Hindawi Publishing Corporation Journal of Allergy Volume 2010, Article ID 238573, 5 pages doi:10.1155/2010/238573

Research Article

In Vitro Determination of the Allergenic Potential of Egg White in Processed Meat

Sabine Hildebrandt,¹ Larsen Schütte,¹ Stefan Stoyanov,² Günther Hammer,² Hans Steinhart,¹ and Angelika Paschke¹

¹ Institute of Biochemistry and Food Chemistry, University of Hamburg, Grindelallee 117, 20146 Hamburg, Germany

Correspondence should be addressed to Angelika Paschke, angelika.paschke@uni-hamburg.de

Received 21 December 2009; Accepted 13 January 2010

Academic Editor: Fabienne Ranceé

Copyright © 2010 Sabine Hildebrandt et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Hen's egg white has been reported as a causative agent of allergic reactions, with ovalbumin, conalbumin, ovomucoid, and lysozyme being the major allergens. However, little is known about the effects of processing with heat and high pressure on the allergenicity of egg white proteins as ingredients in meat. For this purpose, the allergenic characteristics of such treated preparations were studied. The IgE-binding capacity was analyzed by EAST inhibition in raw and processed meat preparations using sera from patients with hen's egg specific IgE. Increasing heat treatment as well as the application of high pressure decreased IgE binding, which is a measure of allergenic potential. The combined application of heat (70°C) and high pressure had synergistic effects in reducing the allergenic potential nearly twice as the sum of the single treatments conducted separately.

1. Introduction

Major allergens of egg white (ovalbumin, conalbumin, ovomucoid, and lysozyme), which rank among the most frequent initiators of food hypersensitivities in children and adults [1], are well characterized. The European Union project REDALL (Reduced Allergenicity of Processed Foods (Containing Animal Allergens), QLK1-CT-2002-02687), supports the development of technologies to reduce the allergenicity of products containing egg white. Many meat products contain additives and ingredients that may possess a risk for consumers in food allergies. This is especially true for egg which is used as a thickener in meat preparations. During food processing, the allergenicity of hen's egg may be altered by mincing and heating associated with industrial preparation of the final products. Moreover, chemical reactions during food processing between natural food ingredients and food additives can occur. Despite these potential sources of protein interaction, only a few allergens do not survive processing. Heat treatment has been recognized as a way of reducing allergenicity and boiled hen's egg has been reported as less allergenic [2–5]. However, severe food hypersensitivity reactions are also described for heat treated hen's egg [6]. Indeed, researchers have demonstrated the stability of ovomucoid against heat denaturation [7–11]. High pressure can also be used to denature proteins [12]. This study investigates the effects of heat and pressure on the binding of IgE to egg white proteins in processed food, particularly in meat preparations as an indication of allergenicity.

A study accomplished by Schöberl [13] demonstrates that after high pressure treatment with >300 MPa raw meat samples showed an inactivation of enzymes, a consolidation of texture (caused by coagulation of dissolved sarcoplasmatic proteins), and a loss of the native red color (caused by autoxidation of myoglobine to brown metmyoglobine). However, the effect of high pressure on the allergenic potential of food is barely researched so far. Since studies by Jankiewicz et al. [14] and Scheibenzuber [15] showed alterations of the allergenic potential of different foods caused by >300 MPa and 600 MPa, respectively, the high pressure treatments

² Bundesforschungsanstalt für Ernährung und Lebensmittel, Institute of Meat Technology, E.-C. Baumann-Straße 20, 95326 Kulmbach, Germany

progressed within the scope of these investigations were accomplished with 600 MPa.

2. Materials and Methods

2.1. Chemicals and Patient Sera. Phosphate buffered saline (PBS, 150 mM NaCl, 10 mM K₂HPO₄ at pH 7.4) was prepared as described by Bernhisel-Broadbent et al. [16]. If not otherwise mentioned, all chemicals were of analytical grade.

Patient sera were collected from 12 patients with egg allergy and a positive enzyme allergosorbent test (EAST, Spez. IgE ELISA RV 5, Allergopharma, Reinbek, Germany), class 2, 3, or 4 for egg white, and pooled. Patients were procured by the Technical University of Munich (Department of Dermatology and Allergology), the University Hospital of Zurich (Allergiestation, Dermatologische Klinik), the Macedonio Melloni Hospital of Milan (Department of Pediatrics), and the Medical University of Vienna (Department of Pediatrics and Juvenile Medicine). Two healthy patients without egg allergy were deployed as negative controls.

2.2. Samples. The processed meat matrix similar to a cooked sausage-batter with beef and fat from pork was established and provided by Bundesforschungsanstalt für Ernährung und Lebensmittel (Institute of Meat Technology, Kulmbach, Germany). Meat products were made from beef (shoulder, 56%), pork (back fat, 24%) and other technologically active substances. These are ice (18.3%), nitrite curing salt (1.65% NaNO₂, in combination with 99.4 to 99.5% of table salt), dried egg powder (1%) and sodium ascorbate (0.05%). A sausage batter-system was used, which is near to industrial standards with the exception that besides fat no pork is used and that no spices are added.

Heat treatments were accomplished in the same way than it is usually done for the industrial production of fresh products (70° C) and different canned meat products (F = 1, F = 3, F = 12). Products like these are already on the market and well accepted by the consumer. The F-value is a measurement for the total quantity of heat that induces harmful effects for microorganisms. The F-value equals the time of heat treatment in minutes, which is required to reduce a specific bacteria count to an accepted end value [17].

The following processing steps are demonstrated in Figure 1: (0): Standard: pasteurized dried egg powder (homogenized, pasteurized, spray-dried whole egg powder from chicken, provided by the company OVOBEST, Germany, EC-No: D/NI-EP-003/EWG). (1): grounded beef and back fat of pigs. (2): chopped meat batches with 1% of dried egg powder. (3): heating of (2) to 70°C (as in products labeled with "best before ..." plus day and month, when still "best"). (4): heating of (2) to an *F*-value of 1 (110°C, 2 atm). (5): heating of (2) to an *F*-value of 3 (110°C, 2 atm, as in products labeled with "best before ..." plus the year, when still "best"). (6): heating of (2) to an *F*-value of 12 (110°C, 2 atm). (7): high pressure treatment of (2) (600 MPa, 20°C, 10 minutes, in an autoclave). (8): combination of (7) and (3) consecutively. (9): combination of (7) and (5)

consecutively. (10): combination of (7) and (6) consecutively. (11): combination of (6) and (7) consecutively.

- 2.3. Protein Determination. Protein concentrations were determined relatively according to the method of Bradford [18], using bovine serum albumin as standard and Bradford reagent consisting of Coomassie Brilliant Blue G-250 and phosphoric acid.
- 2.4. Protein Extraction. Protein sample extracts have been carried out by mixing 1 g of the sample with 5 ml of PBS solution according to Leduc et al. [19] in a laboratory blender (Waring, New Hartford, USA) for 5 minutes. The homogenate was extracted for 1 hour on a laboratory shaker (Bühler, Tübingen, Germany) at 4° C and centrifuged for 30 minutes with $950 \times g$ (Sigma, Osterode, Germany).
- 2.5. East Inhibition. EAST inhibition experiments are applied to compare the IgE binding potency of the different preparations. An additional homologous inhibition is carried out to check the allergenic activity of the native egg extract. The results are expressed as % inhibition. Extract potencies are quantitatively compared by estimating the protein concentration responsible for a 50% inhibition of the IgE binding to the solid phase (C_{50}) from the inhibition graphs.

For EAST inhibition assay standard egg protein extract (PBS extracts see above) was linked to bromocyane activated paper discs (Schleicher & Schüll, Dassel, Germany) using a modified method from Ceska and Lundkvist [20]. $50 \mu L$ of patients sera pool (diluted 1:2), previously incubated with different concentrations of the particular beef preparation or egg standard extract, were subsequently added to the discs and incubated for 3 hours at room temperature in cavities of a microtiter plate (Minisorb, 96 cavity, Nunc, Roskilde, Denmark). Allergopharma (Reinbek, Germany) test kit (Spez. IgE ELISA RV 5) was used for EAST inhibition according to the manufacturer's recommendations with modifications. Free binding sites were blocked with ethanolamine for 1 hour. Dilution series of the inhibitor extracts (extract (0) to (11)) were prepared in 7 steps (undiluted, 1:10, 1:100, 1:1000, 1:10000, 1:100000, 1:1000000). Potato protein was used to check nonspecific inhibition. A total of 50 µL of diluted pool serum was added and incubated for 1 hour at 37°C in the dark. After 3 washes with 1% Tween 20 in PBS, 50 µL of antihuman IgE alkaline phosphatase conjugate (Allergopharma, Reinbek, Germany, diluted 1:200 in incubation buffer) were added and incubated for 1.5 hours at 37°C in the dark. The plate was washed again and the bound enzyme activity was stained with 200 µL of staining solution (containing p-nitrophenylphosphate (PNPP)) for 1 hour at 37°C in the dark. After the addition of stopping solution (100 µL, 1 M NaOH) absorbance was measured at 405 nm. All EAST inhibition experiments were performed in duplicate and data were given in mean values.

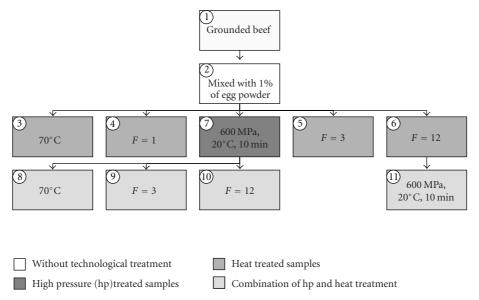


FIGURE 1: Steps of the technological process: (0): Standard: dried egg powder. (1): grounded beef and back fat of pigs. (2): chopped meat batches with 1% of dried egg powder. (3): heating of (2) to 70°C. (4): heating of (2) to an *F*-value of 1 (110°C, 2 atm). (5): heating of (2) to an *F*-value of 3 (110°C, 2 atm). (6): heating of (2) to an *F*-value of 12 (110°C, 2 atm). (7): high pressure treatment of (2) (600 MPa, 20°C, 10 minutes). (8): combination of (7) and (3) consecutively. (9): combination of (7) and (5) consecutively. (10): combination of (6) and (7) consecutively.

3. Results

IgE binding of the protein extracts obtained after the different process steps was determined by the competitive EAST inhibition test using pool serum from hen's egg white allergic patients. The results are depicted in Figure 2 and the corresponding C₅₀ values are shown in Table 1. The binding of IgE is an indication of potential allergenicity. In this assay, the degree of allergenicity was measured on the basis of inhibition of IgE by the total proteins in the extract from beef preparations. In this case, the higher the inhibition (means the lower the C_{50} -value), the more allergenic the extract is. Results showed that—as expected for the negative control (sample (1)) no C₅₀-value could be measured. As sample (2) has a total egg white protein content of only 1%, it is expected that the C50-value of this sample should be 1% of that measured for the pure dried egg white powder (sample (0)). The results shown in Table 1 are in accordance with this expectation, as sample (0) has a C₅₀value of 1.3 μ g/ml and sample (1) a C₅₀-value of 145 μ g/ml. An increasing intensity of heating of the samples resulted in lower inhibitions (and higher C₅₀-values) which indicates a lower allergenic potential of the samples. Heating to 70°C (sample (3)) resulted in an increasing of the C₅₀-value of 1.5 fold of the unheated sample (2). After heating to an Fvalue of 1 the inhibition curve was such flat that a C_{50} -value could not be measured with this EAST inhibition method. This is also considered for heated samples (5) and (6).

High pressure treatment (sample (7)) resulted in an increasing of the C_{50} -value of more than 3fold of the unheated sample (2). By additional heating to 70° C of the high pressure treated sample (sample (8)) the C_{50} -value increased again of more than 2.5 fold in comparison to

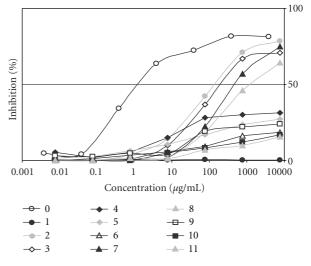


FIGURE 2: East inhibition with the different process step samples as inhibitors. (0): Standard: dried egg powder. (1): grounded beef and back fat of pigs. (2): chopped meat batches with 1% of dried egg powder. (3): heating of (2) to 70°C. (4): heating of (2) to an *F*-value of 1 (110°C, 2 atm). (5): heating of (2) to an *F*-value of 3 (110°C, 2 atm). (6): heating of (2) to an *F*-value of 12 (110°C, 2 atm). (7): high pressure treatment of (2) (600 MPa, 20°C, 10 minutes). (8): combination of (7) and (3) consecutively. (9): combination of (7) and (5) consecutively. (10): combination of (7) and (6) consecutively. (11): combination of (6) and (7) consecutively.

sample (7) that was high pressure treated only. The inhibition curves of the samples (9) to (11) (combinations of high pressure treatment and heating to an F-value of 3 or 12)) are flat and a C_{50} -value could not be measured with this

Sample no.	C_{50} -value ($\mu g m L^{-1}$)
0	1.3
1	_a
2	145
3	214
4	_
5	-
6	_
7	485
8	1286
9	-
10	_
11	_

^aNo C₅₀-value could be measured.

EAST inhibition method. Here, the inhibition curves of the samples with additional high pressure treatment are even more flat than that of the samples that are only heat treated. A comparison of samples (10) and (11) shows no obvious differences in the run of the curves whereas sample (11) that was at first heat treated with an F-value of 12 and than treated with high pressure tends to result in a slightly more flat curve than sample (10) where the treatings happened vice versa.

4. Discussion

For technological reasons—such as viscosity, cohesion, emulsification, and foam formation—whole egg powder is added to different foods during the manufacturing process. Leduc et al. [19] and Kato et al. [21] showed that the food matrix can influence the allergenic potential of the food ingredients during technological treatment. Therefore, within the scope of these investigations the allergenic potential of a model food was examined. For that purpose a meat preparation with an additive of powdered whole egg (see experimental section) for the production of cooked sausages was used as a model food. For technological treatments different high pressure and/or heat treatments were used (see experimental section).

The results of the investigations of the powdered whole egg used as an additive in meat preparations for the production of boiled sausages showed that an increasing heat treatment as well as a high pressure treatment results in a clear alteration of the egg proteins and their allergenic properties. Studies showed that the longer the heating time, the stronger the reduction of IgE-binding properties, which is in accordance with the findings of this study [22–24]. After heat treatment of the sample with 70°C a reduction of the allergenic potential of 1.5fold in comparison to the unheated sample could be measured by EAST inhibition method. All other heat treatments (F = 1, F = 3, F = 12) resulted in a very strong reduction of the allergenic potential in such a way that it could no longer be measured by EAST inhibition method. Nevertheless, a tendency of the

decreasing of the allergenic potential from F = 1 through F = 13 to F = 12 can be derived from the curve progressions of the EAST inhibition method (Figure 2). EAST inhibition results also showed that high pressure treatment of the sample caused a 3.3fold reduction of the egg allergenic potential in comparison to the untreated sample. Thus high pressure treatment resulted in a 2.5fold stronger reduction of the allergenic potential of the egg sample than heat treatment to 70°C, but clearly weaker as all other heat treatments (F = 1, F = 3, F = 12). However, a high pressure induced increasing of antigenicity of beta-lactoglobulin accomplished by ELISA methods with specific monoclonal hen's antibodies could be shown by Illgner [25]. This can be referred to high pressure induced increase of the protein surface but it is not known if these results have clinical relevance. Nevertheless, these different results demonstrate that the effects of high pressure treatment through the allergenic potential of food can be different for diverse allergens in diverse matrices. By the combination of heat and high pressure treatment a higher reduction of the allergenic potential of the sample could be accomplished than caused by the single treatments. The high pressure treatment of the sample that was preheated to 70°C resulted in a 8.9fold stronger reduction in comparison of the untreated sample. Consequently it became clear that the combination of both treatments caused a nearly 2fold stronger reduction of the allergenic potential as the sum of the single treatments conduced separately (1.3fold + 3.3fold = 4.6fold). For all other combinations of heat and high pressure treatments these synergistic effects could not be measured in detail as the C₅₀-values of the EAST inhibition method could not be quantified because of the very low allergenic potential of the samples. To investigate if the order of the technological treatments has an influence of the allergenic potential EAST inhibition curves of samples (10) and (11) (see Figure 2) were compared. Here, no significant differences in the allergenic potential could be shown. Nevertheless, the reduction of the allergenic potential tends to be a little stronger for sample (11) that was at first heat treated and than treated with high pressure.

The sensitivity of the EAST inhibition assay is highly dependent on the used patient's sera. In this case a reduction of the allergenic potential of about 1000fold compared to the native egg extract could still be measured as elucidated in Table 1.

5. Conclusions

The aim of this study was to determine whether heat, high pressure, or a combination of both treatments influences the allergenicity of hen's egg white allergens in meat preparations. Heat treatment with an *F*-value of 3 or 12 brought about a significant decrease in allergenicity measured by EAST inhibition. Heat treatment with 70°C caused a decrease of about 1.5fold in allergenic potential measured with EAST inhibition. High pressure treatment resulted in reductions in IgE-binding properties of more than 3fold. The combined application of heat (70°C) and high pressure produced synergistic effects reducing allergenic potential by nearly

twice as much as the sum of the single treatments conducted separately.

Acknowledgments

This research project was supported by The European Union project REDALL (Reduced Allergenicity of Processed Foods (Containing Animal Allergens), QLK1-CT-2002-02687). The authors would like to thank Knut Brockow and Johannes Ring from the Technical University of Munich (Department of Dermatology and Allergology), Barbara Ballmer-Weber, Manjula Dutta and Barbara Theler from the University Hospital of Zurich (Allergiestation, Dermatologische Klinik), Alessandro Fiocchi, Luigi Terracciano and Teresita Sarratud from the Macedonio Melloni Hospital of Milan (Department of Pediatrics), and Radvan Urbanek, Zsolt Szepfalusi, Veronika Kirchlechner and Eleonora Dehlink from the Medical University of Vienna (Department of Pediatrics and Juvenile Medicine) for providing the human sera. Special thanks go to Bettina Jacobsen for the technical support.

References

- [1] B. Ratner and S. Untracht, "Egg allergy in children; incidence and evaluation in relation to chick-embryo-propagated vaccines," *American Journal of Diseases of Children*, vol. 83, no. 3, pp. 309–316, 1952.
- [2] H. J. Peng, Z. N. Chang, L. C. Tsai, S. N. Su, H. D. Shen, and C. H. Chang, "Heat denaturation of egg-white proteins abrogates the induction of oral tolerance of specific TH2 immune responses in mice," *Scandinavian Journal of Immunology*, vol. 48, pp. 491–496, 1998.
- [3] P. McLaughlan, K. J. Anderson, E. M. Widdowson, and R. R. A. Coombs, "Effect of heat on the anaphylacticsensitising capacity of cow's milk, goat's milk, and various infant formulae fed to guinea-pigs," Archives of Disease in Childhood, vol. 56, pp. 165–171, 1981.
- [4] R. R. A. Coombs and P. McLaughlan, "Allergenicity of food proteins and its possible modification," *Annals of Allergy*, vol. 53, pp. 592–596, 1984.
- [5] Y. H. Lee, "Food-processing approaches to altering allergenic potential of milk-based formula," *Journal of Pediatrics*, vol. 121, pp. 47–50, 1992.
- [6] M. Besler, H. Steinhart, and A. Paschke, "Allergens of animal origin. Stability and allergenicity of processed foods," *Internet Symposium on Food Allergens*, vol. 2, no. 4, pp. 171–184, 2000.
- [7] S. Vieths, A. Jankiewicz, H. Aulepp, and D. Haustein, "Allergy to heated and processed foods," in *Proceedings of the 8th International Paul Ehrlich Seminar*, Gustav Fischer, Jena, Lübeck, Ulm, Stuttgart, Germany, 1997.
- [8] F. C. Stevens and R. E. Feeney, "Chemical modification of avian ovomucoids," *Biochem*, vol. 2, pp. 1346–1352, 1963.
- [9] H. F. Deutsch and J. T. Morton, "Immunochemical properties of heated ovomucoid," *Archives of Biochemistry and Biophysics*, vol. 64, pp. 19–25, 1956.
- [10] T. Matsuda, K. Watanabe, and R. Nakamura, "Immunochemical studies on thermal denaturation of ovomucoid," *Biochimica et Biophysica Acta*, vol. 707, no. 1, pp. 121–128, 1982.

[11] T. Matsuda, K. Watanabe, and R. Nakamura, "Immunochemical and physical properties of peptic-digested ovomucoid," *Journal of Agricultural and Food Chemistry*, vol. 31, no. 5, pp. 942–946, 1983.

- [12] F. Bonomi, S. Iametti, C. Castagna, G. F. Dall'Aglio, and P. P. Rovere, "Proteolysis of bovine β-lactoglobulin and chicken egg ovalbumin during treatment at high pressure," in *Proceedings of the International Conference on High Pressure Bioscience and Technology*, pp. 289–292, Heidelberg, Germany, 1999.
- [13] H. Schöberl, Physikalisch-chemische und strukturelle Auswirkung einer hydrostatischen Hochdruckbehandlung auf Lebensmittel, Dissertation, Technische Universität München, 1999.
- [14] A. Jankiewicz, W. Baltes, K. W. Bögl, et al., "Influence of food processing on the immunochemical stability of celery allergens," *Journal of the Science of Food and Agriculture*, vol. 75, no. 3, pp. 359–370, 1997.
- [15] M. Scheibenzuber, Molekulare und klinische Auswirkungen einer Hochdruckbehandlung von allergenen Lebensmitteln, Dissertation, Technische Universität München, 2002.
- [16] J. Bernhisel-Broadbent, H. M. Dintzis, R. Z. Dintzis, and H. A. Sampson, "Allergenicity and antigenicity of chicken egg ovomucoid (Gal d III) compared with ovalbumin (Gal d I) in children with egg allergy and in mice)," *Journal of Allergy and Clinical Immunology*, vol. 93, pp. 1047–1059, 1994.
- [17] A. Ziermann, Mikrobiologische Kriterien für Milch, Milchprodukte und andere Lebensmittel in Europa, Inaugural-Dissertation, 2005.
- [18] M. M. Bradford, "A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein dye binding," *Analytical Biochemistry*, vol. 72, pp. 248–254, 1976.
- [19] V. Leduc, C. Demeulemester, B. Polack, C. Guizard, L. Le Guern, and G. Peltre, "Immunochemical detection of egg-white antigens and allergens in meat products," *Allergy*, vol. 54, no. 5, pp. 464–472, 1999.
- [20] M. Ceska and U. Lundkvist, "A new and simple radioimmunoassay method for the determination of IgE," *Immuno-chemistry*, vol. 9, no. 10, pp. 1021–1030, 1972.
- [21] Y. Kato, E. Oozawa, and T. Matsuda, "Decrease in antigenic and allergenic potentials of ovomucoid by heating in the presence of wheat flour: dependence on wheat variety and intermolecular disulfide bridges," *Journal of Agricultural and Food Chemistry*, vol. 49, no. 8, pp. 3661–3665, 2001.
- [22] E. J. Jeong, "A study on the allergenicity of egg protein," Han'guk Sikp'um Yongyang Hakhoechi, vol. 11, pp. 228–236, 1998.
- [23] J. Anet, J. F. Back, R. S. Baker, D. Barnett, R. W. Burley, and E. H. Howden, "Allergens in the white and yolk of hen's egg: a study of IgE binding by egg proteins," *International Archives of Allergy and Applied Immunology*, vol. 77, no. 3, pp. 364–371, 1985.
- [24] A. Urisu, H. Ando, Y. Morita, et al., "Allergenic activity of heated and ovomucoid-depleted egg white," *Journal of Allergy and Clinical Immunology*, vol. 100, pp. 171–176, 1997.
- [25] S. Illgner, Einfluss von hydrostatischem druck und temperatur auf die antigenität von beta-Lactoglobulin, Diplomarbeit, Fachhochschule Weihenstephan, 2000.