


REVIEW ARTICLE

Microsatellite instability: A 2024 update

Hiroyuki Yamamoto^{1,2}  | Yoshiyuki Watanabe^{2,3} | Hiroyuki Arai⁴ | Kumiko Umemoto⁴ | Keisuke Tateishi² | Yu Sunakawa⁴

¹Department of Bioinformatics, St. Marianna University Graduate School of Medicine, Kawasaki, Japan

²Department of Gastroenterology, St. Marianna University School of Medicine, Kawasaki, Japan

³Department of Internal Medicine, Kawasaki Rinko General Hospital, Kawasaki, Japan

⁴Department of Clinical Oncology, St. Marianna University School of Medicine, Kawasaki, Japan

Correspondence

Hiroyuki Yamamoto, Department of Bioinformatics, St. Marianna University Graduate School of Medicine, 2-16-1 Sugao, Miyamae-ku, Kawasaki, Kanagawa 216-8511, Japan.

Email: h-yama@marianna-u.ac.jp

Abstract

Deficient mismatch repair (dMMR) results in microsatellite instability (MSI), a pronounced mutator phenotype. High-frequency MSI (MSI-H)/dMMR is gaining increasing interest as a biomarker for advanced cancer patients to determine their eligibility for immune checkpoint inhibitors (ICIs). Various methods based on next-generation sequencing (NGS) have been developed to assess the MSI status. Comprehensive genomic profiling (CGP) testing can precisely ascertain the MSI status as well as genomic alterations in a single NGS test. The MSI status can be also ascertained through the liquid biopsy-based CGP assays. MSI-H has thus been identified in various classes of tumors, resulting in a greater adoption of immunotherapy, which is hypothesized to be effective against malignancies that possess a substantial number of mutations and/or neoantigens. NGS-based studies have also characterized MSI-driven carcinogenesis, including significant rates of fusion kinases in colorectal cancers (CRCs) with MSI-H that are targets for therapeutic kinase inhibitors, particularly in *MLH1*-methylated CRCs with wild-type *KRAS/BRAF*. *NTRK* fusion is linked to the colorectal serrated neoplasia pathway. Recent advances in investigations of MSI-H malignancies have resulted in the development of novel diagnostic or therapeutic techniques, such as a synthetic lethal therapy that targets the Werner gene. DNA sensing in cancer cells is required for antitumor immunity induced by dMMR, opening up novel avenues and biomarkers for immunotherapy. Therefore, clinical relevance exists for analyses of MSI and MSI-H-associated genomic alterations in malignancy. In this article, we provide an update on MSI-driven carcinogenesis, with an emphasis on unique landscapes of diagnostic and immunotherapeutic strategies.

KEYWORDS

DNA mismatch repair, fusion kinases, immunotherapy, microsatellite instability, next-generation sequencing

1 | INTRODUCTION

Deficient mismatch repair (dMMR) results in microsatellite instability (MSI), a pronounced mutator phenotype (Figure 1). High-frequency MSI (MSI-H)/dMMR is gaining increasing interest as

a biomarker for advanced cancer patients to determine their eligibility for immune checkpoint inhibitors (ICIs). In this article, we provide an update on MSI-driven carcinogenesis, with an emphasis on unique landscapes of diagnostic and immunotherapeutic strategies.

This is an open access article under the terms of the [Creative Commons Attribution-NonCommercial-NoDerivs](https://creativecommons.org/licenses/by-nc-nd/4.0/) License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made.

© 2024 The Authors. *Cancer Science* published by John Wiley & Sons Australia, Ltd on behalf of Japanese Cancer Association.

validated tissue-based testing of MSI, particularly in patients whose cancers shed an abundance of ctDNA.¹⁴

Microsatellite instability status can be determined in the real world with FDA-approved liquid biopsy tests such as Foundation Medicine's FoundationOne Liquid CDx and Guardant360 CDx (Figure 2). ctDNA-based MSI testing with Guardant360 was greatly concordant with tissue-based MSI analysis, allowing for a more accurate assessment of MSI status in conjunction with CGP. This improves immunotherapy accessibility for patients diagnosed with advanced cancer who do not have a tissue-based CGP test.¹²

2.3 | Microsatellite instability prediction using artificial intelligence (AI) based on tumor histomorphology

High-frequency MSI/dMMR is gaining increasing interest as a biomarker for ICI eligibility in advanced malignancies. AI technology is being used to forecast MSI/dMMR status based on histomorphological traits on hematoxylin–eosin slides. A systematic review found that AI-based systems exhibited exceptional performance, as evidenced by the highest standard of 0.972 in colorectal cancer (CRC).¹⁵ The performance of AI-based systems was comparatively subpar as evidenced by the highest standards of 0.81 for gastric cancer (GC) and 0.82 for endometrial cancer. Cases identified as MSI-H by AI-based systems, nevertheless, require direct molecular confirmation via PCR-based testing or NGS due to substantial limitations.^{15,16}

3 | HIGH-FREQUENCY MSI-DRIVEN CARCINOGENESIS

3.1 | BRAF V600E mutations in CRC

In the setting of CpG island methylator phenotype–high (CIMP-H), the majority of sporadic MSI-H cases evolve by epigenetic silencing of *MLH1* via promoter hypermethylation^{17,18} (Figure 1). A minority of sporadic MSI-H tumors that did not have *MLH1* methylation had somatic mutations in DNA MMR genes, indicating an alternate pathway to DNA MMR inactivation (Figure 1). The latter malignancies were linked to *KRAS* mutations rather than *BRAF* mutations.

BRAF V600E mutations are strongly correlated with *MLH1* promoter-methylated sporadic CRCs with MSI-H¹⁹ (Figure 1). Therefore, Lynch syndrome (LS) can be essentially ruled out in patients who are positive for the *BRAF* V600E variants and/or *MLH1* promoter methylation.¹⁹ As a diagnostic process for LS, the Japanese Society for Cancer of the Colon and Rectum (JSCCR) guidelines recommend tests for the *BRAF* V600E variants in CRC cases with MSI-H or loss of *MLH1* and *PMS2* expression.¹⁹ After that, *MLH1* promoter methylation testing is recommended for those with *BRAF* V600E wild type.¹⁹

Moreover, CRCs with *BRAF* V600E mutations can be divided into two groups based on gene expression: *BRAF* mutant 1 (BM1) or *BRAF* mutant 2 (BM2)²⁰ (Figure 1). BM1 is distinguished by the activation

of the *KRAS/AKT* pathway, the dysregulation of mTOR/4EBP, the activation of epithelial to mesenchymal transition (EMT), and immune infiltration. In contrast, BM2 exhibits dysregulation of the cell cycle checkpoints. BM1 is additionally distinguished by elevated levels of phosphorylated AKT and 4EBP1, whereas BM2 exhibits decreased cyclin D1 levels and elevated CDK1 levels.

Among the 218 *BRAF*-mutant CRCs, 69 were categorized as BM1 and 149 as BM2. MSI-H enrichment in BM2 was marginally greater than in BM1 (43% vs. 65% in BM2). Four subtypes comprise the CRC Subtyping Consortium (CRCSC) subtype consensus: consensus molecular subtype (CMS) CMS1 (defined by immune patterns and MSI-H), CMS2 (CIN and activation of WNT), CMS3 (metabolic pattern), and CMS4 (EMT)²¹ (Figure 3). A minority of *BRAF*-mutant patients were identified in CMS4 (17%), CMS3 (5%), and CMS2 (2%), while CMS1 accounted for the overwhelming majority (70%) of *BRAF*-mutant patients. Interestingly, CMS1 *BRAF* mutants were found to be distributed in both BM1 and BM2, whereas all *BRAF*-mutated CMS4 were BM1. Additional research is required to elucidate the clinical significance of this *BRAF* classification, particularly with regard to diverse targeted therapy combinations. Individuals with the BM1 or CMS4 subtype, which are both associated with increased MAP3K8 expression, may benefit more from triplet therapy as opposed to doublet therapy.

3.2 | ARID1A deficiency and a CIMP

If *MLH1* is disrupted in the presence of CIMP-H, the tumor transforms into MSI-H; conversely, if *MLH1* is not silenced, the tumor progresses along the CIN pathway and transforms into CIMP-H MSS CIN malignancies.²² *BRAF* V600E mutations were nearly ubiquitous in CIMP-H ascending colon tumors but were absent in CIMP-H GCs. Moreover, *BRAF* V600E mutations were not found in a subset of CIMP-H CRCs with comparable DNA methylation signatures. Therefore, *BRAF* V600E mutation is probably one of the minor mechanisms by which CIMP is induced (Figure 1). On the contrary, CIMP-H could potentially create a favorable milieu for the *BRAF* V600E mutation by inhibiting pathways implicated in apoptosis and senescence induced by oncogenes.²³

CpG island methylator phenotype is thought to be caused by genetic abnormalities in the tricarboxylic acid cycle (TCA) cycle, that is, *IDH1* or *IDH2* mutations in glioblastoma multiforme (GBM) and acute myelogenous leukemia (AML).²⁴ TCA cycle gene mutations are uncommon in CIMP-H GCs and CRCs, in contrast to GBM and AML. Even in AML, *IDH* mutations can only explain roughly half of the instances of CIMP-H. The cause of CIMP-H cases in which *IDH* mutations are absent is still unknown.

ARID1A is a member of a core component of the SWI/SNF chromatin remodeling complexes. Regarding the relationship between *ARID1A* and MSI-H, *ARID1A* promotes MMR and reportedly recruits MSH2 to chromatin during DNA replication.²⁵ *ARID1A* mutation is linked to CIMP and EBV infection in GCs, as well as CRCs and uterine endometrial malignancies. Loss of function of *ARID1A* was

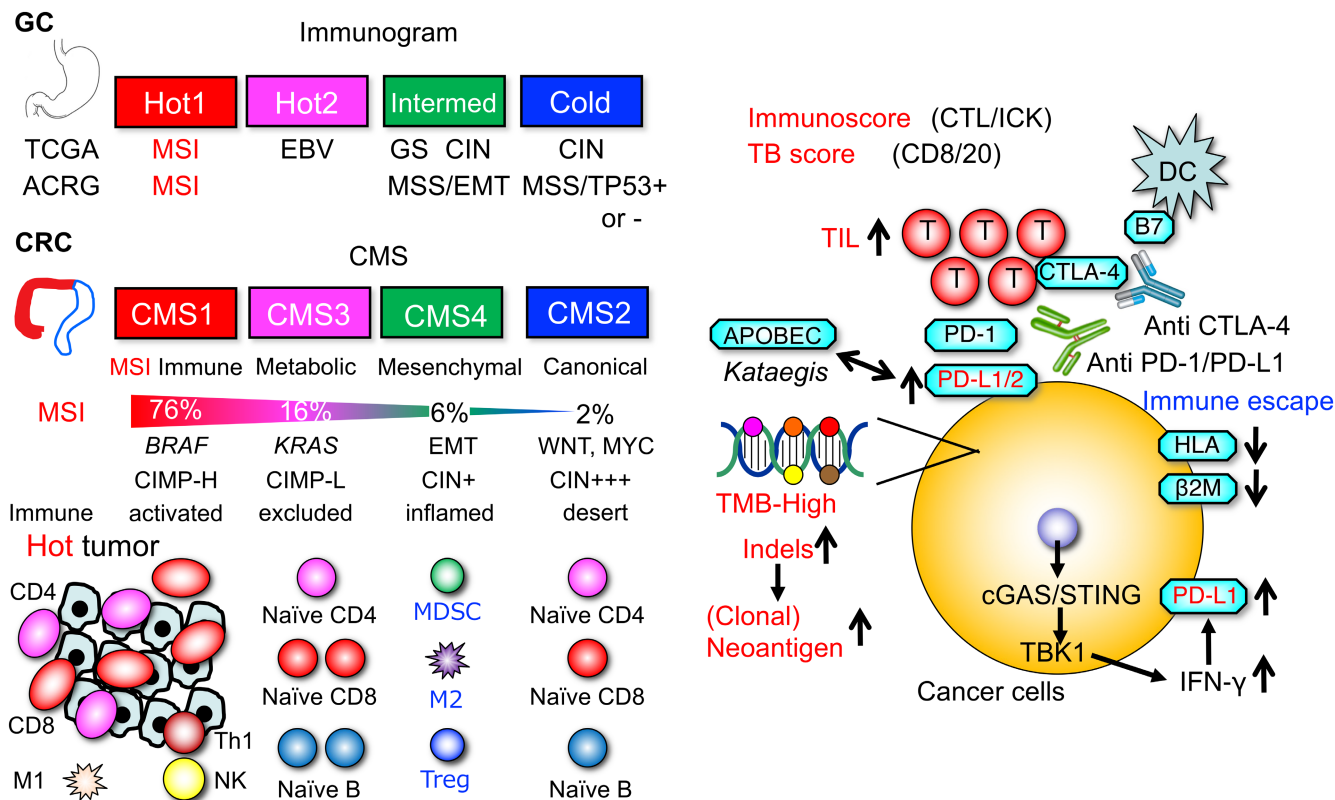


FIGURE 3 Immune microenvironment and immunotherapy in cancer. Upper left: classification of gastric cancer (GC) based on immunogram. ACRG, Asian Cancer Research Group; EMT, epithelial to mesenchymal transition; GS, genomically stable. Lower left: classification of colorectal cancer (CRC) based on CMS. Right: The immune response against high-microsatellite instability (MSI-H) cancers can be interfered with immune evasion mechanisms, such as inactivation of HLA and B2M. APOBEC3 alterations, APOBEC3 overexpression, and kataegis play roles in the overexpression of PD-L1/2. cGAS, cyclic GMP-AMP synthase; CTL/ICK, tumor-infiltrating T cell/immune checkpoint; DC, dendritic cell; TBK1, TANK binding kinase 1; TIL, tumor-infiltrating lymphocyte.

strongly associated with aberrant DNA methylation induction.²⁶ In 293FT cell lines and GES1 gastric epithelial cell lines, aberrant DNA methylation at genomic regions containing acquired and pre-existing H3K27me3 was induced by *ARID1A* knockout. Thus, aberrant DNA methylation induced by *ARID1A* mutation is one of the prospective mechanisms by which CIMP is induced (Figure 1).

3.3 | Frequent fusion kinases in *MLH1*-methylated CRCs with MSI-H

Colorectal cancers with *MLH1* promoter methylation (called MM) displayed clinicopathological characteristics different to CRCs with germline mutations (LS or LS-associated) or Lynch-like somatic MMR alterations (Figure 1). Frequent mutations in *BRAF/RNF43* as well as more insertions and deletions were found in *MLH1*-methylated sporadic MSI-H CRCs than in Lynch-like or LS-associated MSI-H cancers.²⁷ Importantly, 11 fusion kinases were only found in *MLH1*-methylated MSI-H CRCs with wild-type *KRAS/BRAF* and were linked to a short post-relapse survival²⁷ (Figure 1). Sato et al. created a simple approach for identifying *MLH1*-methylated cancers and tested it on a validation cohort of 28 CRCs with MSI-H, detecting 16 *MLH1*-methylated cancers

and two fusion kinases.²⁷ By employing the novel approach for identifying *MLH1*-methylated tumors, MSI-H CRC patients can be efficiently categorized as LS or sporadic cases that are potential candidates for fusion kinase carriers, which are therapeutic targets of kinase inhibitors.

Through the utilization of variant-specific PCR, 5'/3'-end expression imbalance, and RNA-based NGS, actionable gene rearrangements in malignancies with MSI were comprehensively analyzed.²⁸ A total of 58/471 (12.3%) CRCs, 4/69 (5.8%) GCs, and 3/65 (4.6%) endometrial malignancies exhibited gene fusions (8 *ALK*; 24 *NTRK1*; 2 *NTRK2*; 19 *NTRK3*; 12 *RET*). Gene rearrangements occurred most frequently in 53 (26%) of 204 CRCs with wild-type *KRAS/NRAS/BRAF*. Analysis of gene rearrangements of tyrosine kinase is especially viable in MSI-H CRCs with wild-type *KRAS/NRAS/BRAF*, although these events also occur to a moderate degree in other types of malignancies harboring MSI-H.

3.4 | *NTRK* fusions in the colorectal serrated neoplasia pathway

Fusions of *NTRK* gene are promising tissue-agnostic therapeutic targets in a variety of cancers. Immunohistochemical positivity

of TRK was found in 19% (11/58) of *MLH1*-methylated CRCs with MSI-H, 17% (4/23) of sessile serrated lesions with dysplasia, and 4% (5/132) of sessile serrated lesions but not in any of the hyperplastic polyps, traditional serrated adenomas, conventional adenomas, or MSS CRCs²⁹ (Figure 4). The 11 MSI-H CRCs with TRK positivity all had CIMP-H, methylated *MLH1*, wild-type *KRAS*/*BRAF*, and *NTRK1* or *NTRK3* fusion. Therefore, *NTRK*-rearranged CRCs exclusively develop via the colorectal serrated neoplasia pathway and can originate from *KRAS*/*BRAF* wild-type non-dysplastic SSLs before reaching complete development of CRCs with MSI-H/CIMP-H (Figure 4).

3.5 | Identification of Werner (WRN) in MSI-H tumors as a synthetic lethal therapeutic target

Four research groups independently determined, through the use of distinct functional screening techniques, that MSI-H malignancies rely on the WRN helicase to maintain genome stability and ensure cellular survival.^{30–34} Expanded TA-dinucleotide repeats were identified as prevalent sites of DNA breakage in MSI cancer cells when WRN was deficient.³⁵ These findings uncovered the mechanism underlying the synthetic lethality that exists between MSI and WRN deficiency and identified a novel biomarker for MSI (Figure 4).

The propensity of TA repeats to aggregate into cruciform structures elucidates the reason why downregulation of WRN results in preferential disruption of these DNA sequences in MSI cells. This provides a mechanistic foundation for the synthetic-lethal interaction involving TA repeats, which has been identified as a promising approach to precision cancer therapy.³⁶

In MSI CRC cells, WRN depletion significantly increased the expression of TP53 and PUMA, its apoptotic downstream target.³⁷ WRN depletion-induced apoptosis was eliminated by TP53 or PUMA deletion in MSI CRC cells. Notably, the *TP53* locus remains wild type in most MSI CRCs.³⁸ These findings suggest that TP53/

PUMA-mediated apoptosis plays a crucial role in susceptibility to WRN depletion for MSI CRC, and it endorses WRN as a viable treatment target for MSI CRCs with wild-type *TP53*.

4 | UNIQUE LANDSCAPES OF IMMUNOTHERAPEUTIC STRATEGIES

4.1 | Gastric cancer is classified immunologically using an immunogram

Because cancer-immune system interactions are complex, biomarker combinations will be needed to forecast each patient's response to ICI therapy and devise approaches to surmount resistance to therapy (Figure 3). A comprehensive framework called "Immunogram" has been proposed in order to encompass nearly all pertinent immunological variables (Figure 3). A method for converting transcriptomic data to immunogram scores was devised by Kobayashi et al.³⁹ Axis 1 (innate immunity), 2 (priming and activation), 3 (T cells), 4 (interferon gamma response), 5 (inhibitory molecules), 6 (inhibitory Treg cells), 7 (inhibitory myeloid-derived suppressor cells (MDSCs)), 8 (recognition of tumor cells), 9 (proliferation), and 10 (glycolysis) are the 10 molecular profiles comprising this immunogram.

Four subtypes of immune signatures of GCs were identified by immunogram analysis: Hot1, Hot2, Intermediate, and Cold (Figure 3). An impaired T cell signature was observed in Hot tumors, whereas an exclusion signature was noted in cold tumors.⁴⁰ The T cell dysfunction was more pronounced in Hot1 tumors as opposed to Hot2 tumors. Survival was superior in Hot2 subtypes in their cohort as well as TCGA cohort. Moreover, metabolic and cell adhesion pathways were distinct between Hot 1 and Hot 2 malignancies.⁴¹ Hence, it is imperative to incorporate active metabolism modulation into treatment strategies for Hot 1 tumors.

While these immunological subtypes and TCGA molecular subtypes shared some degree of overlap, histological or molecular

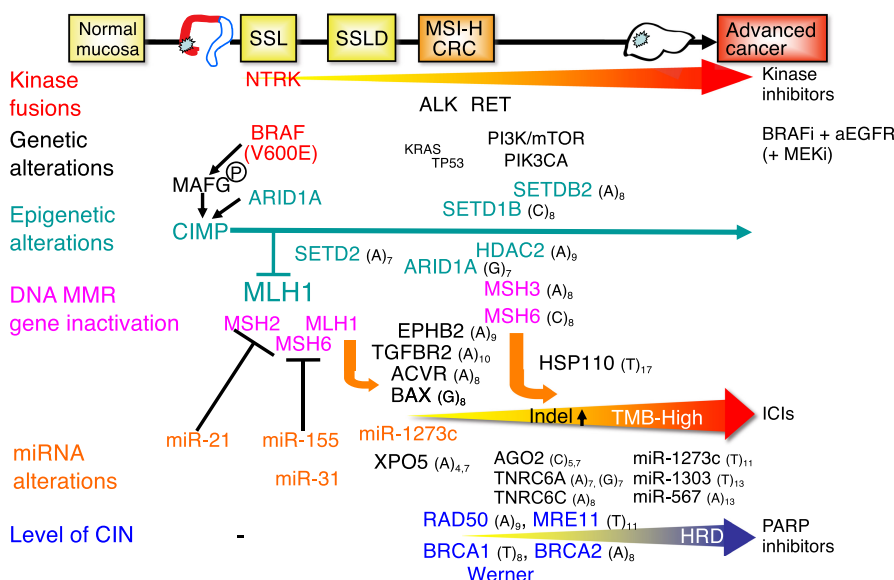


FIGURE 4 Carcinogenesis of sporadic high-microsatellite instability (MSI-H) colorectal cancers (CRCs). A model for MSI-H CRC progression is presented based on genetic, epigenetic, and miRNA alterations with target therapies. Mutations that promote cancer cell growth are considered to be the driving force of MSI-H carcinogenesis and are referred to as driver mutations or Real Common Target genes. SSL, sessile serrated lesions; SSLD, serrated lesions with dysplasia.

subtyping alone did not provide sufficient information to predict intratumoral immune responses in GC (Figure 3). Thus, molecular and immunological classifications complement each other in predicting ICI therapy responses and have the potential to be biomarkers for the treatment of GCs.

4.2 | Enhancement of mutability and ICI-induced antitumor immunity by ARID1A deficiency

ARID1A is a gene that experiences frequent mutations in cancer.^{42,43} *ARID1A* inactivation, on the other hand, impeded MMR and increased mutagenesis. A correlation was observed between *ARID1A* deficiency and the MSI-H genomic signature (preponderant C>T mutation pattern), and an increased burden of tumor mutations across a range of human cancer types. Thus, *ARID1A* deficiency relates to cancer's defective MMR (dMMR) and mutator phenotype, and it may interact favorably with ICI therapy (Figure 1).

4.3 | The accumulation of gene mutations results in decreased expression of HLA-ABC genes, which contributes to immune evasion in CRC

Understanding antitumor immunity is critical for effective cancer immune treatment (Figure 3). The accumulation of gene mutations leads to a decrease in HLA-ABC gene expression and could result in immune evasion in CRC (Figure 3). The impact of defective mutations in the *HLA-ABC* genes on tumor immunity was investigated by Kawazu et al.⁴⁴ A total of 114 MSI-H CRC samples were subjected to long-read sequencing to identify mutations in the *HLA-ABC* loci. There were 101 truncating mutations and 61 allele losses found in 57 and 21 tumors, respectively. Immunological subclassification also identified a subtype of tumor characterized by reduced lymphocyte infiltration, which was partially attributable to nongenetic alterations in the *HLA-ABC* genes (Figure 5). This group of tumors consistently showed the highest TMB, implying that the immunogenic effects of tumor mutations were mitigated by alterations that reduced immunoreactivity. Various genetic and epigenetic changes, including *RFX5* frameshift mutations and *PSMB8* promoter methylation, contributed to a lower HLA-ABC gene expression (Figure 5). Lastly, patients afflicted with these tumors had a shorter survival period than those with other forms of tumors. The insights gained from these results regarding the immune evasion mechanisms of MSI-H CRC contribute to the advancement and patient selection of cancer immunotherapy.

4.4 | The majority of beta 2 microglobulin (B2M)-deficient/mutant CRCs benefit clinically from ICI therapy and are MSI-H

As ICI therapy becomes standard for advanced MSI-H CRC, ICI response and resistance predictors have become increasingly

significant (Figure 3). In melanoma, loss of expression-associated *B2M* mutations have been linked to acquired immunotherapy resistance.⁴⁵ *B2M* mutations have also been linked to primary resistance in patients with immunotherapy-naïve CRC.^{46,47} Middha et al.⁴⁸ investigated the association of mutations of *B2M* with expression of *B2M* as well as MHC class I, tumor-infiltrating lymphocytes (TILs), molecular correlates, and immunotherapy response in CRC. *B2M* mutations were identified in 24% (44/182) of immunotherapy-naïve MSI-H CRCs; in 93% of these cases, *B2M* loss was observed, although loss of MHC class I was not typically observed. After controlling for MSI status, *B2M* mutations were not related to expression of MHC class I, TIL level, PD-L1 expression, or *KRAS* or *BRAF* mutation. Among the 13 patients having *B2M*-mutated CRC, partial response, stable disease, and pseudoprogression were observed in five, six, and one patient, respectively. Accordingly, the fact that 11 (85%) of 13 CRCs harboring *B2M* mutations exhibited some clinical benefit from ICIs indicates that *B2M* mutations do not serve as a predictor of primary ICI resistance in CRC. Despite loss of *B2M*, recognition of neoantigen was functional, as evidenced by abundance of TILs in *B2M*-defective MSI-H CRC. Thus, loss of *B2M* in CRC with MSI-H may not be enough to result in primary immunotherapy resistance. To determine whether the proportion and period of response differ by status of *B2M* mutations, larger, prospective studies are required.

4.5 | The common frameshift mutation landscape of MSI-H malignancies supports immunoediting in tumor progression

On cancer cells, the immune system is capable of identifying and eliminating cancer cells, particularly those that contain abundant mutation-induced neoantigens. Neoantigens of this nature are prevalent in MSI-H malignancies. Ballhausen et al. established a method for quantifying frameshift mutations in CRC and endometrial cancer with MSI-H.⁴⁹ Counterselection of cell clones having marked immunogenic frameshift peptides is suggested by the negative correlation between the frequency of frameshift mutations and the expected immunogenicity of the resulting peptides (Figure 3). The aforementioned correlation was not observed in tumors harboring *B2M* mutations. Conversely, coding microsatellite mutation patterns were found to be associated with HLA-A*02:01 status. Significantly, specific outlier mutations that were associated with frameshift peptides that have been functionally verified as immunogenic were prevalent in MSI-H malignancies. This suggests that these mutations may have played a driving role in the progression of MSI tumors.

Vaccine candidates that are nascent due to shared mutations hold great potential in the prevention of MSI-H malignancies⁴⁹ (Figure 6). A first phase I/IIa trial established the safety and immunological efficacy of a vaccine containing frameshift peptides (<https://clinicaltrials.gov/show/NCT01461148>). The landscapes of frameshift mutations in MSI-H tumors show that frameshift mutations that yield highly immunogenic neoantigens are being negatively selected. This demonstrates the viability of immunoediting in human nonviral

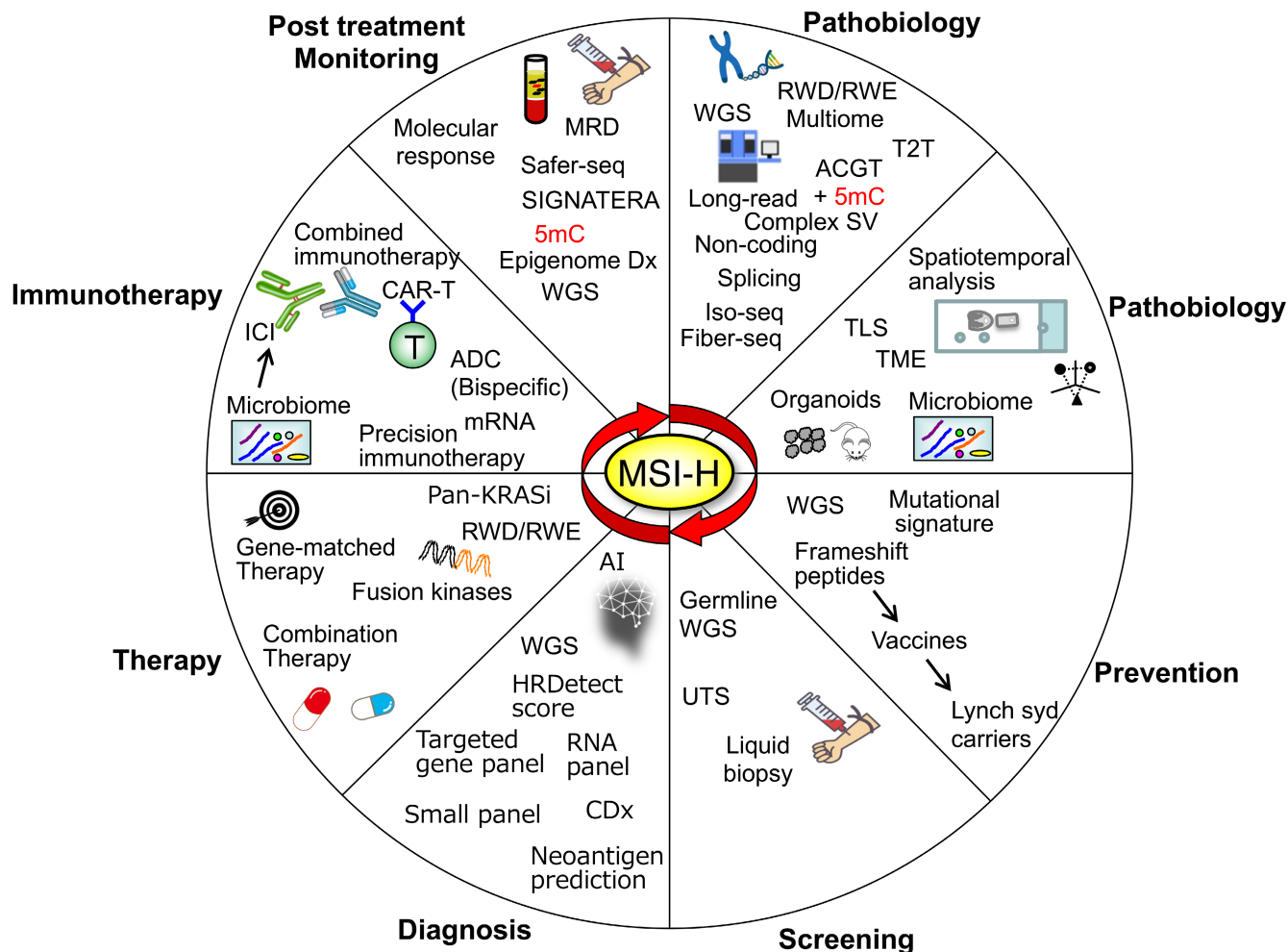


FIGURE 6 Overview of current and future pathobiology of high-microsatellite instability (MSI-H) and precision medicine. MSI-H-driven tumorigenesis, focusing on a novel landscape for diagnostic and therapeutic approach, is shown. Abbreviations in this figure are summarized with brief explanations in [Table 1](#).

MSI-H/dMMR tumors have been identified as notable indicators for responsiveness to ICIs. The presence of cancer-associated (clonal) neoantigens, dense immune cell infiltration, and a greater mutational burden are characteristics of these biomarkers that elicit a robust immune response. Pembrolizumab and nivolumab, with or without ipilimumab, are suggested therapeutic options for chemotherapy-resistant patients with metastatic MSI-H/dMMR CRCs. The FDA has granted full approval to pembrolizumab for the treatment of patients with unresectable/metastatic MSI-H/dMMR solid tumors that have advanced following prior treatment.

4.8 | Pembrolizumab as first-line treatment for patients with MSI-H/dMMR mCRC

For patients having mCRC with MSI-H/dMMR, the FDA approved pembrolizumab as the first-line therapy on the basis of the findings of the trial KEYNOTE-177, in which patients were assigned randomly to receive standard chemotherapy or pembrolizumab. Diaz

et al.⁵² recently reported the KEYNOTE-177 study's final overall survival analysis. While pembrolizumab demonstrated persistent antitumor activity and fewer treatment-related side events when compared with chemotherapy, there was no statistically significant distinction in overall survival between the two treatment cohorts. Pembrolizumab is supported as an efficacious first-line therapy in patients having mCRC with MSI-H/dMMR by these findings. Yoshino et al.'s⁵³ findings provide additional evidence in favor of pembrolizumab as first-line treatment for Asian patients having mCRC with MSI-H/dMMR.

5 | CONCLUSIONS

We provided an update on MSI-driven carcinogenesis, with an emphasis on unique landscapes of diagnostic and immunotherapeutic strategies. As shown in [Figure 6](#) and [Table 1](#), the current and future innovations in research of MSI-H pathogenesis will lead to the development of novel diagnostic, therapeutic, and preventive strategies in clinical settings.

TABLE 1 Abbreviations used in Figure 6 with brief explanations.

Abbreviations	Full spelling	Brief explanations
ADC	Antibody–drug conjugate	A substance made up of a monoclonal antibody chemically linked to a drug
CDx	Companion diagnostics	A medical device which provides information that is essential for the safe and effective use of a corresponding drug or biological product
Fiber-seq	Fiber sequencing	A method for analyzing regions with open chromatin that takes advantage of PacBio's ability to call methylated bases
HRDetect	Homologous recombination deficiency	A WGS-based classifier designed to predict BRCA1/2 deficiency based on six mutational signatures
Iso-seq	Iso-sequencing	Sequencing full-length cDNA using PacBio SMRT sequencing technology
MRD	Minimal residual disease	Cancer cells remaining after treatment that cannot be detected by those same scans or tests
RWD	Real-world data	Data relating to patient health status and/or the delivery of health care routinely collected from a variety of sources
RWE	Real-world evidence	The clinical evidence about the usage and potential benefits or risks of a medical product derived from analysis of RWD
Safer-seq	Safer-sequencing	A method for detecting rare mutations in blood in a highly efficient manner
SIGNATERA		A sensitive ctDNA test for colorectal cancer that can identify relapse sooner than standard tools
SV	Structural variation	A genetic alteration of more than 50 base pairs, including deletions, duplications, insertions, inversions, and translocations, as well as complex rearrangements
T2T	Telomere-to-telomere	The first complete, gapless sequence of a human genome
TLS	Tertiary lymphoid structures	Ectopic lymphocyte aggregates that form at sites of chronic inflammation, including cancers, in nonlymphoid tissues
TME	Tumor microenvironment	The normal cells, molecules, and blood vessels that surround and feed a tumor cell
UTS	Universal tumor screening	A strategy to identify high-risk individuals by testing all colorectal tumors for molecular features suggestive of Lynch syndrome
WGS	Whole genome sequencing	A comprehensive method for analyzing entire genomes

AUTHOR CONTRIBUTIONS

Hiroyuki Yamamoto: Conceptualization; visualization; writing – original draft; writing – review and editing. **Yoshiyuki Watanabe:** Data curation; visualization; writing – review and editing. **Hiroyuki Arai:** Data curation; visualization; writing – review and editing. **Kumiko Umemoto:** Data curation; visualization; writing – review and editing. **Keisuke Tateishi:** Supervision; writing – review and editing. **Yu Sunakawa:** Supervision; writing – review and editing.

ACKNOWLEDGMENTS

Not applicable.

FUNDING INFORMATION

This work was supported by JSPS KAKENHI Grant Number 22H02975.

CONFLICT OF INTEREST STATEMENT

The authors declare no conflict of interest.

ETHICS STATEMENTS

Approval of the research protocol by an Institutional Reviewer Board: N/A.

Informed Consent: N/A.

Registry and the Registration No. of the study/trial: N/A.

Animal Studies: N/A.

ORCID

Hiroyuki Yamamoto  <https://orcid.org/0000-0002-0954-6806>

REFERENCES

- Ruiz-Bañobre J, Goel A. DNA mismatch repair deficiency and immune checkpoint inhibitors in gastrointestinal cancers. *Gastroenterology*. 2019;156:890-903.
- Ganesh K, Stadler ZK, Cercek A, et al. Immunotherapy in colorectal cancer: rationale, challenges and potential. *Nat Rev Gastroenterol Hepatol*. 2019;16:361-375.
- Yamamoto H, Imai K. Microsatellite instability: an update. *Arch Toxicol*. 2015;89:899-921.
- Yamamoto H, Imai K. An updated review of microsatellite instability in the era of next-generation sequencing and precision medicine. *Semin Oncol*. 2019;46:261-270.
- Yamamoto H, Watanabe Y, Maehata T, Imai K, Itoh F. Microsatellite instability in cancer: a novel landscape for diagnostic and therapeutic approach. *Arch Toxicol*. 2020;94:3349-3357.
- Nakamura Y, Kawazoe A, Lordick F, Janjigian YY, Shitara K. Biomarker-targeted therapies for advanced-stage gastric and gastro-oesophageal junction cancers: an emerging paradigm. *Nat Rev Clin Oncol*. 2021;18:473-487.
- Foltz SM, Liang WW, Xie M, Ding L. MIRMMR: binary classification of microsatellite instability using methylation and mutations. *Bioinformatics*. 2017;33:3799-3801.

8. Wang C, Liang C. MSIPred: a python package for tumor microsatellite instability classification from tumor mutation annotation data using a support vector machine. *Sci Rep.* 2018;8:17546.
9. Hirotsu Y, Nagakubo Y, Amemiya K, Oyama T, Mochizuki H, Omata M. Microsatellite instability status is determined by targeted sequencing with MSICall in 25 cancer types. *Clin Chim Acta.* 2020;502:207-213.
10. Fujimoto A, Fujita M, Hasegawa T, et al. Comprehensive analysis of indels in whole-genome microsatellite regions and microsatellite instability across 21 cancer types. *Genome Res.* 2020;30:334-346.
11. Mayrhofer M, De Laere B, Whittington T, et al. Cell-free DNA profiling of metastatic prostate cancer reveals microsatellite instability, structural rearrangements and clonal hematopoiesis. *Genome Med.* 2018;10:85.
12. Willis J, Lefterova MI, Artyomenko A, et al. Validation of microsatellite instability detection using a comprehensive plasma-based genotyping panel. *Clin Cancer Res.* 2019;25:7035-7045.
13. Nakamura Y, Okamoto W, Denda T, et al. Clinical validity of plasma-based genotyping for microsatellite instability assessment in advanced GI cancers: SCRUM-Japan GOZILA substudy. *JCO Precis Oncol.* 2022;6:e2100383.
14. Nakamura Y, Shitara K. Development of circulating tumour DNA analysis for gastrointestinal cancers. *ESMO Open.* 2020;5:e000600.
15. Park JH, Kim EY, Luchini C, et al. Artificial intelligence for predicting microsatellite instability based on tumor histomorphology: a systematic review. *Int J Mol Sci.* 2022;23:2462.
16. Echle A, Ghaffari Laleh N, Quirke P, et al. Artificial intelligence for detection of microsatellite instability in colorectal cancer—a multicentric analysis of a pre-screening tool for clinical application. *ESMO Open.* 2022;7:100400.
17. Herman JG, Umar A, Polyak K, et al. Incidence and functional consequences of hMLH1 promoter hypermethylation in colorectal carcinoma. *Proc Natl Acad Sci USA.* 1998;95:6870-6875.
18. Leung SY, Yuen ST, Chung LP, Chu KM, Chan AS, Ho JC. hMLH1 promoter methylation and lack of hMLH1 expression in sporadic gastric carcinomas with high-frequency microsatellite instability. *Cancer Res.* 1999;59:159-164.
19. Tomita N, Ishida H, Tanakaya K, et al. Japanese Society for Cancer of the Colon and Rectum (JSCCR) guidelines 2020 for the clinical practice of hereditary colorectal cancer. *Int J Clin Oncol.* 2021;26:1353-1419.
20. Barras D, Missiaglia E, Wirapati P, et al. BRAF V600E mutant colorectal cancer subtypes based on gene expression. *Clin Cancer Res.* 2017;23:104-115.
21. Guinney J, Dienstmann R, Wang X, et al. The consensus molecular subtypes of colorectal cancer. *Nat Med.* 2015;21:1350-1356.
22. Fang M, Ou J, Hutchinson L, Green MR. The BRAF oncoprotein functions through the transcriptional repressor MAFK to mediate the CpG Island methylator phenotype. *Mol Cell.* 2014;55:904-915.
23. Hinoue T, Weisenberger DJ, Pan F, et al. Analysis of the association between CIMP and BRAF in colorectal cancer by DNA methylation profiling. *PLoS One.* 2009;4:e8357.
24. Kelly AD, Kroeger H, Yamazaki J, et al. A CpG island methylator phenotype in acute myeloid leukemia independent of IDH mutations and associated with a favorable outcome. *Leukemia.* 2017;31:2011-2019.
25. Shen J, Ju Z, Zhao W, et al. ARID1A deficiency promotes mutability and potentiates therapeutic antitumor immunity unleashed by immune checkpoint blockade. *Nat Med.* 2018;24:556-562.
26. Yamada H, Takeshima H, Fujiki R, et al. ARID1A loss-of-function induces CpG island methylator phenotype. *Cancer Lett.* 2022;532:215587.
27. Sato K, Kawazu M, Yamamoto Y, et al. Fusion kinases identified by genomic analyses of sporadic microsatellite instability-high colorectal cancers. *Clin Cancer Res.* 2019;25:378-389.
28. Mulikdjan RS, Saitova ES, Preobrazhenskaya EV, et al. ALK, ROS1, RET and NTRK1–3 gene fusions in colorectal and non-colorectal microsatellite-unstable cancers. *Int J Mol Sci.* 2023;24:13610.
29. Kim JH, Hong JH, Choi YL, et al. NTRK oncogenic fusions are exclusively associated with the serrated neoplasia pathway in the colorectum and begin to occur in sessile serrated lesions. *J Pathol.* 2021;255:399-411.
30. van Wietmarschen N, Nathan WJ, Nussenzweig A. The WRN helicase: resolving a new target in microsatellite unstable cancers. *Curr Opin Genet Dev.* 2021;71:34-38.
31. Chan EM, Shibue T, McFarland JM, et al. WRN helicase is a synthetic lethal target in microsatellite unstable cancers. *Nature.* 2019;568:551-556.
32. Lieb S, Blaha-Ostermann S, Kamper E, et al. Werner syndrome helicase is a selective vulnerability of microsatellite instability-high tumor cells. *elife.* 2019;8:e43333.
33. Kategaya L, Perumal SK, Hager JH, Belmont LD. Werner syndrome helicase is required for the survival of cancer cells with microsatellite instability. *iScience.* 2019;13:488-497.
34. Behan FM, Iorio F, Picco G, et al. Prioritization of cancer therapeutic targets using CRISPR-Cas9 screens. *Nature.* 2019;568:511-516.
35. van Wietmarschen N, Sridharan S, Nathan WJ, et al. Repeat expansions confer WRN dependence in microsatellite-unstable cancers. *Nature.* 2020;586:292-298.
36. Mengoli V, Ceppi I, Sanchez A, et al. WRN helicase and mismatch repair complexes independently and synergistically disrupt cruciform DNA structures. *EMBO J.* 2023;42:e111998.
37. Hao S, Tong J, Jha A, et al. Synthetic lethality of Werner helicase and mismatch repair deficiency is mediated by p53 and PUMA in colon cancer. *Proc Natl Acad Sci USA.* 2022;119:e2211775119.
38. Rampino N, Yamamoto H, Ionov Y, et al. Somatic frameshift mutations in the BAX gene in colon cancers of the microsatellite mutator phenotype. *Science.* 1997;275:967-969.
39. Kobayashi Y, Kushihara Y, Saito N, Yamaguchi S, Kakimi K. A novel scoring method based on RNA-Seq immunograms describing individual cancer-immunity interactions. *Cancer Sci.* 2020;111:4031-4040.
40. Sato Y, Wada I, Odaira K, et al. Integrative immunogenomic analysis of gastric cancer dictates novel immunological classification and the functional status of tumor-infiltrating cells. *Clin Transl Immunol.* 2020;9:e1194.
41. Saito N, Kobayashi Y, Nagaoka K, et al. Two distinct phenotypes of immunologically hot gastric cancer subtypes. *Biochem Biophys Res.* 2021;28:101167.
42. Wilson BG, Roberts CW. SWI/SNF nucleosome remodellers and cancer. *Nat Rev Cancer.* 2011;11:481-492.
43. Wu JN, Roberts CW. ARID1A mutations in cancer: another epigenetic tumor suppressor? *Cancer Discov.* 2013;3:35-43.
44. Kawazu M, Ueno T, Saeki K, et al. HLA class I analysis provides insight into the genetic and epigenetic background of immune evasion in colorectal cancer with high microsatellite instability. *Gastroenterology.* 2022;162:799-812.
45. Sade-Feldman M, Jiao YJ, Chen JH, et al. Resistance to checkpoint blockade therapy through inactivation of antigen presentation. *Nat Commun.* 2017;8:1136.
46. Janikovits J, Müller M, Krzykalla J, et al. High numbers of PDCD1(PD-1)-positive T cells and B2M mutations in microsatellite-unstable colorectal cancer. *Onco Targets Ther.* 2017;7:e1390640.
47. Grasso CS, Giannakis M, Wells DK, et al. Genetic mechanisms of immune evasion in colorectal cancer. *Cancer Discov.* 2018;8:730-749.
48. Middha S, Yaeger R, Shia J, et al. Majority of B2M-mutant and -deficient colorectal carcinomas achieve clinical benefit from immune checkpoint inhibitor therapy and are microsatellite instability-high. *JCO Precis Oncol.* 2019;3:PO.18.00321.
49. Ballhausen A, Przybilla MJ, Jendrusch M, et al. The shared frameshift mutation landscape of microsatellite-unstable cancers

- suggests immunoediting during tumor evolution. *Nat Commun.* 2020;11:4740.
50. Lu C, Guan J, Lu S, et al. DNA sensing in mismatch repair-deficient tumor cells is essential for anti-tumor immunity. *Cancer Cell.* 2021;39:96-108.
 51. Guan J, Lu C, Jin Q, et al. MLH1 deficiency-triggered DNA hyperexcision by exonuclease 1 activates the cGAS-STING pathway. *Cancer Cell.* 2021;39:109-121.
 52. Diaz LA Jr, Shiu KK, Kim TW, et al. Pembrolizumab versus chemotherapy for microsatellite instability-high or mismatch repair-deficient metastatic colorectal cancer (KEYNOTE-177): final analysis of a randomised, open-label, phase 3 study. *Lancet Oncol.* 2022;23:659-670.
 53. Yoshino T, Andre T, Kim TW, et al. Pembrolizumab in Asian patients with microsatellite-instability-high/mismatch-repair-deficient colorectal cancer. *Cancer Sci.* 2023;114:1026-1036.

How to cite this article: Yamamoto H, Watanabe Y, Arai H, Umemoto K, Tateishi K, Sunakawa Y. Microsatellite instability: A 2024 update. *Cancer Sci.* 2024;115:1738-1748. doi:[10.1111/cas.16160](https://doi.org/10.1111/cas.16160)