



# The Role of Metabolic Dysfunction in T-Cell Exhaustion During Chronic Viral Infection

### Kehong Zheng<sup>1</sup>, Xiaojun Zheng<sup>2\*</sup> and Wei Yang<sup>2,3\*</sup>

<sup>1</sup> Department of General Surgery, Zhujiang Hospital, Southern Medical University, Guangzhou, China, <sup>2</sup> Research Department of Medical Sciences, Guangdong Provincial People's Hospital, Guangdong Academy of Medical Sciences, Guangzhou, China, <sup>3</sup> Department of Pathology, School of Basic Medical Sciences, Southern Medical University, Guangzhou, China

### **OPEN ACCESS**

### Edited by:

Paul "Li-Hao" Huang, Fudan University, China

### Reviewed by:

Qiuju Han, Shandong University, China Martin J. Richer, Indiana University Bloomington, United States

#### \*Correspondence:

Wei Yang yanglab@smu.edu.cn Xiaojun Zheng Zhengxiaojun\_gz@163.com

#### Specialty section:

This article was submitted to T Cell Biology, a section of the journal Frontiers in Immunology

Received: 25 December 2021 Accepted: 07 March 2022 Published: 31 March 2022

#### Citation:

Zheng K, Zheng X and Yang W (2022) The Role of Metabolic Dysfunction in T-Cell Exhaustion During Chronic Viral Infection. Front. Immunol. 13:843242. doi: 10.3389/firmmu.2022.843242 T cells are important components of adaptive immunity that protect the host against invading pathogens during infection. Upon recognizing the activation signals, naïve and/or memory T cells will initiate clonal expansion, trigger differentiation into effector populations and traffic to the inflamed sites to eliminate pathogens. However, in chronic viral infections, such as those caused by human immunodeficiency virus (HIV), hepatitis B and C (HBV and HCV), T cells exhibit impaired function and become difficult to clear pathogens in a state known as T-cell exhaustion. The activation and function persistence of T cells demand for dynamic changes in cellular metabolism to meet their bioenergetic and biosynthetic demands, especially the augmentation of aerobic glycolysis, which not only provide efficient energy generation, but also fuel multiple biochemical intermediates that are essential for nucleotide, amino acid, fatty acid synthesis and mitochondria function. Changes in cellular metabolism also affect the function of effectors T cells through modifying epigenetic signatures. It is widely accepted that the dysfunction of T cell metabolism contributes greatly to T-cell exhaustion. Here, we reviewed recent findings on T cells metabolism under chronic viral infection, seeking to reveal the role of metabolic dysfunction played in T-cell exhaustion.

### Keywords: chronic viral infection, T-cell exhaustion, metabolism, PD-1, glycolysis

Abbreviations: HIV, human immunodeficiency virus; HBV, hepatitis B; HCV, hepatitis C; LCMV, lymphocytic choriomeningitis virus; PD-1, programmed death-1; CTLA-4, cytotoxic T lymphocyte antigen-4; YIM-3, immunoglobulin domain and mucin domain-containing protein 3; LAG-3, lymphocyte activation gene 3 protein; PD-L1, programmed death ligand-1; TCR, T-cell receptor; OXPHOS, oxidative phosphorylation; FAO, fatty acid oxidation; PI3K, phosphoinositide 3-kinase; AKT, protein kinase B; mTORC1, mechanistic target of rapamycin complex 1; TCA, tricarboxylic acid; HIF1 $\alpha$ , hypoxia-inducible factor 1-alpha; SREBP, sterol regulatory element binding proteins; IRF4, interferon regulatory factor 4; GLUT, glucose transporter; ROS, reactive oxygen species; IFN- $\gamma$ , interferon- $\gamma$ ; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; Treg, regulatory T; GS, glutamate synthase; RORs, Retinoic acid receptor-related orphan receptors; Acat, acetyl-CoA acetyltransferase; DCs, dendritic cells; IL, interleukin; miRNAs, microRNAs; PGC1 $\alpha$ , peroxisome proliferator-activated receptor gamma coactivator 1-alpha; ITIM, immunotyrosine inhibitory motif; ITSM, immunotyrosine switch motif; Blimp-1, PR domain zinc-finger protein 1.

1

# INTRODUCTION

T cells are crucial components in the pathogen-specific adaptive immune responses. Naïve and/or memory T cells can recognize the specific pathogen via T-cell receptors (TCRs) during infection, leading to their robust proliferation and differentiation into effector T cells, which can directly clear pathogens and modulating the local immune response by secreting cytokines (1-5). Following the activation of T cells, dynamic changes were initiated in cellular metabolism, which is known as metabolic reprogramming, to meet their increased energy and anabolic demands. The alteration of metabolism can directly influence the epigenetic status by regulating the activity of histone- and DNA- modifying enzymes, and the production of essential substrates for epigenetic modification (6-8). In addition, metabolic reprogramming can promote the generation of biochemical intermediates (9-13), as well as the biogenesis and oxidative phosphorylation (OXPHOS) of mitochondrial (14-16), which are essential for the proper function of effector T cells. Thus, metabolic reprogramming is fundamental to the proliferation, differentiation and function performance of T cells.

In the context of chronic viral infection, the continuous stimulation of specific pathogens makes effector T cells into a dysfunctional state, which is known as T-cell exhaustion (3, 17-19). The state of exhaustion is characterized by stepwise and progressive loss of T cell function, which is distinct from T cell anergy and senescence (17, 20-23). Although more and more researches start revealing the features of CD4<sup>+</sup> T cell exhaustion in chronic viral infection (24, 25), in-depth and comprehensive understanding of exhaustion in CD4<sup>+</sup> T cells are still demanded. In contrast to CD4<sup>+</sup> T cells, the features of CD8<sup>+</sup> T cell exhaustion have been much better characterized in chronic viral infection (24, 25). T-cell exhaustion is mainly due to the continuous overstimulation of pathogen, which results in increasing expression of immunosuppressive cytokines and receptor, as well as decreasing the number of help T cells (26). During the state of exhaustion, dynamic changes of metabolism were detected in effector T cell, such as the disturbance of glycolytic metabolism and the reverse of its dependence on fatty acid oxidation (FAO) and OXPHOS for energy generation (26, 27). It has been widely accepted that metabolic dysfunction is tightly associated with the development and progression of T-cell exhaustion (3, 24, 26, 28). In this review, we will focus on the metabolic dysfunction in T-cell exhaustion during chronic viral infection and seeking potential strategies to reverse T-cell exhaustion.

# THE ROLE OF METABOLIC REPROGRAMMING IN T CELL ACTIVATION AND FUNCTION

Naïve and memory T cells rely primarily on OXPHOS and FAO for energy supply to maintain survival and housekeeping functions (1, 28). Once recognizing of a specific antigen by T

cell receptor (TCR), immature T cells will rapidly initiate the activation of phosphoinositide 3-kinase (PI3K)/protein kinase B (AKT)/mechanistic target of rapamycin complex 1(mTORC1) signaling pathway (29), which can subsequently promote the activity of various transcriptional factor including MYC (30, 31), hypoxia-inducible factor 1-alpha (HIF1α) (32, 33), sterol regulatory element binding proteins (SREBPs) (13, 34) and interferon regulatory factor 4 (IRF4) (35, 36) to trigger the reprogramming of metabolism. Engagement of co-stimulatory receptors, such as CD28, is also crucial for the TCR-mediated activation of PI3K/AKT/mTORC1 axis by enhancing the recruitment of PI3K catalytic subunit p1108 (37, 38). It has been well acknowledged that metabolic reprogramming of T cells not only generates efficient energy, but also fuels multiple biochemical intermediates for nucleotide, amino acid, fatty acid synthesis and mitochondrial function (Figure 1), which are essential to T cell proliferation, differentiation and proper function performance (6, 8, 39).

The enhancement of aerobic glycolysis (also known as "Warburg Effect", a distinct form of cellular metabolism, which enable cells getting energies from converting glucose-derived pyruvate to lactate under normoxic conditions, rather than conventionally entering the TCA cycle in mitochondria) is an important events of metabolic reprogramming during CD4<sup>+</sup> and CD8<sup>+</sup> T cells activation and functioning (1, 6, 26, 40). TCR/ CD28-based activation of PI3K/AKT/mTORC1 signaling can promote the activity of transcriptional factors like MYC and HIF1 $\alpha$ , which directly increase the expression of glucose transporter (GLUT) family members and multiple glycolytic enzymes that are vital for facilitating and sustaining the T-cell glycolysis (41-45). Although glycolysis is a relatively inefficient process of ATP production, ATP production by glycolysis is almost 100 times faster than OXPHOS and can sufficiently satisfy the robust energy demand for T cell activation (40, 46). In addition to ATP generation, upregulation of glycolysis also significantly enhances the production of various intermediates including glucose-6-phosphate, glyceraldehyde-3-phosphate and 3-phosphoglcerate, which are crucial precursors for biomolecules synthesis including nucleic acids, lipids and amino acids (1, 6). Moreover, increased pyruvate production could prevent CD4<sup>+</sup> T cells from apoptosis through maintaining the mitochondrial membrane potential and reactive oxygen species (ROS) level (14). Despite the priority of converting pyruvate into lactate, abundant pyruvate in the cytoplasm produced by the augmentation of aerobic glycolysis can also enter mitochondria for precursor biosynthesis and ATP production, which would also promotes the activation and functions of T cells (1, 6, 42). Beyond the conversion in glycolytic pathways, the alteration of certain glycolytic enzymes during metabolic reprogramming can also regulate the functions of effector T cells through its nonglycolytic functions (16, 47). For example, both  $CD4^+$  and  $CD8^+$ T cells significantly reduce the expression and secretion of interferon- $\gamma$  (IFN- $\gamma$ ) in the absence of glucose, which attributes to the reduced binding of glyceraldehyde-3-phosphate dehydrogenase (GAPDH) to IFN- $\gamma$  mRNA (16, 48). The recruitment of enolase-1, a glycolytic enzyme, to the promoter



FIGURE 1 | Metabolic reprogramming of effector 1 cells in acute viral affection. Upon recognition of specific antigen via 1CH/CD28 receptor, 1 cells rapidly initiate the signaling of Pl3K/AKT/mTORC1 axis and upregulate the expression of its downstream factors including MYC, HIF1α, SREBPs and PGC1α, reprogramming the expression of metabolic transporters and enzymes. Increase expression of glucose transporter family members and glycolytic enzymes dramatically promote the activity of aerobic glycolysis in effector T cells, which not only provide efficient energy generation, but also fueling multi biochemical intermediates that are necessary for nucleotide, amino acid, fatty acid synthesis and mitochondrial function. The reprogramming of amino acid metabolism, especially the changes of glutamine, serine, arginine and leucine metabolism, plays an important role in the activation and functional maintenance of T cells. Augmentation of glutamine metabolism can promote the acetylation reactions or lipid synthesis by acting as an anaplerosis source in the TCA cycle. Increasing level of serine promotes the activity of nuclear acid biosynthesis in effector T cells by fueling the precursor for one-carbon metabolism. In addition, high level of leucine and arginine is required for the full activation of mTORC1 activity in both CD4<sup>+</sup> and CD8<sup>+</sup> T cells. TCR, T-cell receptor; PI3K, phosphoinositide 3-kinase; AKT, protein kinase B; mTORC1, mechanistic target of rapamycin complex 1; HIF1α, hypoxia-inducible factor 1-alpha; SREBPs, sterol regulatory element binding proteins; PGC1α, peroxisome proliferator-activated receptor gamma coactivator 1-alpha; TCA, tricarboxylic acid.

of FoxP3 is essential for the suppressive function of regulatory T (Treg) cells by regulating the transcription level of FoxP3 (47).

The metabolic reprogramming of amino acids, especially changes in glutamine, serine, arginine and leucine metabolism, is important for the activation and function of T cells (49–51). Glutamine is the most abundant amino acid in human body and is an essential nutrient for proliferation (31). Once the pathogen is recognized by TCR in T cells, glutamine uptake is increased by a CD28-dependent upregulation of glutamine transporters. Then, glutamine is further deaminated into glutamate by glutamate synthase (GS) in a MYC-dependent pathway (31, 49). Glutamate can be converted to  $\alpha$ -ketoglutarate, which enters the tricarboxylic acid (TCA) cycle as an anaplerosis source to meet the biosynthetic and energy demands for T cells activation, especially under a glucose-restricted condition (52). Glutamate

can also promote T cell activation by influencing the metabolism of other biomolecules, including nucleotide, fatty acids and amino acids (53). Beyond the fueling of glutamine for serine biosynthesis, serine uptake in both CD4<sup>+</sup> and CD8+ T cells are increased during activation. Serine is an important precursor for one-carbon metabolism and plays a crucial role in the proliferation of effector T cells (50). Activation of T cells can also increase the uptake of various neutral amino acids, especially leucine and arginine. Studies have demonstrated that glutamine, leucine and arginine are crucial for the full activation of mTORC1 activity in both CD4<sup>+</sup> and CD8<sup>+</sup> T cells (6, 11, 12, 54).

>Fatty acid oxidation (FAO) is a major energy acquisition approach for naïve and/or memory T cells. However, lipid metabolism shifts from catabolism to anabolism in activated T cells to fulfill the boost biosynthetic demands for clonal expansion and differentiation, as well as different T-cell functions (1, 6). TCRmediated activation of AKT/PI3K/mTORC1 signaling significantly promotes cholesterol and fatty acid synthesis by upregulating the transcription of many lipogenic enzymes in a sterol regulatory element-binding protein (SREBP)-dependent manner. Deletion of SREBP significantly reduces the clonal expansion and cytokines production of effector T cells during viral infection (13, 55). Studies have demonstrated that the augmentation of cholesterol generation is a hallmark of lipid metabolism during T cell activation and functions, by supporting the synthesis of cellular and organelle membrane (13, 55). Retinoic acid receptor-related orphan receptors (RORs)-induced downregulation of cholesterol-metabolizing enzymes dramatically inhibits the proliferation and survival of CD8<sup>+</sup> T cells (56). In addition, inhibition of acetyl-CoA acetyltransferase (Acat) 1 and 2, enzymes involving in cholesterol esterification and storage, can further promote the proliferation and functions of CD8+ T cells by increasing cholesterol levels (57). The enhanced glycolysis and glutamine metabolism in activated T cells also promote fatty acid synthesis by fueling glyceraldehyde-3phosphate and acetyl-CoA production respectively (1, 53). Notably, metabolic reprogramming of fatty acid synthesis is distinct in the subsets of CD4+ T cells, and tightly linked to their different cell fates (58, 59). Effector CD4<sup>+</sup> T cells appear to rely more on glucose-derived carbons for fatty acid synthesis, while regulatory T (Treg) cells call for exogenous fatty acid uptake. Moreover, high expression of acetyl-CoA carboxylase 1, a key enzyme in fatty acid synthesis, can drive the differentiation of CD4<sup>+</sup> T cells toward Treg cells, other than the Th17 phenotype both in vitro and in vivo (58, 60).

## CHRONIC VIRAL INFECTION AND T-CELL EXHAUSTION

Generally, adaptive immune responses can successfully clear the virus in acute infections by effector T cells. However, effector T cells fail to clear the pathogens in the context of particular viral infections, including those caused by lymphocytic choriomeningitis virus (LCMV) clone-13, human immunodeficiency virus (HIV), hepatitis B virus (HBV) and hepatitis C virus (HCV), and lead to chronic infection. Persistent infection progressively makes effector T cells progressively into a dysfunctional state that known as T-cell exhaustion, which is characterized by a sequential loss of effector cytokines, upregulation of co-inhibitory receptors, impaired proliferative ability and dysfunctional metabolism (17, 24, 26, 61). Although exhausted CD4<sup>+</sup> and CD8<sup>+</sup> T cells share some common features in chronic viral infection, their exhaustion profiles are distinct. Studies in chronic LCMV infection model revealed that CD4<sup>+</sup> T cells are not merely undergoing exhaustion, but also initiate another pattern of differentiation, which is distinct from exhausted CD8<sup>+</sup> T cells (62). Since exhausted T cells are heterogeneous, the precise features of exhaustion in CD4<sup>+</sup> and CD8<sup>+</sup> T cells are not comprehensively defined, "T-cell exhaustion" is still a broad term for a state which the effector T cells are dysfunction caused by chronic viral infection (63). Following intensive studies will give a better understanding of T-cell exhaustion.

The T-cell exhaustion caused by chronic viral infection is mainly driven by continuous overstimulation of TCR signaling and the subsequently increased expression of many inhibitory and co-inhibitory receptors, including programmed death-1 (PD-1), cytotoxic T lymphocyte antigen-4 (CTLA-4), immunoglobulin domain and mucin domain-containing protein 3 (TIM-3), 2B4 (CD244), lymphocyte activation gene 3 protein (LAG-3) and CD160 (3, 26, 61). The augmentation of PD-1/PD-L1 pathway can block the TCR/CD28 mediated PI3K signaling, dramatically inhibit glycolysis and glutamine utilization in effector T cells and revert its dependent on FAO and OXPHOS for energy generation, to trigger the T-cell exhaustion. Notably, the co-expression of multiple co-inhibitory receptors is essential for PD-1-induced Tcell exhaustion (3). Chronic viral infection can also indirectly induce T-cell exhaustion by altering the function of helper cells like CD4<sup>+</sup> T cells and dendritic cells (DCs) by secreting cytokines like interleukin (IL)-2 (64). Moreover, some chronic infections such as HBV may alter the metabolism of infected host cells, as well as uninfected adjacent cells. The metabolic alteration of the environment could also lead to the exhaustion of T cells by altering the available nutrients and certain microRNA (miRNAs) levels (26, 65, 66).

# METABOLIC DYSFUNCTION IN T-CELL EXHAUSTION

It has been well characterized previously that metabolic reprogramming is essential for the activation and function of effector T cells (26, 30, 31, 46). Under chronic viral infection, persistent stimulation of antigen leads to up-regulation of PD-1 and other co-inhibitory receptors in exhausted T cells. Recent studies have highlighted the crucial role of co-inhibitory receptors, especially PD-1, in T cell exhaustion by regulating the transcription of multiple metabolism associated transporters and enzymes, resulting in metabolic alteration of glucose, amino acids and fatty acids (Figure 2) (3, 26, 61). Of note, the decrease of glycolysis and mitochondrial respiration in T cells occurs at the early stage in chronic infection, even before T cell dysfunction occurs, suggesting that metabolic dysfunction is an early driver, rather than a consequence, of T cell exhaustion (27, 67, 68). Therefore, targeting metabolism might be an efficient strategy for revising T cell exhaustion in chronic viral infection.

Down-regulation of glycolysis is a key feature of metabolic alteration in exhausted T cells, which occurs at the early stage of T cell exhaustion. Studies with LCMV clone-13 infection model demonstrated that, upon the chronic stimulation of TCR signaling, effector T cells would increase the expression of multiple co-inhibitory receptors, including PD-1 and CTLA-4 (69). The cytoplasmic tail of PD-1 contains an immunotyrosine inhibitory motif (ITIM) and an immunotyrosine switch motif (ITSM), which can attenuate the downstream signal transduction of TCR by recruiting phosphatases, particularly SHP-2 (70, 71). PD-1 ligation would attenuate PI3K/AKT signaling and the activity of mTORC1, block glucose uptake through downregulating Glut1 expression, and inhibit glycolysis



**FIGURE 2** | Metabolic alteration of exhausted T cells in chronic viral infection. During chronic viral infection, continuous antigen stimulation *via* TCR signaling significantly increase the expression of multiple inhibitory and co-inhibitory receptors including PD-1, CTLA-4, TIM-3 and LAG-3, which subsequently block the signaling of PI3K/AKT/mTORC1 axis and decrease the expression of its downstream factor, inducing metabolic dysfunction in exhausted T cells. Elevated expression of Blimp-1 in exhausted T cells also induce metabolic dysfunction in exhausted T cells by repressing the expression of PGC1α. It is well accepted that the shifting from glycolysis to FAO for energy generation is an important trigger of T-cell exhaustion. In addition, augmentation of FAO in mitochondria can lead to mitochondrial depolarization, impairment of mitochondrial biosynthesis and production of intolerable level of ROS, which are tightly associated with the dysfunction in exhausted T cells. TCR, T-cell receptor; PI3K, phosphoinositide 3-kinase; AKT, protein kinase B; mTORC1, mechanistic target of rapamycin complex 1; PD-1, programmed death-1; CTLA-4, cytotoxic T lymphocyte antigen-4; TIM-3, immunoglobulin domain and mucin domain-containing protein 3; LAG-3, lymphocyte activation gene 3 protein; PGC1α, peroxisome proliferator-activated receptor gamma coactivator 1-alpha; FAO, fatty acid oxidation; ROS, reactive oxygen species.

by suppressing the expression of hexokinase 2 (HK2), a key enzyme of glycolysis catalyzing the conversion of glucose to glucose-6-phosphate (27, 68, 72). Moreover, CTLA-4, which is also upregulated in exhausted T cells with chronic infection, inhibits glycolysis by a similar mechanism as PD-1 (27, 68).

Augmentation of lipolysis and fatty acid oxidation is another key feature of metabolic alteration in exhausted T cells, but mitochondrial function is compromised. PD-1 ligation in T cells could promote the expression of carnitine palmitoyl transferase 1A (CPT1A), a key regulator of lipid metabolism, to promote FAO in mitochondria in the early stage of exhaustion. Inhibition of CPT1A can reduce mitochondrial respiration by about 50% in early exhausted T cells, suggesting early exhausted T cells switch the energy pathway to FAO when glycolysis is restricted (27, 68). In addition, enhanced expression of PD-1 in exhausted T cells can also promote mitochondrial depolarization by reducing the transcriptional expression of peroxisome proliferatoractivated receptor gamma coactivator 1-alpha (PGC1 $\alpha$ ), a key transcriptional regulator involved in the regulation of energy generation and mitochondrial biogenesis genes, and mitochondrial transcription factor A (mtTFA) (68, 73). Overexpression of PGC1 $\alpha$  can promote metabolic fitness by enhanced glycolysis and mitochondrial respiration, and reduce depolarized mitochondria the in early exhausted T cells (17, 27, 68, 73). Recent studies also revealed that continuous TCR stimulation under hypoxia promoted expression of Blimp-1, another transcriptional repressor of PGC1 $\alpha$ , which induced the exhaustion of  $\text{CD8}^{\scriptscriptstyle +}$  T cells by repressing PGC1 $\alpha\text{-dependent}$ mitochondrial reprogramming (74, 75). Despite little is known for the role cholesterol metabolism in chronic infection, it's well accepted that cholesterol metabolism is crucial in T cell function and exhaustion in cancer. Cholesterol accumulation can promote

the expression of co-inhibitory receptors, like PD-1, by increasing endoplasmic reticulum (ER) stress, subsequently initiating the exhaustion of CD8<sup>+</sup> T cell in tumor microenvironment. Inhibiting ER stress sensor XBP1 or reducing cholesterol can effectively restore the function of effector T cells (76, 77). HCV infection induced a genotype-specific hypocholesterolemia through interfering with late cholesterol synthesis (78), suggesting altered cholesterol level of environment in HCV infection. HIV infection induced SREBP activation could induce TFII-I expression of CD4<sup>+</sup> T cells. Blocking SREBP by Oxysterols suppress HIV replication in CD4<sup>+</sup> T cells (79, 80). These results implicate that cholesterol metabolism may play a crucial role in T cell exhaustion during chronic viral infection, which deserves further study.

Recent study found that increasing expression of coinhibitory receptors also altered the amino acid metabolism in effector T cells. Increased expression of both PD-1 and CTLA-4 significantly reduced the transcriptional expression of key glutamine transporters, SNAT1 and SNAT2, which led to the inhibition of glutamine uptake and glutaminolysis in CD4<sup>+</sup> T cells (27). Moreover, PD-1 can also inhibit the utilization of the branched chain amino acids (BCAAs), including valine, isoleucine and leucine, which are important for the growth and function of effector T cells by being incorporated into proteins and/or metabolized for energy generation (27, 68).

In addition, a gene-expression profiles study found that a number of energy metabolism and citric-acid cycle associated genes were transcriptionally downregulated in exhausted CD8+ T cells, including Acas2l, Sdha, Adcy7, Pdha1, and Acadm, suggesting more metabolic pathway may involve in the energy depression in exhausted T cells during chronic infection (81). However, mechanisms that how these metabolism related genes involved in T cell exhaustion still require further investigation.

# CONCLUSION

T-cell exhaustion is a broad definition of dysfunctional effector T cells in chronic viral infection, since the features of, as well as the

### REFERENCES

- Bantug GR, Galluzzi L, Kroemer G, Hess C. The Spectrum of T Cell Metabolism in Health and Disease. *Nat Rev Immunol* (2018) 18:19–34. doi: 10.1038/nri.2017.99
- Masopust D, Schenkel JM. The Integration of T Cell Migration, Differentiation and Function. Nat Rev Immunol (2013) 13:309–20. doi: 10.1038/nri3442
- Saeidi A, Zandi K, Cheok YY, Saeidi H, Wong WF, Lee CYQ, et al. T-Cell Exhaustion in Chronic Infections: Reversing the State of Exhaustion and Reinvigorating Optimal Protective Immune Responses. *Front Immunol* (2018) 9:2569. doi: 10.3389/fimmu.2018.02569
- Speiser DE, Ho PC, Verdeil G. Regulatory Circuits of T Cell Function in Cancer. Nat Rev Immunol (2016) 16:599–611. doi: 10.1038/nri.2016.80
- 5. Suarez-Fueyo A, Bradley SJ, Tsokos GC. T Cells in Systemic Lupus Erythematosus. *Curr Opin Immunol* (2016) 43:32–8. doi: 10.1016/j.coi.2016.09.001
- Dimeloe S, Burgener AV, Grahlert J, Hess C. T-Cell Metabolism Governing Activation, Proliferation and Differentiation; a Modular View. *Immunology* (2017) 150:35–44. doi: 10.1111/imm.12655

pathways to, T-cell exhaustion are still not fully understood. Over the past decades, studies have demonstrated the importance of metabolic reprogramming in the activation and function of effector T cells. Although the characteristic of metabolic alterations in exhausted T cells is still not comprehensively investigated, recent studies highlighted the crucial roles of co-inhibitory receptors mediated in glycolytic and lipidic metabolic dysfunction during T-cell exhaustion, suggesting co-inhibitory receptor could be a promising target to improve anti-virus function of T cells. In addition to coinhibitory receptors, whether there are other mechanisms regulating T cell metabolism in chronic infection still deserves more researches. Since metabolic dysfunction in exhausted T cells is complicated and heterogeneous, it requires more studies to clarify the features, as well as the underlying mechanisms, of metabolic dysfunction in exhausted T cells, including the alterations of glucose, lipid and amino acid metabolism, aiming to provide new insights with potential and promising target in anti-virus immunotherapy.

# **AUTHOR CONTRIBUTIONS**

All authors listed have made a substantial, direct and intellectual contribution to the work, and approved it for publication.

# FUNDING

The work has also been funded by National Key R&D Program of China (No. 2018YFA0800404 to WY), National Natural Science Foundation of China (NSFC, No. 81822036 and 31770931 to WY, No. 82103399 to KZ, No. 31800730 to XZ), Guangdong Natural Science Funds for Distinguished Young Scholar (No. 2017A030306030 to WY),Guangdong Basic and Applied Basic Research Foundation (No. 2020A1515010212 to KZ, No. 2020A1515011246 to XZ), Science and Technology Program of Guangzhou (No. 202102020903 to KZ).

- Gray SM, Kaech SM, Staron MM. The Interface Between Transcriptional and Epigenetic Control of Effector and Memory CD8(+) T-Cell Differentiation. *Immunol Rev* (2014) 261:157–68. doi: 10.1111/imr.12205
- MacIver NJ, Michalek RD, Rathmell JC. Metabolic Regulation of T Lymphocytes. Annu Rev Immunol (2013) 31:259–83. doi: 10.1146/annurevimmunol-032712-095956
- Gubser PM, Bantug GR, Razik L, Fischer M, Dimeloe S, Hoenger G, et al. Rapid Effector Function of Memory CD8+ T Cells Requires an Immediate-Early Glycolytic Switch. *Nat Immunol* (2013) 14:1064–72. doi: 10.1038/ni.2687
- Procaccini C, Carbone F, Di Silvestre D, Brambilla F, De Rosa V, Galgani M, et al. The Proteomic Landscape of Human Ex Vivo Regulatory and Conventional T Cells Reveals Specific Metabolic Requirements. *Immunity* (2016) 44:406–21. doi: 10.1016/j.immuni.2016.01.028
- Long X, Ortiz-Vega S, Lin Y, Avruch J. Rheb Binding to Mammalian Target of Rapamycin (mTOR) Is Regulated by Amino Acid Sufficiency. J Biol Chem (2005) 280:23433–6. doi: 10.1074/jbc.C500169200
- 12. Sancak Y, Bar-Peled L, Zoncu R, Markhard AL, Nada S, Sabatini DM. Ragulator-Rag Complex Targets Mtorc1 to the Lysosomal Surface and Is

Necessary for Its Activation by Amino Acids. Cell (2010) 141:290–303. doi: 10.1016/j.cell.2010.02.024

- Kidani Y, Elsaesser H, Hock MB, Vergnes L, Williams KJ, Argus JP, et al. Sterol Regulatory Element-Binding Proteins Are Essential for the Metabolic Programming of Effector T Cells and Adaptive Immunity. *Nat Immunol* (2013) 14:489–99. doi: 10.1038/ni.2570
- Dimeloe S, Mehling M, Frick C, Loeliger J, Bantug GR, Sauder U, et al. The Immune-Metabolic Basis of Effector Memory CD4+ T Cell Function Under Hypoxic Conditions. J Immunol (2016) 196:106–14. doi: 10.4049/ jimmunol.1501766
- Gerriets VA, Kishton RJ, Nichols AG, Macintyre AN, Inoue M, Ilkayeva O, et al. Metabolic Programming and PDHK1 Control CD4+ T Cell Subsets and Inflammation. J Clin Invest (2015) 125:194–207. doi: 10.1172/JCI76012
- Chang CH, Curtis JD, Maggi LB Jr, Faubert B, Villarino AV, O'Sullivan D, et al. Posttranscriptional Control of T Cell Effector Function by Aerobic Glycolysis. *Cell* (2013) 153:1239–51. doi: 10.1016/j.cell.2013.05.016
- Wherry EJ. T Cell Exhaustion. Nat Immunol (2011) 12:492–9. doi: 10.1038/ ni.2035
- Schietinger A, Greenberg PD. Tolerance and Exhaustion: Defining Mechanisms of T Cell Dysfunction. *Trends Immunol* (2014) 35:51–60. doi: 10.1016/j.it.2013.10.001
- Tan A, Koh S, Bertoletti A. Immune Response in Hepatitis B Virus Infection. Cold Spring Harb Perspect Med (2015) 5:a021428. doi: 10.1101/cshperspect.a021428
- Kasakovski D, Xu L, Li Y. T Cell Senescence and CAR-T Cell Exhaustion in Hematological Malignancies. J Hematol Oncol (2018) 11:91. doi: 10.1186/ s13045-018-0629-x
- Akbar AN, Henson SM. Are Senescence and Exhaustion Intertwined or Unrelated Processes That Compromise Immunity? Nat Rev Immunol (2011) 11:289–95. doi: 10.1038/nri2959
- Brenchley JM, Karandikar NJ, Betts MR, Ambrozak DR, Hill BJ, Crotty LE, et al. Expression of CD57 Defines Replicative Senescence and Antigen-Induced Apoptotic Death of CD8+ T Cells. *Blood* (2003) 101:2711–20. doi: 10.1182/blood-2002-07-2103
- Dolfi DV, Mansfield KD, Polley AM, Doyle SA, Freeman GJ, Pircher H, et al. Increased T-Bet Is Associated With Senescence of Influenza Virus-Specific CD8 T Cells in Aged Humans. *J Leukoc Biol* (2013) 93:825–36. doi: 10.1189/ jlb.0912438
- 24. Yi JS, Cox MA, Zajac AJ. T-Cell Exhaustion: Characteristics, Causes and Conversion. *Immunology* (2010) 129:474–81. doi: 10.1111/j.1365-2567.2010.03255.x
- Han S, Asoyan A, Rabenstein H, Nakano N, Obst R. Role of Antigen Persistence and Dose for CD4+ T-Cell Exhaustion and Recovery. *Proc Natl Acad Sci USA* (2010) 107:20453–8. doi: 10.1073/pnas.1008437107
- Sears JD, Waldron KJ, Wei J, Chang CH. Targeting Metabolism to Reverse T-Cell Exhaustion in Chronic Viral Infections. *Immunology* (2021) 162:135–44. doi: 10.1111/imm.13238
- Patsoukis N, Bardhan K, Chatterjee P, Sari D, Liu B, Bell LN, et al. PD-1 Alters T-Cell Metabolic Reprogramming by Inhibiting Glycolysis and Promoting Lipolysis and Fatty Acid Oxidation. *Nat Commun* (2015) 6:6692. doi: 10.1038/ ncomms7692
- Pallett LJ, Schmidt N, Schurich A. T Cell Metabolism in Chronic Viral Infection. *Clin Exp Immunol* (2019) 197:143–52. doi: 10.1111/cei.13308
- Chi H. Regulation and Function of mTOR Signalling in T Cell Fate Decisions. Nat Rev Immunol (2012) 12:325–38. doi: 10.1038/nri3198
- Yecies JL, Manning BD. Transcriptional Control of Cellular Metabolism by mTOR Signaling. *Cancer Res* (2011) 71:2815–20. doi: 10.1158/0008-5472.CAN-10-4158
- Wang R, Dillon CP, Shi LZ, Milasta S, Carter R, Finkelstein D, et al. The Transcription Factor Myc Controls Metabolic Reprogramming Upon T Lymphocyte Activation. *Immunity* (2011) 35:871–82. doi: 10.1016/ j.immuni.2011.09.021
- 32. Shi LZ, Wang R, Huang G, Vogel P, Neale G, Green DR, et al. HIF1alpha-Dependent Glycolytic Pathway Orchestrates a Metabolic Checkpoint for the Differentiation of TH17 and Treg Cells. J Exp Med (2011) 208:1367–76. doi: 10.1084/jem.20110278
- Shehade H, Acolty V, Moser M, Oldenhove G. Cutting Edge: Hypoxia-Inducible Factor 1 Negatively Regulates Th1 Function. J Immunol (2015) 195:1372–6. doi: 10.4049/jimmunol.1402552

- Bensinger SJ, Bradley MN, Joseph SB, Zelcer N, Janssen EM, Hausner MA, et al. LXR Signaling Couples Sterol Metabolism to Proliferation in the Acquired Immune Response. *Cell* (2008) 134:97–111. doi: 10.1016/ j.cell.2008.04.052
- 35. Yao S, Buzo BF, Pham D, Jiang L, Taparowsky EJ, Kaplan MH, et al. Interferon Regulatory Factor 4 Sustains CD8(+) T Cell Expansion and Effector Differentiation. *Immunity* (2013) 39:833–45. doi: 10.1016/ j.immuni.2013.10.007
- 36. Man K, Miasari M, Shi W, Xin A, Henstridge DC, Preston S, et al. The Transcription Factor IRF4 Is Essential for TCR Affinity-Mediated Metabolic Programming and Clonal Expansion of T Cells. *Nat Immunol* (2013) 14:1155–65. doi: 10.1038/ni.2710
- Jacobs SR, Herman CE, Maciver NJ, Wofford JA, Wieman HL, Hammen JJ, et al. Glucose Uptake Is Limiting in T Cell Activation and Requires CD28-Mediated Akt-Dependent and Independent Pathways. *J Immunol* (2008) 180:4476–86. doi: 10.4049/jimmunol.180.7.4476
- Frauwirth KA, Riley JL, Harris MH, Parry RV, Rathmell JC, Plas DR, et al. The CD28 Signaling Pathway Regulates Glucose Metabolism. *Immunity* (2002) 16:769–77. doi: 10.1016/S1074-7613(02)00323-0
- Chapman NM, Boothby MR, Chi H. Metabolic Coordination of T Cell Quiescence and Activation. *Nat Rev Immunol* (2020) 20:55–70. doi: 10.1038/s41577-019-0203-y
- Reina-Campos M, Scharping NE, Goldrath AW. CD8(+) T Cell Metabolism in Infection and Cancer. Nat Rev Immunol (2021) 21:718–38. doi: 10.1038/ s41577-021-00537-8
- Hoffmann M, Pantazis N, Martin GE, Hickling S, Hurst J, Meyerowitz J, et al. Exhaustion of Activated CD8 T Cells Predicts Disease Progression in Primary HIV-1 Infection. *PloS Pathog* (2016) 12:e1005661. doi: 10.1371/ journal.ppat.1005661
- Menk AV, Scharping NE, Moreci RS, Zeng X, Guy C, Salvatore S, et al. Early TCR Signaling Induces Rapid Aerobic Glycolysis Enabling Distinct Acute T Cell Effector Functions. *Cell Rep* (2018) 22:1509–21. doi: 10.1016/ j.celrep.2018.01.040
- 43. Kunkl M, Sambucci M, Ruggieri S, Amormino C, Tortorella C, Gasperini C, et al. CD28 Autonomous Signaling Up-Regulates C-Myc Expression and Promotes Glycolysis Enabling Inflammatory T Cell Responses in Multiple Sclerosis. *Cells* (2019) 8:575. doi: 10.3390/cells8060575
- Zeng H, Chi H. mTOR Signaling and Transcriptional Regulation in T Lymphocytes. *Transcription* (2014) 5:e28263. doi: 10.4161/trns.28263
- Finlay DK, Rosenzweig E, Sinclair LV, Feijoo-Carnero C, Hukelmann JL, Rolf J, et al. PDK1 Regulation of mTOR and Hypoxia-Inducible Factor 1 Integrate Metabolism and Migration of CD8+ T Cells. J Exp Med (2012) 209:2441–53. doi: 10.1084/jem.20112607
- Kaech SM, Cui W. Transcriptional Control of Effector and Memory CD8+ T Cell Differentiation. Nat Rev Immunol (2012) 12:749–61. doi: 10.1038/nri3307
- De Rosa V, Galgani M, Porcellini A, Colamatteo A, Santopaolo M, Zuchegna C, et al. Glycolysis Controls the Induction of Human Regulatory T Cells by Modulating the Expression of FOXP3 Exon 2 Splicing Variants. *Nat Immunol* (2015) 16:1174–84. doi: 10.1038/ni.3269
- Cham CM, Gajewski TF. Glucose Availability Regulates IFN-Gamma Production and P70s6 Kinase Activation in CD8+ Effector T Cells. J Immunol (2005) 174:4670–7. doi: 10.4049/jimmunol.174.8.4670
- Sikalidis AK. Amino Acids and Immune Response: A Role for Cysteine, Glutamine, Phenylalanine, Tryptophan and Arginine in T-Cell Function and Cancer? *Pathol Oncol Res* (2015) 21:9–17. doi: 10.1007/s12253-014-9860-0
- Ma EH, Bantug G, Griss T, Condotta S, Johnson RM, Samborska B, et al. Serine Is an Essential Metabolite for Effector T Cell Expansion. *Cell Metab* (2017) 25:345–57. doi: 10.1016/j.cmet.2016.12.011
- Angajala A, Lim S, Phillips JB, Kim JH, Yates C, You Z, et al. Diverse Roles of Mitochondria in Immune Responses: Novel Insights Into Immuno-Metabolism. Front Immunol (2018) 9:1605. doi: 10.3389/fimmu.2018.01605
- Campbell C, Marchildon F, Michaels AJ, Takemoto N, van der Veeken J, Schizas M, et al. FXR Mediates T Cell-Intrinsic Responses to Reduced Feeding During Infection. *Proc Natl Acad Sci USA* (2020) 117:33446–54. doi: 10.1073/ pnas.2020619117
- Altman BJ, Stine ZE, Dang CV. From Krebs to Clinic: Glutamine Metabolism to Cancer Therapy. Nat Rev Cancer (2016) 16:619–34. doi: 10.1038/ nrc.2016.71

- Shi H, Chapman NM, Wen J, Guy C, Long L, Dhungana Y, et al. Amino Acids License Kinase Mtorc1 Activity and Treg Cell Function *via* Small G Proteins Rag and Rheb. *Immunity* (2019) 51:1012–27.e7. doi: 10.1016/j.immuni.2019.10.001
- Fu X, Xu M, Zhang H, Li Y, Li Y, Zhang C. Staphylococcal Enterotoxin C2 Mutant-Directed Fatty Acid and Mitochondrial Energy Metabolic Programs Regulate CD8(+) T Cell Activation. J Immunol (2020) 205:2066–76. doi: 10.4049/jimmunol.2000538
- 56. Cai Z, Ishibashi T, Kozai M, Mita H, Wang S, Takada K, et al. ROR Agonist Hampers the Proliferation and Survival of Postactivated CD8(+) T Cells Through the Downregulation of Cholesterol Synthesis-Related Genes. *Immunol Cell Biol* (2021) 99:288–98. doi: 10.1111/imcb.12406
- 57. Yang W, Bai Y, Xiong Y, Zhang J, Chen S, Zheng X, et al. Potentiating the Antitumour Response of CD8(+) T Cells by Modulating Cholesterol Metabolism. *Nature* (2016) 531:651–5. doi: 10.1038/nature17412
- Lochner M, Berod L, Sparwasser T. Fatty Acid Metabolism in the Regulation of T Cell Function. *Trends Immunol* (2015) 36:81–91. doi: 10.1016/j.it.2014.12.005
- Michalek RD, Gerriets VA, Jacobs SR, Macintyre AN, MacIver NJ, Mason EF, et al. Cutting Edge: Distinct Glycolytic and Lipid Oxidative Metabolic Programs Are Essential for Effector and Regulatory CD4+ T Cell Subsets. *J Immunol* (2011) 186:3299–303. doi: 10.4049/jimmunol.1003613
- Berod L, Friedrich C, Nandan A, Freitag J, Hagemann S, Harmrolfs K, et al. *De* Novo Fatty Acid Synthesis Controls the Fate Between Regulatory T and T Helper 17 Cells. *Nat Med* (2014) 20:1327–33. doi: 10.1038/nm.3704
- Fenwick C, Joo V, Jacquier P, Noto A, Banga R, Perreau M, et al. T-Cell Exhaustion in HIV Infection. *Immunol Rev* (2019) 292:149–63. doi: 10.1111/ imr.12823
- Crawford A, Angelosanto JM, Kao C, Doering TA, Odorizzi PM, Barnett BE, et al. Molecular and Transcriptional Basis of CD4(+) T Cell Dysfunction During Chronic Infection. *Immunity* (2014) 40:289–302. doi: 10.1016/ j.immuni.2014.01.005
- Blank CU, Haining WN, Held W, Hogan PG, Kallies A, Lugli E, et al. Defining "T Cell Exhaustion'. Nat Rev Immunol (2019) 19:665–74. doi: 10.1038/s41577-019-0221-9
- Ross SH, Cantrell DA. Signaling and Function of Interleukin-2 in T Lymphocytes. Annu Rev Immunol (2018) 36:411–33. doi: 10.1146/annurevimmunol-042617-053352
- Masson JJ, Billings HW, Palmer CS. Metabolic Reprogramming During Hepatitis B Disease Progression Offers Novel Diagnostic and Therapeutic Opportunities. *Antivir Chem Chemother* (2017) 25:53–7. doi: 10.1177/ 2040206617701372
- 66. Teng CF, Hsieh WC, Wu HC, Lin YJ, Tsai HW, Huang W, et al. Hepatitis B Virus Pre-S2 Mutant Induces Aerobic Glycolysis Through Mammalian Target of Rapamycin Signal Cascade. *PloS One* (2015) 10:e0122373. doi: 10.1371/ journal.pone.0122373
- Staron MM, Gray SM, Marshall HD, Parish IA, Chen JH, Perry CJ, et al. The Transcription Factor FoxO1 Sustains Expression of the Inhibitory Receptor PD-1 and Survival of Antiviral CD8(+) T Cells During Chronic Infection. *Immunity* (2014) 41:802–14. doi: 10.1016/j.immuni.2014.10.013
- Bengsch B, Johnson AL, Kurachi M, Odorizzi PM, Pauken KE, Attanasio J, et al. Bioenergetic Insufficiencies Due to Metabolic Alterations Regulated by the Inhibitory Receptor PD-1 Are an Early Driver of CD8(+) T Cell Exhaustion. *Immunity* (2016) 45:358–73. doi: 10.1016/j.immuni.2016.07.008
- Keir ME, Butte MJ, Freeman GJ, Sharpe AH. PD-1 and Its Ligands in Tolerance and Immunity. Annu Rev Immunol (2008) 26:677–704. doi: 10.1146/annurev.immunol.26.021607.090331
- Okazaki T, Maeda A, Nishimura H, Kurosaki T, Honjo T. PD-1 Immunoreceptor Inhibits B Cell Receptor-Mediated Signaling by Recruiting Src Homology 2-Domain-Containing Tyrosine Phosphatase 2 to

Phosphotyrosine. Proc Natl Acad Sci USA (2001) 98:13866-71. doi: 10.1073/pnas.231486598

- Parry RV, Chemnitz JM, Frauwirth KA, Lanfranco AR, Braunstein I, Kobayashi SV, et al. CTLA-4 and PD-1 Receptors Inhibit T-Cell Activation by Distinct Mechanisms. *Mol Cell Biol* (2005) 25:9543–53. doi: 10.1128/ MCB.25.21.9543-9553.2005
- Schurich A, Pallett LJ, Jajbhay D, Wijngaarden J, Otano I, Gill US, et al. Distinct Metabolic Requirements of Exhausted and Functional Virus-Specific CD8 T Cells in the Same Host. *Cell Rep* (2016) 16:1243–52. doi: 10.1016/ j.celrep.2016.06.078
- Schank M, Zhao J, Wang L, Nguyen LNT, Cao D, Dang X, et al. Oxidative Stress Induces Mitochondrial Compromise in CD4 T Cells From Chronically HCV-Infected Individuals. *Front Immunol* (2021) 12:760707. doi: 10.3389/ fimmu.2021.760707
- 74. Scharping NE, Rivadeneira DB, Menk AV, Vignali PDA, Ford BR, Rittenhouse NL, et al. Mitochondrial Stress Induced by Continuous Stimulation Under Hypoxia Rapidly Drives T Cell Exhaustion. Nat Immunol (2021) 22:205–15. doi: 10.1038/s41590-020-00834-9
- 75. Shin H, Blackburn SD, Intlekofer AM, Kao C, Angelosanto JM, Reiner SL, et al. A Role for the Transcriptional Repressor Blimp-1 in CD8(+) T Cell Exhaustion During Chronic Viral Infection. *Immunity* (2009) 31:309–20. doi: 10.1016/j.immuni.2009.06.019
- Ma X, Bi E, Lu Y, Su P, Huang C, Liu L, et al. Cholesterol Induces CD8(+) T Cell Exhaustion in the Tumor Microenvironment. *Cell Metab* (2019) 30:143– 156 e5. doi: 10.1016/j.cmet.2019.04.002
- Ma X, Yi Q. Cholesterol Induces T Cell Exhaustion. Aging (Albany NY) (2019) 11:7334–5. doi: 10.18632/aging.102305
- Clark PJ, Thompson AJ, Vock DM, Kratz LE, Tolun AA, Muir AJ, et al. Hepatitis C Virus Selectively Perturbs the Distal Cholesterol Synthesis Pathway in a Genotype-Specific Manner. *Hepatology* (2012) 56:49–56. doi: 10.1002/hep.25631
- Taylor HE, Linde ME, Khatua AK, Popik W, Hildreth JE. Sterol Regulatory Element-Binding Protein 2 Couples HIV-1 Transcription to Cholesterol Homeostasis and T Cell Activation. J Virol (2011) 85:7699–709. doi: 10.1128/JVI.00337-11
- Moog C, Aubertin AM, Kirn A, Luu B. Oxysterols, But Not Cholesterol, Inhibit Human Immunodeficiency Virus Replication In Vitro. *Antivir Chem Chemother* (1998) 9:491–6. doi: 10.1177/095632029800900605
- Wherry EJ, Ha SJ, Kaech SM, Haining WN, Sarkar S, Kalia V, et al. Molecular Signature of CD8+ T Cell Exhaustion During Chronic Viral Infection. *Immunity* (2007) 27:670–84. doi: 10.1016/j.immuni.2007.09.006

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2022 Zheng, Zheng and Yang. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.1