



Behavioural, Pharmacokinetic, Metabolic, and Hyperthermic Profile of 3,4-Methylenedioxypyrovalerone (MDPV) in the Wistar Rat

Rachel R. Horsley^{1*}, Eva Lhotkova¹, Katerina Hajkova^{1,2,3}, Barbara Feriancikova², Michal Himl⁴, Martin Kuchar^{1,2} and Tomas Páleníček^{1*}

¹ Department of Experimental Neurobiology, National Institute of Mental Health, Klecany, Czechia, ² Forensic Laboratory of Biologically Active Compounds, Department of Chemistry of Natural Compounds, University of Chemistry and Technology, Prague, Czechia, ³ Department of Analytical Chemistry, University of Chemistry and Technology, Prague, Czechia, ⁴ Department of Organic Chemistry, University of Chemistry and Technology, Prague, Czechia

OPEN ACCESS

Edited by:

Aviv M. Weinstein, Ariel University, Israel

Reviewed by:

Michael H. Baumann, National Institute on Drug Abuse (NIH), United States Carla Cannizzaro, Università degli Studi di Palermo, Italy

*Correspondence:

Rachel R. Horsley rachel.horsley@nudz.cz Tomas Páleníček tomas.palenicek@nudz.cz

Specialty section:

This article was submitted to Addictive Disorders, a section of the journal Frontiers in Psychiatry

Received: 15 September 2017 Accepted: 03 April 2018 Published: 24 April 2018

Citation:

Horsley RR, Lhotkova E, Hajkova K, Feriancikova B, Himl M, Kuchar M and Páleníček T (2018) Behavioural, Pharmacokinetic, Metabolic, and Hyperthermic Profile of 3,4-Methylenedioxypyrovalerone (MDPV) in the Wistar Rat. Front. Psychiatry 9:144. doi: 10.3389/fpsyt.2018.00144

3,4-methylenedioxypyrovalerone (MDPV) is a potent pyrovalerone cathinone that is substituted for amphetamines by recreational users. We report a comprehensive and detailed description of the effects of subcutaneous MDPV (1-4 mg/kg) on pharmacokinetics, biodistribution and metabolism, acute effects on thermoregulation under isolated and aggregated conditions, locomotion (open field) and sensory gating (prepulse inhibition, PPI). All studies used male Wistar rats. Pharmacokinetics after single dose of 2 mg/kg MDPV was measured over 6 h in serum, brain and lungs. The biotransformation study recorded 24 h urinary levels of MDPV and its metabolites after 4 mg/kg. The effect of 2 mg/kg and 4 mg/kg on body temperature (°C) was measured over 12 h in group- vs. individually-housed rats. In the open field, locomotion (cm) and its spatial distribution were assessed. In PPI, acoustic startle response (ASR), habituation, and PPI were measured (AVG amplitudes). In behavioural experiments, 1, 2, or 4 mg/kg MDPV was administered 15 or 60 min prior to testing. Thermoregulation and behavioural data were analysed using factorial analysis of variance (ANOVA). Peak concentrations of MDPV in sera, lung and brain tissue were reached in under 30 min. While negligible levels of metabolites were detected in tissues, the major metabolites in urine were demethylenyl-MDPV and demethylenyl-methyl-MDPV at levels three-four times higher than the parent drug. We also established a MDPV brain/serum ratio \sim 2 lasting for \sim 120 min, consistent with our behavioural observations of locomotor activation and disrupted spatial distribution of behaviour as well as moderate increases in body temperature (exacerbated in group-housed animals). Finally, 4 mg/kg induced stereotypy in the open field and transiently disrupted PPI. Our findings, along with previous research suggest that MDPV is rapidly absorbed, readily crosses the blood-brain barrier and is excreted primarily as metabolites. MDPV acts as a typical stimulant

1

with modest hyperthermic and psychomimetic properties, consistent with a primarily dopaminergic mechanism of action. Since no specific signs of acute toxicity were observed, even at the highest doses used, clinical care and harm-reduction guidance should be in line with that available for other stimulants and cathinones.

Keywords: 3 4-methylenedioxypyrovalerone, behaviour, hyperthermia, locomotion, MDPV, pharmacokinetics, sensory gating, wistar rat

INTRODUCTION

Background and Context

"Novel psychoactive substances" (NPSs) are synthetic compounds that produce similar subjective psychological and behavioural effects to pre-existing illicit recreational drugs, however, they may circumvent legal controls owing to differences in their chemical structures [1]. In recent years, NPSs have proliferated across the globe [2, 3] as a result of their perceived "legality," widespread availability [4] and of changes in socio-cultural attitudes where "sensible" recreational drug-taking has become increasingly normalised [5]. Unsurprisingly, rates of NPS dependence and toxic reactions/fatalities have risen simultaneously ([6, 7]) resulting in the increasing prominence of NPS (mis)use on public health agendas [8]. "Bath salts" is the colloquial street name for a class of NPSs where the principle psychoactive ingredient is a synthetic derivative of [9, 10]. Despite their common origin, cathinone derivatives have heterogeneous effects and mechanisms of action and it remains difficult to reliably predict the behavioural and physiological effects of NPSs based on their structural similarities [11]. Since controlled studies are not usually possible in humans, animal studies provide useful indicators of effects in vivo [12]. Studying effects (on an NPS-by-NPS basis) in animal models and in vitro continues to be a central approach oriented to obtaining knowledge with translational relevance to humans (e.g., harm reduction, clinical care). Here, we present data pertinent to further understanding of the acute pharmacokinetic, metabolic, behavioural, and thermoregulatory effects of the high-potency pyrovalerone cathinone 3,4-methylenedioxypyrovalerone (MDPV).

Mechanisms of Action

MDPV belongs to a sub-class of stimulating cathinones "pyrrolidinophenones" characterised by a complex chemical structure containing a pyrrolidine ring and an alkyl side-chain extending from the α -carbon [13]. Pyrrolidinophenones' stimulant effects reflect their inhibition of monoamine transporter sites where they are highly selective for the dopamine and norepinephrine transporter (DAT and NET), with particularly powerful effects on DAT, and only a negligible effect on the serotonin transporter (SERT) ([4, 14–20], but see [21]). To date, no evidence has been found (in rodents, or humans) that pyrrolidinophenones act as transporter substrate "releasers" [4, 14–16, 20] nor is there evidence that MDPV is active at non-transporter sites [16, 20]. Data from human embryonic kidney (HEK) cells expressing human transporters *in vitro* show a DAT/SERT inhibition ratio ranging from 100 to 300 [20, 22] and in rat brain synaptosomes, a ratio of 806 [18] for MDPV, values which far exceed those for other psychostimulants (e.g., amphetamine and cocaine), as well as most other pyrrolidinophenones [18, 20, 22]. MDPV is also much more effective at DAT than NET inhibition than other psychostimulants, e.g., for cocaine it is 50 times more potent at inhibiting DAT and ten times more potent at inhibiting NET [14, 18, 23, 24].

Pharmacokinetics and Metabolism

Recreational users report that the desirable subjective effects of MDPV include euphoria, sensory and physical stimulation [25, 26], similar to amphetamine (to which MDPV is chemically related; [1, 7, 27]). Recreational users typically ingest MDPV orally or by insufflation [4, 25]. Typical low-moderate active dosages for human recreational use range between 3 and 10 mg [25] with reported onset of subjective effects within 30 min and a duration of 120-210 min, with after-effects up to 8 h, [25, 28]. In vivo studies in rats that examined the concentrations of MDPV in plasma after subcutaneous (sc.) 0.5-3 mg/kg MDPV showed maximal plasma concentrations 10-20 min after administration, which then declined rapidly with an estimated elimination halflife of \sim 80 min [29, 30]. Only one previous study, however, has reported on the penetration of MDPV into the brain (after 1 mg/kg sc.), showing peak concentrations by 25 min in Sprague-Dawley rats [30].

In vitro studies of human liver microsomes have shown that MDPV is metabolised first to 3,4-dihydroxypyrovalerone (demethylenyl-MDPV) via O-demethylenation of its 3,4-methylenedioxy ring, and then to 4-hydroxy-3-(demethylenyl-methyl-MDPV) methoxypyrovalerone bv O-methylation; these phase II metabolites are excreted in urine in conjugated form ([31], but see [32]). More recently it has been shown, that in rats, concentrations of demethylenyl-MDPV and demethylenyl-methyl-MDPV in plasma peak more slowly than MDPV, at around 3-4h [29]. Demethylenyl-methyl-MDPV is the most abundant urinary metabolite in both species, humans and rats [32].

Toxicity and Thermoregulation

Human data on MDPV and pyrrolidinophenone toxicity is often confounded because "bath salts" commonly contain multiple cathinones and unknown contaminants [1, 7, 33]. However, since MDPV has been implicated more frequently in cases of serious intoxication [7, 34, 35] and confirmed in a number of fatalities [7, 36–39] than other cathinones, it may have higher risk of acute toxicity. Acute toxic effects of MDPV specifically involve over-stimulation of the cardiovascular system and the central nervous system (CNS), resulting in agitation, hyperthermia, and tachycardia [13, 26], and in one fatal case (where urine levels of MDPV were 670 ng/mL), coagulopathy, acidosis, rhabdomyolysis and anoxic brain injury were observed [38]. Psychiatric symptoms such as paranoia, delirium and hallucinations may be present, and serotonin syndrome, deterioration of muscle tissue and kidney failure have also been reported [26, 28, 37, 40–42]. Prominent in cathinone-related toxidrome (in humans) is hyperthermia; however, prior research on the effects of MDPV specifically on thermoregulation in rodents has been inconsistent. MDPV can elevate body temperature under conditions of higher ambient temperatures or social interaction, [21, 43, 44], however effects can be negligible, even under such conditions [45].

Behavioural Effects

MDPV is a highly reinforcing psychostimulant with notable addictive potential [20, 22, 46-51]. Increased extracellular dopamine (DA), particularly in the nucleus accumbens (NAcc), is an indicator of the reinforcement potential of drugs, and the accompanying locomotor response itself can be used as a marker of this. MDPV's stimulatory effects are most likely mediated by striatal extracellular DA ([29, 30], see [4]). MDPV has been shown to increase both locomotion in rodents ([14, 45, 52-55], see also [50], for a recent review) as well as increased NAcc DA [4, 14, 29]. Moreover, locomotor stimulation positively correlates with MDPV concentrations in the striatum [30], in plasma, and with extracellular DA concentrations [56] in the NAcc [4, 29]. The open field test is sensitive to stimulatory/sedative effects of centrally acting drugs, as well as having some sensitivity to emotionality (manifested as changes in the temporo-spatial characteristics of locomotor behaviour). Although effects of MDPV have been characterised in a number of other locomotor tests (activity cage, running wheel and rotorod), there is only one study of its effects in the open field using (Sprague-Dawley) rats, which showed 0.5 mg/kg intraperitoneally (ip.) increased locomotion. In mice, lower doses (1-3 mg/kg) MDPV increased locomotion and exploration were reported, with ataxia, hyperactivity and stereotypies emerging at doses 3-30 mg/kg [18, 43, 49, 53, 57]. Locomotor stimulation at lower doses and gross motor effects/stereotypies at higher doses are consistent with the inverted "U" curvilinear dose response identified for MDPV [45, 58].

Prepulse inhibition (PPI) of the acoustic startle response (ASR) is a behavioural operationalisation of sensorimotor gating that reflects pre-attentional filtering of redundant information [59]. PPI is useful in assessing the psychomimetic properties of drugs and is a behavioural endophenotype of psychosis [60]. Stimulants and cathinones can disrupt PPI, but usually only at higher doses ([61–68], but see [69]). As already described, MDPV use (in humans) can, like other cathinones, result in psychotic symptoms such as hallucinations, however to date, MDPV has not been tested in PPI, nor have any other pyrrolidinophenones.

Aims and Predictions

The aim of the present manuscript is to provide a detailed characterisation of the effects of subcutaneous (sc.) MDPV

in the range 1-4 mg/kg with regard to pharmacokinetics (in sera, brain and lung), biotransformation (urine and sera), acute systemic toxicity (thermoregulation), locomotion (open field), and sensory gating (PPI). The study was performed as a part of an experimental series performed in our laboratory enabling direct comparisons with other NPSs belonging to cathinones, phenethylamines as well as aminoindanes (e.g., [66, 70, 71]). Based on these findings and methods as well as more recent ones presented by others [22, 31, 72], we expected MDPV to peak within first hour after administration in serum and brain, and due to its lipophilicity to accumulate into lung tissue. Additionally, levels of major metabolites were evaluated in tissues, sera as well as in urine. In order to simulate the typical environmental situations in which human MDPV use occurs, we measured body temperature under isolated and aggregated conditions with the prediction that aggregation will lead to more pronounced hyperthermic effects. In order to evaluate the characteristic inverted U locomotor dose-response curve we recorded open field activity at three MDPV doses at two testing-onsets following drug administration. Finally, we expected that MDPV, owing to its powerful effects on DAT, will disrupt PPI more potently than other ring-substituted cathinones tested in our laboratory.

METHOD

Design

For the pharmacokinetic study, samples of blood, whole brains and lungs were collected 30, 60, 120, 240, or 480 min after administration of 2 mg/kg MDPV. Concentrations of MDPV in sera and tissues were calculated as ng/ml or ng/g. The brain/serum ratio was calculated as mean brain concentration /mean serum concentration per sampling time point. The biotransformation study screened for MDPV metabolites and quantified MDPV and major metabolites in urine, sera (ng/mL) and tissues (ng/g) collected over 24 h after 4 mg/kg MDPV.

Open field and PPI experiments used a 4×3 factorial design with MDPV treatment (1, 2, 4 mg/kg or vehicle) and testingonset (15 or 60 min) as independent factors. For the open field, trajectory length (cm, corrected for 3 cm deviations), thigmotaxis (measured as $\Sigma f_{peripheral zones} / \Sigma f_{all zones}$, where f = frequencyof line crossings) and T_{centre} (calculated as Σ_{time} in the central zones) were measured. A repeated measures factor was also included in the open field experimental design in order to measure trajectory length over six 5 min time bins (4 \times 3 \times 6 mixed design). In the PPI experiment, all measures were derived from average startle amplitudes (AVG), and were as follows: % habituation (percentage reduction in ASR from six baseline trials, to the final six trials), ASR (mean ASR was derived from pulse alone trials) and % PPI (calculated as: [100-(mean prepulsepulse trials/mean pulse alone trials)*100]) The thermoregulation study measured rectal temperature (°C) and used a 3 \times 2 \times 13 mixed factorial design with MDPV treatment (2, 4 mg/kg or VEH) and home-cage condition (group or individually caged) as independent factors and measurement time points as a repeated measures factor.

Animals

Male outbred Wistar rats (Velaz, Czech Republic) weighing 180–250 g were housed in pairs at $22 \pm 2^{\circ}$ C on a 12/12 h light/dark cycle and with *ad libitum* water and standard diet. Rats were acclimatised for 7–10 days prior to testing, during which they were weighed twice and handled four times. All tests were conducted under standard conditions: humidity 30–70% and temperature $22 \pm 2^{\circ}$ C. All studies were carried out in accordance with the principles of laboratory animal care of the National Committee for the Care and Use of Laboratory Animals (Czech Republic), and according to Guidelines of the European Union (86/609/EU). The protocol was approved by the National Committee for the Care and Use of Laboratory Animals (Czech Republic) under the number: MEYSCR-27527/2012-31.

Across all experiments, 193 rats were used. For open field and PPI experiments, each had a total sample size of 60 (n = 10). Naïve rats were used for each experiment with the exception that (to reduce animal use) 40 rats from the open field were used for pharmacokinetic sampling (n = 8). For thermoregulation studies, N = 60 (n = 10). For the biotransformation study the total number of rats was three. As previously described, data from vehicle (VEH) control animals for PPI, open field and temperature studies were collected twice per annum as part of a series of standardised NPS studies undertaken in our laboratory (e.g., see [73]).

MDPV Doses

MDPV and other analytical standards were synthesised inhouse at the Department of Organic Chemistry, University of Chemistry and Technology, Prague. In all studies, MDPV was dissolved in physiological saline in a volume of 2 ml/kg, and was always administered sc. as a single bolus. MDPV has been shown to be behaviourally active in locomotor tests in rodents at between 0.5 and 30 mg/kg across ip. and sc. routes. Based on previous behavioural research 1, 2, and 4 mg/kg doses were selected for behavioural testing with the expectation that 1 and 2 mg/kg would produce mild stimulatory effects similar to those sought by people. 4 mg/kg was selected to represent more extreme use, with stereotyped behaviours expected. Pharmacokinetic, biotransformation and thermoregulation studies tested 2 and/or 4 mg/kg were tested; these higher doses were chosen in order to increase the likelihood of detectable effects. The dose rage tested was similar to that used by Novellas et al. [30] where they argued that doses up to 3 mg/kg were within a range used by consumers.

Pharmacokinetics: Determination of MDPV in Serum and Tissues

After completing behavioural testing in the open field, rats were decapitated at 30, 60, 120, 240, or 480 min after administration of 2 mg/kg MDPV, and blood, whole brains and lungs were collected. Separated sera and tissues were stored at -20° C until the toxicological analyses. Sample preparation, LC and MS conditions, method of validation and calibration were based on Meyer et al. [32] and similar to other previously published methods (e.g., [31, 72]). For a full description of pharmacokinetic methods, please refer to Horsley et al. [73].

Biotransformation Study: Determination of MDPV and Its Main Metabolites in Urine

Rats were administered 4 mg/kg MDPV, were placed individually into metabolic cages (Harvard Apparatus, USA) and their 24h urine fractions were collected. During collection urine was maintained below 4°C throughout. Samples were subsequently stored at -40° C until analysis. The methods used to the screening of MDPV metabolites, and quantification of MDPV and its metabolites has been described more fully in Horsley et al. [73].

Systemic Toxicity: Thermoregulation

Rats were housed singly vs. five to a home-cage and rectal temperature was measured for 10 s using a digital thermometer. The first three measurements were drug-free, and were taken hourly, 07:00–09:00 h (inclusive) whereupon 2, 4 mg/kg MDPV or VEH was administered. Thereafter, observations were at 0.5 h intervals 09.30–11.00 h, before resuming hourly measurements 12.00–17.00 h. For a fuller description, see e.g., Páleníček et al. [70, 71].

Behaviour: Open Field and PPI Open Field

Rats were administered MDPV at 1, 2, 4 mg/kg or VEH, and then 15 or 60 min post-administration they were placed individually into the centre of the open field apparatus (a $68 \times 68 \times 30$ cm square black plastic arena) and their behaviour was video-recorded for 30 min. Ethovision Colour-Pro version 3.1.1, (Noldus, Netherlands) was used for behavioural capture and pre-processing. During pre-processing, the arena was virtually divided into 5 × 5 identical square zones with 16 located around the periphery and 9 centrally in order to derive thigmotaxis and T_{centre} variables (see section Design for calculations). The procedures used were the same as those used previously, e.g., Páleníček et al. [70, 71].

PPI

Two days before test, rats were acclimatised to the startle chamber (SR-LAB, San Diego Instruments, California, USA) with a drugfree five min pre-exposure to five pulse alone stimuli (115 dB/20 ms) over 75 dB continuous white noise. On the test day, 15 or 60 min prior to testing, rats were administered 1, 2, 4 mg/kg MDPV or VEH, placed into the startle chamber and acclimatised for five min to a continuous 75 dB white noise. They were then presented with six 125 dB/40 ms duration pulse alone trials, followed by 60 pseudorandomised trials of the following: (A) pulse alone: 40 ms 125 dB; (B) prepulse-pulse: 20 ms 83 dB or 91 dB prepulse, a variable (30, 60 or 120 ms) inter-stimulus interval (ISI: mean 70 ms), then 40 ms 125 dB pulse; (C) 60 ms no stimulus. Finally, six pulse alone trials were delivered. There were 72 trials in total with inter-trial intervals (ITIs) of 4-20 s (mean ITI = 12.27 s). A fuller description can be found in our previously published work (e.g., [70, 71]).

Statistics

Statistical analyses of behavioural and thermoregulation data were conducted using IBM SPSS version 22. Unless stated

otherwise, default alpha was set at p = 0.05, and tests were two tailed. For open field, PPI, and temperature analyses we used factorial analysis of variance (ANOVA). For repeated measured ANOVAs, where Mauchly's test of sphericity was significant Greenhouse-Geisser corrections were used. Planned pairwise comparisons (to follow up significant main effects and interactions involving MDPV) used independent *t*-tests. In order to limit inflation of type 1 error the number of comparisons was restricted to those necessary to test the primary hypotheses (MDPV vs. VEH, no between dose comparisons were made), and in the thermoregulation study, comparisons were focused on early time points with observable mean differences. Where Levene's test for equality of variance was significant, corrected statistics are presented. Corrected degrees of freedom are rounded to the nearest whole number for presentational purposes.

RESULTS

Pharmacokinetics, Metabolism, and Hyperthermic Response

MDPV in Serum and Tissue After 2 mg/kg MDPV

Maximal MDPV (**Figure 1**) concentrations in sera were likely attained before the first measurement, since we saw the highest levels at 30 min (140 ng/mL). The same applied to tissues where influx was not detectably delayed compared to serum since maximal concentrations were also observed at 30 min: 263.97 ng/g in brain, and 531.58 ng/g in lung. Until 2 h, the brain/serum ratio was between 1.8 and 2.5, and at 4–6 h the brain/serum ratio was \sim 4 however, by this time levels in serum and tissue were very low, and almost undetectable by 6 h.

MDPV and Metabolites in Urine, Sera, and Tissues After 4 mg/kg MDPV

MDPV itself was present in urine but was extracted primarily as metabolites. We adopted the nomenclature of Meyer et



al. [32] for the most abundant substances detected which were 4-hydroxy-3-methoxypyrovalerone (demethylenyl-methyl-MDPV), followed by 3,4-dihydroxypyrovalerone (demethylenyl-MDPV) (see **Table 1** and **Figure 2** for quantification and structures). All other metabolites were in a minority.

In tissues, the metabolites demethylenyl-MDPV, demethylenyl-methyl-MDPV, demethylenyl-oxo-MDPV, and oxo-MDPV) were present at very low levels. A maximum value of 8 ng/ml demethylenyl-methyl-MDPV in the brain was observed, and 27 ng/ml in the lungs). In serum demethylenyl-oxo-MDPV was present at low levels (maximum value 23.8 ng/ml), and oxo-MDPV was almost undetectable (maximum value 2.8 ng/ml).

Rectal Temperature After 2 or 4 mg/kg MDPV

There were significant main effects on rectal temperature of drug and time, minimum $F_{(2, 54)} = 44.08$, p = 0.0001, but no main effect of home-cage, $F_{(1, 54)} = 0.19$, p = 0.66. All interactions, including the three-way drug x home-cage x time interaction were significant, minimum $F_{(7, 383)} = 2.79$, p = 0.001. **Figure 3** shows that the groups had equivalent body temperatures at the 8.00 h baseline temperature measurement, maximum $t_{(38)} = 0.41$, p = 0.68. VEH rats' temperatures remained steady at about 37.7–38°C, throughout the temporal observation, irrespective of housing condition.

After 2 or 4 mg/kg temperature increased in both individuallyand (more dramatically in) group-housed rats, peaking at 9.30 h and then diminishing. After 2 mg/kg, temperature was significantly elevated in group- and individually-housed rats until 12.00 noon, minimum $t_{(18)} = 2.32$, p = 0.03 (with some fluctuating differences thereafter). Compared to 2 mg/kg the increase in temperature after 4 mg/kg was longer-lasting: significant elevations were seen in group- and individuallyhoused rats at all-time measurements up until 14.00 h (except 10.00 h, individually-housed), minimum $t_{(18)} = 1.86$, p = 0.04, one tailed (with some fluctuating differences thereafter).

Behaviour: Open Field and PPI

Locomotor Stimulation in the Open Field

The main effect of testing-onset and the bins x testing-onset interaction were not significant, maximum $F_{(3, 183)} = 2.12$, p = 0.11. All other main effects and interactions, including the three-way drug × testing-onset × bins interaction were significant, minimum $F_{(8, 183)} = 3.23$, p = 0.001.

 TABLE 1 | The urinary metabolites of phase II were screened using

 UHPLC-QTOF(MS) and the following glucuronides were confirmed: glucuronide

 demethylenyl-methyl-MDPV and glucuronide demethylenyl-MDPV.

	Concentration in urine (ng/mL)		
	Demethylenyl-MDPV	Demethylenyl-methyl-MDPV	MDPV
Urine 1	704	398	122
Urine 2	586	790	227
Urine 3	284	145	74



At both 15 and 60 min testing-onsets the 4 mg/kg treated rats were indistinguishable from VEH (except in bin 1, 60 min), maximum $t_{(13)} = 1.69$, p = 0.11). At both testing-onsets, rats showed a normal pattern of locomotor habituation (progressively diminishing activity over the session) which was not disturbed by any dose MDPV. However, 1 and 2 mg/kg treated rats were significantly more active than VEH at both the 15 min testing-onset, minimum $t_{(11)} = 2.98$, p = 0.00 (**Figure 4A**), and at the 60 min testing-onset after 1 mg/kg (bins 4–6: minimum $t_{(18)} = 2.54$, p = 0.02), and after 2 mg/kg (bins 2–5: minimum $t_{(18)} = 2.11$, p = 0.05 (**Figure 4B**).

Spatial Distribution of Locomotor Behaviour in the Open Field

There was no main effect of testing-onset on T_{centre} , $F_{(1,72)} = 0.00$, p = 0.95, but the main effect for drug, and drug x testing-onset interaction were significant, minimum

 $F_{(3,72)} = 7.13$, p = 0.00, **Figure 5A**. At 15 min 1 mg/kgtreated rats spent significantly more time in the centre (than VEH), and 4 mg/kg -treated rat significantly less, minimum $t_{(10)} = 3.92$, p = 0.00, with 2 mg/kg having no significant effect, $t_{(12)} = 0.47$, p = 0.64. At 60 min, 1 and 2 mg/kg increased time in the centre, minimum $t_{(18)} = 2.26$, p = 0.04, with 4 mg/kg having no effect, $t_{(18)} = 1.87$, p = 0.07.

There was no main effect of testing-onset on thigmotaxis, $F_{(1,72)} = 2.76$, p = 0.10, however there was a significant main effect of drug, and drug x testing-onset interaction, minimum $F_{(3,72)} = 5.77$, p = 0.00, **Figure 5B**. At 15 min, 2 and 4 mg/kg increased, and 1 mg/kg decreased thigmotaxis, minimum $t_{(10)} = 2.27$, p = 0.05. At 60 min, 4 mg/kg again increased thigmotaxis, $t_{(18)} = 2.55$, p = 0.02, but 1 and 2 mg/kg had no significant effect, maximum $t_{(18)} = 0.77$, p = 0.45.



FIGURE 3 | Mean rectal temperature (°C) over 10 h in rats (n = 10) housed individually (solid lines) or in groups of five (dashed lines). 4 mg/kg MDPV (black markers), 2 mg/kg MDPV (grey markers) or VEH (white markers) was administered sc. at 09.00 (black dotted vertical line). Error bars show ±1 standard error of the mean. Asterisks indicate significant differences from VEH at minimum p < 0.05.



FIGURE 4 | Mean trajectory length (cm/5 min over 30 min) tested 15 min (A) or 60 min (B) after drug administration: sc. MDPV 1 mg/kg (light grey), 2 mg/kg (mid-grey) or 4 mg/kg (black) vs. VEH (white). Shown alongside 5 min bins (on the x axis) are minutes elapsed since drug administration. Error bars show ± 1 standard error of the mean. Asterisks indicate significant differences from VEH for the 1 and 2 mg/kg groups, at minimum p < 0.05. Example trajectory patterns are shown in (C).

Prepulse Inhibition

Habituation and ASR data showed no significant main effects or interactions, maximum $F_{(3, 72)} = 2.25$, p = 0.24. PPI data showed no main effect of the drug treatment, $F_{(3, 72)} = 2.21$, p = 0.09, but the main effect of testing-onset, and the drug

treatment × testing-onset interaction were significant, minimum $F_{(3, 72)} = 2.73$, p = 0.05, **Figure 6**. In the 15 min testing-onset group, 4 mg/kg significantly disrupted PPI, $t_{(18)} = 2.70$, p = 0.02, and 1 and 2 mg/kg had a marginally significant effect, maximum $t_{(18)} = 1.95$, p = 0.07. At 60 min testing-onset, none of the





MDPV doses significantly affected PPI, maximum $t_{(18)} = 1.67$, p = 0.11.

DISCUSSION

As expected, our pharmacokinetic findings were in line with previous results for serum and brain, and we reported new data on lungs; after 2 mg/kg, MDPV reaches its peak concentrations in serum and tissues in under 30 min and declines rapidly thereafter. Our biotransformation study detected a number of metabolites in sera and tissues, however at very low levels. In urine, concentrations of metabolites exceeded those of MDPV itself, and we confirmed that demethylenyl-methyl-MDPV the most abundant. As previous studies have shown we found a small increase in temperature in individually-housed rats, which was exacerbated by crowded cage conditions. In line with existing findings, our behavioural effects were systematically related to drug concentrations/dose, and showed good correspondence with the known pharmacokinetic time-course for MDPV. As expected, in the open field, lower doses generally stimulated locomotion and exploration, whereas the higher dose initially induced stereotypy/gross suppression of motor behaviour, consistent with stimulant-typical psychomimesis. We presented additional original data which supported this: 4 mg/kg can induce transient psychomimesis shown as disrupted PPI.



Pharmacokinetics and Metabolism

The highest concentrations of MDPV in serum, brain, and lung tissue were recorded at 30 min followed by a rapid decline, suggesting that in all cases, the actual peaks were prior to our first measurement. Previous pharmacokinetic data in rats (which were not published at the time of our data collection) showed peak plasma concentrations of MDPV at 10–20 min post-administration [29, 30]. It is therefore almost certain that our value of 140 ng/mL for peak serum concentration is an underestimation. Even so, concentrations in this range in the plasma of human MDPV users are more typical of cases of acute intoxication and overdose (in excess of 50–300 ng/mL) than of recreational use where milder effects are associated with much lower blood levels at \sim 10– 50 ng/mL [74].

Novellas et al. [30] reported that MDPV reached the striatum of the brain around 5 min after sc. MDPV (1 mg/kg) in rats and peaked 20-25 min after that, suggesting that our observed peak brain concentration value at 30 min (263.97 ng/g) was likely close to the actual value. From 30 min and the following 2 h, the brain concentration was approximately twice that in the sera. This is consistent with the brain/serum ratio of 2.21 reported by Novellas et al. [30], and reflects MDPV's lipid solubility and capacity to cross the blood-brain barrier. Evidence suggests a kinetic profile of fast onset and relatively short duration, consistent with our short-lasting peak behavioural effects, and with estimates of the onset and duration of subjective effects in humans [25, 28]. MDPV's short duration may have a causal role in re-dosing and escalating use (shown in human and rodent studies), and together with fast penetration of the brain [30] these characteristics are indicative of MDPV's addictive potential [36].

We observed levels of MDPV that were approximately four times higher in lungs than in sera. Rapid transition of substances (such as drugs) from blood to lungs is characteristic of parenterally administered cationic compounds with a lipophilic profile [75]. The rapidity and magnitude of accumulation is partly a consequence of the relatively large volume of blood flowing to the lungs in compared to other organs, such as the brain, where molecules become trapped by lysosomes and mitochondria and accumulate in lung tissue as gradually eluting pools [75]. In consequence, accumulation in, and elimination from lung tissue is usually rapid, which was supported by our data, where lung concentrations of MDPV declined more quickly than in the brain. The relevance (if any) of drug accumulation in the lungs for MDPV toxicity is unclear; pyrrolidinophenone toxicity primarily involve CNS and cardiovascular related problems [13, 56].

As expected, MDPV itself was detected in urine at only very low concentrations, instead MDPV was detected as two major metabolites demethylenyl-MDPV and demethylenylmethyl-MDPV. These urinary metabolites were at concentrations approximately three to four times higher than the parent drug. Other metabolites were detected in urine, serum, brain and lung but again at very low concentrations. Taken together with pharmacokinetic findings here and previous studies [30-32, 72], MDPV appears to be cleared from sera and tissues relatively rapidly (here almost undetectable by 6 h), and by 24h only very small amounts of MDPV in the form of metabolites remain in the body the majority having been excreted in urine. Toxicological tests that seek to confirm MDPV in blood or tissues after 6 h should search for a range of MDPV metabolites rather than MDPV itself and use the most sensitive methods available. Our sample size for the biotransformation study was small, and there was variability in the data, both of which may have contributed to the observation of lower concentrations, and few metabolites than other studies have shown previously.

Thermoregulation

As in our pharmacokinetic study, we found peak temperatures at our earliest measurement time-point (30 min postadministration), therefore, we cannot exclude the possibility that temperatures were increased prior to this. MDPV increased body temperature for several hours up to a maximum of $\sim 0.5^{\circ}$ C in rats housed individually at \sim 22°C. This small increase is in agreement with previous work using individually-housed Wistar rats where MDPV's effects on body temperature (at a similar dose range) were negligible even at 30°C ambient temperatures [45]. A number of findings have been published now that demonstrate that environmental crowding and hotter ambient temperatures can exacerbate drug-induced hyperthermia [70, 73, 76, 77]. We found longer-lasting increases in temperature in group-housed rats, up to 2°C. MDPV increases brain and body temperature in mice at high ambient temperatures and under conditions of social interaction [21, 43, 44], perhaps due to combined DATmediated hyperactivity and NET-mediated sympathomimetic effects [20, 22], although recent research has also implicated 5-HT [21]. If environmental/social conditions can potentiate both locomotor and temperature effects simultaneously, these circumstances might increase the risk of toxic reactions. On the other hand, although MDPV toxicity and fatalities in humans have been documented [7, 34-39, 41], they remain relatively uncommon given the prevalence of cathinone use. Moreover, the doses tested here equate to very high human doses (in excess of ~ 20 mg, [78]), that are in the range where adverse reactions would be expected [25].

Locomotor Behaviour

As expected, the locomotor stimulant action of MDPV across the three doses (1, 2, and 4 mg/kg) was consistent with the inverted "U" behavioural dose-response of dopaminergic agonists (such that progressively larger doses resulted in progressively increasing, then decreasing activity). In our behavioural studies, the 15 min testing-onset synchronised with peak serum and brain concentrations as well as their initial decline. Effects over testing-onsets showed that locomotor effects related to declining systemic levels of MDPV, e.g., effects of 1 mg/kg at 15 min were equivalent to residual effects of 2 mg/kg at 60 min. Other pyrrolidine-containing synthetic cathinone derivatives, α -pyrrolidinopropiobutiophenone (α -PBP), and α pyrrolidinopentiophenone (α -PVP), as well as MDPV, show this inverted "U" curvilinear relationship with locomotor stimulation in rats ([14, 45, 52, 58] but see [53] for effects in mice). The normal pattern of locomotor habituation was not disturbed by lower doses of MDPV, however at 4 mg/kg (which induced some hypolocomotion at 15 min) by 80-90 min post-drug administration, locomotion appeared to be increasing. Had testing continued, hyperlocomotion may have been observed as brain drug concentrations diminished, as previously reported [45, 52]. MDPV's potent stimulatory effects most likely relate to its potent DAT inhibition, coupled with its lack of capacity to increase extracellular 5-HT which can attenuate DA-induced locomotor effects [4].

Rats usually avoid the aversive centre of the open field (which is brighter more and open) preferring to spend time in the

periphery (next to the arena walls). The spatial characteristics of locomotor behaviour in this paradigm can provide some indication of drug effects on emotionality with increased exploration of the centre suggesting decreased emotionality (anxiolytic-like effects) and vice versa. [79-81]. In our experience, effects on T_{centre} and thigmotaxis are usually (but not always) the inverse of one another, however, whilst related, T_{centre} may be more sensitive to effects on emotionality, and thigmotaxis to locomotor stereotypy. MDPV has been shown previously to increase exploration in mice [53], as well as induce stereotypy (at a range of doses) in rats and mice [30, 45, 53, 72]. Here, as the dose/systemic levels of MDPV declined below \sim 2 mg/kg there was increased exploration of the centre (unconfounded by thigmotaxis), possibly indicative of anxiolytic effects (perhaps equating to positive affect or euphoria in humans at acute lower doses; [25]). At \sim 2 mg/kg or more, there was only increased thigmotaxis indicative of locomotor activation and/or emerging stereotypy. Stimulant-typical stereotypies emerge in a systematic manner [30, 72, 82], which was shown here, and, with MDPV, are reversible with the typical antipsychotic (a DA D2 antagonist), haloperidol (e.g., [30]), which is indicative of its DA-ergic origin and psychomimetic potential.

Sensorimotor Gating and Psychomimesis (PPI)

Only 4 mg/kg MDPV disrupted PPI significantly and only transiently so (at the 15 min testing-onset), when plasma levels of MDPV were at their peak, and peak brain concentration was developing. By 60 min, MDPV levels in sera and brain had declined by \sim 30-40% to around 50 and 150 ng/mL respectively, and PPI was no longer significantly disturbed. A specific disruption of PPI (i.e., in the absence of significant baseline drug effects on habituation or ASR that might confound interpretation) reflects dysfunctional sensorimotor gating, and is an index of the psychomimetic properties of drugs [60]. Evidence to date has shown that other (ring-substituted) cathinones and stimulants can disrupt PPI, but usually only at higher doses [61-69]. Although different cathinones and stimulants operate via different DA-ergic mechanisms (i.e., DAT inhibition and/or nonexocytotic efflux) the net result is an increase extracellular DA. 5-HT-ergic compounds more reliably disturb PPI (e.g., [73, 83, 84]), however it would appear that hyperdopaminergia alone is sufficient to disrupt PPI, since MDPV has negligible effects on 5-HT [14, 85]. Moreover, cathinone-induced PPI deficits and stimulant-typical psychomimesis can be reversed with typical and atypical antipsychotics [30, 86] which have DA D2 receptor antagonism in common [87], suggesting a role for this receptor subtype specifically [63]. Our findings indicate that the duration of psychological symptoms is likely to be short, which suggests that in cases of acute MDPV intoxication, adverse psychological effects may pass relatively quickly.

CONCLUSIONS

In conclusion, we confirmed MDPV's typical DA-ergic psychostimulant drug profile: at lower doses, it modestly

increased locomotion and exploration; at higher doses it produced stimulant-typical psychomimetic and gross motor effects/stereotypy. Behavioural findings have relevance to recreational users who, in striving for stimulatory effects may take large amounts of MDPV, unaware that this will only delay rather than increase stimulation (which is modest, in any case) and induce adverse psychological and physical effects instead, with risks of toxicity (especially if taken in combination with other drugs) increasing with dose. However, our lowest dose of MDPV, 1 mg/kg, has been calculated as equivalent to 10 mg human dose and our highest dose is equivalent to 30 mg or more [78]. Thus even our lowest dose, whilst within the range used by people, was nevertheless a high dose, in the range associated with unwanted effects [25]. Responsible and occasional use of sensible doses of MDPV alone is probably relatively low-risk for most healthy people. However, users should be aware that MDPV's acute effects may create interactions between their activity levels, environmental and social circumstances and potentiate temperature dysregulation. Moreover, it is important, in a world where NPSs proliferate [3, 36] and where "sensible" recreational drug-taking is increasingly normalised [4, 5] that even where acute use appears low risk, the possible effects of long-term, chronic or binge use of MDPV on psychological and physical health are kept in mind. These are simply, as yet,

REFERENCES

- 1. Shanks KG, Dahn T, Behonick G, Terrell A. Analysis of first and second generation legal highs for synthetic cannabinoids and synthetic stimulants by ultra-performance liquid chromatography and time of flight mass spectrometry. *J Anal Toxicol.* (2012) **36**:360–71. doi: 10.1093/jat/bks047
- European Monitoring Centre for Drugs and Drug Addiction. EMCDDA -European Monitoring Centre for Drugs and Drug Addiction and Europol European Drug Report. Trends and Developments, Luxembourg (2017).
- United Nations Office of Drugs and Crime (UNDOC). World Drug Report. (2015). Available online at: http://www.unodc.org/documents/wdr2015/ World_Drug_Report_2015.pdf
- Baumann MH, Bukhari MO, Lehner KR, Anizan S, Rice KC, Concheiro M, et al. Neuropharmacology of 3,4-Methylenedioxypyrovalerone (MDPV), its metabolites, and related analogs. *Curr Topics Behav Neurosci.* (2017) 32:93–117. doi: 10.1007/7854_2016_53
- Parker H, Williams L, Aldridge J. The normalization of 'Sensible' recreational drug use: further evidence from the North West England longitudinal study. *Sociology* (2002) 36:941–64. doi: 10.1177/003803850203600408
- Murphy CM, Dulaney AR, Beuhler MC, Kacinko S. "Bath salts" and "plant food" products: the experience of one regional US poison center. J Med Toxicol. (2013) 9:42–8. doi: 10.1007/s13181-012-0243-1
- Spiller HA, Ryan ML, Weston RG, Jansen J. Clinical experience with and analytical confirmation of "bath salts" and "legal highs" (synthetic cathinones) in United States. *Clin Toxicol (Phila)*. (2011) 49:499–505. doi: 10.3109/15563650.2011.590812
- Baumann MH. Awash in a sea of 'bath salts': implications for biomedical research and public health. *Addiction* (2014) 109:1577–9. doi: 10.1111/add.12601
- Harvey EL, Baker LE. Differential effects of 3, 4-methylenedioxypyrovalerone (MDPV) and 4-methylmethcathinone (mephedrone) in rats trained to discriminate MDMA or a d-amphetamine+MDMA mixture. *Psychopharmacology* (2016) 233:673–80. doi: 10.1007/s00213-015-4142-4
- Winder GS, Stern N, Hosanagar A. Are "bath salts" the next generation of stimulant abuse? J Subst Abuse Treat. (2013) 44:42–5. doi: 10.1016/j.jsat.2012.02.003

unknown, and data will only emerge in years to come. The short-lasting behavioural effects and pharmacokinetics shown here are consistent with existing evidence that MDPV has characteristics that encourage risky use (such as re-dosing, binge or poly-drug use, 36) as well as significant addictive potential.

AUTHOR CONTRIBUTIONS

All authors made a substantial contribution to the conception or design of the work; or the acquisition, analysis, or interpretation of data for the work. All authors were involved in drafting the work or revising it critically for important intellectual contents. All authors gave final approval for the current version of the work to be published. All authors agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

FUNDING

This work was funded by a specific university research (project MSMT No 20-SVV/2017) and by the Ministry of Interior of the Czech Republic (project VG20172020056).

- Green AR, King MV, Shortall SE, Fone KC. The preclinical pharmacology of mephedrone; not just MDMA by another name. *Br J Pharmacol.* (2014) 171:2251–68. doi: 10.1111/bph.12628
- Ellefsen KN, Concherio M, Huestis MA. Synthetic cathinone pharmacokinetics, analytical methods, and toxicological findings from human performance and postmortem cases. *Drug Metab Rev.* (2016) 48:237–65. doi: 10.1080/03602532.2016.1188937
- Zaitsu K, Katagi M, Tsuchihashi H, Ishii A. Recently abused synthetic cathinones, α-pyrrolidinophenone derivatives: a review of their pharmacology, acute toxicity, and metabolism. *Forensic Toxicol.* (2014) 32:1. doi: 10.1007/s11419-013-0218-1
- Baumann MH, Partilla JS, Lehner KR, Thorndike EB, Hoffman AF, Holy M, et al. Powerful cocaine-like actions of 3,4-methylenedioxypyrovalerone (MDPV), a principal constituent of psychoactive 'bath salts' products. *Neuropsychopharmacology* (2013) 38:552–62. doi: 10.1038/npp.2012.204
- Cameron KN, Kolanos R, Solis E Jr, Glennon RA, De Felice LJ. Bath salts components mephedrone and methylenedioxypyrovalerone (MDPV) act synergistically at the human dopamine transporter. *Br J Pharmacol.* (2013) 168:1750–7. doi: 10.1111/bph.12061
- Eshleman AJ, Wolfrum KM, Hatfield MG, Johnson RA, Murphy KV, Janowsky A. Substituted methcathinones differ in transporter and receptor interactions. *Biochem Pharmacol.* (2013) 85:1803–15. doi: 10.1016/j.bcp.2013.04.004
- López-Arnau R, Martínez-Clemente J, Pubill D, Escubedo E, Camarasa J. Comparative neuropharmacology of three psychostimulant cathinone derivatives: butylone, mephedrone and methylone. *Br J Pharmacol.* (2012) 167:407–20. doi: 10.1111/j.1476-5381.2012.01998.x
- Marusich JA, Antonazzo KR, Wiley JL, Blough BE, Partilla JS, Bauman MH. Pharmacology of novel synthetic stimulants structurally related to the "bath salts" constituent 3,4-methylenedioxypyrovalerone (MDPV). *Neuropharmacol.* (2014) 87:206–13. doi: 10.1016/j.neuropharm.2014.02.016
- Meltzer PC, Butler D, Deschamps JR, Madras BK. 1-(4-Methylphenyl)-2pyrrolidin-1-yl-pentan-1-one (Pyrovalerone) analogues: a promising class of monoamine uptake inhibitors. *J Med Chem.* (2006) 49:1420–32. doi: 10.1021/jm050797a
- 20. Simmler LD, Buser TA, Donzelli M, Schramm Y, Dieu LH, Huwyler J, et al. Pharmacological characterization of designer cathinones *in*

vitro. Br J Pharmacol. (2013) 168:458–70. doi: 10.1111/j.1476-5381.2012. 02145.x

- Gannon BM, Galindo KI, Rice KC, Collins GT. Individual differences in the relative reinforcing effects of 3,4-methylenedioxypyrovalerone (MDPV) under fixed and progressive ratio schedules of reinforcement in rats. J Pharmacol Exp Ther. (2017) 361:181–9 doi: 10.1124/jpet.116.239376
- Liechti M. Novel psychoactive substances (designer drugs): overview and pharmacology of modulators of monoamine signaling. *Swiss Med Wkly*. (2015) 145:40–3. doi: 10.4414/smw.2015.14043
- Baumann MH, Ayestas MA Jr, Partilla JS, Sink JR, Shulgin AT, Daley PF, et al. The designer methcathinone analogs, mephedrone and methylone, are substrates for monoamine transporters in brain tissue. *Neuropsychopharmacology* (2012) 37:1192–203. doi: 10.1038/npp.2011.304
- Buenrostro-Jáuregui M, Ciudad-Roberts A, Moreno J, Muñoz-Villegas P, López-Arnau R, Pubill D, et al. Changes in CREB and deltaFosB are associated with the behavioural sensitization induced by methylenedioxypyrovalerone. J Psychopharmacol. (2016) 30:707–12. doi: 10.1177/0269881116645300
- 25. Dancesafe (2017). MDPV. Available online at: https://dancesafe.org/mdpv/
- Ross EA, Reisfield GM, Watson MC, Chronister CW, Goldberger BA. Psychoactive "bath salts" intoxication with methylenedioxypyrovalerone. *Am J Med.* (2012) 125:854–8. doi: 10.1016/j.amjmed.2012.02.019
- 27. Kelly JP. Cathinone derivatives: a review of their chemistry, pharmacology and toxicology. *Drug Test Anal.* (2011) **3**:439–53. doi: 10.1002/dta.313
- Joksovic P, Mellos N, van Wattum PJ, Chiles C. "Bath salts"-induced psychosis and serotonin toxicity. J Clin Psychiatry (2012) 73:1125. doi: 10.4088/JCP.12cr07819
- Anizan S, Concheiro M, Lehner KR, Bukhari MO, Suzuki M, Rice KC, et al. Linear pharmacokinetics of 3,4-methylenedioxypyrovalerone (MDPV) and its metabolites in the rat: relationship to pharmacodynamic effects. *Addict Biol.* (2016) 21:339–47. doi: 10.1111/adb.12201
- Novellas J, López-Arnau R, Carbó ML, Pubill D, Camarasa J, Escubedo E. Concentrations of MDPV in rat striatum correlate with the psychostimulant effect. J Psychopharmacol. (2015) 29:1209–18. doi: 10.1177/0269881115598415
- 31. Strano-Rossi S, Cadwallader AB, de la Torre X, Botrè F. Toxicological determination and *in vitro* metabolism of the designer drug methylenedioxypyrovalerone (MDPV) by gas chromatography/mass spectrometry and liquid chromatography/quadrupole time-of-flight mass spectrometry. *Rapid Commun Mass Spectrom*. (2010) 24:2706–14. doi: 10.1002/rcm.4692
- 32. Meyer MR, Du P, Schuster F, Maurer HH. Studies on the metabolism of the α-pyrrolidinophenone designer drug methylenedioxy-pyrovalerone (MDPV) in rat and human urine and human liver microsomes using GC-MS and LC-high-resolution MS and its detectability in urine by GC-MS. *J Mass Spectrom*. (2010) 45:1426–42. doi: 10.1002/jms.1859
- Marinetti LJ, Antonides HM. Analysis of synthetic cathinones commonly found in bath salts in human performance and postmortem toxicology: method development, drug distribution and interpretation of results. J Anal Toxicol. (2013) 37:135–46. doi: 10.1093/jat/bks136
- Froberg BA, Levine M, Beuhler MC, Judge BS, Moore PW, Engebretsen KM, et al. Acute methylenedioxypyrovalerone toxicity. J Med Toxicol. (2015) 11:185–94. doi: 10.1007/s13181-014-0446-8
- Karch SB. Cathinone neurotoxicity (The "3Ms"). Curr Neuropharmacol. (2015) 13:21-5. doi: 10.2174/1570159X13666141210225009
- European Monitoring Centre for Drugs and Drug Addiction. EMCDDA-Europol Joint Report on a New Psychoactive Substance: MDPV (3,4methylenedioxypyrovalerone) (2014). Available online at: http://www.emcdda. europa.eu/system/files/publications/819/TDAS14001ENN_466653.pdf
- 37. Kesha K, Boggs CL, Ripple MG, Allan CH, Levine B, Jufer-Phipps R, et al. Methylenedioxypyrovalerone ("bath salts"), related death: case report and review of the literature. *J Forensic Sci.* (2013) 58:1654–9. doi: 10.1111/1556-4029.12202
- Murray BL, Murphy CM, Beuhler MC. Death following recreational use of designer drug "bath salts" containing 3,4-Methylenedioxypyrovalerone (MDPV). J Med Toxicol. (2012) 8:69–75. doi: 10.1007/s13181-011-0196-9
- Wyman JF, Lavins ES, Engelhart D, Armstrong EJ, Snell KD, Boggs PD., et al. Postmortem tissue distribution of MDPV following lethal intoxication by "bath salts". J Anal Toxicol. (2013) 37:182–5. doi: 10.1093/jat/bkt001

- Mugele J, Nanagas KA, Tormoehlen LM. Serotonin syndrome associated with MDPV use: a case report. Ann Emerg Med. (2012) 60:100–2. doi: 10.1016/j.annemergmed.2011.11.033
- Penders TM, Gestring RE, Vilensky DA. Intoxication delirium following use of synthetic cathinone derivatives. *Am J Drug Alc Abuse*. (2012) 38:616–7. doi: 10.3109/00952990.2012.694535
- Prosser, J. M., and Nelson, L. S. The toxicology of bath salts: a review of synthetic cathinones. J Med Toxicol. (2012) 8:33–4. doi: 10.1007/s13181-011-0193-z
- 43. Fantegrossi WE, Gannon BM, Zimmerman SM, Rice KC. *In vivo* effects of abused 'bath salt' constituent 3,4-methylenedioxypyrovalerone (MDPV) in mice: drug discrimination, thermoregulation, and locomotor activity. *Neuropsychopharmacology* (2012) 38:563–73. doi: 10.1038/npp.2012.233
- Kiyatkin EA, Ren S. MDMA, Methylone, and MDPV: drug-induced brain hyperthermia and its modulation by activity state and environment. *Curr Topics Behav Neurosci.* (2016) 32:183–208. doi: 10.1007/7854_2016_35
- 45. Aarde SM, Huang PK, Creehan KM, Dickerson TJ, Taffe M A. The novel recreational drug 3,4-methylenedioxypyrovalerone (MDPV) is a potent psychomotor stimulant: self-administration and locomotor activity in rats. *Neuropharmacology* (2013) 71:130–40. doi: 10.1016/j.neuropharm.2013.04.003
- 46. Bonano JS, Glennon RA, De Felice LJ, Banks ML, Negus SS. Abuse-related and abuse-limiting effects of methcathinone and the synthetic "bath salts" cathinone analogs methylenedioxypyrovalerone (MDPV), methylone and mephedrone on intracranial self-stimulation in rats. *Psychopharmacology* (*Berl*). (2014) 231:199–207. doi: 10.1007/s00213-013-3223-5
- Gannon BM, Russell LN, Modi MS, Rice KC, Fantegrossi WE. Effects of orally self-administered bath salt constituent 3,4methylenedioxypyrovalerone (MDPV) in mice. *Drug Alc Depend.* (2017) 179:408–15. doi: 10.1016/j.drugalcdep.2017.06.031
- Gannon BM, Williamson A, Rice KC, Fantegrossi WE. Role of monoaminergic systems and ambient temperature in *bath salts* constituent 3,4-methylenedioxypyrovalerone (MDPV) elicited hyperthermia and locomotor stimulation in mice. *Neuropsychopharmacology* (2017) 30422–7. doi: 10.1016/j.neuropharm.2017.09.004
- Karlsson L, Andersson M, Kronstrand R, Kugelberg FC. Mephedrone, Methylone and 3,4-Methylenedioxypyrovalerone (MDPV) induce conditioned place preference in mice. *Basic Clin Pharmacol Toxicol.* (2014) 115:411–6. doi: 10.1111/bcpt.12253
- Moreno-Rius J, Pubill D, Escubedo E, Garcia JC, Miquel M. Locomotor activating effects and addiction-like features of MDPV as assessed in preclinical studies: a review. *Agora de Salut.* (2017) 4.25:239–46. doi: 10.6035/AgoraSalut.2017.4.25
- Watterson LR, Kufahl PR, Nemirovsky NE, Sewalia K, Grabenauer M, Thomas BF, et al. Potent rewarding and reinforcing effects of the synthetic cathinone 3,4-methylenedioxypyrovalerone (MDPV). *Addict Biol.* (2014) 19:165–74. doi: 10.1111/j.1369-1600.2012.00474.x
- Huang PK, Aarde SM, Angrish D, Houseknecht KL, Dickerson TJ, Taffe MA. Contrasting effects of d-methamphetamine, 3,4-methylenedioxymethamphetamine, 3,4-methylenedioxypyrovalerone, and 4-methylmethcathinone on wheel activity in rats. *Drug Alc Depend*. (2012) **126**:168–75. doi: 10.1016/j.drugalcdep.2012.05.011
- Marusich JA, Grant KR, Blough BE, Wiley JL. Effects of synthetic cathinones contained in "bath salts" on motor behavior and a functional observational battery in mice. *Neurotoxicology* (2012) 33:1305–13. doi: 10.1016/j.neuro.2012.08.003
- Schindler CW, Thorndike EB, Goldberg SR, Lehner KR, Cozzi NV, Brandt SD, et al. Reinforcing and neurochemical effects of the "bath salts" constituents 3,4-methylenedioxypyrovalerone (MDPV) and 3,4-methylenedioxy-Nmethylcathinone (methylone) in male rats. *Psychopharmacology (Berl)* (2016) 233:1981–90. doi: 10.1007/s00213-015-4057-0
- Watterson LR, Kufahl PR, Taylor SB, Nemirovsky NE, Olive MF. Sensitization to the motor stimulant effects of 3,4-methylenedioxypyrovalerone (MDPV) and cross-sensitization to methamphetamine in rats. *J Drug Alcohol Res.* (2016) 5:235967. doi: 10.4303/jdar/235967
- 56. Schindler CW, Thorndike EB, Suzuki M, Rice KC, Baumann MH. Pharmacological mechanisms underlying the cardiovascular effects of the bath

salt constituent 3,4-methylenedioxypyrovalerone (MDPV). Br J Pharmacol. (2016) 173:3492–501. doi: 10.1111/bph.13640

- Berquist MD, Traxler HK, Mahler AM, Baker LE. Sensitization to the locomotor stimulant effects of "bath salt" constituents, 4methylmethcathinone (4-MMC) and 3,4-methylenedioxypyrovalerone (MDPV), in male Sprague-Dawley rats. *Drug Alc Depend*. (2016) 164:128–34. doi: 10.1016/j.drugalcdep.2016.05.001
- Gatch MB, Taylor CM, Forster MJ. Locomotor stimulant and discriminative stimulus effects of 'bath salt' cathinones. *Behav Pharmacol.* (2013) 24:437–47. doi: 10.1097/FBP.0b013e328364166d
- Swerdlow NR, Geyer MA. Using an animal model of deficient sensorimotor gating to study the pathophysiology and new treatments of schizophrenia. *Schizophr Bull.* (1998) 24:285–301. doi: 10.1093/oxfordjournals.schbul.a033326
- Geyer MA, Krebs-Thomson K, Braff DL, Swerdlow NR. Pharmacological studies of prepulse inhibition models of sensorimotor gating deficits in schizophrenia: a decade in review. *Psychopharmacology (Berl)*. (2001) 156:117–54. doi: 10.1007/s002130100811
- Hadamitzky M, Markou A, Kuczenski R. Extended access to methamphetamine self-administration affects sensorimotor gating in rats. *Behav Brain Res.* (2011) 217:386–90. doi: 10.1016/j.bbr.2010.11.009
- Mansbach RS, Geyer MA, Braff DL. Dopaminergic stimulation disrupts sensorimotor gating in the rat. *Psychopharmacology (Berl*). (1988) 94:507–14. doi: 10.1007/BF00212846
- Ralph RJ, Varty GB, Kelly MA, Wang YM, Caron MG, Rubinstein M, et al. The dopamine D-2, but not D-3 or D-4, receptor subtype is essential for the disruption of prepulse inhibition produced by amphetamine in mice. J Neurosci. (1999) 19:4627–33. doi: 10.1523/JNEUROSCI.19-11-04627.1999
- Ralph RJ, Paulus MP, Geyer MA. Strain-specific effects of amphetamine on prepulse inhibition and patterns of locomotor behavior in mice. *J Pharmacol Exp Ther.* (2001) 298:148–55. Available online at: http://jpet.aspetjournals. org/content/298/1/148/tab-article-info
- Sills TL. Amphetamine dose dependently disrupts prepulse inhibition of the acoustic startle response in rats within a narrow time window. *Brain Res Bull.* (1999) 48:445–8. doi: 10.1016/S0361-9230(99)00036-2
- Štefková K, Židková M, Horsley RR, Pinterová N, Šíchová K, Uttl, L., et al. Pharmacokinetic, ambulatory, and hyperthermic effects of 3,4methylenedioxy-N-methylcathinone (methylone) in rats. *Front Psychiatry* (2017) 8:232. doi: 10.3389/fpsyt.2017.00232
- Swerdlow NR, Mansbach RS, Geyer MA, Pulvirenti L, Koob GF, Braff DL. Amphetamine disruption of prepulse inhibition of acoustic startle is reversed by depletion of mesolimbic dopamine. *Psychopharmacology (Berl)*. (1990) 100:413–6. doi: 10.1007/BF02244616
- Wan FJ, Swerdlow NR. Sensorimotor gating in rats is regulated by different dopamine-glutamate interactions in the nucleus accumbens core and shell subregions. *Brain Res.* (1996) 722:168–76. doi: 10.1016/0006-8993(96)00209-0
- Shortall SE, Macerola AE, Swaby RT, Jayson R, Korsah C, Pillidge KE, et al. Behavioural and neurochemical comparison of chronic intermittent cathinone, mephedrone and MDMA administration to the rat. *Eur Neuropsychopharmacol.* (2013) 23:1085–95. doi: 10.1016/j.euroneuro.2012.09.005
- Páleníček T, Balikova M, Rohanova M, Novak T, Horacek J, Fujakova M, et al. Behavioral, hyperthermic and pharmacokinetic profile of paramethoxymethamphetamine (PMMA) in rats. *Pharmacol Biochem Behav*. (2011) **98**:130–9. doi: 10.1016/j.pbb.2010.12.011
- Páleníček T, Lhotkova E, Zidkova M, Balikova M, Kuchar M, Himl M, et al. Emerging toxicity of 5,6-methylenedioxy-2-aminoindane (MDAI): pharmacokinetics, behaviour, thermoregulation and LD50 in rats. *Prog Neuropsychopharmacol Biol Psychiatry* (2016) 69:49–59. doi: 10.1016/j.pnpbp.2016.04.004
- 72. Anizan S, Ellefsen K, Concheiro M, Suzuki M, Rice KC, Baumann MH, et al. 3,4-Methylenedioxypyrovalerone (MDPV) and metabolites quantification in human and rat plasma by liquid chromatography-high resolution mass spectrometry. *Anal Chim Acta* (2014) **827**:54–63. doi: 10.1016/j.aca.2014.04.015

- 73. Horsley RR, Lhotkova E, Hajkova K, Jurasek B, Kuchar M, Páleníček T. Detailed pharmacological evaluation of methoxetamine (MXE), a novel psychoactive ketamine analogue-Behavioural, pharmacokinetic and metabolic studies in the Wistar rat. *Brain Res Bull.* (2016) **126**:102–10. doi: 10.1016/j.brainresbull.2016.05.002
- 74. Baselt RC. Disposition of Toxic Drugs and Chemicals in Man. Seal Beach, CA: Biomedical Publications (2014).
- Upton RN, Doolette DJ. Kinetic aspects of drug disposition in the lungs. *Clin Exp Pharmacol Physiol.* (1999) 26:381–91. doi: 10.1046/j.1440-1681.1999.03048.x
- Green AR, O'Shea E, Colado MI. A review of the mechanisms involved in the acute MDMA (ecstasy)-induced hyperthermic response. *Eur J Pharmacol.* (2004) 500:3–13. doi: 10.1016/j.ejphar.2004.07.006
- 77. Green AR, Sanchez V, O'shea E, Saadat KS, Elliott JM, Colado MI. Effect of ambient temperature and a prior neurotoxic dose of 3,4methylenedioxymethamphetamine (MDMA) on the hyperthermic response of rats to a single or repeated ('binge' ingestion) low dose of MDMA. *Psychopharmacology (Berl)* (2004) **173**:264–9. doi: 10.1007/s00213-003-1725-2
- Reagan-Shaw S, Nihal M, Ahmad N. Dose translation from animal to human studies revisited. J Faseb. (2008) 22:659–61. doi: 10.1096/fj.07-9574LSF
- Archer J. Tests for emotionality in rats and mice: a review. *Animal Behav.* (1973) 21:205–35. doi: 10.1016/S0003-3472(73)80065-X
- Walsh RN, Cummins RA. The Open-Field Test: a critical review. *Psychol Bull.* (1976) 83:482–504. doi: 10.1037/0033-2909.83.3.482
- Prut L, Belsung C. The open-field paradigm to measure the effects of drugs on anxiety-like behaviors: a review. *Eur J Pharmacol.* (2003) 463:3–33. doi: 10.1016/S0014-2999(03)01272-X
- Kuczenski R, Segal DS., Aizenstein ML. Amphetamine, cocaine, and fencamfamine: relationship between locomotor and stereotypy response profiles and caudate and accumbens dopamine dynamics. *J Neurosci.* (1991) 11:2703–12. doi: 10.1523/JNEUROSCI.11-09-02703.1991
- Vollenweider FX, Remensberger S, Hell D, Geyer MA. Opposite effects of 3,4 methylenedioxymethamphetamine (MDMA) on sensorimotor gating in rats versus healthy humans. *Psychopharmacology* (1999) 143:365–72. doi: 10.1007/s002130050960
- Palenicek T, Fujakova M, Brunovsky M, Horacek J, Gorman I, Balikova M, et al. Behavioral, neurochemical and pharmaco-EEG profiles of the psychedelic drug 4-bromo-2,5-dimethoxyphenethylamine (2C-B) in rats. *Psychopharmacology* (2013) 225:75–93. doi: 10.1007/s00213-012-2 797-7
- Vaugeois JM, Bonnet JJ, Duterte-Boucher D, Costentin J. In vivo occupancy of the striatal dopamine uptake complex by various inhibitors does not predict their effects on locomotion. Eur J Pharmacol. (1993) 230:195–201. doi: 10.1016/0014-2999(93)90802-O
- Banjaw MY, Fendt, M, Schmidt WJ. Clozapine attenuates the locomotor sensitisation and the prepulse inhibition deficit induced by a repeated oral administration of Catha edulis extract and cathinone in rats. *Behav Brain Res.* (2005) 160:365–73. doi: 10.1016/j.bbr.2005. 01.002
- Kapur S, Mamo D. Half a century of antipsychotics and still a central role for dopamine D2 receptors. *Prog Neuropsychopharmacol Biol Psychiatry* (2003) 27:1081–90. doi: 10.1016/j.pnpbp.2003.09.004

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2018 Horsley, Lhotkova, Hajkova, Feriancikova, Himl, Kuchar and Páleníček. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.