


Dual-Functional Peptide Driven Liposome Codelivery System for Efficient Treatment of Doxorubicin-Resistant Breast Cancer

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Background: The active-targeted drug delivery systems had attracted more and more attention to efficiently overcome multidrug resistance (MDR) in cancer treatments. The aim of the work was to develop a multifunctional nano-structured liposomal system for co-delivery of doxorubicin hydrochloride (DOX) and celecoxib (CEL) to overcome doxorubicin resistance in breast cancer.

Methods: A functional hybrid peptide (MTS-R₈H₃) with unique cellular penetrability, endo-lysosomal escape and mitochondrial targeting ability was successfully synthesized using solid phase synthesis technology. The peptide modified targeted liposomes (DOX/CEL-MTS-R₈H₃ lipo) for co-delivery of DOX and CEL were formulated to overcome the chemoresistance in MCF/ADR cells.

Results: DOX/CEL-MTS-R₈H₃ lipo showed nanosized shape and displayed high stability for one month. The cytotoxicity effect of the co-delivery of DOX and CEL through peptide modified liposomes had remarkable treatment efficacy on killing MCF/ADR cells. Targeted liposome exhibited greater cellular entry ability about 5.72-fold stronger than DOX solution. Moreover, as compared with unmodified liposomes, the presence of MTS-R₈H₃ peptide entity on liposome surface enhanced the mitochondrial-targeting ability and achieved effective reactive oxygen species (ROS) production with significant inhibition of P-gp efflux activity.

Conclusion: The study suggested that the DOX/CEL-MTS-R₈H₃ lipo is a promising strategy for overcoming drug resistance in breast cancer treatments with high targeting inhibition efficiency.

Keywords: doxorubicin hydrochloride, celecoxib, co-delivery, targeting liposomes, multidrug resistance

Introduction

Cancer remains the major public cause of death around the world and considered the 2nd leading cause of death among American people. The three most public cancers for women are breast, lung, and colorectum, which together represent one-half of all cancer cases. Among them, breast cancer alone accounts for 30% of all new cancer diagnoses in women and around 15% of all deaths.¹ The combination of two or more chemotherapeutic agents with different mechanisms have been proved to be more effective than a single chemotherapy.² As it was well known, the application of drug delivery systems provided numerous benefits as compared with single agents. Among various drug delivery systems, liposomes have attracted

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growing scientific interest because of their abilities to enhance drug bioavailability. The liposomes are attained cancer-targeting and drug resistance reversion ability through the enhanced permeability and retention effect (EPR), surface modifications and endocytosis. Previously reported studies proved that administration of peptide-targeted liposome containing chemotherapeutic drugs showed better synergistic therapeutic effect in the management of metastasis and chemoresistance than individual use of conventional liposome loaded with chemotherapeutic drug.^{3,4}

Doxorubicin hydrochloride (DOX) is the frequently administered chemotherapy for the treatment of several types of cancer. However, cardiotoxicity in clinical rehearsal is the main side effect for DOX application. Moreover, the emergence of multiple drug resistance (MDR) in cancer cells decreases DOX efficiency. Among different MDR mechanisms, the P-glycoprotein overexpression (P-gp), an ATP-dependent efflux pump with wide xenobiotic compounds specificity, to pump drugs out of cells is one of the most popular reasons for MDR.

Besides most of the preclinical and clinical studies presented that COX-2 inhibitors specifically celecoxib (CEL) improves the chemotherapeutic effect of anticancer drugs,⁵⁻⁷ accordingly, it was proved that celecoxib enhances the growth inhibitory and apoptosis effect of doxorubicin in different cancer cell line,⁸⁻¹⁰ Moreover, celecoxib plays an important role in down-regulation of P-glycoprotein overexpression (P-gp),^{11,12} mitochondrial dysfunction and potentiation of ROS production.¹³ Mitochondrion is a vital multifunction organelle necessary for cellular activity. So, alteration of mitochondrial functions (ie depolarization of membrane potential, inhibition ATP synthesis, production of reactive oxygen species and cytochrome c release) is considered as the significant underlying cause of a variety of human diseases including cancer.¹⁴⁻¹⁷ One commonly used approach to achieve effective intracellular entree is to conjugate the therapeutic agent with a cell penetrating peptide (CPP) such as octaarginine (R8) which primarily enter cells through adsorption-mediated endocytosis routes.¹⁸⁻²⁰

In our previous work, we found that co-administration of DOX/CEL with liposomes was better and significantly enhanced the inhibition rate against B16 cells and MGC80-3 cells more than single-drug-loaded liposomes. Furthermore, dermarollers treatment prior to gel application strongly improved the topical anti-tumor efficacy against melanoma xenograft mice model.²¹ Despite the

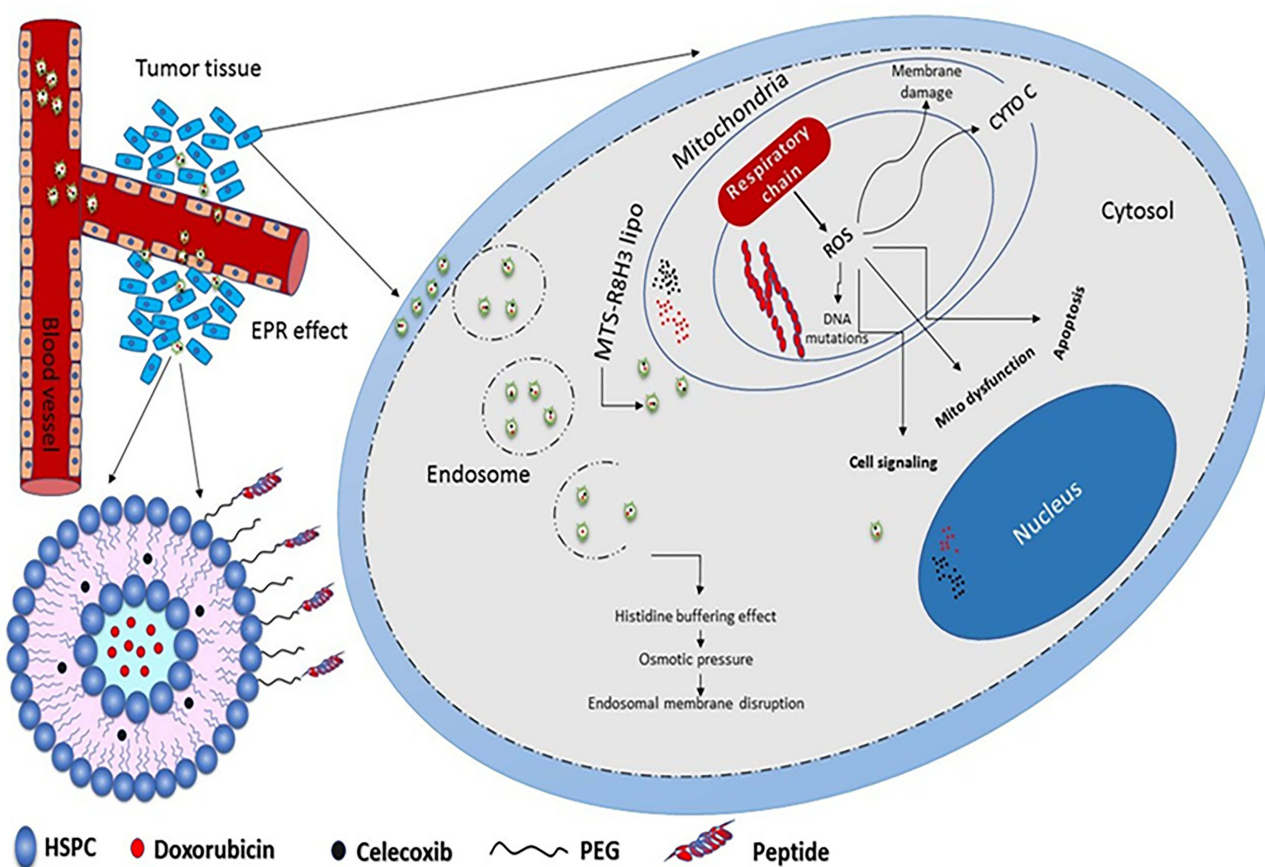
established connections between conventional DOX/CEL liposomes treatment and melanoma cell death and apoptosis, it has yet to be determined whether the co-drug loaded liposomes can improve the therapeutic efficacy in drug-resistance breast cancer treatment by further promoting the targeting ability of liposomes.

The aim of the present study was to develop a multifunctional nano-structured liposomal system with ideal biocompatibility and biodegradability for co-delivery of DOX and CEL to overcome several challenges, including side effects of DOX, lower solubility of CEL, lack of specificity, limited stability, and a high capacity for disparate cargos. Furthermore, our main purpose was to get and enhance the therapeutic benefits resulted from the synergistic effect of DOX and CEL against breast cancer. To realize this potential, as shown in [Scheme 1](#), peptide (MTS-R₈H₃) targeted liposomes with improved cell internalization, lysosomal escape and active mitochondrial targeting ability for multiple drug delivery were designed and evaluated.

Materials and Methods

Materials

All amino acids were purchased from A Peptide Co., Ltd (Shanghai, China). Hydrogenated soya bean phosphatidylcholine (HSPC), 1,2-distearoyl-sn-glycero-3-phosphoethanolamine-N- [carboxy (polyethylene glycol)-2000] (sodium salt) DSPE-PEG (2000) were kindly gifted from Lipoid GmbH. Celecoxib (CEL) was supplied by energy chemical company. Doxorubicin hydrochloride (DOX) was supplied by Beijing Huafeng United Technology Co. Ltd. (Beijing, China). N-hydroxysuccinimide (NHS), 1-ethyl-3-(3-dimethyl aminopropyl) carbodiimide (EDC), and Cholesterol (CH) were purchased from Aladdin Regent Co. (Shanghai, China). RPMI 1640 medium, fetal bovine serum (FBS), penicillin-streptomycin solution, and 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) were obtained from Life Technologies™. 5,5',6,6'-tetrachloro-1,1',3,3'-tetraethylbenzimidazolcarbocyanine iodide (JC-1) and 4',6-diamidino-2-phenylindole (DAPI) were supplied by Shanghai Beyotime Biotechnology Co. Ltd. (Suzhou, China). MCF/ADR human breast cancer cells were bought from the Type Culture Collection of the Chinese Academy of Sciences (Shanghai, China). Chloroform and all other chemicals were of analytical grade.



Scheme 1 A schematic illustration of the pathway of DOX/CEL-loaded targeting liposomes, targeted liposomes were taken up into tumor cells by endocytosis pathway and escaped from lysosomes into mitochondria and nucleus. Peptide: MTS-R₈H₃.

Abbreviation: Cyto C, Cytochrome C.

Cell Cultures

MCF/ADR human breast cancer cells were grown in RPMI 1640 medium containing 1 µg/mL doxorubicin HCl, 10% FBS, 100 U/mL penicillin and 100 mg/mL streptomycin. The cells were cultured at 37 °C and 5% CO₂ in a humidified incubator.

Synthesis of the Multifunctional MTS-R₈H₃ Peptide

A functional hybrid peptide conjugation approach that incorporates CPP, histidines, and MTS into one molecule with a sequence of MLRAALSTARRGPRLSRLLHHH RRRRRRRR was employed to improve the cellular penetrability, endo-lysosomal escape and mitochondrial targeting efficacy in human MCF/ADR cells. All amino acids conjugates were prepared using standard 9-fluorenylmethoxycarbonyl (Fmoc) solid phase synthesis approaches. The amino acids, Fmoc-Arg (Pbf)-OH, Fmoc-Gly-OH, Fmoc-Met-OH, Fmoc-Leu-OH, Fmoc-Ala-OH, Fmoc-Ser (But)-OH, Fmoc-Pro-OH, Fmoc-Thr (But)-OH,

Fmoc-His (Trt)-OH, etc. added in a reaction tube containing Wang resin with a degree of substitution of 1.03 mmol/g. Fmoc deprotections were carried out using a 20% piperidine DMF solution (15 mL/g) for 5 min. Acetylation was performed on α-amino groups of N-terminus amino acids using a 20% acetic anhydride in DMF (N, N-dimethylformamide) solution with 100 µL DIEA (diisopropylethylamine). After pumping out the piperidine solution, some resins were taken and washed three times with ethanol, followed by addition of detection reagents (Phenol reagent, pyridine reagent, ninhydrin reagent) to monitor the reaction. The resin was washed twice using DMF (10 mL/g), methanol (10 mL/g), dichloromethane (10 mL/g), and drained for 10 min. Completed peptides were separated from the resin using the cleavage cocktail (95% trifluoroacetic acid; 1% water; 2% ethanedithiol and 2% triisopropylsilane) for 120min. Finally, the lysate was dried with nitrogen, washed six times with diethyl ether, and then evaporated at room temperature. The crude product was purified by high-performance liquid

chromatography (HPLC, Shimadzu, Japan) and then lyophilized to obtain the MTS-R₈H₃ peptide. The peptide was confirmed using HPLC and mass spectrum (MS, Waters, USA).

Synthesis of DSPE-PEG2000-MTS-R₈H₃ Conjugates

DSPE-PEG2000-MTS-R₈H₃ conjugate was synthesized and purified according to an established procedure with some modifications.³ Firstly, DSPE-PEG2000 was allowed to react with NHS/EDC in DMF for 4h in an ice bath with gentle stirring to activate the carboxylic group of DSPE-PEG2000. The peptide was added to the above solution in an appropriate molar ratio (DSPE-PEG2000: NHS: EDC: peptide is 2:4:4:1) and stirred for 24 h at room temperature under nitrogen protection. The resulting solution was dialyzed in deionized water with a cellulose ester membrane filter (MWCO=3500 Da) for 48 h to remove un-reacted species. Finally, the dialysate was concentrated and lyophilized to obtain DSPE-PEG2000- MTS-R₈H₃. The conjugate was collected and identified by Fourier transform infrared spectra (Bruker Instruments, Beijing, China), and ¹H NMR spectrometry (AVANCE III, 400 MHz, Bruker, Germany).

Preparation and Characterization of the Liposomes

The liposomes were formulated according to the widely used thin film hydration method and post-insertion technique.^{23,24} The lipid phase was composed of hydrogenated soya bean phosphatidylcholine 60 mmol and cholesterol with HSPC/CH molar ratio 1:1. These components were dissolved in chloroform in a round-bottom flask together with an appropriate amount of CEL. Chloroform was slowly evaporated using a rotary evaporator under vacuum forming a thin lipid film on the flask wall that was further dried in a vacuum chamber overnight. The lipid film was then hydrated by ammonium sulfate solution to form multilamellar vesicle suspension that was further subjected to ultrasonication using a probe sonicator for 10 min at 40% (JY92-IIN, Ningbo Scientz Biotechnology Co., Ltd., China) to prepare CEL loading liposomes (CEL lipo). Based on the above procedure, the pH gradient system of the liposomes was constructed by removing untrapped ammonium sulfate through dialysis against a buffer solution. DOX was loaded by mixing the drug solution at 60 °C for 15 min with the liposome's

suspension. Free DOX was separated by the dialysis method to prepare DOX/CEL loading liposomes (DOX/CEL lipo). DOX-loaded liposomes (DOX lipo) were prepared using the above method, but without adding CEL to the lipid components.

For preparing of the peptide-modified DOX/CEL-MTS-R₈H₃ liposomes (targeting lipo), blank liposomes composed of HSPC and CH were first prepared, and the procedure was the same as described above. DSPE-PEG2000-MTS-R₈H₃ at 4 mol % of HSPC was then added into the preformed liposome suspension under stirring for 1 h at 60 °C in the dark.²⁵ The mixture was then dialyzed against phosphate buffer using semi-permeable dialysis membrane (MWCO 14000 Da) for 4 h to remove unconjugated ligands.

The particle size and zeta potential of the liposomes were measured by dynamic light scattering (DLS) using a Zetasizer Nano ZS apparatus (Malvern Instruments, UK). The morphology of the liposomes was determined by transmission electron microscopy (TEM) on a JEM-2100 (JEOL, Japan).

The encapsulation efficiency was tested by ultraviolet spectrophotometry. The unencapsulated DOX and CEL were removed by means of ultrafiltration using centrifugal filter tubes with an Mwt cut-off of (3000 MWCO) (Millipore, Suzhou, China). Liposome suspension was placed into the filter tubes and centrifuged at 5000 rpm and 4 °C for 20 min. The drug solution collected at the bottom of the filter tube was analyzed by ultraviolet spectrophotometric assay (UV-2550, Shimadzu, Japan) to determine DOX and CEL content at 486 nm and 256 nm respectively.

In vitro Drug Release Study

In vitro DOX and CEL released from liposomes was determined using a semi-permeable dialysis membrane (MWCO 14000 Da). The dialysis membrane was sited between the donor and receptor compartments of modified Franz diffusion cells. After separating unencapsulated drugs, 1mL of liposome formulations was applied on the dialysis membrane surface. The receptor compartment was filled with 20 mL fresh PBS buffer medium (pH 7.4) containing ammonium sulfate²⁶ and 1% sodium dodecyl sulfate²⁷ at 37 °C and 90 rpm. At predetermined time intervals, 2 mL of the sample was withdrawn from the receptor medium and substituted with the same amount of buffer phosphate to keep perfect sink conditions. The drug concentrations were measured by ultraviolet

spectrophotometric assay at 486 nm and 256 nm, respectively. The results were the average of the three runs, and the cumulative percent released of each drug was plotted at a certain time.

Assessment of Liposome Stability

Different drug-loaded liposomes made from hydrogenated soya bean phosphatidylcholine formulated under the same conditions were stored at 4 °C for 1 month. Liposomal size, PDI and entrapment efficiency were the parameters chosen to indicate the physical stability of liposomes.

Cytotoxicity Studies

The cytotoxicity of different liposomal formulations against MCF/ADR cells was evaluated by MTT assay. MCF/ADR cells were seeded in 96-well plate at a density of 8×10^3 cells per well, and the cells were adhered by overnight culture. The cell culture medium was discarded and replaced with fresh medium containing different drug formulations. For the cytotoxic effect of empty liposome and empty targeted liposome, 100 μL of 0.3 mmol/mL (correspond to the highest lipid concentration used in drug-loaded liposomes for viability test) was added to one set of 96-well plate to incubate with MCF/ADR cells. After 48 hours of incubation, 100 μL of MTT solution was added. MTT was removed after 4 h incubation. Finally, 100 μL of DMSO was added to dissolve the formazan blue-violet crystals, formed by the living cells, before measurement at 580 nm with a microplate reader (Multiskan MK3, Thermo, USA).

Cellular Uptake Analysis

Cellular uptake of free DOX, DOX lipo, DOX/CEL lipo, and targeting lipo by MCF/ADR cells was studied by qualitative fluorescence imaging and quantitative flow cytometry. The cells were seeded into a laser confocal dish at a density of 8×10^4 cells per well and cultured overnight for adherence. After that, the culture medium was replaced with different drug formulations with a DOX and CEL concentrations of 5 and 20 $\mu\text{g} \cdot \text{mL}^{-1}$ respectively and then the cells were incubated for 1 h, 2 h, and 4 h. After being washed with phosphate-buffered saline (PBS), 200 μL of 4% paraformaldehyde was added into each well to fix cells for 20 min. Then the nuclei were stained with DAPI (20 $\mu\text{g} \cdot \text{mL}^{-1}$) for 15 min, followed by

washing the cells three times using PBS. Finally, 500 μL of PBS was added to each well. The cell uptake was observed using confocal laser scanning microscopy (CLSM, Nikon A1R, Japan).

For flow cytometry analysis, MCF/ADR cells were seeded in 6-well plates at 2×10^5 cells per well, and the cells were adhered by overnight incubation at 37°C and 5% CO_2 . Then the medium was replaced with different formulations with DOX and CEL concentrations of 5 and 20 $\mu\text{g} \cdot \text{mL}^{-1}$ respectively. The plates were incubated for 1 h, 2 h, and 4 h. Followed by washing the cells three times with PBS, 200 μL of trypsin enzyme was added to each well. After being digested, the cells were collected and centrifuged. The resulting cells were suspended in 500 μL of PBS and the fluorescence intensity was evaluated by flow cytometry (Becton Dickinson FACSCalibur, USA).

Subcellular Localization of the Liposomes

The distribution of different liposomal formulations in MCF/ADR tumor cells was observed by confocal laser scanning microscopy (CLSM, Leica, TCS SP8, Germany). MCF/ADR cells were seeded onto a glass bottom cell culture dish (ϕ 15 mm) at 20×10^5 cells per well and cultured overnight for adherence. Then, the culture medium was removed, and preparations were added with DOX and CEL concentrations of 5 and 20 $\mu\text{g} \cdot \text{mL}^{-1}$ respectively. The culture plates were incubated for 1 h, 2 h, and 4 h, and the cells were washed with PBS and stained by lysotracker green (50 nm) for 30 min and DAPI for 20 min to visualize the endo-lysosomes and the nucleus respectively, followed by addition of 200 μL of 4% paraformaldehyde into each well to fix cells. Finally, the plates washed three times using PBS and then observed by confocal laser scanning microscopy (CLSM, Nikon A1R, Japan).

For further tracking the internalization and the mitochondrial targeting ability of different liposomal formulations in MCF/ADR tumor cells, the cells were treated with liposomal preparations at a concentration of 5 $\mu\text{g} \cdot \text{mL}^{-1}$ and 20 $\mu\text{g} \cdot \text{mL}^{-1}$ for DOX and CEL, respectively. After incubation for 1 h, 2 h, and 4 h, the cells were rinsed with cold PBS twice, and mitochondrion staining was performed with Mitotracker Green for 30 min at 37 °C. Then, the cells were counterstained with DAPI (blue fluorescence) for 20 min to visualize the cellular nuclei and observed by CLSM.

Mitochondrial Membrane Potential Assay (MMP)

Mitochondrial membrane-permeable lipophilic cationic fluorochromes are applied as probes of $\Delta\psi$, they permeate cells and their fluorescence reflects the change in MMP.²⁸ Most of the recently published literature using 5,5',6,6'-tetrachloro-1,1',3,3'-tetraethyl benzimidazole carbocyanine iodide (JC-1) to evaluate $\Delta\psi$,^{29,30} JC-1 is a lipophilic fluorochrome that is used to assess the status of the $\Delta\psi$, it can exist in two different states, monomers green (~525 nm) or aggregates red (~590 nm), each with a different emission spectrum. MCF-7/ADR cells were cultured in 6-well plates (1×10^6 cell/well) and incubated for 24 h, and then the cells were incubated with different drug formulations in 5% CO₂ incubator at 37°C for 4 h. after incubation time the cells were washed three times and stained with JC-1 working solution for another 30 min. Finally, the cells were collected, centrifuged and washed thrice with PBS. MMP was measured as the ratio of red/green fluorescence intensity. The fluorescence of green (emission=538 nm) and red (emission=590 nm) was analyzed by flow cytometry (Becton Dickinson FACSCalibur, USA).

Production of Reactive Oxygen Species (ROS)

Dichlorofluorescein diacetate (DCFH-DA) assay was performed to measure ROS generation.^{13,31} DCFH-DA can readily enter cells and be cleaved by esterase to yield DCFH, nonfluorescent polar product. ROS in cells promote the oxidation of DCFH to yield the fluorescent product dichlorofluorescein. MCF/ADR cells were incubated in a six-well plate and exposed to different formulations for 1h, 2h, and 4h. After the pretreatment period, cells were incubated with DCFH-DA at 37 °C in the culture medium for 30 min. After incubation, the cells were washed by PBS three times. Each well was digested by 200 μ L of trypsin enzyme, re-suspended in 500 μ L PBS, and immediately submitted to flow analysis using a flow cytometer (Becton Dickinson FACSCalibur, USA).

Analysis of P-gp Efflux Activity

The intracellular rhodamine 123 accumulation was taken as an index to evaluate the efflux pump activity of P-gp.³² MCF/ADR cells were cultured in six-well plates at initial densities of 5×10^6 /well and incubated for 24 h. Then, cells were treated with different DOX formulations for 12 and

24 h, after incubation time, the medium was discarded, and the cells rinsed three times with PBS. Cells were incubated with Rh 123 (5 μ g/mL) for 30 min in darkness at 37 °C. Finally, cells were washed twice with PBS. The cells were digested, centrifuged, and suspended with 1mL PBS. The fluorescence as an indicator for intracellular Rh123 accumulation was analyzed by flow cytometry (Becton Dickinson FACSCalibur, USA).

Statistical Analysis

The measurements were repeated at least three times. All reported values present mean values \pm standard deviation. The data were subjected to analysis of variance using one-way ANOVA (SPSS 17.0, Chicago, IL, USA), a value of $P < 0.05$ was considered as a minimal level of significance.

Results and Discussion

Synthesis and Characterization of MTS-R₈H₃ and DSPE-PEG-MTS-R₈H₃

Solid phase synthesis approach was used to prepare MTS-R₈H₃ peptide. The purity of the MTS-R₈H₃ conjugate was above 95% according to analytical HPLC analysis ([Supplementary Materials Figure S1](#)). The multiply charged ions of ESI-MS showed that the mass of the conjugate was 3799.57 Da ([Supplementary Materials Figure S2](#)), which was agreed with the calculated exact masses of the peptide.

DSPE-PEG-MTS-R₈H₃ was produced via a nucleophilic substitution reaction, in which the terminal amino group on MTS-R₈H₃ peptides reacted with the carboxylic group of DSPE-PEG in anhydrous DMF ([Figure 1](#)). The resulting product was characterized by ¹H NMR spectra and IR. As shown in [Figure 2A](#), the peaks at 0.843 ppm and 1.226 ppm were belonged to methyl and methylene protons in DSPE, while the peaks at 3.509 ppm and 6.5–8.5 ppm were attributed to PEG and MTS-R₈H₃, respectively. In the Fourier transform infrared spectrum ([Figure 2B](#)), the peak at 1106 cm⁻¹ was the stretching vibration of C–O–C in the PEG segment and the peak at 1645 cm⁻¹ was the stretching vibration of C(=O)–N in MTS-R₈H₃. All the data confirmed that the conjugation reaction successfully proceeded.

Characterization of DOX/CEL Liposomes

Drug-loaded targeting liposomes were formulated using thin film hydration method with pH gradient and post insertion techniques. Liposomes were prepared in the

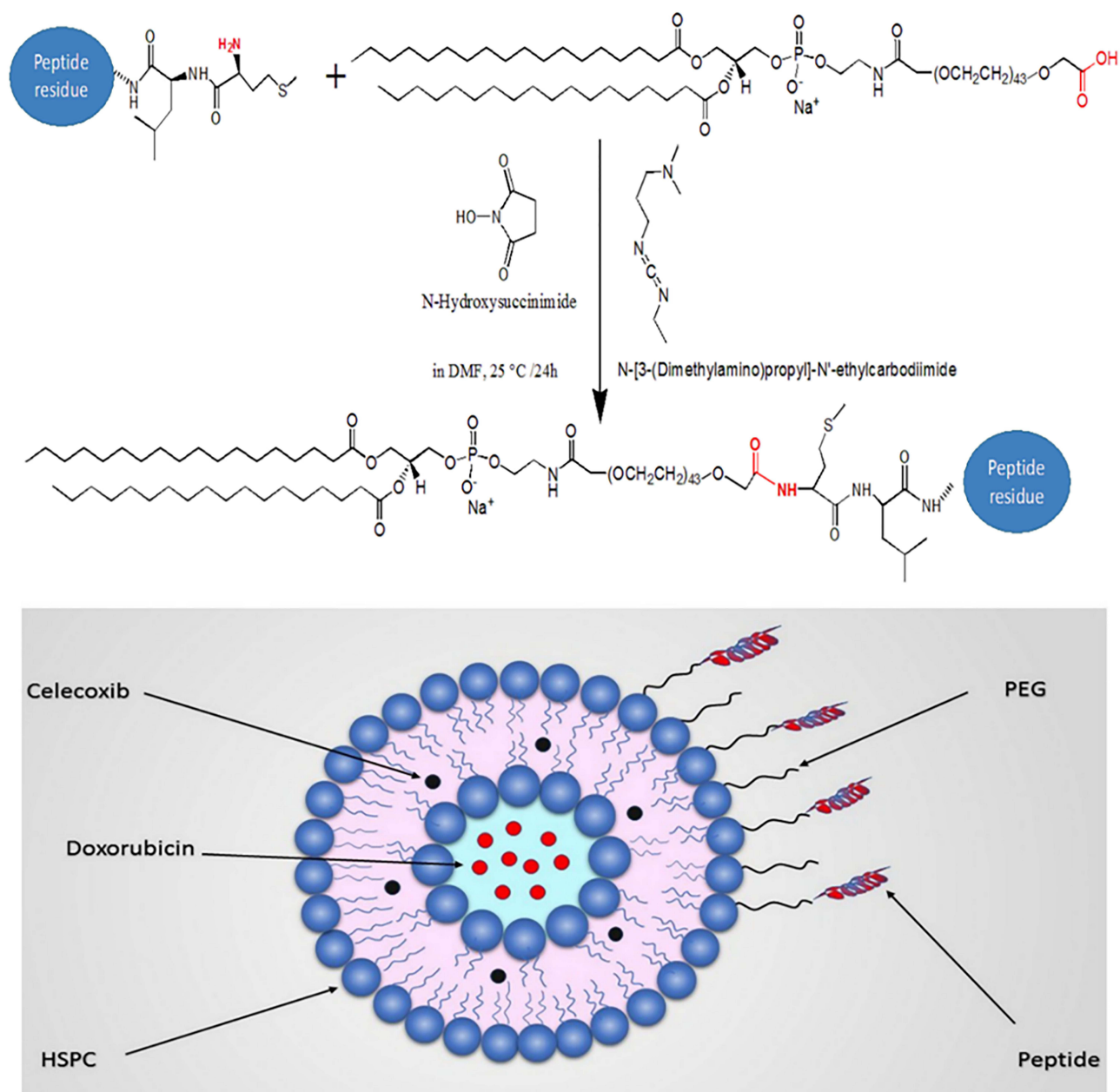


Figure 1 Synthesis scheme of peptide (MTS-R₈H₃) and the illustration of liposome composition.

presence of DSPE-PEG2000 to promote conjugation of MTS-R₈H₃ peptides to liposomal surfaces as an active targeting ligand. The average diameter for all liposomes was approximately similar, in a range of 100–150 nm, and had a highly homogeneous distribution with a PDI less than 0.2 (Table 1). Nanoparticles with such homogeneity have been reported to endow the liposomal delivery system for higher therapeutic effect through EPR effect in vivo.^{33,34} Conjugation of MTS-R₈H₃ peptides increased the vesicle size to approximately 176.16 ± 1.68 nm, indicating the presence of targeting ligands on the

liposome's surfaces. Particle sizes of the targeting liposome were further investigated by TEM imaging. Liposomes displayed nanosized, spherical morphologies and uniform distribution on TEM grids (Figure 3A). Table 1 lists the measured particle size, PDI, zeta potential and entrapment efficiency of liposomes. In all preparations, the encapsulation efficiencies of doxorubicin and celecoxib were approximate >96% consistent with the previously published results about the liposomal formulation of doxorubicin hydrochloride with Palm Oil (EE % about 99%).³⁵

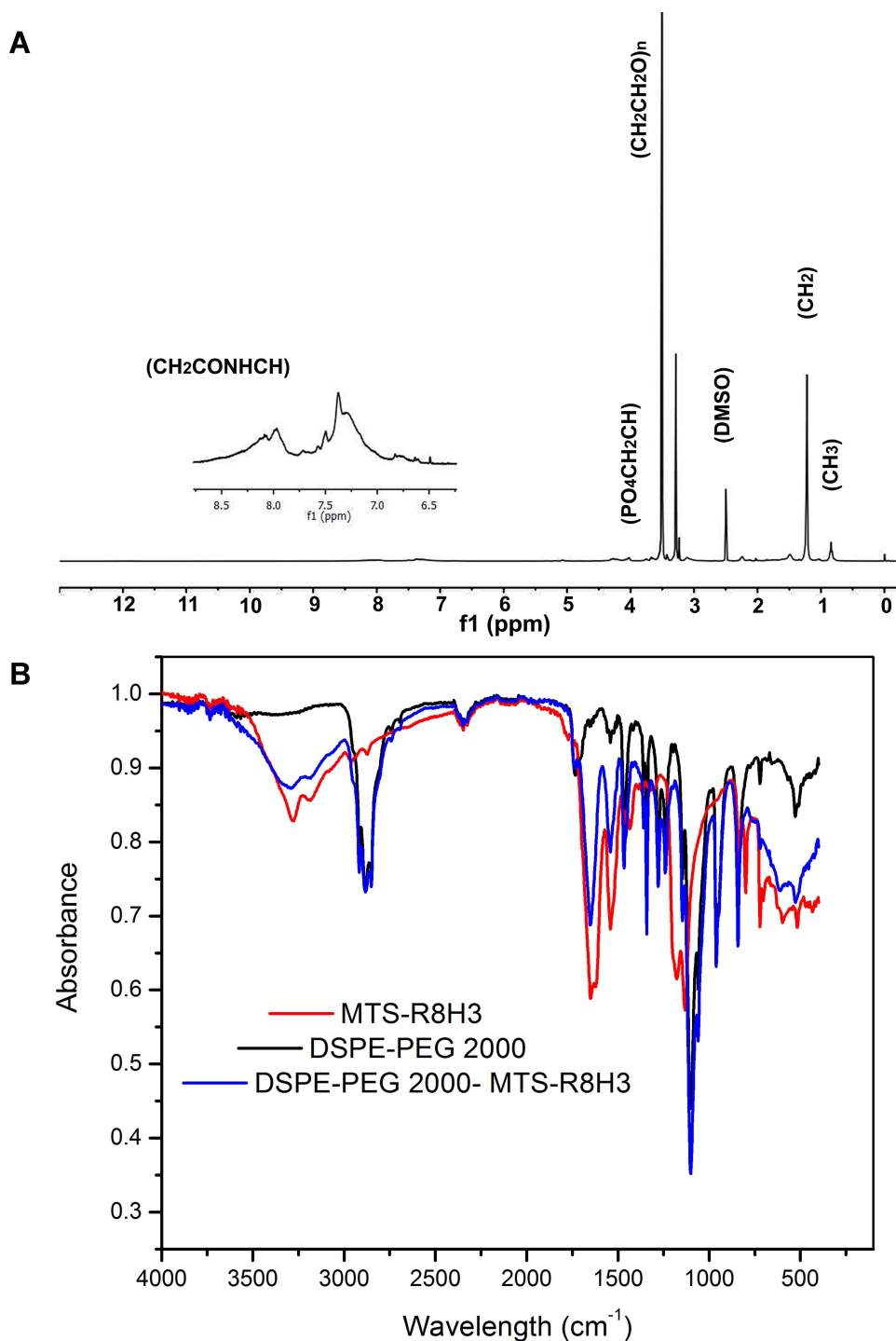


Figure 2 $^1\text{H-NMR}$ spectroscopy (A) and Fourier transform infrared spectra (B) of DSPE-PEG 2000, MTS-R₈H₃, and DSPE-PEG 2000- MTS-R₈H₃.

Drug-Release Studies

In vitro drug release study was performed to evaluate the liposomal drug release property in PBS (pH 7.4) containing 1% w/v sodium lauryl sulfate. As shown in Figure 3C, not a large burst release was observed initially, and only about 40% of total DOX and CEL were released in the first 12 h,

followed by a lower drug release phase within the later 72 h. Presence of ammonium sulfate in the receptor medium enhanced the release of DOX from the internal part of the liposomal vesicle.³⁶ Drug release from the targeted liposomal formulation displayed similar and sustained release manners without significant initial burst release.

Table I The Size, PDI, Zeta-Potential and Encapsulation Efficiency of Different Liposomes. Values are Means \pm Standard Deviation for Three Independent Experiments

| Drug Formulations | Encapsulation Efficiency % | | Particle Size (nm) | PDI | Zeta Potential |
|-------------------|----------------------------|---------------------|--------------------|-------------------|------------------|
| | Doxorubicin | Celecoxib | | | |
| DOX lipo | 99.2% \pm 0.003 | | 112.43 \pm 3.48 | 0.127 \pm 0.016 | -2.24 \pm 0.43 |
| DOX/CEL lipo | 98.37% \pm 0.037 | 99.69% \pm 0.0014 | 141.6 \pm 4.015 | 0.179 \pm 0.014 | -3.15 \pm 0.09 |
| Targeting lipo | 96.37% \pm 0.316 | 97.69% \pm 0.158 | 176.16 \pm 1.68 | 0.193 \pm 0.001 | -4.01 \pm 0.81 |

Stability Study

Physical stability is a critical issue that must be considered during preparation design and development. The liposomal physical instability might be apparent after an increase in vesicle size as a result of the fusion or aggregation of unstable particles during preparation processing or upon storage for a long time. Accordingly, this instability leads to short half-life due to the quick uptake by the reticuloendothelial system, resulting in rapid body clearance and consequently decreased therapeutic efficacy of the loaded drug. So, small and uniform liposomal size preparation is the most vital factor for developing useful and effective pharmaceutical products.³⁷

The stability of liposomes was evaluated at 4 °C and protected from direct light for one month. The data showed that liposomes did not display significant changes in particle size and PDI with a little change in the entrapment efficiency of both drugs (Figure 3B and Table 2). This long-term stability resulted from the incorporation of saturated phospholipids and cholesterol that lead to decrease in the lipid bilayer defect, improve the lateral packing of the acyl chains and consequently, minimizing leakage of liposomal contents.³⁸

Cytotoxicity Studies of Liposomes

MTT assay was applied to evaluate the cytostatic efficacy of DOX and CEL on MCF/ADR cells. Tumor cells were exhibited to the liposomal preparation of CEL, DOX, DOX/CEL and targeted liposome for 24 h. As shown in Figure 4, it was found that the liposomal formulations of each drug alone or in combination could significantly inhibit the proliferation in a dose-dependent manner ($P < 0.05$).

The IC_{50} values of DOX and CEL on MCF/ADR cells were evaluated, respectively. As shown in Table S1, the IC_{50} of DOX and CEL as a single drug-loaded liposome showed the highest concentration to kill 50% of MCF/ADR cells, while the co-delivery and peptide modified targeting liposomes only reached approximately quarter

of the concentration needed from each drug alone. Besides, the targeting liposomes group showed a significant decrease in the minimum inhibitory concentrations in comparison with other groups. This indicated that the cytotoxicity effect of the co-delivery of DOX and CEL through peptide modified liposomes had remarkable treatment efficacy on killing MCF/ADR cells.

Cellular Uptake

Firstly, we used flow cytometry to study the cellular uptake efficiency of the four designed DOX formulations: DOX solu, DOX lipo, DOX/CEL lipo, and targeting lipo. Figure 5 reveals obviously that the targeted lipo showed greater cellular entry ability in comparison to other tested formulations, as proved by its strongest intensity. The mean fluorescence intensity for targeting liposome was about 1.41, 2.74, and 5.72-fold stronger than that for DOX/CEL lipo, DOX lipo, and DOX solu respectively after 4 h incubation, showing a highly significant difference ($P < 0.001$). The observation of the higher fluorescence signal for targeted liposome is due to the unique peptide structure composed from MTS (mitochondrial targeting signal), polyarginine (R_8), and three histidine (H_3). It was reported that the presence of octaarginine had a remarkable capability to assist in cellular internalization.^{39,40}

Assumed that flow cytometry only refers to the overall fluorescence intensity, we did confocal imaging to investigate further the intracellular distribution of the liposomes. As shown in Figure S3, DOX was mainly concentrated in the cytoplasm after incubation for 1 h, but more drugs were entered into the nucleus in DOX-loaded liposome groups. For all groups, the fluorescence intensity of DOX increased after co-incubated for 2 h and 4 h. It was worth observing that the DOX of liposome groups remained in the cytoplasm after co-incubation for 4 h, which indicated that the drugs were released in a controlled manner from the liposomes. And it was not difficult to see that cells incubated with targeted liposomes

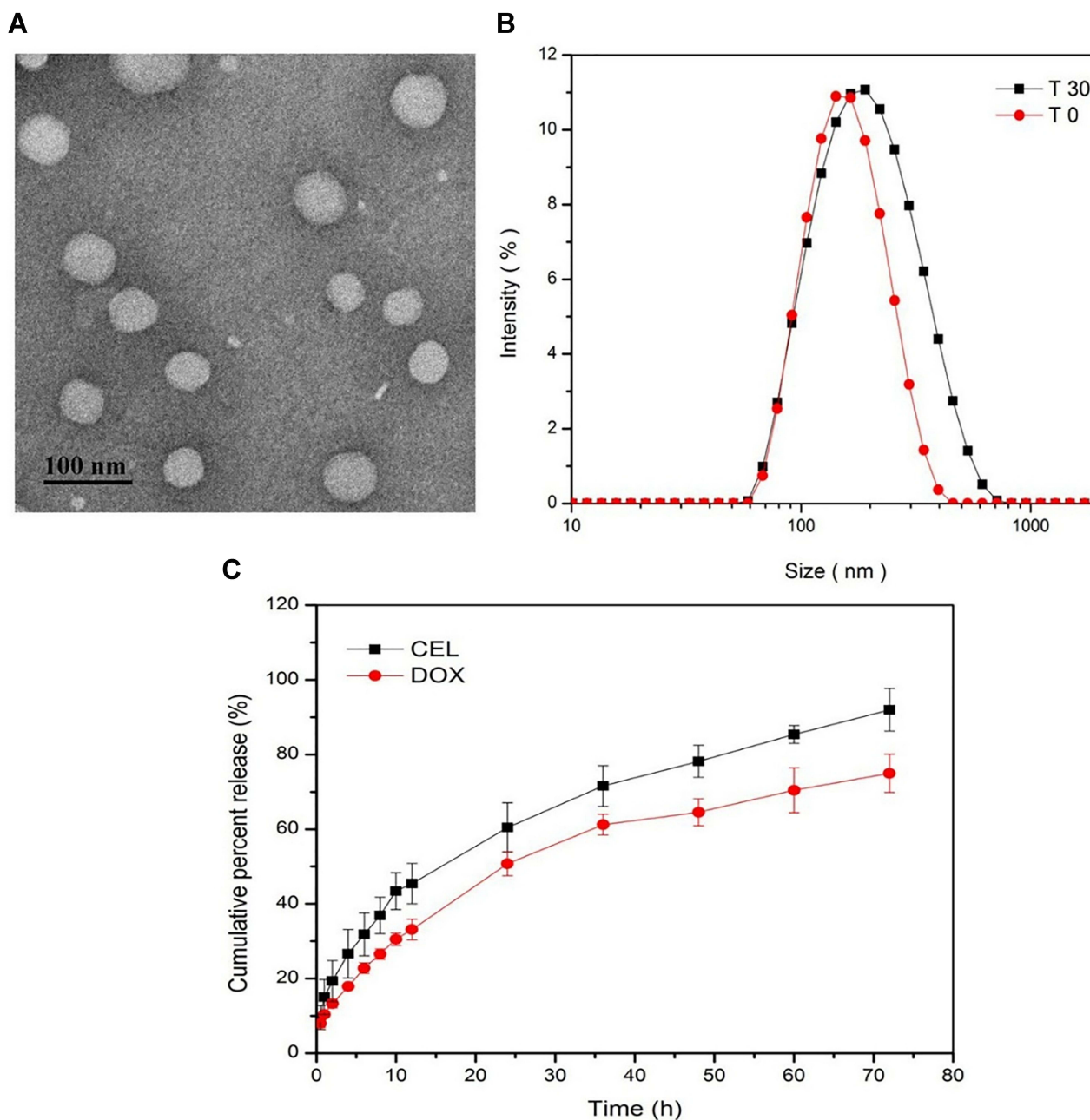


Figure 3 (A) Transmission electron micrographs of targeted liposome. (B) Size distribution of targeted liposome at preparation time (T0) and after one month of storage at 4 °C (T2), and (C) In vitro drug release profiles in phosphate buffer over 72 h at 37 °C. (C) The release profile of DOX and CEL from the liposome suspension. The results are represented as means \pm SD.

showed intense red fluorescence in the cytosol, suggesting that it had superior intracellular access and more drugs were entered into the nucleus with time. With the data obtained from flow cytometry and fluorescence microscopy, it possibly concluded that the presence of MTS-R₈H₃ peptide on the liposome surface enhanced the

cell-specific recognition ability and achieved effective cellular internalization.

Subcellular Localization

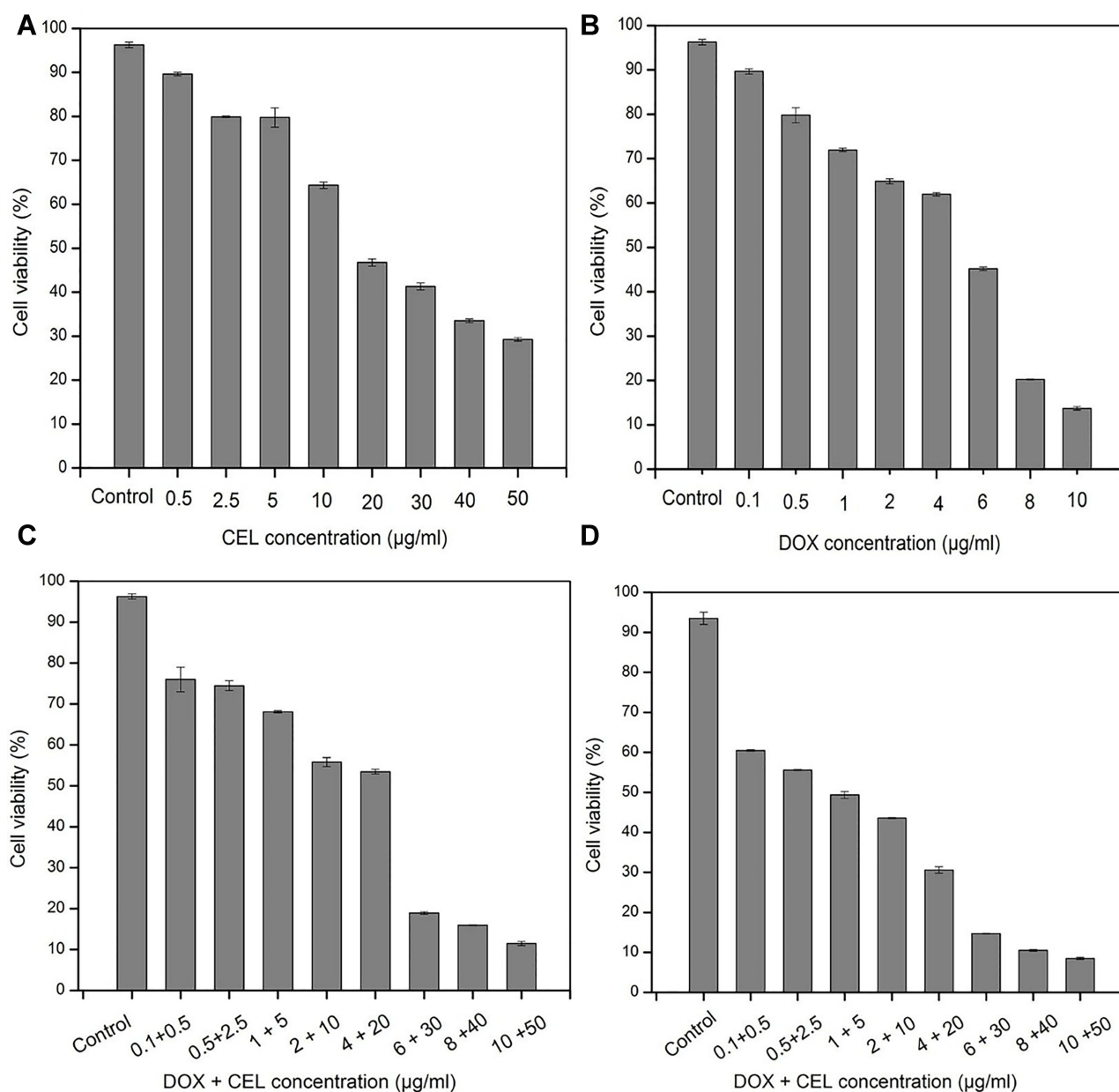
The subcellular localization of different formulations in MCF/ADR cells was assessed by CLSM at different times in order to prove if the peptide modified

Table 2 Stability Study and Characteristics of Loaded Liposomes After 1 Months of Storage at 4 °C. Values are Means \pm Standard Deviation for Three Independent Experiments

| Drug Formulations | Encapsulation Efficiency % | | Particle Size (nm) | PDI | Zeta Potential |
|-------------------|----------------------------|--------------------|--------------------|------------------|-------------------|
| | Doxorubicin | Celecoxib | | | |
| DOX lipo | 98.35% \pm 0.076 | 97.1% \pm 0.0834 | 104.03 \pm 1.4 | 0.124 \pm 1.4 | -1.7 \pm 0.2 |
| DOX/CEL lipo | 96.62% \pm 0.0516 | 95.4% \pm 0.321 | 151.93 \pm 2.18 | 0.28 \pm 0.02 | -2.46 \pm 0.11 |
| Targeting lipo | 95.73% \pm 0.64 | | 191.13 \pm 3.9 | 0.35 \pm 0.004 | -4.57 \pm 0.134 |

liposomes were still capable of escaping the lysosomes and targeting the mitochondria. As shown in Figure 6, after the cells treated with DOX solution and DOX

liposomes, the red fluorescence was increased in the cytoplasm following 2 h incubation. After a prolonged incubation of 4 h, it was observed that strong yellow

**Figure 4** Cytotoxicity against MCF/ADR human breast cancer cells treated with CEL lipo (A), DOX lipo (B), DOX/CEL lipo (C) and targeting lipo (D). Data are reported as the mean \pm standard deviation (n = 3).

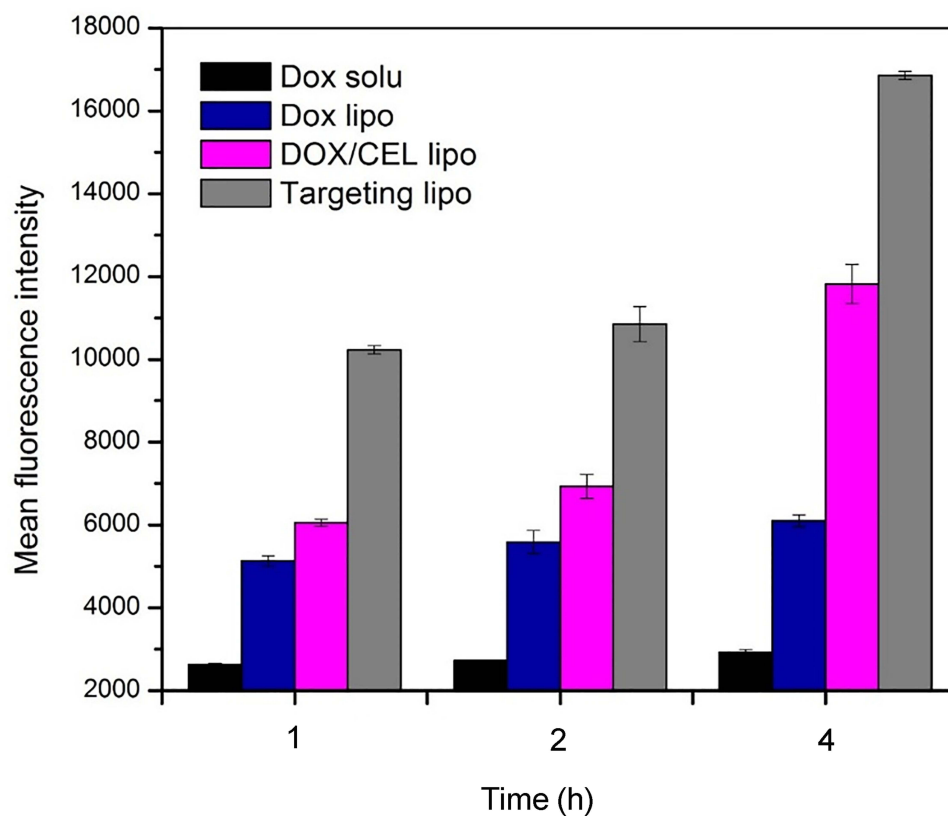


Figure 5 Time-dependent cell uptake of different DOX formulations in MCF/ADR human breast cancer cells. Data are reported as the mean \pm standard deviation ($n = 3$).

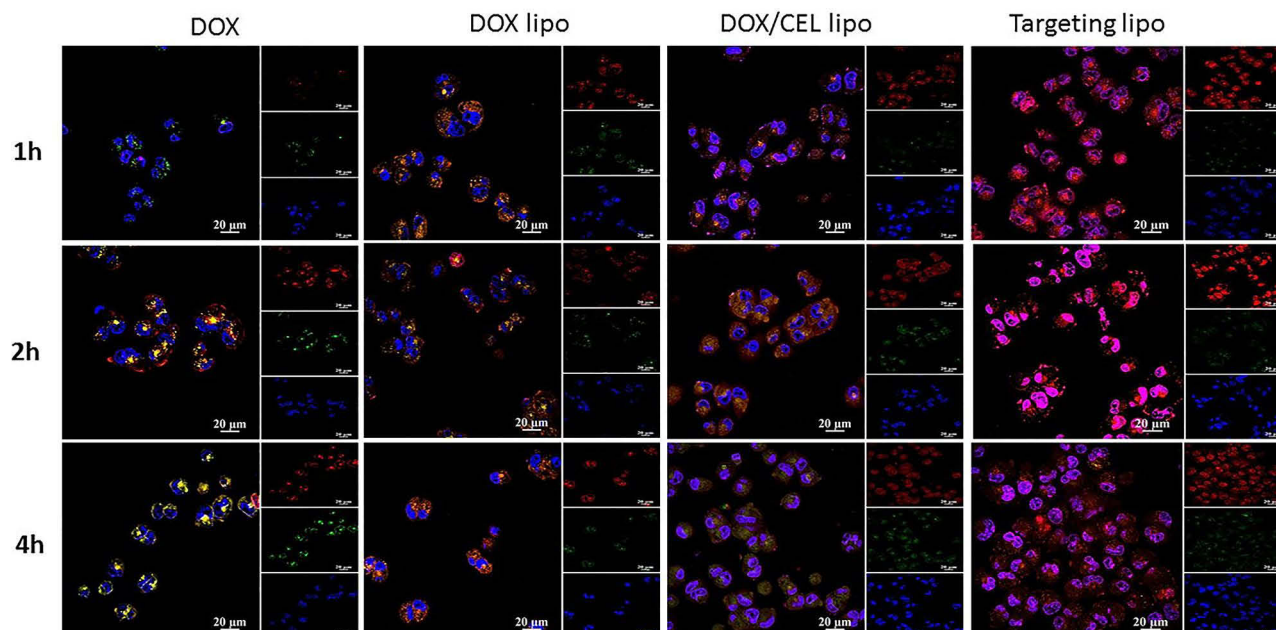


Figure 6 CLSM images of MCF/ADR human breast cancer cells after incubation at different times with DOX, DOX lipo, DOX/CEL lipo, and targeting lipo. DOX was visualized as the red fluorescence, nuclei labeled with DAPI were shown as the blue fluorescence and green fluorescence indicates lysotracker green labeled-mitochondria. Scale bars correspond to 20 μm in all images.

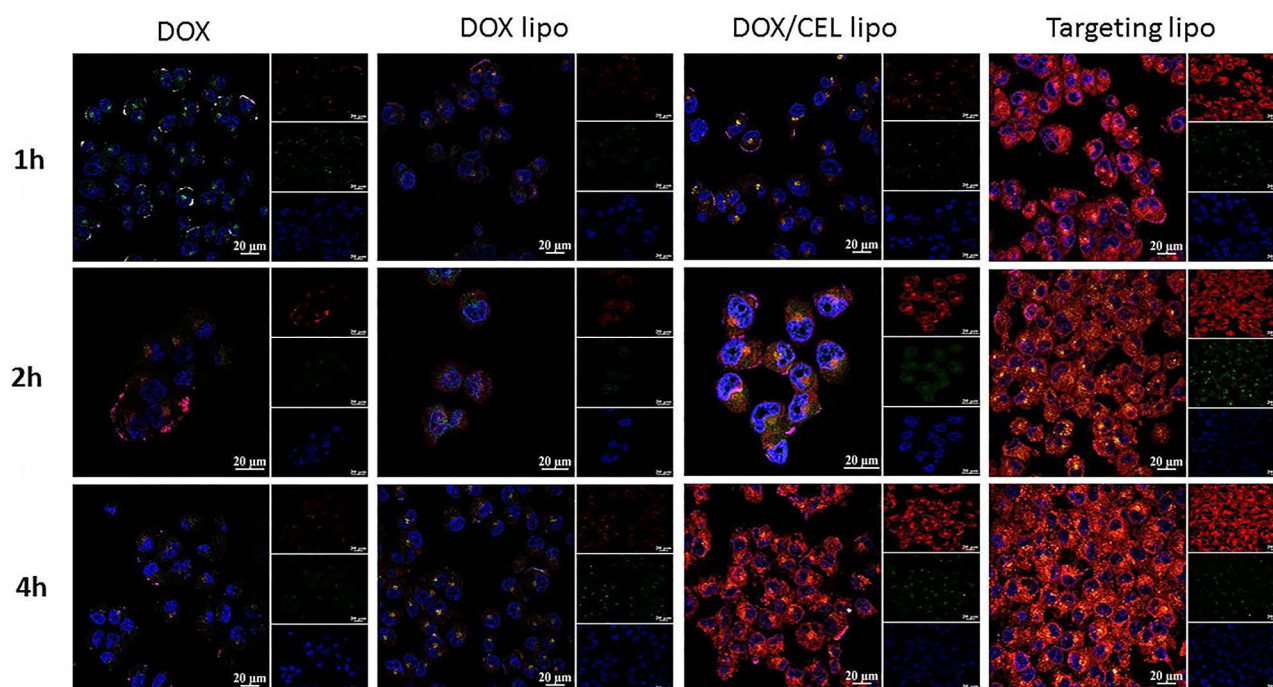


Figure 7 Mitochondrial targeting ability of different DOX formulations in MCF-ADR cells at different times. DOX was visualized as the red fluorescence, nuclei labeled with DAPI were shown as the blue fluorescence and green fluorescence indicates Mitotracker Green labeled-mitochondria. Scale bars correspond to 20 μm in all images.

fluorescence was clearly appeared (overlap of green and red fluorescence in the lysosomes) as a result of DOX trapping in endo-lysosomes. On the other hand, after incubation with DOX/CEL lipo and targeting lipo, the yellow fluorescence was gradually disappeared and the purple fluorescence (overlap of red and blue fluorescence) became more intense after one-hour incubation especially in case of peptide-modified liposomes. This phenomenon was probably attributed to the presence of the histidine-rich molecule used for modification of the liposomes. It was protonation of the imidazole ring of the histidine amino acid to produce a buffering effect, resulting in increasing of the osmotic pressure and disruption of the endosomal membrane and subsequently distribution of targeting liposomes to the mitochondria and nucleus.^{41,42}

MCF/ADR cells were incubated with different DOX liposomal formulations to track the mitochondrial targeting ability of the liposomes. As shown in **Figure 7**, the red fluorescence was mostly dispersed in the cytoplasm or localized around the nucleus. As the incubation time increases, the red fluorescence around the nucleus became strengthened. After 4 h of co-incubation, in case of DOX solu and DOX lipo still did not generate yellow fluorescence or purple fluorescence in the overlay chart (the yellow fluorescence generated from the overlap of the

red fluorescence (DOX) with the green fluorescence (Mitotracker)), indicating that DOX mainly accumulate in the lysosomes but does not transport to mitochondria and nucleus. After 2 h of co-incubation with DOX/CEL lipo, the red fluorescence increased, but the yellow fluorescence still weak, suggesting that part of the DOX/CEL lipo could be transported from the cytoplasm to the nucleus. On the other hand, the yellow fluorescence became shiny enough to be clearly seen after incubation of the cancer cells with the targeting liposome for 1 h and extremely increased over time. And this back to the presence of MTS molecule in the designed peptide as a mitochondrial targeting entity which is a well-studied natural sequence from the mitochondrial oriented protein aldehyde dehydrogenase (ALDH),^{43,44} which enables the liposome to migrate directly to the mitochondria and consequently increasing the yellow color intensity. That is why DOX/CEL-MTS- R_8H_3 modified liposomes display the greatest cytotoxicity among all tested formulations.

Drug-Induced Changes in Mitochondrial Transmembrane Potential

Mitochondrial changes, including disruptions in the mitochondrial membrane potential (MMP), are the vital events during the formulations-induced apoptosis. Apoptosis is

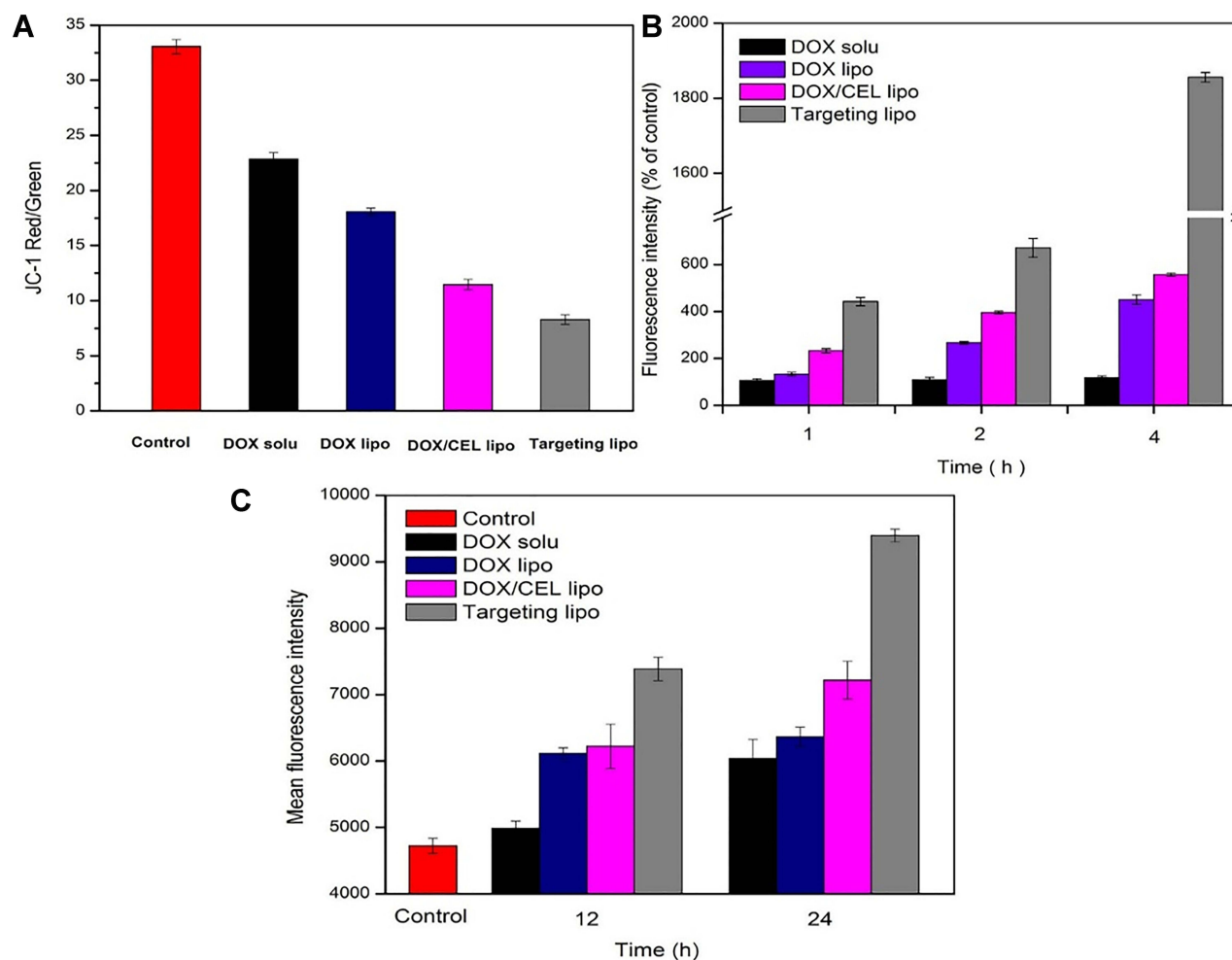


Figure 8 (A) Mitochondrial membrane potential expressed as JC-1 aggregate to monomer ratio in MCF/ADR cells incubated for 4h at 37°C. (B) ROS production by different dox formulations. (C) The intracellular accumulation of rhodamine 123 in MCF/ADR cells. The cells were incubated with 5 µg/mL Rh 123 for 30 min, and the intracellular fluorescence was measured to assess the pump function of P-gp. Data are reported as the mean ± standard deviation (n = 3) (doxorubicin (DOX), celecoxib (CEL), solution (solu), liposome (lipo)).

Abbreviation: ROS, reactive oxygen species.

commonly associated with mitochondrial membrane depolarization. JC-1 dye has been approved to be more specific for the $\Delta\psi$ than other fluorescent dyes such as rhodamine 123 or DiOC6(3).^{45,46} Mitochondria with normal MMP concentrate JC-1 into aggregates (red fluorescence), while JC-1 forms monomers (green fluorescence) in the depolarized state.⁴⁷ Different DOX formulations were incubated with MCF/ADR cells for 4 h at 37 °C. As shown in Figure 8A, the different liposomes were ranked according to the red/green ratios as follow: DOX solu > DOX lipo > DOX/CEL lipo > and targeting lipo. This indicated that the peptide modified targeting liposomes could be internalized into the tumor cells for targeting

the mitochondria, releasing the included DOX and CEL cargoes, resulting in significant depolarization of the MMP.

Determination of the ROS

Reactive oxygen species production was induced by treatment with different liposomes (Figure 8B). MCF/ADR cells treated with DOX/CEL co-loaded targeted liposomes displayed the highest ROS production ability (18.56 fold more than the control group), which was caused by the mitochondrial targeting sequence that enabled DOX liposome accumulation in the mitochondria. CEL at low concentrations in combination with DOX can abruptly induce

excessive production of superoxide through disrupting the cellular respiration and decreasing the mitochondrial membrane potential thereby inducing ROS production directly from the mitochondria, consequently, down-regulation of the multidrug resistance (MDR) transporter P-gp occurring and enhancing the sensitivity of MCF/ADR cancer cells to DOX.⁴⁸

Evaluation of P-gp Efflux Assay

Intracellular accumulation of rhodamine 123 (Rh123) was applied to evaluate the function of membrane P-gp. After cells treatment with different DOX formulations, DOX solu, DOX lipo, DOC/CEL lipo, and Targeting lipo, cells were incubated with Rh 123 for 30 min. The fluorescence intensity of Rh 123 was measured by FCM. As shown in Figure 8C. The data indicated that the mean Rh 123 fluorescence intensity value rank was: Targeting lipo > DOX/CEL lipo > DOX lipo > DOX solu. The intracellular accumulation of Rh 123 in case of DOX/CEL co-loaded liposome was significantly ($p < 0.05$) higher than that of single drug loaded liposomes in a time-dependent manner. Furthermore, the peptide modified liposomes exert the highest inhibitory effect on rhodamine efflux ($p < 0.001$) compared with other DOX formulations. These results indicated that our targeting liposomes could effectively induce greater drug payload to be delivered to the target cell and effectively inhibit doxorubicin-induced overexpression and function of P-gp in MCF/ADR cells. Previously published literature reported that the overexpression of the multicellular prostate tumor spheroids for P-gp-mediated MDR is revoked under elevated intracellular ROS level conditions.⁴⁹ And these data consistent with the previous findings that celecoxib can rapidly enhance the mitochondrial superoxide production from cancer cells within minutes leading to extensive ROS-dependent apoptosis in murine melanoma B16F10 cells.⁵⁰ But a deeper understanding of this accumulation mechanism is urgently needed and could lead to significantly more effective drug delivery in anticancer therapy.

Conclusions

In this research, efficient DOX/CEL-targeting liposomes were successfully prepared as a novel drug delivery carrier due to the targeting ability and therapeutic efficiency to breast cancer cells. Liposomes displayed a nanometric size of approximately 100–150 nm in diameter and spherically shaped vesicles with high encapsulation efficiency and

showed prominent storage stability for one month. DOX/CEL targeted liposomes achieved a significant reduction in cell viability in comparison with the other drug-loaded liposomes. MTS-R₈H₃ modified DOX/CEL liposome significantly increased the in vitro intracellular retention and accumulation of doxorubicin and achieved the best lysosomal escape and mitochondrial targeting ability with the highest ROS production efficiency and significant inhibition of P-gp efflux activity in MCF/ADR cells. These results indicated that MTS-R₈H₃ modified DOX/CEL liposomes can be used as a promising drug delivery system for preventing of chemoresistance of human breast cancer cell line and should be considered for future trials.

Data Sharing Statement

All data associated with this study are present in the paper and the [Supplementary Materials](#).

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Disclosure

The authors declare that they have no conflicts of interest for this work.

References

1. Siegel RL, Miller KD, Jemal A. Cancer statistics, 2020. *CA Cancer J Clin*. 2020;70(1):7–30. doi:10.3322/caac.21590
2. Mielczarek L, Krug P, Mazur M, et al. In the triple-negative breast cancer MDA-MB-231 cell line, sulforaphane enhances the intracellular accumulation and anticancer action of doxorubicin encapsulated in liposomes. *Int J Pharm*. 2019;558:311–318. doi:10.1016/j.ijpharm.2019.01.008
3. Zou LL, Ding WP, Zhang YY, et al. Peptide-modified vemurafenib-loaded liposomes for targeted inhibition of melanoma via the skin. *Biomaterials*. 2018;182:1–12. doi:10.1016/j.biomaterials.2018.08.013
4. Tang J, Wang Q, Yu Q, et al. A stabilized retro-inverso peptide ligand of transferrin receptor for enhanced liposome-based hepatocellular carcinoma-targeted drug delivery. *Acta Biomater*. 2019;83:379–389. doi:10.1016/j.actbio.2018.11.002
5. Xu HB, Shen FM, Lv QZ. Celecoxib enhanced the cytotoxic effect of cisplatin in chemo-resistant gastric cancer xenograft mouse models

- through a cyclooxygenase-2-dependent manner. *Eur J Pharmacol.* 2016;776:1–8. doi:10.1016/j.ejphar.2016.02.035
6. Tian JY, Guo FG, Chen YY, et al. Nanoliposomal formulation encapsulating celecoxib and genistein inhibiting COX-2 pathway and Glut-1 receptors to prevent prostate cancer cell proliferation. *Cancer Lett.* 2019;448:1–10. doi:10.1016/j.canlet.2019.01.002
 7. Kim H, Yim G, Kim Y. Celecoxib and paclitaxel synergistically induce apoptosis in the human ovarian cancer cell line OVCAR-3. *Gynecol Oncol.* 2013;130(1):126–132. doi:10.1016/j.ygyno.2013.04.362
 8. Meng XY, Zhang Q, Zheng GB, et al. Doxorubicin combined with celecoxib inhibits tumor growth of medullary thyroid carcinoma in xenografted mice. *Oncol Lett.* 2014;7(6):2053–2058. doi:10.3892/ol.2014.2050
 9. Chen C, Xu W, Wang CM. Combination of celecoxib and doxorubicin increases growth inhibition and apoptosis in acute myeloid leukemia cells. *Leuk Lymphoma.* 2013;54(11):2517–2522. doi:10.3109/10428194.2013.781170
 10. Kozlu S, Sahin A, Ultav G, et al. Development and in vitro evaluation of doxorubicin and celecoxib co-loaded bone targeted nanoparticles. *J Drug Deliv Sci Technol.* 2018;45:213–219. doi:10.1016/j.jddst.2018.02.004
 11. Fantappiè O, Solazzo M, Lasagna N, Platini F, Tessitore L, Mazzanti R. P-glycoprotein mediates celecoxib-induced apoptosis in multiple drug-resistant cell lines. *Cancer Res.* 2007;67(10):4915–4923.
 12. Huang L, Wang C, Zheng W, et al. Effects of celecoxib on the reversal of multidrug resistance in human gastric carcinoma by downregulation of the expression and activity of P-glycoprotein. *Anticancer Drugs.* 2007;18(9):1075–1080. doi:10.1097/CAD.0b013e3281c49d7a
 13. Sung MW, Lee DY, Park SW, et al. Celecoxib enhances the inhibitory effect of 5-FU on human squamous cell carcinoma proliferation by ROS production. *Laryngoscope.* 2016;127(4):E117–E123.
 14. Soukupová K, Rudolf E. Suppression of proliferation and activation of cell death by sodium selenite involves mitochondria and lysosomes in chemoresistant bladder cancer cells. *J Trace Elem Med Biol.* 2019;52:58–67. doi:10.1016/j.jtemb.2018.11.009
 15. Guo Z, Sevrioukova IF, Denisov IG, et al. Heme binding biguanides target cytochrome P450-dependent cancer cell mitochondria. *Cell Chem Biol.* 2017;24(10):1259–1275.e6. doi:10.1016/j.chembiol.2017.08.009
 16. Li SY, Cheng H, Xie BR, et al. Mitochondria targeted cancer therapy using ethidium derivatives. *Mater Today Chem.* 2017;6:34–44. doi:10.1016/j.mtchem.2017.09.002
 17. Zhao YJ, Sun LM, Wang RR, et al. The effects of mitochondria-associated long noncoding RNAs in cancer mitochondria: new players in an old arena. *Crit Rev Oncol Hematol.* 2018;131:76–82. doi:10.1016/j.critrevonc.2018.08.005
 18. Copolovici DM, Langel K, Eriste E, et al. Cell-penetrating peptides: design, synthesis, and applications. *ACS Nano.* 2014;8(3):1972–1994. doi:10.1021/nn4057269
 19. Walrant A, Cardon S, Burlina F, et al. Membrane crossing and membranotropic activity of cell-penetrating peptides: dangerous liaisons? *Acc Chem Res.* 2017;50(12):2968–2975. doi:10.1021/acs.accounts.7b00455
 20. Freire JM, Veiga AS, Inês R, et al. Nucleic acid delivery by cell penetrating peptides derived from dengue virus capsid protein: design and mechanism of action. *Febs J.* 2014;281(1):191–215. doi:10.1111/febs.12587
 21. Ahmed KS, Shan XT, Mao J, et al. Derma roller[®] microneedles-mediated transdermal delivery of doxorubicin and celecoxib co-loaded liposomes for enhancing the anticancer effect. *Mater Sci Eng C Mater Biol Appl.* 2019;99:1448–1458. doi:10.1016/j.msec.2019.02.095
 22. Yu J, Li W, Yu D. Atrial natriuretic peptide modified oleate adenosine prodrug lipid nanocarriers for the treatment of myocardial infarction: in vitro and in vivo evaluation. *Drug Des Devel Ther.* 2018;12:1697–1706. doi:10.2147/DDDT.S166749
 23. Iden DL, Allen TM. In vitro and in vivo comparison of immunoliposomes made by conventional coupling techniques with those made by a new post-insertion approach. *Biochim Biophys Acta Biomembr.* 2001;1513(2):207–216. doi:10.1016/S0005-2736(01)00357-1
 24. Song SX, Liu D, Peng JJ, et al. Peptide ligand-mediated liposome distribution and targeting to EGFR expressing tumor in vivo. *Int J Pharm.* 2008;363(1):155–161. doi:10.1016/j.ijpharm.2008.07.012
 25. Li CL, Cui JX, Wang CX, et al. Development of pegylated liposomal vinorelbine formulation using “post-insertion” technology. *Int J Pharm.* 2010;391(1):230–236. doi:10.1016/j.ijpharm.2010.03.004
 26. Silverman L, Barenholz Y. In vitro experiments showing enhanced release of doxorubicin from doxil[®] in the presence of ammonia may explain drug release at tumor site. *Nanomedicine.* 2015;11(7):1841–1850. doi:10.1016/j.nano.2015.06.007
 27. Fetih G, Fathalla D, El-Badry M. Liposomal gels for site-specific, sustained delivery of celecoxib: in vitro and in vivo evaluation. *Drug Dev Res.* 2014;75(4):257–266. doi:10.1002/ddr.21179
 28. Scaduto RC, Grotjohann LW. Measurement of mitochondrial membrane potential using fluorescent rhodamine derivatives. *Biophys J.* 1999;76(1):469–477. doi:10.1016/S0006-3495(99)77214-0
 29. Szwed M, Laroche-Clary A, Robert J, et al. Efficacy of doxorubicin-transferrin conjugate in apoptosis induction in human leukemia cells through reactive oxygen species generation. *Cell Oncol.* 2016;39(2):107–118. doi:10.1007/s13402-015-0256-2
 30. Yin J, Guo JB, Zhang Q, et al. Doxorubicin-induced mitophagy and mitochondrial damage is associated with dysregulation of the PINK1/parkin pathway. *Toxicol in Vitro.* 2018;51:1–10. doi:10.1016/j.tiv.2018.05.001
 31. Wang JS, Ho FM, Kang HC, et al. Celecoxib induces heme oxygenase-1 expression in macrophages and vascular smooth muscle cells via ROS-dependent signaling pathway. *Naunyn Schmiedebergs Arch Pharmacol.* 2011;383(2):159–168. doi:10.1007/s00210-010-0586-6
 32. Ma M, Yang X, Zhao L, et al. Celecoxib enhances sensitivity to chemotherapy drugs of T-cell lymphoma. *Oncol Lett.* 2018;15(4):4649–4656.
 33. Lisa S, Tejaswi V, Fatemeh M, et al. Advances and challenges of liposome assisted drug delivery. *Front Pharmacol.* 2015;6(127):27–31.
 34. Kalyane D, Raval N, Maheshwari R, et al. Employment of enhanced permeability and retention effect (EPR): nanoparticle-based precision tools for targeting of therapeutic and diagnostic agent in cancer. *Mater Sci Eng C Mater Biol Appl.* 2019;98:1252–1276. doi:10.1016/j.msec.2019.01.066
 35. Sabeti B, Noordin MI, Mohd S, et al. Development and characterization of liposomal doxorubicin hydrochloride with palm oil. *Biomed Res Int.* 2014;2014:765426. doi:10.1155/2014/765426
 36. Silverman L, Barenholz Y. In vitro experiments showing enhanced release of doxorubicin from doxil in the presence of ammonia may explain drug release at tumor site. *Nanomedicine.* 2015;11(7):31–52.
 37. Ahmed KS, Hussein SA, Ali AH, et al. Liposome: composition, characterisation, preparation, and recent innovation in clinical applications. *J Drug Target.* 2018;27:1–58.
 38. Farzaneh H, Ebrahimi NM, Mashreghi M, et al. A study on the role of cholesterol and phosphatidylcholine in various features of liposomal doxorubicin: from liposomal preparation to therapy. *Int J Pharm.* 2018;551(1):300–308. doi:10.1016/j.ijpharm.2018.09.047
 39. Khalil IA, Kogure K, Futaki S, et al. Octaarginine-modified liposomes: enhanced cellular uptake and controlled intracellular

- trafficking. *Int J Pharm.* 2008;354(1):39–48. doi:10.1016/j.ijpharm.2007.12.003
40. Kitagishi H, Hata DS, Itakura T, et al. Cellular uptake of octaarginine-conjugated tetraarylporphyrin included by per-O-methylated β -cyclodextrin. *Org Biomol Chem.* 2013;11(19):3203–3211. doi:10.1039/c3ob27248f
41. Varkouhi AK, Scholte M, Storm G, et al. Endosomal escape pathways for delivery of biologicals. *J Control Release.* 2011;151(3):220–228. doi:10.1016/j.jconrel.2010.11.004
42. Moreira C, Oliveira H, Pires LR, et al. Improving chitosan-mediated gene transfer by the introduction of intracellular buffering moieties into the chitosan backbone. *Acta Biomater.* 2009;5(8):2995–3006. doi:10.1016/j.actbio.2009.04.021
43. Wang Y, Weiner H. The presequence of rat liver aldehyde dehydrogenase requires the presence of an alpha-helix at its N-terminal region which is stabilized by the helix at its C termini. *J Biol Chem.* 1993;268(7):4759–4765. doi:10.1016/S0021-9258(18)53462-1
44. Lin R, Zhang P, Cheetham AG, et al. Dual peptide conjugation strategy for improved cellular uptake and mitochondria targeting. *Bioconjug Chem.* 2015;26(1):71–77. doi:10.1021/bc500408p
45. Perry SW, Norman JP, Barbieri J, et al. Mitochondrial membrane potential probes and the proton gradient: a practical usage guide. *Biotechniques.* 2011;50(2):98–115. doi:10.2144/000113610
46. Anthony M, Hong Y, Kemp BK, et al. Evaluation of fluorescent dyes for the detection of mitochondrial membrane potential changes in cultured cardiomyocytes. *Cardiovasc Res.* 2000;1:126–138.
47. Qiu L, Qiao MX, Chen Q, et al. Enhanced effect of pH-sensitive mixed copolymer micelles for overcoming multidrug resistance of doxorubicin. *Biomaterials.* 2014;35(37):9877–9887. doi:10.1016/j.biomaterials.2014.08.008
48. Wartenberg M, Ling FC, Schallenberg M, et al. Down-regulation of intrinsic P-glycoprotein expression in multicellular prostate tumor spheroids by reactive oxygen species. *J Biol Chem.* 2001;276(20):17420–17428. doi:10.1074/jbc.M100141200
49. Wartenberg M, Hoffmann E, Schwindt H, et al. Reactive oxygen species-linked regulation of the multidrug resistance transporter p-glycoprotein in Nox-1 overexpressing prostate tumor spheroids. *FEBS Lett.* 2005;579(20):4541–4549. doi:10.1016/j.febslet.2005.06.078
50. Pritchard R, Rodríguez-Enríquez S, Pacheco-Velázquez SC, et al. Celecoxib inhibits mitochondrial O₂ consumption, promoting ROS dependent death of murine and human metastatic cancer cells via the apoptotic signalling pathway. *Biochem Pharmacol.* 2018;154:318–334. doi:10.1016/j.bcp.2018.05.013

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