#### REVIEW



# The therapeutic potential of bone marrow-derived macrophages in neurological diseases

Kai Zhou<sup>1</sup> | Jinming Han<sup>2</sup> | Yafeng Wang<sup>1,3</sup> | Yiran Xu<sup>4</sup> | Yaodong Zhang<sup>1</sup> | Changlian Zhu<sup>4,5</sup> |

#### Correspondence

Kai Zhou, Henan Neurodevelopment Engineering Research Center for Children, Children's Hospital Affiliated to Zhengzhou University, Zhengzhou, China. Email: kaizhoubusi@gmail.com

Changlian Zhu, Henan Key Laboratory of Child Brain Injury and Henan Pediatric Clinical Research Center, The Third Affiliated Hospital and Institute of Neuroscience, Zhengzhou University, Zhengzhou 450052, China (or) Center for Brain Repair and Rehabilitation, Institute of Neuroscience and Physiology, University of Gothenburg, Gothenburg 40530. Sweden.

Email: changlian.zhu@neuro.gu.se

#### **Funding information**

Swedish Governmental grants to scientists working in health care, Grant/Award Number: ALFGBG-965197; the National Nature Science Foundation of China, Grant/Award Number: U21A20347; the Swedish Cancer Foundation, Grant/Award Number: 20-1121-PjF; Swedish Childhood Cancer Foundation, Grant/Award Number: PR2018-0082 and PR2021-0020

#### **Abstract**

Circulating monocytes are precursors of both tissue macrophages and dendritic cells, and they can infiltrate the central nervous system (CNS) where they transform into bone marrow-derived macrophages (BMDMs). BMDMs play essential roles in various CNS diseases, thus modulating BMDMs might be a way to treat these disorders because there are currently no efficient therapeutic methods available for most of these neurological diseases. Moreover, BMDMs can serve as promising gene delivery vehicles following bone marrow transplantation for otherwise incurable genetic CNS diseases. Understanding the distinct roles that BMDMs play in CNS diseases and their potential as gene delivery vehicles may provide new insights and opportunities for using BMDMs as therapeutic targets or delivery vehicles. This review attempts to comprehensively summarize the neurological diseases that might be treated by modulating BMDMs or by delivering gene therapies via BMDMs after bone marrow transplantation.

#### KEYWORDS

bone marrow transplantation, dysfunctional microglia, gene delivery, gene therapy, macrophage, monocyte

# 1 | INTRODUCTION

Monocytes differentiate from monoblasts in the bone marrow before moving into the bloodstream, and they account for 2%-8% of

the total leukocytes in the blood.<sup>1</sup> Over 50% of monocytes are reserved in the spleen and accumulate rapidly after injury.<sup>2</sup> Three types of circulating monocytes in humans have been identified based on the expression of CD14 and CD16 and are classified as CD14<sup>high</sup>/CD16<sup>-</sup>.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2022 The Authors. CNS Neuroscience & Therapeutics published by John Wiley & Sons Ltd.

<sup>&</sup>lt;sup>1</sup>Henan Neurodevelopment Engineering Research Center for Children, Children's Hospital Affiliated to Zhengzhou University, Zhengzhou, China

<sup>&</sup>lt;sup>2</sup>Department of Neurology, Xuanwu Hospital, Capital Medical University, Beijing, China

<sup>&</sup>lt;sup>3</sup>Department of Hematology and Oncology, Children's Hospital Affiliated to Zhengzhou University, Henan, Children's Hospital, Zhengzhou Children's Hospital, Zhengzhou, China

<sup>&</sup>lt;sup>4</sup>Henan Key Laboratory of Child Brain Injury and Henan Pediatric Clinical Research Center, The Third Affiliated Hospital and Institute of Neuroscience, Zhengzhou University, Zhengzhou, China

<sup>&</sup>lt;sup>5</sup>Centre for Brain Repair and Rehabilitation, Institute of Neuroscience and Physiology, Sahlgrenska Academy, University of Gothenburg, Gothenburg, Sweden

CD14<sup>low</sup>/CD16<sup>high</sup>, and CD14<sup>high</sup>/CD16<sup>low</sup> monocytes.<sup>3</sup> In addition, two subsets of circulating monocytes have been identified in mice, namely CX3CR1<sup>high</sup>/CCR2<sup>-</sup>/Ly6C<sup>low</sup>/PD-L1<sup>+</sup> and CX3CR1<sup>low</sup>/CCR2<sup>+</sup>/Ly6C<sup>high</sup>/PD-L1<sup>-</sup> monocytes, and they can be recruited to both inflamed and noninflamed sites or can patrol along the blood vessels, respectively.<sup>1,4,5</sup> Moreover, monocytes can differentiate into tissue macrophages or dendritic cells after infiltrating into the tissue.<sup>5,6</sup>

Circulating monocytes cannot infiltrate into the brain under physiological conditions due to the blood-brain barrier (BBB). However, CX3CR1 low/CCR2+/Lv6C high/PD-L1 monocytes can penetrate the brain and become bone marrow-derived macrophages (BMDMs) under some disease conditions in which the BBB is compromised. 7-10 However, the cellular and molecular mechanisms regulating monocyte infiltration are largely unknown. Cell adhesion molecules such as intercellular adhesion molecule-1 and vascular cell adhesion molecule-1 expressed in endothelial cells have been shown to play critical roles in monocyte infiltration, and these molecules have been used to regulate monocyte infiltration and neuroinflammation. 11-13 Interestingly, previous studies have suggested that the process of monocyte infiltration does not require BBB damage. Instead, C-C motif chemokine receptor 2 (CCR2) was noted to be the entry ticket for monocyte infiltration into the CNS. Thus, CCR2 is widely used as a therapeutic target for inhibiting monocyte infiltration into the brain.<sup>7,9,14,15</sup>

Similar to the resident microglia, BMDMs express cellular markers, such as CD11b, lba-1, and CX3CR1. However, BMDMs and microglia display distinct functions in disease conditions. <sup>7,16</sup> Moreover, chimeric mouse models, bone marrow transplantation (BMT), single cell sequencing, and the discovery of new microglial-specific markers have made it possible to distinguish BMDMs and microglia. <sup>17-21</sup> BMDMs play paradoxical roles in CNS pathology and recovery, <sup>22,23</sup> and here we summarize the complex roles that BMDMs play in various CNS diseases and discuss how BMDMs might serve as a potential therapeutic target for treating such disorders.

# 2 | BMDMS PLAY COMPLEX ROLES IN VARIOUS CNS DISEASES

BMDMs can promote neuroinflammation and exacerbate neurodegeneration in various CNS diseases, but they can also compensate for dysfunctional microglia and can remove toxic substances and cellular debris, thus protecting the brain from further injury (Figure 1). These dual roles of BMDMs have been reported in various CNS disease conditions as described below.

#### 2.1 | Viral encephalitis

Acute viral infection and the following chronic inflammatory responses cause various behavioral deficits, including motor, psychiatric, and cognitive dysfunctions.<sup>24</sup> In addition, the infiltration of monocytes is a hallmark of viral infections in the CNS.<sup>25</sup>

Inhibiting monocyte infiltration using CCR2 knockout (KO) mice increased the mortality and duration of West Nile virus (WNV) encephalitis, indicating a beneficial role of BMDMs in treating this disease.<sup>26</sup> In support of this, the depletion of monocytes using clodronate-loaded liposomes increased the mortality in a low virus dose-induced WNV model, which might be due to the loss of phagocytosis of the virus particles.<sup>27</sup> In contrast, inhibiting monocyte infiltration by blocking adhesion molecule binding or by using an antibody against C-C motif chemokine ligand 2 (CCL2) revealed a pathologic role for BMDMs in a lethal WNV model, which might be due to the proinflammatory response of BMDMs.<sup>28,29</sup> Moreover, CCR2 KO decreased hippocampal neuronal death by decreasing BMDM-derived proinflammatory cytokines in a Theiler's virus model. 30 Thus, BMDMs appear to play contrasting roles in viral encephalitis, which might be due to the different viral encephalitis models that have been studied and to the different ways of inhibiting the infiltration of monocytes.

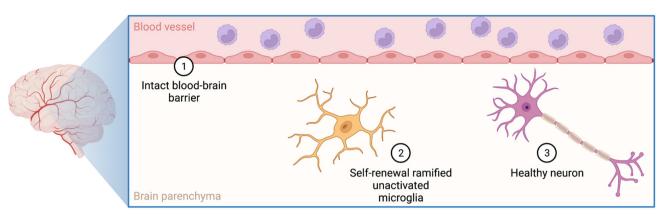
# 2.2 | Multiple sclerosis (MS)

In an experimental autoimmune encephalomyelitis (EAE) mouse model, electron microscopy showed that BMDMs were attached to nodes of Ranvier and initiated demyelination in the brain.<sup>31</sup> Moreover, BMDMs were found to secrete large amounts of proinflammatory cytokines and to participate in the inflammatory response in the EAE model.<sup>32</sup> A strong correlation between BMDMs and the paralytic stage of EAE was also noted.<sup>33</sup> Furthermore, decreasing monocyte infiltration by blocking endothelial adhesion molecules exerted antiinflammatory effects and reduced the pathological process of MS, 13,34 and CCR2 KO mice failed to develop pathological lesions in the CNS. indicating the indispensable role of BMDMs in EAE pathology. 35,36 Conversely, a clinical study suggested that foamy macrophages consisting of both microglia and BMDMs are antiinflammatory in patients with MS.<sup>37</sup> However, that study lacked evidence to show that they could accurately distinguish between microglia and BMDMs. Taken together, consistent results support the detrimental roles of BMDMs in animal models of MS. However, caution should be used when interpreting the clinical results because of the limitations of the methods for distinguishing microglia from BMDMs.

## 2.3 | Traumatic brain injury (TBI)

TBI comprises primary and secondary injury with disruption of the BBB. After the primary injury, circulating monocytes are recruited to the injury site where they might secrete proinflammatory cytokines, reactive oxygen species, and proapoptotic proteins that aggravate the neuronal damage. Moreover, CCR2 KO and CCR2 selective antagonists diminish the TBI-induced brain injury and cognitive dysfunctions. Similar results were obtained in the C-C motif chemokine ligand 2 (CCL2) KO and liposome-encapsulated clodronate-induced monocyte depletion model. Territories, infiltrating monocytes are more robust in aged TBI mice compared with young TBI mice, and CCR2

# **Healthy Brain**



# AD and other CNS disorders

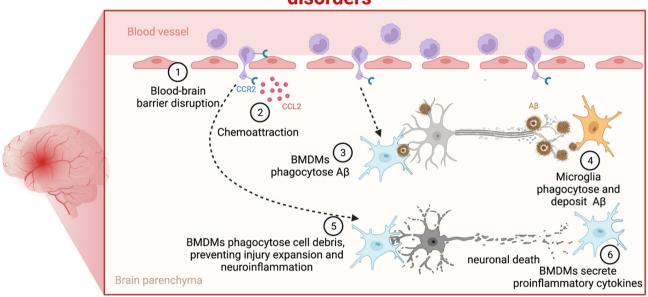


FIGURE 1 Monocytes in the healthy brain and in AD and other CNS disorders. The top panel illustrates how monocytes circulate in the blood vessels of the healthy brain, with no infiltration into the brain parenchyma. The bottom panel shows how circulating monocytes infiltrate into the brain parenchyma due to BBB disruption and chemoattraction in AD and other CNS disorders.

KO prevents chronic injury-induced cognitive decline in the aged TBI mice. 46 Together, these results indicate that BMDMs play pathological roles in TBI, and therefore, inhibiting the infiltration of monocytes might be an effective strategy to alleviate brain injury.

# 2.4 | Spinal cord injury (SCI)

SCI usually causes irreversible disability because of the permanent loss of neurons and the inability for them to regrow. Circulating monocytes infiltrate the injury site after SCI, and BMDMs may help spinal cord recovery after SCI through the expression of antiinflammatory cytokine IL-10. Moreover, eliminating BMDMs by antigenmediated depletion or by exposure to diphtheria toxin reduced the

physical recovery after SCI.<sup>47</sup> Other studies also demonstrated a protective role for BMDMs by resolving neuroinflammation and removing myeline or cellular debris in various animal models of SCI.<sup>48–51</sup> Conversely, BMDMs are necessary for forming the fibrotic scar that inhibits axon regeneration.<sup>52</sup> Moreover, monocyte depletion by liposome-encapsulated clodronate alleviates SCI-induced behavioral deficits.<sup>53–55</sup> Taken together, these findings indicate the complex roles of BMDMs in SCI.

# 2.5 | Alzheimer's disease (AD)

Amyloid- $\beta$  (A $\beta$ ) plays a central role in the pathophysiology of AD, and accumulation of A $\beta$  in the extracellular space can be neurotoxic

and further induce tau pathology, leading to neurodegeneration. Therefore, targeting A $\beta$  has been used as a therapeutic strategy for early stage AD. <sup>56</sup> Previous studies revealed that microglia are responsible for A $\beta$  deposition, <sup>57-60</sup> and a recent study demonstrated that microglia are A $\beta$  carriers and can transport A $\beta$  to healthy brain tissues. <sup>61</sup> Conversely, microglia can also degrade and remove A $\beta$  (Figure 1).

Similarly, monocytes play dual roles in the progression of AD.<sup>64</sup> However, BMDMs have been shown to play a beneficial role in early AD pathology, and one study demonstrated that BMDMs, rather than microglia, can eliminate A $\beta$  by phagocytosis<sup>65</sup> (Figure 1). Consistent with this, inhibiting the infiltration of monocytes (CCR2 KO) aggravates the progression of AD. 66,67 Moreover, transplantation of wild-type bone marrow cells expressing CCR2 can decrease Aβ accumulation and prevent cognitive decline in the CCR2 KO AD mouse model.<sup>68</sup> All of these findings indicate that BMDMs play neuroprotective roles in controlling the progression of AD, and BMDMs are thus therapeutic targets for AD.<sup>69</sup> Several approaches targeting BMDMs have shown positive therapeutic effects. For example, systematic administration of macrophage colony-stimulating factor increased the number of BMDMs in the brain, thus decreasing the AB deposition and further rescuing social deficits and cognitive decline.<sup>70</sup> Taken together, these findings suggest that promoting BMDMs can be an effective treatment for the early stages of AD.

## 2.6 | Ischemic stroke

Ischemic stroke accounts for the majority of stroke cases, and the BBB can be severely compromised after ischemic stroke. This leads to the infiltration of circulating monocytes to the injury sites, and BMDMs can remove cellular debris,<sup>71</sup> supporting angiogenesis,<sup>72</sup> and resolve ongoing neuroinflammation. 73-75 Moreover, impaired monocyte infiltration due to CCR2 KO or to treatment with CCR2 antagonists can impair angiogenesis, increase the extent of tissue damage, and exacerbate behavioral deficits.<sup>74-76</sup> Furthermore. macrophage colony-stimulating factor mobilizes circulating monocytes, increases the infiltration of BMDMs into the injury sites, and protects the brain from secondary injury.<sup>77</sup> Interestingly, both Ly6C<sup>high</sup> and Ly6C<sup>low</sup> monocytes in mice can infiltrate into the brain after ischemic stroke and play distinctly protective roles in disease recovery. 78,79 Notably, one study demonstrated that BMDMs play distinct roles in different stages of ischemic stroke. In the acute phase, BMDMs are proinflammatory and can worsen the brain injury, and CCR2-deficiency decreases acute injury. However, at later phases, BMDMs gradually switch to antiinflammatory states and are essential for brain recovery, and CCR2-deficient mice show increased mortality rates and increased delayed neurological injury. 80 Moreover, modulating BMDMs state from proinflammatory to antiinflammatory can promote brain recovery. 81,82 Because BMDMs play distinct roles in different phases of ischemic stroke, different intervention strategies may need to be applied at different phases.

# 2.7 | Retinal degeneration

Monocytes were shown to infiltrate into the retina, proliferate, and become BMDMs in an N-methyl-N-nitrosourea-induced retinal damage mouse model. BMDMs made up 15% of the total microglia 7 days after the injection of N-methyl-N-nitrosourea, and they were explicitly located in the injury sites, which suggests a role in the phagocytosis of cell debris and in resolving inflammation. In an inherited retinal degeneration mouse model, 80% of the microglia were replaced by BMDMs, and decreasing the recruitment of BMDMs into the degenerating retina by inhibiting stromal-derived factor 1 or by systemic depletion of circulating monocytes resulted in the acceleration of retinal degeneration. Moreover, systemic administrations of granulocyte colony-stimulating factor 1 and erythropoietin have been shown to slow down retinal degeneration by synergistically stimulating bone marrow stem cells and circulating monocytes. 84,85

A study demonstrated that BMDMs are neuroprotective in a mouse model of glutamate toxicity in the eye. <sup>86</sup> The authors showed that the existence of BMDMs in the retina can protect retinal ganglion cells from retinal insult, and they showed the antiinflammatory action of BMDMs by regulating the accumulation of other immune cells. Finally, the authors provided evidence that BMDMs support progenitor cell renewal after retinal injury. Moreover, BMDMs can promote the clearing o of A $\beta$  from the retina, prevent neuron loss in AD models, and encourage vascularization after hypoxic retinopathy. <sup>87</sup>

Different strategies have been investigated to facilitate BMDM infiltration into the retina. For example, insulin-like growth factor 1 can break down the blood-retina barrier and increase the infiltration of BMDMs into the retina. Reference introduced introduced by intravitreally injected bone marrow cells can also engraft into the retina and transform into microglialike cells or BMDMs. All of these findings indicate that BMDMs play protective roles against different types of retinal degeneration and that stimulating the infiltration of monocytes and bone marrow cells is a promising treatment strategy for these hard-to-treat diseases.

In summary, BMDMs plays complex roles in various CNS diseases. The controversial results in different studies may arise from different models, differences in disease severity in preclinical models, different pathological phases, and the accuracy of distinguishing BMDMs from microglia. Moreover, both subsets of monocytes can infiltrate into the brain in some disease conditions, such as after ischemic stroke, and these subsets of monocytes-derived BMDMs may have distinct effects. Thus, studying these subsets separately may provide more precise treatment strategies for these diseases. It is important to note that many of these conclusions are based on the inhibition of monocyte infiltration, and caution should be taken when interpreting related results because CCR2 KO may also inhibit the process of microglial migration.

# 3 | BMDMS AS A VEHICLE FOR CNS GENE DELIVERY

Microglia are CNS-resident immune cells and constitute approximately 10% of the total cells in the brain.<sup>89</sup> Apart from immune

surveillance, microglia play vital roles in CNS development and functions, such as synaptic pruning, neurogenesis, and the prevention of excitotoxicity. Dysfunctional microglia caused by gene mutations accelerate the progression of various CNS diseases pathologies, and they can also directly cause CNS diseases, e.g., colony-stimulating factor 1 receptor (CSF-1R)-related leukoencephalopathy. Moreover, gene mutations in the brain are not only limited to microglia, but also occur in other cell types in some diseases, such as in lysosomal storage disorders (LSDs), and the correct or missing genes delivered by BMDM provide functional proteins to both microglia and neurons. Therefore, introducing therapeutic genes delivered by BMDMs is a promising therapeutic strategy for CNS-related diseases (Figure 2).

The BBB acts as a barrier to gene therapy delivery into the CNS system, and intraparenchymal injection into specific parts of the brain is currently used in most preclinical and clinical trials. However, this is an invasive approach, and the delivery efficiency and distribution are not optimal. Moreover, multiple dosing is limited when using intraparenchymal injections. Importantly, BMDMs have been proposed as a drug delivery system for the CNS. 87.99 BMDMs can infiltrate into the brain injury sites and can widely replace resident microglia, and thus they can be used to deliver therapeutic proteins and additional nutrients to the CNS.87.100 (Figure 2).

BMDMs expressing normal genes or engineered BMDMs with therapeutic genes can be delivered by BMT. However, it is important to note that BMDMs can only graft into the brain parenchyma under special conditions following BMT. For example, whole-body irradiation can significantly suppress the immune system and ablate the bone marrow niche, thus providing enough space for the engraftment of donor cells. 9,101 However, irradiation-induced cell death and tissue damage may lead to other severe diseases. 102 Another way to support BMDM engraftment is to use chemotherapies such as busulfan and cyclophosphamide prior to BMT. 8,103

Monocytes can infiltrate specifically into the injury sites under some disease conditions and then disappear upon recovery, and thus, they do not contribute to the microglial pool. <sup>33,77,103</sup> Moreover,

the numbers of BMDMs are much lower than resident microglia or dysfunctional microglia in CNS diseases models or after BMT.<sup>104</sup> Therefore, increasing the microglial replacement rate by BMDMs is a critical step in delivering therapeutic genes.

BMDMs can only replace resident microglia when a microglial niche is deprived of microglia, and after microglial depletion the surviving microglia compete with the BMDMs for the empty microglial niche. 105 One study demonstrated that microglia can be widely replaced by BMDMs after efficient microglia depletion using a CSF-1R inhibitor. 106 Moreover, efficient microglial depletion and BMT can achieve up to 80% microglial replacement by BMDMs. 107 Furthermore, clinical trials of cancer therapy using CSF-1R inhibitors have indicated that microglia can also be depleted in humans. 108 However, large doses of these drugs are needed to achieve efficient depletion, and the safety concerns regarding extensive microglial depletion have limited their clinical use. Nonetheless, administration of a CX3CR1 inhibitor has been proposed to achieve efficient microglial replacement by BMDMs, even with only partial microglia depletion. 109 In summary, combining BMT, microglia depletion, and CX3CR1 inhibition can be a strategy for achieving widespread microglial replacement and optimal gene therapy delivery into the CNS.

# 3.1 | Amyotrophic lateral sclerosis (ALS)

The results from a chimeric mouse study showed that microglia with Cu/Zn superoxide dismutase (SOD1) mutations are neurotoxic and contribute to the ALS pathology. Moreover, wild-type BMT has protective effects in the SOD1 ALS mouse model due to the significant engraftment of BMDMs in the brain. Furthermore, as gene carriers engineered bone marrow cells successfully delivered neuroprotective glutamate transporters to pathological lesions in the SOD1 ALS mouse model and restored motor functions. Many other cell therapies using different cell sources, including umbilical cord blood cells and mesenchymal stem cells, and given through various administration routes (intracerebroventricular, intraspinal,

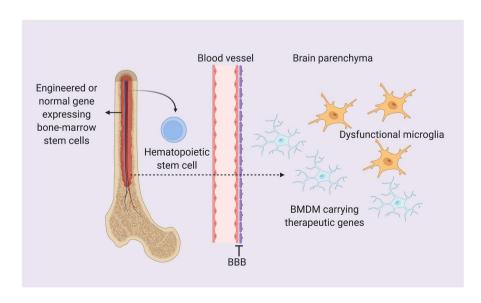


FIGURE 2 BMT delivers therapeutic genes to the brain parenchyma via BMDMs. Bone-marrow stem cells, especially hematopoietic stem cells carrying therapeutic genes, infiltrate into the brain parenchyma via the BBB and transform into BMDMs. As a result, BMDMs express the correct genes and replace the dysfunctional microglia.

and intramuscular injections) have also shown modest beneficial effects.  $^{113-116}$ 

LSDs make up a group of inherited metabolic disorders with a deficiency in lysosomal hydrolases that break down macromolecules (including proteins, lipids, carbohydrates, and nucleic acid) in the cells. <sup>117</sup> In patients with LSDs, these macromolecules can accumulate and eventually become toxic to the CNS or other organs. CNS manifestations of LSDs can be ameliorated by BMDM engraftment followed by BMT with cells expressing the deficient hydrolase. <sup>95,101</sup> Another study indicated that the failure of BMT in patients with overt neurological symptoms may be due to the rapid turnover of residential microglia and insufficient BMDM engraftment. Thus, combining microglial depletion and BMT can be a solution to increase the engraftment of BMDMs in these conditions. <sup>101</sup>

Metachromatic leukodystrophy (MLD) is an LSD resulting from deficiency in the lysosomal enzyme arylsulfatase A and causes myelin degeneration in the CNS and peripheral nervous system. Currently, there is no available treatment for this disease; however, gene therapies via intraparenchymal injections of virus vectors carrying correct genes and ex vivo-engineered cells might be viable options. However, the need for multiple doses and the invasiveness of the procedures limits the use of these methods in clinical practice. Engineered bone marrow cells can engraft into the CNS and achieve widespread distribution of BMDMs that express exogenous genes. <sup>118</sup> For example, transplantation of engineered bone marrow-expressing transduced arylsulfatase A was engrafted in the CNS, thus preventing neuropathological progression and rescuing behavioral deficiencies in a mouse model of MLD. <sup>100,119</sup>

BMT is also widely used in other LSD disorders such as globoid cell leukodystrophy.  $^{120,121}$  Glycolipid storage diseases are inherited disorders in which enzymes for the degradation of glycosphingolipids are missing, which eventually leads to CNS degeneration.  $^{122}$  Sandhoff disease is a glycolipid storage disease characterized by a lack of lysosomal  $\beta$ -hexosaminidase and the subsequent accumulation of  $\beta$ -hexosaminidase in the CNS. The introduction via BMT of BMDMs expressing the wild-type gene has a neuroprotective effect in a mouse model of Sandhoff disease.  $^{123}$  Another example of glycolipid storage disease is GM1-gangliosidosis, and BMT with engineered BMCs expressing lysosomal  $\beta$ -galactosidase can rescue the pathological features of this disease.  $^{124}$  Taken together, BMT of BMDMs with corrected gene expression might serve as a powerful tool for treating LSDs.

### 3.2 | Rett syndrome

Rett syndrome is a rare genetic and developmental neurological disorder caused by mutation in the *MECP2* gene (encoding a methyl-CpG-binding protein). Rett syndrome is frequently noted in girls and causes progressive motor neuron loss. Patients with Rett syndrome usually develop movement deficiency, loss of speech, and even breathing difficulties. One study revealed that *MECP2* gene mutations in microglia can be neurotoxic, indicating the contribution of dysfunctional microglia in the progression of Rett syndrome. Moreover, BMDMs with normal *MECP2* gene expression delivered by BMT had neuroprotective effects and halted disease progression. Furthermore, BMT without BMDMs infiltration by shielding the head during irradiation

TABLE 1 BMDMs play different roles in various central nervous system diseases

The Paragray and references in various central net rocks system also assess			
Disease model	BMDMs beneficial or detrimental	Possible mechanisms	Methods for blocking BMDMs methods
Low dose WNV model	Beneficial	Phagocytosis virus particles	CCR2 KO or monocyte depletion
Lethal WNV model	Detrimental	Proinflammatory	Blocking adhesion molecules or antibody against CCL2
Theiler's virus model	Detrimental	Secreting proinflammatory cytokines	CCR2 KO
MS	Detrimental	Initiating demyelination and secreting proinflammatory cytokines	Blocking adhesion molecules or CCR2 KO
TBI	Detrimental	Secreting proinflammatory cytokines, reactive oxygen species, and proapoptotic proteins	CCR2 KO or CCL2 KO or CCR2 selective antagonists
SCI	Beneficial	Secreting antiinflammatory cytokines and removing myeline or cellular debris	Monocyte depletion
	Detrimental	Inhibiting axon regeneration	Monocyte depletion
Early AD	Beneficial	Phagocytosing $A\beta$	CCR2 KO
Acute ischemic stroke	Detrimental	Proinflammatory	CCR2 KO
Late ischemic stroke	Beneficial	Antiinflammatory	CCR2 KO
Retinal degeneration	Beneficial	Phagocytosis of cell debris and Antiinflammatory	Inhibiting SDF1 or monocyte depletion

Abbreviations: AD, Alzheimer's disease; BMDMs, Bone marrow-derived macrophages; CCL2, C-C motif chemokine ligand 2; CCR2, C-C motif chemokine receptor 2; MS, Multiple sclerosis; SCI, Spinal cord injury; SDF1, Stromal-derived factor 1; TBI, Traumatic brain injury; WNV, West Nile virus.

showed no protective effect.<sup>128</sup> These findings suggest that BMT with corrected gene expression is an effective treatment for Rett syndrome.

### 3.3 | Trichotillomania

Trichotillomania is an obsessive-compulsive disorder that commonly affects juveniles. Apart from hair and skin damage, patients with trichotillomania usually develop psychiatric disorders, including emotional stress and social anxiety. The exact cause of trichotillomania is still not clear, but three mouse models (Hoxb8 KO, Sapap3 KO, and Slitrk5 KO) with elevated grooming behaviors have been developed to study the pathophysiology of trichotillomania. Only microglia express detectable Hoxb8 in the brain, and thus they are the most affected cells in the brain of the Hoxb8 mutant mouse. Therefore, infiltration of BMDMs with regular Hoxb8 gene expression after BMT can rescue pathological grooming behaviors, indicating a potential therapeutic effect. 133,134

#### 3.4 X-linked adrenoleukodystrophy (ALD)

ALD is a peroxisomal membrane transporter protein encoded by the *ABCD1* gene located on the X chromosome. *ABCD1* gene mutations can cause ALD with progressive demyelination, which is lethal. BMT has shown beneficial effects in these patients, and BMT with no suitable donor cells has also shown beneficial results for patients by using engineered cells. <sup>135,136</sup> The potential mechanisms of BMT for preventing CNS demyelination remain unclear, but the effect is likely due to the engraftment of BMDMs expressing normal ALD. <sup>136</sup>

#### 4 | CONCLUSION

Similar to microglia, BMDMs play dual roles in various CNS diseases, and in different disease conditions the evidence supporting the beneficial or detrimental effects of BMDMs are substantial and consistent (Table 1). For example, BMDMs play protective roles in the early stage of AD by phagocytosing A $\beta$ , and in retinal degeneration by phagocytosing cell debris and resolving inflammation, while they can be harmful in MS and TBI by secreting proinflammatory cytokines. In addition, BMDMs play opposite roles in the early and late development of ischemic stroke. Therefore, promoting or inhibiting BMDMs within a suitable time window may slow disease progression and facilitate brain recovery.

Gene therapies are powerful therapeutic tools for genetic diseases, but their delivery into the CNS remains challenging. BMDMs have proven to be a powerful tool for delivering therapeutic genes to injury sites and throughout the CNS. However, BMT after either whole-body irradiation or chemotherapy may cause severe side effects and thus should only be used in the clinic in the case of

life-threatening conditions. Moreover, the number of BMDMs is relatively low following BMT. However, combining BMT with microglial depletion can significantly increase the replacement rate of dysfunctional microglia, thus exerting potentially therapeutic effects on neurological diseases.

#### **AUTHOR CONTRIBUTIONS**

K.Z. wrote the manuscript draft and prepared the figures. All other authors edited and revised the manuscript. All authors read and approved the final manuscript for publication.

#### **ACKNOWLEDGEMENTS**

We would like to thank BioRender.com for providing the template and tools for creating the figures. Figure 1 was Adapted from "Fibrinogen-Activated Microglia Contribute to Tissue Damage in Multiple Sclerosis", by BioRender.com (2020). Retrieved from https://app.biorender.com/biorender-templates. Figure 2 was created with BioRender.com.

#### **FUNDING INFORMATION**

This study was supported by the National Nature Science Foundation of China (U21A20347), the Swedish Childhood Cancer Foundation (PR2018-0082, PR2021-0020), the Swedish Cancer Foundation (20-1121-PjF), and Swedish Governmental grants to scientists working in health care (ALFGBG-965197).

#### **CONFLICT OF INTEREST**

The authors declare no competing interest.

#### DATA AVAILABILITY STATEMENT

Data sharing is not applicable to this article as no new data were created or analyzed in this study.

#### ORCID

Kai Zhou https://orcid.org/0000-0003-1026-757X

Jinming Han https://orcid.org/0000-0002-6084-3275

Changlian Zhu https://orcid.org/0000-0002-5029-6730

#### **REFERENCES**

- Geissmann F, Jung S, Littman DR. Blood monocytes consist of two principal subsets with distinct migratory properties. *Immunity*. 2003;19(1):71-82.
- Swirski FK, Nahrendorf M, Etzrodt M, et al. Identification of splenic reservoir monocytes and their deployment to inflammatory sites. Science. 2009;325(5940):612-616.
- Ziegler-Heitbrock L, Ancuta P, Crowe S, et al. Nomenclature of monocytes and dendritic cells in blood. Blood. 2010;116(16):e74-e80.
- Bianchini M, Duchêne J, Santovito D, et al. PD-L1 expression on nonclassical monocytes reveals their origin and immunoregulatory function. Sci Immunol. 2019;4(36):eaar3054.
- Geissmann F, Manz MG, Jung S, Sieweke MH, Merad M, Ley K. Development of monocytes, macrophages, and dendritic cells. *Science*. 2010;327(5966):656-661.
- Sunderkotter C, Nikolic T, Dillon MJ, et al. Subpopulations of mouse blood monocytes differ in maturation stage and inflammatory response. J Immunol. 2004;172(7):4410-4417.

- Lund H, Pieber M, Parsa R, et al. Competitive repopulation of an empty microglial niche yields functionally distinct subsets of microglia-like cells. Nat Commun. 2018;9(1):4845.
- Sailor KA, Agoranos G, López-Manzaneda S, et al. Hematopoietic stem cell transplantation chemotherapy causes microglia senescence and peripheral macrophage engraftment in the brain. *Nat Med.* 2022;28(3):517-527.
- Mildner A, Schmidt H, Nitsche M, et al. Microglia in the adult brain arise from Ly-6ChiCCR2+ monocytes only under defined host conditions. Nat Neurosci. 2007;10(12):1544-1553.
- Yang H, Ni W, Wei P, et al. HDAC inhibition reduces white matter injury after intracerebral hemorrhage. J Cereb Blood Flow Metab. 2021:41(5):958-974.
- Dietrich J-B. The adhesion molecule ICAM-1 and its regulation in relation with the blood-brain barrier. J Neuroimmunol. 2002;128(1-2):58-68.
- Turowski P, Adamson P, Greenwood J. Pharmacological targeting of ICAM-1 signaling in brain endothelial cells: potential for treating neuroinflammation. Cell Mol Neurobiol. 2005;25(1):153-170.
- Floris S, Ruuls SR, Wierinckx A, et al. Interferon-β directly influences monocyte infiltration into the central nervous system. J Neuroimmunol. 2002;127(1–2):69-79.
- Prinz M, Priller J. Tickets to the brain: role of CCR2 and CX3CR1 in myeloid cell entry in the CNS. J Neuroimmunol. 2010;224(1-2):80-84.
- Varvel NH, Neher JJ, Bosch A, et al. Infiltrating monocytes promote brain inflammation and exacerbate neuronal damage after status epilepticus. Proc Natl Acad sci U S A. 2016;113(38):E5665-E5674.
- Lyu J, Xie D, Bhatia TN, Leak RK, Hu X, Jiang X. Microglial/macrophage polarization and function in brain injury and repair after stroke. CNS Neurosci Ther. 2021;27(5):515-527.
- Larochelle A, Bellavance MA, Michaud JP, Rivest S. Bone marrow-derived macrophages and the CNS: an update on the use of experimental chimeric mouse models and bone marrow transplantation in neurological disorders. *Biochim Biophys Acta*. 2016;1862(3):310-322.
- Bennett ML, Bennett FC, Liddelow SA, et al. New tools for studying microglia in the mouse and human CNS. Proc Natl Acad sci U S A. 2016;113(12):E1738-E1746.
- Butovsky O, Jedrychowski MP, Moore CS, et al. Identification of a unique TGF-β-dependent molecular and functional signature in microglia. *Nat Neurosci.* 2014;17(1):131-143.
- 20. Koso H, Tsuhako A, Lai CY, et al. Conditional rod photoreceptor ablation reveals Sall1 as a microglial marker and regulator of microglial morphology in the retina. *Glia*. 2016;64(11):2005-2024.
- Xu J, Chen Z, Yu F, et al. IL-4/STAT6 signaling facilitates innate hematoma resolution and neurological recovery after hemorrhagic stroke in mice. Proc Natl Acad sci U S A. 2020;117(51):32679-32690.
- Quarta A, Berneman Z, Ponsaerts P. Functional consequences of a close encounter between microglia and brain-infiltrating monocytes during CNS pathology and repair. J Leukoc Biol. 2021;110(1):89-106.
- Gopinath A, Collins A, Khoshbouei H, Streit WJ. Microglia and other myeloid cells in central nervous system health and disease. J Pharmacol Exp Ther. 2020;375(1):154-160.
- Klein RS, Garber C, Howard N. Infectious immunity in the central nervous system and brain function. *Nat Immunol*. 2017;18(2):132-141.
- Chhatbar C, Prinz M. The roles of microglia in viral encephalitis: from sensome to therapeutic targeting. *Cell Mol Immunol*. 2021;18(2):250-258.
- Lim JK, Obara CJ, Rivollier A, Pletnev AG, Kelsall BL, Murphy PM. Chemokine receptor Ccr2 is critical for monocyte accumulation and survival in West Nile virus encephalitis. J Immunol. 2011;186(1):471-478.

- Ben-Nathan D, Huitinga I, Lustig S, van Rooijen N, Kobiler D. West Nile virus neuroinvasion and encephalitis induced by macrophage depletion in mice. *Arch Virol*. 1996;141(3–4):459-469.
- Getts DR, Terry RL, Getts MT, et al. Ly6c+ "inflammatory monocytes" are microglial precursors recruited in a pathogenic manner in West Nile virus encephalitis. J Exp Med. 2008:205(10):2319-2337.
- Getts DR, Terry RL, Getts MT, et al. Targeted blockade in lethal West Nile virus encephalitis indicates a crucial role for very late antigen (VLA)-4-dependent recruitment of nitric oxide-producing macrophages. J Neuroinflammation. 2012;9(1):1-8.
- Käufer C, Chhatbar C, Bröer S, et al. Chemokine receptors CCR2 and CX3CR1 regulate viral encephalitis-induced hippocampal damage but not seizures. Proc Natl Acad sci U S A. 2018;115(38):E8 929-e8938
- 31. Yamasaki R, Lu H, Butovsky O, et al. Differential roles of microglia and monocytes in the inflamed central nervous system. *J Exp Med*. 2014;211(8):1533-1549.
- 32. Valentin-Torres A, Savarin C, Hinton DR, Phares TW, Bergmann CC, Stohlman SA. Sustained TNF production by central nervous system infiltrating macrophages promotes progressive autoimmune encephalomyelitis. *J Neuroinflammation*. 2016;13:46.
- Ajami B, Bennett JL, Krieger C, McNagny KM, Rossi FM. Infiltrating monocytes trigger EAE progression, but do not contribute to the resident microglia pool. *Nat Neurosci.* 2011;14(9):1142-1149.
- Steinman L. Blocking adhesion molecules as therapy for multiple sclerosis: natalizumab. Nat Rev Drug Discov. 2005;4(6):510-518.
- Fife BT, Huffnagle GB, Kuziel WA, Karpus WJ. CC chemokine receptor 2 is critical for induction of experimental autoimmune encephalomyelitis. J Exp Med. 2000;192(6):899-905.
- Izikson L, Klein RS, Charo IF, Weiner HL, Luster AD. Resistance to experimental autoimmune encephalomyelitis in mice lacking the CC chemokine receptor (CCR)2. J Exp Med. 2000;192(7):1075-1080.
- Boven LA, Van Meurs M, Van Zwam M, et al. Myelin-laden macrophages are anti-inflammatory, consistent with foam cells in multiple sclerosis. *Brain*. 2006;129(Pt 2):517-526.
- 38. Alam A, Thelin EP, Tajsic T, et al. Cellular infiltration in traumatic brain injury. *J Neuroinflammation*. 2020;17(1):328.
- Chen Y, Hallenbeck JM, Ruetzler C, et al. Overexpression of monocyte chemoattractant protein 1 in the brain exacerbates ischemic brain injury and is associated with recruitment of inflammatory cells. J Cereb Blood Flow Metab. 2003;23(6):748-755.
- Morganti JM, Jopson TD, Liu S, et al. CCR2 antagonism alters brain macrophage polarization and ameliorates cognitive dysfunction induced by traumatic brain injury. J Neurosci. 2015;35(2):748-760.
- 41. Gyoneva S, Kim D, Katsumoto A, Kokiko-Cochran ON, Lamb BT, Ransohoff RM. Ccr2 deletion dissociates cavity size and tau pathology after mild traumatic brain injury. *J Neuroinflammation*. 2015;12:228.
- 42. Hsieh CL, Niemi EC, Wang SH, et al. CCR2 deficiency impairs macrophage infiltration and improves cognitive function after traumatic brain injury. *J Neurotrauma*. 2014;31(20):1677-1688.
- Makinde HM, Cuda CM, Just TB, Perlman HR, Schwulst SJ. Nonclassical monocytes mediate secondary injury, neurocognitive outcome, and neutrophil infiltration after traumatic brain injury. J Immunol. 2017;199(10):3583-3591.
- 44. Makinde HM, Just TB, Cuda CM, Bertolino N, Procissi D, Schwulst SJ. Monocyte depletion attenuates the development of posttraumatic hydrocephalus and preserves white matter integrity after traumatic brain injury. PLoS One. 2018;13(11):e0202722.
- 45. Semple BD, Bye N, Rancan M, Ziebell JM, Morganti-Kossmann MC. Role of CCL2 (MCP-1) in traumatic brain injury (TBI): evidence from severe TBI patients and CCL2-/- mice. *J Cereb Blood Flow Metab*. 2010;30(4):769-782.

- Chou A, Krukowski K, Morganti JM, Riparip LK, Rosi S. Persistent infiltration and impaired response of peripherally-derived monocytes after traumatic brain injury in the aged brain. Int J Mol sci. 2018;19(6):1616.
- Shechter R, London A, Varol C, et al. Infiltrating blood-derived macrophages are vital cells playing an anti-inflammatory role in recovery from spinal cord injury in mice. PLoS Med. 2009;6(7):e1000113.
- 48. Wang J, Wang J, Wang J, Yang B, Weng Q, He Q. Targeting microglia and macrophages: a potential treatment strategy for multiple sclerosis. *Front Pharmacol.* 2019;10:286.
- 49. GrandPré T, Nakamura F, Vartanian T, Strittmatter SM. Identification of the Nogo inhibitor of axon regeneration as a reticulon protein. *Nature*. 2000;403(6768):439-444.
- Kotter MR, Li WW, Zhao C, Franklin RJ. Myelin impairs CNS remyelination by inhibiting oligodendrocyte precursor cell differentiation. J Neurosci. 2006;26(1):328-332.
- Milich LM, Ryan CB, Lee JK. The origin, fate, and contribution of macrophages to spinal cord injury pathology. *Acta Neuropathol*. 2019;137(5):785-797.
- Zhu Y, Soderblom C, Krishnan V, Ashbaugh J, Bethea JR, Lee JK. Hematogenous macrophage depletion reduces the fibrotic scar and increases axonal growth after spinal cord injury. *Neurobiol Dis*. 2015;74:114-125.
- Popovich PG, Guan Z, Wei P, Huitinga I, van Rooijen N, Stokes BT.
   Depletion of hematogenous macrophages promotes partial hind-limb recovery and neuroanatomical repair after experimental spinal cord injury. Exp Neurol. 1999;158(2):351-365.
- Iannotti CA, Clark M, Horn KP, van Rooijen N, Silver J, Steinmetz MP. A combination immunomodulatory treatment promotes neuroprotection and locomotor recovery after contusion SCI. Exp Neurol. 2011;230(1):3-15.
- Lee SM, Rosen S, Weinstein P, van Rooijen N, Noble-Haeusslein LJ. Prevention of both neutrophil and monocyte recruitment promotes recovery after spinal cord injury. J Neurotrauma. 2011;28(9):1893-1907.
- 56. Hampel H, Hardy J, Blennow K, et al. The amyloid-beta pathway in Alzheimer's disease. *Mol Psychiatry*. 2021;26(10):5481-5503.
- Frackowiak J, Potempska A, LeVine H, Haske T, Dickson D, Mazur-Kolecka B. Extracellular deposits of a beta produced in cultures of Alzheimer disease brain vascular smooth muscle cells. J Neuropathol Exp Neurol. 2005;64(1):82-90.
- Parhizkar S, Arzberger T, Brendel M, et al. Loss of TREM2 function increases amyloid seeding but reduces plaque-associated ApoE. *Nat Neurosci.* 2019;22(2):191-204.
- Venegas C, Kumar S, Franklin BS, et al. Microglia-derived ASC specks cross-seed amyloid-beta in Alzheimer's disease. *Nature*. 2017;552(7685):355-361.
- Ziegler-Waldkirch S, d'Errico P, Sauer JF, et al. Seed-induced Aβ deposition is modulated by microglia under environmental enrichment in a mouse model of Alzheimer's disease. EMBO J. 2018:37(2):167-182.
- 61. d'Errico P, Ziegler-Waldkirch S, Aires V, et al. Microglia contribute to the propagation of Abeta into unaffected brain tissue. *Nat Neurosci.* 2022;25(1):20-25.
- 62. Lee CY, Landreth GE. The role of microglia in amyloid clearance from the AD brain. J Neural Transm (Vienna). 2010;117(8):949-960.
- Liu Y, Walter S, Stagi M, et al. LPS receptor (CD14): a receptor for phagocytosis of Alzheimer's amyloid peptide. *Brain*. 2005;128(Pt 8):1778-1789.
- 64. Hohsfield LA, Humpel C. Migration of blood cells to beta-amyloid plaques in Alzheimer's disease. *Exp Gerontol.* 2015;65:8-15.
- 65. Simard AR, Soulet D, Gowing G, Julien JP, Rivest S. Bone marrowderived microglia play a critical role in restricting senile plaque formation in Alzheimer's disease. *Neuron*. 2006;49(4):489-502.
- El Khoury J, Toft M, Hickman SE, et al. Ccr2 deficiency impairs microglial accumulation and accelerates progression of Alzheimerlike disease. Nat Med. 2007;13(4):432-438.

- 67. Naert G, Rivest S. CC chemokine receptor 2 deficiency aggravates cognitive impairments and amyloid pathology in a transgenic mouse model of Alzheimer's disease. *J Neurosci.* 2011;31(16):6208-6220.
- Naert G, Rivest S. Hematopoietic CC-chemokine receptor 2 (CCR2) competent cells are protective for the cognitive impairments and amyloid pathology in a transgenic mouse model of Alzheimer's disease. Mol Med. 2012:18:297-313.
- 69. Naert G, Rivest S. A deficiency in CCR2+ monocytes: the hidden side of Alzheimer's disease. *J Mol Cell Biol.* 2013;5(5):284-293.
- Boissonneault V, Filali M, Lessard M, Relton J, Wong G, Rivest S. Powerful beneficial effects of macrophage colony-stimulating factor on beta-amyloid deposition and cognitive impairment in Alzheimer's disease. *Brain*. 2009;132(Pt 4):1078-1092.
- 71. Zhang W, Zhao J, Wang R, et al. Macrophages reprogram after ischemic stroke and promote efferocytosis and inflammation resolution in the mouse brain. CNS Neurosci Ther. 2019:25(12):1329-1342.
- Wang R, Liu Y, Ye Q, et al. RNA sequencing reveals novel macrophage transcriptome favoring neurovascular plasticity after ischemic stroke. J Cereb Blood Flow Metab. 2020;40(4):720-738.
- Ritzel RM, Patel AR, Grenier JM, et al. Functional differences between microglia and monocytes after ischemic stroke. J Neuroinflammation. 2015;12:106.
- Kronenberg G, Uhlemann R, Richter N, et al. Distinguishing features of microglia- and monocyte-derived macrophages after stroke. Acta Neuropathol. 2018;135(4):551-568.
- Chu HX, Broughton BR, Kim HA, Lee S, Drummond GR, Sobey CG. Evidence that Ly6C(hi) monocytes are protective in acute ischemic stroke by promoting M2 macrophage polarization. Stroke. 2015;46(7):1929-1937.
- Pedragosa J, Miró-Mur F, Otxoa-de-Amezaga A, et al. CCR2 deficiency in monocytes impairs angiogenesis and functional recovery after ischemic stroke in mice. J Cereb Blood Flow Metab. 2020;40(1\_suppl):S98-S116.
- Lampron A, Pimentel-Coelho PM, Rivest S. Migration of bone marrow-derived cells into the central nervous system in models of neurodegeneration. J Comp Neurol. 2013;521(17):3863-3876.
- Kim E, Yang J, Beltran CD, Cho S. Role of spleen-derived monocytes/macrophages in acute ischemic brain injury. J Cereb Blood Flow Metab. 2014;34(8):1411-1419.
- ElAli A, Jean LeBlanc N. The role of monocytes in ischemic stroke pathobiology: new avenues to explore. Front Aging Neurosci. 2016;8:29.
- Fang W, Zhai X, Han D, et al. CCR2-dependent monocytes/ macrophages exacerbate acute brain injury but promote functional recovery after ischemic stroke in mice. *Theranostics*. 2018;8(13):3530-3543.
- Lei X, Li H, Li M, et al. The novel Nrf2 activator CDDO-EA attenuates cerebral ischemic injury by promoting microglia/macrophage polarization toward M2 phenotype in mice. CNS Neurosci Ther. 2021;27(1):82-91.
- 82. Cai M, Sun S, Wang J, et al. Sevoflurane preconditioning protects experimental ischemic stroke by enhancing anti-inflammatory microglia/macrophages phenotype polarization through GSK-3β/ Nrf2 pathway. *CNS Neurosci Ther.* 2021;27(11):1348-1365.
- 83. Kaneko H, Nishiguchi KM, Nakamura M, Kachi S, Terasaki H. Characteristics of bone marrow-derived microglia in the normal and injured retina. *Invest Ophthalmol Vis sci.* 2008;49(9):4162-4168.
- 84. Sasahara M, Otani A, Oishi A, et al. Activation of bone marrowderived microglia promotes photoreceptor survival in inherited retinal degeneration. *Am J Pathol.* 2008;172(6):1693-1703.
- 85. Heeschen C, Aicher A, Lehmann R, et al. Erythropoietin is a potent physiologic stimulus for endothelial progenitor cell mobilization. *Blood*. 2003;102(4):1340-1346.

- 86. London A, Itskovich E, Benhar I, et al. Neuroprotection and progenitor cell renewal in the injured adult murine retina requires healing monocyte-derived macrophages. *J Exp Med*. 2011;208(1):23-39.
- 87. Jin N, Gao L, Fan X, Xu H. Friend or foe? Resident microglia vs bone marrow-derived microglia and their roles in the retinal degeneration. *Mol Neurobiol*. 2017:54(6):4094-4112.
- Haurigot V, Villacampa P, Ribera A, et al. Increased intraocular insulin-like growth factor-l triggers blood-retinal barrier breakdown. J Biol Chem. 2009:284(34):22961-22969.
- 89. Aguzzi A, Barres BA, Bennett ML. Microglia: scapegoat, saboteur, or something else? *Science*. 2013;339(6116):156-161.
- Colonna M, Butovsky O. Microglia function in the central nervous system during health and neurodegeneration. Annu Rev Immunol. 2017;35:441-468
- 91. Han J, Sarlus H, Wszolek ZK, Karrenbauer VD, Harris RA. Microglial replacement therapy: a potential therapeutic strategy for incurable CSF1R-related leukoencephalopathy. *Acta Neuropathol Commun*. 2020;8(1):217.
- Konno T, Kasanuki K, Ikeuchi T, Dickson DW, Wszolek ZK. CSF1Rrelated leukoencephalopathy: a major player in primary microgliopathies. *Neurology*. 2018;91(24):1092-1104.
- Tipton PW, Kenney-Jung D, Rush BK, et al. Treatment of CSF1Rrelated leukoencephalopathy: breaking new ground. Mov Disord. 2021;36(12):2901-2909.
- Prinz M, Priller J. Microglia and brain macrophages in the molecular age: from origin to neuropsychiatric disease. *Nat Rev Neurosci*. 2014;15(5):300-312.
- Krivit W, Sung JH, Shapiro EG, Lockman LA. Microglia: the effector cell for reconstitution of the central nervous system following bone marrow transplantation for lysosomal and peroxisomal storage diseases. *Cell Transplant*. 1995;4(4):385-392.
- 96. Pastores GM. Therapeutic approaches for lysosomal storage diseases. Ther Adv Endocrinol Metab. 2010;1(4):177-188.
- Prinz M, Priller J, Sisodia SS, Ransohoff RM. Heterogeneity of CNS myeloid cells and their roles in neurodegeneration. *Nat Neurosci*. 2011;14(10):1227-1235.
- Bennett ML, Bennett FC. The influence of environment and origin on brain resident macrophages and implications for therapy. *Nat Neurosci.* 2020;23(2):157-166.
- 99. Zhang J, Shi XQ, Echeverry S, Mogil JS, De Koninck Y, Rivest S. Expression of CCR2 in both resident and bone marrow-derived microglia plays a critical role in neuropathic pain. *J Neurosci.* 2007;27(45):12396-12406.
- Biffi A, De Palma M, Quattrini A, et al. Correction of metachromatic leukodystrophy in the mouse model by transplantation of genetically modified hematopoietic stem cells. J Clin Invest. 2004;113(8):1118-1129.
- Capotondo A, Milazzo R, Politi LS, et al. Brain conditioning is instrumental for successful microglia reconstitution following hematopoietic stem cell transplantation. *Proc Natl Acad sci U S A*. 2012;109(37):15018-15023.
- 102. Duran-Struuck R, Dysko RC. Principles of bone marrow transplantation (BMT): providing optimal veterinary and husbandry care to irradiated mice in BMT studies. *J Am Assoc Lab Anim sci.* 2009;48(1):11-22.
- Bellavance MA, Gosselin D, Yong VW, Stys PK, Rivest S. Patrolling monocytes play a critical role in CX3CR1-mediated neuroprotection during excitotoxicity. *Brain Struct Funct*. 2015;220(3):1759-1776.
- 104. Ginhoux F, Lim S, Hoeffel G, Low D, Huber T. Origin and differentiation of microglia. *Front Cell Neurosci.* 2013;7:45.
- 105. Guilliams M, Thierry GR, Bonnardel J, Bajenoff M. Establishment and maintenance of the macrophage niche. *Immunity*. 2020;52(3):434-451.
- 106. Hohsfield LA, Najafi AR, Ghorbanian Y, et al. Effects of long-term and brain-wide colonization of peripheral bone marrow-derived myeloid cells in the CNS. J Neuroinflammation. 2020;17(1):279.

- Xu Z, Rao Y, Huang Y, et al. Efficient strategies for microglia replacement in the central nervous system. Cell Rep. 2020;33(8):108443.
- Cannarile MA, Weisser M, Jacob W, Jegg AM, Ries CH, Rüttinger
   D. Colony-stimulating factor 1 receptor (CSF1R) inhibitors in cancer therapy. J Immunother Cancer. 2017;5(1):53.
- 109. Zhou K, Han J, Lund H, et al. An overlooked subset of Cx3cr1(wt/wt) microglia in the Cx3cr1(CreER-Eyfp/wt) mouse has a repopulation advantage over Cx3cr1(CreER-Eyfp/wt) microglia following microglial depletion. J Neuroinflammation. 2022;19(1):20.
- Clement AM, Nguyen MD, Roberts EA, et al. Wild-type nonneuronal cells extend survival of SOD1 mutant motor neurons in ALS mice. Science. 2003;302(5642):113-117.
- 111. Corti S, Locatelli F, Donadoni C, et al. Wild-type bone marrow cells ameliorate the phenotype of SOD1-G93A ALS mice and contribute to CNS, heart and skeletal muscle tissues. *Brain*. 2004;127(Pt 11):2518-2532.
- Ohashi N, Terashima T, Katagi M, et al. GLT1 gene delivery based on bone marrow-derived cells ameliorates motor function and survival in a mouse model of ALS. Sci Rep. 2021;11(1):12803.
- 113. Bigini P, Veglianese P, Andriolo G, et al. Intracerebroventricular administration of human umbilical cord blood cells delays disease progression in two murine models of motor neuron degeneration. *Rejuvenation Res.* 2011;14(6):623-639.
- 114. Gubert F, Decotelli AB, Bonacossa-Pereira I, et al. Intraspinal bone-marrow cell therapy at pre- and symptomatic phases in a mouse model of amyotrophic lateral sclerosis. Stem Cell Res Ther. 2016;7:41.
- 115. Gubert F, Bonacossa-Pereira I, Decotelli AB, et al. Bone-marrow mononuclear cell therapy in a mouse model of amyotrophic lateral sclerosis: functional outcomes from different administration routes. *Brain Res.* 2019;1712:73-81.
- 116. Vasques JF, Teixeira Pinheiro LC, de Jesus Gonçalves RG, Mendez-Otero R, Gubert F. Cell-based research and therapy for amyotrophic lateral sclerosis: Promises and challenges. In: Araki T, ed. Amyotrophic Lateral Sclerosis. Exon Publications; 2021.
- 117. Platt FM, d'Azzo A, Davidson BL, Neufeld EF, Tifft CJ. Lysosomal storage diseases. *Nat Rev Dis Primers*. 2018;4(1):27.
- Priller J, Flügel A, Wehner T, et al. Targeting gene-modified hematopoietic cells to the central nervous system: use of green fluorescent protein uncovers microglial engraftment. *Nat Med*. 2001;7(12):1356-1361.
- 119. Biffi A, Capotondo A, Fasano S, et al. Gene therapy of metachromatic leukodystrophy reverses neurological damage and deficits in mice. *J Clin Invest*. 2006;116(11):3070-3082.
- 120. Krivit W, Aubourg P, Shapiro E, Peters C. Bone marrow transplantation for globoid cell leukodystrophy, adrenoleukodystrophy, metachromatic leukodystrophy, and hurler syndrome. *Curr Opin Hematol.* 1999;6(6):377-382.
- Peters C, Steward CG. Hematopoietic cell transplantation for inherited metabolic diseases: an overview of outcomes and practice guidelines. *Bone Marrow Transplant*. 2003;31(4):229-239.
- 122. Neufeld EF. Lysosomal storage diseases. Annu Rev Biochem. 1991;60:257-280.
- 123. Wada R, Tifft CJ, Proia RL. Microglial activation precedes acute neurodegeneration in Sandhoff disease and is suppressed by bone marrow transplantation. *Proc Natl Acad sci U S A*. 2000;97(20):10954-10959.
- 124. Sano R, Tessitore A, Ingrassia A, d'Azzo A. Chemokine-induced recruitment of genetically modified bone marrow cells into the CNS of GM1-gangliosidosis mice corrects neuronal pathology. *Blood*. 2005;106(7):2259-2268.
- Kyle SM, Vashi N, Justice MJ. Rett syndrome: a neurological disorder with metabolic components. *Open Biol*. 2018;8(2):170216.
- 126. Feldman D, Banerjee A, Sur M. Developmental dynamics of Rett syndrome. *Neural Plast*. 2016;2016:6154080-6154089.

- 127. Amir RE, Van den Veyver IB, Wan M, Tran CQ, Francke U, Zoghbi HY. Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2. *Nat Genet*. 1999;23(2):185-188.
- Derecki NC, Cronk JC, Lu Z, et al. Wild-type microglia arrest pathology in a mouse model of Rett syndrome. *Nature*. 2012;484(7392):105-109.
- 129. Grant JE, Chamberlain SR. Trichotillomania. *Am J Psychiatry*. 2016;173(9):868-874.
- 130. Greer JM, Capecchi MR. Hoxb8 is required for normal grooming behavior in mice. *Neuron*. 2002;33(1):23-34.
- 131. Welch JM, Lu J, Rodriguiz RM, et al. Cortico-striatal synaptic defects and OCD-like behaviours in Sapap3-mutant mice. *Nature*. 2007;448(7156):894-900.
- 132. Shmelkov SV, Hormigo A, Jing D, et al. Slitrk5 deficiency impairs corticostriatal circuitry and leads to obsessive-compulsive-like behaviors in mice. *Nat Med.* 2010;16(5):598-602.
- 133. Chen SK, Tvrdik P, Peden E, et al. Hematopoietic origin of pathological grooming in Hoxb8 mutant mice. *Cell*. 2010;141(5):775-785.

- Frick LR, Williams K, Pittenger C. Microglial dysregulation in psychiatric disease. Clin Dev Immunol. 2013;2013:608654.
- 135. Cartier N, Hacein-Bey-Abina S, Bartholomae CC, et al. Hematopoietic stem cell gene therapy with a lentiviral vector in X-linked adrenoleukodystrophy. *Science*. 2009;326(5954):818-823.
- Cartier N, Aubourg P. Hematopoietic stem cell transplantation and hematopoietic stem cell gene therapy in X-linked adrenoleukodystrophy. *Brain Pathol.* 2010;20(4):857-862.

How to cite this article: Zhou K, Han J, Wang Y, Xu Y, Zhang Y, Zhu C. The therapeutic potential of bone marrow-derived macrophages in neurological diseases. *CNS Neurosci Ther.* 2022;28:1942-1952. doi: 10.1111/cns.13964