

Designing Inhibitors of M2 Proton Channel against H1N1 Swine Influenza Virus

Qi-Shi Du^{1,2,4*}, Ri-Bo Huang^{1,2}, Shu-Qing Wang³, Kuo-Chen Chou⁴

1 Guangxi Academy of Sciences, Nanning, Guangxi, China, **2** College of Life Science and Technique, Guangxi University, Nanning, Guangxi, China, **3** School of Pharmaceutical Sciences, Tianjin Medical University, Tianjin, China, **4** Gordon Life Science Institute, San Diego, California, United States of America

Abstract

Background: M2 proton channel of H1N1 influenza A virus is the target protein of anti-flu drugs amantadine and rimantadine. However, the two once powerful adamantane-based drugs lost their 90% bioactivity because of mutations of virus in recent twenty years. The NMR structure of the M2 channel protein determined by Schnell and Chou (Nature, 2008, 451, 591–595) may help people to solve the drug-resistant problem and develop more powerful new drugs against H1N1 influenza virus.

Methodology: Docking calculation is performed to build the complex structure between receptor M2 proton channel and ligands, including existing drugs amantadine and rimantadine, and two newly designed inhibitors. The computer-aided drug design methods are used to calculate the binding free energies, with the computational biology techniques to analyze the interactions between M2 proton channel and adamantane-based inhibitors.

Conclusions: 1) The NMR structure of M2 proton channel provides a reliable structural basis for rational drug design against influenza virus. 2) The channel gating mechanism and the inhibiting mechanism of M2 proton channel, revealed by the NMR structure of M2 proton channel, provides the new ideas for channel inhibitor design. 3) The newly designed adamantane-based inhibitors based on the modeled structure of H1N1-M2 proton channel have two pharmacophore groups, which act like a “barrel hoop”, holding two adjacent helices of the H1N1-M2 tetramer through the two pharmacophore groups outside the channel. 4) The inhibitors with such binding mechanism may overcome the drug resistance problem of influenza A virus to the adamantane-based drugs.

Citation: Du Q-S, Huang R-B, Wang S-Q, Chou K-C (2010) Designing Inhibitors of M2 Proton Channel against H1N1 Swine Influenza Virus. PLoS ONE 5(2): e9388. doi:10.1371/journal.pone.0009388

Editor: Anil Kumar Tyagi, University of Delhi, India

Received: October 20, 2009; **Accepted:** January 12, 2010; **Published:** February 23, 2010

Copyright: © 2010 Du et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This work is financially supported by the National High-tech Research and Development Program (‘863’) of China under the project 2006AA020103, and by the Chinese National Science Foundation (NSFC) under the project 30970562. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: qishi_du@yahoo.com.cn

Introduction

Recently, the outbreak of H1N1 influenza A virus is a pandemic of a new strain of influenza virus [1] identified in April 2009, commonly referred to as “swine flu”. Within only four months, the pandemic has caused many deaths from the first detected country Mexico to almost all countries of the world (<http://www.who.int/csr/disease/swineflu/>). The H1N1 influenza virus is quite familiar to us because it had caused the 1918–1919 Spain pandemic that had infected 5% of the world population and resulted in 20–50 million deaths worldwide [1]. In July 2009 the WHO (World Health Organization) enhanced the warning to phase 6, meaning that the spread of H1N1 influenza virus has become a serious global pandemic. It was anticipated that a stronger outbreak might occur in the coming winter. The even worse news is that cases were reported that several strains of H1N1 influenza A viruses were resistant to oseltamivir (Tamiflu).

Although an influenza virus only possesses eight genes (far less than the estimated 25,000 that a human being has), its simplicity has not stopped it from wreaking havoc on human beings for centuries. “The only thing predictable about influenza is its unpredictability” [2]. Influenza A virus has the ability to undergo

changes by the mechanisms of antigenic drift and shift, resulting in new evolving virus strains, which may be extremely toxic and drug-resistant [3–5]. Given that influenza shifts may occur every 20–30 years, the danger of future influenza A pandemics highlights the need to develop more effective drugs. The threat of an impending influenza pandemic, possibly through the mutations of the present avian strain H5N1 or swine strain H1N1, has triggered a global effort to develop more effective antiviral drugs. However, during the past several decades many efforts in developing anti influenza drugs have almost been futile due to the rapid mutations of the influenza virus, resulting in the persistent resistance to the existing drugs.

The M2 protein [6–9] from influenza A virus is a pH-sensitive proton channel that mediates acidification of the interior of viral particles entrapped and replication in endosomes [10]. Since the M2 protein was found, it has been the main target for finding drugs against influenza A virus. The adamantane-based drugs, amantadine and rimantadine [11–13], which target the M2 channel, had been used for many years as the first-choice antiviral drugs against community outbreaks of influenza A viruses. However, the once powerful drugs lost their effectivity quickly due to mutations and evolutions of influenza A viruses. Recent

reports show that the resistance of influenza A virus to the adamantane-based drugs in humans, birds and pigs has reached more than 90% [3,4].

To solve the drug-resistance problem, a reliable molecular structure of M2 proton channel is absolutely necessary [14,15]. Very recently, using high-resolution nuclear magnetic resonance (NMR) spectroscopy, Schnell and Chou [16] for the first time successfully determined the solution structure of M2 proton channel. They reported an unexpected mechanism of its inhibition by the flu-fighting adamantane drug family. According to the novel mechanism, rimantadine binds at four equivalent sites near the “tryptophan gate” on the lipid-facing side of the channel and stabilizes the closed conformation of the pore. This is completely different from the traditional view but more reasonable in the sense of energetics [17–19].

The new discovery of M2 proton channel structure has brought us the light, by which the drug-resistance problem may be solved, and more powerful adamantane-based drugs may be developed. This is because if we can understand how the drug blocks the channel and how mutations evade the effect of the drug, we can come up with better approaches to block it [20].

Based on such a rationale as well as the high-resolution NMR structure of M2 proton channel [16], the present study was initiated in an attempt to solve the drug resistant problem and to design more effective adamantane-based drugs by conducting molecular modeling and docking studies.

Materials and Methods

Until September 4, 2009, a total of 48 amino acid sequences of H1N1-M2 proton channel proteins are deposited in the website NCBI (www.ncbi.nlm.nih.gov). However, no experimental 3D

(three-dimensional) structure of H1N1-M2 protein is reported so far. To develop its 3D structure, the H1N1-M2 protein sequence with the NCBI code of GQ385303 (www.ncbi.nlm.nih.gov) is used in this study. The sequence is isolated in Toronto from a H1N1 virus strain in July 2009. To build the three dimensional structure of GQ385303, the high-resolution NMR structure of M2 proton channel [16] with the PDB code of 2RLF (www.rcsb.org/pdb) was adopted as a template, which was determined for the M2 channel isolated from the Udorn strain of human influenza virus.

The sequence alignment is performed between the targeted protein (GQ385303) and the template (2RLF). The result of alignment is shown in **Fig. 1**, where the characters highlighted in red indicate the three functional residues (pH sensor His37, channel gate Trp41, and channel lock Asp44) [16–18,21,22] of the M2 channel, which are highly conserved in the M2 proteins. Those residues with the light-blue frame are the possible binding sites (Thr43, Asp44, and Arg45) of the inhibitors; while those with the green frame are different between the two sequences.

Thus, using the structural bioinformatics tools [23], the 3D structure of the H1N1-M2 (GQ385383) channel was developed by following the same procedures as elaborated in references [24–26]. The computed 3D structure of H1N1-M2 channel (GQ385383) is shown in **Fig. 2A**. Subsequently, the Auto Dock program [27] with the MMFF94 force field and atomic partial charges [28] was utilized to dock the ligands to the H1N1-M2 channel. The starting point of the docking calculations was the position at which the drug rimantadine is located in the NMR structure of 2RLF [16]. A cubic box with side length of 30 Å surrounding the ligand was used for the docking calculations. A total of 25 docking conformations with the lowest binding energies were recorded.

Position	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	
2RLF																		ARG	SER	ILE	
GQ385303	MET	SER	LEU	LEU	THR	GLU	VAI	GLU	THR	PRO	THR	ARG	SER	GLU	TRP	GLU	CYS	ARG	CYS	SER	
Position	21	22	23	24	25	26	27	28	29	30	31	32	33	34	35	36	37	38	39	40	
2RLF	ASP	SER	SER	ASP	PRO	LEU	VAL	VAL	ALA	ALA	SER	IEL	IEL	GLY	ILE	LEU	HIS	LEU	ILE	LEU	
GQ385303	ASP	SER	SER	ASP	PRO	LEU	VAL	ILE	ALA	ALA	ASN	IEL	IEL	GLY	ILE	LEU	HIS	LEU	ILE	LEU	
Position	41	42	43	44	45	46	47	48	49	50	51	52	53	54	55	56	57	58	59	60	
2RLF	TRP	ILE	LEU	ASP	ARG	LEU	PHE	PHE	LYS	SER	ILE	TYR	ARG	PHE	PHE	GLU	HIS	GLY	LEU	LYS	
GQ385303	TRP	ILE	THR	ASP	ARG	LEU	PHE	PHE	LYS	CYS	ILE	TYR	ARG	ARG	PHE	LYS	TYR	GLY	LEU	LYS	
Position	61	62	63	64	65	66	67	68	69	70	71	72	73	74	75	76	77	78	79	80	
2RLF																					
GQ385303	ARG	GLY	PRO	SER	THR	GLU	GLY	VAL	PRO	GLU	SER	MET	ARG	GLU	GLU	TYR	GLN	GLN	GLU	GLN	
Position	81	82	83	84	85	86	87	88	89	90	91	92	93	94	95	96	97				
2RLF																					
GQ385303	GLN	SER	ALA	VAL	ASP	VAL	ASP	ASP	GLY	HIS	PHE	VAL	ASN	ILE	GLU	LEU	GLU				

Figure 1. The sequence alignments between the M2 protein isolated from H1N1 influenza A virus (NCBI code: GQ385383) and the M2 channel isolated from the Udorn strain of human influenza virus. The latter 3D structure has been determined by NMR [16] with PDB code of 2RLF, and can be used to serve as a template to model the former. The GQ385383 is a complete M2 sequence consisting of 97 residues, while 2RLF only contains a segment of 43 structure-defined residues (18–60). The red codes highlight the functional residues: pH sensor His37, channel gate Trp41, and channel lock Asp44. Those residues, which are different between the two M2 sequences, are framed by a box drawn in green line, while those framed with a box drawn in light-blue line indicate the possible binding sites (Thr43, Asp44, and Arg45) of inhibitors. The functional residue Asp44 is the first binding site, and the Arg45 or Thr43 is the possible second binding site. Thr43 is a natural mutation Leu43Thr in the H1N1-M2 proton channel.

doi:10.1371/journal.pone.0009388.g001

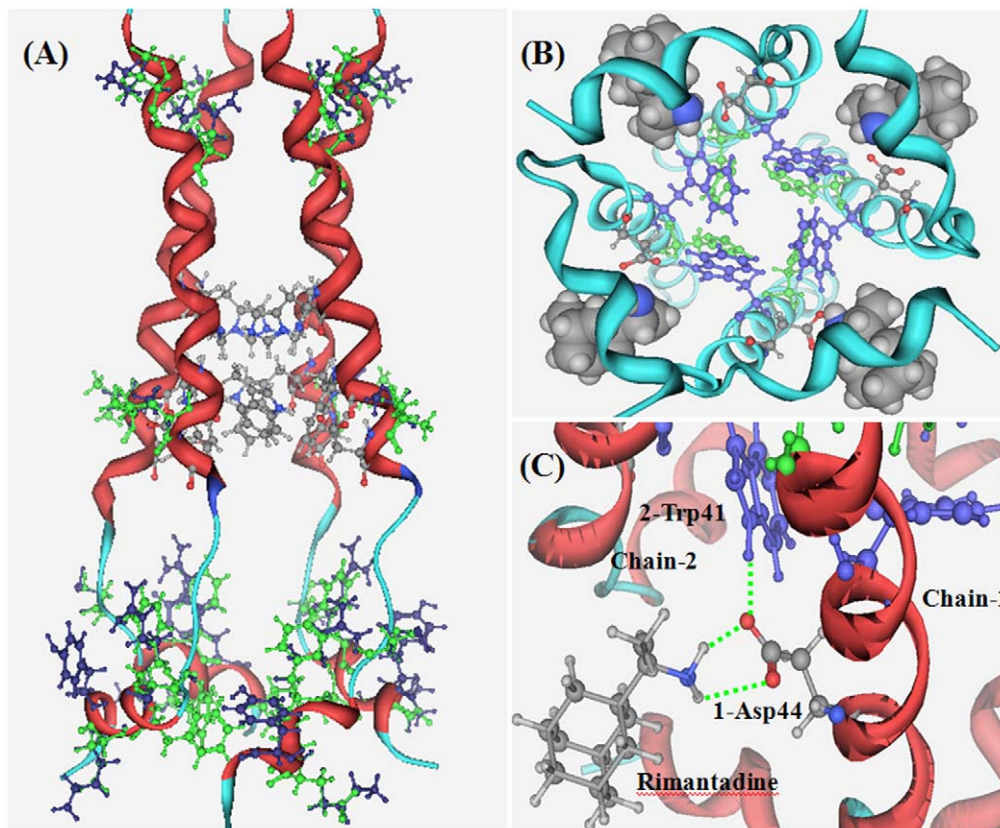


Figure 2. The computational three-dimensional structure of 2009-H1N1 M2 proton channel (NCBI code: GQ385303) and the template 2RLF with ligand rimantadine. (A) Superposition of the homology model of 2009-H1N1 M2 proton channel with its template, the NMR structure with the PDB code of 2RLF [16]. The two M2 proteins are highly homologous, and no differences are found on their backbones. The three functional residues of the M2 channel are rendered in ball-and-stick drawing with chemical element color; they are located at the middle of the channel. The different residues between two M2 proteins are shown in green for 2RLF and blue for GQ385383. (B) A close-up view from the bottom of the M2 channel. The three functional residues are rendered in green for pH sensor His37, blue for channel gate Trp41, and chemical element color for channel lock Asp44. The ligand rimantadine binds at four equivalent sites near the channel lock Asp44 on the lipid-facing side of the channel and stabilizes the closed conformation of the pore. (C) The complex structure between the inhibitor rimantadine and the receptor M2 proton channel. The channel lock Asp44 holds the channel gate Trp41 through a hydrogen bond keeping it in the closed conformation. The ligand rimantadine forms two hydrogen bonds (green dotted lines) with the Asp44 of M2 proton channel. doi:10.1371/journal.pone.0009388.g002

Results

Before the 3D NMR structure of M2 protein [16] was published, it was traditionally thought that the adamantane-based inhibitors were bound inside the channel and hence physically blocking the pore of channel [6]. With the novel allosteric inhibition mechanism revealed by the NMR structure [16], we can conduct the structure-based inhibitor design and find effective drugs against the H1N1-M2 channel in a completely different strategy [19,29], as illustrated below.

Computed Structure of H1N1 M2 Proton Channel

By following the procedures as described in the Materials and Methods section, the 3D structure of H1N1-M2 (GQ385383) was developed as shown in **Fig. 2A**, where for facilitating comparison the template structure is also given in a superposition manner with the targeted structure. The structural data can be provided by authors upon inquire. As we can see from the figure, the backbones of the two structures are almost the same as expected because the sequences of the two proteins are highly homologous. The three functional residues (His37, Trp41, and Asp44) of the M2 channel are rendered in ball-and-stick drawing with the

chemical element color. They are located at the middle of the channel. The different residues between the two channel structures are shown in green for 2RLF and blue for GQ385383. Most mutated residues in H1N1-M2 protein are at the top and bottom of the channel, and they have little effects on the functional residues (His37, Trp41, and Asp44). However, one mutation, namely Leu43Thr, is found around the active region: the non polar residue Leu43 of 2RLF is replaced by the polar residue Thr43 in the H1N1-M2 protein. A close-up view from the bottom of the M2 channel is given in **Fig. 2B** to show the position and orientation of the three functional residues and the position of ligand rimantadine. The three functional residues are rendered in green for pH sensor His37, blue for channel gate Trp41, and chemical element color for channel lock Asp44, respectively. Ligand rimantadine binds at four equivalent sites near the channel lock Asp44 on the lipid-facing side of the channel. The Asp44 residue is not only the channel lock, but also the proton exit of the M2 channel. Therefore, drug binding at this position not only can hold the channel in the closed conformation but also can simply block the proton exits. [22].

Illustrated in **Fig. 2C** is the gating and inhibiting mechanism of the M2 proton channel [16–18,22]. The channel lock Asp44 holds

the channel gate Trp41 through a hydrogen bond, keeping it in the closed conformation in the middle or higher pH (~ 7.5) condition. But in the lower pH environment, the pH sensor His37 and the indole amine of Trp41 are protonated, so as to weaken the hydrogen bond between Trp41 and Asp44, making it easily broken by the repulsive interaction between the positively charged His37 residues of two adjacent helix chains. After rimantadine binds at the Asp44 through two hydrogen bonds between amino group of rimantadine and carboxyl group of Asp44, the pK_a value of Asp44 is lowered by the two hydrogen bonds [30,31]. Therefore, in the lower pH environment, it becomes difficult for Asp44 to be protonated. That is why in the acidic condition the ligand rimantadine can help Asp44 to keep the channel in the closed conformation.

Inhibitors with Two Pharmacophore Groups

Almost all existing M2 channel inhibitors have only one pharmacophore group, and in most cases it is an amino group [19]. The structures of several adamantane-based drugs are shown in **Fig. 3**, where the inhibitors A1 and A2 are commercially available drugs amantadine and rimantadine, respectively, in

which the pharmacophore substitutes are at the position 3 of adamantane [13,32,33]. Actually, the pharmacophore substitutes also can be put on the position 2 of adamantane [11,12,19], and in many cases the substitutes on position 2 can give better results than on the position 3 [19].

The M2 channel is a tetramer consisting of four identical helices. As shown in **Fig. 4A**, the adamantane-based inhibitor with one pharmacophore group can only bind on one helix of the tetrameric M2 channel. If an additional pharmacophore group is added into the adamantane-based inhibitor, it will be able to bind at two sites of two neighboring helices. Thus, the inhibitor will be able to hold the M2 channel in the closed conformation with a more effective way, just like a hoop in fastening a barrel, as shown by **Fig. 4B**. Actually only one such inhibitor molecule would be sufficient to block the M2 channel because the hoop-like inhibitor can block half of the channel and stop the conductance of proton (H_3O^+) flow.

Structure-Based Inhibitor Design for M2 Protein

The discussions for the adamantane-based inhibitors with more than one pharmacophore groups can be found in references

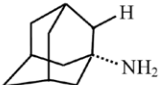
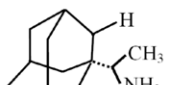
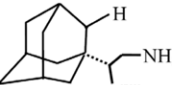
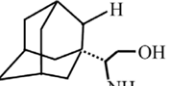
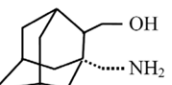
Mol	Structure	R ₁	R ₂	Expt	Compt
				(μ M)	(kCal/mol)
				EC ₅₀	ΔE_{bind}
A1		H ₂	NH ₂	49.1	-5.96
A2		H ₂	$\begin{matrix} \text{CH}_3 \\ \text{NH}_2 \end{matrix}$	19.1	-6.76
A3		H ₂	$\begin{matrix} \text{NH}_2 \\ \text{NH}_2 \end{matrix}$	18.3	-7.90
A4		H ₂	$\begin{matrix} \text{OH} \\ \text{NH}_2 \end{matrix}$	—	-9.78
A5		—OH	—NH ₂	—	-9.31

Figure 3. Summary of the existing adamantane-based drugs and newly designed inhibitors with two pharmacophore groups. A1 and A2 are the commercially available drugs amantadine and rimantadine. A3 was reported in ref [13]. A4 and A5 are inhibitors with two pharmacophore groups designed in this study. doi:10.1371/journal.pone.0009388.g003

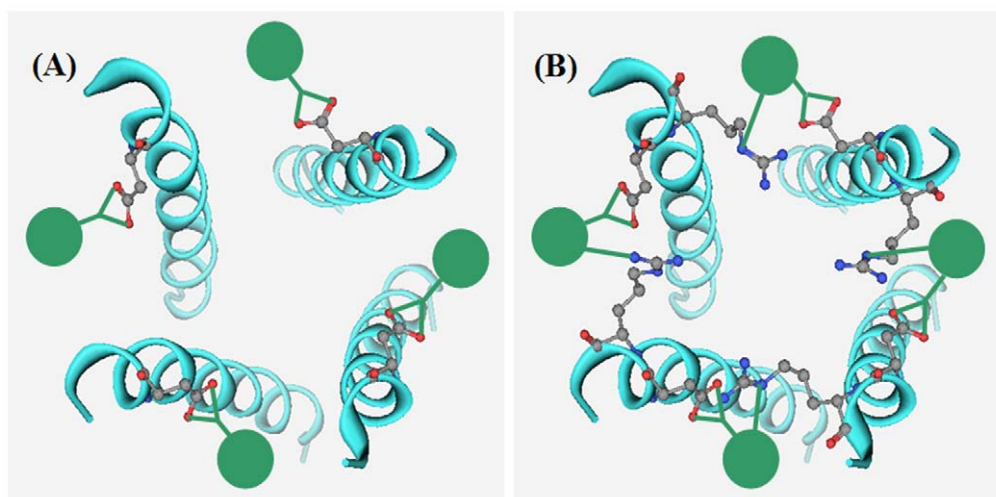


Figure 4. The binding models between M2 proton channel and adamantane-based inhibitors with one and two pharmacophore groups. (A) The adamantane-based inhibitors with one pharmacophore group can bind on only one helix of the tetrameric M2 channel. (B) With an additional pharmacophore group added into the adamantane-based inhibitors, the inhibitors can bind at the two sites of two neighboring helices of the M2 channel, and hence can hold the M2 channel in closed conformation more effectively, just like a barrel hoop. Actually only one such inhibitor is sufficient to block the M2 channel because it can block half of the channel and stop the conduction of proton (H_3O^+) flow effectively. doi:10.1371/journal.pone.0009388.g004

[12,13]. However, before a reliable 3D structure of M2 channel is available, this kind of design was lack of a footing since we did not know the binding site and the interaction mechanism of the second pharmacophore. The inhibitor A3 in **Fig. 3** has two amino groups and possesses the bioactivity slightly higher than that of rimantadine [12,13]. However, based on our docking studies, the A3 inhibitor could not effectively bind at the two neighboring helices. The inhibitor with two pharmacophore groups needs two binding sites on the M2 channel. The first binding site is unchangeably the carboxyl group of the Asp44, while the second binding site could be either the amino group of Arg45 or the hydroxyl group of Thr43 of the neighboring helix. These two residues are the closest residues to the first binding site Asp44.

With two pharmacophore groups, the inhibitors A4 and A5 in **Fig. 3** were designed based on the structure of H1N1-M2 proton channel. On the subsite of inhibitor A4 corresponding to the position 3 of adamantane, there is an amino group and a hydroxyl group. Docking calculation gives an illustration for the interactions between the designed inhibitor A4 and the M2 proton channel. The amino group of inhibitor A4 binds at the 1-Asp44 of Chain-1 through two hydrogen bonds, while the second pharmacophore hydroxyl group forms two hydrogen bonds with the amino group of 2-Arg45 of Chain-2. The detailed interactions between M2 channel and the ligand A4 are shown in **Fig. 5**. It is through the two binding sites that the inhibitor A4 holds the Chain-1 tightly with its adjacent Chain-2 of the tetrameric M2 proton channel. Actually, the inhibitor A4 was firstly synthesized by Clariana et al. in 2000 [34] not as an anti flu drug, but as a reagent for studying peptide-receptor interactions. As for inhibitor A4, no bioactivity data to M2 proton channel was reported.

The designed inhibitor A5 has two pharmacophore groups: a hydroxyl group on position 2 and an amino group on position 3 of adamantane. Illustrated in **Fig. 6** is a close view of the interactions between the designed inhibitor A5 and the M2 proton channel. The amino group of inhibitor A5 binds at the carboxyl group of 1-Asp44 of Chain-1 through two hydrogen bonds, while the second pharmacophore hydroxyl group forms two hydrogen bonds with the amino group of 2-Arg45 of Chain-2. Therefore, it is in the

same way as inhibitor A4 that the inhibitor A5 holds the Chain 1 tightly with its adjacent Chain 2 of the tetrameric M2 proton channel.

Energetic Analysis for M2 Channel-Inhibitor Interactions

During the molecular docking of the inhibitors to the tetrameric M2 proton channel, a total of 25 conformations with the strongest binding energies were recorded for each of the five inhibitors in **Fig. 3**, where the most favorable binding energy for each of the five inhibitors are also given. It can be seen from the **Fig. 3** that $\Delta E_{A1} < \Delta E_{A2} < \Delta E_{A3} < \Delta E_{A5} < \Delta E_{A4}$, indicating that the binding free energies of A4 and A5 are much stronger than those of A1

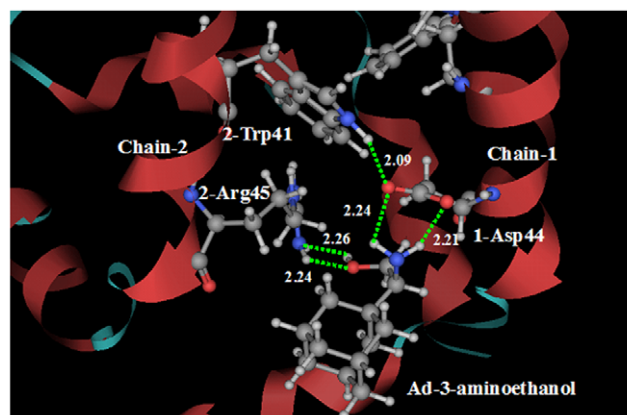


Figure 5. Illustration to show the interactions between the designed inhibitor A4 (Ad—3—aminoethanol) and the M2 proton channel. The designed adamantane-based inhibitor A4 has two pharmacophore groups. The amino group binds at the 1-Asp44 of Chain-1 through two hydrogen bonds (green dotted lines), while the second pharmacophore hydroxyl group forms two hydrogen bonds with the 2-Arg45 of Chain-2. It is through the two binding sites that the inhibitor A4 holds the Chain 1 tightly with its adjacent Chain 2 of the tetrameric M2 proton channel. doi:10.1371/journal.pone.0009388.g005

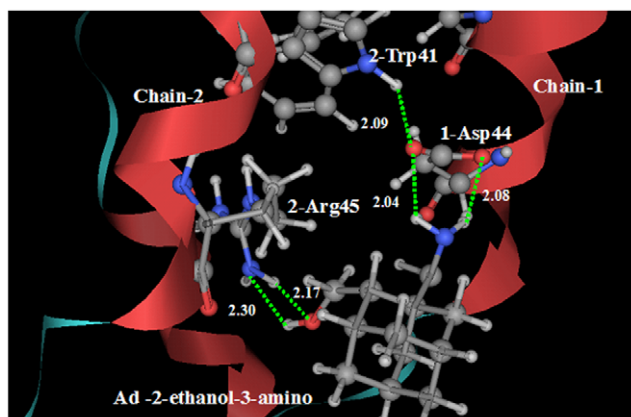


Figure 6. Illustration to show the interactions between the designed inhibitor A5 (Ad-(2-ethanol)-(3-amino)) and the M2 proton channel. The designed adamantane-based inhibitor A5 has two pharmacophore groups. The amino group of inhibitor A5 binds at the 1-Asp44 of Chain-1 through two hydrogen bonds (green dotted lines), while the second pharmacophore hydroxyl group forms two hydrogen bonds with the 2-Arg45 of Chain-2. It is through the two binding sites that the inhibitor A5 holds the Chain-1 tightly with its adjacent Chain-2 of the tetrameric M2 proton channel.
doi:10.1371/journal.pone.0009388.g006

and A2 due to the contributions of the additional pharmacophore, namely the hydroxyl group, which forms two additional hydrogen bonds with the receptor. Although A3 inhibitor also possesses two amino groups and the second amino group can form hydrogen bonds with the amino group of Arg45, the hydrogen bonds between the two amino groups are easily broken in acidic condition.

Discussion

The NMR structure of M2 proton channel [16] provides a reliable structural basis for rational drug design against influenza virus. The channel gating mechanism and the inhibiting mechanism of M2 proton channel, revealed by the NMR structure of M2 proton channel, stimulate the new idea and strategy for channel inhibitor design. The two inhibitors (A4 and A5) of M2 proton channel, designed in this study, are hopefully the potential drugs for the 2009-H1N1 swine flu.

References

- Trifonov V, Khiabani H, Rabadan R (2009) Geographic dependence, surveillance, and origins of the 2009 influenza A (H1N1) virus. *N Engl J Med* 361: 115–119.
- Shinde V, Bridges CB, Uyeki TM, Shu B, Balish A, et al. (2009) Triple-reassortant swine influenza A (H1) in humans in the United States, 2005–2009. *N Engl J Med* 360: 2616–2625.
- Bright RA, Shay DK, Shu B, Cox NJ, Klimov AI (2006) Adamantane resistance among influenza A viruses isolated early during the 2005–2006 influenza season in the United States. *J Am Med Assoc* 295: 891–894.
- Deyde VM, Xu X, Bright RA, Shaw M, Smith CB, et al. (2007) Surveillance of resistance to adamantanes among influenza A(H3N2) and A(H1N1) viruses isolated worldwide. *J Infect Dis* 196: 249–257.
- Smith BJ, McKimm-Breshkin JL, McDonald M, Fernley RT, Varghese JN, et al. (2002) Structural studies of the resistance of influenza virus neuraminidase to inhibitors. *J Med Chem* 45: 2207–2212.
- Sugrue RJ, Hay AJ (1991) Structural characteristics of the M2 protein of influenza A viruses: evidence that it forms a tetrameric channel. *Virology* 180: 617–624.
- Holsinger LJ, Nichani D, Pinto LH, Lamb RA (1994) Influenza A Virus M2 Ion Channel Protein: a Structure-Function Analysis. *J Viology* 68: 1551–1563.
- Ayton GS, Voth GA (2007) Multiscale simulation of transmembrane proteins. *J Struct Biol* 157: 570–578.
- Hay AJ (1992) The action of adamantanamines against influenza A viruses: inhibition of the M2 ion channel protein. *Semin Virol* 3: 21–30.
- Kumpf RA, Dougherty DA (1993) A mechanism for ion selectivity in potassium channels: computational studies of cation-Tr interactions. *Science* 261: 1708–1710.
- Kolocouris N, Zoidis G, Foscolos GB, Fytas G, Prathalingham SR, et al. (2007) Design and synthesis of bioactive adamantane spiro heterocycles. *Bioorg Med Chem Lett* 17: 4358–4362.
- Stamatiou G, Kolocouris A, Kolocouris N, Fytas G, Foscolos GB, et al. (2001) Novel 3-(2-Adamantyl)pyrrolidines with Potent Activity Against Influenza A Virus-Identification of Aminoadamantane Derivatives Bearing Two Pharmacophoric Amine Groups. *Bioorg Med Chem Lett* 11: 2137–2142.
- Smith BJ, Fytas G, Kolocouris A, Fytas C, Kolocouris N, et al. (2007) Influence of an additional 2-amino substituent of the 1-aminoethyl pharmacophore group on the potency of rimantadine against influenza virus A. *Bioorg Med Chem Lett* 17: 692–696.
- Hu J, Fu R, Nishimura K, Zhang L, Zhou HX, et al. (2006) Histidines, heart of the hydrogen ion channel from influenza A virus: toward an understanding of conductance and proton selectivity. *Proc Natl Acad Sci USA* 103: 6865–6870.
- Hu F, Fu R, Cross TA (2007) The chemical and dynamic influence of the drug amantadine on the M2 proton channel transmembrane domain. *Biophys J* 93: 276–283.

The adamantane-based drugs (amantadine and rimantadine) are not like the ordinary drug molecules from the viewpoint of Lipinski's "the rule of five" [35], a rule of thumb to evaluate druglikeness, or to determine if a chemical compound with a certain pharmacological or biological activity has properties that would make it a likely orally active drug in humans. This is because amantadine and rimantadine possesses very few pharmacophore groups and very few hydrogen-bond-forming elements. Rimantadine can hold one of the four helices in the tetrameric M2 proton channel by one, and only one, pharmacophore amino group. An additional pharmacophore group is needed to hold the adjacent helix of the tetrameric channel so as to strengthen its closed conformation for blocking the proton conductance. The high-resolution NMR structure of the M2 proton channel [16] and the gating and inhibiting mechanism revealed therefrom has made it possible to rationally design new and more powerful drugs against influenza viruses. It is the second pharmacophore group in the inhibitors A4 and A5 that might significantly enhance their ability in inhibiting the M2 channel in comparison with amantadine and rimantadine.

The M2 proton channel is a membrane protein; while the adamantane-based inhibitors are detergent-like compounds with a hydrophilic head and a hydrophobic body, possessing the ability to penetrate the bilayer lipid membrane. Although the second additional hydrophilic pharmacophore group of A4 and A5 inhibitors can enhance their inhibition ability to the M2 proton channel, it might lower their ability in penetrating membrane. Therefore, a series of follow-up experiments are needed along this direction to find an optimal inhibitor by taking into account these two aspects.

Acknowledgments

We wish to thank the two anonymous reviewers for their constructive comments, which are very helpful for strengthening the presentation of this study.

Author Contributions

Conceived and designed the experiments: QSD RBH KCC. Performed the experiments: QSD RBH SQW. Analyzed the data: QSD RBH SQW KCC. Wrote the paper: QSD KCC. Calculations: QSD SQW. Drew figures: SQW.

16. Schnell JR, Chou JJ (2008) Structure and mechanism of the M2 proton channel of influenza A virus. *Nature* 451: 591–595.
17. Huang RB, Du QS, Wang CH, Chou KC (2008) An in-depth analysis of the biological functional studies based on the NMR M2 channel structure of influenza A virus. *Biochem Biophys Res Commun* 377: 1243–1247.
18. Du QS, Huang RB, Wang CH, Li XM, Chou KC (2009) Energetic analysis of the two controversial drug binding sites of the M2 proton channel in influenza A virus. *J Theor Biol* 259: 159–164.
19. Wei H, Wang CH, Du QS, Meng J, Chou KC (2009) Investigation into Adamantane-Based M2 Inhibitors with FB-QSAR. *Medicinal Chemistry* 5: 305–317.
20. Morton CC (2008) With proton channel structure, flu fighters record hit against infection. *Harvard Focus News*, http://focus.hms.harvard.edu/2008/022208/structural_biology.shtml.
21. Takeuchi H, Okada A, Miura T (2003) Roles of the histidine and tryptophan side chains in the M2 proton channel from influenza A virus. *FEBS Lett* 552: 35–38.
22. Pielak RM, Schnell JR, Chou JJ (2009) Mechanism of drug inhibition and drug resistance of influenza A M2 channel. *Proc. Natl Acad Sci USA* 106: 7379–7384.
23. Chou KC (2004) structural bioinformatics and its impact to biomedical science. *Curr Med Chem* 11: 2105–2134.
24. Chou KC, Jones D, Heinrikson RL (1997) Prediction of the tertiary structure and substrate binding site of caspase-8. *FEBS Letters* 419: 49–54.
25. Chou KC (2004) Insights from modelling the 3D structure of the extracellular domain of alpha7 nicotinic acetylcholine receptor. *Biochem Biophys Res Commun* 319: 433–438.
26. Chou KC (2004) Molecular therapeutic target for type-2 diabetes. *J Proteome Res* 3: 1284–1288.
27. Morris GM, Goodsell DS, Halliday RS, Huey R, Hart WE, et al. (1998) Automated docking using a Lamarckian Genetic Algorithm and empirical binding free energy function. *J Comput Chem* 19: 1639–1662.
28. Cornell WD, Cieplak P, Bayly CI, Gould IR, Merz Jr KM, et al. (1995) A second generation force field for the simulation of proteins and nucleic acids. *J Am Chem Soc* 117: 5179–5197.
29. Borman S (2008) Flu virus proton channel analyzed: structures of key surface protein suggest different drug mechanisms. *Chem Eng News* 86: 53–54.
30. Li H, Robertson AD, Jensen JH (2004) The determinants of carboxyl pKa values in turkey ovomucoid third domain. *Proteins* 55: 689–704.
31. Li H, Robertson AD, Jensen JH (2005) Very fast empirical prediction and rationalization of protein pKa values. *Proteins* 61: 704–721.
32. Kolocouris A, Tataridis D, Fytas G, Mavromoustakos T, Foscolos GB, et al. (1999) Synthesis of 2-(2-adamantyl) piperidines and structure anti-influenza virus A activity relationship study using a combination of NMR spectroscopy and molecular modeling. *Bioorg Med Chem Lett* 9: 3465–70.
33. Turner JV, Maddalena DJ, Agatonovic-Kustrin S (2003) Bioavailability Prediction Based on Molecular Structure for a Diverse Series of Drugs. *Pharmaceutical Research* 21: 68–82.
34. Clariana J, Garcia-Granda S, Gotor V, Gutiérrez-Fernández A, et al. (2000) Preparation of (R)-(1-adamantyl)glycine and (R)-2-(1-adamantyl)-2-aminoethanol: a combination of cobalt-mediated b-ketoester alkylation and enzyme-based aminoalcohol resolution. *Tetrahedron: Asymmetry* 11: 4549–4557.
35. Lipinski CA, Lombardo F, Dominy BW, Febej PJ (2001) Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv Drug Deliv Rev* 46: 3–26.