# Nematode Sodium Calcium Exchangers: A Surprising Lack of Transport



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**ABSTRACT:**  $Na^+/Ca^{2+}$  exchangers are low-affinity, high-capacity transporters that rapidly transport calcium against a gradient of  $Na^+$  ions.  $Na^+/Ca^{2+}$  exchangers are divided into three groups based upon substrate specificity:  $Na^+/Ca^{2+}$  exchangers (NCX),  $Na^+/Ca^{2+}/K^+$  exchangers (NCKX), and  $Ca^{2+}/cation$  exchangers (NCLX). In mammals, there are three NCX genes, five NCKX genes, and a single NCLX gene. The genome of the nematode *Caenorhabditis elegans* contains 10 Na<sup>+</sup>/Ca<sup>2+</sup> exchanger genes: three NCX, five NCLX, and two NCKX genes. In a previous study, we characterized the structural and taxonomic specializations within the family of  $Na^+/Ca^{2+}$  exchangers across the phylum Nematoda and observed a complex picture of  $Na^+/Ca^{2+}$  exchanger evolution across diverse nematode species. We noted multiple cases of putative gene gain and loss and, most surprisingly, did not detect members of the NCLX type of exchangers within subsets of nematode species. In this commentary, we discuss these findings and speculate on the functional outcomes and physiology of these observations. Our data highlight the importance of studying diverse systems in order to get a deeper understanding of the evolution and regulation of  $Ca^{2+}$  signaling critical for animal function.

KEYWORDS: Nematode, NCX, NCLX, sodium calcium exchanger, evolution

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## **Short Commentary**

Maintaining calcium homeostasis is necessary for the development and survival of all animals. In multicellular organisms, calcium ions can enter the cytosol via the voltage-gated calcium channel at a rate of one million ions per second,<sup>1</sup> and from there, they can target hundreds of effector proteins that engage pathways involved in muscular contraction, neurotransmission, cell differentiation, and energy metabolism to name but a few (see Ref. 2 for an excellent discussion of calcium signaling). Many of the target proteins for calcium ions in the cytosol function in a buffering capacity to chelate free calcium in order to prevent activation of pathways including that of apoptotic pathways linked to excessive free calcium. In order to set appropriate calcium levels inside cells, a superfamily of antiporters, called the sodium calcium exchangers, utilize the sodium gradient to transport calcium across various membranes. This superfamily is composed of the Na<sup>+</sup>/Ca<sup>2+</sup> exchangers (NCX), which exchange sodium for calcium, Na<sup>+</sup>/Ca<sup>2+</sup>/K<sup>+</sup> exchangers (NCKX), which exchange sodium for potassium and calcium, and Ca<sup>2+</sup>/cation exchangers (also called NCLX), which exchange sodium or lithium for calcium.<sup>3-5</sup> In mammals, there are three genes encoding various isoforms of NCX transporters, five NCKX genes, and a single NCLX gene. The mammalian NCX gene products (NCX1-3)

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are highly expressed in cardiac muscle, skeletal muscle, and the central nervous system<sup>6-8</sup>; the mammalian NCKX genes (NCKX1-5) are also widely expressed in various cells including photoreceptor cells, retinal ganglion cells, platelets, vascular smooth muscles, uterus, brain tissue, intestine, lungs, thymus, and epidermal cells.<sup>9-12</sup> The mammalian NCLX is expressed in all tissues examined including the brain, thymus, heart, skeletal muscles, lungs, kidneys, intestines, and testes and has been shown to localize to the inner membrane of mitochondria.<sup>13,14</sup> Disruption of Na<sup>+</sup>/Ca<sup>2+</sup> exchanger activity has been linked to apoptosis due to excessive accumulation of cytosolic calcium: in cardiomyocytes, overstimulation of  $G\alpha q$  protein downregulates NCX causing apoptosis,15 and in oligodendrocytes, glutamatemediated excitotoxicity can reverse NCX activity, which causes mitochondrial dysfunction and apoptosis.<sup>16</sup> Considering the central role of Na<sup>+</sup>/Ca<sup>2+</sup> exchangers and also the significant conservation between these exchangers in Caenorhabditis elegans and other animals,<sup>17</sup> it was most unusual that in a recent genome-wide study,<sup>18</sup> we failed to identify representatives of the NCLX in several species of nematodes.

The nematode *C. elegans* contains 10 Na<sup>+</sup>/Ca<sup>2+</sup> exchanger genes: three NCX (*ncx-1*, *ncx-2*, *ncx-3*), two NCKX (*ncx-4*, *ncx-5*), and five NCLX (*ncx-6*, *ncx-7*, *ncx-8*, *ncx-9*, *ncx-10*).

These exchangers are highly conserved with their counterparts in mammals and flies, <sup>3,17,19</sup> and these exchangers are widely expressed in numerous types of excitable cells and tissues in C. elegans.<sup>17</sup> In an attempt to understand taxonomic specializations within this superfamily of transporters, we studied this family in a diverse group of nematode species that encompassed Clade III, Clade IV, and Clade V representatives.<sup>18</sup> Clade designations are as described by Blaxter et al.<sup>20</sup> Clade III is thought to contain the most ancient species of the three clades we examined.<sup>21</sup> After reconstructing the phylogenetic and evolutionary relationships between these exchangers across each nematode species, we observed several putative examples of gene gain and loss, but most surprising was the apparent absence of NCLX from the Clade III nematode taxa that we sampled (Fig. 1). We detected between one and five NCLX within Clade V and also Clade IV nematode taxa that we examined but did not find evidence of NCLX from any of the three Clade III species these were Brugia malayi, Loa loa, and Ascaris suum. Furthermore, based upon a tip from one of our anonymous reviewers during the review process of our original paper,<sup>18</sup> we have since observed that members of the Clade I group also appear to lack NCLX: these include Trichinella spiralis, Trichuris muris, Trichuris suis, and Trichuris trichiura. This is



most unusual considering that Clade I taxa contain some of the most basal nematode species.<sup>21</sup> Together, these findings reveal a dynamic evolutionary history of Na<sup>+</sup>/Ca<sup>2+</sup> transporters in nematodes that have been shaped by a series of putative gene loss and gain events. This is most surprising considering the central role mediated by Na<sup>+</sup>/Ca<sup>2+</sup> transporters in controlling calcium dynamics.<sup>3,22</sup>

The NCLX is considered to be the primary regulator of mitochondrial Ca<sup>2+</sup> efflux, while mitochondrial Ca<sup>2+</sup> influx is regulated predominantly by the mitochondrial calcium uniporter (MCU).14,23 MCU is a highly selective ion channel that facilitates the uptake of cytoplasmic Ca<sup>2+</sup> ions into mitochondria and does not require adenosine triphosphate (ATP) hydrolysis or the exchange of other ions for conducting Ca<sup>2+</sup> transport. Together, calcium handling via NCLX and MCU is critical for shaping intracellular calcium signaling dynamics, mitochondrial ATP production, and regulation of cell death pathways.<sup>24-27</sup> We investigated if Clade III nematode taxa contain sequences for the MCU channel. While the Clade III taxa lack NCLX, their genomes contain a putative sequence for the MCU channel (eg, BMA-MICU-1). The presence of MCU but the absence of NCLX in this clade poses a unique challenge to the understanding of how mitochondrial calcium dynamics are regulated



**Figure 1.** Representative tree of the phylogenetic relationships between nematode species belonging to Clades I, III, IV, and V. **Note:** The types of sodium calcium exchangers are color coded red (NCKX), blue (NCX), and green (NCLX), and the number of colored lines represents the numbers of each exchanger detected from the genome of each nematode species in our analysis.<sup>18</sup>



in some organisms. It is estimated that steady-state calcium transients are reduced by as much as 36% due to mitochondrial Ca<sup>2+</sup> uptake.<sup>28</sup> However, prolonged buildup of Ca<sup>2+</sup> in the mitochondria is a trigger for cellular apoptotic pathways, and Ca<sup>2+</sup> efflux is consequentially of utmost importance for maintaining cell and organismal health.<sup>29</sup> It is unworkable that mitochondrial Ca2+-clearing mechanisms do not exist in these nematode species lacking the NCLX, but rather it is possible that mitochondrial Ca2+ efflux is regulated entirely by the NCX variants found within these organisms. Members of the NCX subfamily have been recently implicated in mitochondrial Na<sup>+</sup>/Ca<sup>2+</sup> exchange. Human NCX1 is functional in the mitochondria of neuronal and glial cells and colocalizes with the excitatory amino acid carrier 1 to promote glutamate uptake in mitochondria.<sup>30</sup> The human NCX2 and NCX3 localize and function at the mitochondria of dopaminergic neurons.<sup>31</sup> Furthermore, NCX3 activity in the outer mitochondrial membrane is essential for promoting cell survivability in neurons during hypoxic conditions.<sup>32</sup> NCLX exhibits several functional differences to that of NCX, in particular, NCLX exhibits specific sensitivity to the inhibitor CGP 3715733 and also exhibits substrate specificity to Li<sup>+</sup>, which is not shared by NCX. However, both NCX and NCLX mediate Na<sup>+</sup>/Ca<sup>2+</sup> exchange, and the Li<sup>+</sup> specificity of NCLX is not considered physiological. Therefore, we speculate that in Clade III taxa,

a lack of NCLX may be overcome by mitochondrial localization and functioning of the NCX, which may function to handle plasma membrane exchange and also exchange at the mitochondrion.

It is quite striking that within the phylum Nematoda, there exists such remarkable diversity within the number of Na<sup>+</sup>/Ca<sup>2+</sup> exchangers found within each species. This apparent diversity suggests that mechanisms regulating calcium homeostasis are precisely tailored to meet the varying physiological demands of individual species over their lifetime. Yet, the important regulatory domains of these exchangers appear to be highly conserved across nematodes, flies, and humans<sup>17,18,34</sup> and even fungi in the case of NCLX.<sup>22</sup> Our observations on the absence of NCLX in Clade III nematodes highlight the importance of studying diverse systems in order to get a deeper understanding of the evolution and regulation of Ca<sup>2+</sup> signaling critical for animal biology.

### **Author Contributions**

Conceived the concepts: VS, DMO. Analyzed the data: VS, DMO. Wrote the first draft of the manuscript: VS, DMO. Contributed to the writing of the manuscript: VS, DMO. Agree with manuscript results and conclusions: VS, DMO. Jointly developed the structure and arguments for the paper: VS, DMO. Made critical revisions and approved final version: VS, DMO. Both authors reviewed and approved of the final manuscript.

#### REFERENCES

- 1. Clapham DE. Calcium signaling. Cell. 2007;131(6):1047-58.
- 2. Clapham DE. Calcium signaling. Cell. 1995;80(2):259-68.
- Cai X, Lytton J. The cation/Ca(2+) exchanger superfamily: phylogenetic analysis and structural implications. *Mol Biol Evol*. 2004;21(9):1692–703.
- Baker PF, Blaustein MP, Hodgkin AL, Steinhardt RA. The influence of calcium on sodium efflux in squid axons. J Physiol. 1969;200(2):431–58.
- Blaustein MP. Barbiturates block sodium and potassium conductance increases in voltage-clamped lobster axons. J Gen Physiol. 1968;51(3):293–307.
- Minelli A, Castaldo P, Gobbi P, Salucci S, Magi S, Amoroso S. Cellular and subcellular localization of Na+-Ca2+ exchanger protein isoforms, NCX1, NCX2, and NCX3 in cerebral cortex and hippocampus of adult rat. *Cell Calcium*. 2007;41(3):221–34.
- Nicoll DA, Quednau BD, Qui Z, Xia YR, Lusis AJ, Philipson KD. Cloning of a third mammalian Na+-Ca2+ exchanger, NCX3. J Biol Chem. 1996; 271(40):24914–21.
- Li Z, Matsuoka S, Hryshko LV, et al. Cloning of the NCX2 isoform of the plasma membrane Na(+)-Ca2+ exchanger. J Biol Chem. 1994;269(26):17434–9.
- Altimimi HF, Szerencsei RT, Schnetkamp PP. Functional and structural properties of the NCKX2 Na(+)-Ca(2+)/K(+) exchanger: a comparison with the NCX1 Na(+)/Ca(2+) exchanger. Adv Exp Med Biol. 2013;961:81–94.
- Lee SH, Kim MH, Park KH, Earm YE, Ho WK. K+-dependent Na+/Ca2+ exchange is a major Ca2+ clearance mechanism in axon terminals of rat neurohypophysis. *J Neurosci.* 2002;22(16):6891–9.
- Lytton J, Li XF, Dong H, Kraev A. K+-dependent Na+/Ca2+ exchangers in the brain. Ann NY Acad Sci. 2002;976:382–93.
- Yang H, Yoo YM, Jung EM, Choi KC, Jeung EB. Uterine expression of sodium/ potassium/calcium exchanger 3 and its regulation by sex-steroid hormones during the estrous cycle of rats. *Mol Reprod Dev.* 2010;77(11):971–7.
- Palty R, Hershfinkel M, Sekler I. Molecular identity and functional properties of the mitochondrial Na+/Ca2+ exchanger. J Biol Chem. 2012;287(38): 31650-7.
- Palty R, Silverman WF, Hershfinkel M, et al. NCLX is an essential component of mitochondrial Na+/Ca2+ exchange. *Proc Natl Acad Sci U S A*. 2010;107(1): 436–41.
- Miyamoto S, Howes AL, Adams JW, Dorn GW II, Brown JH. Ca2+ dysregulation induces mitochondrial depolarization and apoptosis: role of Na+/Ca2+ exchanger and AKT. J Biol Chem. 2005;280(46):38505-12.
- Chen H, Kintner DB, Jones M, et al. AMPA-mediated excitotoxicity in oligodendrocytes: role for Na(+)-K(+)-Cl(-) co-transport and reversal of Na(+)/Ca(2+) exchanger. J Neurochem. 2007;102(6):1783–95.
- Sharma V, He C, Sacca-Schaeffer J, et al. Insight into the family of Na+/Ca2+ exchangers of *Caenorhabditis elegans*. *Genetics*. 2013;195(2):611–9.
- He C, O'Halloran DM. Analysis of the Na+/Ca2+ exchanger gene family within the phylum Nematoda. *PLoS One*. 2014;9(11):e112841.
- Szerencsei RT, Tucker JE, Cooper CB, et al. Minimal domain requirement for cation transport by the potassium-dependent Na/Ca-K exchanger. Comparison with an NCKX paralog from *Caenorbabditis elegans*. J Biol Chem. 2000;275(1):669-76.
- Blaxter ML, De Ley P, Garey JR, et al. A molecular evolutionary framework for the phylum Nematoda. *Nature*. 1998;392(6671):71–5.
- Coghlan, A. Nematode genome evolution. September 07, 2005. WormBook, ed. The C. elegans Research Community, WormBook, doi/10.1895/wormbook.1.15.1, http://www.wormbook.org.
- 22. Cai X, Clapham DE. Ancestral Ca2+ signaling machinery in early animal and fungal evolution. *Mol Biol Evol*. 2012;29(1):91–100.
- Perocchi F, Gohil VM, Girgis HS, et al. MICU1 encodes a mitochondrial EF hand protein required for Ca(2+) uptake. *Nature*. 2010;467(7313):291-6.
- Budd SL, Nicholls DG. A reevaluation of the role of mitochondria in neuronal Ca2+ homeostasis. J Neurochem. 1996;66(1):403–11.
- Scorrano L, Oakes SA, Opferman JT, et al. BAX and BAK regulation of endoplasmic reticulum Ca2+: a control point for apoptosis. *Science*. 2003; 300(5616):135–9.
- Rizzuto R, Bernardi P, Pozzan T. Mitochondria as all-round players of the calcium game. J Physiol. 2000;529(pt 1):37–47.
- Jouaville LS, Ichas F, Holmuhamedov EL, Camacho P, Lechleiter JD. Synchronization of calcium waves by mitochondrial substrates in *Xenopus laevis* oocytes. *Nature*. 1995;377(6548):438–41.
- Maack C, Cortassa S, Aon MA, Ganesan AN, Liu T, O'Rourke B. Elevated cytosolic Na+ decreases mitochondrial Ca2+ uptake during excitation-contraction coupling and impairs energetic adaptation in cardiac myocytes. *Circ Res.* 2006;99(2):172–82.
- Orrenius S, Gogvadze V, Zhivotovsky B. Calcium and mitochondria in the regulation of cell death. *Biochem Biophys Res Commun.* 2015;460(1):72–81.
- Magi S, Lariccia V, Castaldo P, et al. Physical and functional interaction of NCX1 and EAAC1 transporters leading to glutamate-enhanced ATP production in brain mitochondria. *PLoS One*. 2012;7(3):e34015.

3



- Wood-Kaczmar A, Deas E, Wood NW, Abramov AY. The role of the mitochondrial NCX in the mechanism of neurodegeneration in Parkinson's disease. *Adv Exp Med Biol.* 2013;961:241–9.
- Scorziello A, Savoia C, Sisalli MJ, et al. NCX3 regulates mitochondrial Ca(2+) handling through the AKAP121-anchored signaling complex and prevents hypoxia-induced neuronal death. *J Cell Sci.* 2013;126(pt 24):5566–77.
- Cox DA, Conforti L, Sperelakis N, Matlib MA. Selectivity of inhibition of Na(+)-Ca2+ exchange of heart mitochondria by benzothiazepine CGP-37157. *J Cardiovasc Pharmacol.* 1993;21(4):595-9.
- 34. Sharma V, O'Halloran D. Recent structural and functional insights into the family of sodium calcium exchangers. *Genesis*. 2014;52(2):93.