

Genome Sequence of *Rickettsia hoogstraalii*, a Geographically Widely Distributed Tick-Associated Bacterium

Erwin Sentausa, Khalid El Karkouri, Thi-Tien Nguyen, Aurélie Caputo, Didier Raoult, Pierre-Edouard Fournier

Aix-Marseille Université, URMITE, UM63, CNRS 7278, IRD 198, INSERM U1095, Faculté de Médecine, Marseille, France

***Rickettsia hoogstraalii* is a tick-associated member of the spotted fever group rickettsiae that is geographically widely distributed. We report here the draft genome of *R. hoogstraalii* strain Croatica^T (=DSM 22243 = UTMB 00003), which was isolated from *Haemaphysalis sulcata* ticks collected in Croatia.**

Received 30 September 2014 Accepted 2 October 2014 Published 6 November 2014

Citation Sentausa E, El Karkouri K, Nguyen T-T, Caputo A, Raoult D, Fournier P-E. 2014. Genome sequence of *Rickettsia hoogstraalii*, a geographically widely distributed tick-associated bacterium. *Genome Announc.* 2(6):e01171-14. doi:10.1128/genomeA.01171-14.

Copyright © 2014 Sentausa et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Pierre-Edouard Fournier, pierre-edouard.fournier@univ-amu.fr.

The genus *Rickettsia* consists of Gram-negative obligate intracellular bacteria that are associated with arthropods. The spotted fever group of this genus is made of species causing tick-borne rickettsioses, which are among the oldest known vector-borne diseases. *Rickettsia hoogstraalii* is a spotted fever group of rickettsiae that was described in 2010 (1). This species is closely related to *Rickettsia felis*. Although its pathogenesis in vertebrate hosts is unknown, *R. hoogstraalii* causes a cytopathic effect in Vero, CCE3, and ISE6 cells. Originally isolated in 2006 from *Haemaphysalis sulcata* ticks from Croatia (2) and *Carios capensis* ticks from the United States (3), it has been detected in other tick species in different parts of the world, including Japan (4), Spain (5), Cyprus (6), Ethiopia (7), Turkey (8), and the western Indian Ocean (9). Here, we briefly describe the genome sequencing of *R. hoogstraalii* strain Croatica^T (= DSM 22243^T = UTMB 00003^T).

The genome was sequenced using MiSeq technology (Illumina, San Diego, CA, USA) with a mate-pair strategy. SPAdes-3.1.0 (10) was used to perform a *de novo* assembly of the reads, and the best assembly with a *k*-mer value of 127 was chosen for annotation. Potential coding sequences (CDSs) were predicted using AMI-Gene (11), and the assignment of protein functions was performed by searching against the RickBase (12), GenBank, and Pfam (13) databases using BLASTp (14), while ribosomal RNAs, tRNAs, and other RNAs were identified using BLASTn, tRNAscanSE version 1.21 (15), and RNAmmer 1.2 (16). Orthologous genes between the chromosomes of *R. hoogstraalii* and *R. felis* strain URRWXCal2 (GenBank accession no. NC_007109.1) were identified using OrthoMCL (17), with a BLASTp *E* value cutoff of 1×10^{-5} and the default MCL inflation parameter of 1.5.

The draft genome of *R. hoogstraalii* Croatica^T consists of two contigs of 1,444,049 nucleotides and 40,763 nucleotides, respectively, with an average genome coverage of 326-fold and a G+C content of 32.38%. The shorter contig is a putative plasmid with an identity match of 83% (35% coverage, *E* value 0.0) to Plasmid01 from *Rickettsia australis* strain Cutlack (accession no. CP003339.1) when aligned using BLASTn. The chromosome contains 1,824 CDSs and, like other rickettsiae, 3 noncontiguous

rRNAs (5S, 16S, and 23S rRNA), 33 tRNAs, and 3 other RNAs. In addition, the plasmid contains 70 CDSs but no RNAs.

Compared to the *R. felis* chromosome, several genes are lacking in *R. hoogstraalii*, including genes encoding a putative esterase and a putative hydrolase/acyltransferase of the α/β hydrolase superfamily, a toxin of a toxin-antitoxin system that contains a PIN domain for nucleic acid binding (*vapC2*), a site-specific DNA methylase, a superfamily I DNA and RNA helicase, a sugar kinase from the ribokinase family, three guanosine polyphosphate pyrophosphohydrolases/synthetases (*spoT5*, *spoT8*, and *spoT10*), the cell surface antigen Sca11, a major facilitator superfamily (MFS)-type permease (*proP*), an alkylated DNA repair protein, the DNA-damage-inducible protein J (*relB2*), the MnhF subunit of a multisubunit Na⁺/H⁺ antiporter, a glutamine amidotransferase-like protein, and a penicillin acylase.

Nucleotide sequence accession numbers. The genome and plasmid sequences have been deposited in DDBJ/EMBL/GenBank under accession numbers [CCXM01000001](https://www.ncbi.nlm.nih.gov/nuccore/CCXM01000001) and [CCXM01000002](https://www.ncbi.nlm.nih.gov/nuccore/CCXM01000002), respectively.

ACKNOWLEDGMENT

This research was funded by the Mediterranean-Infection Foundation.

REFERENCES

- Duh D, Punda-Polic V, Avsic-Zupanc T, Bouyer D, Walker DH, Popov VL, Jelovsek M, Gracner M, Trilar T, Bradaric N, Kurtti TJ, Strus J. 2010. *Rickettsia hoogstraalii* sp. nov., isolated from hard- and soft-bodied ticks. *Int. J. Syst. Evol. Microbiol.* 60:977–984. <http://dx.doi.org/10.1099/ijs.0.011049-0>.
- Duh D, Punda-Polić V, Trilar T, Petrovec M, Bradarić N, Avsic-Zupanc T. 2006. Molecular identification of *Rickettsia felis*-like bacteria in *Haemaphysalis sulcata* ticks collected from domestic animals in southern Croatia. *Ann. N. Y. Acad. Sci.* 1078:347–351. <http://dx.doi.org/10.1196/annals.1374.068>.
- Mattila JT, Burkhardt NY, Hutcheson HJ, Munderloh UG, Kurtti TJ. 2007. Isolation of cell lines and a rickettsial endosymbiont from the soft tick *Carios capensis* (Acari: Argasidae: Ornithodorinae). *J. Med. Entomol.* 44:1091–1101. [http://dx.doi.org/10.1603/0022-2585\(2007\)44\[1091:IOCLAA\]2.0.CO;2](http://dx.doi.org/10.1603/0022-2585(2007)44[1091:IOCLAA]2.0.CO;2).
- Kawabata H, Ando S, Kishimoto T, Kurane I, Takano A, Nogami S, Fujita H, Tsurumi M, Nakamura N, Sato F, Takahashi M, Ushijima Y,

- Fukunaga M, Watanabe H. 2006. First detection of *Rickettsia* in soft-bodied ticks associated with seabirds, Japan. *Microbiol. Immunol.* 50: 403–406. <http://dx.doi.org/10.1111/j.1348-0421.2006.tb03807.x>.
5. Márquez FJ. 2008. Spotted fever group *Rickettsia* in ticks from southeastern Spain natural parks. *Exp. Appl. Acarol.* 45:185–194. <http://dx.doi.org/10.1007/s10493-008-9181-7>.
 6. Chochlakis D, Ioannou I, Sandalakis V, Dimitriou T, Kassinis N, Papadopoulos B, Tselentis Y, Psaroulaki A. 2012. Spotted fever group *Rickettsiae* in ticks in Cyprus. *Microb. Ecol.* 63:314–323. <http://dx.doi.org/10.1007/s00248-011-9926-4>.
 7. Pader V, Nikitorowicz BJ, Abdissa A, Adamu H, Tolosa T, Gashaw A, Cutler RR, Cutler SJ. 2012. *Candidatus Rickettsia hoogstraalii* in Ethiopian *Argas persicus* ticks. *Ticks Tick Borne Dis.* 3:338–345. <http://dx.doi.org/10.1016/j.ttbdis.2012.10.021>.
 8. Orkun O, Karaer Z, Cakmak A, Nalbantoglu S. 2014. Spotted fever group rickettsiae in ticks in Turkey. *Ticks Tick Borne Dis.* 5:213–218. <http://dx.doi.org/10.1016/j.ttbdis.2012.11.018>.
 9. Dietrich M, Lebarbenchon C, Jaeger A, Le RC, Bastien M, Lagadec E, McCoy KD, Pascalis H, Le CM, Dellagi K, Tortosa P. 2014. *Rickettsia* spp. in seabird ticks from western Indian Ocean islands, 2011–2012. *Emerg. Infect. Dis.* 20:838–842. <http://dx.doi.org/10.3201/eid2005.131088>.
 10. Nurk S, Bankevich A, Antipov D, Gurevich A, Korobeynikov A, Lapidus A, Prjibelsky A, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, McLean J, Lasken R, Clingenpeel S, Woyke T, Tesler G, Alekseyev M, Pevzner P. 2013. Assembling genomes and mini-metagenomes from highly chimeric reads, p 158–170. *In* Deng M, Jiang R, Sun F, Zhang X (ed), *Research in computational molecular biology*. Springer Verlag, Berlin, Germany.
 11. Bocs S, Cruveiller S, Vallenet D, Nuel G, Médigue C. 2003. AMiGene: annotation of Microbial genes. *Nucleic Acids Res.* 31:3723–3726. <http://dx.doi.org/10.1093/nar/gkg590>.
 12. Blanc G, Ogata H, Robert C, Audic S, Suhre K, Vestris G, Claverie J-M, Raoult D. 2007. Reductive genome evolution from the mother of *Rickettsia*. *PLoS Genet.* 3:e14. <http://dx.doi.org/10.1371/journal.pgen.0030014>.
 13. Punta M, Coghill PC, Eberhardt RY, Mistry J, Tate J, Boursnell C, Pang N, Forslund K, Ceric G, Clements J, Heger A, Holm L, Sonnhammer ELL, Eddy SR, Bateman AFinn RD. 2012. The Pfam protein families database. *Nucleic Acids Res.* 40:D290–D301. <http://dx.doi.org/10.1093/nar/gkr1065>.
 14. Altschul SF, Madden TL, Schäffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ. 1997. Gapped BLAST and psi-blast: a new generation of protein database search programs. *Nucleic Acids Res.* 25:3389–3402. <http://dx.doi.org/10.1093/nar/25.17.3389>.
 15. Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res.* 25: 955–964. <http://dx.doi.org/10.1093/nar/25.5.0955>.
 16. Lagesen K, Hallin P, Rødland EA, Stærfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res.* 35:3100–3108. <http://dx.doi.org/10.1093/nar/gkm160>.
 17. Li L, Stoekert CJ, Roos DS. 2003. OrthoMCL: identification of ortholog groups for eukaryotic genomes. *Genome Res.* 13:2178–2189. <http://dx.doi.org/10.1101/gr.1224503>.