MITOGENOME ANNOUNCEMENT

Taylor & Francis Taylor & Francis Group

OPEN ACCESS Check for updates

The mitochondrial genome of an Endangered freshwater snail *Koreoleptoxis nodifila* (Caenogastropoda: Semisulcospiridae) from South Korea

Eun Hwa Choi^a*, Na Rae Choi^a* and Ui Wook Hwang^{a,b,c}

^aDepartment of Biology Education, Teachers College & Institute for Phylogenomics and Evolution, Kyungpook National University, Daegu, South Korea; ^bInstitute for Korean Herb-Bio Convergence Promotion, Kyungpook National University, Daegu, South Korea; ^cBiomedical Convergence Science and Technology, Kyungpook National University, Daegu, South Korea

ABSTRACT

The mitochondrial genome of the Endangered freshwater snail *Koreoleptoxis nodifila* (Caenogastropoda: Semisulcospiridae) from South Korea is determined and characterized in detail. It is 15,737 bp in length being composed of 13 protein-coding genes (PCGs), 22 transfer *RNA* genes (tRNAs), two ribosomal *RNA* genes (rRNAs), and one control region. It has a base composition of 31.23% for A, 16.29% for G, 17.84% for C, and 34.64% for T. The phylogenetic trees reconstructed based on the maximum-likelihood (ML) method and Bayesian inference (BI) confirmed that *K. nodifila* belongs to the Semisulcospiridae clade in the monophyletic caeonogastropod superfamily Cerithioidea.

ARTICLE HISTORY

Received 15 December 2020 Accepted 7 March 2021

KEYWORDS

Koreoleptoxis nodifila; Semisulcospiridae; Caeonogastropoda; mitochondrial genome; phylogeny

The family Semisulcospiridae, a member of Cerithioidea, is speciose and widely distributed throughout eastern Asia, including the Korean Peninsula, the Russian Far East, Japan, southern China, and Taiwan (Davis 1969; Chiu et al. 2017) as well as western North America (Strong and Köhler 2009). In South Korea, a total of seven species from the three semisulcospirid Koreanomelania, Koreoleptoxis, and genera Semisulcospira have been recorded so far (Ko et al. 2001; Lee et al. 2001). Recently, the genus Koreanomelania Burch & Jung, 1988 has been synonymized with the genus Koreoleptoxis Burch & Jung, 1988 based on the analyses of morphological data and molecular data, such as mitochondrial COI and 16S rRNA (Köhler 2017).

During the last decades, mitochondrial genomes have been considered useful phylogenetic markers in mollusks (Lee et al. 2012; Zeng et al. 2015; Hilgers et al. 2016; Cha et al. 2018; Kim and Lee 2018; Jiang et al. 2019; Lee et al. 2019; Tenorio et al. 2020). Here, we characterized a nearly complete mitochondrial genome of the Endangered freshwater snail *Koreoleptoxis nodifila* (Martens 1886). The sample used for this study was collected from Yongtan-ri, Jeongseon-eup, Jeongseon-gun, Gangwon-do, South Korea (37°23'35.3"N, 128°36'00.4"E). The specimen was deposited under the voucher number FNRVIV000000082 in the National Institute of Biological Resources (NIBR), Ministry of Environment, South Korea. Genomic DNA was extracted from the muscle tissue using a DNeasy Blood & Tissue kit (Qiagen, Hilden, Germany). Sequencing was performed from an Illumina paired-end library and 150 bp paired-end reads were generated through the Illumina NextSeg500 platform, resulting in a total of 81,273,381 reads (not published). For data assembly, sequencing errors were discarded using the error correction module of SOAPec version 2.02 (Luo et al. 2012). And then, mitochondrial genome assembly of K. nodifila was conducted using SOAPdenovo2 version 2.04-r240 (Luo et al. 2012). Reconstruction of the mitochondrial genome was performed with MITObim (Hahn et al. 2013) using the Illumina reads. The seed sequences were extended by mapping reads iteratively using MITObim. Finally, after conducting manual curation, the annotation of mitochondrial genes, such as protein-coding genes (PCGs), tRNA genes, and rRNA genes was carried out using the MITOS web server (Bernt et al. 2013). The mitogenome of K. globus (GenBank accession no. LC006055) was used as a reference for the annotation.

The mitochondrial genome of *K. nodifila* (GenBank accession no. NC_046494), which is 15,737 bp in length, exhibits the same gene components and gene order as those previously known from the family Semisulcospiridae, such as *Semisulcospira coreana* (Martens 1886) (Kim and Lee 2018), *S. libertina* (Zeng et al. 2015), and *Koreoleptoxis globus* (Martens 1886) (GenBank accession number LC006055; unpublished data). It contains a total of 37 genes including 13 PCGs (*COX1-3, ND1-6, ND4L, CYTB, ATP6*, and *ATP8*), two rRNAs (*16S rRNA* and *12S rRNA*), 22 tRNAs, and one control region, of which 16 genes are located on the positive strand and the remaining on the negative strand. The overall A + T content

© 2021 The Author(s). Published by Informa UK Limited, trading as Taylor & Francis Group.

CONTACT Ui Wook Hwang 🔯 uwhwang1@gmail.com 🗈 Department of Biology, Teachers College & Institute for Phylogenomics and Evolution, Kyungpook National University, Daegu 41655, South Korea

^{*}These authors have contributed equally to this work.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.



Figure 1. A maximum-likelihood tree reconstructed with the amino acid sequences of 13 mitochondrial PCGs showing relationships among 42 caenogastropod species. *Pleuropoma jana* (Neritimorpha) and *Haliotis rubra* (Vetigastropoda) are used as outgroups. The monophylies of Caenogastropoda and Hypsogastropoda are indicated with an arrow, respectively. The dark yellow box indicates the monophyly of the superfamily Cerithioidea including this result of *Koreoleptoxis nodifila*. Branch supports are bootstrapping values in percent (BP) obtained the ultrafast bootstrap method using IQ-TREE webserver and Bayesian posterior probability (BPP) inferred from Bayesian inference using MrBayes version 3.2.7a in order. The nodes exhibiting both 100 BP and 1.00 BPP mark an asterisk (*). Major habitats of the taxa examined here are depicted beside each taxon name.

of the *K. nodifila* mitogenome is 65.87%: 64.66% for PCGs and 65.85% for tRNAs and rRNAs. The control region is located between *trnF* and *trnC*. Its size was predicted to be about 3 kb, out of which only 399 bp were sequenced. The control region sequencing could not be completed due to the A + T - rich hairpin structures and too long A + T - dinucleotide repeats. This is also observed in the

semisulcospirid *K. globus* and the pachychilid *Tylomelania sar-asinorum* (Hilgers et al. 2016).

The maximum likelihood (ML) and Bayesian inference (BI) trees were reconstructed based on the amino acid sequences of the 13 PCGs from 42 caenogastropod species (Figure 1). The best-fitting model mtZOA + F + I + G4 for the ML analysis was selected using the implemented function in IQ-Tree (

Trifinopoulos et al. 2016). We also performed BI analysis using MrBayes version 3.2.7a (Ronguist et al. 2012) under the best model mtREV + I + G4, each with four MCMC chains for 1,000,000 generations. Sampling was done at the end of every 1000th generation with 25% of the initial trees discarded as burnin. Both ML and BI trees strongly supported the monophyly of the subclass Caenogastropoda (100% BP - bootstrapping values in percent and 1.0 BPP - Bayesian posterior probability). It also confirmed that K. nodifila belongs to the Semisulcospiridae (100% BP and 1.0 BPP) in the monophyletic superfamily Cerithioidea (100% BP and 1.0 BPP) which includes Semisulcospiridae (Semisulcospira and Koreoleptoxis), Pachychilidae (Tylomelania), and Turritellidae (Turritella). Hypsogastropoda appeared as a monophyletic group (100% BP and 1.0 BPP), with exception of the superfamily Vermetoidea which placed at the basal branch of Caenogastropoda. Due to the unexpected placement of the superfamily Vermetoidea, the monophyly of the order Littorinimorpha was not supported, but was polyphyletic instead. The peculiar positioning of Vermetoidea, which is not in line with morphological evidence (Sigwart and Lindberg 2015), may be interpreted as a spurious result caused by the long-branch attraction artifact or some unique mitochondrial genome characteristics (i.e. Jiang et al. 2019). Within Hypsogastropoda excepting Vermetoidea, phylogenetic analyses did not recover Littorinimorpha as a monophyletic group, but rather suggested that this group is paraphyletic with respect to the order Neogastropoda. Whether the Littorinimorpha in this sense really is not a monophyletic groups needs further investigation. In addition, Architaenioglossa was not recovered as a monophyletic group because Cochlostomatidae was not placed in this clade. Compared to other studies addressing caenogastropod phylogeny based on mitochondrial genomes (Osca et al. 2015; Lee et al. 2019), the results presented here may be considered better-resolved because the monophyly of Viviparidae and Ampullariidae was recovered, although with somewhat low support in ML only (67% BP). Unexpectedly, within the order Neogastropoda, the monophylies of Conoidea and Volutoidea were not supported in the ML and BI analyses. These also need to be examined in detail in future studies.

Acknowledgments

The authors thank Dr. Jun Sang Lee, who works for the Department of Life Sciences and Biotechnology, College of Natural Sciences, Soonchunhyang University, Asan, South Korea, for his generous sampling and species identification of the specimen used in this study.

Disclosure statement

No potential conflict of interest was reported by the author(s).

Funding

This research was supported by Kyungpook National University Development Project Research Fund, 2018.

Data availability statement

The data that support the findings of this study are openly available in GenBank of NCBI at https://www.ncbi.nlm.nih.gov/nuccore/NC_046494. The associated BioProject, SRA, and BioSample numbers are PRJNA693879, SRR13516403, and SAMN17487625, respectively.

References

- Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritzsch G, Pütz J, Middendorf M, Stadler PF. 2013. MITOS: improved de novo metazoan mitochondrial genome annotation. Mol Phylogenet Evol. 69(2): 313–319.
- Cha SH, Lee JH, Choi EH, Jang KH, Lim YJ, Kim SG, Ryu SH, Lee YS, Hwang UH. 2018. The complete mitochondrial genome of a freshwater mussel *Nodularia douglasiae* (Bivalvia: Unionidae). Mitochondrial DNA B Resour. 3(2):945–946.
- Chiu YW, Bor H, Kuo PH, Hsu KC, Tan MS, Wang WK, Lin HD. 2017. Origins of *Semisulcospira libertina* (Gastropoda: Semisulcospiridae) in Taiwan. Mitochondrial DNA A DNA Mapp Seq Anal. 28(4):518–525.
- Davis GM. 1969. A taxonomic study of some species of *Semisulcospira* in Japan (Mesogastropoda: Pleuroceridae). Malacologia. 7:211–294.
- Hahn C, Bachmann L, Chevreux B. 2013. Reconstructing mitochondrial genomes directly from genomic next-generation sequencing reads a baiting and iterative mapping approach. Nucleic Acids Res. 41(13): e129–e129.
- Hilgers L, Grau JH, Pfaender J, von Rintelen T. 2016. The complete mitochondrial genome of the viviparous freshwater snail *Tylomelania sarasinorum* (Caenogastropoda: Cerithioidea). Mitochondrial DNA B Resour. 1(1):330–331.
- Jiang D, Zheng X, Zeng X, Kong L, Li Q. 2019. The complete mitochondrial genome of *Harpago chiragra* and *Lambis lambis* (Gastropoda: Stromboidea): implications on the Littorinimorpha phylogeny. Sci Rep. 9(1):17683.
- Kim Y, Lee S. 2018. The complete mitochondrial genome of freshwater snail, *Semisulcospira coreana* (Pleuroceridae: Semisulcospiridae). Mitochondrial DNA B Resour. 3(1):259–260.
- Ko JH, Lee JS, Kwon OK. 2001. Study on radulae of seven species of the Family Pleuroceridae in Korea. Korean J Malacol. 17(2):105–115.
- Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B. 2017. Partition finder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. Mol Biol Evol. 34(3):772–773.
- Lee JH, Choi EH, Kim SK, Ryu SH, Hwang UW. 2012. Mitochondrial genome of the cockscomb pearl mussel *Cristaria plicata* (Bivalvia, Unionoida, Unionidae). Mitochondrial DNA. 23(1):39–41.
- Lee JS, Ko JH, Kwon OK. 2001. Isozyme variation in two species of freshwater Pleurocerid snails in Korea: *Koreanomelania nodifila* and *Koreoleptoxis globus ovalis*. Korean J Malacol. 17(2):117–123.
- Lee SY, Lee HJ, Kim YK. 2019. Comparative analysis of complete mitochondrial genomes with Cerithioidea and molecular phylogeny of the freshwater snail, *Semisulcospira gottschei* (Caenogastropoda, Cerithioidea). Int J Biol Macromol. 135:1193–1201.
- Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y, et al. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. Gigascience. 1(1):18.
- Osca D, Templado J, Zardoya R. 2015. Caenogastropod mitogenomics. Mol Phylogenet Evol. 93:118–128.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient bayesian phylogenetic inference and model choice across a large model space. Syst Biol. 61(3):539–542.
- Sigwart JD, Lindberg DR. 2015. Consensus and confusion in molluscan trees: evaluating morphological and molecular phylogenies. Syst Biol. 64(3):384–395.
- Strong EE, Köhler F. 2009. Morphological and molecular analysis of 'Melania' jacqueti Dautzenberg and Fischer, 1906: from anonymous

orphan to critical basal offshoot of the Semisulcospiridae (Gastropoda: Cerithioidea). Zool Scr. 38(5):483–502.

- Tenorio MJ, Abalde S, Pardos-Blas JR, Zardoya R. 2020. Taxonomic revision of West African cone snails (Gastropoda: Conidae) based upon mitogenomic studies: implications for conservation. Eur J Taxon. 663: 1–89.
- Trifinopoulos J, Nguyen LT, von Haeseler A, Minh BQ. 2016. W-IQ-TREE: a fast online phylogenetic tool for maximum likelihood analysis. Nucleic Acids Res. 44(W1):W232–W235.
- Zeng T, Yin W, Xia R, Fu C, Jin B. 2015. Complete mitochondrial genome of a freshwater snail, *Semisulcospira libertina* (Cerithioidea: Semisulcospiridae). Mitochondrial DNA. 26(6):897–898.