Genome-Wide Association Study of Gene by Smoking Interactions in Coronary Artery Calcification

Linda M. Polfus¹, Jennifer A. Smith², Lawrence C. Shimmin¹, Lawrence F. Bielak², Alanna C. Morrison¹, Sharon L. R. Kardia², Patricia A. Peyser², James E. Hixson²*

1 Division of Epidemiology, Human Genetics and Environmental Sciences, The University of Texas Health Science Center at Houston, Houston, Texas, United States of America, 2 Department of Epidemiology, University of Michigan, Ann Arbor, Michigan, United States of America

Abstract

Many GWAS have identified novel loci associated with common diseases, but have focused only on main effects of individual genetic variants rather than interactions with environmental factors (GxE). Identification of GxE interactions is particularly important for coronary heart disease (CHD), a major preventable source of morbidity and mortality with strong non-genetic risk factors. Atherosclerosis is the major cause of CHD, and coronary artery calcification (CAC) is directly correlated with guantity of coronary atherosclerotic plaque. In the current study, we tested for genetic variants influencing extent of CAC via interaction with smoking (GxS), by conducting a GxS discovery GWAS in Genetic Epidemiology Network of Arteriopathy (GENOA) sibships (N = 915 European Americans) followed by replication in Framingham Heart Study (FHS) sibships (N = 1025 European Americans). Generalized estimating equations accounted for the correlation within sibships in strata-specific groups of smokers and nonsmokers, as well as GxS interaction. Primary analysis found SNPs that showed suggestive associations ($p \le 10^{-5}$) in GENOA GWAS, but these index SNPs did not replicate in FHS. However, secondary analysis was able to replicate candidate gene regions in FHS using other SNPs (+/-250 kb of GENOA index SNP). In smoker and nonsmoker groups, replicated genes included TCF7L2 ($p = 6.0 \times 10^{-5}$) and WWOX ($p = 4.5 \times 10^{-6}$); and TNFRSF8 $(p = 7.8 \times 10^{-5})$, respectively. For GxS interactions, replicated genes included TBC1D4 $(p = 6.9 \times 10^{-5})$ and ADAMTS9 $(P = 7.1 \times 10^{-5})$. Interestingly, these genes are involved in inflammatory pathways mediated by the NF- κ B axis. Since smoking is known to induce chronic and systemic inflammation, association of these genes likely reflects roles in CAC development via inflammatory pathways. Furthermore, the NF-κB axis regulates bone remodeling, a key physiological process in CAC development. In conclusion, GxS GWAS has yielded evidence for novel loci that are associated with CAC via interaction with smoking, providing promising new targets for future population-based and functional studies of CAC development.

Citation: Polfus LM, Smith JA, Shimmin LC, Bielak LF, Morrison AC, et al. (2013) Genome-Wide Association Study of Gene by Smoking Interactions in Coronary Artery Calcification. PLoS ONE 8(10): e74642. doi:10.1371/journal.pone.0074642

Editor: Dan E. Arking, Johns Hopkins University, United States of America

Received April 3, 2013; Accepted August 5, 2013; Published October 3, 2013

Copyright: © 2013 Polfus et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work was supported by grants from the National Heart, Lung, and Blood Institute of the National Institutes of Health (R01HL091988 to J.E.H., R01HL87660 to S.L.R.K.), and in part by a training fellowship from the Burroughs Wellcome Fund - The Houston Laboratory and Population Science Training Program in Gene-Environment Interaction of the Brown Foundation Institute of Molecular Medicine of UTHealth Medical School (BWF Grant No. 1008200 to L.M.P). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: James.E.Hixson@uth.tmc.edu

Introduction

Recent genome-wide association studies (GWAS) have identified numerous novel loci associated with common diseases and their risk factors. However, GWAS have typically focused only on main effects of individual genetic variants, rather than interactions with other genes (epistasis) and with environmental factors (GxE interactions). Although GxE interactions provide a well-established paradigm for progression of complex chronic diseases, more precise biological and statistical characterization of this interplay remains elusive [1]. Identification of GxE interactions is particularly important for coronary heart disease (CHD), a major preventable source of morbidity and mortality with strong nongenetic risk factors such as physical activity, diet, and smoking.

Atherosclerosis is the major cause of CHD, and extent of coronary atherosclerosis is the most powerful predictor of subsequent clinical events. Non-invasive imaging of coronary artery calcification (CAC) has emerged as a useful method to assess CHD risk. The quantity of CAC, measured by computed tomography (CT), is heritable [2,3]. and correlates directly with the quantity of coronary atherosclerotic plaque. Furthermore, CAC scores predict all cause mortality [4] and coronary outcomes in asymptomatic individuals as shown in a cohort of over 10,000 individuals followed for 5 years [5,6]. A CAC score >100 has demonstrated clinical relevance representing the transition from mild to moderate coronary atherosclerosis [7,8]. Additionally, a CAC score >100 is associated with a 7 fold increased risk for myocardial infarction (MI) and CHD death after adjusting for traditional risk factors [9].

A recent CAC GWAS conducted by the Cohorts for Heart and Aging Research in Genomic Epidemiology consortium (CHARGE) included five independent cohorts for discovery and three cohorts for replication [10]. The strongest SNP associations for CAC quantity and score >100 were found on chromosome 9p21 (top SNPs near *CDKN2A* and *CDKN2B*) and within the *PHACTR1* gene on chromosome 6p24. These same regions are associated with early CHD. To date, no GWAS has investigated associations between GxE interactions and CAC. Cigarette smoking, a major risk factor for CHD, is associated with CAC. In a study of over 30,000 asymptomatic adults, Hoff et al. reported a significant, independent association between having ever smoked and CAC score >100 (OR = 1.8 in men and 1.5 in women) [11]. Recently, North et al. found several chromosomal regions with evidence for linkage with CAC quantities only in nonsmokers (chromosomes 4 and 6) or only in smokers (chromosomes 11 and 13), with significant genotype by smoking interactions (p<0.05) [12].

In the current study, we used GWAS to test for genes that interact with smoking for CAC score >100 in European Americans from the Genetic Epidemiology Network of Arteriopathy study (GENOA). Our primary analysis included GWAS analysis of GENOA subgroups stratified by smoking status, as well as genome-wide tests for variants that show significant Gene by Smoking interactions (GxS). Primary analysis also attempted to replicate GENOA index SNPs that showed suggestive associations ($p < 10^{-5}$) in FHS. For secondary analysis, we tested SNPs located in candidate gene regions (+/-250 kb of GENOA index SNP) for associations with CAC in the Framingham Heart Study (genebased strategy).

Methods

Ethics Statement

These studies received approval from Institutional Review Boards (University of Texas Health Science Center at Houston IRB, University of Michigan Health Sciences and Behavioral Sciences IRB, Boston University IRB), and study participants gave written informed consents.

Characteristics of Study Cohorts

The discovery cohort consisted of sibships of European ancestry who participated in the GENOA study as part of the NHLBI Family Blood Pressure Program (FBPP) (FBPP Investigators, 2002). Sibships containing at least two individuals diagnosed with essential hypertension before age 60 years were recruited from Rochester, Minnesota. All other siblings were invited to participate regardless of hypertensive status. Exclusion criteria were secondary hypertension, alcoholism or drug abuse, pregnancy, insulindependent diabetes mellitus, or active malignancy. GENOA sibships recruited during Phase 1 (N = 1,583, during 1995 to 2000) were invited to participate in Phase 2 (N = 1,241, during 2000 to 2004) and received electron beam CT scans of the heart. Other information collected by the GENOA study included demographic, environmental, anthropometric, and physiological data. Participant measurements of blood pressure and other clinical and physiological data have been described elsewhere [10]. Individuals with a history of coronary revascularization (n = 83)were excluded from measurement of CAC. Participants with a history of myocardial infarction (n = 19), stroke (n = 27), positive angiogram (n = 31), missing data (n = 2), or self-reporting as Hispanic ancestry (n = 2) were excluded from analyses. Of the 1,077 GENOA participants with CAC and risk factor measures, 915 participants (539 women and 376 men in 421 sibships) had genotype data.

The replication study participants were European Americans from the Offspring Cohort of the Framingham Heart Study (FHS) that participated in the Offspring Exam 7 (n = 1,314) conducted between 1998 and 2001. FHS participants were excluded from the analyses if considered to have a cardiovascular disease (CVD) event prior to Exam 7 (N = 160) yielding the final number of siblings with valid genotype data to 1,025 participants in 431 sibships. A CVD event was defined as the occurrence of coronary death, myocardial infarction, stable or unstable angina pectoris, atherothrombotic stroke, intermittent claudication, or cardiovascular death, and hospitalized coronary insufficiency. Risk factors were assessed from participants undergoing a routine physical examination, anthropometry, and laboratory collection during offspring examination [10].

GWAS Genotyping

GENOA participants were genotyped with the Affymetrix Genome-Wide Human SNP Array 6.0. A total of 669,293 SNPs were genotyped in the 915 participants after passing quality control measures including exclusion of SNPs with a call rate <95%, or a minor allele frequency (MAF) <0.01. These measured SNP genotypes were used for imputation (CEU HapMap haplotypes, release 22, build 36) by Markov Chain modeling (MACH version 1.0), yielding 2,171,047 SNPs for subsequent GWAS analyses (excluded SNPs with MAF <0.05) [13,14].

A total of 8,481 FHS participants were genotyped using the Affymetrix GeneChip Human Mapping 500K Array and an additional 50K Affymetrix gene-focused Molecular Inversion Probe (MIP) array. After excluding SNPs with a call rate <97% or MAF <0.01, 378,163 SNPs were used for imputation (CEU HapMap haplotypes, release 22, build 36) with MACH version 1.0.15, yielding 2,543,887 SNPs.

CAC Measurements

GENOA participants were imaged with an Imatron C-150 electron beam CT scanner (Imatron Inc.) as previously described (O'Donnell et al., 2011). Scan results were initially reviewed by a radiologist for technical quality, and then scored by a radiologic technologist. A focus of CAC was considered to exist if there were at least four contiguous pixels \geq 130 Hounsfield Units (HU) in density. The CAC score is a calculation based on the number, area, and density of CAC foci summed from the four major epicardial arteries using the method of Agatston et al. (1990).

For FHS participants, a multidetector CT exam was conducted between 2002 to 2005 with a calcified lesion in the coronary arteries defined as an area of at least 3 contiguous pixels >130 HUs with the use of 3-dimensional connectivity criteria (6 points). Agatson scores developed for electron beam CT scans were modified for multidetector CT scans as described previously (Parikh 2007). A CAC score cutpoint of 100 to define a qualitative outcome was used in all analyses and is referred to as CAC.

Categorization of smoking status

Smokers (current and previous) and nonsmokers (never) were classified based on self-report. Previous smokers were categorized by having smoked more than 100 cigarettes in a lifetime according to the National Health Interview Survey [15]. Self-reports were used for classification since biochemical measures (e.g., cotinine levels) of smoking amounts were not available for GENOA participants. A dichotomous categorization of smoking status rather than a quantitative measure (e.g., pack-years) was chosen due to the inherent high dimensionality of GWAS analysis (2.1 million SNPs with 4.2 million main effects and interaction variables per SNP).

Data Analysis

In order to account for the correlation among sibships, the Genome-Wide Association Analysis with Family (GWAF) package was utilized within R statistical software version 2.10 employing a generalized estimating equations (GEE) model [16]. Adjustment for additional covariates in the GEE model included age, sex, body

Table 1. Stratified descriptive statistics by gender and smoking for the discovery cohort (Genetic Epidemiology Network of Arteriopathy Study, GENOA) and the replication cohort (Framingham Heart Study).

	GENOA					
	Males (N = 376)			Females (N = 539)		
	Smoker (N=219)	Non-Smoker (N = 157)	P value	Smoker (N = 207)	Non-Smoker (N = 332)	P value
CAC>100 n(%) ^a	103 (69.1)	64 (54.7)	0.228	46 (30.9)	53 (45.3)	0.068
CAC≤100 n(%)	116 (41.9)	93 (25.0)		161 (58.1)	279 (75.0)	
CAC score	340.8	270.5		150.1	83.5	
Log (CAC +1) ^b	3.82±2.6	3.55±2.5	0.30	2.67±2.5	1.96±2.2	0.05
Age (years)	58.7±9.7	58.0±10.9	0.530	56.6±9.4	58.7±10.5	0.197
LDL-C (mg/dL)	113.3±27.7	122.8±31.8	0.002	116.3±30.5	115.9±32.0	0.868
BMI (kg/m ²)	30.8±5.1	30.7±5.1	0.829	30.5±7.6	30.8±6.6	0.685
Pulse Pressure	54.2±14.1	53.2±14.9	0.536	56.0±14.4	59.7±16.4	0.008
Diabetes n(%)	41 (18.7)	19 (12.1)	0.084	24 (11.6)	39 (11.8)	0.957
Hypertensive Meds n(%)	140 (63.9)	102 (65.0)	0.835	136 (65.7)	225 (67.8)	0.619
Lipid Lowering Meds n(%)	77 (35.2)	43 (27.4)	0.111	43 (20.8)	67 (20.2)	0.868
	FRAMINGHAM HEA	ART STUDY				
	Males (N = 459)			Females (N = 566)		
	Smoker (N = 259)	Non-Smoker (N = 200)	P value	Smoker (N = 309)	Non-Smoker (N = 257)	P value
CAC>100 n(%)	136 (52.5)	80 (0.40)	0.008	66 (21.4)	50 (19.5)	0.576
CAC≤100 n(%)	123 (47.5)	120 (0.60)		243 (78.6)	207 (80.5)	
CAC score	467.9	238.2		106.5	81.3	
Log (CAC +1)	4.31±2.51	3.22±2.55	<0.001	2.30±2.37	1.99±2.27	0.106
Age (years)	63.8±8.8	60.9±9.4	0.001	62.6±8.1	64.1±9.3	0.038
LDL-C (mg/dL)	119.5±31.3	122.9±29.0	0.229	121.9±36.1	119.6±30.7	0.408
BMI (kg/m²)	28.9±4.3	28.1±4.2	0.057	27.8±5.7	27.2±5.3	0.195
Pulse Pressure	50±13.1	48.5±14.3	0.258	49.6±15.2	51.0±15.0	0.257
Diabetes n(%)	21 (8.1)	11 (5.5)	0.277	17 (5.5)	18 (7.0)	0.460
Hypertensive Meds n(%)	80 (30.9)	58 (29.0)	0.662	86 (27.8)	74 (28.8)	0.802
Lipid Lowering Meds n(%)	39 (15.1)	30 (15 0)	0.986	37 (12 0)	39 (15 2)	0.266

CAC, coronary artery calcification; LDL-C, Low Density Lipoprotein-Cholesterol; BMI, body mass index.

^aFor categorical variables, the number and percent are presented (Chi-square P-values).

^bFor continuous variables, the mean and standard deviation are presented (t-test P-values).

doi:10.1371/journal.pone.0074642.t001

mass index (BMI), pulse pressure, diabetes, systolic blood pressure (SBP), use of anti-hypertensive medications, use of lipid lowering medications, and LDL-cholesterol. To assess population stratification in GENOA GWAS, the first ten principal components (PC) were calculated using Eigenstrat [17]. However, none of the PCs were significant (p<0.05) in GEE models (p = 0.82 for the first PC), so were not used in subsequent analyses. To assess interactions of genotype by smoking (GxS) status, subgroups of smokers/nonsmokers were analyzed separately. Results from the stratified analyses were then used to test for interaction in the combined group. Effect differences for smoking were assessed by testing if beta coefficients differed from zero for associations of individual SNPs with CAC. The test for interaction approximates a 2 sample t-test with the following null hypotheses:

$H_0: \beta_{smokers} = \beta_{nonsmokers}$

where $\beta_{smokers}$ is the beta coefficient for the SNP in smokers and $\beta_{nonsmokers}$ is the beta coefficient for the SNP in nonsmokers.

For all 2.1 million discovery SNPs, the interaction test statistic was calculated from the following equation as adapted from Heid et al. [18].

$$(\beta_{\rm smokers} - \beta_{\rm nonsmokers}) - \sqrt{(se_{\rm smokers}^2 + se_{\rm nonsmokers}^2)}$$

Primary analysis started with GxS GWAS in GENOA sibships, followed by testing of index SNPs that showed suggestive evidence for association ($p \le 10^{-5}$) in FHS sibships. A genome-wide significance threshold based on Bonferroni correction was used to account for multiple testing of 2.1 million SNPs ($p \le 2.3 \times 10^{-8}$). Analyses in FHS were carried out within the corresponding strata-specific/interaction subgroups as described in GENOA. Second-ary analysis used a gene-based approach that tested multiple SNPs in FHS sibships that were located in associated gene regions (+/-250 kb of GENOA index SNPs with GWAS $p \le 10^{-5}$). We also considered genes containing SNPs with p-values between 10^{-4} and 10^{-5} , and that were near genes (≤ 250 kb) with functions

related to CAC (NCBI MeSH search). For secondary analysis in FHS, we accounted for multiple testing of correlated SNPs within each gene region using LD-based Bonferroni corrections with the number of LD blocks centered around the GENOA discovery SNP (significance threshold of p≤0.05/number of blocks in each gene region) [19]. LD blocks were delimited by SNPs in strong LD (95% with D'≥0.70) according to Haploview (CEU HapMap Build 22) [20].

Results

GENOA study participants were in 421 sibships ranging in size from 1 to 10, with the majority of sibships consisting of size 2 (44%) or size 3 (21%). The general characteristics of the GENOA participants stratified by gender and smoking status are presented in Table 1. Overall, more participants were female (59%) than male (41%), and males had a higher proportion of smokers (58%)compared to females (38%). Participants in all categories had similar ages (mean age of 58.1 years). Males had higher mean CAC scores than females, regardless of smoking status. Smokers had higher mean CAC scores than nonsmokers in both men (341 versus 271) and women (150 versus 84). Table 1 also shows general characteristics for the stratified FHS participants. Overall, FHS participants were older than GENOA participants, with lower use of hypertensive and lipid-lowering medications, and lower frequency of diabetes. Like GENOA, smokers had higher mean CAC scores in both men (468 versus 238) and women (107 versus 81).

Table 2 presents a list of index SNPs and their nearest genes that reached $p \le 10^{-5}$ separately for smokers and nonsmokers in GENOA, and for GxS interactions. Primary analysis also included attempts to replicate our discovery results (index SNPs with $p\le 10^{-5}$) in the FHS cohort. Table 2 shows corresponding p values for GENOA index SNPs in FHS sibships. Overall, only one SNP (*COLEC11* rs12990669 in GENOA smokers, $p = 1.4 \times 10^{-8}$) reached genome-wide significance levels ($p\le 2.3 \times 10^{-8}$) after Bonferroni corrections for multiple testing in GENOA or FHS (Table 2). However, this signal may represent a false positive result as other SNPs in *COLEC11* did not show associations.

We conducted secondary analysis to investigate gene regions containing SNPs that showed suggestive associations in GENOA ($p \le 10^{-5}$) by testing multiple SNPs in FHS sibships within +/-250 kb of GENOA index SNPs [21]. We also included genes containing SNPs with p-values between 10^{-4} and 10^{-5} in GENOA with known functions relevant to CAC (Table 2). Table 3 shows the results for FHS subgroups for SNPs that reached significance thresholds using an LD-based Bonferroni approach to correct for multiple testing ($p \le 0.05$ /number of LD blocks in each gene region), thus avoiding potential overcorrection for correlated SNPs [19].

In GENOA, rs8047995 in intron 8 of the gene for WW domaincontaining oxidoreductase (*WWOX*) showed associations only in smokers ($p = 4.5 \times 10^{-6}$). A different variant in intron 8 of *WWOX* (rs10492908) also showed associations only in smokers in FHS ($p = 1.7 \times 10^{-3}$). Also in GENOA, rs10128255 in the gene for transcription factor 7 like-2 (*TCF7L2*) showed associations only in smokers (Tables 2 and 3, $p = 6.0 \times 10^{-5}$), and another *TCF7L2* SNP (rs11196175) showed associations in FHS smokers (Table 3 $p = 4.0 \times 10^{-4}$). In GENOA nonsmokers, we found associations with rs11569850 ($p = 7.8 \times 10^{-5}$) in the gene for tumor necrosis factor receptor superfamily, member 8 (*TNFRSF8*).

For GxS interactions, rs4410439 in the gene for ADAM metallopeptidase thrombospondin type 1 motif, 9 (*ADAMTS9*) showed associations in GENOA (Table 3, $p = 7.1 \times 10^{-5}$). We also

found associations with ADAMTS9 (rs4688504) in the FHS cohort $(p = 1.0 \times 10^{-4})$. Another variant (rs1560540) that showed associations for GxS interaction in GENOA ($p = 6.9 \times 10^{-5}$) was located in the gene for TBC1 domain family member 4 (TBC1D4). We found stronger associations with TBC1D4 (rs1062087) in the FHS cohort ($p = 4.2 \times 10^{-7}$). Figure 1 shows the stratified effects for genotypes for ADAMST9 (rs4410439) and TBC1D4 (rs1560540) from the GxS interaction results in Table 3. Panel A shows the additive genotype effects (odds ratios obtained by exponentiation of the beta coefficients) for each smoking strata used to calculate the interaction test for rs4410439 in ADAMST9. For smokers $(\beta = 0.59, SE = 0.17, p = 4.5 \times 10^{-4})$, the odds ratio (OR) for reference AA genotypes was set at 1, the OR for AC genotypes was 1.80 (95% confidence interval of 1.47-2.13), and the OR for CC genotypes was 3.23 (0.35–7.40). For nonsmokers ($\beta = -0.43$, SE = 0.21, $p = 3.8 \times 10^{-2}$), the OR for AC genotypes was 0.65 (0.24-1.06) and the OR for CC genotypes was 0.42 (0.06-2.49). Likewise, Panel B shows the additive genotype effects by smoking strata for rs1560540 in *TBC1D4*. For smokers ($\beta = 0.69$, SE = 0.20, $p = 4.7 \times 10^{-4}$), the OR for reference GG genotypes was set at 1, the OR for GA genotypes was 1.99 (1.60-2.38), and the OR for AA genotypes was 3.96 (3.58–4.35). For nonsmokers ($\beta = -0.36$, SE = 0.19, $p = 5.8 \times 10^{-2}$), the OR for GA genotypes was 0.69 (0.32–1.07) and the OR for AA genotypes was 0.48 (0.10–0.86). For both genes, the direction of effects are opposite for the smokers versus nonsmokers (Bs of opposite signs), the hallmark of GxE interaction.

We compared the results of the GxS interaction GWAS with the recent GWAS meta-analysis for main SNP effects on CAC quantity [10]. We interrogated the SNPs with the strongest associations in the GWAS meta-analysis including rs1333049 (*CDKN2B*), rs9349379 and rs2026458 (*PHACTRI*), rs3809346 (*COL4A2*), rs6783981 (*SERPINI*), rs17676451 (*HAL*), rs6604023 (*CDC7*), and rs8001186 (*IRS2*). However, we did not find significant associations with any of these loci for GxS interaction (GENOA discovery cohort) with nominal p-values ranging from 0.09 to 0.49.

Discussion

We used a genome-wide approach to investigate GxS interactions to identify genetic variants associated with CAC exclusively in either smokers or nonsmokers, and with GxS interactions in the GENOA cohort. Our primary analysis used GWAS in GENOA, followed by attempts to replicate associated index SNPs ($p \le 10^{-5}$) in FHS. However, these tests did not yield SNP associations that met genome-wide significance in GENOA, or standard SNPbased replication in FHS (Table 2). Primary analyses were likely limited by the relatively small sample size of the discovery GENOA cohort that reduced statistical power for main effects in GWAS results within strata. In secondary analysis, we used a genebased approach, testing multiple SNPs in FHS within associated gene regions (+/-250 kb of GENOA index SNPs) [19]. We found many instances of SNPs within gene regions that showed significant associations in FHS after correction for multiple testing using numbers of LD blocks in each region (Table 3). Overall, these genes were not represented by identical SNPs in the two cohorts, likely due to differences in allele frequencies or functional differences of SNPs in different regions of the same gene. In some instances, we observed different direction of effects (beta-coefficients) for different SNPs in the same gene (Table 3). Using theoretical modeling, Lin and coworkers demonstrated valid "flipflop" associations may occur even for identical SNPs due to correlations with other causal variants that differ among cohorts

Another and the problem of the										
International Associational Associad Associational Associational	Smokers									
Optimizationality 0100020120120120120120120120120120120120	Locus			Association						Nearby Genes
1 682441 (c) <th>SNP</th> <th>Ъ</th> <th>Position (Mb)</th> <th>Allele (+/-)</th> <th>MAF</th> <th>Beta</th> <th>SE</th> <th>GENOA P value</th> <th>FHS P Value</th> <th>Relative position (-upstream,+downstream)</th>	SNP	Ъ	Position (Mb)	Allele (+/-)	MAF	Beta	SE	GENOA P value	FHS P Value	Relative position (-upstream,+downstream)
(1) <td>rs12990669</td> <td>2</td> <td>3649241</td> <td>(C/G)</td> <td>0.07</td> <td>- 1.79</td> <td>0.32</td> <td>1.36×10^{-8}</td> <td>0.71</td> <td>COLEC11</td>	rs12990669	2	3649241	(C/G)	0.07	- 1.79	0.32	1.36×10^{-8}	0.71	COLEC11
(1) (3) <td>rs2190305</td> <td>7</td> <td>28859450</td> <td>(G/A)</td> <td>0.19</td> <td>-0.98</td> <td>0.20</td> <td>1.24×10^{-6}</td> <td>0.06</td> <td>CREBS</td>	rs2190305	7	28859450	(G/A)	0.19	-0.98	0.20	1.24×10^{-6}	0.06	CREBS
(3) (3) (3) (3) (1) <td>rs10131267</td> <td>14</td> <td>30868736</td> <td>(G/A)</td> <td>0.08</td> <td>- 1.40</td> <td>0.29</td> <td>1.77×10^{-6}</td> <td>0.44</td> <td>intergenic</td>	rs10131267	14	30868736	(G/A)	0.08	- 1.40	0.29	1.77×10^{-6}	0.44	intergenic
(6) (5) <td>rs9574536</td> <td>13</td> <td>80585091</td> <td>(C/T)</td> <td>0.34</td> <td>1.41</td> <td>0.30</td> <td>2.44×10^{-6}</td> <td>0.67</td> <td>intergenic</td>	rs9574536	13	80585091	(C/T)	0.34	1.41	0.30	2.44×10^{-6}	0.67	intergenic
(3) (4) (3973) (1) (3973) (397	rs8047995	16	78949736	(C)(0.32	1.13	0.25	4.51×10^{-6}	0.66	XOMM
(1) <td>rs7158225</td> <td>14</td> <td>84397780</td> <td>(T/C)</td> <td>0.21</td> <td>-0.89</td> <td>0.20</td> <td>5.34×10^{-6}</td> <td>0.69</td> <td>intergenic</td>	rs7158225	14	84397780	(T/C)	0.21	-0.89	0.20	5.34×10^{-6}	0.69	intergenic
(373606) (1) (469/7) (6/1) (0) (0) (0) (0) (0) (0) (0) (37968646) (1) (377160) (10) (3701 (10) (3701 (10) (3701 (10) (37011 (37011 (37011 <td>rs17608293</td> <td>18</td> <td>13378636</td> <td>(A/G)</td> <td>0.38</td> <td>0.83</td> <td>0.18</td> <td>6.01×10^{-6}</td> <td>0.77</td> <td>C180RF1</td>	rs17608293	18	13378636	(A/G)	0.38	0.83	0.18	6.01×10^{-6}	0.77	C180RF1
(5)(5	rs7926081	11	4676977	(d/T)	0.49	-0.83	0.19	8.51×10^{-6}	0.92	intergenic
67111361200716661(10)(10)(11)<	rs7988945	13	51496320	(A/G)	0.39	0.96	0.22	8.94×10^{-6}	0.44	RNASEH2B
\$	rs7411138	-	240716461	(C/A)	0.10	1.34	0.33	5.66×10^{-5}	0.10	GREM2
10114/1335(a)(a)(a)(a)(a)(a)(a)(a)(a)(a)(a)319914000(TG)(TG)(1)(1)(1)(1)(1)(1)(1)(a)1319914000(TG)(1)(1)(1)(1)(1)(1)(1)(1)(a)108193748(a)(1)(1)(1)(1)(1)(1)(1)(1)(a)108193748(a)(1)(1)(1)(1)(1)(1)(1)(1)(a)108193748(1)(1)(1)(1)(1)(1)(1)(1)(1)(a)108193748(1)(1)(1)(1)(1)(1)(1)(1)(1)(a)108193748(1)(1)(1)(1)(1)(1)(1)(1)(1)(a)1111111111(1)(1)(1)(1)(1)(1)(1)(a)11111111111111(1)(1)(1)(1)(1)(1)(a)111111111111111111(1)(1)(1)(1)(1)(1)(a)1111111111111111111111(1)(1)(1)(1)(1)(1)(1)(1)(1)(1)(1	rs7089541	10	68558800	(A/G)	0.19	-0.96	0.24	5.77×10^{-5}	0.54	CTNNA3
(5) (5) (1) <td>rs10128255</td> <td>10</td> <td>114742835</td> <td>(G/A)</td> <td>0.36</td> <td>0.82</td> <td>0.20</td> <td>5.96×10^{-5}</td> <td>0.03</td> <td>TCF7L2</td>	rs10128255	10	114742835	(G/A)	0.36	0.82	0.20	5.96×10^{-5}	0.03	TCF7L2
6723452186073923(Wc)022-0.07800.073800.103	rs16824684	m	154914090	(T/G)	0.19	- 1.00	0.25	6.04×10^{-5}	0.45	<i>MME</i> (12593)
(313) <th< td=""><td>rs7234352</td><td>18</td><td>60739233</td><td>(A/G)</td><td>0.22</td><td>-0.87</td><td>0.22</td><td>8.50×10^{-5}</td><td>0.23</td><td>BCL2 (51346)</td></th<>	rs7234352	18	60739233	(A/G)	0.22	-0.87	0.22	8.50×10^{-5}	0.23	BCL2 (51346)
6378103 10 810127 (TC) 016 023 7.3.×10 ⁻⁵ 0.96 G47.3 15320235 8 25566.5 (GA) 0.05 313 0.79 7.4.×10 ⁻⁵ 0.70 CM01 15370235 8 25566.5 (GA) 0.05 313 0.79 7.4.×10 ⁻⁵ 0.70 CM01 1732054 6 474712 (AC) 0.20 1.21 0.32 2.31×10 ⁻⁷ 0.40 CM01 1.24 1732054 5 3136544 (AC) 0.20 1.21 0.32 2.31×10 ⁻⁷ 0.44 intergenic 1.48 16403330 16 (T) 0.20 1.21 0.23 2.31×10 ⁻⁷ 0.44 intergenic 1.48 1.48 1.48 1.48 1.48 1.48 1.48 1.48 1.48 1.48 1.48 1.44 1.48 1.48 1.44 1.48 1.44 1.48 1.44 1.48 1.44 1.44 1.44 1.44 1.44	rs17819063	16	53873428	(G/A)	0.12	- 1.33	0.32	4.20×10^{-5}	0.70	FTO
(513902) (5) (5) (1) (2) (1) (2) (2) (2) (2) (2) (2) Montrolex A	rs3781093	10	8101927	(T/C)	0.16	-0.91	0.23	7.23×10^{-5}	0.96	GATA3
Monsumera Sup GFIO Mol Mole (µ-1)	rs17390295	8	2958662	(G/A)	0.05	3.13	0.79	7.46×10^{-5}	0.70	CSMD1
NP Ch Destinor (MD) Allel (4-1) NA Er Cen out	Nonsmokers									
rs432695 6 4674712 (A/G) 0.20 121 0.23 231×10 ⁻⁷ 0.54 integenic rs4867326 5 31361704 (CT) 0.25 12.8 0.25 4,74×10 ⁻⁷ 0.61 integenic rs4867326 5 3136544 (CT) 0.25 12.8 0.25 4,94×10 ⁻⁷ 0.60 integenic rs6873705 5 3136544 (A/G) 0.25 1-127 0.25 4,94×10 ⁻⁷ 0.60 intergenic rs6803186 17 76549152 (C/G) 0.13 -1.12 0.20 2,93×10 ⁻⁶ 0.61 intergenic rs11649339 16 7427730 (A/G) 0.13 -1.12 0.30 2,78×10 ⁻⁶ 0.69 intergenic rs1173732 4 2389546 (A/G) 0.13 -1.12 0.30 2,78×10 ⁻⁶ 0.69 intergenic rs1173732 18 2339556 (T/G) 0.13 2,13×10 ⁻⁶ 0.69 0.69 intergenic	SNP	Chr	Position (Mb)	Allele (+/–)	MAF	Beta	SE	GENOA P value	FHS P value	Relative position (-upstream,+downstream)
54867326 5 31361704 (CT) 0.25 1.28 0.25 4.74 × 10 ⁻⁷ 0.61 Integenic 76873705 5 3136554 (MG) 0.26 -1.27 0.25 4.94 × 10 ⁻⁷ 0.60 Integenic 76873705 17 7654915 (CG) 0.26 -1.27 0.25 4.94 × 10 ⁻⁷ 0.60 Integenic 76807186 17 7654915 (CG) 0.42 0.33 2.05 × 10 ⁻⁶ 0.61 Integenic 7611649339 16 74272730 (A/C) 0.13 -1.14 0.30 2.05 × 10 ⁻⁶ 0.69 Integenic 761164933 1 3698640 (GA) 0.13 -1.14 0.30 2.93 × 10 ⁻⁶ 0.69 Integenic 7611737432 1 3898640 (GA) 0.13 -1.14 0.30 2.93 × 10 ⁻⁶ 0.65 Integenic 761173732 13 333756 18 0.16 0.14 0.62 Integenic 71773726 1	rs432695	9	4674712	(A/G)	0.20	1.21	0.23	2.31×10^{-7}	0.54	intergenic
r56373705 5 3136544 (AG) 0.26 -1.27 0.25 4.94 × 10^{-7} 0.60 integenic r8080186 17 75549152 (CG) 0.42 0.33 0.20 2.05 × 10^{-6} 0.61 DMM17 r811649330 16 7427300 (A'C) 0.13 -1.29 0.27 2.42 × 10^{-6} 0.99 intergenic r81164933 1 3698640 (G/A) 0.13 -1.24 0.30 2.05 × 10^{-6} 0.99 intergenic r81173743 1 3698640 (G/A) 0.13 -1.41 0.30 2.33 × 10^{-6} 0.45 0.46 intergenic r81173743 1 35397564 (T/G) 0.10 -1.38 0.30 2.33 × 10^{-6} 0.45 0.45 intergenic r81173743 13 53397564 (T/G) 0.39 -303 2.33 × 10^{-6} 0.45 0.45 intergenic r82053734 18 4228493 (G/A) 0.09 1.41 0.30	rs4867326	5	31361704	(C/T)	0.25	1.28	0.25	4.74×10^{-7}	0.61	intergenic
rs8808186 17 76549152 (C/G) 0.42 0.33 0.20 205×10 ⁻⁶ 0.61 D/MH17 rs11649339 16 7427730 (A/C) 013 -1.29 0.27 2.42×10 ⁻⁶ 0.99 <i>integenic</i> rs4462101 1 3698640 (G/A) 0.13 -1.141 0.30 2.78×10 ⁻⁶ 0.99 <i>integenic</i> rs11737432 4 2876776 (G/A) 0.10 -1.38 0.30 2.38×10 ⁻⁶ 0.99 <i>integenic</i> rs11737432 4 2876776 (G/A) 0.10 -1.38 0.30 2.38×10 ⁻⁶ 0.45 <i>integenic</i> rs117373726 18 288557 (G/A) 0.39 -0.96 0.21 3.81×10 ⁻⁶ 0.65 <i>integenic</i> rs17735726 18 4828657 (G/A) 0.09 0.31 0.31 0.32 0.32×10 ⁻⁶ 0.65 <i>integenic</i> rs17735726 18 4828657 (G/A) 0.09 0.710 ⁻⁶ 0.65 0.66	rs6873705	5	31366544	(A/G)	0.26	-1.27	0.25	4.94×10^{-7}	0.60	intergenic
I164339 I6 7427230 (AC) 0.13 -1.29 0.27 2.42×10 ⁻⁶ 0.99 intergenic I4462101 1 3689640 (G/A) 0.13 -1.41 0.30 2.78×10 ⁻⁶ 0.99 intergenic I5173742 4 2876776 (G/A) 0.13 -1.41 0.30 2.78×10 ⁻⁶ 0.99 intergenic I5173742 13 5339754 (G/A) 0.10 -1.38 0.30 2.93×10 ⁻⁶ 0.45 intergenic I51735726 18 88557 (G/A) 0.39 -0.96 0.21 381×10 ⁻⁶ 0.65 intergenic I51735726 18 4828557 (G/A) 0.39 -0.96 0.61 0.66 intergenic I51735726 18 48286557 (G/A) 0.09 0.13 0.30 6.61×10 ⁻⁶ 0.90 intergenic I51735726 18 4288657 (G/A) 0.09 0.13 0.21×10 ⁻⁶ 0.90 0.81 0.81 0.81	rs8080186	17	76549152	(C/G)	0.42	0.93	0.20	2.05×10^{-6}	0.61	DNAH17
F4462101 1 3698640 (G/A) 0.13 -1.41 0.30 2.78×10 ⁻⁶ 0.99 CF3R (-40761) f31173742 4 2876776 (G/A) 0.10 -1.38 0.30 2.38×10 ⁻⁶ 0.99 CF3R (-40761) f3173742 4 2837574 (G/A) 0.10 -1.38 0.30 2.93×10 ⁻⁶ 0.45 intergenic f502348 13 53397564 (T/G) 0.39 -0.96 0.21 3.81×10 ⁻⁶ 0.65 intergenic f51737576 18 4828557 (G/A) 0.09 -1.37 0.30 6.61×10 ⁻⁶ 0.90 MAPK4 (28361) f51735726 18 4828557 (G/A) 0.09 -1.37 0.30 6.61×10 ⁻⁶ 0.90 MAPK4 (28361) f51872734 4 2248933 (G/A) 0.09 -1.43 0.32 7.25×10 ⁻⁶ 0.85 GPR1 25 f51345251 7 103548958 (T/A) 0.09 0.14 0.25×10 ⁻⁶ 0.20 0.20	rs11649339	16	74272730	(A/C)	0.13	-1.29	0.27	2.42×10^{-6}	0.99	intergenic
Iritification Image: Construction Const	rs4462101	-	36989640	(G/A)	0.13	- 1.41	0.30	2.78×10^{-6}	0.99	CSF3R (-40761)
r5622348 13 53397564 (T/G) 0.39 -0.06 0.21 381×10^{-6} 0.62 intergenic r517735726 18 4286557 (G/A) 0.09 -1.37 0.30 6.61 \times 10^{-6} 0.90 MAP4 (28361) r51687234 4 22484933 (G/A) 0.05 -1.91 0.42 6.97 \times 10^{-6} 0.85 GPR125 r5134521 7 22248527 (G/T) 0.09 -1.43 0.32 7.25 \times 10^{-6} 0.85 GPR125 r5465945 7 103548958 (T/A) 0.19 0.143 0.32 7.25 \times 10^{-6} 0.20 RAPGEF5 r540608 1 2314069 0.24 2.62 \times 10^{-6} 0.73 0.20 RAPGEF5	rs11737432	4	28767776	(G/A)	0.10	-1.38	0.30	2.93×10^{-6}	0.45	intergenic
If 17735726 18 48286557 (G/A) 0.09 -1.37 0.30 6.61 × 10 ⁻⁶ 0.90 MAPK4 (28361) If 16872734 4 22484933 (G/A) 0.05 -1.91 0.42 6.97 × 10 ⁻⁶ 0.85 GPR125 If 345251 7 22248527 (G/T) 0.09 -1.43 0.32 7.25 × 10 ⁻⁶ 0.20 RAPGEF5 Is 4656945 7 103548958 (T/A) 0.15 1.65 0.37 9.21 × 10 ⁻⁶ 0.73 RAPGEF5 Is 4306080 1 25135004 (C/T) 0.27 -1.00 0.24 2.62 × 10 ⁻⁵ 0.93 CIIC4	rs622348	13	53397564	(T/G)	0.39	-0.96	0.21	3.81×10^{-6}	0.62	intergenic
rs16872734 4 22484933 (G/A) 0.05 -1.91 0.42 6.97×10 ⁻⁶ 0.85 GPR125 rs1345251 7 22248527 (G/T) 0.09 -1.43 0.32 7.25×10 ⁻⁶ 0.20 RAPGEF5 rs6465945 7 103548958 (T/A) 0.15 1.65 0.37 9.21×10 ⁻⁶ 0.73 RELN rs4306080 1 25135004 (C/T) 0.27 -1.00 0.24 2.62×10 ⁻⁵ 0.39 CLC4	rs17735726	18	48286557	(G/A)	0.09	-1.37	0.30	6.61×10^{-6}	06.0	MAPK4 (28361)
rs134521 7 22248527 (G/T) 0.09 -1.43 0.32 7.25×10 ⁻⁶ 0.20 RAPGEF5 rs6465945 7 103548958 (T/A) 0.15 1.65 0.37 9.21×10 ⁻⁶ 0.73 RELN rs4306080 1 25135004 (C/T) 0.27 -1.00 0.24 2.62×10 ⁻⁵ 0.98 CL/C4	rs16872734	4	22484933	(G/A)	0.05	- 1.91	0.42	6.97×10^{-6}	0.85	GPR1 25
rs6465945 7 103548958 (T/A) 0.15 1.65 0.37 9.21×10 ⁻⁶ 0.73 RELN rs4306080 1 25135004 (C/T) 0.27 -1.00 0.24 2.62×10 ⁻⁵ 0.98 CLIC4	rs1345251	7	22248527	(G/T)	0.09	- 1.43	0.32	7.25×10^{-6}	0.20	RAPGEF5
rs4306080 1 25135004 (C/T) 0.27 -1.00 0.24 2.62×10 ⁻⁵ 0.98 CLIC4	rs6465945	7	103548958	(T/A)	0.15	1.65	0.37	9.21×10^{-6}	0.73	RELN
	rs4306080	-	25135004	(C/T)	0.27	- 1.00	0.24	2.62×10^{-5}	0.98	CLIC4

ionsmokers									
ЧN	Chr	Position (Mb)	Allele (+/-)	MAF	Beta	SE	GENOA P value	FHS P value	Relative position (-upstream,+downstream)
9521695	13	110981007	(C/G)	0.49	- 1.11	0.27	4.75×10^{-5}	0.10	C0L4A2
2727551	7	151391653	(A/G)	0.07	1.86	0.46	5.16×10^{-5}	0.09	PRKAG2
2784773	10	81919800	(T/C)	0.34	-0.91	0.23	6.84×10^{-5}	0.57	ANXA11
12602288	17	68573878	(C/A)	0.49	-0.75	0.19	6.91×10^{-5}	0.26	intergenic
11790127	6	18624300	(C/T)	0.10	1.38	0.35	7.95×10^{-5}	0.45	ADAMTSL1
11569850	-	12164391	(G/C)	0.09	-1.36	0.35	7.83×10^{-5}	0.97	TNFRSF8
5703976	-	21836934	(C/T)	0.11	1.24	0.31	5.86×10^{-5}	0.88	ALPL
893009	17	64708358	(C/G)	0.46	0.81	0.21	8.59×10^{-5}	0.37	PRKCA
ene by Smokii	ng Interactic	suc							
٩	Chr	Position (Mb)	Allele (+/–)	MAF	Beta	SE	GENOA P value	FHS P value	Relative position (-upstream,+downstream)
6872734	4	22484933	(G/A)	0.05	3.62	0.73	3.95×10^{-7}	0.24	GPR125
395540	12	61446746	(T/C)	0.26	1.44	0.30	9.27×10^{-7}	0.18	intergenic
99877	8	130490213	(G/T)	0.32	-1.36	0.29	1.63×10^{-6}	0.34	intergenic
0096362	8	95083587	(G/T)	0.14	- 1.83	0.40	2.21×10^{-6}	0.49	intergenic
843942	£	30729636	(G/A)	0.37	-1.24	0.27	3.00×10^{-6}	0.13	TGFBR2
6951358	15	68153732	(G/C)	0.25	- 1.43	0.32	4.00×10^{-6}	0.02	intergenic
776377	15	68103387	(C/T)	0.29	1.32	0.30	4.39×10^{-6}	0.26	MAP2K5
144284	14	44433152	(T/C)	0.07	2.47	0.57	7.54×10^{-6}	0.48	intergenic
574536	13	80585091	(C/T)	0.34	1.81	0.42	9.44×10^{-6}	0.30	intergenic
462101	-	36989640	(G/A)	0.13	1.78	0.43	1.83×10^{-5}	0.24	CSF3R (-40761)
679195	e	187880447	(C/T)	0.39	-1.13	0.28	3.43×10^{-5}	0.08	Грр
410439	e	64655724	(A/C)	0.63	1.02	0.27	7.05×10^{-5}	0.28	ADAMTS9
560540	13	75882495	(G/A)	0.52	-1.05	0.28	6.86×10^{-5}	0.14	TBC1D4
618157	15	39844315	(G/A)	0.09	-2.22	0.57	5.31×10^{-5}	0.18	THBS1 (-28812)
008465	8	1722760	(T/G)	0.15	-1.52	0.38	3.36×10^{-5}	0.40	CLN8
565497	17	78912633	(A/T)	0.57	-1.13	0.27	1.17×10^{-5}	0.29	RPTOR
867326	5	31361704	(T/C)	0.75	1.32	0.32	2.31×10^{-5}	0.46	CDH6 (32451)
032769	20	44938851	(C/J)	0.22	1.52	0.36	1.26×10^{-5}	0.16	CDH22 (-1714)
0514838	6	123746796	(T/C)	0.10	- 1.83	0.48	6.40×10^{-5}	0.14	CS

cohort.
discovery
e GENOA
CAC in th
ited with
Ps associa
aining SN
enes cont
=HS for g
cation in l
ised replic
of gene-ba
: results c
y analysis
Secondar
ble 3.

				-					
SNP Chr	Position (Mb)	Allele (+/–)	MAF	Beta	SE	P value	# of blocks (threshold)	Relative position (-upstream,+downstream)	Cohort
rs8047995 16	78949736	(G/C)	0.32	1.13	0.25	4.51×10^{-6}		Хомм	GENOA
rs10492908 16	79006863	(A/G)	0.05	1.4	0.45	1.70×10^{-3}	50 (1.0×10 ⁻³)	ХОММ	FHS
rs10128255 10	114742835	(G/A)	0.36	0.82	0.20	6.0×10^{-5}		TCF7L2	GENOA
rs11196175 10	114736614	(C/T)	0.33	0.72	0.21	4.0×10^{-4}	30 (1.7×10 ⁻³)	TCF7L2	FHS
Nonsmokers									
SNP Chr	Position (Mb)	Allele (+/–)	MAF	Beta	SE	P value	# of blocks (threshold)	Relative position (-upstream,+downstream)	Cohort
rs2727551 7	151391653	(A/G)	0.07	1.86	0.46	5.2×10^{-5}		PRKAG2	GENOA
rs2727527 7	151347462	(A/C)	0.3	-0.67	0.20	9.0×10^{-4}	27 (1.9×10 ⁻³)	PRKAG2	FHS
rs11569850 1	12164391	(C/C)	60.0	-1.36	0.35	7.8×10^{-5}		TNFRSF8	GENOA
rs4304595 1	11946389	(A/C)	0.31	-0.78	0.21	2.0×10^{-4}	21 (2.4×10 ⁻³)	PLOD1 (upstream)	FHS
Gene by Smoking Intera	ction								
SNP Chr	Position (Mb)	Allele (+/-)	MAF	Beta	SE	P value	# of blocks (threshold)	Relative position (-upstream,+downstream)	Cohort
rs4462101 1	36989640	(G/A)	0.13	1.78	0.43	1.8×10^{-5}		CSF3R (-40761)	GENOA
rs7528341 1	37141174	(C/T)	0.21	0.95	0.29	4.0×10^{-4}	30 (1.7×10 ⁻³)	FTLP18 (44716)	FHS
rs1679195 3	187880447	(C/T)	0.39	-1.13	0.28	3.4×10^{-5}		ГРР	GENOA
rs9871228 3	187795325	(G/T)	0.23	-1.36	0.42	6.2×10^{-4}	38 (1.3×10 ⁻³)	LPP (-75747)	FHS
rs4410439 3	64655724	(A/C)	0.37	1.02	0.27	7.1×10^{-5}		ADAMTS9	GENOA
rs4688504 3	64684941	(C/T)	0.47	0.78	0.21	1.0×10^{-4}	30 (1.7×10 ⁻³)	ADAMTS9	FHS
rs1560540 13	75882495	(G/A)	0.48	1.05	0.28	6.9×10^{-5}		TBC1D4	GENOA
rs1062087 13	75884216	(C/T)	0.12	1.91	0.38	4.2×10^{-7}	40 (1.3×10 ⁻³)	TBC1D4	FHS
rs7008465 8	1722760	(1/C)	0.15	-1.52	0.38	3.4×10^{-5}		CLN8	GENOA
rs3758014 8	1849275	(A/G)	0.28	-1.97	0.52	7.8×10^{-5}	45 (1.1×10^{-3})	ARHGEF10	FHS

Panel A

Panel B



Figure 1. GxS interaction effects stratified by smoking status for ADAMST9 (rs4410439) (Panel A) and TBC1D4 (rs1560540) (Panel B) genotypes. The Figure shows the additive genotype effects (odds ratios) for each smoking strata used to calculate interaction tests (blue bars for nonsmokers and red bars for smokers). The odds ratios on the y-axis are plotted on the log scale with error bars for 95% confidence intervals, and the genotypes are shown on the x-axis. doi:10.1371/journal.pone.0074642.g001

because of interaction effects or differences in LD patterns caused by sampling variation within ethnic groups or evolutionary history between ethnic groups [22]. In general, replication in GxE GWAS may prove challenging, given the complex nature of traits like CAC that are influenced by numerous interactions among multiple loci and environmental factors, as well as differences among cohorts in LD structure and environmental exposures.

Our GWAS studies of GxS interactions identified several genes that showed concordance of results in GENOA and FHS that are involved in diverse cellular processes such as inflammation and osteogenesis that are relevant to CAC. In particular, Inflammation is a likely mediator of GxS interactions, since many of the deleterious effects of smoking are due to induction of inflammatory responses, contributing to chronic diseases such as CHD [23]. Inflammatory markers are also well known risk factors for type 2 diabetes (T2D), providing a likely physiological connection between development of CHD and T2D [24]. Recent in vitro experiments in human umbilical vein endothelial cells demonstrated that nicotine stimulates cellular inflammatory response via activation of the NF- κ B transcription factor axis by a second messenger pathway [25]. In a rat model, exposure to cigarette smoke caused changes in levels of inflammatory markers including NF- κ B in cardiac tissues [26]. In addition to inflammation, the NF-KB axis plays a central role in CAC quantity and bone remodeling by induction of osteoclast differentiation [27,28].

In GENOA and FHS, the gene for transcription factor 7 like-2 (*TCF7L2*) showed associations only in smokers (Table 3). *TCF7L2* (chromosome 10) has shown associations with T2D in a previous GWAS [29], and may impair pancreatic beta-cell function with effects on blood glucose homeostasis [30]. *TCF7L2* has shown associations with angiographically determined CHD in diabetic and non-diabetic patients [31], as well as with CVD, ischemic stroke, peripheral artery disease, and all-cause mortality [32]. *TCF7L2* encodes the Tcf-4 transcription factor in the Wnt

Signalling pathway, directly regulating beta-catenin, a major activator of the NF- κ B axis [33]. Activation of the NF- κ B axis may provide a clue concerning the role of *TCF7L2* in CAC development, since osteogenesis and bone remodeling are regulated by NF- κ B [27,28].

The gene for WW domain-containing oxidoreductase (*WWOX*) also showed associations only in smokers from both GENOA and FHS (Table 3). *WWOX* (chromosome16) is an established tumor suppressor gene that is associated with CHD, bone development, and higher methylation levels in smokers [34]. In previous population-based studies, variants in *WWOX* have shown associations with CHD and left ventricular mass [35,36]. Ablation of *WWOX* in knock-out mouse strains (*Wwox -/-*) caused development of osteosarcomas, as well as osteopenia and bone growth retardation [34,37].

In GENOA nonsmokers, we found associations in the gene for tumor necrosis factor receptor superfamily, member 8 (*TNFRSF8*) (Table 3). Like *TCF7L2*, this gene may also influence CAC via the NF- κ B axis that regulates inflammatory response and bone remodeling [27,28]. *TNFRSF8* is a member of the TNF-receptor superfamily that play key roles in signaling pathways that regulate NF- κ B activation via interaction with TNF cytokines. In FHS, the associated variant in this chromosomal region was in an intergenic region near *PLOD1* (rs4304595, p = 2.0 × 10⁻⁴) (Table 3).

For GxS interactions, rs1062087 within the gene for TBC1 domain family member 4 (*TBC1D4*) showed stronger association in FHS than the SNP identified in GENOA (Table 3). Interestingly, rs1062087 is a nonsynonymous variant (Ile818Val) that is located in the same LD block with rs1560540 that showed associations in GENOA. *TBC1D4* encodes the AS160 protein that mediates insulin homeostasis by regulating glucose uptake in fat and muscle cells via GLUT4 glucose transporters. Inflammatory markers (TNF- α , IL-1, IL-6) are associated with a reduction of AS160 activities, resulting in increased insulin resistance [38].

Another variant (rs4410439) that showed associations for GxS interactions in GENOA (P = 7.1×10^{-5}) was located in the gene for ADAM metallopeptidase thrombospondin type 1 motif, 9 (*ADAMTS9*) (Table 3). We also found associations with *ADAMTS9* (rs4688504) in the FHS cohort (p = 1.0×10^{-4}) (Table 3). *ADAMTS9* encodes a metalloprotease that is involved in thrombosis, cleaving veriscan, proteoglycans, and aggrecan. In transgenic mice that were heterozygous for an inactivated allele carrying the *LacZ* reporter gene (*Adamts9*^{+LacZ}), *ADAMTS9* haploinsufficiency altered cardiovascular development and allostasis, resulting in valvular and aortic anomalies [39]. As with *TCF7L2*, both *TBC1D4* and *ADAMTS9* have shown associations in GWAS meta-analysis for T2D [29,40]. Such genes may provide new insights into the well-known relationship between CHD and T2D, perhaps mediated by inflammatory processes.

Perhaps the future best use of existing GWAS data from epidemiological cohorts is the identification of loci involved in interactions (gene by gene, gene by environment) that underlie complex diseases such as CHD and their risk factors. Our results demonstrate that such interactions (i.e., gene by smoking) may be generalizable among cohorts, given that many of the genes identified in GENOA also showed significant associations in FHS. These interactions are likely to reflect the role of particular metabolic or physiological pathways that include many genes, and that interact with environmental factors such as smoking. In our

References

- Sing CF, Stengard JH, Kardia SL (2003) Genes, environment, and cardiovascular disease. Arterioscler Thromb Vasc Biol 23: 1190–1196.
- Peyser PA, Bielak LF, Chu JS, Turner ST, Ellsworth DL, et al. (2002) Heritability of coronary artery calcium quantity measured by electron beam computed tomography in asymptomatic adults. Circulation 106: 304–308.
- Turner ST, Peyser PA, Kardia SL, Bielak LF, Sheedy PF III, et al. (2006) Genomic loci with pleiotropic effects on coronary artery calcification. Atherosclerosis 185:340–346.
- Shaw LJ, Raggi P, Schisterman E, Berman DS, Callister TQ (2003) Prognostic value of cardiac risk factors and coronary artery calcium screening for all-cause mortality. Radiology 228: 826–833.
- Arad Y, Spadaro LA, Goodman K, Newstein D, Guerci AD (2000) Prediction of coronary events with electron beam computed tomography. J Am Coll Cardiol 36: 1253–1260.
- O'Malley PG, Taylor AJ, Jackson JL, Doherty TM, Detrano RC (2000) Prognostic value of coronary electron-beam computed tomography for coronary heart disease events in asymptomatic populations. Am J Cardiol 85: 945–948.
- Rumberger JA (2010) Using noncontrast cardiac CT and coronary artery calcification measurements for cardiovascular risk assessment and management in asymptomatic adults. Vasc Health Risk Manag 6: 579–591.
- Arad Y, Goodman KJ, Roth M, Newstein D, Guerci AD (2005) Coronary calcification, coronary disease risk factors, C-reactive protein, and atherosclerotic cardiovascular disease events: the St. Francis Heart Study. J Am Coll Cardiol 46: 158–165.
- Detrano R, Guerci AD, Carr JJ, Bild DE, Burke G, et al. (2008) Coronary calcium as a predictor of coronary events in four racial or ethnic groups. N Engl J Med 358: 1336–1345.
- O'Donnell CJ, Kavousi M, Smith AV, Kardia SL, Feitosa MF, et al. (2011) Genome-wide association study for coronary artery calcification with follow-up in myocardial infarction. Circulation 124: 2855–2864.
- Hoff JA, Daviglus ML, Chomka EV, Krainik AJ, Sevrukov A, et al. (2003) Conventional coronary artery disease risk factors and coronary artery calcium detected by electron beam tomography in 30,908 healthy individuals. Ann Epidemiol 13: 163–169.
- North KE, Carr JJ, Borecki IB, Kraja A, Province M, et al. (2007) QTL-specific genotype-by-smoking interaction and burden of calcified coronary atherosclerosis: the NHLBI Family Heart Study. Atherosclerosis 193: 11–19.
- Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR (2010) MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. Genet Epidemiol 34: 816–834.
- Li Y, Willer C, Sanna S, Abccasis G (2009) Genotype imputation. Annu Rev Genomics Hum Genet 10: 387–406.
- Botman SL (2000) Design and estimation for the National Health Interview Survey, 1995–2004. Vital Health Stat 2 130: 1–31.
- Chen MH, Yang Q (2010) GWAF: an R package for genome-wide association analyses with family data. Bioinformatics 26: 580–581.

study, we found that many of the replicated genes were involved in inflammatory pathways mediated by the NF- κ B axis. In addition, three of the loci associated with CAC also showed associations in GWAS meta-analysis of T2D [40], a chronic disease with altered inflammatory pathways and increased CHD risk. Since smoking may cause chronic and systemic inflammation, association of these genes in GENOA likely reflect their roles in CAC development and progression via participation in inflammatory pathways. Interestingly, the NF- κ B axis also regulates bone remodeling, providing a link between inflammation and pathways of osteogenesis involved in development and progression of CAC. Additional genetic studies will be required for further tests of these genes in other human populations, as well as functional studies to understand how these genes influence gene by smoking interactions.

Acknowledgments

We thank the participants and staff of the GENOA and FHS studies for their important contributions.

Author Contributions

Conceived and designed the experiments: LMP ACM PAP SLRK. Performed the experiments: LMP JAS. Analyzed the data: LMP JAS LCS ACM. Wrote the paper: LMP JEH PAP LCS LFB JAS.

- Price AL, Patterson NJ, Plenge RM, Weinblatt ME, Shadick NA, et al. (2006) Principal components analysis corrects for stratification in genome-wide association studies. Nat Genet 38: 904–909.
- Heid IM, Jackson AU, Randall JC, Winkler TW, Qi L, et al (2010) Metaanalysis identifies 13 new loci associated with waist-hip ratio and reveals sexual dimorphism in the genetic basis of fat distribution. Nat Genet 42: 949–960.
- Johnson RC, Nelson GW, Troyer JL, Lautenberger JA, Kessing BD, et al. (2010) Accounting for multiple comparisons in a genome-wide association study (GWAS). BMC Genomics 11: 724.
- Gabriel SB, Schaffner SF, Nguyen H, Moore JM, Roy J, et al. (2002) The structure of haplotype blocks in the human genome. Science 296: 2225–2229.
- Ramos E, Chen G, Shriner D, Doumatey A, Gerry NP, et al. (2011) Replication of genome-wide association studies (GWAS) loci for fasting plasma glucose in African-Americans. Diabetologia 54: 783–788.
- Lin PI, Vance JM, Pericak-Vance MA, Martin ER (2007) No gene is an island: the flip-flop phenomenon. Am J Hum Genet 80: 531–538.
- Roivainen M, Viik-Kajander M, Palosuo T, Toivanen P, Leinonen M, et al. (2000) Infections, inflammation, and the risk of coronary heart disease. Circulation 101: 252–257.
- Hu FB, Meigs JB, Li TY, Rifai N, Manson JE (2004) Inflammatory markers and risk of developing type 2 diabetes in women. Diabetes 53: 693–700.
- Ueno H, Pradhan S, Schlessel D, Hirasawa H, Sumpio BE (2006) Nicotine enhances human vascular endothelial cell expression of ICAM-1 and VCAM-1 via protein kinase C, p38 mitogen-activated protein kinase, NF-κB, and AP-1. Cardiovasc Toxicol 6: 39–50.
- Gokulakrisnan A, Jayachandran DB, Thirunavukkarasu C (2011) Attenuation of the cardiac inflammatory changes and lipid anomalies by (-)-epigallocatechingallate in cigarette smoke-exposed rats. Mol Cell Biochem 354: 1–10.
- Venuraju SM, Yerramasu A, Corder R, Lahiri A (2010) Osteoprotegerin as a predictor of coronary artery disease and cardiovascular mortality and morbidity. J Am Coll Cardiol 55: 2049–2061.
- Baud'huin M, Lamoureux F, Duplomb L, Redini F, Heymann D (2007) RANKL, RANK, osteoprotegerin: key partners of osteoimmunology and vascular diseases. Cell Mol Life Sci 64: 2334–2350.
- Strawbridge RJ, Dupuis J, Prokopenko I, Barker A, Ahlqvist E, et al. (2011) Genome-wide association identifies nine common variants associated with fasting proinsulin levels and provides new insights into the pathophysiology of type 2 diabetes. Diabetes 60: 2624–2634.
- Loos RJ, Franks PW, Francis RW, Barroso I, Gribble FM, et al. (2007) TCF7L2 polymorphisms modulate proinsulin levels and beta-cell function in a British Europid population. Diabetes 56: 1943–1947.
- Muendlein A, Saely CH, Geller-Rhomberg S, Sonderegger G, Rein P, et al. (2011) Single nucleotide polymorphisms of TCF7L2 are linked to diabetic coronary atherosclerosis. PLoS One 6: e17978.
- Bielinski SJ, Pankow JS, Folsom AR, North KE, Boerwinkle E (2008) TCF7L2 single nucleotide polymorphisms, cardiovascular disease and all-cause mortality: the Atherosclerosis Risk in Communities (ARIC) study. Diabetologia 51: 968– 970.

- Wang X, Xiao Y, Mou Y, Zhao Y, Blankesteijn WM, et al. (2002) A role for the beta-catenin/T-cell factor signaling cascade in vascular remodeling. Circ Res 90: 340–347.
- Aqeilan RI, Croce CM (2007) WWOX in biological control and tumorigenesis. J Cell Physiol 212: 307–310.
- Saez ME, Gonzalez-Perez A, Martinez-Larrad MT, Gayan J, Real LM, et al. (2010) WWOX gene is associated with HDL cholesterol and triglyceride levels. BMC Med Genet 11: 148.
- Vasan RS, Glazer NL, Felix JF, Lieb W, Wild PS, et al. (2009) Genetic variants associated with cardiac structure and function: a metaanalysis and replication of genome-wide association data. JAMA 302: 168–178.
- Del Mare S, Kurek KC, Stein GS, Lian JB, Aqeilan RI (2011) Role of the WWOX tumor suppressor gene in bone homeostasis and the pathogenesis of osteosarcoma. Am J Cancer Res 1: 585–594.
- Levinger I, Levinger P, Trenerry MK, Feller JA, Bartlett JR, et al. (2011) Increased inflammatory cytokine expression in the vastus lateralis of patients with knee osteoarthritis. Arthritis Rheum 63: 1343–1348.
- Kern CB, Wessels A, McGarity J, Dixon LJ, Alston E, et al. (2010) Reduced versican cleavage due to Adamts9 haploinsufficiency is associated with cardiac and aortic anomalies. Matrix Biol 29: 304–316.
- Zeggini E, Scott LJ, Saxena R, Voight BF, Marchini JL, et al. (2008) Metaanalysis of genome-wide association data and large-scale replication identifies additional susceptibility loci for type 2 diabetes. Nat Genet 40: 638–645.