFERENTIATION AND CELL PROLIFERATION

Colon, prostate, breast, and ovarian cancer

A role for vitamin D in the pathogenesis of cancer was proposed in 1980[30] after it was observed that colon cancer rates were higher in the northern rather than the southern United States. The association of vitamin D deficiency with cancer, including breast[31], prostate[32], and colon cancer[33] was attributed to the ability of vitamin D to differentiation cells[34] and to suppress cell proliferative[35] along with other effects [36,37].

Immunity, autoimmunity, and inflammation

The risk of type 1 diabetes was reduced by vitamin D supplement in a birth-cohort study from Finland[38]. Furthermore, a polymorphism in the vitamin D receptor was associated with increased risk of type 1 diabetes[39]. Not unexpectedly, a role of vitamin D deficiency in the pathogenesis of type 1 diabetes was proposed[40]. In addition, the association of vitamin D deficiency with multiple sclerosis[41], systemic lupus erythematosus[42], and other autoimmune diseases[43] was attributed to the immunomodulatory and anti-inflammatory effects of vitamin D[44]. Furthermore, vitamin D plays an important role in the maintenance of B cell homeostasis[45], and vitamin D replacement may reduce B cell-mediated autoimmune disorders.

The role of vitamin D in the treatment of tuberculosis was appreciated with the observation that sun exposure altered the clinical presentation of tuberculosis[46]. Subsequently, vitamin D was administered as part of the treatment of tuberculosis [47]. Vitamin D deficiency was frequently observed in patient with untreated tuberculosis[48]. It is now known that Toll-like receptors up-regulate expression of the vitamin D receptor and the vitamin D-1-hydroxylase genes, leading to induction of the antimicrobial peptide cathelicidin and killing of intracellular Mycobacterium tuberculosis[49]. Thus, the role of vitamin D in fighting infection is established[50]. Further, vitamin D deficiency is associated with acute respiratory tract infection[51], bacterial vaginosis[52], pneumonia[53], foot infection in diabetics[54], chronic hepatitis C infection[55], and human immunodeficiency virus infection[56]. Recently, vitamin D deficiency was recognized as a risk factors for COVID-19 infection[57-61]. Thus, vitamin D could play a role in fighting infection.

An association between vitamin D receptor polymorphism and the severity of coronary artery disease was reported[62]. Deficiency was also noted to associate with an increased risk of myocardial infraction[63], hypertension[64], and stroke[65]. The mechanism proposed to account for these associations included activation of the renin-angiotensin system[66], coronary calcification[67], platelet activation and aggregation [68], increased proinflammatory cytokines[69], and vascular endothelial dysfunction [65].

Fuel metabolism

In patients with vitamin D deficiency and diabetes, vitamin D supplementation improved beta cell function and glucose tolerance[70]. An association between vitamin D deficiency and glucose intolerance and beta cell dysfunction was observed in east London Asians[71]. Similarly, alternations in vitamin D metabolism in obese subjects manifesting as low 25-hydroxyvitamin D is well-recognized[72]. This topic will be reviewed in this article.

Neuropsychiatric disorders

Vitamin D deficiency was reported to be associated with depression[73], schizophrenia [74], autism[75], and Parkinson's disease[76]. Various mechanisms have been reported to support a role of vitamin D in neuropsychiatric disorders. Vitamin D has a protective effect on dopaminergic neurons[77]. Vitamin D deficiency could result in altered synaptic plasticity through its effect on perineuronal nets leading to cognitive deficits[78]. Vitamin D deficiency alters brain protein expression in rats[79]. Furthermore, immunohistochemical study revealed the expression of vitamin D receptor and 1 α -hydroxylase (CYP27B1) in various regions of human brain with the strong expression in the hypothalamus and in the large (presumably dopaminergic) neurons within the substantia nigra[80]. Thus, vitamin D deficiency could play a role in the pathogenesis of various neuropsychiatric disorders.

VITAMIN D REPLACEMENT THERAPY

Source of vitamin D

Vitamin D is available in two forms: ergocalciferol (vitamin D₂) and cholecalciferol (vitamin D₃). Ergocalciferol comes from plants in the form of ergosterol (provitamin D₂). Ergosterol is an important component of mushrooms. Through ultraviolet b (UVB) irradiation, which can occur within mushroom or artificially, it becomes ergocalciferol [81]. Cholecalciferol comes from animals and people through the biosynthesis of cholesterol to 7-dehydrocholesterol (Provitamin D₃). Again, through UVB irradiation, this intermediate becomes cholecalciferol. Thus, dietary intake and sun exposure are the major determinants of serum 25-hydroxyvitamin D levels.

Sun, mainly UVB irradiation, plays an important role in biosynthesis of vitamin D. Since 7-dehydrocholesterol can be synthesized from cholesterol, theoretically vitamin D supplementation is not required once sun exposure is adequate. Skin color is a key determinant of vitamin D synthesis[82]. Vitamin D has been proposed to play a role in human evolution and migration away from equator by affecting skin color through the development of depigmented and tannable skin *via* genetic pathways under positive selection[1,83]. Sun exposure is highly effective in raising serum 25-hydroxyvitamin D concentration, while its effects diminish significantly on donning clothing and using sun screen[84]. In this regard, more body surface area exposure is more effective than longer exposure time[85]. However, the efficacy of sun exposure to increase serum 25-hydroxyvitamin D concentrations diminishes with the degree of skin tanning[86]. Thus, minimized sun exposure time for 5 min to 30 min (depending on time of day, season, latitude, and skin pigmentation) with maximize body surface exposure is recommended[9]. However, increased risk of sun-mediated skin cancer makes this approach to prevent vitamin D deficiency less optimum[87].

Vitamin D can be obtained through dietary intake. However, except for cod liver oil, vitamin D content in naturally occurring food is relatively low, even in mushrooms (Table 2). Although ergosterol is highly abundant in the membrane of mushrooms, mushroom are cultivated under shadow without UVB irradiation[81]. Thus, dietary intake of vitamin D is inadequate and vitamin D supplement is often needed to avoid deficiency.

Comparison of metabolism of vitamin D₂ vs vitamin D₃

It is estimated that 65% of vitamin D is present as vitamin D while 35% is in the form of 25-hydroxyvitamin D. As well, almost 75% of vitamin D is in adipose tissue, while 25-hydroxyvitamin D is distributed 20% in muscle, 30% in serum, 35% in fat, and 15% in other tissues[88]. The metabolism of vitamin D₃ and vitamin D₂ is summarized in Table 3. Vitamin D binding protein transports the various forms of vitamin D in circulation, including vitamin D, 25-hydroxyvitamin D, and 1,25-dihydroxyvitamin D [89]. Each vitamin D binding protein molecule has one binding site for vitamin D and/or its metabolites. The relative affinity of vitamin D binding protein to vitamin D₃

Table 2 Vitamin D content of selected foods

Food	Per serving		Percent DV
	IU	µg	
Cod liver oil, 1 tablespoon	1360	34.00	170
Trout (rainbow), farmed, cooked, 3 ounces	645	16.13	81
Salmon (sockeye), cooked, 3 ounces	570	14.25	71
Mushrooms, white, raw, sliced, exposed to UV light, 1/2 cup	366	9.15	46
Milk, 2% milkfat, vitamin D fortified, 1 cup	120	3.00	15
Soy, almond, and oat milks, vitamin D fortified, various brands, 1 cup	100-144	2.50-3.60	13-18
Ready-to-eat cereal, fortified with 10% of the DV for vitamin D, 1 serving	80	2.00	10
Sardines (Atlantic), canned in oil, drained, 2 sardines	46	1.15	6
Egg, 1 large, scrambled (Vitamin D is in the yolk)	44	1.10	6
Liver, beef, braised, 3 ounces	42	1.05	5
Tuna fish (light), canned in water, drained, 3 ounces	40	1.00	5
Cheese, cheddar, 1 ounce	12	0.30	2
Mushrooms, portabella, raw, diced, ½ cup	4	0.10	1
Chicken breast, roasted, 3 ounces	4	0.10	1
Beef, ground, 90% lean, broiled, 3 ounces	1.7	0.04	0

Adapted from: <https://ods.od.nih.gov/factsheets/VitaminD-HealthProfessional/#en25>. The Food and Drug Administration developed daily values (DVs) to help consumers compare the nutrient contents of foods and dietary supplements within the context of a total diet. The DV for vitamin D on the new Nutrition Facts and Supplement Facts labels used for the values in Table 2 is 20 µg (800 IU) for adults and children aged 4 years and older. Foods providing 20% or more of the DV are considered to be high sources of a nutrient, but foods providing lower percentages of the DV also contribute to a healthful diet. DV: Daily value.

Table 3 Comparison of transportation and metabolism of vitamin D₃ vs D₂

Ref.	Symbol	Name (chromosome location)	Function	D3/D2
Haddad <i>et al</i> [90], 1993	VBP	Vitamin D binding protein (4q12-q13)	Vitamin D transportation	1.14
Holmberg <i>et al</i> [91], 1986	CYP2R1	25-hydroxylase (11p15.2)	Conversion of vitamin D to 25-hydroxy vitamin D	5.0
Zarei <i>et al</i> [93], 2016	CYP27B1	1alpha-hydroxylase (12q13.1-q13.3)	Conversion of 25(OH)D to 1,25(OH)2D	2.4
Jones <i>et al</i> [94], 1980	VDR	Vitamin D receptor (7q36)	Receptor for vitamin D	1.3

is 1.14 times stronger than to vitamin D₂[90]. 25-hydroxylase (CYP2R1) catalyzes 25-hydroxylation of vitamin D₃ 5 times more efficiently than vitamin D₂[91]. Thus, after administration of a single oral dose of vitamin D₃ and vitamin D₂, a more sustainable and prolonged increase in serum 25-hydroxyvitamin D₃ concentration is observed compared to serum 25-hydroxyvitamin D₂ concentration[92]. 1alpha-hydroxylase (CYP27B1) converts 25-hydroxyvitamin D₃ to 1,25-dihydroxyvitamin D₃ 2.4-time more efficiently than 25-hydroxyvitamin D₂[93]. In receptor binding assays, 1,25-dihydroxyvitamin D₃ has 1.3 times more receptor affinity than 1,25-dihydroxyvitamin D₂[94]. These data indicate that vitamin D₃ is more biologically potent than vitamin D₂.

Comparison of biological potency of vitamin D₂ vs vitamin D₃

Vitamin D₂ and vitamin D₃ were reported to have similar efficacy in raising serum 25-hydroxyvitamin D concentration[95]. However, other studies demonstrated that vitamin D₃ was more efficacious at raising serum 25(OH)D concentrations than vitamin D₂[96-100]. This finding was confirmed by a meta-analysis of the randomized

control trials[101]. Furthermore, 25-hydroxyvitamin D₃ has a longer half-life compared to 25-hydroxyvitamin D₂ (15.1 ± 3.1 d *vs* 13.9 ± 2.6 d, $P = 0.001$, mean \pm STD)[18]. In comparison to oral vitamin D₂, oral vitamin D₃ achieves a higher serum concentration of 1,25-dihydroxyvitamin D[100,102] and a more effective suppression of serum parathyroid hormone concentration[97]. Physicians preferring use of vitamin D₂ should be aware of its markedly lower potency and shorter duration of action when compared to vitamin D₃. Thus, vitamin D₃ is the preferred form of vitamin D for replacement therapy.

OPTIMAL SERUM 25-HYDROXYVITAMIN D CONCENTRATION

Minimal serum 25-hydroxyvitamin D concentration

The primary function of vitamin D is to maintain calcium homeostasis. The minimal serum 25-hydroxyvitamin D concentration for health was defined based on the serum parathyroid hormone response to replacement therapy with ergocalciferol[103]. A serum 25-hydroxyvitamin D concentration of 50 nmol/L (20 ng/mL) was recommended since no further changes in serum parathyroid hormone levels were found in subjects with a serum 25-hydroxyvitamin D level of 50 nmol/L (≥ 20 ng/mL). In 2010, the United States Institute of Medicine adapted this value as a target for ensuring good bone health[104]. However, based on a larger observational study with 1569 subjects in France, serum parathyroid hormone concentration were noted to still decrease when the serum 25-hydroxyvitamin D rose to 78 nmol/L (31 ng/mL)[105]. Furthermore, a serum 25-hydroxyvitamin level of 75 nmol/L (30 ng/mL) is a recognized threshold for intestinal calcium absorption[106]. As shown in Table 4, many professional organizations and agencies have since adapted 75 nmol/L (30 ng/mL) as the minimal acceptable serum 25-hydroxyvitamin D concentration recognizing this may have beneficial effects beyond bone health, targeting beyond bone health while the Institute of Medicine define the minimal 25-hydroxyvitamin D concentration 50 nmol/L (20 ng/mL) on bone health with a public health interest.

Maximal serum 25-hydroxyvitamin D concentration

The maximal allowed serum 25-hydroxyvitamin D concentration is defined by the appearance of adverse effects. Although the Institute of Medicine does not define maximal serum 25-hydroxyvitamin D concentration[104], a warning against elevated serum 25-hydroxyvitamin D concentrations is stated. This warning is based upon the observed association of increasing mortality with serum 25-hydroxyvitamin D concentration > 125 nmol/L (50 ng/mL)[107] by limiting the maximal daily vitamin D allowance (Table 4). This notion was further supported by the finding of increased cardiovascular mortality with serum 25-hydroxyvitamin D > 125 nmol/L (50 ng/mL)[108]. In addition, a progressive decline in bone mineral density with serum 25-hydroxyvitamin D greater than 125 nmol/L (50 ng/mL) was observed in a United States population[109]. Conversely, bone mineral density improved after discontinuation of vitamin D supplementation in patients with a serum 25-hydroxyvitamin D concentration greater than 50 ng/mL[110]. Although vitamin D supplementation increased calcium absorption without a threshold effect[111], reanalysis of the data revealed a diminished response (per 1000 IU of vitamin D in Table 5) with increasing dose of vitamin D supplement suggesting a threshold effect of vitamin D on calcium absorption[112], something noted by others[106]. We reported lack of improvement in insulin sensitivity in individuals with a serum 25-hydroxyvitamin D concentration > 125 nmol/L (50 ng/mL)[113]. Although hypercalcemia from vitamin D intoxication occurs mainly when the serum 25-hydroxyvitamin D concentration is > 374 nmol/L (150 ng/mL)[114], serum 25-hydroxyvitamin D concentrations > 75 nmol/L (50 ng/mL) could be either harmful or lack beneficial effect.

Comparison of daily replacement vs intermittent replacement of vitamin D

The observation that a single oral dose of vitamin D₃ 2.5 mg (100000 IU) can maintain serum 25-hydroxyvitamin D above the target goal[115] provides a unique dosing strategy of vitamin D replacement therapy with greater adherence. It could even ensure 100% compliance if given by or under the direct supervision of a health care provider. Weekly[103], monthly[116], biyearly[117], and even yearly[118] schedules were reported in various trials leading to initiation of more convenient dosing schedule at less frequent intervals in clinical practice. To reduce the dosing frequency, a much higher dose of vitamin D is required which is predicted to cause a short-term spike (> 75 nmol/L or 50 ng/mL) in serum 25-hydroxyvitamin D concentration shortly

Table 4 Recommended daily vitamin D intake as promulgated by selected organizations and agencies

Organization	Daily intake		Goal	
	IU	µg	ng/mL	nmol/L
Institute of Medicine	600-800	15-20	> 20 (20-50)	> 50 (50-125)
Agency of Healthcare Research and Quality, Department of Health and Human Services	> 1000	> 25	> 30	> 75
Office of Dietary Supplements, NIH	600-800	15-20	20-50	50-125
National Osteoporosis Foundation	800-1000	20-25	> 30	> 75
American Association of Clinical Endocrinologists	1000-2000	25-50	30-60	75-150
Endocrine Society	1500-2000	37.5-50	30-100	75-250

Table 5 Diminished response of intestinal calcium absorption in response to increasing vitamin D supplementation

Daily vitamin D supplementation		Observed increase in calcium absorption	Estimated increase in calcium absorption per 1000 IU (25 µg)
IU	µg		
800	20	3.90%	4.88%
2000	50	5.00%	2.50%
4000	100	6.70%	1.68%

after oral administration. In addition to the adverse effects as described in the above section 4.2, increased falls and fracture are observed with annual vitamin D replacement therapy. These mainly occur within the first 3 mo after oral administration of 12.5 mg vitamin D₃[118]. Furthermore, the associations of high-dose vitamin D treatment with gastrointestinal complaints[119], increased bone turnover markers [120], hypercalcemia[121], hypercalciuria[122], and increased urinary magnesium loss [123] have been reported. Similar levels of serum 25-hydroxyvitamin D concentration were achieved at the end of a 56-d trial from daily (1500 IU/d), weekly (10500 IU/wk), and monthly (45000 IU/4 wk) replacement therapy. Excessive serum 25-hydroxyvitamin D concentration was not observed in those on the daily regimen but was observed in individuals on the weekly regimen and was still more common in those on monthly regimen[124]. Thus, high-dose vitamin D replacement therapy results in excessive serum 25-hydroxyvitamin D concentration.

A Lysine (K) amino acid polymorphism, in replacement of Threonine (T), at position 436 of vitamin D binding protein is associated with increased affinity of vitamin D and is associated a 416% elevation in serum 25-hydroxyvitamin D concentration if high-dose (4000 IU) vitamin D₃ replacement therapy is given as opposed to low-dose (600 IU) vitamin D₃ replacement therapy. Individuals carrying the TT SNP showed only a 136% increase in circulating vitamin[125]. Since the K allele is a minor allele and KK genotype accounts for less than few percent of population, the KK subjects may account for the excessive serum 25-hydroxyvitamin D-associated complications noted in certain studies. Given the above, daily vitamin D supplementation would seem to be most physiological and safest way to correct vitamin D deficiency and avoid the possible adverse effects associated with the excessive serum 25-hydroxyvitamin D concentration.

Factors affecting serum 25-hydroxyvitamin D concentration

Various genetic loci are associated with serum 25-hydroxyvitamin D concentration [126] with 4 major loci identified (Table 6). These are all key proteins involved in the transportation and metabolism of vitamin D. Race and ethnicity were noted to have significant impact on serum 25-dihydroxyvitamin D concentration[127], again implicating a genetic influence[126] including skin color[128].

Seasonable variations in serum 25-hydroxyvitamin D concentrations related to sun exposure are well described[126]. Consistent with this, latitude has a significant impact on serum 25-dihydroxyvitamin D concentration[129]. Living closer to the equator and increasing sun exposure can improve vitamin D levels. However, the increased risk of skin cancer from sun exposure should be balanced employing maximum skin

Table 6 Major loci associated with changes in serum 25-hydroxyvitamin D concentration

Chromosome	SNP	Gene symbol	Protein	P value
4p12	rs2282679	GC	Vitamin D binding protein	1.9×10^{-109}
11q12	rs12785878	DHCR7	7-dehydrocholesterol reductase	2.1×10^{-27}
11p15	rs10741657	CYP2R1	1-alpha-hydroxylase	3.3×10^{-20}
20q13	rs6013897	CYP24A1	1,25-dihydroxyvitamin D3 24-hydroxylase	6.0×10^{-10}

Adapted from Wang *et al*[126]. SNP: Single nucleotide polymorphism.

exposure area with decreased exposure time[85]. Dietary supplementation also corrects deficiency. Obesity is associated with a lower serum 25-hydroxyvitamin D concentration[72] while weight reduction with loss of adipose tissue is associated with improvement in serum 25-hydroxyvitamin D concentration[130]. These findings indicate that vitamin D status may be improved through modification of lifestyle.

Practical recommendations for vitamin D replacement therapy

As showed in Table 4, the recommended vitamin D supplement varies between organizations and agencies. The reasons for this relate to the purpose of vitamin D supplementation, visive calcemic *vs* non-calcemic effects. For calcemic effects, bone health is the goal of supplementation and is maximized through using a conservative daily vitamin D to achieve the minimal serum 25-hydroxyvitamin D concentration while avoiding possible adverse effects associated with overreplacement. A public health approach to this is displayed in Table 7. In contrast, a more personized approach is rationale when the target is to promote the non-calcemic effects of vitamin D.

We recommend using vitamin D₃ instead of vitamin D₂ for the rationale as discussed in the sections 3.2 and 3.3. We are in favor of daily replacement therapy and against intermittent mega dose replacement. This is supported by the recommendations of the Endocrine Society for indefinitely intermittent mega dose replacement [131]. It has been estimated that supplement with cholecalciferol 1000 IU (50 µg) daily will increase serum 25-hydroxyvitamin D concentration by 10 ng/mL[132]. Since vitamin D is a fat soluble, replacement therapy can be further enhanced by taking it with the largest meal of day[133]. We recommend vitamin D₃ 1000 IU daily for achievement of an initial serum 25-hydroxyvitamin D concentration between 51 nmol/L (21 ng/mL and 75 nmol/L (30 ng/mL); 2000 IU daily for between 26 nmol/L (11 ng/mL) and 50 nmol/L (20 ng/mL); and 5000 IU for equal or less than 25 nmol/L (10 ng/mL). Serum 25-hydroxyvitamin concentration should be measured within 3 mo for assessment and, if indicated, dose adjustment. We are targeting serum 25-hydroxyvitamin D concentration between 75 nmol/L (30 ng/mL) and 125 nmol/L (50 ng/mL).

VITAMIN D AND DIABETES PREVENTION

Vitamin D diabetes prevention trials

To date, eight clinical trials employed vitamin D to reduce prediabetes progression to overt diabetes (Table 8). Only two studies[134,135] demonstrated positive results. Although these two studies had small sample size, they recruited true vitamin D deficient (25-hydroxyvitamin D < 50 nmol/L or 20 ng/mL) subjects and achieved final 25-hydroxyvitamin D concentration at 89-90 nmol/L, after intervention for 1 year and 6 mo, respectively. Of note, the study in India[134] was a randomized open label study demonstrating an odds ratio of 0.31 [95% confidence intervals (CI): 0.11-0.90]. The study in Iran was a randomized placebo control study[135] revealing an odds ratio of 0.06 (95% CI: 0.01-0.51). Because of relatively small sample sizes of both studies, the CI were very wide. Additional studies with similar initial and final 25-hydroxyvitamin D concentration (< 50 nmol/L and 90-100 nmol/L, respectively) and much larger sample sizes are required to confirm these data.

Two negative studies[136,137] were noted to have similar initial 25-hydroxyvitamin D concentrations (25-42 nmol/L). The negative results could be due to the relatively short interventions (8-16 wk) and small sample sizes. The study in Holland only

Table 7 Vitamin D supplementation versus vitamin D replacement therapy

	Vitamin D supplement	Vitamin D replacement therapy
Target goal	Bone health	Beyond bone health
Target 25-hydroxyvitamin D level	> 20 ng/mL (50 nmol/L)	> 30 ng/mL (75 nmol/L)
Initial testing for 25-hydroxyvitamin D level	No	Yes
Concern of over-replacement	Yes	Yes
Follow-up testing for 25-hydroxyvitamin D level	No	Yes
Dose adjustment	No	Yes
Approach	Public health	Individualized

Table 8 Preventive trials of vitamin D supplementation to prevent the development of type 2 diabetes

Ref.	Country Race/ethnicity	n	Placebo control		n	Intervention		Dose	Frequency	Duration	Diabetes prevention
			25(OH)D nmol/L			25(OH)D nmol/L					
			Initial	Final		Initial	Final				
Dutta <i>et al</i> [134], 2014 ¹	IndiaAsian Indian	49	45	44	55	43	89	1500 µg	Weekly X 8, monthly	1 yr	Positive ²
Niroomand <i>et al</i> [135], 2019	IranIranian	83	32	40	83	31	90	1250 µg	Weekly for 3 mo, monthly	6 mo	Positive ³
Wagner <i>et al</i> [136], 2016 ⁴	Sweden	22	47	46	21	42	83	750 µg	weekly	8 wk	Negative
Oosterwerff <i>et al</i> [137], 2014	HollandNon-Western	65	22	23	65	25	60	30 µg	daily	16 wk	Negative
Barengolts <i>et al</i> [141], 2015 ⁵	United States African American	86	35	50	87	37	120	1250 µg	weekly	12 m	Negative
Davidson <i>et al</i> [139], 2013 ⁶	United States Latino and African American	53	55	60	56	55	167	2222 µg	weekly	12 mo	Negative
Jorde <i>et al</i> [140], 2016	Norway	255	61	64	256	60	110	500 µg	weekly	5 yr	Negative
Pittas <i>et al</i> [138], 2019	United States mixed	1212	70	72	1211	69	136	100 µg	daily	24 mo	Negative

¹This study was an open label randomized design, instead of randomized placebo-control design as other studies.

²Intervention is associated with significantly lower progression to diabetes (11% *vs* 27%; $P = 0.04$) and higher reversal to normoglycemia (43% *vs* 20%; $P = 0.02$).

³The rate of progression toward diabetes was significantly lower in the intervention group (3% *vs* 28%; $P = 0.002$).

⁴Median 25-hydroxyvitamin D was provided, rather than mean 25-hydroxyvitamin D as in other studies.

⁵Ergocalciferol was used, rather than cholecalciferol in other studies.

⁶Weekly dose of cholecalciferol was adjusted to titrate serum 25-hydroxyvitamin D between 162 nmol/L and 200 nmol/L.

achieved a final suboptimal 25-hydroxyvitamin D concentration of 60 nmol/L.

The other four studies[138-141] had a final 25-hydroxyvitamin D concentration > 100 nmol/L which might not be optimal for glucose metabolism. Among them, the study in African American[141] was the only study that recruited true vitamin D deficient subjects (initial 25-hydroxyvitamin D 37 nmol/L). Of note, ergocalciferol was used which could be less effective biologically as discussed above in 3.2 and 3.3. Enrollment of non-vitamin D deficient (25-hydroxyvitamin D < 50 nmol/L) subjects [138-140] could further reduce the chance of finding any effect. Furthermore, the study in Norway had a significant dropout rate in the interventional group with only 45% of participants completing the planned 5-year visit. The largest intervention trial[138] included more than 1000 subjects in each group. To be able to apply to the general population in the United States, this study did not target vitamin D deficient subjects and allowed the participants to take additional vitamin D up to 25 µg daily. Therefore, it had the highest initial 25-hydroxyvitamin D among these studies, 70 nmol/L in the

control group and 69 nmol/L in the interventional group, which might diminish the power of this study to detect the beneficial effect of vitamin D. Regardless of the negative results in most studies, the beneficial effect of vitamin D supplementation cannot be completely excluded, especially in subjects with vitamin D deficiency (25-hydroxyvitamin D < 50 nmol/L).

The effects of vitamin D supplement on parameters of glucose metabolism

Various parameters of glucose metabolism were reported in most of above-mentioned studies, except one[138]. After vitamin D intervention for 1 year, the study from India [134] observed improvement in fasting and 2-hr post-challenge glucose concentrations, insulin sensitivity by Homeostasis Model (HOMA) insulin resistance index, QUICKI, and 1/fasting insulin concentration while no impact on HbA1c and beta cell function by HOMA. Following vitamin D supplement for 6 mo, the study from Iran[135] reported the improvement in the HOMA insulin resistance index and marginal improvement in fasting insulin concentration ($P = 0.05$) and 2-hour post-challenge blood glucose concentration ($P = 0.07$) with no impact on fasting blood glucose concentration.

After an 8-wk intervention, the study from Sweden[136] assessed insulin sensitivity and beta cell function using the hyperglycemic clamp. They observed a significant improvement in disposition index based on the first phase insulin response ($P = 0.005$) and marginal improvement in first phase insulin response ($P = 0.06$), insulin sensitive index ($P = 0.09$), disposition index based on the second phase insulin response ($P = 0.06$), and A1c ($P = 0.06$) but no impact on the second phase insulin response and fasting and 2-hr post-challenge blood glucose concentration.

In contrast, the study from Holland[137] evaluated glucose metabolism parameters based on the 75-g glucose tolerance test following intervention for 16 wk. They reported negative results, finding no effects upon insulin area under curve, glucose area under curve, insulin sensitivity by composite insulin sensitivity index, Stumvoll index, insulin resistance index by HOMA, and beta cell function by insulinogenic index. Of note, the final 25-hydroxyvitamin D concentration was only 60 nmol/L which could be suboptimal for glucose metabolism. Similarly, after the vitamin D supplementation for 5 years, the study from Norway[140] observed no impact on fasting and 2-hr post-challenge serum glucose concentration, fasting and post challenge serum insulin concentration, fasting serum C-peptide concentration, HbA1c, and insulin sensitivity by HOMA insulin resistance index and QUICKI.

Following a 12-mo intervention, the study involving Latino and African Americans [139] observed a significant improvement in HbA1c but no effects on fasting and 2-hr post-challenge blood glucose concentration, beta cell function by the ratio of insulin and glucose area under curve, Stumvoll first and second insulin response, insulinogenic index, insulin sensitivity index by HOMA insulin resistance index and composite insulin sensitivity index, and oral disposition index. However, a significant improvement in composite insulin sensitivity index but not Matsuda index, insulinogenic index, C-peptidogenic index, and HbA1c was noted.

Excepting two studies[137,140] with negative results, favorable outcomes on parameters of glucose metabolism were reported in five studies[134-136,139,141] suggesting some benefits to supplementation under these conditions.

Summary of vitamin D and diabetes prevention

In vitamin D deficient (25-hydroxyvitamin D < 50 nmol/L) prediabetic subjects, vitamin D supplement appears to be effective in reduction of the development of overt diabetes. However, there appears to be no benefit in vitamin D sufficient subjects, which was noted in a study from Norway[142]. Based on pooled data from four intervention trials, in subjects without vitamin D deficiency there is no improvement in glucose metabolism with high dose vitamin D supplementation and if anything, the effect is negative[143]. This notion is consistent with the observed threshold effect of vitamin D on bone health and lack of benefit in subjects with baseline 25-hydroxyvitamin D level ≥ 75 nmol/L (30 ng/mL)[14,15].

LABORATORY EVIDENCE SUPPORTING THE EFFECT OF VITAMIN D ON GLUCOSE AND FUEL HOMEOSTASIS

Beta cell function

Functional beta cell studies: The important role of vitamin D on insulin secretion has

been noted in laboratory animals since 1980. Insulin secretion was reduced by about 50% in isolated perfused islets from vitamin D-deficient rats compared to controls [144]. Interestingly, 1,25-dihydroxyvitamin D₃ was noted in cell nuclei in the islets of langerhans [145]. Furthermore, administration of 1,25-dihydroxyvitamin D₃ to vitamin D-deficient rats improved insulin secretion significantly when compared to controls [146]. Vitamin D deficiency impaired both phases of insulin release in rats while correction of hypocalcemia failed to reverse the defect in insulin release [147]. Vitamin D, but not calcium, was essential for normal insulin secretion from the perfused rat pancreas [148]. The positive effect of single dose of 1,25-dihydroxyvitamin D₃ on insulin secretion was apparent at 8 h in perfused rat pancreata, peaked at 14 h, and then decreased to pretreatment baseline values by 36 h [149]. Dietary vitamin D₃ supplementation improved impaired glucose tolerance and insulin secretion in the vitamin D-deficient rats [150]. A dose-dependent effect from parenteral 1,25-dihydroxyvitamin D on insulin secretion and glucose metabolism was observed within 3 h and remained effective up to 20 h in the vitamin D-deficient rats [151]. The role of vitamin D on insulin synthesis and secretion was supported by studies in vitamin D receptor knockout mice. Insulin secretory capacity was reduced by 60% in vitamin D receptor knockout mice [152] with increased post-challenged blood glucose but normal fasting blood glucose concentration and reduced insulin mRNA levels in pancreatic islets but normal pancreatic beta cell mass, islet architecture, and islet neogenesis when compared to wild type mice. Thus, vitamin D plays an important role in pancreatic insulin synthesis and secretion *in vivo*.

Mechanistic studies of beta cell function: Although the essential role of vitamin D on insulin secretion has been established in vitamin D depleted laboratory animal, details of the underlying molecular mechanism remain to be defined. Employing a proteomic approach, treatment with 1,25-dihydroxyvitamin D₃ resulted in 31 differentially expressed proteins in INS-1 beta-like cells [153] with 29 upregulated, some of which were implicated in insulin granule motility and insulin exocytosis as well as regulation of ions. Pretreatment of INS1E cells with 1,25-dihydroxyvitamin D or 25-hydroxyvitamin D and glucose resulted in 526 and 181 differentially expressed genes, respectively [154].

Several molecular mechanisms were proposed to account for the effects of vitamin D on beta cells, including changes in the local pancreatic islet renin-angiotensin system [155], restoration of GLUT2 expression [156], enhancement of IP3 and AMPA receptor expression [157], vitamin D-binding protein-induced beta cell dedifferentiation [158], reduction of oxidative damage [159], reduced cholinergic pancreatic effects [160], enhanced transcriptional regulation of voltage-gated calcium channels [161], and elevation of PPAR-γ expression [162]. However, further studies are required to confirm the proposed mechanisms.

Insulin sensitivity

Functional studies of insulin sensitivity: In contrast to beta cell function, there are fewer studies of insulin sensitivity. Dietary supplementation of vitamin D improved insulin sensitivity, hepatic steatosis, and myocardial fibrosis in Western diet fed rats [163]. In dietary-induced obese mice, vitamin D receptor activation in liver macrophages improved insulin sensitivity with reduction of hepatic inflammation and steatosis [164]. Vitamin D treatment improved insulin resistance index in a nongenetic model of type 2 diabetes [165]. However, vitamin D status were not reported in these studies.

Mechanistic studies of insulin sensitivity: Chronic central administration of 1,25-dihydroxyvitamin D₃ dramatically reduced body weight, putatively by lowering food intake, in obese rodents [166]. Treatment with vitamin D increased mitochondrial function and insulin sensitivity, in part, through upregulation of perilipin 2, a perilipin protein upregulated with 1,25-dihydroxyvitamin D treatment [167]. In skeletal myocytes, vitamin D reduced insulin resistance by altering lipid partitioning and lipid droplet packaging in favor of lipid turnover [168]. FGF-23 knockout mice are hypoglycemic with profoundly increased peripheral insulin sensitivity and improved subcutaneous glucose tolerance. Ablation of vitamin D signaling in these mice normalized subcutaneous glucose tolerance tests and insulin sensitivity [169]. Caveolin-1 protein, which is necessary for vitamin D signaling, could play a role in vitamin D-induced insulin sensitivity in skeletal muscle [170]. In cultured rat osteoblasts, 1,25-dihydroxyvitamin D₃ treatment increased expression of the insulin and vitamin D receptors, and elevated osteocalcin levels under high glucose exposure [171], which may in turn improve insulin sensitivity.

However, the results of vitamin D receptor knockout mice were less uniform. Skeletal muscle-specific vitamin D receptor knockout mice developed insulin resistance and glucose intolerance accompanied by increased expression and activity of FOXO1[172]. Deletion of macrophage vitamin D receptor promoted insulin resistance and monocyte cholesterol transport and accelerated atherosclerosis[173]. In contrast, deletion of the vitamin D receptor gene in endothelial cells improved glucose tolerance and insulin sensitivity in skeletal muscle and reduced expression and secretion of insulin in pancreatic islets[174]. Together these data indicate that vitamin D has positive and negative effects on insulin sensitivity that are cell and organ specific.

CONCERNS ARISING WITH REPORTED STUDIES

Lack of true vitamin D deficient subjects

Due to publicity and potential non-calcemic benefits of vitamin D supplementation, the sale of vitamin D supplements increased significantly and taking vitamin D supplements is common. Thus, there are less true vitamin D deficient subjects available for inclusion in clinical trials. As well, a general lack of funding support for large trials impedes addressing the ability of researchers to address the gaps in knowledge surrounding vitamin D and its beneficial effects.

Lack of beneficial effects from suboptimal replacement and detrimental effects of over-replacement

To obtain the maximal effect of vitamin D, serum 25-hydroxyvitamin D concentration should be maintained in an optimal range, namely between 75 nmol/L (30 ng/mL) and 125 nmol/L (50 ng/mL). Inadequate vitamin D replacement therapy will reduce the chance to observe the expected beneficial effect of vitamin D while adverse effects associated with excessive serum 25-hydroxyvitamin D concentration will also cloud data interpretation. Although mega doses of vitamin D given intermittently could improve compliance in a study protocol, the predicted wide swings in serum 25-hydroxyvitamin D concentrations will confound outcomes. It is important in clinical studies to use a proper daily dose to avoid these pitfalls.

Inadequate sample size

The Diabetes Prevention Program demonstrated a 58% (95%CI: 48%-66%) reduction in the incidence of diabetes in the lifestyle intervention group (cumulative incidence of diabetes 14.4% in 1079 participants) and a 31% reduction in diabetes (95%CI: 17%-43%) in the metformin treated group (cumulative incidence of diabetes 21.7% in 1073 participants) when compared to the placebo (cumulative incidence of diabetes 28.9% in 1082 participants)[175]. Insulin sensitivity improved by 61.8% in the lifestyle intervention group and 28.3% in the metformin group[176]. This study can be employed to calculate a sample size sufficient for assessing the effects of vitamin D intervention.

Based on the non-linear relationship of serum 25-hydroxyvitamin D concentration and insulin sensitivity index as we reported[113], we constructed Table 9. Assuming a linear relationship between improvement in insulin sensitivity and reduction of diabetes from the Diabetes Prevention Program[175,176], we calculated the required sample size to detect the reduction of diabetes incidence after vitamin D replacement therapy in a population similar to that of the Diabetes Prevention Program[175] with a power of 0.80 to detect the proposed difference and a type I error rate, alpha, of 0.05 in a clinical trial of 3 years. Starting with a baseline serum 25-hydroxyvitamin D of 25 ng/mL (10 ng/mL), 170 subjects would be needed. Such a study cohort size is not excessive. However, if the baseline serum 25-hydroxyvitamin D is equal or greater than 50 nmol/L (20 ng/mL) the cohort size needed increases markedly. These calculations suggest that all studies to date are flawed secondary to inadequate sample size.

It has been frustrating to confound the published negative reports while ample evidence supports the benefit of vitamin D. Accordingly, we propose these guidelines [177]. Future studies into the effects of vitamin D supplementation need to ensure the proper selection of study subjects, adequate vitamin D replacement to achieve an optimal serum 25-hydroxyvitamin D concentrations, avoidance over-placement to eliminate detrimental effects, and adequate sample size to detect the proposed effects.

Table 9 Calculated sample size requirement to detect an improvement in insulin sensitivity based on a baseline serum 25-hydroxyvitamin D concentration of 40 ng/mL (100 nmol/L) and a power of 0.80 and alpha of 0.05

Initial serum 25-hydroxy-vitamin D concentration		Estimated insulin sensitivity index($\mu\text{M}/\text{min}/\text{m}^2/\text{pM}$)	Improvement in insulin sensitivity index with postintervention Serum 25-hydroxyvitamin D concentration 40 ng/mL (100 nmol/L)	Diabetes reduction based on the Diabetes Prevention Program	Sample size
ng/mL	nmol/L				
10	25	4.1326	0.8664	0.4361	340
15	37	5.4144	0.4246	0.2118	1602
20	50	6.2812	0.2280	0.1121	5934
25	62	6.8674	0.1232	0.0589	21878
30	75	7.2638	0.0619	0.0278	99260
35	87	7.5319	0.0241	0.0086	1041162

THE ISSUES THAT NEED TO BE ADDRESSED BY THE FUTURE STUDIES

Optimal serum 25-hydroxyvitamin D concentration for glucose metabolism

Table 4 summarizes the recommended serum vitamin D concentrations from several institutions and agencies. As appreciated, studies on bone health[14,15] showed no additional benefit in the subjects with serum 25-hydroxyvitamin D > 75 nmol/L (30 ng/mL) and this agrees with the effects upon diabetes prevention. However, increased all-cause mortality[107] and cardiovascular mortality[108] occurred prior to the 125 nmol/L (50 ng/mL) threshold, implying a much lower maximum dose for optimal serum 25-hydroxyvitamin D concentration. The question remains whether the same relationship applies to glucose homeostasis.

Detrimental effects on glucose metabolism for serum 25-hydroxyvitamin D concentrations above a maximum threshold

The detrimental effects noted in individuals with serum 25-hydroxyvitamin D concentration above a maximum threshold was observed in a cross-sectional study[109]. Further, improvement in bone density after discontinuation of vitamin D supplementation in osteoporotic patients with elevated serum 25-hydroxyvitamin D concentration was reported[110]. Elevated serum 25-hydroxyvitamin D concentrations were also associated with increased falls and fracture[118]. These reports suggest that assessment of negative effects from elevated serum 25-hydroxyvitamin D concentration may be uncovered with additional study.

Diabetes prevention in vitamin D deficit subjects

Although various evidence suggests the benefit of vitamin D on glucose metabolism, published diabetes prevention trials are not convincing and suffer from improper designed and execution. To address this issue, a well-designed and well-conducted randomized, placebo-control trial to test the effects of vitamin D to limit development of diabetes is warranted, by selecting true vitamin D deficient subjects, achieving optimal but not excessive serum 25-hydroxyvitamin concentration, and enrolling adequate number of subjects. Properly monitoring serum 25-hydroxyvitamin D concentrations is required during the study.

CONCLUSION

The role of vitamin D in glucose metabolism and fuel homeostasis is supported by a number of observational studies. We reported that serum 25-hydroxyvitamin D concentration accounted for 21.2% of the variation in insulin sensitivity index in univariate analysis and 6.1% by itself among 42% with other covariates in multivariate analysis[178]. We also reported that serum 25-hydroxyvitamin D concentration accounted for 8.2% of the variation in beta cell function in univariate analysis and 4.5% by itself among 25.5% with other covariates in multivariate analysis[179]. Although the intervention studies have failed to provide concordant data for multiple reasons, laboratory studies revealed a number of molecular mechanisms that underlie the effect

of vitamin D supporting the important role of the vitamin in glucose metabolism and fuel homeostasis. Since the independent contributions of vitamin D to insulin sensitivity[178] and beta cell function[179] are relatively small, vitamin D deficiency could be the last straw that breaks camel's back in polygenetic and multifactorial diseases, such as diabetes, obesity, and hyperlipidemia.

ACKNOWLEDGEMENTS

A special acknowledgement is due to Chiu-Tien Chiu, MD, PhD for his unconditional support to KCC. We are in debt to Jeffrey Isenberg MD, MPH for critical reading and editing of the manuscript.

REFERENCES

- 1 **Diamond J.** Evolutionary biology: geography and skin colour. *Nature* 2005; **435**: 283-284 [PMID: 15902239 DOI: 10.1038/435283a]
- 2 **Platt BS.** Sir Edward Mellanby, G.B.E., K.C.B., F.R.S. *Nature* 1955; **175**: 530-532 [PMID: 14370159 DOI: 10.1038/175530a0]
- 3 **McCullum EV, Pitz W, Simmonds N, Becker JE, Shipley PG, Bunting RW.** The effect of additions of fluorine to the diet of the rat on the quality of the teeth. 1925. Studies on experimental rickets. XXI. An experimental demonstration of the existence of a vitamin which promotes calcium deposition. 1922. The effect of additions of fluorine to the diet of the rat on the quality of the teeth. 1925. *J Biol Chem* 2002; **277**: E8 [PMID: 11991957]
- 4 **DeLuca HF.** History of the discovery of vitamin D and its active metabolites. *Bonekey Rep* 2014; **3**: 479 [PMID: 24466410 DOI: 10.1038/bonekey.2013.213]
- 5 **Askew FA, Bourdillon RB, Webster TA.** The production of vitamin D in a glow discharge. *Biochem J* 1932; **26**: 814 [PMID: 16744889 DOI: 10.1042/bj0260814]
- 6 **DeLuca HF.** Current concepts. Vitamin D. *N Engl J Med* 1969; **281**: 1103-1104 [PMID: 4309963 DOI: 10.1056/NEJM196911132812006]
- 7 **Holick MF.** Noncalcemic actions of 1,25-dihydroxyvitamin D₃ and clinical applications. *Bone* 1995; **17**: 107S-111S [PMID: 8579891 DOI: 10.1016/8756-3282(95)00195-j]
- 8 **Mitchell F.** Vitamin-D and COVID-19: do deficient risk a poorer outcome? *Lancet Diabetes Endocrinol* 2020; **8**: 570 [PMID: 32445630 DOI: 10.1016/S2213-8587(20)30183-2]
- 9 **Holick MF.** Vitamin D deficiency. *N Engl J Med* 2007; **357**: 266-281 [PMID: 17634462 DOI: 10.1056/NEJMr070553]
- 10 **Aloia J, Fazzari M, Islam S, Mikhail M, Shieh A, Katumuluwa S, Dhaliwal R, Stolberg A, Usera G, Ragolia L.** Vitamin D Supplementation in Elderly Black Women Does Not Prevent Bone Loss: A Randomized Controlled Trial. *J Bone Miner Res* 2018; **33**: 1916-1922 [PMID: 29905969 DOI: 10.1002/jbmr.3521]
- 11 **Aspray TJ, Chadwick T, Francis RM, McColl E, Stamp E, Prentice A, von Wilamowitz-Moellendorff A, Schoenmakers I.** Randomized controlled trial of vitamin D supplementation in older people to optimize bone health. *Am J Clin Nutr* 2019; **109**: 207-217 [PMID: 30624670 DOI: 10.1093/ajcn/nqy280]
- 12 **Burt LA, Billington EO, Rose MS, Raymond DA, Hanley DA, Boyd SK.** Effect of High-Dose Vitamin D Supplementation on Volumetric Bone Density and Bone Strength: A Randomized Clinical Trial. *JAMA* 2019; **322**: 736-745 [PMID: 31454046 DOI: 10.1001/jama.2019.11889]
- 13 **Reid IR, Bolland MJ, Grey A.** Effects of vitamin D supplements on bone mineral density: a systematic review and meta-analysis. *Lancet* 2014; **383**: 146-155 [PMID: 24119980 DOI: 10.1016/S0140-6736(13)61647-5]
- 14 **Reid IR, Horne AM, Mihov B, Gamble GD, Al-Abuwi F, Singh M, Taylor L, Fenwick S, Camargo CA, Stewart AW, Scragg R.** Effect of monthly high-dose vitamin D on bone density in community-dwelling older adults substudy of a randomized controlled trial. *J Intern Med* 2017; **282**: 452-460 [PMID: 28692172 DOI: 10.1111/joim.12651]
- 15 **Macdonald HM, Reid IR, Gamble GD, Fraser WD, Tang JC, Wood AD.** 25-Hydroxyvitamin D Threshold for the Effects of Vitamin D Supplements on Bone Density: Secondary Analysis of a Randomized Controlled Trial. *J Bone Miner Res* 2018; **33**: 1464-1469 [PMID: 29665087 DOI: 10.1002/jbmr.3442]
- 16 **Glossmann HH.** Origin of 7-dehydrocholesterol (provitamin D) in the skin. *J Invest Dermatol* 2010; **130**: 2139-2141 [PMID: 20445550 DOI: 10.1038/jid.2010.118]
- 17 **Nicoloff JT, Low JC, Dussault JH, Fisher DA.** Simultaneous measurement of thyroxine and triiodothyronine peripheral turnover kinetics in man. *J Clin Invest* 1972; **51**: 473-483 [PMID: 4110897 DOI: 10.1172/jci106835]
- 18 **Jones KS, Assar S, Harnpanich D, Bouillon R, Lambrechts D, Prentice A, Schoenmakers I.** 25(OH)D₂ half-life is shorter than 25(OH)D₃ half-life and is influenced by DBP concentration and genotype. *J Clin Endocrinol Metab* 2014; **99**: 3373-3381 [PMID: 24885631 DOI: 10.1210.2014-0311]

- 10.1210/jc.2014-1714]
- 19 **Berry D**, Hyppönen E. Determinants of vitamin D status: focus on genetic variations. *Curr Opin Nephrol Hypertens* 2011; **20**: 331-336 [PMID: 21654390 DOI: 10.1097/MNH.0b013e328346d6ba]
- 20 **Fagerberg L**, Hallström BM, Oksvold P, Kampf C, Djureinovic D, Odeberg J, Habuka M, Tahmasebpour S, Danielsson A, Edlund K, Asplund A, Sjöstedt E, Lundberg E, Szgyarto CA, Skogs M, Takanen JO, Berling H, Tegel H, Mulder J, Nilsson P, Schwenk JM, Lindskog C, Danielsson F, Mardinoglu A, Sivertsson A, von Feilitzen K, Forsberg M, Zwahlen M, Olsson I, Navani S, Huss M, Nielsen J, Ponten F, Uhlén M. Analysis of the human tissue-specific expression by genome-wide integration of transcriptomics and antibody-based proteomics. *Mol Cell Proteomics* 2014; **13**: 397-406 [PMID: 24309898 DOI: 10.1074/mcp.M113.035600]
- 21 **Lee S**, Clark SA, Gill RK, Christakos S. 1,25-Dihydroxyvitamin D3 and pancreatic beta-cell function: vitamin D receptors, gene expression, and insulin secretion. *Endocrinology* 1994; **134**: 1602-1610 [PMID: 8137721 DOI: 10.1210/endo.134.4.8137721]
- 22 **Zhang H**, Shen Z, Lin Y, Zhang J, Zhang Y, Liu P, Zeng H, Yu M, Chen X, Ning L, Mao X, Cen L, Yu C, Xu C. Vitamin D receptor targets hepatocyte nuclear factor 4a and mediates protective effects of vitamin D in nonalcoholic fatty liver disease. *J Biol Chem* 2020; **295**: 3891-3905 [PMID: 32051143 DOI: 10.1074/jbc.RA119.011487]
- 23 **Bischoff-Ferrari HA**, Borchers M, Gudat F, Dürmüller U, Stähelin HB, Dick W. Vitamin D receptor expression in human muscle tissue decreases with age. *J Bone Miner Res* 2004; **19**: 265-269 [PMID: 14969396 DOI: 10.1359/jbmr.2004.19.2.265]
- 24 **Yuzbashian E**, Asghari G, Hedayati M, Zarkesh M, Mirmiran P, Khalaj A. Determinants of vitamin D receptor gene expression in visceral and subcutaneous adipose tissue in non-obese, obese, and morbidly obese subjects. *J Steroid Biochem Mol Biol* 2019; **187**: 82-87 [PMID: 30412764 DOI: 10.1016/j.jsbmb.2018.11.004]
- 25 **Bland R**, Markovic D, Hills CE, Hughes SV, Chan SL, Squires PE, Hewison M. Expression of 25-hydroxyvitamin D3-1alpha-hydroxylase in pancreatic islets. *J Steroid Biochem Mol Biol* 2004; **89-90**: 121-125 [PMID: 15225758 DOI: 10.1016/j.jsbmb.2004.03.115]
- 26 **Vuica A**, Ferhatović Hamzić L, Vukojević K, Jerić M, Puljak L, Grković I, Filipović N. Aging and a long-term diabetes mellitus increase expression of 1 α -hydroxylase and vitamin D receptors in the rat liver. *Exp Gerontol* 2015; **72**: 167-176 [PMID: 26471398 DOI: 10.1016/j.exger.2015.10.005]
- 27 **Srikuea R**, Zhang X, Park-Sarge OK, Esser KA. VDR and CYP27B1 are expressed in C2C12 cells and regenerating skeletal muscle: potential role in suppression of myoblast proliferation. *Am J Physiol Cell Physiol* 2012; **303**: C396-C405 [PMID: 22648952 DOI: 10.1152/ajpcell.00014.2012]
- 28 **Wamberg L**, Christiansen T, Paulsen SK, Fisker S, Rask P, Rejnmark L, Richelsen B, Pedersen SB. Expression of vitamin D-metabolizing enzymes in human adipose tissue -- the effect of obesity and diet-induced weight loss. *Int J Obes (Lond)* 2013; **37**: 651-657 [PMID: 22828938 DOI: 10.1038/ijo.2012.112]
- 29 **Hewison M**, Zehnder D, Bland R, Stewart PM. 1alpha-Hydroxylase and the action of vitamin D. *J Mol Endocrinol* 2000; **25**: 141-148 [PMID: 11013342 DOI: 10.1677/jme.0.0250141]
- 30 **Garland CF**, Garland FC. Do sunlight and vitamin D reduce the likelihood of colon cancer? *Int J Epidemiol* 1980; **9**: 227-231 [PMID: 7440046 DOI: 10.1093/ije/9.3.227]
- 31 **Garland FC**, Garland CF, Gorham ED, Young JF. Geographic variation in breast cancer mortality in the United States: a hypothesis involving exposure to solar radiation. *Prev Med* 1990; **19**: 614-622 [PMID: 2263572 DOI: 10.1016/0091-7435(90)90058-r]
- 32 **Ahonen MH**, Tenkanen L, Teppo L, Hakama M, Tuohimaa P. Prostate cancer risk and prediagnostic serum 25-hydroxyvitamin D levels (Finland). *Cancer Causes Control* 2000; **11**: 847-852 [PMID: 11075874 DOI: 10.1023/a:1008923802001]
- 33 **Garland CF**, Comstock GW, Garland FC, Helsing KJ, Shaw EK, Gorham ED. Serum 25-hydroxyvitamin D and colon cancer: eight-year prospective study. *Lancet* 1989; **2**: 1176-1178 [PMID: 2572900 DOI: 10.1016/s0140-6736(89)91789-3]
- 34 **Abe E**, Miyaura C, Sakagami H, Takeda M, Konno K, Yamazaki T, Yoshiki S, Suda T. Differentiation of mouse myeloid leukemia cells induced by 1 alpha,25-dihydroxyvitamin D3. *Proc Natl Acad Sci U S A* 1981; **78**: 4990-4994 [PMID: 6946446 DOI: 10.1073/pnas.78.8.4990]
- 35 **Peehl DM**, Skowronski RJ, Leung GK, Wong ST, Stamey TA, Feldman D. Antiproliferative effects of 1,25-dihydroxyvitamin D3 on primary cultures of human prostatic cells. *Cancer Res* 1994; **54**: 805-810 [PMID: 7508338]
- 36 **Masuda S**, Jones G. Promise of vitamin D analogues in the treatment of hyperproliferative conditions. *Mol Cancer Ther* 2006; **5**: 797-808 [PMID: 16648549 DOI: 10.1158/1535-7163.MCT-05-0539]
- 37 **Samuel S**, Sitrin MD. Vitamin D's role in cell proliferation and differentiation. *Nutr Rev* 2008; **66**: S116-S124 [PMID: 18844838 DOI: 10.1111/j.1753-4887.2008.00094.x]
- 38 **Hyppönen E**, Läärä E, Reunanen A, Järvelin MR, Virtanen SM. Intake of vitamin D and risk of type 1 diabetes: a birth-cohort study. *Lancet* 2001; **358**: 1500-1503 [PMID: 11705562 DOI: 10.1016/S0140-6736(01)06580-1]
- 39 **Chang TJ**, Lei HH, Yeh JI, Chiu KC, Lee KC, Chen MC, Tai TY, Chuang LM. Vitamin D receptor gene polymorphisms influence susceptibility to type 1 diabetes mellitus in the Taiwanese population. *Clin Endocrinol (Oxf)* 2000; **52**: 575-580 [PMID: 10792336 DOI: 10.1046/j.1365-2265.2000.00985.x]
- 40 **Zella JB**, DeLuca HF. Vitamin D and autoimmune diabetes. *J Cell Biochem* 2003; **88**: 216-222

- [PMID: 12520517 DOI: 10.1002/jcb.10347]
- 41 **Munger KL**, Zhang SM, O'Reilly E, Hernán MA, Olek MJ, Willett WC, Ascherio A. Vitamin D intake and incidence of multiple sclerosis. *Neurology* 2004; **62**: 60-65 [PMID: 14718698 DOI: 10.1212/01.wnl.0000101723.79681.38]
 - 42 **Kamen DL**, Cooper GS, Bouali H, Shaftman SR, Hollis BW, Gilkeson GS. Vitamin D deficiency in systemic lupus erythematosus. *Autoimmun Rev* 2006; **5**: 114-117 [PMID: 16431339 DOI: 10.1016/j.autrev.2005.05.009]
 - 43 **Maruotti N**, Cantatore FP. Vitamin D and the immune system. *J Rheumatol* 2010; **37**: 491-495 [PMID: 20080911 DOI: 10.3899/jrheum.090797]
 - 44 **Murdaca G**, Tonacci A, Negrini S, Greco M, Borro M, Puppo F, Gangemi S. Emerging role of vitamin D in autoimmune diseases: An update on evidence and therapeutic implications. *Autoimmun Rev* 2019; **18**: 102350 [PMID: 31323357 DOI: 10.1016/j.autrev.2019.102350]
 - 45 **Chen S**, Sims GP, Chen XX, Gu YY, Chen S, Lipsky PE. Modulatory effects of 1,25-dihydroxyvitamin D3 on human B cell differentiation. *J Immunol* 2007; **179**: 1634-1647 [PMID: 17641030 DOI: 10.4049/jimmunol.179.3.1634]
 - 46 The sun cure for surgical tuberculosis. *Br Med J* 1923; **2**: 111-112 [PMID: 20771236]
 - 47 **Klip W**. The tuberculostatic action of vitamin D2. *Antonie Van Leeuwenhoek* 1952; **18**: 217-226 [PMID: 14944180 DOI: 10.1007/BF02538610]
 - 48 **Davies PD**, Brown RC, Woodhead JS. Serum concentrations of vitamin D metabolites in untreated tuberculosis. *Thorax* 1985; **40**: 187-190 [PMID: 3872485 DOI: 10.1136/thx.40.3.187]
 - 49 **Liu PT**, Stenger S, Li H, Wenzel L, Tan BH, Krutzik SR, Ochoa MT, Schaubert J, Wu K, Meinken C, Kamen DL, Wagner M, Bals R, Steinmeyer A, Zügel U, Gallo RL, Eisenberg D, Hewison M, Hollis BW, Adams JS, Bloom BR, Modlin RL. Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science* 2006; **311**: 1770-1773 [PMID: 16497887 DOI: 10.1126/science.1123933]
 - 50 **Zasloff M**. Fighting infections with vitamin D. *Nat Med* 2006; **12**: 388-390 [PMID: 16598282 DOI: 10.1038/nm0406-388]
 - 51 **Laaksi I**, Ruohola JP, Tuohimaa P, Auvinen A, Haataja R, Pihlajamäki H, Ylikomi T. An association of serum vitamin D concentrations < 40 nmol/L with acute respiratory tract infection in young Finnish men. *Am J Clin Nutr* 2007; **86**: 714-717 [PMID: 17823437 DOI: 10.1093/ajcn/86.3.714]
 - 52 **Bodnar LM**, Krohn MA, Simhan HN. Maternal vitamin D deficiency is associated with bacterial vaginosis in the first trimester of pregnancy. *J Nutr* 2009; **139**: 1157-1161 [PMID: 19357214 DOI: 10.3945/jn.108.103168]
 - 53 **Inamo Y**, Hasegawa M, Saito K, Hayashi R, Ishikawa T, Yoshino Y, Hashimoto K, Fuchigami T. Serum vitamin D concentrations and associated severity of acute lower respiratory tract infections in Japanese hospitalized children. *Pediatr Int* 2011; **53**: 199-201 [PMID: 21648117 DOI: 10.1111/j.1442-200x.2010.03224.x]
 - 54 **Tiwari S**, Pratyush DD, Gupta B, Dwivedi A, Chaudhary S, Rayicherla RK, Gupta SK, Singh SK. Prevalence and severity of vitamin D deficiency in patients with diabetic foot infection. *Br J Nutr* 2013; **109**: 99-102 [PMID: 22715859 DOI: 10.1017/S0007114512000578]
 - 55 **Terrier B**, Jehan F, Munteanu M, Geri G, Saadoun D, Sène D, Poynard T, Souberbielle JC, Cacoub P. Low 25-hydroxyvitamin D serum levels correlate with the presence of extra-hepatic manifestations in chronic hepatitis C virus infection. *Rheumatology (Oxford)* 2012; **51**: 2083-2090 [PMID: 22908327 DOI: 10.1093/rheumatology/kes209]
 - 56 **Pinzone MR**, Di Rosa M, Malaguarnera M, Madeddu G, Focà E, Ceccarelli G, d'Ettorre G, Vullo V, Fisichella R, Cacopardo B, Nunnari G. Vitamin D deficiency in HIV infection: an underestimated and undertreated epidemic. *Eur Rev Med Pharmacol Sci* 2013; **17**: 1218-1232 [PMID: 23690192]
 - 57 **Abrishami A**, Dalili N, Mohammadi Torbati P, Asgari R, Arab-Ahmadi M, Behnam B, Sanei-Taheri M. Possible association of vitamin D status with lung involvement and outcome in patients with COVID-19: a retrospective study. *Eur J Nutr* 2021; **60**: 2249-2257 [PMID: 33123774 DOI: 10.1007/s00394-020-02411-0]
 - 58 **Panagiotou G**, Tee SA, Ihsan Y, Athar W, Marchitelli G, Kelly D, Boot CS, Stock N, Macfarlane J, Martineau AR, Burns G, Quinton R. Low serum 25-hydroxyvitamin D (25[OH]D) levels in patients hospitalized with COVID-19 are associated with greater disease severity. *Clin Endocrinol (Oxf)* 2020; **93**: 508-511 [PMID: 32621392 DOI: 10.1111/cen.14276]
 - 59 **Merzon E**, Tworowski D, Gorohovski A, Vinker S, Golan Cohen A, Green I, Frenkel-Morgenstern M. Low plasma 25(OH) vitamin D level is associated with increased risk of COVID-19 infection: an Israeli population-based study. *FEBS J* 2020; **287**: 3693-3702 [PMID: 32700398 DOI: 10.1111/febs.15495]
 - 60 **Carpagnano GE**, Di Lecce V, Quaranta VN, Zito A, Buonamico E, Capozza E, Palumbo A, Di Gioia G, Valerio VN, Resta O. Vitamin D deficiency as a predictor of poor prognosis in patients with acute respiratory failure due to COVID-19. *J Endocrinol Invest* 2021; **44**: 765-771 [PMID: 32772324 DOI: 10.1007/s40618-020-01370-x]
 - 61 **Meltzer DO**, Best TJ, Zhang H, Vokes T, Arora V, Solway J. Association of Vitamin D Status and Other Clinical Characteristics With COVID-19 Test Results. *JAMA Netw Open* 2020; **3**: e2019722 [PMID: 32880651 DOI: 10.1001/jamanetworkopen.2020.19722]
 - 62 **Van Schooten FJ**, Hirvonen A, Maas LM, De Mol BA, Kleinjans JC, Bell DA, Durrer JD. Putative susceptibility markers of coronary artery disease: association between VDR genotype, smoking, and

- aromatic DNA adduct levels in human right atrial tissue. *FASEB J* 1998; **12**: 1409-1417 [PMID: 9761785 DOI: 10.1096/fasebj.12.13.1409]
- 63 **Giovannucci E**, Liu Y, Hollis BW, Rimm EB. 25-hydroxyvitamin D and risk of myocardial infarction in men: a prospective study. *Arch Intern Med* 2008; **168**: 1174-1180 [PMID: 18541825 DOI: 10.1001/archinte.168.11.1174]
 - 64 **Scragg R**, Sowers M, Bell C. Serum 25-hydroxyvitamin D, ethnicity, and blood pressure in the Third National Health and Nutrition Examination Survey. *Am J Hypertens* 2007; **20**: 713-719 [PMID: 17586404 DOI: 10.1016/j.amjhyper.2007.01.017]
 - 65 **Pilz S**, Dobnig H, Fischer JE, Wellnitz B, Seelhorst U, Boehm BO, März W. Low vitamin d levels predict stroke in patients referred to coronary angiography. *Stroke* 2008; **39**: 2611-2613 [PMID: 18635847 DOI: 10.1161/STROKEAHA.107.513655]
 - 66 **Xiang W**, Kong J, Chen S, Cao LP, Qiao G, Zheng W, Liu W, Li X, Gardner DG, Li YC. Cardiac hypertrophy in vitamin D receptor knockout mice: role of the systemic and cardiac renin-angiotensin systems. *Am J Physiol Endocrinol Metab* 2005; **288**: E125-E132 [PMID: 15367398 DOI: 10.1152/ajpendo.00224.2004]
 - 67 **Watson KE**, Abrolat ML, Malone LL, Hoeg JM, Doherty T, Detrano R, Demer LL. Active serum vitamin D levels are inversely correlated with coronary calcification. *Circulation* 1997; **96**: 1755-1760 [PMID: 9323058 DOI: 10.1161/01.cir.96.6.1755]
 - 68 **Sultan M**, Twito O, Tohami T, Ramati E, Neumark E, Rashid G. Vitamin D diminishes the high platelet aggregation of type 2 diabetes mellitus patients. *Platelets* 2019; **30**: 120-125 [PMID: 29313404 DOI: 10.1080/09537104.2017.1386298]
 - 69 **Schleithoff SS**, Zittermann A, Tenderich G, Berthold HK, Stehle P, Koerfer R. Vitamin D supplementation improves cytokine profiles in patients with congestive heart failure: a double-blind, randomized, placebo-controlled trial. *Am J Clin Nutr* 2006; **83**: 754-759 [PMID: 16600924 DOI: 10.1093/ajcn/83.4.754]
 - 70 **Kumar S**, Davies M, Zakaria Y, Mawer EB, Gordon C, Olukoga AO, Boulton AJ. Improvement in glucose tolerance and beta-cell function in a patient with vitamin D deficiency during treatment with vitamin D. *Postgrad Med J* 1994; **70**: 440-443 [PMID: 8029165 DOI: 10.1136/pgmj.70.824.440]
 - 71 **Boucher BJ**, Mannan N, Noonan K, Hales CN, Evans SJ. Glucose intolerance and impairment of insulin secretion in relation to vitamin D deficiency in east London Asians. *Diabetologia* 1995; **38**: 1239-1245 [PMID: 8690178 DOI: 10.1007/BF00422375]
 - 72 **Bell NH**, Epstein S, Greene A, Shary J, Oexmann MJ, Shaw S. Evidence for alteration of the vitamin D-endocrine system in obese subjects. *J Clin Invest* 1985; **76**: 370-373 [PMID: 2991340 DOI: 10.1172/JCI11971]
 - 73 **Ganji V**, Milone C, Cody MM, McCarty F, Wang YT. Serum vitamin D concentrations are related to depression in young adult US population: the Third National Health and Nutrition Examination Survey. *Int Arch Med* 2010; **3**: 29 [PMID: 21067618 DOI: 10.1186/1755-7682-3-29]
 - 74 **Zhu JL**, Luo WW, Cheng X, Li Y, Zhang QZ, Peng WX. Vitamin D deficiency and Schizophrenia in Adults: A Systematic Review and Meta-analysis of Observational Studies. *Psychiatry Res* 2020; **288**: 112959 [PMID: 32335466 DOI: 10.1016/j.psychres.2020.112959]
 - 75 **Wang T**, Shan L, Du L, Feng J, Xu Z, Staal WG, Jia F. Serum concentration of 25-hydroxyvitamin D in autism spectrum disorder: a systematic review and meta-analysis. *Eur Child Adolesc Psychiatry* 2016; **25**: 341-350 [PMID: 26514973 DOI: 10.1007/s00787-015-0786-1]
 - 76 **Lv Z**, Qi H, Wang L, Fan X, Han F, Wang H, Bi S. Vitamin D status and Parkinson's disease: a systematic review and meta-analysis. *Neurol Sci* 2014; **35**: 1723-1730 [PMID: 24847960 DOI: 10.1007/s10072-014-1821-6]
 - 77 **Smith MP**, Fletcher-Turner A, Yurek DM, Cass WA. Calcitriol protection against dopamine loss induced by intracerebroventricular administration of 6-hydroxydopamine. *Neurochem Res* 2006; **31**: 533-539 [PMID: 16758362 DOI: 10.1007/s11064-006-9048-4]
 - 78 **Mayne PE**, Burne THJ. Vitamin D in Synaptic Plasticity, Cognitive Function, and Neuropsychiatric Illness. *Trends Neurosci* 2019; **42**: 293-306 [PMID: 30795846 DOI: 10.1016/j.tins.2019.01.003]
 - 79 **Almeras L**, Eyles D, Benech P, Laffite D, Villard C, Patatian A, Boucraut J, Mackay-Sim A, Mgrath J, Féron F. Developmental vitamin D deficiency alters brain protein expression in the adult rat: implications for neuropsychiatric disorders. *Proteomics* 2007; **7**: 769-780 [PMID: 17295352 DOI: 10.1002/pmic.200600392]
 - 80 **Eyles DW**, Smith S, Kinobe R, Hewison M, Mgrath JJ. Distribution of the vitamin D receptor and 1 alpha-hydroxylase in human brain. *J Chem Neuroanat* 2005; **29**: 21-30 [PMID: 15589699 DOI: 10.1016/j.jchemneu.2004.08.006]
 - 81 **Phillips KM**, Ruggio DM, Horst RL, Minor B, Simon RR, Feeney MJ, Byrdwell WC, Haytowitz DB. Vitamin D and sterol composition of 10 types of mushrooms from retail suppliers in the United States. *J Agric Food Chem* 2011; **59**: 7841-7853 [PMID: 21663327 DOI: 10.1021/jf104246z]
 - 82 **Libon F**, Cavalier E, Nikkels AF. Skin color is relevant to vitamin D synthesis. *Dermatology* 2013; **227**: 250-254 [PMID: 24134867 DOI: 10.1159/000354750]
 - 83 **Jablonski NG**, Chaplin G. Colloquium paper: human skin pigmentation as an adaptation to UV radiation. *Proc Natl Acad Sci U S A* 2010; **107** Suppl 2: 8962-8968 [PMID: 20445093 DOI: 10.1073/pnas.0914628107]
 - 84 **Holick MF**. Vitamin D: the underappreciated D-lightful hormone that is important for skeletal and cellular health. *Curr Opin Endocrinol Diabetes* 2002; **9**: 87-98
 - 85 **Barger-Lux MJ**, Heaney RP. Effects of above average summer sun exposure on serum 25-

- hydroxyvitamin D and calcium absorption. *J Clin Endocrinol Metab* 2002; **87**: 4952-4956 [PMID: 12414856 DOI: 10.1210/jc.2002-020636]
- 86 **Rockell JE**, Skeaff CM, Williams SM, Green TJ. Association between quantitative measures of skin color and plasma 25-hydroxyvitamin D. *Osteoporos Int* 2008; **19**: 1639-1642 [PMID: 18408879 DOI: 10.1007/s00198-008-0620-4]
 - 87 **Epstein JH**. Photocarcinogenesis, skin cancer, and aging. *J Am Acad Dermatol* 1983; **9**: 487-502 [PMID: 6355213 DOI: 10.1016/s0190-9622(83)70160-x]
 - 88 **Heaney RP**, Horst RL, Cullen DM, Armas LA. Vitamin D3 distribution and status in the body. *J Am Coll Nutr* 2009; **28**: 252-256 [PMID: 20150598 DOI: 10.1080/07315724.2009.10719779]
 - 89 **Bouillon R**, Schuit F, Antonio L, Rastinejad F. Vitamin D Binding Protein: A Historic Overview. *Front Endocrinol (Lausanne)* 2019; **10**: 910 [PMID: 31998239 DOI: 10.3389/fendo.2019.00910]
 - 90 **Haddad JG**, Matsuoka LY, Hollis BW, Hu YZ, Wortsman J. Human plasma transport of vitamin D after its endogenous synthesis. *J Clin Invest* 1993; **91**: 2552-2555 [PMID: 8390483 DOI: 10.1172/JCI116492]
 - 91 **Holmberg I**, Berlin T, Ewerth S, Björkhem I. 25-Hydroxylase activity in subcellular fractions from human liver. Evidence for different rates of mitochondrial hydroxylation of vitamin D2 and D3. *Scand J Clin Lab Invest* 1986; **46**: 785-790 [PMID: 3026027 DOI: 10.3109/00365518609084051]
 - 92 **Armas LA**, Hollis BW, Heaney RP. Vitamin D2 is much less effective than vitamin D3 in humans. *J Clin Endocrinol Metab* 2004; **89**: 5387-5391 [PMID: 15531486 DOI: 10.1210/jc.2004-0360]
 - 93 **Zarei A**, Hulley PA, Sabokbar A, Javaid MK, Morovat A. 25-Hydroxy- and 1 α ,25-Dihydroxycholecalciferol Have Greater Potencies than 25-Hydroxy- and 1 α ,25-Dihydroxyergocalciferol in Modulating Cultured Human and Mouse Osteoblast Activities. *PLoS One* 2016; **11**: e0165462 [PMID: 27893751 DOI: 10.1371/journal.pone.0165462]
 - 94 **Jones G**, Byrnes B, Palma F, Segev D, Mazur Y. Displacement potency of vitamin D2 analogs in competitive protein-binding assays for 25-hydroxyvitamin D3, 24,25-dihydroxyvitamin D3, and 1,25-dihydroxyvitamin D3. *J Clin Endocrinol Metab* 1980; **50**: 773-775 [PMID: 6965943 DOI: 10.1210/jcem-50-4-773]
 - 95 **Holick MF**, Biancuzzo RM, Chen TC, Klein EK, Young A, Bibuld D, Reitz R, Salameh W, Ameri A, Tannenbaum AD. Vitamin D2 is as effective as vitamin D3 in maintaining circulating concentrations of 25-hydroxyvitamin D. *J Clin Endocrinol Metab* 2008; **93**: 677-681 [PMID: 18089691 DOI: 10.1210/jc.2007-2308]
 - 96 **Trang HM**, Cole DE, Rubin LA, Pierratos A, Siu S, Vieth R. Evidence that vitamin D3 increases serum 25-hydroxyvitamin D more efficiently than does vitamin D2. *Am J Clin Nutr* 1998; **68**: 854-858 [PMID: 9771862 DOI: 10.1093/ajcn/68.4.854]
 - 97 **Romagnoli E**, Mascia ML, Cipriani C, Fassino V, Mazzei F, D'Erasmo E, Carnevale V, Scillitani A, Minisola S. Short and long-term variations in serum calcitropic hormones after a single very large dose of ergocalciferol (vitamin D2) or cholecalciferol (vitamin D3) in the elderly. *J Clin Endocrinol Metab* 2008; **93**: 3015-3020 [PMID: 18492750 DOI: 10.1210/jc.2008-0350]
 - 98 **Heaney RP**, Recker RR, Grote J, Horst RL, Armas LA. Vitamin D(3) is more potent than vitamin D(2) in humans. *J Clin Endocrinol Metab* 2011; **96**: E447-E452 [PMID: 21177785 DOI: 10.1210/jc.2010-2230]
 - 99 **Glendenning P**, Chew GT, Seymour HM, Gillett MJ, Goldswain PR, Inderjeeth CA, Vasikaran SD, Taranto M, Musk AA, Fraser WD. Serum 25-hydroxyvitamin D levels in vitamin D-insufficient hip fracture patients after supplementation with ergocalciferol and cholecalciferol. *Bone* 2009; **45**: 870-875 [PMID: 19631774 DOI: 10.1016/j.bone.2009.07.015]
 - 100 **Shieh A**, Chun RF, Ma C, Witzel S, Meyer B, Rafison B, Swinkels L, Huijs T, Pepkowitz S, Holmquist B, Hewison M, Adams JS. Effects of High-Dose Vitamin D2 Versus D3 on Total and Free 25-Hydroxyvitamin D and Markers of Calcium Balance. *J Clin Endocrinol Metab* 2016; **101**: 3070-3078 [PMID: 27192696 DOI: 10.1210/jc.2016-1871]
 - 101 **Tripkovic L**, Lambert H, Hart K, Smith CP, Bucca G, Penson S, Chope G, Hyppönen E, Berry J, Vieth R, Lanham-New S. Comparison of vitamin D2 and vitamin D3 supplementation in raising serum 25-hydroxyvitamin D status: a systematic review and meta-analysis. *Am J Clin Nutr* 2012; **95**: 1357-1364 [PMID: 22552031 DOI: 10.3945/ajcn.111.031070]
 - 102 **Cipriani C**, Romagnoli E, Pepe J, Russo S, Carlucci L, Piemonte S, Nieddu L, McMahon DJ, Singh R, Minisola S. Long-term bioavailability after a single oral or intramuscular administration of 600,000 IU of ergocalciferol or cholecalciferol: implications for treatment and prophylaxis. *J Clin Endocrinol Metab* 2013; **98**: 2709-2715 [PMID: 23766519 DOI: 10.1210/jc.2013-1586]
 - 103 **Malabanan A**, Veronikis IE, Holick MF. Redefining vitamin D insufficiency. *Lancet* 1998; **351**: 805-806 [PMID: 9519960 DOI: 10.1016/s0140-6736(05)78933-9]
 - 104 **Institute of Medicine Committee to Review Dietary Reference Intakes for Vitamin D, Calcium**. Dietary Reference Intakes for Calcium and Vitamin D. Ross AC, Taylor CL, Yaktine AL, Del Valle HB, editors. Washington (DC): National Academies Press (US), 2011
 - 105 **Chapuy MC**, Preziosi P, Maamer M, Arnaud S, Galan P, Hercberg S, Meunier PJ. Prevalence of vitamin D insufficiency in an adult normal population. *Osteoporos Int* 1997; **7**: 439-443 [PMID: 9425501 DOI: 10.1007/s001980050030]
 - 106 **Heaney RP**. Functional indices of vitamin D status and ramifications of vitamin D deficiency. *Am J Clin Nutr* 2004; **80**: 1706S-1709S [PMID: 15585791 DOI: 10.1093/ajcn/80.6.1706S]
 - 107 **Melamed ML**, Michos ED, Post W, Astor B. 25-hydroxyvitamin D levels and the risk of mortality in the general population. *Arch Intern Med* 2008; **168**: 1629-1637 [PMID: 18695076 DOI: 10.1001/archinternmed.2008.1629-1637]

- 10.1001/archinte.168.15.1629]
- 108 **Durup D**, Jørgensen HL, Christensen J, Tjønneland A, Olsen A, Halkjær J, Lind B, Heegaard AM, Schwarz P. A Reverse J-Shaped Association Between Serum 25-Hydroxyvitamin D and Cardiovascular Disease Mortality: The CopD Study. *J Clin Endocrinol Metab* 2015; **100**: 2339-2346 [PMID: 25710567 DOI: 10.1210/jc.2014-4551]
 - 109 **Bischoff-Ferrari HA**, Giovannucci E, Willett WC, Dietrich T, Dawson-Hughes B. Estimation of optimal serum concentrations of 25-hydroxyvitamin D for multiple health outcomes. *Am J Clin Nutr* 2006; **84**: 18-28 [PMID: 16825677 DOI: 10.1093/ajcn/84.1.18]
 - 110 **Adams JS**, Lee G. Gains in bone mineral density with resolution of vitamin D intoxication. *Ann Intern Med* 1997; **127**: 203-206 [PMID: 9245225 DOI: 10.7326/0003-4819-127-3-199708010-00004]
 - 111 **Aloia JF**, Dhaliwal R, Shieh A, Mikhail M, Fazzari M, Ragolia L, Abrams SA. Vitamin D supplementation increases calcium absorption without a threshold effect. *Am J Clin Nutr* 2014; **99**: 624-631 [PMID: 24335055 DOI: 10.3945/ajcn.113.067199]
 - 112 **Huang J**, Ou HY, Karnchanasorn R, Chiu KC. Clinical implication of vitamin D threshold. *Am J Clin Nutr* 2014; **100**: 295-296 [PMID: 24951577 DOI: 10.3945/ajcn.114.087171]
 - 113 **Ou HY**, Karnchanasorn R, Lee LZ, Chiu KC. Interaction of BMI with vitamin D and insulin sensitivity. *Eur J Clin Invest* 2011; **41**: 1195-1201 [PMID: 21434896 DOI: 10.1111/j.1365-2362.2011.02525.x]
 - 114 **Heaney RP**. Nutrition and chronic disease. *Mayo Clin Proc* 2006; **81**: 297-299 [PMID: 16529131 DOI: 10.4065/81.3.297]
 - 115 **Ilahi M**, Armas LA, Heaney RP. Pharmacokinetics of a single, large dose of cholecalciferol. *Am J Clin Nutr* 2008; **87**: 688-691 [PMID: 18326608 DOI: 10.1093/ajcn/87.3.688]
 - 116 **Ghazi AA**, Hosseini F, Ardakani E, Ghazi S, Hedayati M, Azizi F. Effects of different doses of oral cholecalciferol on serum 25(OH)D, PTH, calcium and bone markers during fall and winter in schoolchildren. *Eur J Clin Nutr* 2010; **64**: 1415-1422 [PMID: 20823895 DOI: 10.1038/ejcn.2010.169]
 - 117 **Carnes J**, Quinn S, Nelson M, Jones G, Winzenberg T. Intermittent high-dose vitamin D corrects vitamin D deficiency in adolescents: a pilot study. *Eur J Clin Nutr* 2012; **66**: 530-532 [PMID: 22190133 DOI: 10.1038/ejcn.2011.204]
 - 118 **Sanders KM**, Stuart AL, Williamson EJ, Simpson JA, Kotowicz MA, Young D, Nicholson GC. Annual high-dose oral vitamin D and falls and fractures in older women: a randomized controlled trial. *JAMA* 2010; **303**: 1815-1822 [PMID: 20460620 DOI: 10.1001/jama.2010.594]
 - 119 **Leventis P**, Kiely PD. The tolerability and biochemical effects of high-dose bolus vitamin D2 and D3 supplementation in patients with vitamin D insufficiency. *Scand J Rheumatol* 2009; **38**: 149-153 [PMID: 18991184 DOI: 10.1080/03009740802419081]
 - 120 **Rossini M**, Gatti D, Viapiana O, Fracassi E, Idolazzi L, Zanoni S, Adami S. Short-term effects on bone turnover markers of a single high dose of oral vitamin D₃. *J Clin Endocrinol Metab* 2012; **97**: E622-E626 [PMID: 22298802 DOI: 10.1210/jc.2011-2448]
 - 121 **von Restorff C**, Bischoff-Ferrari HA, Theiler R. High-dose oral vitamin D3 supplementation in rheumatology patients with severe vitamin D3 deficiency. *Bone* 2009; **45**: 747-749 [PMID: 19539796 DOI: 10.1016/j.bone.2009.06.012]
 - 122 **Tellioglu A**, Basaran S, Guzel R, Seydaoglu G. Efficacy and safety of high dose intramuscular or oral cholecalciferol in vitamin D deficient/insufficient elderly. *Maturitas* 2012; **72**: 332-338 [PMID: 22613271 DOI: 10.1016/j.maturitas.2012.04.011]
 - 123 **Witham MD**, Dove FJ, Sugden JA, Doney AS, Struthers AD. The effect of vitamin D replacement on markers of vascular health in stroke patients - a randomised controlled trial. *Nutr Metab Cardiovasc Dis* 2012; **22**: 864-870 [PMID: 21194910 DOI: 10.1016/j.numecd.2010.11.001]
 - 124 **Ish-Shalom S**, Segal E, Salganik T, Raz B, Bromberg IL, Vieth R. Comparison of daily, weekly, and monthly vitamin D3 in ethanol dosing protocols for two months in elderly hip fracture patients. *J Clin Endocrinol Metab* 2008; **93**: 3430-3435 [PMID: 18544622 DOI: 10.1210/jc.2008-0241]
 - 125 **Fu L**, Yun F, Oczak M, Wong BY, Vieth R, Cole DE. Common genetic variants of the vitamin D binding protein (DBP) predict differences in response of serum 25-hydroxyvitamin D [25(OH)D] to vitamin D supplementation. *Clin Biochem* 2009; **42**: 1174-1177 [PMID: 19302999 DOI: 10.1016/j.clinbiochem.2009.03.008]
 - 126 **Wang TJ**, Zhang F, Richards JB, Kestenbaum B, van Meurs JB, Berry D, Kiel DP, Streeten EA, Ohlsson C, Koller DL, Peltonen L, Cooper JD, O'Reilly PF, Houston DK, Glazer NL, Vandenput L, Peacock M, Shi J, Rivadeneira F, McCarthy MI, Anneli P, de Boer IH, Mangino M, Kato B, Smyth DJ, Booth SL, Jacques PF, Burke GL, Goodarzi M, Cheung CL, Wolf M, Rice K, Goltzman D, Hidioglou N, Ladouceur M, Wareham NJ, Hocking LJ, Hart D, Arden NK, Cooper C, Malik S, Fraser WD, Hartikainen AL, Zhai G, Macdonald HM, Forouhi NG, Loos RJ, Reid DM, Hakim A, Dennison E, Liu Y, Power C, Stevens HE, Jaana L, Vasani RS, Soranzo N, Bojunga J, Psaty BM, Lorentzon M, Forouhi T, Harris TB, Hofman A, Jansson JO, Cauley JA, Uitterlinden AG, Gibson Q, Jarvelin MR, Karasik D, Siscovick DS, Econs MJ, Kritchevsky SB, Florez JC, Todd JA, Dupuis J, Hyppönen E, Spector TD. Common genetic determinants of vitamin D insufficiency: a genome-wide association study. *Lancet* 2010; **376**: 180-188 [PMID: 20541252 DOI: 10.1016/S0140-6736(10)60588-0]
 - 127 **Hsu S**, Hoofnagle AN, Gupta DK, Gutierrez OM, Peralta CA, Shea S, Allen NB, Burke G, Michos ED, Ix JH, Siscovick D, Psaty BM, Watson KE, Kestenbaum B, de Boer IH, Robinson-Cohen C.

- Race, Ancestry, and Vitamin D Metabolism: The Multi-Ethnic Study of Atherosclerosis. *J Clin Endocrinol Metab* 2020; **105**: e4337-e4350 [PMID: [32869845](#) DOI: [10.1210/clinem/dgaa612](#)]
- 128 **Clemens TL**, Adams JS, Henderson SL, Holick MF. Increased skin pigment reduces the capacity of skin to synthesise vitamin D3. *Lancet* 1982; **1**: 74-76 [PMID: [6119494](#) DOI: [10.1016/s0140-6736\(82\)90214-8](#)]
- 129 **Webb AR**, Kline L, Holick MF. Influence of season and latitude on the cutaneous synthesis of vitamin D3: exposure to winter sunlight in Boston and Edmonton will not promote vitamin D3 synthesis in human skin. *J Clin Endocrinol Metab* 1988; **67**: 373-378 [PMID: [2839537](#) DOI: [10.1210/jcem-67-2-373](#)]
- 130 **Gangloff A**, Bergeron J, Lemieux I, Després JP. Changes in circulating vitamin D levels with loss of adipose tissue. *Curr Opin Clin Nutr Metab Care* 2016; **19**: 464-470 [PMID: [27537278](#) DOI: [10.1097/MCO.0000000000000315](#)]
- 131 **Holick MF**, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, Murad MH, Weaver CM; Endocrine Society. Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2011; **96**: 1911-1930 [PMID: [21646368](#) DOI: [10.1210/jc.2011-0385](#)]
- 132 **Vieth R**. Vitamin D supplementation, 25-hydroxyvitamin D concentrations, and safety. *Am J Clin Nutr* 1999; **69**: 842-856 [PMID: [10232622](#) DOI: [10.1093/ajcn/69.5.842](#)]
- 133 **Mulligan GB**, Licata A. Taking vitamin D with the largest meal improves absorption and results in higher serum levels of 25-hydroxyvitamin D. *J Bone Miner Res* 2010; **25**: 928-930 [PMID: [20200983](#) DOI: [10.1002/jbmr.67](#)]
- 134 **Dutta D**, Mondal SA, Choudhuri S, Maisnam I, Hasanooz Reza AH, Bhattacharya B, Chowdhury S, Mukhopadhyay S. Vitamin-D supplementation in prediabetes reduced progression to type 2 diabetes and was associated with decreased insulin resistance and systemic inflammation: an open label randomized prospective study from Eastern India. *Diabetes Res Clin Pract* 2014; **103**: e18-e23 [PMID: [24456991](#) DOI: [10.1016/j.diabres.2013.12.044](#)]
- 135 **Niroomand M**, Fotouhi A, Irannejad N, Hosseinpanah F. Does high-dose vitamin D supplementation impact insulin resistance and risk of development of diabetes in patients with pre-diabetes? *Diabetes Res Clin Pract* 2019; **148**: 1-9 [PMID: [30583032](#) DOI: [10.1016/j.diabres.2018.12.008](#)]
- 136 **Wagner H**, Alvarsson M, Mannheimer B, Degerblad M, Östenson CG. No Effect of High-Dose Vitamin D Treatment on β -Cell Function, Insulin Sensitivity, or Glucose Homeostasis in Subjects With Abnormal Glucose Tolerance: A Randomized Clinical Trial. *Diabetes Care* 2016; **39**: 345-352 [PMID: [26786573](#) DOI: [10.2337/dc15-1057](#)]
- 137 **Oosterwerff MM**, Eekhoff EM, Van Schoor NM, Boeke AJ, Nanayakkara P, Meijnen R, Knol DL, Kramer MH, Lips P. Effect of moderate-dose vitamin D supplementation on insulin sensitivity in vitamin D-deficient non-Western immigrants in the Netherlands: a randomized placebo-controlled trial. *Am J Clin Nutr* 2014; **100**: 152-160 [PMID: [24898240](#) DOI: [10.3945/ajcn.113.069260](#)]
- 138 **Pittas AG**, Dawson-Hughes B, Sheehan P, Ware JH, Knowler WC, Aroda VR, Brodsky I, Ceglia L, Chadha C, Chatterjee R, Desouza C, Dolor R, Foreyt J, Fuss P, Ghazi A, Hsia DS, Johnson KC, Kashyap SR, Kim S, LeBlanc ES, Lewis MR, Liao E, Neff LM, Nelson J, O'Neil P, Park J, Peters A, Phillips LS, Pratley R, Raskin P, Rasouli N, Robbins D, Rosen C, Vickery EM, Staten M; D2d Research Group. Vitamin D Supplementation and Prevention of Type 2 Diabetes. *N Engl J Med* 2019; **381**: 520-530 [PMID: [31173679](#) DOI: [10.1056/NEJMoa1900906](#)]
- 139 **Davidson MB**, Duran P, Lee ML, Friedman TC. High-dose vitamin D supplementation in people with prediabetes and hypovitaminosis D. *Diabetes Care* 2013; **36**: 260-266 [PMID: [23033239](#) DOI: [10.2337/dc12-1204](#)]
- 140 **Jorde R**, Sollid ST, Svartberg J, Schirmer H, Joakimsen RM, Njølstad I, Fuskevåg OM, Figenschau Y, Hutchinson MY. Vitamin D 20,000 IU per Week for Five Years Does Not Prevent Progression From Prediabetes to Diabetes. *J Clin Endocrinol Metab* 2016; **101**: 1647-1655 [PMID: [26829443](#) DOI: [10.1210/jc.2015-4013](#)]
- 141 **Barengolts E**, Manickam B, Eisenberg Y, Akbar A, Kukreja S, Ciubotaru I. Effect of high-dose vitamin D repletion on glycemic control in african-american males with prediabetes and hypovitaminosis d. *Endocr Pract* 2015; **21**: 604-612 [PMID: [25716637](#) DOI: [10.4158/EP14548.OR](#)]
- 142 **Jorde R**, Figenschau Y. Supplementation with cholecalciferol does not improve glycaemic control in diabetic subjects with normal serum 25-hydroxyvitamin D levels. *Eur J Nutr* 2009; **48**: 349-354 [PMID: [19370371](#) DOI: [10.1007/s00394-009-0020-3](#)]
- 143 **Jorde R**, Strand Hutchinson M, Kjærgaard M, Sneve M, Grimnes G. Supplementation with High Doses of Vitamin D to Subjects without Vitamin D Deficiency May Have Negative Effects: Pooled Data from Four Intervention Trials in Tromsø. *ISRN Endocrinol* 2013; **2013**: 348705 [PMID: [23577264](#) DOI: [10.1155/2013/348705](#)]
- 144 **Norman AW**, Frankel JB, Heldt AM, Grodsky GM. Vitamin D deficiency inhibits pancreatic secretion of insulin. *Science* 1980; **209**: 823-825 [PMID: [6250216](#) DOI: [10.1126/science.6250216](#)]
- 145 **Clark SA**, Stumpf WE, Sar M, DeLuca HF, Tanaka Y. Target cells for 1,25 dihydroxyvitamin D3 in the pancreas. *Cell Tissue Res* 1980; **209**: 515-520 [PMID: [6996826](#) DOI: [10.1007/BF00234764](#)]
- 146 **Clark SA**, Stumpf WE, Sar M. Effect of 1,25 dihydroxyvitamin D3 on insulin secretion. *Diabetes* 1981; **30**: 382-386 [PMID: [7014306](#) DOI: [10.2337/diab.30.5.382](#)]
- 147 **Chertow BS**, Sivitz WI, Baranetsky NG, Clark SA, Waite A, Deluca HF. Cellular mechanisms of insulin release: the effects of vitamin D deficiency and repletion on rat insulin secretion.

- Endocrinology* 1983; **113**: 1511-1518 [PMID: [6352248](#) DOI: [10.1210/endo-113-4-1511](#)]
- 148 **Kadowaki S**, Norman AW. Dietary vitamin D is essential for normal insulin secretion from the perfused rat pancreas. *J Clin Invest* 1984; **73**: 759-766 [PMID: [6323527](#) DOI: [10.1172/JCI111269](#)]
- 149 **Kadowaki S**, Norman AW. Time course study of insulin secretion after 1,25-dihydroxyvitamin D3 administration. *Endocrinology* 1985; **117**: 1765-1771 [PMID: [3899614](#) DOI: [10.1210/endo-117-5-1765](#)]
- 150 **Cade C**, Norman AW. Vitamin D3 improves impaired glucose tolerance and insulin secretion in the vitamin D-deficient rat in vivo. *Endocrinology* 1986; **119**: 84-90 [PMID: [3013599](#) DOI: [10.1210/endo-119-1-84](#)]
- 151 **Cade C**, Norman AW. Rapid normalization/stimulation by 1,25-dihydroxyvitamin D3 of insulin secretion and glucose tolerance in the vitamin D-deficient rat. *Endocrinology* 1987; **120**: 1490-1497 [PMID: [3549262](#) DOI: [10.1210/endo-120-4-1490](#)]
- 152 **Zeitz U**, Weber K, Soegiarto DW, Wolf E, Balling R, Erben RG. Impaired insulin secretory capacity in mice lacking a functional vitamin D receptor. *FASEB J* 2003; **17**: 509-511 [PMID: [12551842](#) DOI: [10.1096/fj.02-0424fje](#)]
- 153 **Pepaj M**, Bredahl MK, Gjerlaugsen N, Bornstedt ME, Thorsby PM. Discovery of novel vitamin D-regulated proteins in INS-1 cells: a proteomic approach. *Diabetes Metab Res Rev* 2015; **31**: 481-491 [PMID: [25449168](#) DOI: [10.1002/dmrr.2629](#)]
- 154 **Bornstedt ME**, Gjerlaugsen N, Olstad OK, Berg JP, Bredahl MK, Thorsby PM. Vitamin D metabolites influence expression of genes concerning cellular viability and function in insulin producing β -cells (INS1E). *Gene* 2020; **746**: 144649 [PMID: [32251702](#) DOI: [10.1016/j.gene.2020.144649](#)]
- 155 **Cheng Q**, Li YC, Boucher BJ, Leung PS. A novel role for vitamin D: modulation of expression and function of the local renin-angiotensin system in mouse pancreatic islets. *Diabetologia* 2011; **54**: 2077-2081 [PMID: [21424540](#) DOI: [10.1007/s00125-011-2100-1](#)]
- 156 **Lahbib A**, Ghodbane S, Louchami K, Sener A, Sakly M, Abdelmelek H. Effects of vitamin D on insulin secretion and glucose transporter GLUT2 under static magnetic field in rat. *Environ Sci Pollut Res Int* 2015; **22**: 18011-18016 [PMID: [26169817](#) DOI: [10.1007/s11356-015-4844-5](#)]
- 157 **Jayanarayanan S**, Anju TR, Smijin S, Paulose CS. Vitamin D3 supplementation increases insulin level by regulating altered IP3 and AMPA receptor expression in the pancreatic islets of streptozotocin-induced diabetic rat. *J Nutr Biochem* 2015; **26**: 1041-1049 [PMID: [26054778](#) DOI: [10.1016/j.jnutbio.2015.04.011](#)]
- 158 **Kuo T**, Damle M, González BJ, Egli D, Lazar MA, Accili D. Induction of α cell-restricted Gc in dedifferentiating β cells contributes to stress-induced β -cell dysfunction. *JCI Insight* 2019; **5**: e128351 [PMID: [31120862](#) DOI: [10.1172/jci.insight.128351](#)]
- 159 **He D**, Wang Y, Liu R, He A, Li S, Fu X, Zhou Z. 1,25(OH)₂D₃ Activates Autophagy to Protect against Oxidative Damage of INS-1 Pancreatic Beta Cells. *Biol Pharm Bull* 2019; **42**: 561-567 [PMID: [30930416](#) DOI: [10.1248/bpb.b18-00395](#)]
- 160 **Guareschi ZM**, Valcanaia AC, Ceglarek VM, Hotz P, Amaral BK, de Souza DW, de Souza TA, Nardelli T, Ferreira TR, Leite NC, Lubackzeuski C, de O Emilio HR, Grassioli S. The effect of chronic oral vitamin D supplementation on adiposity and insulin secretion in hypothalamic obese rats. *Br J Nutr* 2019; **121**: 1334-1344 [PMID: [30924427](#) DOI: [10.1017/S0007114519000667](#)]
- 161 **Kjalarsdottir L**, Tersey SA, Vishwanath M, Chuang JC, Posner BA, Mirmira RG, Repa JJ. 1,25-Dihydroxyvitamin D₃ enhances glucose-stimulated insulin secretion in mouse and human islets: a role for transcriptional regulation of voltage-gated calcium channels by the vitamin D receptor. *J Steroid Biochem Mol Biol* 2019; **185**: 17-26 [PMID: [30071248](#) DOI: [10.1016/j.jsbmb.2018.07.004](#)]
- 162 **Park S**, Kim DS, Kang S. Vitamin D deficiency impairs glucose-stimulated insulin secretion and increases insulin resistance by reducing PPAR- γ expression in nonobese Type 2 diabetic rats. *J Nutr Biochem* 2016; **27**: 257-265 [PMID: [26522682](#) DOI: [10.1016/j.jnutbio.2015.09.013](#)]
- 163 **Mazzone G**, Morisco C, Lembo V, D'Argenio G, D'Armiento M, Rossi A, Giudice CD, Trimarco B, Caporaso N, Morisco F. Dietary supplementation of vitamin D prevents the development of western diet-induced metabolic, hepatic and cardiovascular abnormalities in rats. *United European Gastroenterol J* 2018; **6**: 1056-1064 [PMID: [30228894](#) DOI: [10.1177/2050640618774140](#)]
- 164 **Dong B**, Zhou Y, Wang W, Scott J, Kim K, Sun Z, Guo Q, Lu Y, Gonzales NM, Wu H, Hartig SM, York RB, Yang F, Moore DD. Vitamin D Receptor Activation in Liver Macrophages Ameliorates Hepatic Inflammation, Steatosis, and Insulin Resistance in Mice. *Hepatology* 2020; **71**: 1559-1574 [PMID: [31506976](#) DOI: [10.1002/hep.30937](#)]
- 165 **Sadek KM**, Shaheen H. Biochemical efficacy of vitamin D in ameliorating endocrine and metabolic disorders in diabetic rats. *Pharm Biol* 2014; **52**: 591-596 [PMID: [24251869](#) DOI: [10.3109/13880209.2013.854812](#)]
- 166 **Sisley SR**, Arble DM, Chambers AP, Gutierrez-Aguilar R, He Y, Xu Y, Gardner D, Moore DD, Seeley RJ, Sandoval DA. Hypothalamic Vitamin D Improves Glucose Homeostasis and Reduces Weight. *Diabetes* 2016; **65**: 2732-2741 [PMID: [27217488](#) DOI: [10.2337/db16-0309](#)]
- 167 **Schnell DM**, Walton RG, Vekaria HJ, Sullivan PG, Bollinger LM, Peterson CA, Thomas DT. Vitamin D produces a perilipin 2-dependent increase in mitochondrial function in C2C12 myotubes. *J Nutr Biochem* 2019; **65**: 83-92 [PMID: [30658160](#) DOI: [10.1016/j.jnutbio.2018.11.002](#)]
- 168 **Jefferson GE**, Schnell DM, Thomas DT, Bollinger LM. Calcitriol concomitantly enhances insulin sensitivity and alters myocellular lipid partitioning in high fat-treated skeletal muscle cells. *J Physiol Biochem* 2017; **73**: 613-621 [PMID: [28980208](#) DOI: [10.1007/s13105-017-0595-8](#)]

- 169 **Hesse M**, Fröhlich LF, Zeitz U, Lanske B, Erben RG. Ablation of vitamin D signaling rescues bone, mineral, and glucose homeostasis in Fgf-23 deficient mice. *Matrix Biol* 2007; **26**: 75-84 [PMID: 17123805 DOI: [10.1016/j.matbio.2006.10.003](https://doi.org/10.1016/j.matbio.2006.10.003)]
- 170 **Boucher BJ**. Does vitamin D status contribute to caveolin-1-mediated insulin sensitivity in skeletal muscle? *Diabetologia* 2009; **52**: 2240 [PMID: 19672573 DOI: [10.1007/s00125-009-1478-5](https://doi.org/10.1007/s00125-009-1478-5)]
- 171 **Wu YY**, Yu T, Zhang XH, Liu YS, Li F, Wang YY, Gong P. 1,25(OH)2D3 inhibits the deleterious effects induced by high glucose on osteoblasts through undercarboxylated osteocalcin and insulin signaling. *J Steroid Biochem Mol Biol* 2012; **132**: 112-119 [PMID: 22595150 DOI: [10.1016/j.jsbmb.2012.05.002](https://doi.org/10.1016/j.jsbmb.2012.05.002)]
- 172 **Chen S**, Villalta SA, Agrawal DK. FOXO1 Mediates Vitamin D Deficiency-Induced Insulin Resistance in Skeletal Muscle. *J Bone Miner Res* 2016; **31**: 585-595 [PMID: 26462119 DOI: [10.1002/jbmr.2729](https://doi.org/10.1002/jbmr.2729)]
- 173 **Oh J**, Riek AE, Darwech I, Funai K, Shao J, Chin K, Sierra OL, Carmeliet G, Ostlund RE Jr, Bernal-Mizrachi C. Deletion of macrophage Vitamin D receptor promotes insulin resistance and monocyte cholesterol transport to accelerate atherosclerosis in mice. *Cell Rep* 2015; **10**: 1872-1886 [PMID: 25801026 DOI: [10.1016/j.celrep.2015.02.043](https://doi.org/10.1016/j.celrep.2015.02.043)]
- 174 **Ni W**, Glenn DJ, Gardner DG. Tie-2Cre mediated deletion of the vitamin D receptor gene leads to improved skeletal muscle insulin sensitivity and glucose tolerance. *J Steroid Biochem Mol Biol* 2016; **164**: 281-286 [PMID: 26369613 DOI: [10.1016/j.jsbmb.2015.09.017](https://doi.org/10.1016/j.jsbmb.2015.09.017)]
- 175 **Knowler WC**, Barrett-Connor E, Fowler SE, Hamman RF, Lachin JM, Walker EA, Nathan DM; Diabetes Prevention Program Research Group. Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. *N Engl J Med* 2002; **346**: 393-403 [PMID: 11832527 DOI: [10.1056/NEJMoa012512](https://doi.org/10.1056/NEJMoa012512)]
- 176 **Knowler WC**, Hamman RF, Edelstein SL, Barrett-Connor E, Ehrmann DA, Walker EA, Fowler SE, Nathan DM, Kahn SE; Diabetes Prevention Program Research Group. Prevention of type 2 diabetes with troglitazone in the Diabetes Prevention Program. *Diabetes* 2005; **54**: 1150-1156 [PMID: 15793255 DOI: [10.2337/diabetes.54.4.1150](https://doi.org/10.2337/diabetes.54.4.1150)]
- 177 **Karnchanasorn R**, Ou HY, Chiu KC. Proposed Guidelines for Future Vitamin D Studies. *JAMA Intern Med* 2016; **176**: 280-281 [PMID: 26830241 DOI: [10.1001/jamainternmed.2015.7974](https://doi.org/10.1001/jamainternmed.2015.7974)]
- 178 **Chiu KC**, Chu A, Go VL, Saad MF. Hypovitaminosis D is associated with insulin resistance and beta cell dysfunction. *Am J Clin Nutr* 2004; **79**: 820-825 [PMID: 15113720 DOI: [10.1093/ajcn/79.5.820](https://doi.org/10.1093/ajcn/79.5.820)]
- 179 **Karnchanasorn R**, Ou HY, Chiu KC. Plasma 25-hydroxyvitamin D levels are favorably associated with β -cell function. *Pancreas* 2012; **41**: 863-868 [PMID: 22258069 DOI: [10.1097/MPA.0b013e31823c947c](https://doi.org/10.1097/MPA.0b013e31823c947c)]



Published by **Baishideng Publishing Group Inc**
7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: <https://www.f6publishing.com/helpdesk>

<https://www.wjgnet.com>

