DOI: 10.1002/prp2.598

REVIEW



Mechanistic insights into antiretroviral drug-induced liver injury

Jamie N. Pillaye¹ | Mohlopheni J. Marakalala^{2,3} | Nonhlanhla Khumalo⁴ | Wendy Spearman⁵ | Hlumani Ndlovu¹

¹Division of Chemical and System Biology, Department of Integrative Biomedical Sciences, Faculty of Health Sciences, University of Cape Town, Cape Town, South Africa

²Africa Health Research Institute, Durban, KwaZulu Natal, South Africa

³Division of Infection and Immunity, University College London, London, UK

⁴Hair and Skin Research Lab, Division of Dermatology, Department of Medicine, Groote Schuur Hospital and University of Cape Town, Cape Town, South Africa

⁵Division of Hepatology, Department of Medicine, Groote Schuur Hospital and University of Cape Town, Cape Town, South Africa

Correspondence

Hlumani Ndlovu, Division of Chemical and System Biology, Department of Integrative Biomedical Sciences, University of Cape Town, Faculty of Health Sciences, Cape Town, South Africa. Email: hlumani.ndlovu@uct.ac.za

Funding information

This work was supported by the South African National Research Foundation (NRF) Competitive Support for Unrated Researchers (116260), NRF Equipment Related Travel Grant (120164) to HN; South African Medical Research Council Capacity Development Grant to HN and MJM; and NRF MSc Fellowship to JP.

Abstract

All classes of antiretroviral therapy (ART) have been implicated to induce adverse drug reactions such drug-induced liver injury (DILI) and immune-mediated adverse reactions in Human Immunodeficiency Virus (HIV) infected individuals. Patients that develop adverse drug reactions tend to have prolonged stays in hospital and may require to change to alternative regimens if reactions persist upon rechallenge or if rechallenge is contraindicated due to severity of the adverse reaction. Diagnosis of DILI remains a huge obstacle that delays timely interventions, since it is still based largely on exclusion of other causes. There is an urgent need to develop robust diagnostic and predictive biomarkers that could be used alongside the available tools (biopsy, imaging, and serological tests for liver enzymes) to give a specific diagnosis of DILI. Crucial to this is also achieving consensus in the definition of DILI so that robust studies can be undertaken. Importantly, it is crucial that we gain deeper insights into the mechanism of DILI so that patients can receive appropriate management. In general, it has been demonstrated that the mechanism of ART-induced liver injury is driven by four main mechanisms: mitochondrial toxicity, metabolic host-mediated injury, immune reconstitution, and hypersensitivity reactions. The focus of this review is to discuss the type and phenotypes of DILI that are caused by the first line ART regimens. Furthermore, we will summarize recent studies that have elucidated the cellular and molecular mechanisms of DILI both in vivo and in vitro.

KEYWORDS

antiretroviral therapy, diagnosis, drug-induced liver injury, human immunodeficiency virus, mechanisms

1 | INTRODUCTION

Acquired Immune Deficiency Syndrome (AIDS), caused by Human Immunodeficiency Virus (HIV), is a highly communicable disease that has placed a tremendous burden on national healthcare systems globally but specifically in sub-Saharan Africa. The disease was estimated to affect 37.9 million people worldwide in 2018, directly causing approximately 770 000 deaths.¹ The prevalence of HIV infections is the highest in Eastern and Southern Africa where an estimated 20.6 million people are living with HIV/AIDS. Co-infection

© 2020 The Authors. Pharmacology Research & Perspectives published by British Pharmacological Society and American Society for Pharmacology and Experimental Therapeutics and John Wiley & Sons Ltd

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

with opportunistic infections, predominantly tuberculosis (TB), have further increased the morbidity and mortality rate of HIV-infected individuals. In 2018, 1.5 million individuals died from TB, of which 400 000 were HIV positive, making TB the top killer of people living with HIV/AIDS.

The successful roll out of antiretroviral therapy (ART) has reduced the risk of early mortality due to opportunistic infections and improved both the prognosis as well as the quality of life for HIV-infected patients. There was an estimated 24.5 million people receiving ART worldwide in 2018, a significant increase from 7.7 million people that were accessing ART in 2010.¹ This jump in the number of people accessing ART seems to have been propelled by the implementation of the "test and treat" policy of the World Health Organisation (WHO) in 2015. Consequently, there has also been a 33% decline in AIDS-related deaths since 2010,¹ indicating the success of the current efforts to treat and control the disease.

Unfortunately, some patients receiving ART may develop severe adverse drug reactions such as drug-induced liver injury (DILI). The reported incidence of ART-induced liver injury varies greatly due a number of factors such as a criteria used to define the severity of hepatotoxicity, geographical location, prevalence of chronic viral hepatitis infections, genetic heterogeneity in populations, frequency of liver enzyme determinations, medication prescribing patterns, and other exogenous exposures.²⁻⁴ Hence, the reported incidence of ART-induced DILI ranges from 8%-23% in HIV-infected patients and up to 30% of these patients may require change of regimen or the discontinuation of therapy.⁴⁻⁷ DILI is a common cause of prolonged hospitalization in HIVinfected patients and in severe cases may be fatal. Discontinuation of the offending drug can lead to treatment failure, and the potential emergence of drug resistant pathogens. Studies have shown that efavirenz (EFV), a first line ART drug is a key driver of DILI in HIV-infected individuals taking the first line EFV/Tenofovir disoproxil fumarate/ Emtricitabine fixed-dose combination. The reported risk factors for DILI are female gender, young age, and high CD4 counts.^{8,9}

In this review, we will discuss the different patterns and severity of DILI and describe the types and phenotypes of DILI caused by ART in HIV-infected individuals. We will also provide a summary of the studies that have analysed the molecular mechanisms of DILI with a particular focus on cellular toxicity, immunological responses, and biochemical pathways that have been implicated to drive hepatotoxicity of first line ART. It is important to note that the WHO recently introduced new guidelines that recommended the use of dolutegravir alongside two nucleoside reverse transcriptase inhibitors (NRTIs) as a first line regimen.¹⁰ Although there were some initial concerns about the safety of dolutegravir, particularly in pregnant women,¹¹ subsequent studies have shown it to be safe.^{12,13} The effects of this new regimen will not be the focus of this review.

1.1 | Patterns and severity of DILI

There has been great variability in the criteria used in clinical studies to define DILI and/or the severity of hepatotoxicity. Some studies

have defined DILI as elevations in liver enzymes (alanine aminotransferase [ALT] and aspartate aminotransferase [AST]) two times the upper limit of normal (ULN),^{14,15} while others used an absolute threshold of liver enzymes regardless of the baseline levels.^{16,17} Efforts have been made to achieve consensus in the definition and grading of DILI.¹⁸⁻²¹ The clinical presentations of DILI have been classified biochemically and clinically into three main categories based on *R* value (*R* = (ALT value ÷ ALT ULN) ÷ (ALP value ÷ ALP ULN)). DILI is categorized as hepatocellular when *R* is more than 5 (*R* > 5), cholestatic when *R* is less than 2 (*R* < 2), and mixed when *R* is more than 2 but less than 5 (2 < *R* < 5).^{19,20,22} It is important to mention that ARTs are mainly associated with hepatocellular DILI that usually arise within one year of starting the offending therapy.

The DILI Expert Working Group and AIDS Clinical Trial Group graded DILI severity into four grades based on baseline ALT or alkaline phosphatase (ALP) below the ULN.^{18,21} Grade 1 (mild) is defined by 1.25-2.5x ULN ALT or ALP; Grade 2 (moderate) is characterized by 2.6-5.0x ULN ALT or ALP; Grade 3 (severe) defined by 5.1-10x ULN ALT or ALP; and Grade 4 (severe) is defined by greater than 10x ULN ALT or ALP, or death, or transplantation due to DILI.^{18,21} Incidences of DILI as well as the associated side effects vary between individuals due to their genetic profile and whether they are consuming other drugs. Hence, for the effective management of DILI, a more personalized treatment approach may be required.

All classes of ARTs cause DILI in HIV-infected patients; however, some classes are more toxic than others. Table 1 provides a list of ARTs that are known to cause adverse reactions in HIV positive individuals. Non-nucleoside reverse transcriptase inhibitors (NNRTI) such as nevirapine (NVP) have been shown to induce DILI in 12% of the patients, which is three times more than that caused by EFV.²³ Similar results were reported by Sanne and colleagues, who found that patients treated with a regimen containing NVP had a DILI incidence rate of 17%, while those treated with EFV did not develop DILI.²⁴ This study recruited HIV-infected men and nonpregnant women who were ART naïve, had plasma HIV-1 RNA levels greater than 5000 copies/mL and CD4⁺ cell count that was greater than 200 cells/mm^{3, 24} Importantly, 385 patients were enrolled in the NVP arm while the EFV arm had only 83 patients, hence, introducing a possible bias and skewing of the data. Therefore, these data need to be interpreted with more caution. Although these studies indicate a higher risk of liver toxicity for patients receiving NVP compared to EFV, EFV has been shown to be toxic in both treatment naïve and experienced patients.9,25,26

Nucleoside reverse transcriptase inhibitors, protease inhibitors (PIs), and fusion inhibitors have also been reported to cause DILI in HIV-infected patients. NRTIs such as zidovudine, stavudine, and didanosine caused moderate to severe DILI while emtricitabine, abacavir, and tenofovir induce minor elevations in liver enzymes.²⁷ PIs have been reported to cause DILI in 1%-9.5% of the patients, with a few patients that have been reported to develop severe elevation in liver enzymes.²⁷ There have been a few case reports of liver toxicity with indinavir and tipranavir, particularly in patients with underly-ing diseases such as cirrhosis and chronic hepatitis C infection.^{28,29} **TABLE 1**Summary of the antiretroviraldrugs with a known adverse reaction inHIV positive patients

Drug	Class	Adverse reaction	References
Nevirapine	NNRTI	DILI, hypersensitivity	4-8,23
Efavirenz	NNRTI	DILI, hypersensitivity	4-8,23
Zidovudine	NRTI	DILI	27
Stavudine	NRTI	DILI	27
Didanosine	NRTI	DILI	27
Abacavir	NRTI	Hypersensitivity	75,80-82
Indinavir	PI	DILI	27,29
Tipranivir	PI	DILI	27,29
Aplaviran	CCR5 antagonist	DILI	30

Abbreviations: DILI, drug-induced liver injury; HIV, Human Immunodeficiency Virus; NNRTI, nonnucleoside reverse transcriptase inhibitors; NRTI, nucleoside reverse transcriptase inhibitor; PI, protease inhibitor.

The majority of PI's reported to cause DILI are not constituents of the fixed-dose drug regimen that is composed of EFV, tenofovir and emtricitabine. GlaxoSmithKline halted a Phase 2b and 3 clinical trials for Aplaviran, a CCR5 antagonist, after four men from approximately 300 patients developed severe liver disease.³⁰ The other CC5R antagonists, maraviroc and vicriviroc, appear to be safe. Enfuvirtide, the only approved fusion inhibitor, appears to be well-tolerated and safe.³¹ Similarly, integrase inhibitors such as MK-0518³² and dolute-gravir have been demonstrated to be safe and tolerable in treatment naïve and experienced adults and adolescents.³³⁻³⁶ Moreover, a recent observational study conducted in Botswana showed that dolutegravir-based ART regimen was safe for pregnant women.¹¹

1.2 | Types and major phenotypes of drug induced liver injury

Drug induced liver injury (DILI) has been mainly classified as either direct or idiosyncratic; however, indirect injury is emerging as a third class.^{37,38} Direct hepatotoxicity is attributed to the drugs or metabolites that are intrinsically toxic to the liver, biliary epithelial cells, and the liver vasculature. It is common, predictable, dose-dependent, and reproducible in animal models.³⁷ Direct hepatotoxicity occurs rapidly, with a latency period of one and five days after intake of high therapeutic doses of a drug. This type of injury is associated with elevations in liver enzymes such as ALT and ALP without the accompanying hyperbilirubinemia.³⁹ The elevations in liver enzymes can subside when the offending drug is stopped or dose is reduced—but in some cases can spontaneously resolve, a phenomenon referred to as adaptation.⁴⁰

1.2.1 | Direct hepatotoxicity

Clinically, the most common feature of direct hepatotoxicity is acute hepatic necrosis, and severe cases present with acute hepatic failure characterized by encephalopathy and coagulopathy.^{20,41} Interestingly, histological examination depicts a centrilobular and

panlobular necrosis with little inflammation. It is interesting to note that NNRTIs such as NVP and EFV have been reported to cause similar pathological features with some distinctive features such as inflammation with lymphocytes, eosinophils, and plasma cells. For instance, NVP has been shown to be more toxic causing portal tract expansion and severe inflammatory reactions with eosinophils infiltrating the parenchyma.⁴² Analysis of liver biopsies obtained from patients that received a regimen containing EFV revealed that submassive necrosis was associated with significant morbidity and mortality.^{8,25} Submassive necrosis is characterized by zonal/panzonal necrosis with an "immune-allergic" pattern and recruitment of inflammatory cells, such lymphocytes, plasma cells, and eosinophils.²⁵ A study by Elsharkawy and colleagues identified a first adult case of EFV induced acute liver failure that required liver transplantation within 5 months after initiating therapy.⁴³ Interestingly, this 43-year-old woman was a slow drug metabolizer, which contributed to the severe clinical presentation.⁴³ The first generation NRTI's such as zidovudine, stavudine, and didanosine have been reported to cause nodular regenerative hyperplasia, a noncirrhotic portal hypertension.⁴⁴ Finally, didanosine has been shown to induce hyperlactatemia and lactic acidosis that has an onset within months of initiating therapy in HIV-infected individuals ⁴⁵

1.2.2 | Idiosyncratic hepatotoxicity

Idiosyncratic hepatotoxicity occurs in rare cases (typically 1 to 2000 and 1 to 100 000 patient-exposures) and is usually caused by drugs with little or no intrinsic toxicity to the liver.^{46,47} It is unpredictable, dose-independent, and unreproducible in animal models. Idiosyncratic injury is classified into three categories based on the *R* value; hepatocellular, cholestatic, and mixed.²⁰ Idiosyncratic liver injury frequently manifests as mixed injury with some hepatocellular hepatitis.^{46,48,49} Drugs such as isoniazid, nitrofurantoin, and diclofenac have been incriminated as the common causes of idiosyncratic hepatocellular injury.⁴⁸⁻⁵¹ Idiosyncratic drug-induced injury may be associated with an immune-allergic reaction such as rash,

PILLAYE ET AL.

fever, and eosinophilia-all typical signs of hypersensitivity. 48,49,52 NVP and EFV have been reported to be the most common causes of skin rashes in patients taking first line ART regimens and have an incidence rate of 10%-17%. 53-55 NNRTIs induce a mild macular, maculopapular, or erythematous rash and a severe rash characterized with blistering, moist desquamation, and ulceration.⁵³ Numerous reviews have been written on hypersensitivity reactions to ARTs in HIV-infected individuals.^{19,56-58}

1.2.3 | Indirect hepatotoxicity

Indirect hepatotoxicity is caused by the action of the drug in triggering a new liver condition or worsening an underlying condition such as viral hepatitis than its intrinsic toxicity or idiosyncratic properties.³⁸ Indirect toxicity presents with the characteristics of the underlying condition or predisposition. ART-mediated immune reconstitution inflammatory syndrome has been reported to trigger flares of liver enzymes, spontaneous seroconversion, and exacerbation of hepatitis caused by Hepatitis B and C virus (HBV and HCV).⁵⁹ In fact, numerous studies have reported that HIV-infected patients with an underlying chronic HBV or HCV may develop severe toxicity to ART and need closer monitoring.⁶⁰⁻⁶⁷

1.3 | The role of genetic polymorphisms in **ART-induced liver injury**

Drug metabolizing enzymes 1.3.1

Genotype/

allele

6/6

0101

0102

5701

3505

5801

Gene

CYP2B6

HLA-DRB1

HLA-DRB1

HLA-B

HLA-B

HLA-B

Genetic variations in drug metabolizing enzymes have been associated with risk of adverse drug reaction in patients taking ART (Table 2). A study by Phillips and colleagues found an association between CYP2B6 slow metabolizer genotype with NVP driven hypersensitivity reactions (rash) but not with hepatotoxicity.⁶⁸ This was further demonstrated in a recent study by de Almeida and colleagues who analysed the drug metabolism gene polymorphisms in a Brazilian cohort that received an EFVbased regimen and found that the CYP2B6 slow metabolizer genotype was associated with an increased risk of EFV adverse reaction.⁶⁹ They also showed that CNS adverse effects were associated with CYP3A4

Drugs

Nevirapine

Nevirapine

Nevirapine

Nevirapine

Abacavir

Nevirapine, efavirenz

rs4646437 genotype.^{69,70} In a Thai cohort, the CYP2B6 haplotype *6/*6 was identified in 8.2% of the HIV-infected patients that received an EFVbased regimen and it was associated with high ALP and total bilirubin, suggesting that this genotype may correlate with an increased risk of hepatotoxicity in patients taking an EFV-based ART regimen.⁷¹ Similar findings were observed in a prospective cohort Ethiopian study that revealed an association between the CYP2B6 *6/*6 genotype with ARTinduced liver injury and high plasma concentrations of EFV.⁷² Therefore, it may be necessary to screen individuals for the CYP2B6 genotypes so they could be given therapy that would have minimal adverse effects to encourage compliance treatment or know the most likely CYP2B6 genotypes within a population to decide on pragmatic therapy.

1.3.2 | Cytokines

The immune response plays a critical role in driving some of the adverse drug reactions that are associated with ART in HIV-infected individuals. The severity of DILI is influenced by the presence of toxic molecules and a critical counterbalance of cytokines such as tumor necrosis factor-alpha, interleukin-1beta (IL-1β), IL-10 and IL-1RN.⁶⁸ In fact, some studies have associated polymorphisms in cytokine genes with the development of ART-associated hepatotoxicity⁷³ and hypersensitivity.⁷⁴ A recent study by Singh and colleagues investigated IL-1RN (VNTR) and IL-1 β (-511C/T) polymorphisms in 34 HIV-positive patients with ARV hepatotoxicity, 128 HIV-positive patients without hepatotoxicity, and 152 healthy controls using PCR and PCR-RFLP. They found that IL-1RN 2/2 and 1/3 genotypes were highly represented in patients with hepatotoxicity to a NVP based ARV regimen.⁷⁵⁻⁷⁷ Moreover, *IL*-1 β -511CT and -511TT genotypes enhanced the risk of hepatotoxicity in patients taking the NVP based regimen. Finally, a study by Asensi and colleagues showed that $IL-1\beta$ 3954T allele is associated with lipodystrophy syndrome in patients on ART.^{78,79}

1.3.3 | Human leucocyte antigen

Reference

69,71,105

78,83

84,85

85-87

79

79 75,80-82

The interactions between the T cells and professional antigen presenting cells via the class I or class II human leucocyte antigen (HLA) plays a key role in development of adverse drug reactions. In fact,

> TABLE 2 Genetic variations in Cytochrome P450 and human leucocyte antigen genes that are associated with ART induced adverse reactions

	HLA-C	0401	Nevirapine	Hypersensitivity	
Abbreviations: ART, antiretroviral therapy: DILI, drug-induced liver injury.					

Adverse reaction

Hypersensitivity

Hypersensitivity

DILI

DILI

DILI, hypersensitivity

DILI, hypersensitivity

HLA-B*5701 polymorphism has been linked with abacavir hypersensitivity in HIV-positive individuals.^{75,80-82} while the HLA-DRB1*0101⁷⁸ and HLA-B*3505⁸³⁻⁸⁵ was associated with increased risk of developing NVP-driven hypersensitivity reactions.^{79,83,85-87} Moreover, HLA-C*0401 allele is associated with hypersensitivity to NVP and is carried across most ethnicities.⁸⁵⁻⁸⁷ HLA-DRB*0101 has been associated with a risk of developing NVP induced hepatitis across ethnicities.^{83,85} A study by Phillips and colleagues that recruited 385 South African participants who initiated a NVP containing regimen showed that HLA-B*5801 and HLA-DRB1*0102 were associated with hepatotoxicity.⁷⁹ All these alleles and the adverse reaction that are triggered by ART are summarized in Table 2. Hence, HLA typing applied as a precision medicine tool could significantly reduce a risk of adverse reactions to ART. In fact, this has been implemented for abacavir hypersensitivity and has been hailed as a huge success as revealed in a recent metaanalysis by Stainsby and colleagues.⁸²

1.4 | Molecular mechanisms of ART-driven hepatotoxicity

1.4.1 | Effect of drugs on cellular morphology

The molecular mechanism of hepatotoxicity induced by ART seems to be multifactorial. A number of studies have reported that EFV causes cellular damage indicated by changes in the cells morphology, endoplasmic reticulum (ER), and mitochondrial integrity.⁸⁸⁻⁹⁰ A study by Apostolova and colleagues showed that Hep3B cells treated with EFV displayed a concentration dependent mitochondrial dysfunction and damage compared to control cells that contained smooth mitochondria with complete membranes and distinct and well-formed cristae.^{88,89} Hep3B cells are an immortalized human hepatoma cell line that expresses little or no drug metabolizing enzymes that are required for drug metabolism and toxicity studies.^{88,89} Moreover. Hep3B cells exposed to EFV displayed an abnormal cellular morphology with a swollen phenotype.⁸⁹ A higher concentration of EFV induced severe damage depicted by visible alterations of the mitochondrial outer membranes, reduction in the number of mitochondrial cristae and change in surface structure.⁸⁹

Mitochondrial toxicity is a common pathway linked with the use of NRTIs in HIV positive individuals.⁹¹ The mitochondria is an essential organelle that is critical for energy production, metabolism of glucose and fats, and production of reactive oxygen species (ROS) that could be deleterious to the cells. A study by de Mendoza and colleagues showed that HIV-positive patients taking a stavudine-containing regimen had significantly reduced mitochondrial DNA copy number compared to HIV-positive patients that were on other regimens, and this was associated with elevated lactate levels.⁹² Mitochondrial toxicity can manifest in patients taking NRTI drugs as nonspecific symptoms to lactic acidosis syndrome that may be accompanied by fulminant hepatic failure.^{92,93} Moreover, serum analysis can reveal modest elevations in liver enzymes, with AST greater than ALT. Unfortunately, the mortality rate is high in patients

ASPET - BRITISH PHARMACOLOGICAL 5 of 9

who develop lactic acidosis syndrome, but administration of specific therapy with cofactors can lower the mortality rate.⁹⁴

1.4.2 | Bioenergetics and signaling pathways

Research has implicated the involvement of respiratory chain proteins,^{88,95} abnormal Jun N-terminal kinases (JNK) and BIM-extralong (BimEL) signaling pathways in the development of ARV induced cytotoxicity (Figure 1).⁹⁶ A study by Perier and colleagues showed that some ART drugs inhibit complex I (NADH: ubiquinone oxidoreductase) of the electron transport chain (ETC), resulting in defects in mitochondrial bioenergetics due to reduced flow of electrons and impairment of oxidative phosphorylation,⁹⁷ especially in tissues with high energy demand such as the liver. The inhibition of complex I by EFV has been shown to result in the accumulation of lipids in the cytoplasm of both human hepatic tissue as well as Hep3B cells.⁹⁸ As expected, the inhibition of complex I impairs the respiratory chain resulting in reduced levels of adenosine triphosphate (ATP), increased production of ROS, and continuous activation of adenosine monophosphate-activated protein kinase (AMPK), which is the major regulator of cellular bioenergetics⁹⁸⁻¹⁰⁰ (Figure 1). Interestingly, methylene blue, an alternative electron carrier which can bypass the proximal ETC, can prevent the energy crisis and protect against lethal cell injury associated with the mitochondria targeting drugs.⁹⁶ These results are compatible with the concept that underlying silent mitochondrial dysfunction may be a susceptibility factor contributing to DILI.96

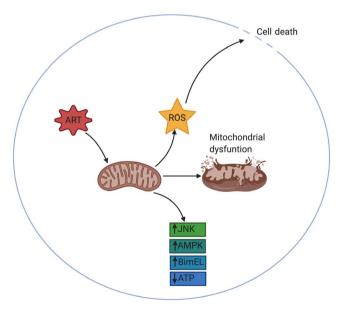


FIGURE 1 Antiretroviral therapy caused mitochondirial dysfunction and increased ROS production in vitro. Antiretroviral therapy, particularly efavirenz has been implicated in causing mitochondrial dysfunction, increased production of reactive oxygen species (ROS), leading to cell death. Moreover, drugs induced increased expression of JNK, AMPK and BimEL. AMPK, adenosine monophosphate-activated protein kinase; BimEL, BIM-extra-long; JNK, Jun N-terminal kinases

Efavirenz is known to increase the activity of AMPK, which results in increased production of ROS and possibly initiates cell death.¹⁰¹? In fact, treating primary mouse hepatocytes with EFV triggered cell death that was associated with complex I inhibition, peroxynitrite formation and ATP depletion.¹⁰¹ Interestingly, deletion of *sirt3* gene encoding Sirtuin-3, a major mitochondrial NAD⁺-dependent deacetylase in mice (Sirt3 knockout mice) protected hepatocytes from EFV-induced cytotoxicity compared to cells from wild-type mice.¹⁰¹ Therefore, induction of stress responses, particularly the production of ROS and peroxynitrite formation is implicated in causing cell death in EFV treated cells.

Another interesting biochemical family of serine/threonine kinase is the JNK. These are important regulators of cellular stress responses including modulation of cell death.¹⁰¹ Studies have reported that EFV treated primary human hepatocytes undergo apoptotic cell death that requires JNK activation and BimEL expression (Figure 1).¹⁰¹ Moreover, these effects can be recapitulated via treatment of hepatocytes using synthetic 8-hydroxy EFV, which is the primary metabolite of EFV.¹⁰² The inhibition of cytochrome P450mediated metabolism markedly decreased the toxicity of EFV in human hepatocytes measured by cell death, caspase-3 activation and ROS formation.¹⁰² The potential role of JNK and BimEL signaling pathways and its involvement in EFV-mediated hepatocyte death makes it a possible target for design and development of drugs to mitigate EFV-induced hepatotoxicity.

A study by Lee et al showed that in murine hepatocytes, the high concentrations of isoniazid alone did not cause acute cell injury.¹⁰² However, there was evidence that acute exposure to isoniazid caused mitochondrial abnormalities, specifically changes in the oxygen consumption rate.⁹⁸ When Rotenone, a potent complex II inhibitor was added with INH, there was a noticeable decrease in hepatotoxicity.⁹⁸ Interestingly, a study by Lee and colleagues also found that neither EFV nor isoniazid were able to induce cell death alone.¹⁰³ However, exposure to a combination of EFV and isoniazid resulted in increased oxidative and nitrosative stress, leading to the formation of membrane permeability transition, and ultimately necrotic cell death.⁷³ These data demonstrated that a combination of drugs causing hepatotoxicity can induce mitochondrial dysfunction, leading to necrotic cell death.

1.4.3 | Effects of drugs on the ER and autophagy

Careful analysis of cellular morphology by transmission electron microscopy (TEM) revealed autophagic degradation of the mitochondria and the ER appeared to be wrapped around the mitochondria, possibly in order to generate a membrane that would later be incorporated into the autophagic vacuoles.⁹⁵ Further evaluation of cellular morphology in Hep3B cells and primary human hepatocytes exposed to EFV by TEM revealed a change in ER morphology and the presence of granular deposits.⁹⁰ The granular deposits may represent an accumulation of unfolded proteins. Therefore, it appears that cellular damage is one of the key mechanisms responsible for hepatotoxicity induced by anti-HIV drugs, particularly EFV.

At cellular level, treating Hep3B cells with lower concentrations of EFV has been shown to induce autophagy as evidenced by the presence of autophagic vacuoles and expression of specific autophagic protein markers such as microtubule-associated protein 1A/1B light chain 3 and Beclin-1.89 Autophagy is an essential lysosomal pathway that is required for maintaining cell function and survival through degradation of proteins, cellular components, and organelles.^{89,104} However, exposing Hep3B cells to higher concentrations of EFV resulted in blockade of autophagic flux which induced autophagic stress, and ultimately promoted severe cellular damage.⁸⁹ Counterintuitively, specific inhibition of autophagy with 3-methyladenine (3MA) in Hep3B cells treated with EFV had the deleterious effect on cell survival/proliferation by promoting apoptosis, which suggests that autophagy may act as an adaptive mechanism of cell survival.⁸⁹ More work is required to elucidate the role of autophagy at cellular level, especially given that ARV regimens are changing with the introduction of new drugs to fixed does combinations.

2 | CONCLUSION

Adverse drug reactions including DILI present an enormous challenge for the treatment and management of HIV-infected patients. This is further compounded by other complexities such as the emergence of drug resistance, introduction of new multidrug regimens, and co-morbidities like TB and viral hepatitis. One cannot underscore a need for a breakthrough in diagnosis of DILI and how this will aid in the management of the adverse reactions so that patients can adhere to their treatment regimen. A good example to cite here is the success that has been achieved by taking a precision medicine approach in dealing with the abacavir hypersensitivity and implementing testing for HLA-B57:01 genotype prior to prescribing the treatment.⁸² More pharmacogenetic and pharmacokinetic studies need to be conducted to identify other critical genotypes that may be contributing to the advent of DILI in this vulnerable group of patients, particularly in light of the changing regimens and doses.

Efforts have been made over the years to achieve consensus in the definition of DILI.^{16,18,20,21} This will facilitate easier interpretation and comparison of future studies whilst also enabling researchers to design robust studies in light of the changing multidrug regimens for HIV. Gaining mechanistic insights into how ART causes DILI is essential for the identification of possible diagnostic targets and also potential therapeutic candidates that could be targeted to ameliorate the adverse reactions. Genetic variations in genes encoding for drug metabolizing enzymes (CYPs) have been associated with hepatotoxicity and hypersensitivity reactions to some ART.^{78,79,83,84,86} The mitochondrial toxicity of ART is not unexpected given the fact that the mitochondria is an energy hub of a cell and plays a critical role in the detoxification of xenobiotics. Recent mechanistic studies have demonstrated that EFV can inhibit oxidative phosphorylation,^{96,98} while triggering ER stress⁸⁸ and activation of AMPK,^{97,99,101} JNK,¹⁰¹ and BimEL¹⁰¹ signaling pathways. Although we have gained some crucial insights into the biochemical underpinnings of cytotoxicity caused by the drugs, more

work is still necessary to comprehensively unravel the critical players and lead to the identification of possible drug targets.

The immune system plays a critical role in augmenting and mediating some of the adverse reactions such as DILI and hypersensitivity reactions. Studies have incriminated polymorphisms in cytokine genes^{68,73,74} and the HLA^{75-77,79-81,86,87} that predispose-HIV-infected individuals to adverse reactions to ART. There is a paucity of studies dissecting the contribution of different immune subsets in the pathogenesis of DILI in HIV-infected patients that are on therapy. Beyond identifying the various subtypes, it would also be necessary to analyse their activation states and key regulatory circuits. Technologies such as single cell RNA sequencing could be deployed to answer these lingering scientific questions.

CONFLICT OF INTEREST

The authors have no conflict of interest to declare.

AUTHOR CONTRIBUTIONS

JP, MJM, NK, WCS, and HN: conception and design of manuscript; JP and HN: manuscript writing; JP, MJM, NK, WCS, and HN: reviewed and edited the manuscript; HN and MJM: provided financial and administrative support; HN: final approval of the manuscript.

ORCID

Hlumani Ndlovu D https://orcid.org/0000-0003-0137-1439

REFERENCES

- Global HIV & AIDS statistics. 2019 fact sheet | UNAIDS [Internet]. https://www.unaids.org/en/resources/fact-sheet.AccessedMarch13, 2020.
- Soriano V, Puoti M, Garcia-Gascó P, et al. Antiretroviral drugs and liver injury. AIDS. 2008;22(1):1-13.
- Lamar ZS, Núnez M. Higher risk of severe drug-induced liver injury among hispanic hiv-infected patients after initiation of highly active antiretroviral therapy. J Int Assoc Phys AIDS Care. 2011;10(3):183-186.
- 4. Kalyesubula R, Kagimu M, Opio KC, et al. Hepatotoxicity from first line antiretroviral therapy: an experience from a resource limited setting. *Afr Health Sci.* 2011;11(1):16-23.
- Yimer G, Gry M, Amogne W, et al. Evaluation of patterns of liver toxicity in patients on antiretroviral and anti-tuberculosis drugs: a prospective four arm observational study in ethiopian patients. *PLoS ONE*. 2014;9(4):e94271.
- Jones M, Núñez M. Liver Toxicity of Antiretroviral Drugs. Semin Liver Dis. 2012;32(2):167-176.
- Núñez M. Clinical syndromes and consequences of antiretroviral-related hepatotoxicity. *Hepatology*. 2010;52(3):1143-1155.
- Sonderup MW, Wainwright H, Hall P, Hairwadzi H, Spearman CWN. A clinicopathological cohort study of liver pathology in 301 patients with human immunodeficiency virus/acquired immune deficiency syndrome. *Hepatology*. 2015;61(5):1721-1729.
- 9. Maggiolo F. Efavirenz: a decade of clinical experience in the treatment of HIV. J Antimicrob Chemother. 2009;64(5):910-928.
- World Health Organization. WHO | Update of Recommendations on First- and Second-Line Antiretroviral Regimens. WHO; 2019.
- Zash R, Jacobson DL, Diseko M, et al. Comparative safety of dolutegravir-based or efavirenz-based antiretroviral treatment started during pregnancy in Botswana: an observational study. *Lancet Global Health*. 2018;6(7):e804-e810.

- Zash R, Holmes L, Diseko M, et al. Neural-tube defects and antiretroviral treatment regimens in Botswana. N Engl J Med. 2019;381(9):827-840.
- Reefhuis J, FitzHarris LF, Gray KM, et al. Neural tube defects in pregnancies among women with diagnosed HIV infection - 15 jurisdictions, 2013–2017. MMWR Morb Mortal Wkly Rep. 2020;69(1):1-5.
- Hernandez LV, Gilson I, Jacobson J, Affi A, Puetz TR, Dindzans VJ. Antiretroviral hepatotoxicity in human immunodeficiency virus-infected patients. *Aliment Pharmacol Ther.* 2001;15(10):1627-1632.
- den Brinker M, Wit FWNM, Wertheim-van Dillen PME, et al. Hepatitis B and C virus co-infection and the risk for hepatotoxicity of highly active antiretroviral therapy in HIV-1 infection. *AIDS*. 2000;14(18):2895-2902.
- 16. Jong E, Conradie F, Berhanu R, et al. Consensus statement: Management of drug-induced liver injury in HIV-positive patients treated for TB. S Afr. J HIV Med. 2013;14(3):113-119.
- 17. Schutz C, Ismail Z, Proxenos CJ, et al. Burden of antituberculosis and antiretroviral drug-induced liver injury at a secondary hospital in South Africa. *S Afr Med J.* 2012;102(6):506-511.
- Aithal GP, Watkins PB, Andrade RJ, et al. Case definition and phenotype standardization in drug-induced liver injury. *Clin Pharmacol Ther.* 2011;89(6):806-815.
- Njuguna C, Gunter H, Swart AM, et al. Medicines Information Centre Division of Clinical Pharmacology University of Cape Town Management of suspected drug-induced rash, kidney injury and liver injury in adult patients on TB treatment and/or antiretroviral treatment. 2018; www.mic.uct.ac.za. Accessed March 18, 2020.
- Danan G, Benichou C. Causality assessment of adverse reactions to drugs-I. A novel method based on the conclusions of international consensus meetings: Application to drug-induced liver injuries. J Clin Epidemiol. 1993;46(11):1323-1330.
- Adverse DAIDS. Event Grading Tables | DAIDS Regulatory Support Center (RSC) [Internet]. https://rsc.niaid.nih.gov/clinical-researchsites/daids-adverse-event-grading-tables. Accessed March 27, 2020.
- Watkins PB, Seeff LB. Drug-induced liver injury: Summary of a single topic clinical research conference. *Hepatology*. 2006;43(3):618-631.
- Martín-Carbonero L, Núñez M, González-Lahoz J, Soriano V. Incidence of liver injury after beginning antiretroviral therapy with efavirenz or nevirapine. *HIV Clin Trials*. 2003;4(2):115-120.
- Sanne I, Mommeja-Marin H, Hinkle J, et al. Severe hepatotoxicity associated with nevirapine use in HIV-infected subjects. J Infect Dis. 2005;191(6):825-829.
- Sonderup MW, Maughan D, Gogela N, et al. Identification of a novel and severe pattern of efavirenz drug-induced liver injury in South Africa. AIDS. 2016;30(9):1483-1485.
- Lammert C, Einarsson S, Saha C, Niklasson A, Bjornsson E, Chalasani N. Relationship between daily dose of oral medications and idiosyncratic drug-induced liver injury: search for signals. *Hepatology*. 2008;47(6):2003-2009.
- Sulkowski MS. Drug-induced liver injury associated with antiretroviral therapy that includes HIV-1 protease inhibitors. *Clin Infect Dis.* 2004;38(Supplement_2):S90-S97.
- Kandula VR, Khanlou H, Farthing C. Tipranavir: a novel second-generation nonpeptidic protease inhibitor. *Expert Rev Anti Infect Ther.* 2005;3(1):9-21.
- 29. Hicks CB, Cahn P, Cooper DA, et al. Durable efficacy of tipranavir-ritonavir in combination with an optimised background regimen of antiretroviral drugs for treatment-experienced HIV-1-infected patients at 48 weeks in the Randomized Evaluation of Strategic Intervention in multi-drug reSistant patients with Tipranavir (RESIST) studies: an analysis of combined data from two randomised open-label trials. *Lancet*. 2006;368(9534):466-475.

- BRITISH PHARMACOLOGICAL 8 of 9 PRP CASPET BRITISH PRP SOLIETY SOLIETY

- 30. Crabb C. GlaxoSmithKline ends aplaviroc trials. *AIDS*. 2006;20(5):641.
- 31. Poveda E, Briz V, Soriano V. Enfuvirtide, the first fusion inhibitor to treat HIV infection. *AIDS Rev.* 2005;7(3):139-147.
- García-Gasco P, Blanco F, Soriano V. Integrase inhibitors. J HIV Ther. 2005;10(4):75-78. Accessed March 26, 2020.
- Viani RM, Alvero C, Fenton T, et al. Safety, pharmacokinetics and efficacy of dolutegravir in treatment-experienced HIV-1 infected adolescents: forty-eight-week results from IMPAACT P1093. *Pediatric Infect Dis J.* Lippincott Williams and Wilkins. 2015;1207-1213.
- Raffi F, Rachlis A, Stellbrink HJ, et al. Once-daily dolutegravir versus raltegravir in antiretroviral-naive adults with HIV-1 infection: 48 week results from the randomised, double-blind, non-inferiority SPRING-2 study. *Lancet*. 2013;381(9868):735-743.
- Cahn P, Pozniak AL, Mingrone H, et al. Dolutegravir versus raltegravir in antiretroviral-experienced, integrase-inhibitor-naive adults with HIV: week 48 results from the randomised, double-blind, non-inferiority SAILING study. *Lancet.* 2013;382(9893):700-708.
- Safety and Efficacy of Dolutegravir in Treatment-Experienced Subjects With Raltegravir-Resistant HIV Type 1 Infection: 24-Week Results of the VIKING Study [Internet]. https://www.ncbi.nlm.nih. gov/pmc/articles/PMC3563307/. Accessed March 26, 2020.
- Zimmerman HJ. Hepatotoxicity: The Adverse Effects of Drugs and Other Chemicals on the Liver. Philadelphia, PA: Lippincott Williams & Wilkins; 1999. ISBN 0-7817-1952-6.
- Hoofnagle JH, Björnsson ES. Drug-induced liver injury types and phenotypes. N Engl J Med. 2019;381(3):264-273.
- Navarro VJ, Senior JR. Drug-related hepatotoxicity. N Engl J Med. 2006;354(7):731-739.
- Watkins PB. Idiosyncratic liver injury: challenges and approaches. Toxicol Pathol. 2005;33(1):1-5.
- 41. Reuben A, Koch DG, Lee WM. Drug-induced acute liver failure: results of a U.S. multicenter, prospective study. *Hepatology*. 2010;52(6):2065-2076.
- Sonderup MW, Wainwright HC. Human immunodeficiency virus infection, antiretroviral therapy, and liver pathology. *Gastroenterol Clin North Am.* 2017;46(2):327-343.
- Elsharkawy AM, Schwab U, McCarron B, et al. Efavirenz induced acute liver failure requiring liver transplantation in a slow drug metaboliser. J Clin Virol. 2013;58(1):331-333.
- Cotte L, Bénet T, Billioud C, et al. The role of nucleoside and nucleotide analogues in nodular regenerative hyperplasia in HIV-infected patients: a case control study. J Hepatol. 2011;54(3):489-496.
- 45. Dragovic G, Jevtovic D. The role of nucleoside reverse transcriptase inhibitors usage in the incidence of hyperlactatemia and lactic acidosis in HIV/AIDS patients. *Biomed Pharmacother*. 2012;66(4):308-311.
- Björnsson ES, Bergmann OM, Björnsson HK, Kvaran RB, Olafsson Sigurdur. Incidence, presentation, and outcomes in patients with drug-induced liver injury in the general population of Iceland. *Gastroenterology*. 2013;144(7):1419-1425.e3.
- Kullak-Ublick GA, Andrade RJ, Merz M, et al. Drug-induced liver injury: recent advances in diagnosis and risk assessment. *Gut.* 2017;66(6):1154-1164.
- Chalasani N, Bonkovsky HL, Fontana R, et al. Features and outcomes of 899 patients with drug-induced liver injury: the DILIN prospective study. *Gastroenterology*. 2015;148(7):1340-1352.e7.
- Andrade RJ, Lucena MI, Fernández MC, et al. Drug-induced liver injury: an analysis of 461 incidences submitted to the Spanish registry over a 10-year period. *Gastroenterology*. 2005;129(2):512-521.
- de Boer YS, Kosinski AS, Urban TJ, et al. Features of autoimmune hepatitis in patients with drug-induced liver injury. *Clin Gastroenterol Hepatol.* 2017;15(1):103-112.e2.

- Schmeltzer PA, Kosinski AS, Kleiner DE, et al. Liver injury from nonsteroidal anti-inflammatory drugs in the United States. *Liver Int*. 2016;36(4):603-609.
- Devarbhavi H, Karanth D, Prasanna KS, Adarsh CK, Patil M. Drug-Induced liver injury with hypersensitivity features has a better outcome: a single-center experience of 39 children and adolescents. *Hepatology*. 2011;54(4):1344-1350.
- 53. Chaponda M, Pirmohamed M. Hypersensitivity reactions to HIV therapy. Br J Clin Pharmacol. 2011;71(5):659-671.
- Luther J, Glesby MJ. Dermatologic adverse effects of antiretroviral Therapy: recognition and management. Am J Clin Dermatol. 2007;8(4):221-233.
- 55. Njuguna C, Stewart A, Mouton JP, et al. Adverse drug reactions reported to a national HIV & tuberculosis health care worker hotline in South Africa: description and prospective follow-up of reports. Drug Saf. 2016;39(2):159-169.
- 56. Peter J, Choshi P, Lehloenya RJ. Drug hypersensitivity in HIV infection. *Curr Opin Allergy Clin Immunol.* 2019;19(4):272-282.
- 57. Stewart A, Lehloenya R, Boulle A, de Waal R, Maartens G, Cohen K. Severe antiretroviral-associated skin reactions in South African patients: a case series and case-control analysis. *Pharmacoepidemiol Drug Saf.* 2016;25(11):1313-1319.
- Lehloenya RJ, Kgokolo M. Clinical presentations of severe cutaneous drug reactions in HIV-infected Africans. *Dermatol Clin.* 2014;32(2):227-235.
- Kim HN, Harrington RD, Shuhart MC, et al. Case report: hepatitis C virus activation in HIV-infected patients initiating highly active antiretroviral therapy. *AIDS Patient Care STDs.* 2007;21(10):718-723.
- 60. Aceti A, Pasquazzi C, Zechini B, et al. Hepatotoxicity development during antiretroviral therapy containing protease inhibitors in patients with HIV: the role of hepatitis B and C virus infection. J Acquir Immune Defic Syndr. 2002;29(1):41-48.
- Servoss JC, Kitch DW, Andersen JW, Reisler RB, Chung RT, Robbins GK. Predictors of antiretroviral-related hepatotoxicity in the Adult AIDS Clinical Trial Group (1989–1999). J Acquir Immune Defic Syndr. 2006;43(3):320-323.
- Wit FWNM, Weverling GJ, Weel J, Jurriaans S, Lange JMA. Incidence of and risk factors for severe hepatotoxicity associated with antiretroviral combination therapy. J Infect Dis. 2002;186(1):23-31.
- Bonfanti P, Landonio S, Ricci E, et al. Risk factors for hepatotoxicity in patients treated with highly active antiretroviral therapy. J Acquir Immune Defic Syndr. 2001;27(3):316-318.
- 64. Savès M, Raffi F, Clevenbergh P, et al. Hepatitis B or hepatitis C virus infection is a risk factor for severe hepatic cytolysis after initiation of a protease inhibitor-containing antiretroviral regimen in human immunodeficiency virus-infected patients. Antimicrob Agents Chemother. 2000;44(12):3451-3455.
- Rodriguez-Rosado R, Garcia-Samaniego J, Soriano V. Hepatotoxicity after introduction of highly active antiretroviral therapy. *AIDS*. 1998;12(10):1256.
- 66. Saves M, Vandentorren S, Daucourt V, et al. Severe hepatic cytolysis: incidence and risk factors in patients treated by antiretroviral combinations Aquitaine Cohort, France, 1996-1998. Groupe dEpidemiologie Clinique de Sida en Aquitaine (GECSA). AIDS (London, England). 1999;13(17):F115-F151.
- 67. Sulkowski MS, Thomas DL, Chaisson RE, Moore RD. Hepatotoxicity associated with antiretroviral therapy in adults infected with human immunodeficiency virus and the role of hepatitis C or B virus infection. *J Am Med Assoc.* 2000;283(1):74-80.
- de Oliveira RR, Helena Barem Rabenhorst S, de Carvalho G, et al. Association of *IL10*, *IL4*, *IFNG* and *CTLA4* gene polymorphisms with efavirenz hypersensitivity reaction in patients infected with human immunodeficiency virus. *Jpn J Infect Dis*. 2017;70(4):430-436.

- de Almeida TB, de Azevedo MCVM, da Pinto JF, et al. Drug metabolism and transport gene polymorphisms and efavirenz adverse effects in Brazilian HIV-positive individuals. J Antimicrob Chemother. 2018;73(9):2460-2467.
- Dhoro M, Ngara B, Kadzirange G, Nhachi C, Masimirembwa C. Genetic variants of drug metabolizing enzymes and drug transporter (ABCB1) as possible biomarkers for adverse drug reactions in an HIV/AIDS cohort in Zimbabwe. *Curr HIV Res.* 2014;11(6):481-490.
- Manosuthi W, Sukasem C, Lueangniyomkul A, et al. CYP2B6 haplotype and biological factors responsible for hepatotoxicity in HIVinfected patients receiving efavirenz-based antiretroviral therapy. *Int J Antimicrob Agents*. Elsevier. 2014;292-296.
- 72. Yimer G, Amogne W, Habtewold A, et al. High plasma efavirenz level and CYP2B66 are associated with efavirenz-based HAARTinduced liver injury in the treatment of naïve HIV patients from Ethiopia: a prospective cohort study. *Pharmacogenomics J.* 2012;12(6):499-506.
- Singh H, Samani D, Nema V, Ghate MV, Gangakhedkar RR. IL-1RN and IL-1β polymorphism and ARV-associated hepatotoxicity. *Mediators Inflammation*. 2018;2018.
- 74. Asensi V, Rego C, Montes AH, et al. IL-1β (+3954C/T) polymorphism could protect human immunodeficiency virus (HIV)-infected patients on highly active antiretroviral treatment (HAART) against lipodystrophic syndrome. *Genet Med.* 2008;10(3):215-223.
- Martin AM, Nolan D, Gaudieri S, et al. Predisposition to abacavir hypersensitivity conferred by HLA-B*5701 and a haplotypic Hsp70-Hom variant. Proc Natl Acad Sci USA. 2004;101(12):4180-4185.
- Illing PT, Vivian JP, Dudek NL, et al. Immune self-reactivity triggered by drug-modified HLA-peptide repertoire. *Nature*. 2012;486(7404):554-558.
- Ostrov DA, Grant BJ, Pompeu YA, et al. Drug hypersensitivity caused by alteration of the MHC-presented self-peptide repertoire. Proc Natl Acad Sci USA. 2012;109(25):9959-9964.
- Martin AM, Nolan D, James I, et al. Predisposition to nevirapine hypersensitivity associated with HLA-DRB1*0101 and abrogated by low CD4 T-cell counts. *AIDS*. 2005;19(1):97-99.
- Phillips E, Bartlett JA, Sanne I, et al. Associations between HLA-DRB1*0102, HLA-B*5801, and hepatotoxicity during initiation of nevirapine-containing regimens in South Africa. J Acquir Immune Defic Syndr. 2013;62(2):e55-e57.
- Mallal S, Phillips E, Carosi G, et al. HLA-B*5701 screening for hypersensitivity to abacavir. N Engl J Med. 2008;358(6):568-579.
- Ma JD, Lee KC, Kuo GM. HLA-B*5701 testing to predict abacavir hypersensitivity. *PLoS Curr.* 2010;2:RRN1203.
- Stainsby CM, Perger TM, Vannappagari V, et al. Abacavir hypersensitivity reaction reporting rates during a decade of HLA-B*5701 screening as a risk-mitigation measure. *Pharmacotherapy*. 2019;39(1):40-54.
- Keane NM, Pavlos RK, McKinnon E, et al. HLA class I restricted CD8⁺ and Class II restricted CD4⁺ T cells are implicated in the pathogenesis of nevirapine hypersensitivity. *AIDS*. 2014;28(13):1891-1901.
- Chantarangsu S, Mushiroda T, Mahasirimongkol S, et al. HLA-B* 3505 allele is a strong predictor for nevirapine-induced skin adverse drug reactions in HIV-infected Thai patients. *Pharmacogenet Genomics*. 2009;19(2):139-146.
- Yuan J, Guo S, Hall D, et al. Toxicogenomics of nevirapine-associated cutaneous and hepatic adverse events among populations of African, Asian, and European descent. *AIDS*. 2011;25(10):1271-1280.
- Pavlos R, McKinnon EJ, Ostrov DA, et al. Shared peptide binding of HLA Class i and II alleles associate with cutaneous nevirapine hypersensitivity and identify novel risk allele. *Sci Rep.* 2017;7(1):8653.
- Carr DF, Chaponda M, Jorgensen AL, et al. Association of human leukocyte antigen alleles and nevirapine hypersensitivity in a Malawian HIV-infected population. *Clin Infect Dis.* 2013;56(9):1330-1339.

- Apostolova N, Gomez-Sucerquia LJ, Alegre F, et al. ER stress in human hepatic cells treated with efavirenz: mitochondria again. J Hepatol. 2013;59(4):780-789.
- Apostolova N, Gomez-Sucerquia LJ, Gortat A, Blas-Garcia A, Esplugues JV. Compromising mitochondrial function with the antiretroviral drug efavirenz induces cell survival-promoting autophagy. *Hepatology*. 2011;54(3):1009-1019.
- Polo M, Alegre F, Funes HA, et al. Mitochondrial (dys)function a factor underlying the variability of efavirenz-induced hepatotoxicity? Br J Pharmacol. 2015;172(7):1713-1727.
- Brinkman K, ter Hofstede HJM, Burger DM, Smeitink JAM, Koopmans PP. Adverse effects of reverse transcriptase inhibitors: mitochondrial toxicity as common pathway. AIDS. 1998;12(14):1735-1744.
- de Mendoza C, de Ronde A, Smolders K, et al. Changes in mitochondrial DNA copy number in blood cells from HIV-infected patients undergoing antiretroviral therapy. *AIDS Res Hum Retroviruses*. 2004;20(3):271-273.
- Coghlan ME, Sommadossi J, Jhala NC, Many WJ, Saag MS, Johnson VA. Symptomatic lactic acidosis in hospitalized antiretroviral-treated patients with human immunodeficiency virus infection: a report of 12 cases. *Clin Infect Dis.* 2001;33(11):1914-1921.
- 94. Falcó V, Rodríguez D, Ribera E, et al.Severe nucleoside-associated lactic acidosis in human immunodeficiency virus-infected patients: report of 12 cases and review of the literature.
- Perier C, Tieu K, Guégan C, et al. Complex I deficiency primes Baxdependent neuronal apoptosis through mitochondrial oxidative damage. Proc Natl Acad Sci USA. 2005;102(52):19126-19131.
- Imaizumi N, Kwang Lee K, Zhang C, Boelsterli UA. Mechanisms of cell death pathway activation following drug-induced inhibition of mitochondrial complex I. *Redox Biol.* 2015;4:279-288.
- Blas-García A, Apostolova N, Ballesteros D, et al. Inhibition of mitochondrial function by efavirenz increases lipid content in hepatic cells. *Hepatology*. 2010;52(1):115-125.
- Lee KK, Boelsterli UA. Bypassing the compromised mitochondrial electron transport with methylene blue alleviates efavirenz/isoniazid-induced oxidant stress and mitochondria-mediated cell death in mouse hepatocytes. *Redox Biol.* 2014;2:599-609.
- Purnell PR, Fox HS. Efavirenz induces neuronal autophagy and mitochondrial alterations. J Pharmacol Exp Ther. 2014;351(2):250-258.
- Wu Y, Geng X-C, Wang J-F, Miao Y-F, Lu Y-L, Li B. The HepaRG cell line, a superior in vitro model to L-02, HepG2 and hiHeps cell lines for assessing drug-induced liver injury. *Cell Biol Toxicol*. 2016;32(1):37-59.
- 101. Bumpus NN. Efavirenz and 8-hydroxyefavirenz induce cell death via a JNK- and BimEL-dependent mechanism in primary human hepatocytes. *Toxicol Appl Pharmacol.* 2011;257(2):227-234.
- Lee KK, Fujimoto K, Zhang C, et al. Isoniazid-induced cell death is precipitated by underlying mitochondrial complex I dysfunction in mouse hepatocytes. *Free Radic Biol Med.* 2013;65:584-594.
- Racanelli V, Rehermann B. The liver as an immunological organ. Hepatology. 2006;43(2 SUPPL. 1):S54-S62.
- 104. Czaja MJ, Ding WX, Donohue TM, et al. Functions of autophagy in normal and diseased liver. *Autophagy*. 2013;9(8):1131-1158.
- 105. Yimer G, Amogne W, Habtewold A, et al. High plasma efavirenz level and CYP2B6*6 are associated with efavirenz-based HAART-induced liver injury in the treatment of naïve HIV patients from Ethiopia: a prospective cohort study. *Pharmacogenomics J.* 2012;12(6):499-506.

How to cite this article: Pillaye JN, Marakalala MJ, Khumalo N, Spearman W, Ndlovu H. Mechanistic insights into antiretroviral drug-induced liver injury. *Pharmacol Res Perspect*. 2020;e00598. https://doi.org/10.1002/prp2.598