

AN ADAPTIVE RADIATION OF FROGS IN A SOUTHEAST ASIAN ISLAND ARCHIPELAGO

David C. Blackburn,^{1,2,3} Cameron D. Siler,^{1,4,5} Arvin C. Diesmos,⁶ Jimmy A. McGuire,⁷ David C. Cannatella,⁴ and Rafe M. Brown^{1,4}

¹Department of Ecology and Evolutionary Biology and Biodiversity Institute, University of Kansas, Lawrence, Kansas 66045 ²Current address: Department of Vertebrate Zoology and Anthropology, California Academy of Sciences, San Francisco, California 94118

³E-mail: dblackburn@calacademy.org

⁴Section of Integrative Biology, University of Texas and Texas Natural Science Center, Austin, Texas 78712

⁵Current address: Department of Biology, University of Oklahoma, Norman, Oklahoma 73072

⁶Herpetology Section, Zoology Division, National Museum of the Philippines, Padre Burgos Avenue, Ermita 1000, Manila, Philippines

⁷Department of Integrative Biology, University of California, Berkeley, California 94720

Received June 28, 2011 Accepted April 10, 2013 Data Archived: Dryad doi:10.5061/dryad.dj342

Living amphibians exhibit a diversity of ecologies, life histories, and species-rich lineages that offers opportunities for studies of adaptive radiation. We characterize a diverse clade of frogs (*Kaloula*, Microhylidae) in the Philippine island archipelago as an example of an adaptive radiation into three primary habitat specialists or ecotypes. We use a novel phylogenetic estimate for this clade to evaluate the tempo of lineage accumulation and morphological diversification. Because species-level phylogenetic estimates for Philippine *Kaloula* are lacking, we employ dense population sampling to determine the appropriate evolutionary lineages for diversification analyses. We explicitly take phylogenetic uncertainty into account when calculating diversification and disparification statistics and fitting models of diversification. Following dispersal to the Philippines from Southeast Asia, *Kaloula* radiated rapidly into several well-supported clades. Morphological variation within *Kaloula* is partly explained by ecotype and accumulated at high levels during this radiation, including within ecotypes. We pinpoint an axis of morphospace related directly to climbing and digging behaviors and find patterns of phenotypic evolution suggestive of ecological opportunity with partitioning into distinct habitat specialists. We conclude by discussing the components of phenotypic diversity that are likely important in amphibian adaptive radiations.

KEY WORDS: Comparative methods, disparity, diversification, ecomorphology, Kaloula, microhylidae.

Comparative studies of radiations reveal the commonalities of evolutionary processes that shaped the Tree of Life. By broadly framing evolutionary radiations as the accumulation of both lineages and phenotypes (Losos and Mahler 2010), comparisons among clades become focused not on the absolute number of lineages or specific types of phenotypes but rather on the tempo and mode of speciation and phenotypic evolution; these themes have long been at the heart of studies of adaptive radiations (Simpson 1953; Givnish 1997; Schluter 2000). Studies of vertebrate radiations reveal broad patterns that include the relative importance of different modes of phenotypic diversification, the roles of character displacement and ecological opportunity, and the frequency of hybridization among lineages (Schluter and McPhail 1992; Grant and Grant 1994, 2006; Schluter 2000; Streelman and Danley 2003; Butler and King 2004; Gavrilets and Losos 2009; Glor 2010; Harmon et al. 2010).

2631 © 2013 The Authors. *Evolution* published by Wiley Periodicals, Inc. on behalf of The Society for the Study of Evolution. This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited. *Evolution* 67-9: 2631–2646

Among the diversity of evolutionary radiations, studies of adaptive radiations focus specifically on lineage accumulation and phenotypic breadth. Adaptive radiations are frequently conceptualized as the rapid accumulation of lineages through speciation accompanied by adaptation to diverse ecological niches (Schluter 2000; Gavrilets and Losos 2009; but see Glor 2010), which is typically manifest through a diversity of corresponding phenotypes (e.g., morphologies, life histories, behaviors). During the past three decades, studies of adaptive radiation have come to differentiate the diversity of lineages in a clade (sometimes called "taxonomic diversity") from breadth of phenotypes, or disparity (for review, see Foote 1997; Wagner 2010). By melding advances in time-calibrated molecular phylogenetics with concepts developed within paleobiology, a synthetic view of adaptive radiations emerges that encompasses both extant and extinct taxa. This synthesis has a broad impact by facilitating discovery of unrecognized adaptive radiations and developing a conceptual framework applicable across evolutionary time scales.

Despite long-standing interest in vertebrate adaptive radiations, most clades of living amphibians continue to receive little attention. Whereas many studies have focused on adaptive radiation in mammals (e.g., Simpson 1953; Springer et al. 1997; Alroy 1999; van Valkenburgh 1999; Madsen et al. 2001; Slater et al. 2010), ample opportunity remains for fundamental studies of amphibian radiations. Consider, for example, that extant mammalian species diversity is surpassed by that of living amphibians, including the species-rich clade comprising neobatrachian frogs (Köhler et al. 2005; Reeder et al. 2007; AmphibiaWeb 2012). The lack of attention to adaptive radiations in living amphibians is surprising. Amphibians exhibit a tight correlation between phenotypes and habitat types (Duellman and Trueb 1986; Stebbins and Cohen 1995; McDiarmid and Altig 1999; Wells 2007; Hillman et al. 2009), and have substantial opportunity for population divergence due to limited dispersal abilities and high philopatry (Vences and Wake 2007). Taken together, these suggest that adaptive radiations may be frequently encountered among clades of living amphibians. Recent summaries (i.e., Glor 2010; Losos and Mahler 2010) focus on a single case of nonadaptive radiation in the salamander genus Plethodon (Kozak et al. 2006), but in fact another salamander clade is a celebrated example of adaptive radiation. Tropical plethodontid salamanders (tribe Bolitoglossini sensu Vieites et al. 2011) represent a radiation into diverse phenotypes related to habitat utilization (Wake 1966, 1987, 2009; Wake and Lynch 1976). The adaptive radiation of this clade may have been facilitated by key innovations in both life history (direct development; e.g., Wake 1966; Chippindale et al. 2004) and trophic morphology (Lombard and Wake 1986); for a recent analysis of the relationship between speciation and phenotypic diversity in plethodontids, see Rabosky and Adams (2012). Admittedly, most claims of adaptive radiation in extant amphibians are largely ad

hoc, such as for Asian toads (Bufonidae; van Bocxlaer et al. 2009), Caribbean frogs (Eleutherodactylus; Hedges 1989a,b), Andean frogs (Telmatobius; Cei 1986), and various lineages of ranoid frogs (Bossuyt and Milinkovitch 2000; van der Meijden et al. 2005; Bossuyt et al. 2006; see also Savage 1973). Few studies of amphibian adaptive radiations have explicitly characterized the relationship between habitat utilization and morphological variation, or examined the accumulation of phenotypic diversity through time (but see Kozak et al. 2005; Setiadi et al. 2011). Unlike other vertebrates in which feeding morphologies may play a prominent role in diversification (Liem 1973; van Valkenburgh 1999; Lovette et al. 2002; van Valkenburgh et al. 2004; Westneat et al. 2005; Collar et al. 2008; but see Lombard and Wake 1986), amphibian biologists have historically emphasized life history and reproductive modes as integral to adaptive radiation (Orton 1953, 1957; Wake 1966; Duellman and Trueb 1986; Callery et al. 2001).

Here we characterize a clade of frogs in the Philippine island archipelago as an adaptive radiation. We provide the most complete inference of phylogenetic relationships of frogs in the genus *Kaloula* (family Microhylidae), assess the relationships between habitat specialization and morphological variation, and characterize the accumulation of lineages and phenotypes. *Kaloula* exhibits habitat specialists that we refer to as ecotypes, including terrestrial ground frogs, scansorial shrub frogs, and arboreal tree-hole specialists (Inger 1954; Diesmos et al. 2002). Phylogenetic inference reveals that a single clade of *Kaloula* diversified rapidly following colonization of the Philippines. We demonstrate that morphological variation within *Kaloula* is partly explained by ecotype and that during its diversification the endemic Philippine clade exhibited high and sustained levels of disparification (sensu Evans et al. 2009).

Materials and Methods Taxa, genes, and phylogenetic analyses

The DNA dataset contains extensive population-level sampling of *Kaloula* (140 individuals) from throughout Southeast Asia, and especially the Philippines, for the mitochondrial 12S and 16S ribosomal RNA genes and intervening transfer RNA for valine (\sim 2400 bp). We estimated phylogenetic relationships using maximum likelihood and Bayesian approaches, and estimated relative divergence times using relaxed clock methods implemented in BEAST version 1.6.2 (Drummond et al. 2011). Our approach capitalizes on the resources for genetic and phenotypic analyses obtained through biodiversity surveys and archived in museum collections. Further details are provided in the Supporting Information.

Analyses of lineage diversification and morphological evolution were conducted on a pruned version of the maximum clade credibility tree (MCCT) and the posterior distribution of trees from BEAST analyses, using a single exemplar per species, candidate species, or subspecies (hereafter referred to simply as "species"). Tests of diversification make assumptions about the speciation and extinction of species, not gene lineages. Including multiple individuals per species could lead to erroneously favoring models in which diversification increased toward the present (Pybus and Harvey 2000). Thus, we were careful to not include multiple individuals per putative species. A phylogeny based on a large sequence dataset, which is subsequently pruned, provides a more robust estimate of topology and branch lengths than does a separate analysis based on a limited number of exemplars for each species.

TEMPORAL PATTERNS OF LINEAGE DIVERSIFICATION

We used two approaches to evaluate whether diversification rates were homogenous through time and specifically whether diversification was rapid following colonization of the Philippines. Both incorporated phylogenetic uncertainty by summarizing diversification across the posterior distribution of trees from BEAST analyses.

First, to test whether temporal patterns of lineageaccumulation remained constant through time, we fit multiple models of diversification using the function Misfits (Burbrink et al. 2012) in R version 2.15 for Mac OS X (R Development Core Team 2008). These models are ranked by Akaike information criterion (AIC) weights for each tree and summarized across the set of post–burn-in trees. We evaluated the fit of the nine coalescent-based models of Morlon et al. (2010), each of which is a unique combination of properties characterizing the probabilities of constant or changing diversity, the presence of extinction, and constant or variable rates of speciation and extinction. For Misfits analyses, we used starting parameter values of $\alpha = 0.00001$ (the exponential variation in speciation rate) and $\lambda = 0.05$ (the speciation rate), assuming complete sampling of Philippine *Kaloula*.

In addition, we estimated the γ -statistic for the constant rates test (Pybus and Harvey 2000) using the APE package for R (Paradis et al. 2004). Because extinction may mislead interpretations of the γ -statistic (Rabosky and Lovette 2008; Morlon et al. 2010), our use of the γ -statistic is informed by results of fitting multiple diversification models that incorporate extinction. Significant negative values of γ suggest that internal nodes of the phylogeny are closer to the root than expected under a Yule purebirth model; in this respect, the Yule model (no extinction) is conservative as it would tend to push nodes closer to the root than a model with extinction. If diversification occurred rapidly following colonization of the Philippines (i.e., little extinction), then γ will be significantly negative ($\gamma < -1.645$; Pybus and Harvey 2000). Although estimations of γ are predicted to differ between gene trees and species trees, gene trees likely still provide accurate estimates of γ as long as the population parameter θ is not too large (Burbrink and Pyron 2011). Because estimates of y should ideally incorporate uncertainty in both topology and branch lengths, we calculated γ based on both the pruned MCCT as well as each of the post-burn-in trees from the BEAST analysis. In addition, because the internode distances are affected by the choice of exemplar tips, we evaluated whether γ for the pruned MCCT was significantly different from a tree in which the same number of tips had been pruned at random. Significance for this was assessed by comparing γ to a null distribution obtained by calculating γ on 1000 replicate trees in which the MCCT is randomly pruned to the same number of tips for our exemplar tree of the endemic Philippines clade (pruned 126 tips). Because sampling within endemic Philippine Kaloula is assumed to be near complete, we assume that incomplete taxon sampling does not affect our estimation of the γ -statistic (Pybus and Harvey 2000). Calculations of γ and associated statistical tests were performed using the gammaStat function of the APE package.

ECOTYPES AND MORPHOLOGICAL VARIATION

To provide a framework for interpreting temporal patterns of ecomorphological diversification within *Kaloula*, we evaluated the relationship between ecotype and morphological variation. We characterized each ecotype qualitatively as a summary of information on general ecology and natural history (e.g., obligate forest species, species common in open or disturbed habitats), as well as microhabitat preference (e.g., ground species, species that perch when active, species exclusively found in tree holes; R. M. Brown, C. D. Siler, and A. C. Diesmos, pers. obs.). We also collected morphometric data to describe the shape of each species. Then, we evaluated the relationship between ecotype class and patterns of morphological variation. We classified species of *Kaloula* into three primary ecotype classes: (1) arboreal tree-hole frogs; (2) scansorial shrub frogs; and (3) ground frogs (see Table 1).

We used a multivariate analysis of continuous variables to characterize morphological variation. For each species, we aimed to collect measurement data for 10 specimens; data collection was limited to males because they are more common in museum collections. Because our analysis revealed ample cryptic diversity within Philippine *Kaloula*, we limited our data collection to populations for which we are confident of the species identity. We also collected data for two *Kaloula* species (*Kaloula borealis* and *Kaloula rugifera*) not included in our phylogeny to more fully characterize the relationship between ecotype and morphological variation within the genus. However, we were unable to include in this analysis two new species (from Panay and Sulawesi, respectively), because voucher specimens could not be accessed. Sampling for each species ranged from 3 to 10 specimens (median: 8). If most morphological variation is partitioned

Table 1. Summary of qualitative categorization general geographic range, ecological type, microhabitat preference, reproduction an
activity patterns, morphological specialization for terrestrial/arboreal habits (degree of finer and toe tip expansion), and overall ecotype
defined here (see text for details).

Taxon	Range	General ecology	al ecology Reproductive characteristics icrohabitat and activity patterns		Ecotype
	Range	and interonabilat	and activity patterns	toe up shape	Leotype
K. taprobanica	Sri Lanka	Terrestrial; forest floor, semifossorial; ephemeral pools	Males call in water following heavy rains	Wide	Ground
K. verrucosa	China	Terrestrial; forest floor, semifossorial; ephemeral pools	Males call in water following heavy rains	Narrow	Ground
K. pulchra	Sundaland	Terrestrial; open, disturbed habitat, semifossorial; ephemeral pools	Males call in water following heavy rains	Narrow	Ground
K. mediolineata	Thailand	Terrestrial; forest floor, semifossorial	Males call in water following heavy rains	Narrow	Ground
<i>K</i> . sp. nov.	Vietnam	Semiarboreal; forest floor and tree trunks	Males call in water following heavy rains	Wide	Ground-tree
<i>K</i> . sp. nov.	Peninsular Malaysia	Semiarboreal; forest floor and tree trunks	Males call in water following heavy rains	Wide	Ground-tree
K. baleata	Java	Terrestrial; open, disturbed habitat, semifossorial	Males call in water following heavy rains	Wide	Ground-tree
<i>K</i> . sp. nov.	Palawan	Arboreal; tree hole and branches	Males call from tree holes	Wide	Tree hole
<i>K</i> . sp. nov.	Sulawesi	Terrestrial; open, disturbed habitat, semifossorial	Males call in water following heavy rains	Wide	Ground
K. walteri	S. Luzon	Terrestrial; montane forest floor and dry stream beds, semifossorial	Males call from gravel and rock crevices in dry season	Narrow	Ground
K. rigida	N. Luzon	Terrestrial; forest floor semifossorial; ephemeral pools	Males call in water following heavy rains	Narrow	Ground
<i>K</i> . sp. nov.	Sibuyan	Scansorial; ephemeral pools	Males call in water following heavy rains	Wide	Shrub
K. c. negrosensis	Negros	Scansorial; ephemeral pools	Males call from low elevated perches above water following heavy rains	Wide	Shrub
K. c. meridionalis	Mindanao	Scansorial; ephemeral pools	Males call from low elevated perches or in water following heavy rains	Wide	Shrub
K. c. conjuncta	S. Luzon	Scansorial, ephemeral pools	Males call in water following heavy rains	Wide	Shrub
<i>K</i> . sp. nov.	Mindoro, Semirara	Scansorial; ephemeral pools	Males call from elevated perches above water	Wide	Shrub
K. picta	Philippines	Terrestrial; open, disturbed habitat, semifossorial, ephemeral pools	Males call in water, year round.	Narrow	Ground
<i>K</i> . sp. nov.	Samar- Leyte	Arboreal; tree holes	Males call from tree holes	Wide	Tree hole
<i>K</i> . sp. nov.	Panay	Arboreal; tree holes	Males call from tree holes	Wide	Tree hole
K. kokacii	Luzon	Arboreal, tree holes	Males call from tree holes	Wide	Tree hole
K. kalingensis	N.W. Luzon	Arboreal; tree holes	Males call from tree holes	Wide	Tree hole
<i>K</i> . sp. nov.	E. Luzon	Arboreal; tree holes	Males call from tree holes	Wide	Tree hole

interspecifically, this sampling will likely result in a low type I error rate (Harmon and Losos 2005).

We measured 15 continuous characters that are relevant to ecomorphology (all measured by D.C.B. to reduce interobserver bias; Lee 1990; Hayek et al. 2001; for further details, see Supporting Information). Frog limb morphology is directly related to performance relevant to ecology (e.g., Emerson 1991). Limb proportions of the fore and hind limbs are intimately related to locomotor mode with walking and burrowing species having shorter limbs and digits than those that jump and climb (Emerson 1988). Similarly, the anatomy of the hands and feet plays an important role in different locomotor modes: foot webbing is important to propulsion in water (Stahmhuis and Nauwelaerts 2005); enlarged metatarsal tubercles on the feet facilitate digging (Emerson 1971, 1976); and expanded digit tips on the hands and feet support larger toe pads that increase surface adhesion for climbing (Emerson and Diehl 1980). Finally, head shape can explain some patterns of dietary type in frogs (Emerson 1985). For each taxon, we calculated the mean value for each measurement to maximize interspecific variation relative to intraspecific variation (Harmon and Losos 2005).

We evaluated the relationship between ecotype and measurement data using a multivariate analysis of variance (MANOVA). We asked whether ecotype classes could be discriminated based on observed morphological variation for the twenty species considered. To focus this analysis on differences in shape, we followed standard procedures (e.g., Garland et al. 1992; Collar and Wainwright 2006; Revell 2009; Mahler et al. 2010) to "sizecorrect" fourteen measurements by calculating their residuals based on a generalized least squares linear regression (see Hansen and Bartoszek 2012) of each variable against snout-vent length (SVL; measure of body size) using the NLME package for R. We conducted a principal components analysis (PCA) based on SVL and the 14 residuals. Principal components analysis was conducted using the prcomp function of the STATS package for R and eigenvalues explaining $\geq 10\%$ of the cumulative variance saved for analysis. We did not incorporate phylogenetic information into this analysis because we aimed to test the relationship observed in the field between ecotype and form that spurred this study, and not the influence of phylogeny per se. We then conducted a MANOVA with ecotype class as the factor (using the manova function of the STATS package in R); significance was assessed using the Wilks' statistic. We also conducted a separate analysis using a simulation-based phylogenetic ANOVA (Garland et al. 1993) as implemented in the phy.anova function in the GEIGER package in R (Harmon et al. 2008); 1000 simulations were used to calculate the phylogenetic P-value. These analyses used the same ecotype classes and the In-transformed measurement data (excluding snout-vent length) as above, as well as the pruned

species-level MCCT from our BEAST analysis; we excluded the two mainland species not included in our phylogenetic analyses.

TEMPORAL PATTERNS OF MORPHOLOGICAL DIVERSIFICATION

Using phylogenetic comparative methods, we evaluated temporal patterns of morphological change across the evolutionary history of Kaloula. Measurements of morphological features related to ecotype are confounded by both body size and shared phylogenetic history. These confounding effects should be accounted for before evaluating temporal patterns of morphological change across a phylogeny. Using the pruned species-level MCCT from our BEAST analysis (representing 18 of the 20 species measured earlier), we performed a phylogenetic principal component analysis on phylogenetically size-corrected shape variables using the method of Revell (2009). By accommodating the nonindependence of observations due to shared evolutionary history, this approach reduces variance of regression coefficients so as to be closer to expected type I error rates (Felsenstein 1985; Rohlf 2006). Principal component axes with eigenvalues comprising >10% of the cumulative variance were used to represent a morphospace. Species values for these phylogeny-corrected principal components scores (PC_{phylo}) provided the data for analyses of morphological evolution using comparative phylogenetic methods.

Temporal patterns of morphological diversification were evaluated using the disparity-through-time (DTT) approach (Harmon et al. 2003). We calculated total disparity as the average Euclidean distance between all points in a morphospace (Ciampaglio et al. 2001), whereas relative disparities for subclades (Foote 1993) were calculated as the disparity in the subclade divided by the total disparity. High relative disparity of a subclade indicates a relatively large contribution to total observed disparity. Using an estimate of phylogeny, the DTT approach calculates mean relative disparity for all subclades in which ancestral lineages are present at times defined by each node in the ultrametric phylogenies (Harmon et al. 2003). Thus, similar to a lineagethrough-time (LTT) plot, mean relative disparity for each node is used to generate a DTT plot. For these DTT analyses, we used the PC_{phylo} scores for each species and the MCCT from BEAST. To evaluate whether the observed DTT differs from expectations under a null hypothesis of Brownian motion character change, we conducted 10,000 simulations of character change; 95% confidence intervals (CI) were constructed for these simulated data (Slater et al. 2010). Analyses were conducted using the GEIGER package. The DTT curve for the observed data was evaluated qualitatively by comparison to the 95% CI for the simulated data as well as plots of average subclade disparity by node for each of the BEAST trees. We quantitatively evaluated the DTT curve by calculating the morphological disparity index (MDI; Harmon et al. 2003). The MDI is the area contained between curves of the observed relative disparity and the median relative disparity from simulations. Values of MDI greater than 0 indicate that most morphological disparity is partitioned within subclades, whereas values less than 0 indicate that most morphological disparity is partitioned between subclades. Although DTT curves and the MDI statistic allow for inferences of phenotypic evolution that circumvent issues related to inferring ancestral states (Harmon et al. 2003), these methods may still be sensitive to uncertainties in topology and branching times. Thus, we calculated MDI for each tree in the posterior samples from BEAST analyses as above for the MCCT (though using only 1,000 simulations for calculating MDI for each tree); for each tree, we calculated MDI using scores for the first three PCphylo axes based on the variancecovariance matrix for that tree. We then obtained a distribution for MDI and calculated a 95% credible interval. Finally, we evaluated whether patterns of evolution differed along each PCphylo by performing all of the above DTT analyses on each PC_{phylo} separately. We then tested the fit of two models of character evolution to each PC_{phylo}: a random walk (modeled as Brownian motion [BM]) and a random walk with a selective optimum (modeled as Ornstein-Uhlenbeck process [OU]; Hansen 1997; Butler and King 2004). This combination of approaches identified whether changes in specific axes of morphological variation were associated with bursts of lineage diversification.

Results

PHYLOGENETIC RELATIONSHIPS

In general, the ingroup relationships among Southeast Asian Kaloula are well resolved (Figs. 1, S1, S2). Both Bayesian and likelihood methods revealed strong support (PP = 1.00; ML NBS = 98%) for a large clade consisting of 12 divergent lineages endemic to the Philippines (Figs. 1, S2; Table 2). Divergence time analyses suggest that an MRCA for this Philippines clade occurred in the Late Miocene (median: 11.9 mya; 95% HPD: 6.1-18.4 mya; see Supporting Information). Individuals from within this endemic Philippines radiation are assigned to five well-resolved clades: (1) Kaloula picta; (2) an undescribed species from Samar and Leyte islands; (3) Kaloula rigida + Kaloula walteri; (4) the "Kaloula kalingensis" clade (consisting of K. kalingensis, Kaloula kokacii, and two undescribed species from Panay Island and eastern Luzon Island, respectively); and (5) the "Kaloula conjuncta" clade (consisting of K. conjuncta conjuncta, Kaloula c. meridionalis, Kaloula c. negrosensis, Kaloula c. stickeli, and two undescribed taxa, one from Mindoro Island and another from Panay and Sibuyan islands). However, both Bayesian and ML analyses provide equivocal support (< 0.50 PP; < 50% ML NBS) for specific relationships between these five clades.

TEMPORAL PATTERNS OF LINEAGE DIVERSIFICATION

Rapid lineage accumulation followed colonization of the Philippines. The best-fit models of diversification for the Philippines radiation are those in which diversification varies through time (Table 3). Of the coalescent-based models, the best-fit models are those with time-varying rates of speciation, with most trees fitting a pattern of expanding diversity, time-varying speciation, and no extinction (Model 6 of Morlon et al. 2010). The observed γ for the endemic Philippines radiation is strongly negative (γ for MCCT: -3.096; 95% credibility interval: -3.874, -2.181). This value of γ is significantly different than expected given random pruning the complete phylogeny (P < 0.01).

CORRELATION OF ECOTYPE AND MORPHOLOGICAL VARIATION

Principal components analysis on the size-corrected data resulted in three PC axes that each accounted for $\geq 10\%$ of the variation. Together, the first three PC axes account for 84% of the variance (PC1: 53.1%; PC2: 20.2%; PC3: 10.6%). The strongest loadings on PC1 (Fig. 2; Table 4) reveal a strong inverse correlation of the widths of finger and toe tips and the length of inner and outer metatarsal tubercles. The strongest loadings on PC2 reveal an inverse correlation between body size (SVL) and the extent of pedal webbing (with larger species having less webbing), whereas the strongest loadings on PC3 indicate a contrasting pattern with body size (SVL) being positively correlated with the extent of pedal webbing and inversely correlated with the size of the inner metatarsal tubercles. Variation in only six of 15 measurements (SVL, finger and toe tip widths, length of the inner and outer metatarsals, and extent of pedal webbing) contributes substantially to explaining patterns of interspecific morphological variation. Ecotype significantly explains patterns of variation in the morphological data. The MANOVA with ecotype as a factor is significant based on species scores on the first three PC axes (Wilks' $\lambda = 0.0650$; P < 0.0001). Examination of scatter plots of PC scores also reveals general correspondence of ecotype with particular regions of morphospace (Fig. 2). The phylogenetic MANOVA is marginally nonsignificant (phylogenetic P = 0.0890), although this analysis included only eighteen of the twenty species that were included in the nonphylogenetic MANOVA.

TEMPORAL PATTERNS OF MORPHOLOGICAL DIVERSIFICATION

In the principal components analysis using the variance– covariance matrix from the phylogeny (MCCT), the first three principal component axes (PC_{phylo}) each have eigenvalues accounting for more than 10% (12%–31%) of the cumulative variation (Table 5). Together these three PC_{phylo} axes explain



Figure 1. Maximum-likelihood phylogram estimated from mitochondrial DNA sequences (12S and 16S ribosomal RNA genes) depicting the phylogenetic relationships of *Kaloula* (Anura: Microhylidae; see also Fig. S1). The gray box indicates the endemic Philippine radiation. Images of each species in the Philippine radiation are provided, except for the new species from Panay.

approximately 70% of the cumulative variation and general patterns of relative loadings are similar to analyses earlier.

Disparity-through-time analyses conducted using the species scores on the PC_{phylo} axes result in different estimates of MDI depending on whether each PC_{phylo} was analyzed separately or all three together (Table 6). Disparity-through-time plots for the scores on the first three axes reveal a significant increase in disparity during a period of time of active cladogenesis within the Philippines radiation (Fig. 3). During this period, average subclade disparity exceeds disparity of the clade as a whole, indicating that species within these subclades are more different from each other, on average, than species in the group as a whole. Separate analysis of scores on each PC_{phylo} axis reveals that this increase in morphological disparity is driven largely by $PC1_{phylo}$. Visual inspection of the DTT plots for each PC_{phylo} reveals differences in the patterns of morphological evolution (Fig. 3). Fitting of BM and OU models of character evolution to these PC_{phylo} using the MCCT further supports these components as representing different evolutionary patterns (Table 6). Observed disparity for $PC1_{phylo}$ is consistent with an OU model of character

Taxon	PP	ML NBS	<i>p</i> -distance
K. pulchra	1.00	100%	0.5%
K. mediolineata	1.00	100%	1.1%
K. sp. nov. Vietnam	1.00	100%	0.2%
K. sp. nov. Palawan	1.00	100%	0.3%
K. baleata Bali and Java	1.00	100%	0.8%
K. sp. nov. Peninsula and	1.00	100%	0.8%
Borneo			
K. sp. nov. Sulawesi	0.42	37%	1.1%
K. sp. nov. Samar and Leyte	1.00	100%	1.3%
K. walteri	1.00	100%	0.3%
K. rigida ¹	1.00	100%	0.5%
K. sp. nov. Panay	1.00	100%	0.0%
K. kalingensis	1.00	100%	2.3%
K. kokacii	1.00	100%	2.9%
K. sp. nov. East Luzon	0.97	75%	3.2%
K. picta	1.00	100%	0.2%
K. sp. nov. Sibuyan	1.00	75%	0.8%
K. conjuncta conjuncta	1.00	100%	0.7%
K. conjuncta meridionalis	1.00	99%	0.1%
K. conjuncta negrosensis	1.00	100%	0.0%

Table 2. Bayesian posterior probabilities (PP), maximum likelihood nonparametric bootstrap support, and average within-clade *p*-distances for recovered clades of *Kaloula*.

¹Calculations exclude ACD 1570.

change, whereas patterns are more equivocal for $PC2_{phylo}$ and $PC3_{phylo}$ (Table 6), although the 95% CIs on the DTT plots (Fig. 3) demonstrate that observed patterns of morphological change on these two axes may be consistent with BM models.

Discussion diversification and disparification

Following colonization of the Philippines island archipelago possibly in the Late Miocene (see Supporting Information), *Kaloula* rapidly diversified into several well-supported clades, each characterized by one of three distinctive ecotypes. In general, the best-fit coalescent models of diversification for the Philippines radiation are those with increasing diversity and changes in the speciation rate through time. With 12 species-level lineages, the species richness of *Kaloula* endemic to the Philippines is approximately equal to that found elsewhere in southern Asia (including the four undescribed species in the *baleata* clade). All ecotypes found within the Philippines radiation are also found in species of *Kaloula* not included in this radiation, which indicates convergent evolution. Because the Philippines radiation diversified into ecotypes found also in species of *Kaloula* from mainland Asia, this suggests constraints on ecotype diversity and/or the existence of several ecomorphological "optima."

During diversification, Philippine Kaloula exhibited high levels of disparification. The DTT analyses indicate that the disparity accumulated during the Philippines radiation lies outside of the 95% CI based on expectations using a BM model (Fig. 3). This inference appears robust to phylogenetic uncertainty (Figs. 4, S3). Furthermore, the morphological disparity index for PC1-3_{phylo} (MDI for MCCT: 0.311; 95% credibility interval: 0.118-0.586) indicates that disparity during the evolution of Kaloula has been on average greater than expected. During the Philippines radiation, accumulation of disparity along a single axis (PC1_{phylo}) exceeds the total average subclade disparity of Kaloula (Fig. 3). Morphological evolution along this axis is bestfit by an OU model (a random walk with a selective optimum) rather than by a Brownian model, and represents a contrast between traits related to climbing and digging. Support for an OU model of change provides evidence of a selective optimum for this suite of traits and the loading on traits related to climbing (length and width of digits and digit tips) and digging (lengths of metatarsal tubercles) is suggestive of morphological tradeoffs involved in vertically partitioning habitats during ecotype evolution. The patterns for PC2_{phylo} and PC3_{phylo} are more equivocal.

ECOLOGICAL OPPORTUNITY AND BIOGEOGRAPHY

The pattern observed for Philippine *Kaloula* supports the suggested role of ecological opportunity as a driving force underlying adaptive radiations (Worthington 1937, Lack 1940; Simpson 1953; Rensch 1959; see also Sepkoski 1984), particularly in island

Table 3. Tests of coalescent-based models of diversification of Morlon et al. (2010). The properties of these models (saturated vs. expanding diversity; constant or varying rates; and extinction) are given; models 4a–d differs in ways in which speciation and extinction are modeled (for details, see Morlon et al. 2010). The two best-fit sets of models are ranked by mean weighted AIC scores. For each model, the frequency of models (as proportions) across the posterior distribution from BEAST analyses are given.

	Model 1	Model 2	Model 3	Model 4a	Model 4b	Model 4c	Model 4d	Model 5	Model 6	AIC w	SD
Diversity	Saturated	Saturated	Expanding								
Rates	Constant	Varying	Constant	Varying	Varying	Varying	Varying	Constant	Varying		
Extinction	Positive	None	None								
Best-fit	0.0	16.7%	0.0	0.0	0.0	0.0	0.0	2.7%	75.2%	0.56	0.10
Next best	0.0	73.3%	0.0	0.0	0.0	0.0	0.0	2.0%	19.2%	0.32	0.32



Figure 2. Relationship between morphological variation and ecotype categories in *Kaloula*. Species scores for the first two principal components (PC1 and PC2) are plotted; see Table 4 for loadings and percent variance explained.

systems (Mayr 1963; Mahler et al. 2010). Although other anuran taxa may already have occupied the Philippines when *Kaloula* arrived, there are few other microhylid frogs that might have limited feeding and foraging opportunities for *Kaloula* (Inger 1954;

Table 4. Loadings and variance explained by principal component analysis on size-corrected shape variables.

PC1	PC2	PC3
-0.25	-0.30	-0.45
0.02	-0.07	-0.05
0.12	0.05	0.15
0.06	0.13	0.08
0.07	0.15	-0.07
0.19	0.17	0.16
0.61	0.14	-0.21
0.07	0.09	0.13
0.09	0.09	0.16
0.08	0.16	0.15
0.15	0.10	0.22
0.36	0.18	0.19
-0.36	0.14	0.54
-0.37	0.20	0.23
0.24	-0.82	0.46
0.53	0.20	0.11
	PC1 -0.25 0.02 0.12 0.06 0.07 0.19 0.61 0.07 0.09 0.08 0.15 0.36 -0.36 -0.36 -0.37 0.24 0.53	PC1 PC2 -0.25 -0.30 0.02 -0.07 0.12 0.05 0.06 0.13 0.07 0.15 0.19 0.17 0.61 0.14 0.07 0.09 0.09 0.09 0.09 0.09 0.036 0.18 -0.36 0.14 -0.37 0.20 0.24 -0.82 0.53 0.20

Alcala and Brown 1998; Brown 2007). The derived feeding system of microhylid frogs is based on a muscular hydrostatic tongue that can be aimed side-to-side without moving the head (Meyers et al. 2004); this is shared only with two African frog families

Table 5. Loadings and variance explained by phylogenetic principal component analysis on phylogenetically size-corrected shape variables.

Trait	PC1 _{phylo}	PC2 _{phylo}	$PC3_{phylo}$
Snout-vent length	-0.17	0.58	0.31
Head width	-0.23	-0.59 ·	-0.63
Snout length	-0.79	-0.04 ·	-0.34
Forearm length	-0.27	0.74	-0.11
Third finger length	-0.44	0.48	0.23
Third finger width	-0.88	0.18	0.34
Third finger-tip width	-0.88	-0.26	0.17
Thigh length	-0.12	0.82 .	-0.41
Crus length	-0.25	0.70 ·	-0.63
Third toe length	-0.40	0.65	-0.24
Third toe width	-0.67	-0.07	0.45
Third toe-tip width	-0.88	-0.08	0.23
Inner metatarsal tubercle lengt	h 0.55	0.57	0.15
Outer metatarsal tubercle lengt	h 0.69	0.38	0.36
Webbing	-0.15	-0.74	-0.19
Variance explained	0.32	0.28	0.13

	MDI	95% CI	Range	Median	BM ln L	OU ln L	BM AICc	OU AICc	ΔAICc	BM w	OU w
PC1 _{phylo}	0.436	0.230-0.675	0.109-0.970	0.420	-73.686	-70.576	152.229	148.998	3.231	0.199	0.801
PC2 _{phylo}	0.311	0.109-0.535	-0.007 - 0.836	0.303	-72.465	-69.768	149.788	147.383	2.405	0.300	0.700
PC3 _{phylo}	0.096	0.058-0.337	-0.032 - 0.497	0.166	-65.328	-64.680	135.513	137.206	1.693	0.429	0.571
$PC1-3_{phylo}$	0.312	0.200-0.457	0.118-0.586	0.323	_	_	_	-	_	-	-

Table 6. Morphological disparity index (MDI) values from disparity-through-time analyses and log-likelihoods, AICc, and Akaike weights (*w*) of Brownian motion (BM) and Ornstein-Uhlenbeck (OU) models of character evolution for each PC_{phylo} axis and all three together for MCCT from BEAST analysis.

Hemisotidae and Brevicipitidae (Nishikawa et al. 1999; Meyers et al. 2004), and skeletal features unique to the family Microhylidae may be related to this feeding system (Trueb et al. 2011). Furthermore, microhylids tend to specialize on small prey, especially ants and termites (Emerson 1971; Das 1996; Parmalee 1999; Wells 2007). The feeding system and ecology of microhylid frogs suggest that ecological opportunities for this family might differ from that of other anurans, including in the Philippines. Aside from Kaloula, there are few microhylid species in the Philippines. Both Chaperina fusca and Kalophrynus pleurostigma are found on islands and the mainland of the continental Sunda Shelf; both species occur on the Mindanao Pleistocene Aggregate Island Complex (PAIC; Brown and Diesmos 2009) and C. fusca is also found on Palawan Island. Two species of Oreophryne are endemic to the Mindanao PAIC. A single species of Microhyla is only present on the southern island of Tawi-tawi. The endemic Philippine radiation of Kaloula occurs on six PAICs (Table 7), and co-occurs with these other microhylids only on the Mindanao and Palawan PAICs. Furthermore, humans might have facilitated the dispersal of the only species (K. picta) in the Philippine radiation that occurs on Palawan Island. Thus, it seems likely that early in the endemic Philippine radiation species of Kaloula potentially competed with other microhylid frogs only on the Mindanao PAIC. If indeed the ecological opportunities available to microhylids do differ from those of other anurans, then there may have been substantial opportunity for diversification within Kaloula following colonization of the archipelago.

We find both a strong correlation between ecotype and morphological diversity and a partitioning of ecotypes into distinct clades during the Philippine radiation. This suggests that these three ecotypes arose early following dispersal into the archipelago with subsequent speciation predominantly mediated by allopatric isolation coupled with conservation of each clade's ancestral ecotype. Within ecotypes of the Philippine radiation, it is clear that dispersal occurred across oceanic barriers: tree-hole frogs occur on three PAICs; shrub frogs on four PAICs; and the broad distribution of the ground frog *K. picta* on many PAICs implies high levels of dispersal, and potentially human-mediated range expansion facilitated by conversion of forests to agriculture (Brown et al. 2010). The remaining ground frogs, *K. rigida* and *K. walteri*, are restricted to the Luzon PAIC. There is a rough positive relationship between PAIC size and the number of ecotypes present, with Luzon, Mindanao, and Negros-Panay PAICs possessing the greatest ecotype diversity (Table 7). Yet, there is no obvious order of colonization of the islands, as observed in other systems (i.e., Hawaiian islands; Roderick and Gillespie 1998; Hormiga et al. 2003).

The relationship between phenotype and colonization ability is of interest in studies of both invasive species (Kolar and Lodge 2001) and radiations in island systems (Losos 1992; Schluter et al. 1997; Gillespie 2004; Cristescu et al. 2010), with recent work drawing parallels between these (Poe et al. 2011). Although ancestral state reconstructions for the ecotype of the MRCA of the endemic Philippine radiation are ambiguous (see Supplementary Materials), limited evidence suggests its MRCA may have been a ground frog. First, the most common ecotype outside of the endemic Philippine radiation is a ground frog (Table 1). Second, it is clear that ground frogs accomplished two other dispersal events across oceanic barriers. These include the colonization of Palawan by the *Kaloula baleata* clade, which are all ground frogs except for the undescribed species endemic to Palawan, and the dispersal of the ground frog *K. picta* throughout the Philippines.

Although there is no evidence for further ecotype diversification following the initial partitioning of ecotypes by clade, morphological evolution within these subclades is above average relative to the total observed disparity in Kaloula. This indicates continued phenotypic diversification subsequent to the evolution of distinct ecotypes, although it remains difficult at present to evaluate the significance of within-ecotype diversification. Although ecotypes cluster in the phylogeny, species of Kaloula forming a community at any given locality in the Philippines are not close relatives. For example, communities at sites on the Luzon PAIC are composed of K. picta (large ground frog), K. conjuncta conjuncta (shrub frog), a kalingensis clade species (treehole frog), and a rigida clade species (small ground frog). Thus, communities of Kaloula species appear to exhibit phylogenetic overdispersion of structural habitat specialist phenotypes (e.g., Emerson and Gillespie 2008).

Phenotypic evolution in the Philippines radiation of *Kaloula* is potentially consistent with more than one pattern of phenotypic



Figure 3. Disparification through relative time in *Kaloula*. Top-left panel shows MCCT chronogram (with shapes corresponding to ecotypes of Fig. 2) and top-right panel shows lineage-through-time plot. Middle and bottom panels show disparity-through-time (DTT) plots for PC1_{phylo}, PC2_{phylo}, PC3_{phylo}, and all three axes together (PC1–3_{phylo}). Solid black lines on DTT plots is observed disparity based on MCCT, gray lines and polygons represent median and 95% confidence intervals from BM simulations, respectively.



Figure 4. Uncertainty in disparification through time in *Kaloula* for PC1_{phylo}. Solid black line on DTT plot is observed disparity based on MCCT; gray polygons represent 95% confidence interval for observed disparity based on post–burn-in trees from BEAST analysis.

change. The evolution of morphological diversity along $PC1_{phylo}$ (Fig. 3) exhibits high levels of variation within subclades and is best-fit by an OU model of change (Table 6). By incorporating selection and drift, support for the OU model provides evidence for stabilizing selection and/or phenotypic constraints (e.g., Lande 1976; Felsenstein 1988; Martins 1994; Hansen 1997; Butler and King 2004; Harmon et al. 2010). Thus, our analysis may provide support that phenotypic diversification in the Philippine radiation was at least partially associated with selective optima related to ecotypes.

ADAPTIVE RADIATIONS IN AMPHIBIANS

Given the general lack of detailed studies of adaptive radiations in amphibians, it is not surprising that few patterns have emerged

specific to amphibians. Although diversification into three ecotypes may not constitute an unusual diversity of adaptive forms (e.g., Warheit et al. 1999; Losos and Miles 2002), phenotypic diversification related to habitat utilization is only one component of an adaptive radiation. We suspect that the axes of disparification in amphibians may differ from those of other vertebrate radiations. The Philippine radiation of Kaloula satisfies the first stage of the "general vertebrate model" of adaptive radiation, with initial diversification into habitat specialists followed by morphological diversification along other axes (Streelman and Danley 2003; see summary in Glor 2010). For many vertebrates, one important axis of subsequent disparification may be related to trophic biology, including diversity of feeding morphologies (Liem 1973; Lovette et al. 2002; van Valkenburgh et al. 2004; Westneat et al. 2005), or strategies related to behaviorally partitioning habitats (e.g., Losos 2009). Although the Philippine radiation of Kaloula might have had an ecological opportunity for colonization and diversification because of differences in feeding from other Philippine frogs, there is no indication of differences among these species of Kaloula. In fact, most adult anuran amphibians are believed to be fairly similar in trophic biology by being both predatory and feeding generalists (Nishikawa 2000). Other than the rare example of robust skulls of snail-eating frogs (Drewes and Roth 1981) and the general observation that species specializing on ants have relatively narrower heads (Toft 1980, 1981; Emerson 1985), there has been little study of the dietary relevance of morphological diversity in structures such as the tongue (Emerson 1985; Grant et al. 1997; but see Trueb and Gans 1983). Resource partitioning is documented in anurans, but this may be due more to differences in habitat and less associated with selectivity of prey types (Toft 1985).

We suggest that for amphibians, especially the order Anura (i.e., frogs and toads), partitioning of the environment in relation to reproductive modes and life histories plays an important role in adaptive radiation. This is suggested both by past discussions of radiations in amphibians that emphasize the importance of life history (Orton 1953, 1957; Duellman and Trueb 1986), and

Table 7. Distribution of ecotypes by Philippine Aggregate Island Complex (PAIC).

PAIC	PAIC area (km ²)	Ecotypes	No. Taxa	Таха
Luzon	147,451	Ground, shrub, tree-hole	7	K. c. conjuncta; K. kalingensis; K. kokacii; K. rigida; K. walteri; K. picta; K. sp. nov. East Luzon
Mindanao	175,430	Ground, shrub, tree-hole	3	<i>K. c. meridionalis; K. picta; K.</i> sp. nov. Samar and Leyte
Mindoro	13,009	Ground, shrub	2	K. c. conjuncta; K. picta
Negros-Panay	59,623	Ground, shrub, tree-hole	3	K. c. negrosensis; K. picta; K. sp. nov. Panay
Palawan	61,198	Ground, tree-hole	2	K. picta; K. sp. nov. Palawan
Romblon	1,407	Ground, shrub	2	K. picta; K. sp. nov. Sibuyan

observations of clades with high levels of life history diversity (Wells 2007). Two promising lines of future inquiry on adaptive radiations in anurans are evaluating (1) the relationships between lineage accumulation and the diversity of reproductive modes and life histories (Gomez-Mestre et al. 2012); and (2) the relationship between diversification and either resource partitioning among tadpoles (Heyer 1973, 1974; Toft 1985) or by different reproductive modes (Salthe and Duellman 1973).

Conclusions

Across frog diversity, it is difficult to characterize any clade as having an unusually great diversity of adaptive forms. Yet, interestingly, the same forms have evolved again and again throughout the > 200 million year history of crown-group Anura. It is clear that there is convergence in ecomorphological form across vast spans of time, with many last sharing a common ancestor more than 50 million years ago and some more than 200 million years ago (Roelants et al. 2007). Examples of remarkable convergence in morphology among taxa that last shared a common ancestor in the Mesozoic include aquatic frogs such as Barbourula (Bombinatoridae) and Conraua (Conrauidae), large terrestrial frogs such as Leptobrachium (Megophyridae) and Astylosternus (Arthroleptidae), small aposematic terrestrial frogs such as the diverse species in the Dendrobatidae and Mantellidae, generalized terrestrial frogs in Strabomantidae and Ceratobatrachidae, burrowing "spade-foot" frogs such as Pelobates (Pelobatidae) and Heleioporus (Limnodynastidae), and fossorial frogs such as Rhinophrynus (Rhinophrynidae), Hemisus (Hemisotidae), and Nasikabatrachus (Nasikabatrachidae). Because of their long evolutionary history, ecomorphological diversity, and life-history characteristics, frogs represent a remarkable research opportunity for studying how the same suites of phenotypes might have evolved at different periods during the Mesozoic and Cenozoic. Studying phenotypic evolution within discrete clades such as our study of the genus Kaloula or even within families (such as the Mantellidae endemic to Madagascar) will be fruitful. However, an even broader net should be cast to examine patterns of convergence and disparification across many clades as there have likely been many periods of rapid diversification and disparification in anurans in both the Mesozoic and Cenozoic. Furthermore, although broad categories of form are recognizable and possibly related to locomotion, diet, and habitat type (Duellman and Trueb 1986; Emerson 1988; Hillman et al. 2009), there is a need for field studies that explicitly relate performance and fitness to morphological and life history features characteristic of the diverse frog ecotypes. As phylogenetic resolution improves within the anuran Tree of Life, studies of frogs promise many opportunities for understanding phenotypic diversification and the partitioning of environments in relation to reproductive modes and life history.

ACKNOWLEDGMENTS

This work is funded by the National Science Foundation grants DEB 0073199, DEB 0743491, and EF 0334952 to RMB,; DEB 0804115 to CDS and RMB; Fulbright and Fulbright-Hayes Fellowships to CDS; DEB 0334952 and DEB 0206729 to DCC; and DEB 0328700 and DEB 0640967 to JAM. RMB thanks the Society for the Study of Amphibians and Reptiles (grants-in-aid fund), American Society of Ichthyologists and Herpetologists (Gaige Fund), and Society of Systematic Biologists (Graduate Student Research Grant) for support of Philippine fieldwork. The Department of Integrative Biology, University of California (Berkeley), and the University of Kansas Biodiversity Institute provided additional logistical and financial support. The authors thank B. Fernandez and J. Fernandez, V. Yngente, and E. Rico for assistance with fieldwork. The authors the Philippine Department of Environment and Natural Resources, Protected Areas and Wildlife Bureau providing research permits, and logistical support critical for this and related studies. The authors are grateful to the following institutions and people for providing loans of specimens and genetic samples: M. Meegaskumbura, Royal Ontario Museum (R. Murphy), Field Museum of Natural History (H. Voris, R. Inger, A. Resetar, and B. Stuart), the Smithsonian (R. Crombie, K. de Queiroz), California Academy of Sciences (J. Vindum, R. Drewes, and A. Leviton), National Museum of the Philippines (R. Sison and V. Papal-latoc), La Sierra University (L. Grismer), National Museum of Natural Science, Taiwan (W.-H. Chou), Forest Research Institute of Malaysia (N. Yaakob), University of Malaysia (I. Das and G. Gee), Philippine Endemic Species Conservation Foundation (M. Gaulke and E. Curio), and the Texas Natural History Collection and Genetic Diversity Collection of the University of Texas at Austin (T. LaDuc). C. Oliveros and N. Barve provided the estimates of PAIC size used in Table 7. The authors thank J. Bollback, F. Burbrink, L. Harmon, D. Mahler, A. Pyron, D. Rabosky, and G. Slater for discussion regarding analyses, and F. Burbrink, A. Pyron, and G. Slater for each sharing R code in development. The authors thank B. Evans, J. Losos, D. Mahler, L. Harmon, J. Esselstyn, and the Brown and McGuire lab groups for a number of thoughtful discussions that contributed to the final form of this research.

LITERATURE CITED

- Alcala, A. C., and W. C. Brown. 1998. Philippine Amphibians: an illustrated field guide. Bookmark Inc., Makati City, Philippines, 116 pp.
- Alroy, J. 1999. The fossil record of North American mammals: evidence for a Paleocene evolutionary radiation. Syst. Biol. 48:107–118.
- AmphibiaWeb. 2012. Available at http://amphibiaweb.org. Accessed November 1, 2012.
- Bossuyt, F., and M. C. Milinkovitch. 2000. Convergent adaptive radiations in Madagascan and Asian ranid frogs reveal covariation between larval and adult traits. Proc. Natl. Acad. Sci. USA 97:6585–6590.
- Bossuyt, F., R. M. Brown, D. M. Hillis, D. C. Cannatella, and M. C. Milinkovitch. 2006. Phylogeny and biogeography of a cosmopolitan frog radiation: Late Cretaceous diversification resulted in continentscale endemism in the family Ranidae. Syst. Biol. 55:579–594.
- Brown, R. M. 2007. Introduction to Robert F. Inger's systematics and zoogeography of Philippine Amphibia. Forward to the reprint of Inger's 1954 monograph. Pp. 1–17 in R. F. Inger. Systematics and zoogeography of Philippine Amphibia. Natural History Publications, Kota Kinabalu, Malaysia.
- Brown, R. M., and A. C. Diesmos. 2009. Philippines, biology. Pp. 723–732 in R. Gillespie and D. Clague, eds. Encyclopedia of islands. University of California Press, Berkeley, CA, 1111 pp.
- Brown, R. M., C W. Linkem, C. D. Siler, J. Sukumaran J. A. Esselstyn, A. C. Diesmos, D. T. Iskandar, D. Bickford, B. J. Evans, J. A. McGuire,

et al. 2010. Phylogeography and historical demography of *Polypedates leucomystax* in the islands of Indonesia and the Philippines: evidence for recent human-mediated range expansion? Mol. Phylogenet. Evol. 57:598–619.

- Burbrink, F. T., and R. A. Pyron. 2011. The impact of gene-tree/species-tree discordance on diversification rate estimation. Evolution 65:1851–1861.
- Burbrink, F. T., X. Chen, E. A. Myers, M. C. Brandley, and R. A. Pyron. 2012. Evidence for determinism in species diversification and contingency in phenotypic evolution during adaptive radiation. Proc. R. Soc. B. 279:4817–4826.
- Butler, M. A., and A. A. King. 2004. Phylogenetic comparative analysis: a modeling approach for adaptive evolution. Am. Nat. 164:683–695.
- Callery, E. M., H. Fang, and R. P. Elinson. 2001. Frogs without polliwogs: evolution of anuran direct development. BioEssays 23:233–241.
- Cei, J. M. 1986. Speciation and adaptive radiation in Andean *Telmatobius* frogs. Pp. 374–386 *in* F. Vuilleumier and M. Monasterio, eds. High altitude tropical biogeography, Oxford Univ. Press, Oxford, U.K.
- Ciampaglio, C. N., M. Kemp, and D. W. McShea. 2001. Detecting changes in morphospace occupation patterns in the fossil record: characterization and analysis of measures of disparity. Paleobiology 27:695–715.
- Chippindale, P. T., R. M. Bonett, A. S. Baldwin, and J. J. Wiens. 2004. Phylogenetic evidence for a major reversal of life-history evolution in plethodontid salamanders. Evolution 58:2809–2822.
- Collar, D. C., and P. C. Wainwright. 2006. Discordance between morphological and mechanical diversity in the feeding mechanism of centrarchid fishes. Evolution 60:2575–2584.
- Cristecu, M. E., S. J. Adamowicz, J. J. Vaillant, and D. G. Haffner. 2010. Ancient lakes revisited: from the ecology to the genetics of speciation. Mol. Ecol. 19:4837–4851.
- Collar, D. C., P. C. Wainwright, and M. E. Alfaro. 2008. Integrated diversification of locomotion and feeding in labrid fishes. Biol. Lett. 4:84–86.
- Das, I. 1996. Resource use and foraging tactics in a south Indian amphibian community. J. South Asian Nat. Hist. 2:1–30.
- Diesmos, A. C., R. M. Brown, and A. C. Alcala. 2002. A new species of narrow-mouthed frog (Amphibia; Anura; Microhylidae: genus *Kaloula*) from the mountains of southern Luzon and Polillo islands, Philippines. Copeia 2002:1037–1051.
- Drewes, R. C., and B. Roth. 1981. Snail-eating frogs from the Ethiopian Highlands—a new anuran specialization. Zool. J. Linn. Soc. 72:267– 287.
- Drummond, A. J., A. Rambaut, and M. Suchard. 2011. BEAST v1.6.2: Bayesian evolutionary analysis sampling trees. Available at http://beast.bio.ed.ac.uk. Accessed November 1, 2011.
- Duellman, W. E., and L. Trueb. 1986. Biology of amphibians. McGraw-Hill Book Co., New York, 670 pp.
- Emerson, B. C., and R. G. Gillespie. 2008. Phylogenetic analysis of community assembly and structure over space and time. Trends Ecol. Evol. 23:619–630.
- Emerson, S. B. 1971. The fossorial frog adaptive zone: a study of convergence and parallelism in the Anura. Unpublished Ph.D. thesis, University of Southern California, Los Angeles, U.S.A.
- ------. 1976. Burrowing in frogs. J. Morphol. 149:437-458.
- ———. 1985. Skull shape in frogs—correlations with diet. Herpetologica 41:177–188.
- ———. 1988. Convergence and morphological constraint in frogs: variation in postcranial morphology. Fieldiana (Zool.) 43:1–19.
- ———. 1991. The ecomorphology of Bornean tree frogs (family Rhacophoridae). Zool. J. Linn. Soc. 101:337–357.
- Emerson, S. B., and D. Diehl. 1980. Toe pad morphology and mechanisms of sticking in frogs. Biol. J. Linn. Soc. 13:199–216.

- Evans, M. E. K., S. A. Smith, R. S. Flynn, and M. J. Donoghue. 2009. Climate, niche evolution, and diversification of the "Bird-Cage" Evening Primroses (*Oenothera*, sections *Anogra* and *Kleinia*). Am. Nat. 173:225– 240.
- Felsenstein, J. 1985. Phylogenies and the comparative method. Am. Nat. 125:1–15.
- ———. 1988. Phylogenies and quantitative characters. Ann. Rev. Ecol. Syst. 19:445–471.
- Foote, M. 1993. Contributions of individual taxa to overall morphological disparity. Paleobiology 19:403–419.
- ———. 1997. The evolution of morphological diversity. Ann. Rev. Ecol. Syst. 28:129–152.
- Garland, T., Jr., P. H. Harvey, and A. R. Ives. 1992. Procedures for the analysis of comparative data using phylogenetically independent contrasts. Syst. Biol. 41:18–32.
- Garland, T., Jr., A. W. Dickerman, C. M. Janis, and J. A. Jones. 1993. Phylogenetic analysis of covariance by computer simulation. Syst. Biol. 42:265–292.
- Gavrilets, S., and J. B. Losos. 2009. Adaptive radiation: contrasting theory with data. Science 323:732–737.
- Gillespie, R. 2004. Community assembly through adaptive radiation in Hawaiian spiders. Science 303:356–359.
- Givnish, T. J. 1997. Adaptive radiations and molecular systematics: issues and approaches. Pp. 1–54 in T. J. Givnish and K. J. Sytsma, ed. Molecular evolution and adaptive radiation. Cambridge Univ. Press, Cambridge, U.K.
- Glor, R. E. 2010. Phylogenetic insights on adaptive radiation. Ann. Rev. Ecol. Evol. Syst. 41:251–270.
- Gomez-Mestre, I., R. A. Pyron, and J. J. Wiens. 2012. Phylogenetic analyses reveal unexpected patterns in the evolution of reproductive modes in frogs. Evolution 66:3687–3700.
- Grant, P. E., and B. R. Grant. 1994. Phenotypic and genetic effects of hybridization in Darwin's finches. Evolution 48:297–316.
- ———. 2006. Evolution of character displacement in Darwin's fiches. Science 313:224–226.
- Grant, T., E. C. Humphrey, and C. W. Myers. 1997. The median lingual process of frogs: a bizarre character of Old World ranoids discovered in South American dendrobatids. Amer. Mus. Novitates 3212:1–40.
- Hansen, T. F. 1997. Stabilizing selection and the comparative analysis of adaptation. Evolution 51:1341–1351.
- Hansen, T. F., and K. Bartoszek. 2012. Interpreting the evolutionary regression: the interplay between observational and biological errors in phylogenetic comparative studies. Syst. Biol. 61:413–425.
- Harmon, L. J., and J. B. Losos. 2005. The effect of intraspecific sample size on type I and type II error rates in comparative studies. Evolution 59:2705–2710.
- Harmon, L. J., J. A. Schulte II, A. Larson, and J. B. Losos. 2003. Tempo and mode of evolutionary radiation in iguanian lizards. Science 301:961– 964.
- Harmon, L. J., J. T. Weir, C. D. Brock, R. E. Glor, and W. Challenger. 2008. GEIGER: investigating evolutionary radiations. Bioinformatics 24:129– 131.
- Harmon, L. J., J. B. Losos, T. J. Davies, R. G. Gillespie, J. L. Gittleman, W. B. Jennings, K. H. Kozak, M. A. McPeek, F. Moreno-Roark, T. J. Near, et al. 2010. Early bursts of body size and shape evolution are rare in comparative data. Evolution 64:2385–2396.
- Hayek, L.-A. C., W. R. Heyer, and C. Gascon. 2001. Frog morphometrics: a cautionary tale. Alytes 18:153–177.
- Hedges, S. B. 1989a. Evolution and biogeography of West Indian frogs of the genus *Eleutherodactylus*: slow-evolving loci and the major groups.

Pp. 305–370 *in* C. Woods, ed. Biogeography of the West Indies: Past, present, and future. Sandhill Crane Press, Gainesville, FL.

- —. 1989b. An island radiation: allozyme evolution in Jamaican frogs of the genus *Eleutherodactylus* (Leptodactylidae). Caribbean J. Sci. 25:123–147.
- Heyer, W. R. 1973. Ecological interactions of frog larvae at a seasonal tropical location in Thailand. J. Herpetol. 7:337–361.
 - ——. 1974. Niche measurements of frog larvae from a seasonal tropical location in Thailand. Ecology 55:651–656.
- Hillman, S. S., P. C. Withers, R. C. Drewes, and S. D. Hillyard. 2009. Ecological and Environmental Physiology of Amphibians. Oxford Univ. Press, Oxford, U.K., 464 pp.
- Hormiga, G., M. Arnedo, and R. G. Gillespie. 2003. Speciation on a conveyor belt: sequential colonization of the Hawaiian Islands by *Orsonwelles* spiders (Araneae, Linyphiidae). Syst. Biol. 52:70–88.
- Inger, R. F. 1954. Systematics and zoogeography of Philippine Amphibia. Fieldiana 33:181–531.
- Köhler, J., D. R. Vieites, R. M. Bonett, F. H. García, F. Glaw, D. Steinke, and M. Vences. 2005. New amphibians and global conservation: a boost in species discoveries in a highly endangered vertebrate group. BioScience 55:693–696.
- Kolar, C. S., and D. M. Lodge. 2001. Progress in invasion biology: predicting invaders. Trends Ecol. Evol. 16:199–204.
- Kozak, K. H., A. Larson, R. M. Bonett, and L. J. Harmon. 2005. Phylogenetic analysis of ecomorphological divergence, community structure, and diversification rates in Dusky Salamanders (Plethodontidae: *Desmognathus*). Evolution 59:2000–2016.
- Kozak, K. H., D. W. Weisrock, and A. Larson. 2006. Rapid lineage accumulation in a non-adaptive radiation: phylogenetic analysis of diversification rates in eastern North American woodland salamanders (Plethodontidae: *Plethodon*). Proc. R. Soc. B 273:539–546.
- Lack, D. 1940. Evolution of the Galapagos finches. Nature 146:324-327.
- Lande, R. 1976. Natural selection and random genetic drift in phenotypic evolution. Evolution 30:314–334.
- Lee, J. C. 1990. Sources of extraneous variation in the study of meristic characters: the effect of size and inter-observer variability. Syst. Zool. 39:31–39.
- Liem, K. F. 1973. Evolutionary strategies and morphological innovations: cichlid pharyngeal jaws. Syst. Zool. 22:425–441.
- Lombard, R. E., and D. B. Wake. 1986. Tongue evolution in the lungless salamanders, family Plethodontidae. IV. Phylogeny of plethodontid salamanders and the evolution of feeding dynamics. Syst. Zool. 35:532–551.
- Losos, J. B. 1992. The evolution of convergent structure in Caribbean *Anolis* communities. Syst. Biol. 41:403–420.
- 2009. Lizards in an evolutionary tree: ecology and adaptive radiation of anoles. Univ. California Press, Berkeley, CA, 507 pp.
- Losos, J. B., and D. L. Mahler. 2010. Adaptive radiation: the interaction of ecological opportunity, adaptation, and speciation. Pp. 381–420 in M. A. Bell, D. J. Futuyma, W. F. Eanes, and J. S. Levinton, eds. Evolution since Darwin: the first 150 years. Sinauer Associates, Sunderland, U.K.
- Losos, J. B., and D. B. Miles. 2002. Testing the hypothesis that a clade has adaptively radiated: iguanid lizard clades as a case study. Am. Nat. 160:147–157.
- Lovette, I. J., E. Bermingham, and R. E. Ricklefs. 2002. Clade specific morphological diversification and adaptive radiation in Hawaiian songbirds. Proc. R. Soc. Lond. B 269:37–42.
- Madsen, O., M. Scally, C. J. Douady, D. J. Kao, R. W. DeBry, R. Adkins, H. M. Amrine, M. J. Stanhope, W. W. de Jong, and M. S. Springer. 2001. Parallel adaptive radiations in two major clades of placental mammals. Nature 409:610–614.

- Mahler, D. L., L. J. Revell, R. E. Glor, and J. B. Losos. 2010. Ecological opportunity and the rate of morphological diversification of Greater Antillean anoles. Evolution 64:2731–2745.
- Martins, E. P. 1994. Estimating the rate of phenotypic evolution from comparative data. Am. Nat. 144:193–209.
- Mayr, E. 1963. Animal species and evolution. Harvard Univ. Press, Cambridge, 797 pp.
- McDiarmid, R. W., and R. Altig, eds. 1999. Tadpoles: the biology of anuran larvae. Univ. Chicago Press, Chicago, 444 pp.
- Meyers, J. J., J. C. O'Reilly, J. A. Monroy, and K. C. Nishikawa. 2004. Mechanism of tongue protraction in microhylid frogs. J. Exp. Biol. 207:21–31.
- Morlon, H., M. D. Potts, and J. B. Plotkin. 2010. Inferring the dynamics of diversification: a coalescent approach. PLoS Biol. 8:e1000493.
- Nishikawa, K. C. 2000. Feeding in frogs. Pp. 117–147 in K. Schwenk, ed. Feeding. Academic Press, San Diego, CA.
- Nishikawa, K. C., W. M. Kier, and K. K. Smith. 1999. Morphology and mechanics of tongue movement in the African pig-nosed frog *Hemisus marmoratum*: a muscular hydrostatic model. J. Exp. Biol. 202: 771–780.
- Orton, G. L. 1953. The systematics of vertebrate larvae. Syst. Zool. 2:63-75.
- 1957. The bearing of larval evolution on some problems in frog classification. Syst. Zool. 6:79–86.
- Paradis, E., J. Claude, and K. Strimmer. 2004. APE: analyses of phylogenetics and evolution in R language. Bioinformatics 20:289–290.
- Parmalee, J. R. 1999. Trophic ecology of a tropical anuran assemblage. Sci. Pap. Nat. Hist. Mus. Univ. Kansas 11:1–59.
- Poe, S., J. T. Giermakowski, I. Latella, E. W. Schaad, E. P. Hulebak, and M. J. Ryan. 2011. Ancient colonization predicts recent naturalization in *Anolis* lizards. Evolution 65:1195–1202.
- Pybus, O. G., and P. H. Harvey. 2000. Testing macro-evolutionary models using incomplete molecular phylogenies. Proc. R. Soc. Lond. B 267:2267– 2272.
- R Development Core Team. 2008. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0.
- Rabosky, D. L., and D. C. Adams. 2012. Rates of morphological evolution are correlated with species richness in salamanders. Evolution 66:1807– 1818.
- Rabosky, D. L., and I. J. Lovette. 2008. Density-dependent diversification in North American wood warblers. Proc. R. Soc. Lond. B 275:2363–2371.
- Reeder, D. M., K. M. Helgen, and D. E. Wilson. 2007. Global trends and biases in new mammal species discoveries. Occ. Papers Mus. Texas Tech Univ. 269:1–35.
- Rensch, B. 1959. Evolution above the species level. John Wiley & Sons, Inc., New York.
- Revell, L. J. 2009. Size-correction and principal components for interspecific comparative studies. Evolution 63:3258–3268.
- Roderick, G. K., and R. G. Gillespie. 1998. Speciation and phylogeography of Hawaiian terrestrial arthropods. Mol. Ecol. 7:519–531.
- Roelants, K., D. J. Gower, M. Wilkinson, S. P. Loader, S. D. Biju, K. Guillaume, L. Moriau, and F. Bossuyt. 2007. Global patterns of diversification in the history of modern amphibians. Proc. Natl. Acad. Sci. USA 104:887–892.
- Rohlf, F. J. 2006. A comment on phylogenetic correction. Evolution 60:1509– 1515.
- Salthe, S. N., and W. E. Duellman. 1973. Quantitative constraints associated with reproductive mode in anurans. Pp. 229–249 *in* J. L. Vial, ed. The evolutionary biology of the anurans. Univ. Missouri Press, Columbia, MI.

- Savage, J. M. 1973. The geographic distribution of frogs: patterns and predictions. Pp. 351–445 in J. L. Vial, ed. The evolutionary biology of the anurans. Univ. Missouri Press, Columbia, MI.
- Schluter, D. 2000. The ecology of adaptive radiation. Oxford Univ. Press, Oxford, U.K., 288 pp.
- Schluter, D., and J. D. McPhail. 1992. Ecological character displacement and speciation in sticklebacks. Am. Nat. 140:85–108.
- Schluter, D., T. Price, A. Ø. Mooers, and D. Ludwig. 1997. Likelihood of ancestor states in adaptive radiation. Evolution 51:1699–1711.
- Sepkoski, J. J., Jr. 1984. A kinetic model of Phanerozoic taxonomic diversity. III. Post-Paleozoic families and mass extinctions. Paleobiology 10:246– 267.
- Setiadi, M. I., J. A. McGuire, R. M. Brown, M. Zubairi, D. T. Iskandar, N. Andayani, J. Supriatna, and B. J. Evans. 2011. Adaptive radiation and ecological opportunity in Sulawesi and Philippine fanged frog (*Lim-nonectes*) communities. Am. Nat. 178:221–240.
- Simpson, G. G. 1953. The major features of evolution. Columbia Univ. Press, New York, 434 pp.
- Slater, G. J., S. A. Price, F. Santini, and M. E. Alfaro. 2010. Diversity versus disparity and the radiation of modern cetacaeans. Proc. R. Soc. Lond. B 277:3097–3104.
- Springer, M. S., J. A. W. Kirsch, and J. A. Chase. 1997. The chronicle of marsupial evolution. Pp 129–161 in T. J. Givnish and K. J. Sytsma, eds. Molecular evolution and adaptive radiation. Cambridge Univ. Press, Cambridge.
- Stamhuis, E. J., and S. Nauwelaerts. 2005. Propulsive force calculations in swimming frogs. II. Application of a vortex ring model to DPIV data. J. Exp. Biol. 208:1445–1451.
- Stebbins, R. C., and N. W. Cohen. 1995. A natural history of amphibians. Princeton Univ. Press, New Jersey, 316 pp.
- Streelman, J. T., and P. D. Danley. 2003. The stages of vertebrate evolutionary radiation. Trends Ecol. Evol. 18:126–131.
- Toft, C. A. 1980. Feeding ecology of thirteen syntopic species of anurans in a seasonal tropical environment. Oecologia 45:131–141.
- ——. A. 1981. Feeding ecology of Panamanian litter anurans: patterns in diet and foraging mode. J. Herpetol. 15:139–144.
- ——. A. 1985. Resource partitioning amphibians and reptiles. Copeia 1985:1–21.
- Trueb, L., and C. Gans. 1983. Feeding specializations of the Mexican burrowing toad, *Rhinophrynus dorsalis* (Anura: Rhinophrynidae). J. Zool. 199:189–208.
- Trueb, L., R. Diaz, and D. C. Blackburn. 2011. Osteology and chondrocranial morphology of *Gastrophryne carolinensis* (Anura: Microhylidae), with a review of the osteological diversity of New World microhylids. Phyllomedusa 10:99–135.

- van Bocxlaer, I., S. D. Biju, S. P. Loader, and F. Bossuyt. 2009. Toad radiation reveals into-India dispersal as a source of endemism in the Western Ghats-Sri Lanka biodiversity hotspot. BMC Evol. Biol. 9:131.
- van der Meijden, A., M. Vences, S. Hoegg, and A. Meyer. 2005. A previously unrecognized radiation of ranid frogs in Southern Africa revealed by nuclear and mitochondrial DNA sequences. Mol. Phylogenet. Evol. 37:674–685.
- van Valkenburgh, B. 1999. Major patterns in the history of carnivorous mammals. Ann. Rev. Earth Planet. Sci. 27:463–493.
- van Valkenburgh, B., X. Wang, and J. Damuth. 2004. Cope's rule, hypercarnivory, and extinction in North American canids. Science 306:101–104.
- Vences, M., and D. B. Wake. 2007. Speciation, species boundaries and phylogeography of amphibians. Pp. 2614–2671 in H. Heatwole and M. J. Tyler, eds. Amphibian biology Vol. 7, systematics. Surrey Beatty & Sons, Chipping Norton, Australia.
- Vieites, D. R., S. N. Román, M. H. Wake, and D. B. Wake. 2011. A multigenic perspective on phylogenetic relationships in the largest family of salamanders, the Plethodontidae. Mol. Phylogenet. Evol. 59:623–635.
- Wagner, P. 2010. Paleontological perspectives on morphological evolution. Pp. 451–478 in M. A. Bell, D. J. Futuyma, W. F. Eanes, and J. S. Levinton, eds. Evolution since Darwin: the first 150 years. Sinauer Associates, Sunderland, U.K.
- Wake, D. B. 1966. Comparative osteology and evolution of the lungless salamanders, family Plethodontidae. Mem. South. Calif. Acad. Sci. 4:1–111.
- ———. 1987. Adaptive radiation of salamanders in Middle American cloud forests. Ann. Mo. Bot. Gard. 74:242–264.
- 2009. What salamanders have taught us about evolution. Ann. Rev. Ecol. Evol. Syst. 40:333–352.
- Wake, D. B., and J. F. Lynch. 1976. The distribution, ecology, and evolutionary history of plethodontid salamanders in tropical America. Sci. Bull. Nat. Hist. Mus. Los Angeles Co. 25:1–65.
- Warheit, K. I., J. D. Forman, J. B. Losos, and D. B. Miles. 1999. Morphological diversification and adaptive radiation: a comparison of two diverse lizard clades. Evolution 53:1226–1234.
- Wells, K. D. 2007. The ecology and behavior of amphibians. Univ. Chicago Press, Chicago, 1148 pp.
- Westneat, M. W., M. E. Alfaro, P. C. Wainwright, D. R. Bellwood, J. R. Grubich, J. L. Fessler, K. D. Clements, and L. L. Smith. 2005. Local phylogenetic divergence and global evolutionary convergence of skull function in reef fishes of the family Labridae. Proc. R. Soc. Lond. B 272:993–1000.
- Worthington, E. B. 1937. On the evolution of the fish in the Great Lakes of Africa. Int. Rev. Hydrobiol. Hydrogr. 35:304–317.

Associate Editor: P. Lindenfors

Supporting Information

Additional Supporting Information may be found in the online version of this article at the publisher's website:

Table S1. Summary of specimens corresponding to genetic samples included in the study.

Figure S1. Maximum-likelihood phylogram estimated from mitochondrial DNA sequences (12S and 16S ribosomal RNA genes) depicting the phylogenetic relationships of other microhylid taxa in relation to *Kaloula*.

Figure S2. Maximum-likelihood phylogram estimated from mitochondrial DNA sequences (12S and 16S ribosomal RNA genes) depicting the phylogenetic relationships of *Kaloula* (Anura: Microhylidae; figure complements Fig. S1).

Figure S3. Uncertainty in disparification through time in *Kaloula*.