



REVIEW ARTICLE

Autophagy-related lncRNAs in tumor progression and drug resistance: A double-edged sword

Yunchao Zhang ^a, Jiayu Tang ^a, Cheng Wang ^c, Qinxu Zhang ^a,
Anqi Zeng ^{b,*}, Linjiang Song ^{a,*}

^a School of Medical and Life Sciences, Chengdu University of Traditional Chinese Medicine, Chengdu, Sichuan 611137, China

^b Institute of Translational Pharmacology and Clinical Application, Sichuan Academy of Chinese Medical Science, Chengdu, Sichuan 610041, China

^c School of Pharmacy, Chengdu University of Traditional Chinese Medicine, Chengdu, Sichuan 611137, China

Received 14 March 2023; received in revised form 21 April 2023; accepted 23 April 2023

Available online 16 June 2023

KEYWORDS

Apoptosis;
Autophagy;
Biomarker;
Cancer;
Drug resistance;
lncRNA

Abstract The incidence and mortality rates of cancer are increasing every year worldwide but the survival rate of cancer patients is still unsatisfactory. Therefore, it is necessary to further elucidate the molecular mechanisms involved in tumor development and drug resistance to improve cancer cure or survival rates. In recent years, autophagy has become a hot topic in the field of oncology research, which plays a double-edged role in tumorigenesis, progression, and drug resistance. Meanwhile, long non-coding RNA (lncRNA) has also been shown to regulate autophagy, and the two-sided nature of autophagy determines the dual regulatory role of autophagy-related lncRNAs (ARlncRNAs). Therefore, ARlncRNAs can be effective therapeutic targets for various cancers. Furthermore, the high abundance and stability of ARlncRNAs in tumor tissues make them promising biomarkers. In this review, we summarized the roles and mechanisms of ARlncRNAs in tumor cell proliferation, apoptosis, migration, invasion, drug resistance, angiogenesis, radiation resistance, and immune regulation. In addition, we described the clinical significance of these ARlncRNAs, including as biomarkers/therapeutic targets and their association with clinical drugs.

© 2023 The Authors. Publishing services by Elsevier B.V. on behalf of KeAi Communications Co., Ltd. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

* Corresponding author.

E-mail addresses: zeng6002aq@163.com (A. Zeng), songlinjiang@cdutcm.edu.cn (L. Song).

Peer review under responsibility of Chongqing Medical University.

Introduction

The incidence and fatality rates of cancer are rising year by year.¹ Notwithstanding a conflation of surgical intervention, chemotherapeutic agents, radiotherapeutic measures, and targeted molecular therapy, the efficacy of these treatments in ameliorating the survival prospects of several cancers remain subpar. Timely detection and intervention can potentially augment the probability of recovery or survival. Consequently, additional molecular network studies are imperative to completely explicate the pathogenic mechanisms underlying the genesis and progression of cancers, as well as drug resistance, and to discern novel biomarkers and therapeutic targets.^{2–10}

Non-coding RNAs that are longer than 200 nucleotides are called long non-coding RNAs (lncRNAs), which can be involved in the physiopathological processes of various tumors.^{11,12} Their roles are exerted by several mechanisms involving: i) lncRNAs regulate target genes downstream of miRNAs by competitively adsorbing and down-regulating miRNAs through base complementation. ii) lncRNAs can bind epigenetically related proteins to modify the post-transcriptional translation of specific proteins. iii) lncRNAs can directly bind chromosomal DNA to repress gene transcription or enhance gene transcription by attracting transcription factors. iv) lncRNAs can act as precursors to miRNAs. v) lncRNAs can interact with proteins and affect their activity. vi) lncRNAs can interact with genomic DNA to form triplexes thereby regulating target gene transcription.^{13–16}

There exist three distinct forms of autophagy, namely, macroautophagy, microautophagy, and chaperone-mediated autophagy.^{17–19} In our study, the term autophagy pertains specifically to macroautophagy, which is extensively researched and holds a significant functional position.¹³ The generation of autophagosomes encompasses four principal phases, namely, initiation, phagocyte nucleation, elongation and closure of autophagosomes, and fusion of autolysosomes.²⁰ These processes involve the participation of several autophagy-related gene (ATG) proteins. Among mammals, there are five core ATG protein subgroups: the Vps34-Beclin 1 class III PI3 kinase complex, Unc-51-like autophagy-activating kinase 1 (ULK1) protein kinase complex, ATG12 coupling system, ATG9-WIPI-1 complex, and the microtubule-associated protein 1 light chain 3 (LC3) coupling system.²¹

Interestingly, autophagy plays a double-edged role in tumorigenesis, progression, and drug resistance. During the nascent phases of neoplasms, autophagy exerts an inhibitory effect on tumorigenesis by enzymatically decomposing oncogenic elements; however, as the malignancies advance, autophagy shields the tumor cells by ameliorating the stressful conditions present in the microenvironment of the tumor.^{6,22} Many complex mechanisms are involved, including the regulation of autophagy-related lncRNAs (ARlncRNAs). The two-sided nature of autophagy determines the dual regulatory role of ARlncRNAs.^{17,23–27} Thus, ARlncRNAs can serve as effective therapeutic targets for a variety of cancers. Additionally, the high abundance of ARlncRNAs in tumor tissues and their circulating stability make them promising biomarkers for the diagnosis and

prognosis of various tumors.^{9,28} In this review, we summarize the roles and mechanisms of ARlncRNAs in tumor cell proliferation and apoptosis, migration, and invasion, drug resistance, angiogenesis, radioresistance, and immune regulation. In addition, we describe the clinical significance of these ARlncRNAs, including as biomarkers/therapeutic targets and their association with clinical pharmacotherapy (Fig. 1).

ARlncRNAs promote tumor drug resistance

ARlncRNAs activate autophagy to promote tumor drug resistance

Numerous ARlncRNAs exhibit up-regulation in diverse carcinomas and foster resistance to chemotherapeutic agents via a variety of mechanisms. The ARlncRNA/miRNA/ATG axis orchestrates the regulation of autophagy-mediated drug resistance, whereby ATG5 serves as a crucial mediator of autophagy. In gastric cancer (GC), the ARlncRNA MALAT1 intensifies cisplatin resistance through the stimulation of autophagy, by down-regulating miR-30b and concomitantly up-regulating ATG5. Unexpectedly, Hu et al found that MALAT1 knockdown exerted no effect on ATG5 mRNA levels in SGC7901/VCR cells.²⁹ The discrepancy between the results of these two reports may be due to the intracellular environment.^{29,30} In contrast, ARlncRNA FEZF1-AS1 specifically targets the up-regulation of ATG5 to increase multi-drug resistance in GC.³¹ ATG7 is an E1-like protease that activates the autophagic ubiquitin-like proteins ATG8 and ATG12. ARlncRNA BLACAT1 promotes ATG7 expression via miR-17, thereby promoting autophagy and facilitating chemoresistance in non-small cell lung cancer (NSCLC).³² ATG14 can act as a potent activator of autophagy. By competitively connecting to miR-188-3p, EIF3J-DT increases the stability of ATG14 mRNA and inhibits its degradation. Consequently, EIF3J-DT enhances the transcription of ATG14 and leads to activating autophagy and chemoresistance.³³ The C-terminal peptide of LC3 protein requires cleavage by mammalian ATG4 homologs, thereby promoting double-membrane autophagosome formation. ARlncRNA KCNQ10T1 targets the miR-34a/ATG4B to enhance oxaliplatin chemoresistance in colon cancer.³⁴

ULK1 is an autophagy-associated gene and a target of miRNA. The lncRNA/miRNA/ULK1 axis can regulate autophagy-mediated drug resistance. ARlncRNA LUCAT1 promotes cisplatin resistance in NSCLC via controlling the miR-514a-3p/ULK1.³⁵ In colorectal cancer (CRC), the ARlncRNA SNHG6 facilitates resistance to 5-Fluorouracil (5-Fu) by sequestering miR-26a-5p and thus augmenting ULK1-mediated autophagy.¹⁴ The ARlncRNA Sox2OT-V7 exerts a dual mechanism to up-regulate ULK1 by acting as a miRNA sponge for either miR-142 or miR-204, thereby augmenting autophagy and contributing to drug resistance. Additionally, ULK phosphorylates Beclin-1 after amino acid withdrawal.²¹ Beclin1 is an essential part of autophagosome formation, and up-regulation of Beclin1 activates autophagy. ARlncRNA HOTAIR down-regulates miR-17-5p expression and enhances Beclin1 expression in RC cells, thus promoting cellular autophagy.³⁶

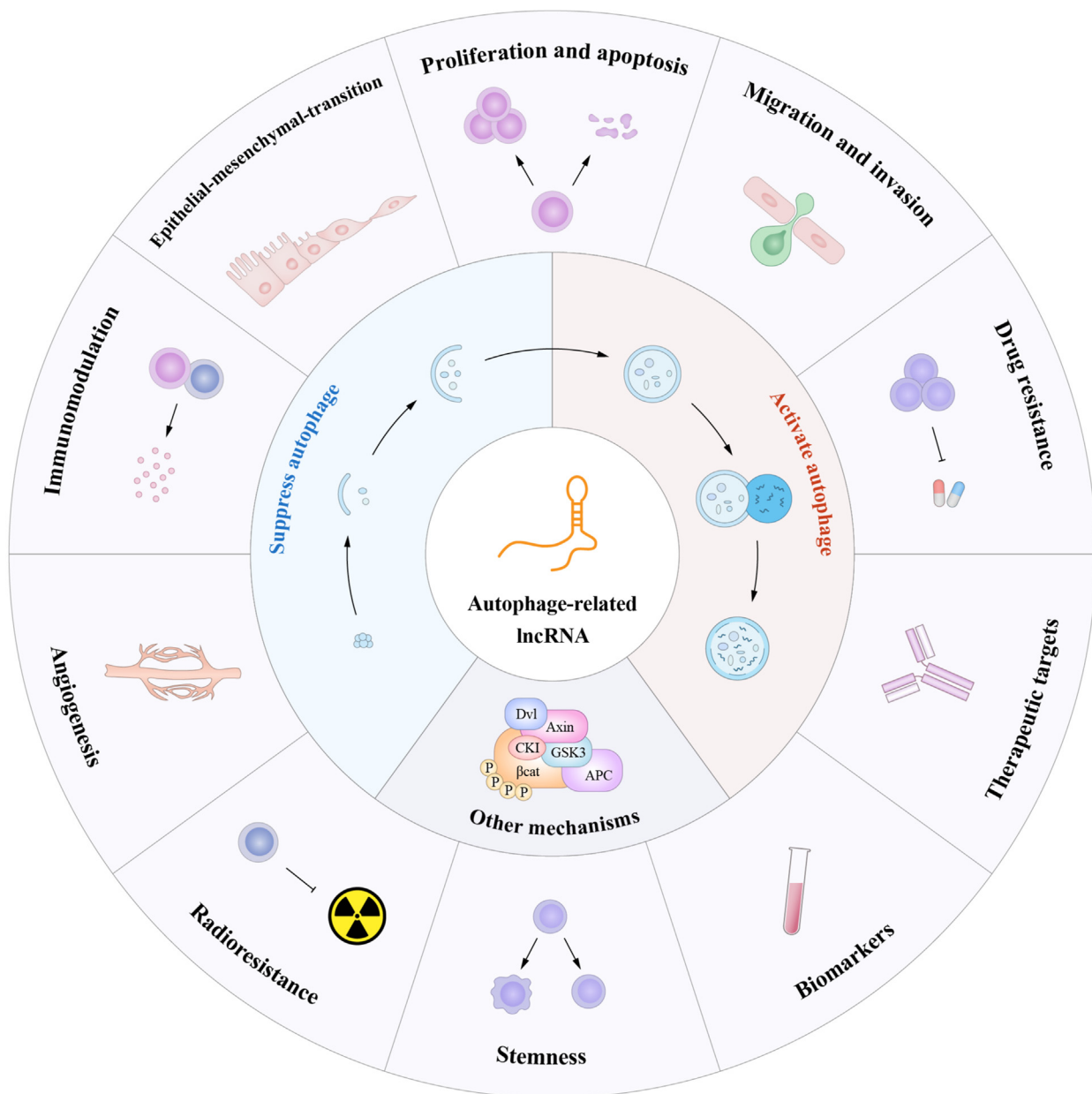


Figure 1 Mechanism, function, and clinical significance of ARlncRNAs.

Drug resistance is not only affected by autophagy but also regulated by other signaling pathways. Some ARlncRNAs can dual-regulate drug resistance through autophagy and other signaling pathways. The ARlncRNA PVT1 is known to augment gemcitabine resistance in pancreatic cancer by coordinated regulation of the miR-619-5p/Pygo2/Wnt/ β -catenin and miR-619-5p/ATG14 pathways. Additionally, there exists a positive feedback loop between the Wnt/ β -catenin signal and PVT1 transcription. It remains to be determined whether this positive feedback loop also plays a role in conferring chemoresistance in other types of cancer.¹³ The DNA damage response is a crucial mechanism for preserving genomic stability. ARlncRNA OTUD6B-AS1 increases the formation of paclitaxel resistance in triple-negative breast cancer by inhibiting DNA repair (up-regulating genomic instability) and promoting autophagy.

Nonetheless, the mechanism by which metadherin regulates autophagy and DNA damage response-related protein stimulation requires more investigation.³⁷

ARlncRNAs inhibit autophagy to promote tumor drug resistance

Conversely, ARlncRNA HOXA11-AS, ROR can promote drug resistance by inhibiting autophagy and thereby promoting drug resistance. So, they can be used as drugs by themselves or as therapeutic targets to improve the outcome of cancer patients. Researchers performed lentiviral transfection of ovarian cancer cells and showed that HOXA11-AS knockdown significantly inhibited tumor cell motility and promoted cellular autophagy and sensitivity to cisplatin.³⁸

The ARlncRNA ROR has been implicated in conferring drug resistance and promoting invasiveness in breast cancer cells.³⁹ Further investigations revealed that the inhibition of ROR significantly augmented the sensitivity of breast cancer cells to tamoxifen by promoting autophagy (up-regulation of LC3 and Beclin 1 expression).⁴⁰ The AKT/mammalian target of rapamycin (mTOR) pathway is considered to be an important signal for the regulation of autophagy.⁴¹ In NSCLC, silencing ROR can reverse cisplatin resistance by inhibiting phosphatidylinositol-3-kinase (PI3K)/protein kinase B (AKT)/mTOR signaling.⁴²

ARlncRNAs inhibit tumor drug resistance

ARlncRNAs activate autophagy to inhibit tumor drug resistance

These ARlncRNAs that impede drug resistance are frequently found to exhibit low expression levels in drug-resistant tumor tissues. This finding holds significant importance in the selection and application of autophagy-targeted therapeutics. The ARlncRNA EGOT-enhanced autophagy sensitizes paclitaxel cytotoxicity by up-regulating inositol 1,4,5-trisphosphate receptor 1 (ITPR1). Mechanistically, EGOT increases autophagosome accumulation through cis- and trans-up-regulating ITPR1. First, EGOT causes pre-ITPR1 accumulating to up-regulate ITPR1 levels in cis by generating pre-ITPR1/EGOT dsRNA. EGOT engages heterogeneous nuclear ribonucleoproteins to increase alternative splicing of pre-ITPR1 in trans via two binding motifs in fragment 2 of exon 1 of EGOT. Researchers also found that hypoxia can up-regulate EGOT, while estrogen directly inhibits it.⁴³ The ARlncRNA GAS5 inhibits tumor growth by inducing autophagy in breast cancer cells. Overexpression of GAS5 up-regulates ULK1/2 in MCF-7 cells without interfering with other autophagy initiation-related proteins or decreasing cell proliferation, migration, or tumor development. The researchers hypothesize that the antitumor effects of GAS5 overexpression are at least partially mediated by autophagy induction. Nevertheless, the mechanism through which GAS5 acts as a negative regulator independent of autophagy is unclear.⁴⁴

ARlncRNAs inhibit autophagy to inhibit tumor drug resistance

The ARlncRNA ACTA2-AS1 suppresses cisplatin resistance in NSCLC via engaging enhancer of Zeste 2 (EZH2) to the TSC2 gene promoter and decreasing tuberous sclerosis complex-2 (TSC2) expression.⁴⁵ By suppressing autophagy, the E2F6-regulated ARlncRNA CRNDE sensitizes GC cells to chemotherapy. Mechanistically, overexpression of CRNDE suppresses autophagy and promotes apoptosis, making GC cells more sensitive to chemotherapeutic treatments. In addition, the classical transcriptional inhibitor E2F6 was shown to up-regulate and suppress CRNDE expression in GC.⁴⁶ CRNDE can also disrupt autophagy-mediated chemoresistance by interacting with and decreasing the protein stability of the downstream target gene serine/arginine-rich splicing factor 6 (SRSF6) and thus decreasing the

selective splicing of phosphatidylinositol binding clathrin-assembly protein (PICALM) mRNA.⁴⁷ The ARlncRNA MEG3 has been observed to enhance sensitivity to vincristine by impeding autophagy in the context of lung cancer chemotherapy. Researchers have demonstrated that MEG3 expression was substantially down-regulated in drug-resistant cells compared to non-resistant cells, as seen *in vitro*. As a result, overexpression of MEG3 markedly suppressed the viability and proliferation of both drug-resistant and non-resistant lung cancer cells.⁴⁸ More information and mechanisms of ARlncRNAs regulating tumor drug resistance are shown in Table 1 and Figure 2.

ARlncRNAs promote tumor cell proliferation and inhibit apoptosis

ARlncRNAs activate autophagy to promote tumor cell proliferation and inhibit apoptosis

Tumor-promoting ARlncRNAs tend to be up-regulated in tissues. The mechanistic target of rapamycin complex 1 (mTORC1) is a major positive modulator of cell proliferation and growth.⁴⁹ Partition-defective 3 (PARD3) is located downstream of mTORC1 and AMP-activated protein kinase (AMPK) and senses amino acid and energy signaling to trigger the initiation of autophagy. ARlncRNA SLCO4A1-AS1 can down-regulate miR-508-3p and thus up-regulate PARD3 expression to promote autophagy and proliferation of CRC cells.⁷ The ARlncRNA MALAT1 can sponge miR-204 in GC to up-regulate transient receptor potential melastatin 3 (TRPM3). TRPM3 has been shown to modulate the expression of LC3A and LC3B, which can activate oncogenic autophagy and facilitate cancer progression. Moreover, the MALAT1/miR-204/LC3B signaling pathway plays a crucial role in *H. pylori*-induced gastric cancer and is involved in the regulation of autophagy during infection. Further research will focus on elucidating the molecular mechanisms by which this axis regulates autophagy and contributes to *H. pylori*-induced tumorigenesis.⁵ In addition, MALAT1 activates autophagy to promote cell proliferation and reduce apoptosis by down-regulating miR-101 expression in CRC cell lines.¹¹ In multiple myeloma, MALAT-1 increases high mobility group box-1 (HMGB1) to enhance autophagy, hence inhibiting cancer cell death.⁵⁰ In NSCLC patients, a disintegrin and metalloproteinase 28 (ADAM28) has been reported to relate to tumor growth and lymphatic metastasis. ARlncRNA NEAT1 induces autophagy by stimulating the Janus kinase (JAK)/signal transducer and activator of transcription (STAT)3 signaling cascade via the miR-128-3p/ADAM28 axis in NSCLC. However, it could not be determined if ADAM28 directly promotes the activation of the JAK2/STAT3 or indirectly regulates the JAK2/STAT3 by targeting other proteins or RNAs (e.g., lncRNA, miRNA, or circRNA). This is a question that must be investigated in future research.⁵¹

ARlncRNAs inhibit autophagy to promote tumor cell proliferation and inhibit apoptosis

Wnt/ β -catenin and PI3K/AKT are essential signals that govern cell growth and metastasis. mTOR, which is

Table 1 ARlncRNAs that regulate tumor drug resistance.

lncRNA	Levels	Cancer Type	Autophagy	Mechanisms of Autophagy	Drug	Drug resistance	Reference
EGOT	↓	Breast cancer	Activates	ITPR1 ↑	Paclitaxel	Inhibits	43
GAS5	↓	Breast cancer	Activates	ULK1/2 ↑	Cisplatin	Inhibits	44
TUG1	↓	Non-small cell lung cancer	Activates	miR-221/PTEN	Cisplatin	Inhibits	88
ACTA2-AS1	↓	Non-small cell lung cancer	Suppresses	Recruits EZH2 to TSC2 gene promoter	Cisplatin	Inhibits	45
CRNDE	↓	Gastric cancer	Suppresses	E2F6-CRNDE axis	Oxaliplatin/5-Fu	Inhibits	46
MEG3	↓	Lung cancer	Suppresses	—	Vincristine	Inhibits	48
CRNDE	↓	Gastric cancer	Suppresses	SRSF6/PICALM	5-FU/oxaliplatin	Inhibits	47
H19	↑	Colorectal cancer	Activates	miR-194-5p/SIRT1	5-Fu	Promotes	89
NEAT1	↑	Colorectal cancer	Activates	miR-34a/HMGB1/ATG9A/ATG4B	5-Fu	Promotes	90
SNHG6	↑	Colorectal cancer	Activates	miR-26a-5p bind to SNHG6 and target ULK1	5-Fu	Promotes	14
UCA1	↑	Colorectal cancer	Activates	miR-23b-3p/ZNF281 Axis	5-Fu	Promotes	91
XIST	↑	Ovarian cancer	Activates	miR-506-3p/FOXPI/AKT/mTOR	Carboplatin	Promotes	41
HULC	↑	Gastric cancer	Activates	FoxM1	Cisplatin	Promotes	80
LUCAT1	↑	Non-small cell lung cancer	Activates	miR-514a-3p/ULK1	Cisplatin	Promotes	35
MALAT1	↑	Gastric cancer	Activates	microRNA-30b/ATG 5	Cisplatin	Promotes	29
TUG1	↑	Colorectal cancer	Activates	miR-195-5p/HDGF/DDX5/β-catenin	Cisplatin	Promotes	92
BLACAT1	↑	Non-small cell lung cancer	Activates	miR-17/ATG7	Cisplatin	Promotes	32
HOTAIR	↑	Non-small cell lung cancer	Activates	ULK1 ↑	Crizotinib	Promotes	93
FEZF1-AS1	↑	Gastric cancer	Activates	ATG5	5-FU/cisplatin	Promotes	31
PCDRlnc1	↑	Prostate cancer	Activates	UHRF1 protein/Beclin-1	Docetaxel	Promotes	94
Sox2OT-V7	↑	Osteosarcoma	Activates	miR-142/ULK1, ATG4A, and ATG5	Doxorubicin	Promotes	21
GBCDRlnc1	↑	Gallbladder cancer	Activates	miR-22/ULK1	Doxorubicin	Promotes	20
MITA1	↑	Non-small-cell lung cancer	Activates	PGK1/ATG5-ATG12 conjugate.	Gefitinib	Promotes	95
ANRIL	↑	Pancreatic cancer	Activates	—	Gemcitabine	Promotes	96
PVT1	↑	Pancreatic cancer	Activates	miR-181a/HMGB1	Gemcitabine	Promotes	13
HOTAIR	↑	Gastrointestinal stromal tumor	Activates	miR-619-5p/ATG14	Imatinib	Promotes	97
KCNQ10T1	↑	Colon cancer	Activates	miR-130a/ATG2B	Oxaliplatin	Promotes	34
EIF3J-DT	↑	Gastric cancer	Activates	miR-34a/Atg4B	Oxaliplatin/5-Fu	Promotes	33
OTUD6B-AS1	↑	Triple negative breast cancer	Activates	ATG14	Paclitaxel	Promotes	37
HOTAIR	↑	Renal cancer	Activates	miR-26a-5p/MTDH	Sunitinib	Promotes	36
HOXA11-AS	↑	Ovarian cancer	Suppresses	miR-17-5p/Beclin1	Cisplatin	Promotes	38
ROR	↑	Breast cancer	Suppresses	ATG	Tamoxifen	Promotes	40
				LC3 and Beclin 1 ↑			

positioned downstream of the PI3K/AKT, is a negative key controller for autophagy. By regulating EZH2, ARlncRNA SNHG1 stimulates Wnt/β-catenin and PI3K/AKT/mTOR to promote prostate cancer cell growth and inhibit apoptosis and autophagy.⁵² In oral squamous cell carcinoma, over-expression of ARlncRNA CASC9 can suppress autophagy-mediated apoptosis through the AKT/mTOR, hence boosting tumor growth.⁵³ The oncogenic factor paired-box 6 (PAX6) of BC belongs to a family of cassette transcription factors

that can influence the behavior of tumor cells. The ARlncRNA DANCER/miR-758/3p-PAX6 axis can inhibit autophagy and apoptosis in mammary cancer cells.⁸ Activation of recombinant nuclear factor I/B (NFIB), one of the main proteins affecting tumor cell differentiation, may affect EC progression. ARlncRNA DRAIC/miR-149-5p/NFIB axis can inhibit autophagy and apoptosis in esophageal cancer.⁵⁴ FK506-binding protein 4 (FKBP4) is a member of the pro-immune protein family, and FKBP4 plays a role in immune

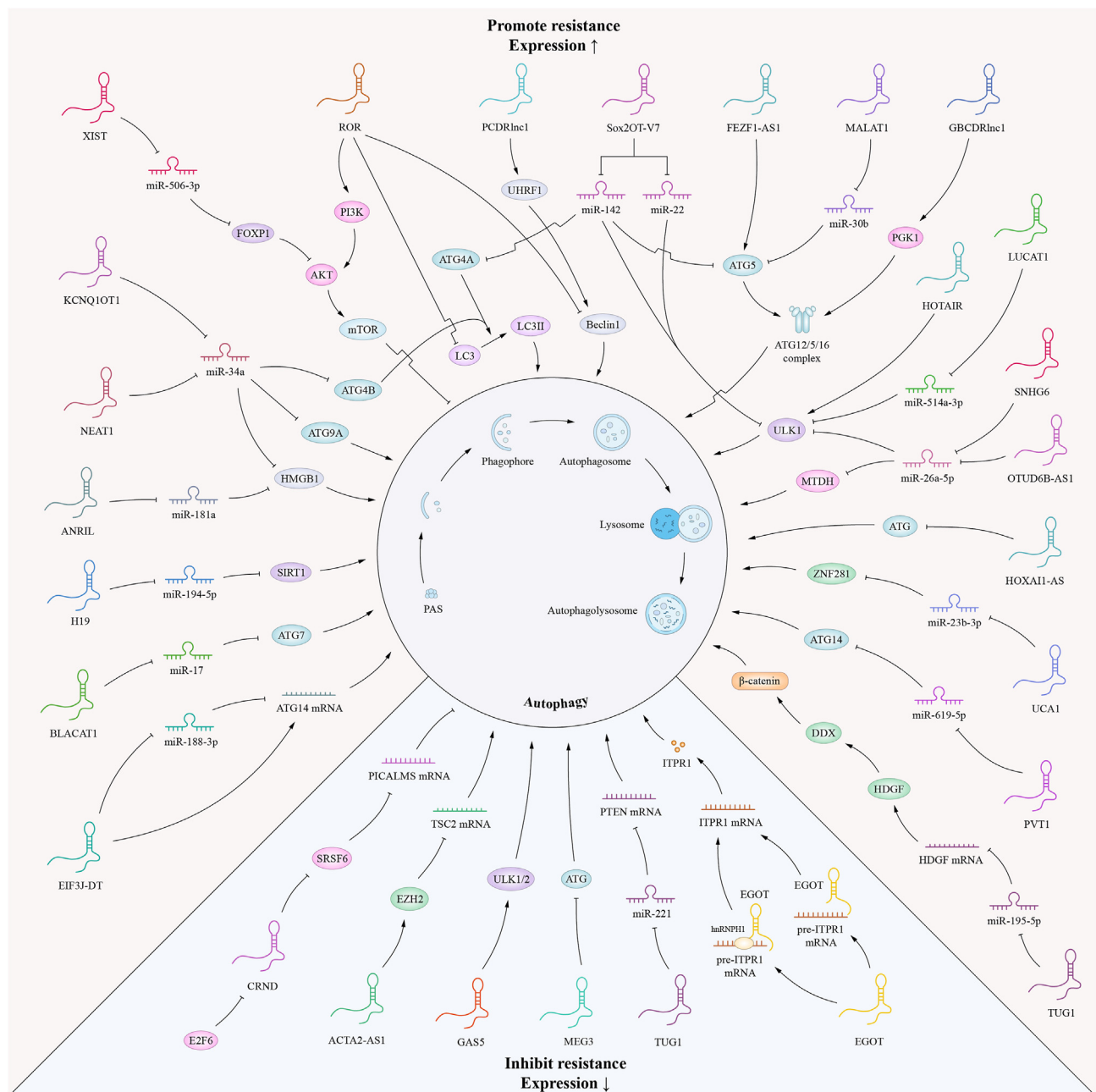


Figure 2 Dual regulatory mechanisms of ARlncRNAs in tumor drug resistance.

regulation and essential cellular processes involving protein folding and transport. ARlncRNA MAFG-AS1 can sponge miR-3612 to increase the expression of FKBP4, which activates cell proliferation and inhibits apoptosis and autophagy.⁵⁵

ARlncRNAs inhibit tumor cell proliferation and promote apoptosis

ARlncRNAs activate autophagy to inhibit tumor cell proliferation and promote apoptosis

These tumor suppressor ARlncRNAs tend to be down-regulated in tissues. Activating transcription factor 2 (ATF2) is

critical in cell development and survival. Researchers found that ATF2 binds to the promoter region of ARlncRNA GAS8-AS1 and activates its expression. GAS8-AS1 could promote papillary thyroid cell autophagy via miR-187-3p/ATG5 and miR-1343-3p/ATG7 thereby inhibiting cancer cell proliferation. However, papillary thyroid patients have various kinds of GAS8-AS1 mutations. Notably, the mutant GAS8-AS1 did not modulate miR-187-3p and miR-1343-3p. The reason for this result may be the difference in secondary structure between the mutant and wild-type GAS8-AS1.^{56,57}

Nuclear factor-kappaB (NF- κ B) is associated with gene transcription in immunity, inflammation, and proliferation. It has been demonstrated that several carcinogens increase proliferation and anti-apoptosis by activating NF- κ B.

Therefore, NF- κ B is anticipated to be a novel cancer therapeutic target. ARlncRNA CASC2 was detected to be lowly expressed in colon cancer cells and inhibited cell viability by down-regulating miR19a and inhibiting the NF- κ B/p65, suggesting that CASC2 may be a prospective factor for the therapeutic therapy of colon cancer.⁵⁸ Tripartite motif protein 16 (TRIM16) is a positive transcriptional regulator of retinoic acid receptor β 2 in retinoid-treated cancer cells and can act as a tumor suppressor.⁵⁹ CASC2 also causes apoptosis and autophagy in colon cancer by regulating the levels of TRIM16 via miR-214.⁶⁰ Interestingly, CASC2 in NSCLC similarly regulates the miR-214/TRIM16 but results in the inhibition of autophagy and promotion of apoptosis in NSCLC.⁵⁹ This different role of ARlncRNAs in different tumors through the same signaling pathway may be caused by differences in the tumor microenvironment and deserves further investigation.

ARlncRNAs inhibit autophagy to inhibit tumor cell proliferation and promote apoptosis

The ARlncRNA CLRN1-AS1 acts as a tumor suppressor in pituitary prolactinomas and inhibits autophagy by suppressing the Wnt/ β -catenin. Functionally, CLRN1-AS1 inhibits cell proliferation, promotes apoptosis, and suppresses autophagy. Mechanistically, CLRN1-AS1 up-regulates dickkopf WNT signaling pathway inhibitor 1 (DKK1) by sponging miR-

217. In addition, they found that the transcriptional repressor upstream of CLRN1-AS1 is Forkhead box P1 (FOXP1).⁶¹ Similarly, the ARlncRNA MEG3 inhibits autophagy and induces apoptosis by sponging miR-93 to down-regulate the PI3K/AKT/mTOR axis in BC progression.⁶² Interestingly, PTC cells can transmit ARlncRNA SNHG9 to neighboring normal cells via exosomes. Consequently, it can suppress the production of the YBOX3 protein, negatively regulate P21, reduce autophagy, and increase death in normal cells. However, this must be validated in animals and the mechanism by which SNHG9 triggers YBOX3 degradation must be elucidated.⁶³ More information and mechanisms of ARlncRNAs regulating tumor proliferation and apoptosis are shown in Table 2 and Figure 3.

ARlncRNAs promote tumor migration and invasion

ARlncRNAs activate autophagy to promote tumor migration and invasion

ARlncRNAs that promote tumor cell migration and invasion are up-regulated in tumor tissues. Matrix infiltration is one of the most important steps in cancer metastasis. Matrix metalloproteinases (MMPs) are a class of zinc-dependent protein hydrolases, and MMP2 mainly hydrolyzes type IV collagen (a key element of the basement membrane) to

Table 2 ARlncRNAs that regulate tumor proliferation and apoptosis.

LncRNA	Levels	Cancer Type	Autophagy	Mechanisms of Autophagy	Role	Reference
LCPAT1	↑	Lung cancer	Activates	—	Oncogene	6
MALAT1	↑	Gastric cancer	Activates	miR-204↓	Oncogene	5
MALAT 1	↑	Colorectal cancer	Activates	miR-101↓	Oncogene	11
MALAT-1	↑	Multiple myeloma	Activates	HMGB1↑	Oncogene	50
NEAT1	↑	Non-small-cell lung cancer	Activates	miR-128-3p/ADAM28	Oncogene	51
SLCO4A1-AS1	↑	Colorectal cancer	Activates	miR-508-3p/PARD3	Oncogene	7
SNHG8	↑	Colorectal cancer	Activates	miR-588/ATG7	Oncogene	98
UCA1	↑	Colorectal cancer	Activates	miR-185-5p-WISP2-Wnt/ β -catenin	Oncogene	99
MSTO2P	↑	Lung cancer	Activates	EZH2↑	Oncogene	100
CASC9	↑	Oral squamous cell carcinoma	Suppresses	AKT/mTOR	Oncogene	53
DANCR	↑	Breast cancer	Suppresses	miR-758-3p/PAX6	Oncogene	8
DRAIC	↑	Esophageal cancer	Suppresses	miR-149-5p/NFIB	Oncogene	54
EGOT	↑	Colon cancer	Suppresses	miR-33a-5p and miR-33b-5p	Oncogene	9
KTN1-AS1	↑	Non-small cell lung cancer	Suppresses	miR-130a-5p/PDPK1	Oncogene	81
LINC00858	↑	Colon cancer	Suppresses	activates WNK2 promoter methylation	Oncogene	101
LINC01207	↑	Pancreatic cancer	Suppresses	miR-143-5p/AGR2	Oncogene	102
MAFG-AS1	↑	Breast cancer	Suppresses	miR-3612/FKBP4	Oncogene	55
PRRT3-AS1	↑	Prostate cancer	Suppresses	PPAR γ /mTOR	Oncogene	103
RHPN1-AS1	↑	Prostate cancer	Suppresses	miR-7-5p/EGFR/PI3K/AKT/mTOR	Oncogene	104
SNHG1	↑	Prostate cancer	Suppresses	Wnt/ β -Catenin and PI3K/AKT/mTOR	Oncogene	52
CASC2	↓	Colon cancer	Activates	miR19a↓	Suppressor	58
CASC2	↓	Colon cancer	Activates	microRNA-214/TRIM16	Suppressor	60
GAS8-AS1	↓	Thyroid Cancer	Activates	miR-187-3p/ATG5 and miR-1343-3p/ATG7	suppressor	57
GAS8-AS1	↓	papillary thyroid cancer	Activates	ATG5↑	suppressor	56
CASC2	↓	non-small cell lung cancer	Suppresses	miR-214/TRIM16	suppressor	59
CLRN1-AS1	↓	pituitary prolactinoma	Suppresses	miR-217/DKK1/Wnt/ β -catenin	suppressor	61
MEG3	↓	bladder cancer	Suppresses	PI3K/AKT/mTOR	suppressor	62

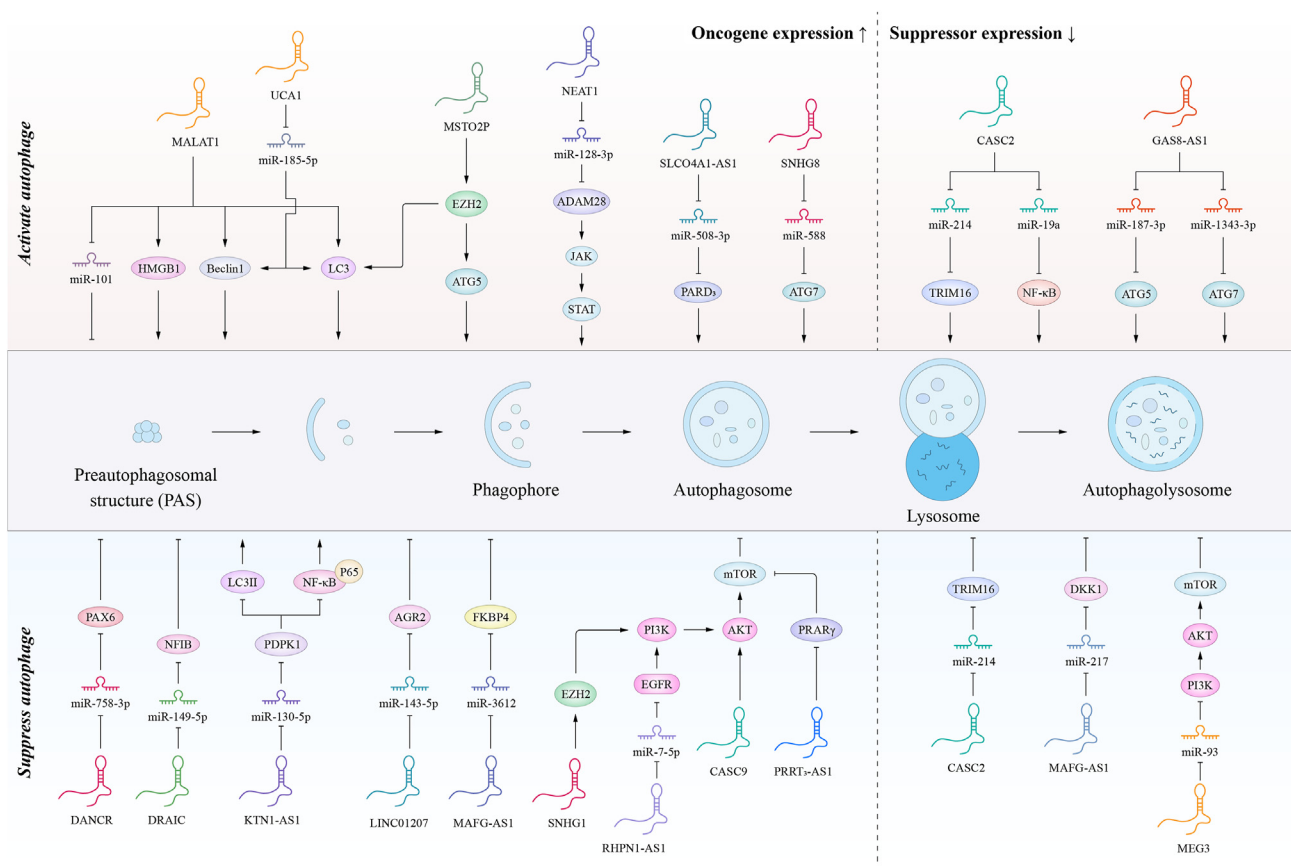


Figure 3 The “double-edged sword” mechanism of ARlncRNAs in tumor proliferation and apoptosis.

facilitate cancer cell invasion and spread. Autophagy stimulation by ARlncRNA SNHG1 increases the migration of basal bladder cancer. On the one hand, SNHG1 directly binds to the PP2A catalytic subunit to inhibit its interaction with c-Jun, which then promotes c-Jun phosphorylation and thus mediates MMP2 transcription; on the other hand, SNHG1 initiates autophagy by inducing autophagy-associated protein abundance and further destabilizes miR-34a, thereby decreasing miR-34a binding to the 3' UTR of MMP2 mRNA, thus promoting the stabilization of MMP2 mRNA.⁶⁴

The Wnt/ β -catenin pathway is recognized as carcinogenic. Functionally, the ARlncRNA SNHG11 promotes cell proliferation, stemness, metastasis, and epithelial–mesenchymal transition in GC via enhancing autophagy. Mechanistically, SNHG11 increases the expression of the transcription of catenin β 1 (CTNNB1) and ATG12 via miR-483-3p/miR-1276 while inhibiting the processing of pre-miR-483/pre-miR-1276. SNHG11 interacts with Cullin 4A (CUL4A) and further stimulates the Wnt/ β -catenin pathway by inducing glycogen synthase kinase 3 (GSK-3) ubiquitination. Intriguingly, SNHG11 regulates autophagy via the ATG12 pathway and not the Wnt/ β -catenin pathway, but it increases malignant activity in GC cells via both channels.¹⁷

ARlncRNAs inhibit autophagy to promote tumor migration and invasion

ARlncRNA CDKN2B-AS1 enhances hepatocellular carcinoma invasion, proliferation, and migration by targeting and inhibiting miR-199a-5p to suppress apoptosis and autophagy, which is expected to be a breakthrough for future diagnosis and treatment. However, the number of patients included in this study was small, and only blood samples were obtained from liver cancer patients for testing. More *in vivo* and *ex vivo* experiments are needed for validation in the future.⁶⁵ STAT3 can induce ARlncRNA HAGLROS overexpression and thus activate mTOR signaling to inhibit autophagy and promote malignant progression in GC cells. HAGLROS regulates mTOR signaling in two ways. On the one hand, HAGLROS can down-regulate miR-100-5p to stimulate mTOR mRNA expression. On the other hand, HAGLROS interacts with mTORC1 to activate mTORC1 signaling to negatively regulate autophagy.⁴⁹ ARlncRNA ADAMTS9-AS1 can also up-regulate mTOR via PI3K/AKT, suppressing the apoptosis and autophagy of bladder cancer cells while boosting their invasion and migration.⁶⁶ In addition, the newly found lncRNA-45 stimulates the mTOR signal to enhance breast tumor progression.⁶⁷

ARlncRNAs inhibit tumor migration and invasion

ARlncRNAs activate autophagy to inhibit tumor migration and invasion

These ARlncRNAs that inhibit tumor migration and invasion tend to be up-regulated in expression. Beclin1 belongs to the first autophagy genes discovered in mammals, and it primarily governs autophagosome formation by building complexes with PI3K. Beclin1 could modulate growth factor signalings, such as the AKT and extracellular signal-regulated kinase routes. ARlncRNA GAS8-AS1 suppresses the development of ovarian cancer by triggering the Beclin1-mediated autophagy pathway.²³ ARlncRNA PTCSC3 inhibits human oral cancer cell proliferation by inducing LC3B-II and Beclin 1 to promote autophagy and up-regulate apoptosis-associated proteins such as Bax.⁶⁸ ARlncRNA ADAMTS9-AS2 promotes autophagy and inhibits the proliferation, migration, and invasion of hepatocellular carcinoma cells by suppressing the PI3K/AKT/mTOR axis. The phosphorylation of AKT and mTOR, the autophagy protein SQSTM1, and the anti-apoptotic protein Bcl-2 were reduced by ADAMTS9-AS2. In contrast, ADAMTS9-AS2 elevated autophagy protein LC3-II, pro-apoptotic protein Bax, and the autophagy regulator Beclin 1. However, this experiment was only conducted on two cell lines; therefore, its experimental outcomes are limited. Further work is required to determine whether the effects of ADAMTS9 on various cellular characteristics are attributable to its impact on the AKT/mTOR.⁶⁹

ARlncRNAs inhibit autophagy to inhibit tumor migration and invasion

A study found that ARlncRNA CPS1-IT1 inhibited the growth and metastasis of hepatocellular carcinoma by reducing HIF-1 α activation and inhibiting epithelial–mesenchymal transition.⁷⁰ Similarly, one study found that ARlncRNA CPS1-IT1 also inhibits cell proliferation, invasion, and metastasis in colorectal cancer.⁷¹ Its intrinsic mechanism is related to the phenomenon of hypoxia in the solid tumor microenvironment. Hypoxia induces autophagy through the HIF-1 α signaling pathway and promotes tumor cell invasion and metastasis. However, ARlncRNA CPS1-IT1 inhibits hypoxia-induced autophagy by suppressing HIF-1 α , thereby suppressing epithelial–mesenchymal transition and CRC metastasis.⁷² More information and mechanisms of ARlncRNAs regulating tumor migration and invasion are shown in Table 3 and Figure 4.

Other regulatory functions of ARlncRNAs

Radiotherapy is the cornerstone of treatment for many types of cancer. However, radiotherapy resistance largely affects the effectiveness of radiotherapy. ARlncRNAs such as HULC and HOTAIR have been found to mediate autophagy and thus promote radiotherapy resistance. Interestingly, HULC binds to Beclin-1, thereby inhibiting Beclin-1

phosphorylation, leading to the inhibition of autophagy through the mTOR pathway to promote radiotherapy resistance. In contrast, HOTAIR promotes autophagy by regulating the miR-93/ATG12 axis in CRC, thereby enhancing radiation resistance. Since they are both up-regulated after radiotherapy, decreased tumor cell survival and cell cycle, and increased apoptosis and radiosensitivity were observed after their knockdown.^{73,74}

Tumor formation and development cannot be separated from angiogenesis and glucose utilization. The highly enriched exosomal ARlncRNA OIP5-AS1 can promote angiogenesis and autophagy in osteosarcomas via miR-153/ATG5. The intrinsic mechanism is that ATG5 regulates angiogenesis in vascular endothelial cells via reactive oxygen species-dependent signal.²⁶ Glycolysis is the main pathway of glucose utilization by cancer cells. MYC mRNA is one of the core regulators of glycolysis, which is regulated by the upstream IGF2BP2. Wang et al found that the ARlncRNA LINRIS binds to the ubiquitination site of IGF2BP2, thereby preventing the degradation of IGF2BP2 via the ubiquitination-autophagy pathway.⁷⁵ As a result, its downstream MYC mRNA was stabilized. *In vivo* experiments demonstrated that LINRIS knockdown decreased tumor development in orthotopic and patient-derived xenograft models.⁷⁵

Immune cells and immune-related mediators are essential in tumor growth in the tumor microenvironment. Cancer-associated fibroblasts are a major source of chemicals released by the tumor microenvironment that drive cancer cell growth. Similarly, variations of inflammatory mediators in cancer cells inside the tumor microenvironment result in the transformation of normal fibroblasts into cancer-associated fibroblasts. MALAT1 was found to decrease autophagic flux and increase IL-6 by regulating the PTEN/AKT/mTOR and SQSTM1/NF- κ B, thereby transforming fibroblasts into cancer-associated fibroblasts to accelerate GC development. Nevertheless, the mechanism behind IL-6-induced cancer-associated fibroblast activation needs further investigation.⁷⁶ Through pattern recognition receptors such as Toll-like receptors and advanced glycosylation end-product-specific receptors (AGERs), carcinogens can activate the innate immune system. AGERs recognize endogenous molecules released during chronic inflammation and recruit epithelial cells for innate immune cell recruitment. Investigators found that up-regulation of ARlncRNA AGER inhibited lung cancer cell proliferation and migration, induced cell cycle arrest in the G0/G1 phases, and promoted apoptosis. ARlncRNA AGER may also promote cytotoxic activity and autophagy of immune effector cells by up-regulating AGER by sponging miR-185. Thus, it is a potential target for tumor immunotherapy.²⁷

The clinical significance of ARlncRNAs

ARlncRNAs as biomarkers

Most current cancer biomarkers are proteins or peptides whose changes in tissue or blood levels reflect cancer progression. However, these biomarkers suffer from shortcomings. On the one hand, they produce a large number of

Table 3 ARlncRNAs that regulate tumor migration and invasion.

LncRNA	Levels	Cancer type	Autophagy	Mechanisms	Role	Reference
CCAT1	↑	Gastric cancer	Activates	miR-140-3p/ATG5	Oncogene	105
FIRRE	↑	Colorectal cancer	Activates	PTBP1	Oncogene	25
LCPAT1	↑	Lung cancer	Activates	RCC2	Oncogene	106
loc146880	↑	Lung cancer	Activates	ROS	Oncogene	24
SNHG1	↑	Basal bladder cancer	Activates	ATG↑	Oncogene	64
SNHG11	↑	Gastric cancer	Activates	miR-483-3p/ATG12 and miR-1276/ATG12	Oncogene	17
JPX	↑	Gastric cancer	Activates	inhibiting miR-197	Oncogene	107
ADAMTS9-AS1	↑	Bladder cancer	Suppresses	activation of PI3K/AKT/mTOR	Oncogene	66
CDKN2B-AS1	↑	Liver cancer	Suppresses	miR-199a-5p	Oncogene	65
DANCR	↑	Gastric cancer	Suppresses	miR-194/AKT2 axis	Oncogene	108
HAGLROS	↑	Gastric cancer	Suppresses	miR-100-5p/mTOR	Oncogene	49
lncRNA-45	↓	Breast cancer	Suppresses	activating the mTOR signaling pathway	Oncogene	67
ADAMTS9-AS2	↓	Liver cancer	Activates	PI3K/AKT/mTOR	Suppressor	69
GAS8-AS1	↓	Ovarian cancer	Activates	binding with Beclin1	Suppressor	23
PTCSC3	↓	Oral cancer	Activates	LC3B-I/Beclin 1	Suppressor	68
CPS1-IT1	↓	Colorectal cancer	Suppresses	inactivation of HIF-1 α	Suppressor	72

false-positive and/or false-negative results, which is partly due to the nature of the biomarker itself. On the other hand, the invasive and inconvenient nature of tissue biopsy hinders its application. Therefore, it is necessary to develop highly sensitive, non-invasive non-protein biomarkers. Due to their great stability and resistance to nuclease-mediated degradation, circulating lncRNAs appear to be more dependable than other circulating nucleic acids.^{9,28}

ARlncRNA CASC9 was substantially expressed in oral squamous cell carcinoma and significantly linked with tumor volume, local lymph node migration, stage of the disease, and overall survival. Therefore, CASC9 could be a potential marker for the diagnosis and prognosis of oral squamous cell carcinoma.⁵³ ARlncRNA EGOT is also significantly expressed in colon cancer patients. Survival time is decreased in patients with high EGOT expression. After EGOT overexpression, the apoptosis rate decreased and the cell growth invasion increased. Moreover, EGOT was found to potentially impede the proliferation and spread of colon cancer via modulating autophagy, making it a potential therapeutic target and diagnostic marker for colon cancer.⁹ In addition, due to its involvement in controlling cancer cell resistance to paclitaxel, EGOT might even function as a promising biomarker of paclitaxel response.⁴³

ARlncRNAs are used to construct prognostic models

Due to the low sensitivity or specificity of various circulating lncRNAs for specific cancer types, the diagnostic accuracy of a number of circulating lncRNAs is quite poor when analyzed individually. It has been reported that a single circulating lncRNA does not outperform the diagnostic capabilities of models consisting of several lncRNAs.²⁸ Therefore, with the development of bioinformatics, more and more prognostic models are being developed. Prediction models based on lncRNAs and clinical characteristics can effectively predict osteosarcoma.⁷⁷ Hang et al constructed an alternative marker for lung adenocarcinoma consisting of 14 ARlncRNAs and accurately

predicted survival, tumor immune microenvironment status, and even the efficacy of immune checkpoint inhibitors in patients with lung adenocarcinoma.⁷⁸ The group with the lowest risk had a higher survival benefit. High-risk scores were negatively connected with abundant peritumor immune cells and stromal cells, as well as a high mutational burden in the tumor. Low-risk patients showed greater PD-1 and CTLA-4 expression and immune checkpoint inhibitors were more effective. Li et al constructed a risk score model based on 18 ARlncRNAs to assess osteosarcoma in breast cancer patients.⁷⁹ They also found that these lncRNAs are participating in the modulation of oxidative phosphorylation, nucleotide excision repair, TGF- β signaling pathway, and multicellular biomolecule metabolism.

ARlncRNAs as promising targets for tumor therapy

Knockdown of proto-oncogene can promote apoptosis and drug sensitivity of tumor cells and inhibit proliferation and metastasis of tumor cells. In an *in vitro* study, the investigators observed that the p-AKT, p-mTOR, and BCL-2 levels were reduced, while the LC3BII/LC3BI ratio was elevated in oral squamous cells with knockdown of ARlncRNA CASC9. These results suggest that CASC9 induces autophagy and death while suppressing cell proliferation.⁵³ In another *in vitro* study, hepatocellular carcinoma cells exhibited weaker viability, invasive and migratory activities, and stronger apoptotic activity after silencing ARlncRNA CDKN2B-AS1 and up-regulating miR-199a-5p.⁶⁵ In *in vivo* studies, inhibition of tumor cell growth was observed after the knockdown of ARlncRNA MALAT-1, HULC, DANCR, and LCPAT1.^{6,50,80,81} By the same token, up-regulation of oncogenes is also an effective anti-tumor strategy. Overexpression of ARlncRNA PTCSC3 leads to a substantial reduction in human oral cancer cell proliferation through the induction of apoptotic cell death.⁶⁸ Fang et al constructed the ARlncRNA GAS8-AS1 plasmid to transfect ovarian cancer cells.²³ After transfection, GAS8-AS1 was up-regulated and inhibited the migratory ability of ovarian

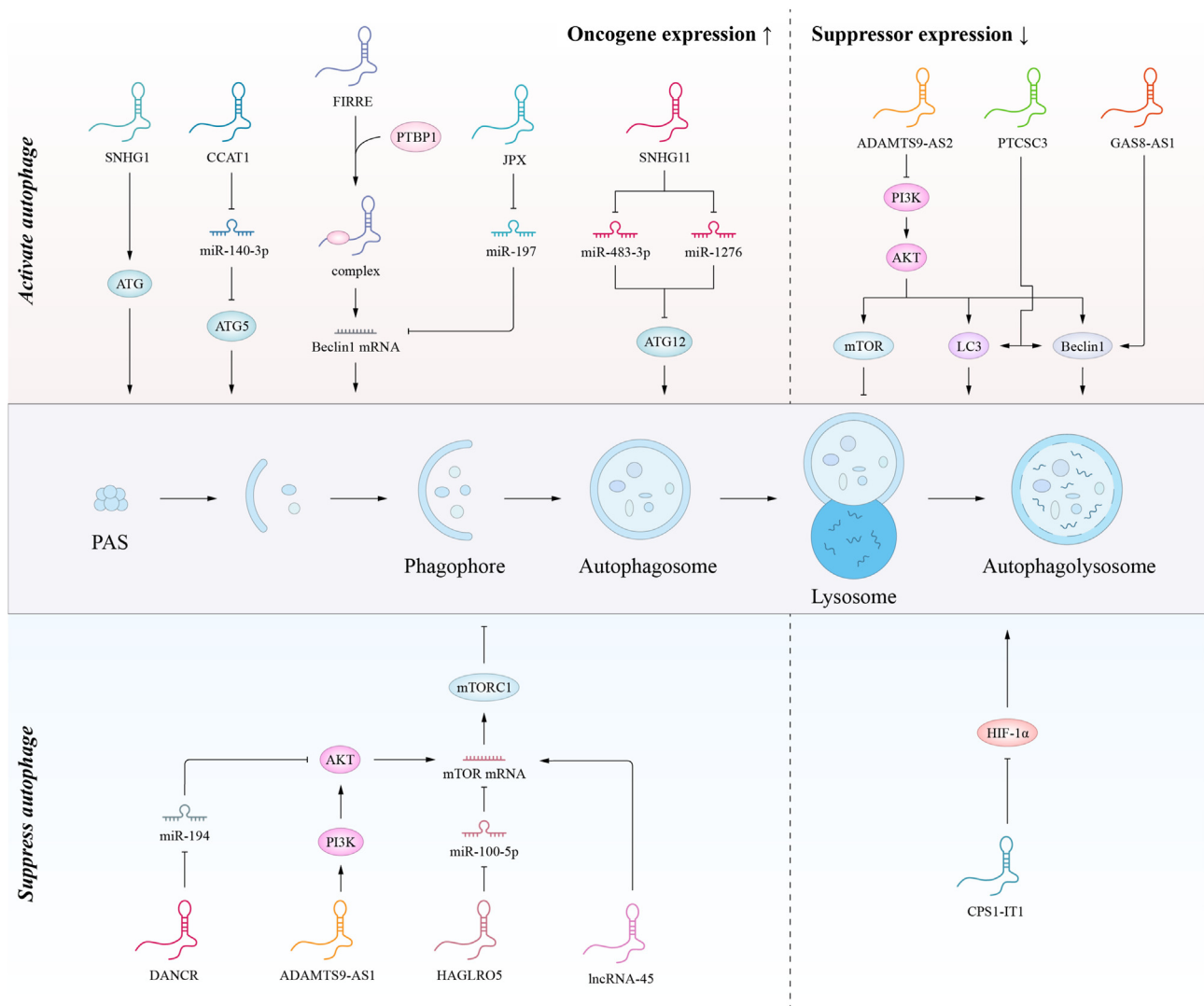


Figure 4 The “double-edged sword” mechanism of ARlncRNAs in tumor migration and invasion.

cancer cells; in addition, ovarian cancer cells were reduced by GAS8-AS1.

The connection between clinical pharmacotherapy and ARlncRNAs

Scientists have discerned several pharmacological agents that modulate ARlncRNA and thereby impede tumorigenesis via multiple signaling pathways. These recent findings regarding drug interventions furnish a foundation for novel antineoplastic therapeutic approaches.

Some antitumor drugs exert antitumor effects while up-regulating certain ARlncRNAs that activate autophagy, which reduces the efficacy of the drugs. Therefore, knocking down the ARlncRNA up-regulation induced by these drugs could exert synergistic antitumor effects. The accumulation of misfolded proteins in the endoplasmic reticulum and the activation of unfolded protein responses may lead to endoplasmic reticulum stress. Continuous or exacerbated endoplasmic reticulum stress can lead tumor cells toward apoptosis.⁸² However, endoplasmic reticulum

stress can be mitigated by autophagy to preserve its homeostasis.⁸³ Researchers found that resveratrol promotes ARlncRNA H19 expression, thereby activating autophagy. Interestingly, knocking down H19 in resveratrol-lowered cells further promoted the impacts of resveratrol on apoptosis, endoplasmic reticulum stress, and cell cycle S-phase arrest, and inhibited cell migration. Therefore, knocking down H19 reverses the resistance of tumor cells to resveratrol treatment.⁸² MALAT1 expression is up-regulated after metformin use in breast cancer, which also implies activation of autophagy. The elevated endoplasmic reticulum stress induced by metformin treatment was further up-regulated after the knockdown of MALAT1 was made. It is suggested that the combination of metformin and MALAT1 knockdown could potentially result in synergistic induction of cellular death.⁸³ Ferroptosis is a type of necrotic cell death that is dependent on iron and is characterized by the occurrence of oxidative damage to phospholipids and an up-regulated expression of unsaturated fatty acids in the cellular membrane. This ultimately results in the formation of lipid peroxidation and a subsequent disturbance in the

structure of the membrane. Researchers discovered that metformin could trigger ferroptosis by blocking H19-mediated autophagy in breast cancer cells.⁸⁴

Furthermore, it is worth noting that certain antitumor drugs have been found to possess the ability to down-regulate ARlncRNA and thereby exhibit antitumor properties. The administration of propofol was observed to promote apoptosis and decrease autophagic activity by inhibiting the MALAT1/miR-30e/ATG5 axis. *In vivo*, co-treatment with propofol and cisplatin resulted in a marked reduction in both the size and weight of tumors in a GC xenograft model.⁸⁵ Apatinib inhibits cancer stem cell properties and malignant biological behavior of breast cancer stem cells by blocking the Wnt/ β -catenin signal through down-regulation of ARlncRNA ROR.⁸⁶ In addition, cisplatin-induced autophagy in HO8910 ovarian cancer cells. Besides, ARlncRNA RP11-135L22.1 overexpression inhibited cisplatin-induced autophagy, thus enhancing the effect of cisplatin on ovarian cancer cells. The combination of cisplatin and RP11-135L22.1 could decrease autophagy, enhance apoptosis, and suppress the activity of ovarian cancer cells.⁸⁷

Conclusions and perspectives

In recent years, autophagy has become a hot topic in the field of tumor research. However, the two-sided nature of autophagy in tumors has led to the dilemma of targeting autophagy for therapy. Interestingly, ARlncRNAs can mediate both sides of autophagy, thus positively or negatively influencing tumorigenesis, progression, and drug resistance. The dualistic nature of ARlncRNA can be attributed to the heterogeneity of tumor microenvironments, as well as the varied mechanisms of action among different ARlncRNAs. Consequently, these factors must be taken into account when considering targeted therapies. Additionally, a deeper investigation into the regulatory mechanisms governing ARlncRNA-mediated molecular networks is warranted.

Numerous investigations have demonstrated that ARlncRNAs function by modulating specific miRNAs that are downstream of them. Nevertheless, a single lncRNA is not restricted to regulating solely one miRNA, and thus, more extensive exploration of the numerous targets or signaling pathways that lie downstream of ARlncRNAs is indispensable. For example, ATG5-mediated autophagy also involves NF- κ B and p53/Rb signaling pathways. Whether ARlncRNAs regulate these pathways requires further study and discussion. Similarly, tumor cell proliferation, apoptosis, migration and invasion, and drug resistance are not only affected by autophagy, but some ARlncRNAs can achieve multiple regulations of tumors through autophagy and other non-autophagic signaling pathways. On the one hand, it is necessary to clarify the regulatory mechanisms of these non-autophagy; on the other hand, it is necessary to clarify the crosstalk between these non-autophagy mechanisms and autophagy, which may rescue the failure of autophagy-targeted therapy. For example, the relationship between Wnt/ β -catenin and autophagy is complex and controversial and deserves further investigation. In addition, autophagy can promote immune escape during cancer cell

development. How the regulation of autophagy in tumor cells by ARlncRNA alters the mechanism of immune system monitoring of tumor cells also deserves further analysis.

Early diagnosis can help improve cancer cure or survival rates. The advantages of ARlncRNA, such as its high stability and relative abundance in circulation, make it a reliable diagnostic and prognostic biomarker. However, the premise is that it needs to be improved; otherwise, there is still the problem of being degraded in circulation. Employing a panel of various ARlncRNAs in model construction may substantially amplify the diagnostic and predictive capabilities. While the study of circulating lncRNA biomarkers is still in its nascent stage, more comprehensive investigations are warranted to uncover circulating biomarkers that can reliably detect cancer in its incipient stages.

In conclusion, ARlncRNAs in tumors are a double-edged sword in mediating tumor development and drug resistance. They mediate autophagy while also affecting tumor cell proliferation, apoptosis, migration, and invasion, drug resistance, angiogenesis, glycolysis, radiation therapy resistance, and immune microenvironment; therefore, they can serve as ideal therapeutic targets and biomarkers. However, further studies are needed to elucidate these molecular mechanisms and the crosstalk between them.

Author contributions

LS, AZ, and QZ contributed to the study's conception. YZ, JT, and CW wrote the manuscript. LS, AZ, and QZ critically revised the manuscript. All authors read and approved the submitted version of the manuscript.

Conflict of interests

The authors declare that there is no conflict of interests.

Funding

The work is supported by the Xinglin project of Chengdu University of Traditional Chinese Medicine, Sichuan, China (No. ZKYY2019, MPRC2021012).

References

1. Sung H, Ferlay J, Siegel RL, et al. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA A Cancer J Clin*. 2021;71(3):209–249.
2. Du J, Wei J, Yang Y, et al. Disappearance of bone metastases in chemotherapy-resistant gastric cancer treated with antigen peptide-pulsed dendritic cell-activated cytotoxic T lymphocyte immunotherapy: a case report. *Oncol Lett*. 2018;16(1):875–881.
3. Katsuda M, Miyazawa M, Ojima T, et al. A double-blind randomized comparative clinical trial to evaluate the safety and efficacy of dendritic cell vaccine loaded with WT1 peptides (TLP0-001) in combination with S-1 in patients with advanced pancreatic cancer refractory to standard chemotherapy. *Trials*. 2019;20(1):242.

4. Matsuda T, Takeuchi H, Sakurai T, et al. Pilot study of WT1 peptide-pulsed dendritic cell vaccination with docetaxel in esophageal cancer. *Oncol Lett.* 2018;16(1):1348–1356.
5. Shao G, Zhao Z, Zhao W, et al. Long non-coding RNA MALAT1 activates autophagy and promotes cell proliferation by downregulating microRNA-204 expression in gastric cancer. *Oncol Lett.* 2020;19(1):805–812.
6. Yu X, Ye X, Lin H, et al. Knockdown of long non-coding RNA LCPAT1 inhibits autophagy in lung cancer. *Cancer Biol Med.* 2018;15(3):228–237.
7. Wang Z, Jin J. LncRNA SLCO4A1-AS1 promotes colorectal cancer cell proliferation by enhancing autophagy via miR-508-3p/PARD3 axis. *Aging.* 2019;11(14):4876–4889.
8. Zhang XH, Li BF, Ding J, et al. LncRNA DANCER-miR-758-3p-PAX6 molecular network regulates apoptosis and autophagy of breast cancer cells. *Cancer Manag Res.* 2020;12:4073–4084.
9. Liu Y, Zhang B, Cao WB, Wang HY, Niu L, Zhang GZ. Study on clinical significance of LncRNA EGOT expression in colon cancer and its effect on autophagy of colon cancer cells. *Cancer Manag Res.* 2020;12:13501–13512.
10. Zhang Y, Liu C, Wu C, Song L. Natural peptides for immunological regulation in cancer therapy: mechanism, facts and perspectives. *Biomed Pharmacother.* 2023;159:114257.
11. Si Y, Yang Z, Ge Q, et al. Long non-coding RNA Malat1 activated autophagy, hence promoting cell proliferation and inhibiting apoptosis by sponging miR-101 in colorectal cancer. *Cell Mol Biol Lett.* 2019;24:50.
12. Li N, Zeng A, Wang Q, Chen M, Zhu S, Song L. Regulatory function of DNA methylation mediated lncRNAs in gastric cancer. *Cancer Cell Int.* 2022;22(1):227.
13. Zhou C, Yi C, Yi Y, et al. LncRNA PVT1 promotes gemcitabine resistance of pancreatic cancer via activating Wnt/ β -catenin and autophagy pathway through modulating the miR-619-5p/Pygo2 and miR-619-5p/ATG14 axes. *Mol Cancer.* 2020;19(1):118.
14. Wang X, Lan Z, He J, et al. LncRNA SNHG6 promotes chemoresistance through ULK1-induced autophagy by sponging miR-26a-5p in colorectal cancer cells. *Cancer Cell Int.* 2019;19:234.
15. Chen Z, Wu H, Zhang M. Long non-coding RNA: an underlying bridge linking neuroinflammation and central nervous system diseases. *Neurochem Int.* 2021;148:105101.
16. Liang Y, Lu Y, Chen Q, et al. Identification of long noncoding RNAs that exert transcriptional regulation by forming RNA-DNA triplexes in prostate cancer. *Int J Mol Sci.* 2023;24(3):2035.
17. Wu Q, Ma J, Wei J, Meng W, Wang Y, Shi M. LncRNA SNHG11 promotes gastric cancer progression by activating the Wnt/ β -catenin pathway and oncogenic autophagy. *Mol Ther.* 2021;29(3):1258–1278.
18. Yorimitsu T, Klionsky DJ. Autophagy: molecular machinery for self-eating. *Cell Death Differ.* 2005;12(Suppl 2):1542–1552.
19. Galluzzi L, Baehrecke EH, Ballabio A, et al. Molecular definitions of autophagy and related processes. *EMBO J.* 2017;36(13):1811–1836.
20. Cai Q, Wang S, Jin L, et al. Long non-coding RNA GBCDRlnc1 induces chemoresistance of gallbladder cancer cells by activating autophagy. *Mol Cancer.* 2019;18(1):82.
21. Zhu K, Yuan Y, Wen J, et al. LncRNA Sox2OT-V7 promotes doxorubicin-induced autophagy and chemoresistance in osteosarcoma via tumor-suppressive miR-142/miR-22. *Aging (Albany NY).* 2020;12(8):6644–6666.
22. Chavez-Dominguez R, Perez-Medina M, Lopez-Gonzalez JS, Galicia-Velasco M, Aguilar-Cazares D. The double-edge sword of autophagy in cancer: from tumor suppression to pro-tumor activity. *Front Oncol.* 2020;10:578418.
23. Fang YJ, Jiang P, Zhai H, Dong JS. LncRNA GAS8-AS1 inhibits ovarian cancer progression through activating Beclin1-mediated autophagy. *OncoTargets Ther.* 2020;13:10431–10440.
24. Deng X, Feng N, Zheng M, et al. PM_{2.5} exposure-induced autophagy is mediated by lncRNA loc146880 which also promotes the migration and invasion of lung cancer cells. *Biochim Biophys Acta Gen Subj.* 2017;1861(2):112–125.
25. Wang Y, Li Z, Xu S, et al. LncRNA FIRRE functions as a tumor promoter by interaction with PTBP1 to stabilize BECN1 mRNA and facilitate autophagy. *Cell Death Dis.* 2022;13(2):98.
26. Li Y, Lin S, Xie X, Zhu H, Fan T, Wang S. Highly enriched exosomal lncRNA OIP5-AS1 regulates osteosarcoma tumor angiogenesis and autophagy through miR-153 and ATG5. *Am J Transl Res.* 2021;13(5):4211–4223.
27. Pan Z, Liu L, Nie W, et al. Long non-coding RNA AGER-1 functionally upregulates the innate immunity gene AGER and approximates its anti-tumor effect in lung cancer. *Mol Carcinog.* 2018;57(3):305–318.
28. Badowski C, He B, Garmire LX. Blood-derived lncRNAs as biomarkers for cancer diagnosis: the good, the bad and the beauty. *NPJ Precis Oncol.* 2022;6(1):40.
29. Hu Y, Yu Y, You S, et al. Long noncoding RNA MALAT1 regulates autophagy associated chemoresistance via miR-23b-3p sequestration in gastric cancer. *Mol Cancer.* 2017;16(1):174.
30. Xi Z, Si J, Nan J. LncRNA MALAT1 potentiates autophagy-associated cisplatin resistance by regulating the microRNA-30b/autophagy-related gene 5 axis in gastric cancer. *Int J Oncol.* 2019;54(1):239–248.
31. Gui Z, Zhao Z, Sun Q, et al. LncRNA FEZF1-AS1 promotes multi-drug resistance of gastric cancer cells via upregulating ATG5. *Front Cell Dev Biol.* 2021;9:749129.
32. Huang FX, Chen HJ, Zheng FX, et al. LncRNA BLACAT1 is involved in chemoresistance of non-small cell lung cancer cells by regulating autophagy. *Int J Oncol.* 2019;54(1):339–347.
33. Luo Y, Zheng S, Wu Q, et al. Long noncoding RNA (lncRNA) EIF3J-DT induces chemoresistance of gastric cancer via autophagy activation. *Autophagy.* 2021;17(12):4083–4101.
34. Li Y, Li C, Li D, Yang L, Jin J, Zhang B. LncRNA KCNQ10T1 enhances the chemoresistance of oxaliplatin in colon cancer by targeting the miR-34a/ATG4B pathway. *OncoTargets Ther.* 2019;12:2649–2660.
35. Shen Q, Xu Z, Xu S. Long non-coding RNA LUCAT1 contributes to cisplatin resistance by regulating the miR-514a-3p/ULK1 axis in human non-small cell lung cancer. *Int J Oncol.* 2020;57(4):967–979.
36. Li D, Li C, Chen Y, et al. LncRNA HOTAIR induces sunitinib resistance in renal cancer by acting as a competing endogenous RNA to regulate autophagy of renal cells. *Cancer Cell Int.* 2020;20:338.
37. Li PP, Li RG, Huang YQ, Lu JP, Zhang WJ, Wang ZY. LncRNA OTUD6B-AS1 promotes paclitaxel resistance in triple negative breast cancer by regulation of miR-26a-5p/MTDH pathway-mediated autophagy and genomic instability. *Aging (Albany NY).* 2021;13(21):24171–24191.
38. Chen Y, Cui Z, Wu Q, Wang H, Xia H, Sun Y. Long non-coding RNA HOXA11-AS knockout inhibits proliferation and overcomes drug resistance in ovarian cancer. *Bioengineered.* 2022;13(5):13893–13905.
39. Chen YM, Liu Y, Wei HY, Lv KZ, Fu P. Linc-ROR induces epithelial-mesenchymal transition and contributes to drug resistance and invasion of breast cancer cells. *Tumour Biol.* 2016;37(8):10861–10870.
40. Li Y, Jiang B, Zhu H, et al. Inhibition of long non-coding RNA ROR reverses resistance to Tamoxifen by inducing autophagy in breast cancer. *Tumour Biol.* 2017;39(6):1010428317705790.

41. Xia X, Li Z, Li Y, Ye F, Zhou X. LncRNA XIST promotes carboplatin resistance of ovarian cancer through activating autophagy via targeting miR-506-3p/FOXp1 axis. *J Gynecol Oncol*. 2022;33(6):e81.
42. Shi H, Pu J, Zhou XL, Ning YY, Bai C. Silencing long non-coding RNA ROR improves sensitivity of non-small-cell lung cancer to cisplatin resistance by inhibiting PI3K/Akt/mTOR signaling pathway. *Tumour Biol*. 2017;39(5):1010428317697568.
43. Xu S, Wang P, Zhang J, et al. Ai-lncRNA EGOT enhancing autophagy sensitizes paclitaxel cytotoxicity via upregulation of ITPR1 expression by RNA-RNA and RNA-protein interactions in human cancer. *Mol Cancer*. 2019;18(1):89.
44. Li G, Qian L, Tang X, Chen Y, Zhao Z, Zhang C. Long non-coding RNA growth arrest-specific 5 (GAS5) acts as a tumor suppressor by promoting autophagy in breast cancer. *Mol Med Rep*. 2020;22(3):2460–2468.
45. Liu X, Zhang X, Du S. Long non-coding RNA ACTA2-AS1 inhibits the cisplatin resistance of non-small cell lung cancer cells through inhibiting autophagy by suppressing TSC2. *Cell Cycle*. 2022;21(4):368–378.
46. Zhang F, Chen Q, Chen P, Liu C, Wang H, Zhao L. The lncRNA CRNDE is regulated by E2F6 and sensitizes gastric cancer cells to chemotherapy by inhibiting autophagy. *J Cancer*. 2022;13(10):3061–3072.
47. Zhang F, Wang H, Yu J, et al. LncRNA CRNDE attenuates chemoresistance in gastric cancer via SRSF6-regulated alternative splicing of PICALM. *Mol Cancer*. 2021;20(1):6.
48. Xia H, Qu XL, Liu LY, Qian DH, Jing HY. LncRNA MEG3 promotes the sensitivity of vincristine by inhibiting autophagy in lung cancer chemotherapy. *Eur Rev Med Pharmacol Sci*. 2018;22(4):1020–1027.
49. Chen JF, Wu P, Xia R, et al. STAT3-induced lncRNA HAGLROS overexpression contributes to the malignant progression of gastric cancer cells via mTOR signal-mediated inhibition of autophagy. *Mol Cancer*. 2018;17(1):6.
50. Gao D, Lv AE, Li HP, Han DH, Zhang YP. LncRNA MALAT-1 elevates HMGB1 to promote autophagy resulting in inhibition of tumor cell apoptosis in multiple myeloma. *J Cell Biochem*. 2017;118(10):3341–3348.
51. Xie Y, Zheng ZW, He HT, Chang ZB. LncRNA NEAT1 induces autophagy through the miR-128-3p/ADAM28 axis to suppress apoptosis of non-small-cell lung cancer. *Kaohsiung J Med Sci*. 2022;38(10):933–949.
52. Chen J, Wang F, Xu H, et al. Long non-coding RNA SNHG1 regulates the Wnt/ β -catenin and PI3K/AKT/mTOR signaling pathways via EZH2 to affect the proliferation, apoptosis, and autophagy of prostate cancer cell. *Front Oncol*. 2020;10:552907.
53. Yang Y, Chen D, Liu H, Yang K. Increased expression of lncRNA CASC9 promotes tumor progression by suppressing autophagy-mediated cell apoptosis via the AKT/mTOR pathway in oral squamous cell carcinoma. *Cell Death Dis*. 2019;10:41.
54. Li F, Zhou X, Chen M, Fan W. Regulatory effect of lncRNA DRAIC/miR-149-5p/NFIB molecular network on autophagy of esophageal cancer cells and its biological behavior. *Exp Mol Pathol*. 2020;116:104491.
55. Gao Z, Zheng G, Gong X, et al. LncRNA MAFG-AS1 deregulated in breast cancer affects autophagy and progression of breast cancer by interacting with miR-3612 and FKBP₄ invitro. *Biochem Biophys Res Commun*. 2022;616:95–103.
56. Qin Y, Sun W, Zhang H, et al. LncRNA GAS8-AS1 inhibits cell proliferation through ATG5-mediated autophagy in papillary thyroid cancer. *Endocrine*. 2018;59(3):555–564.
57. Qin Y, Sun W, Wang Z, et al. ATF2-induced lncRNA GAS8-AS1 promotes autophagy of thyroid cancer cells by targeting the miR-187-3p/ATG5 and miR-1343-3p/ATG7 axes. *Mol Ther Nucleic Acids*. 2020;22:584–600.
58. Zhang P, Pan Y, Sun J, Pan G. Aberrant expression of lncRNA CASC2 mediated the cell viability, apoptosis and autophagy of colon cancer cells by sponging miR-19a via NF- κ B signaling pathway. *Int J Exp Pathol*. 2021;102(3):163–171.
59. Li Q, Chen K, Dong R, Lu H. LncRNA CASC2 inhibits autophagy and promotes apoptosis in non-small cell lung cancer cells via regulating the miR-214/TRIM16 axis. *RSC Adv*. 2018;8(71):40846–40855.
60. Ju B, Liu Z, Nai C, Zhu X. Long non-coding RNA CASC2 induces apoptosis and autophagy in human colon cancer cells via modulation of TRIM16 expression. *Am J Transl Res*. 2020;12(6):2695–2702.
61. Wang C, Tan C, Wen Y, et al. FOXp1-induced lncRNA CLRN1-AS1 acts as a tumor suppressor in pituitary prolactinoma by repressing the autophagy via inactivating Wnt/ β -catenin signaling pathway. *Cell Death Dis*. 2019;10(7):499.
62. Fan X, Huang H, Ji Z, Mao Q. Long non-coding RNA MEG3 functions as a competing endogenous RNA of miR-93 to regulate bladder cancer progression via PI3K/AKT/mTOR pathway. *Transl Cancer Res*. 2020;9(3):1678–1688.
63. Wen D, Liu WL, Lu ZW, Cao YM, Ji QH, Wei WJ. SNHG9, a papillary thyroid cancer cell exosome-enriched lncRNA, inhibits cell autophagy and promotes cell apoptosis of normal thyroid epithelial cell nthy-ori-3 through YBOX3/P21 pathway. *Front Oncol*. 2021;11:647034.
64. Xu J, Yang R, Hua X, et al. lncRNA SNHG1 promotes basal bladder cancer invasion via interaction with PP2A catalytic subunit and induction of autophagy. *Mol Ther Nucleic Acids*. 2020;21:354–366.
65. Xu L, Wu H, Pan J, Chen Z, Du L. Significance of lncRNA CDKN2B-AS1 in interventional therapy of liver cancer and the mechanism under its participation in tumour cell growth via miR-199a-5p. *JAMA Oncol*. 2022;2022:2313416.
66. Yang G, Li Z, Dong L, Zhou F. lncRNA ADAMTS9-AS1 promotes bladder cancer cell invasion, migration, and inhibits apoptosis and autophagy through PI3K/AKT/mTOR signaling pathway. *Int J Biochem Cell Biol*. 2021;140:106069.
67. Qiu J, Guo Y, Wang S, et al. Newly identified lncRNA-45 promotes breast cancer metastasis through activating the mTOR signaling pathway. *Biochem Biophys Res Commun*. 2023;640:40–49.
68. Zhang H, Wang J, Xun W, Wang J, Song W, Wang X. Long non-coding RNA PTCSC3 inhibits human oral cancer cell proliferation by inducing apoptosis and autophagy. *Arch Med Sci*. 2021;17(2):492–499.
69. Li H, Huang H, Li S, Mei H, Cao T, Lu Q. Long non-coding RNA ADAMTS9-AS2 inhibits liver cancer cell proliferation, migration and invasion. *Exp Ther Med*. 2021;21(6):559.
70. Wang TH, Yu CC, Lin YS, et al. Long noncoding RNA CPS₁-IT1 suppresses the metastasis of hepatocellular carcinoma by regulating HIF-1 α activity and inhibiting epithelial-mesenchymal transition. *Oncotarget*. 2016;7(28):43588–43603.
71. Zhang W, Yuan W, Song J, Wang S, Gu X. LncRNA CPS₁-IT1 suppresses cell proliferation, invasion and metastasis in colorectal cancer. *Cell Physiol Biochem*. 2017;44(2):567–580.
72. Zhang W, Yuan W, Song J, Wang S, Gu X. LncRNA CPS₁-IT1 suppresses EMT and metastasis of colorectal cancer by inhibiting hypoxia-induced autophagy through inactivation of HIF-1 α . *Biochimie*. 2018;144:21–27.
73. Chen C, Wang K, Wang Q, Wang X. LncRNA HULC mediates radioresistance via autophagy in prostate cancer cells. *Braz J Med Biol Res*. 2018;51(6):e7080.
74. Liu Y, Chen X, Chen X, et al. Long non-coding RNA HOTAIR knockdown enhances radiosensitivity through regulating microRNA-93/ATG12 axis in colorectal cancer. *Cell Death Dis*. 2020;11(3):175.

75. Wang Y, Lu JH, Wu QN, et al. lncRNA LINRIS stabilizes IGF2BP2 and promotes the aerobic glycolysis in colorectal cancer. *Mol Cancer*. 2019;18(1):174.
76. Wang Z, Wang X, Zhang T, et al. lncRNA MALAT1 promotes gastric cancer progression via inhibiting autophagic flux and inducing fibroblast activation. *Cell Death Dis*. 2021;12(4):368.
77. Duan L, Xia Y, Li C, Lan N, Hou X. Identification of autophagy-related lncRNA to predict the prognosis of colorectal cancer. *Front Genet*. 2022;13:906900.
78. Chen H, Hu Z, Sang M, et al. Identification of an autophagy-related lncRNA prognostic signature and related tumor immunity research in lung adenocarcinoma. *Front Genet*. 2021;12:767694.
79. Li X, Chen J, Yu Q, et al. A signature of autophagy-related long non-coding RNA to predict the prognosis of breast cancer. *Front Genet*. 2021;12:569318.
80. Xin L, Zhou Q, Yuan YW, et al. METase/lncRNA HULC/FoxM1 reduced cisplatin resistance in gastric cancer by suppressing autophagy. *J Cancer Res Clin Oncol*. 2019;145(10):2507–2517.
81. Li C, Zhao W, Pan X, et al. lncRNA KTN1-AS1 promotes the progression of non-small cell lung cancer via sponging of miR-130a-5p and activation of PDPK1. *Oncogene*. 2020;39(39):6157–6171.
82. Li T, Zhang X, Cheng L, et al. Modulation of lncRNA H19 enhances resveratrol-inhibited cancer cell proliferation and migration by regulating endoplasmic reticulum stress. *J Cell Mol Med*. 2022;26(8):2205–2217.
83. Huang Y, Zhou Z, Zhang J, et al. lncRNA MALAT1 participates in metformin inhibiting the proliferation of breast cancer cell. *J Cell Mol Med*. 2021;25(15):7135–7145.
84. Chen J, Qin C, Zhou Y, Chen Y, Mao M, Yang J. Metformin may induce ferroptosis by inhibiting autophagy via lncRNA H19 in breast cancer. *FEBS Open Bio*. 2022;12(1):146–153.
85. Zhang YF, Li CS, Zhou Y, Lu XH. Propofol facilitates cisplatin sensitivity via lncRNA MALAT1/miR-30e/ATG5 axis through suppressing autophagy in gastric cancer. *Life Sci*. 2020;244:117280.
86. Jiang B, Zhu H, Tang L, et al. Apatinib inhibits stem properties and malignant biological behaviors of breast cancer stem cells by blocking Wnt/ β -catenin signal pathway through down-regulating lncRNA ROR. *Anti Cancer Agents Med Chem*. 2022;22(9):1723–1734.
87. Zou SH, Du X, Sun FD, Wang PC, Li M. Cisplatin suppresses tumor proliferation by inhibiting autophagy in ovarian cancer via long non-coding RNA RP11-135L22.1. *Eur Rev Med Pharmacol Sci*. 2018;22(4):928–935.
88. Guo S, Zhang L, Zhang Y, et al. Long non-coding RNA TUG1 enhances chemosensitivity in non-small cell lung cancer by impairing microRNA-221-dependent PTEN inhibition. *Aging (Albany NY)*. 2019;11(18):7553–7569.
89. Wang M, Han D, Yuan Z, et al. Long non-coding RNA H19 confers 5-Fu resistance in colorectal cancer by promoting SIRT1-mediated autophagy. *Cell Death Dis*. 2018;9(12):1149.
90. Liu F, Ai FY, Zhang DC, Tian L, Yang ZY, Liu SJ. lncRNA NEAT1 knockdown attenuates autophagy to elevate 5-FU sensitivity in colorectal cancer via targeting miR-34a. *Cancer Med*. 2020;9(3):1079–1091.
91. Xian Z, Hu B, Wang T, et al. lncRNA UCA1 contributes to 5-fluorouracil resistance of colorectal cancer cells through miR-23b-3p/ZNF281 axis. *OncoTargets Ther*. 2020;13:7571–7583.
92. Xia C, Li Q, Cheng X, Wu T, Gao P, Gu Y. Insulin-like growth factor 2 mRNA-binding protein 2-stabilized long non-coding RNA Taurine up-regulated gene 1 (TUG1) promotes cisplatin-resistance of colorectal cancer via modulating autophagy. *Bioengineered*. 2022;13(2):2450–2469.
93. Yang Y, Jiang C, Yang Y, et al. Silencing of lncRNA-HOTAIR decreases drug resistance of non-small cell lung cancer cells by inactivating autophagy via suppressing the phosphorylation of ULK1. *Biochem Biophys Res Commun*. 2018;497(4):1003–1010.
94. Xie J, Chen X, Wang W, Guan Z, Hou J, Lin J. Long non-coding RNA PCDRlnc1 confers docetaxel resistance in prostate cancer by promoting autophagy. *J Cancer*. 2022;13(7):2138–2149.
95. Hu J, Dong SW, Pei Y, Wang J, Zhang J, Wei XP. lncRNA MITA1 promotes gefitinib resistance by inducing autophagy in lung cancer cells. *Biochem Biophys Res Commun*. 2021;551:21–26.
96. Wang L, Bi R, Li L, Zhou K, Yin H. lncRNA ANRIL aggravates the chemoresistance of pancreatic cancer cells to gemcitabine by targeting inhibition of miR-181a and targeting HMGB1-induced autophagy. *Aging*. 2021;13(15):19272–19281.
97. Zhang J, Chen K, Tang Y, et al. lncRNA-HOTAIR activates autophagy and promotes the imatinib resistance of gastrointestinal stromal tumor cells through a mechanism involving the miR-130a/ATG2B pathway. *Cell Death Dis*. 2021;12(4):367.
98. He C, Fu Y, Chen Y, Li X. Long non-coding RNA SNHG8 promotes autophagy as a ceRNA to upregulate ATG7 by sponging microRNA-588 in colorectal cancer. *Oncol Lett*. 2021;22(2):577.
99. Liu C, Ji L, Song X. Long non coding RNA UCA1 contributes to the autophagy and survival of colorectal cancer cells via sponging miR-185-5p to up-regulate the WISP2/ β -catenin pathway. *RSC Adv*. 2019;9(25):14160–14166.
100. Wang LJ, Sun GZ, Chen YF. lncRNA MSTO2P promotes proliferation and autophagy of lung cancer cells by up-regulating EZH2 expression. *Eur Rev Med Pharmacol Sci*. 2019;23(8):3375–3382.
101. Wu J, Meng X, Gao R, et al. Long non-coding RNA LINC00858 inhibits colon cancer cell apoptosis, autophagy, and senescence by activating WNK2 promoter methylation. *Exp Cell Res*. 2020;396(1):112214.
102. Liu C, Wang JO, Zhou WY, et al. Long non-coding RNA LINC01207 silencing suppresses AGR2 expression to facilitate autophagy and apoptosis of pancreatic cancer cells by sponging miR-143-5p. *Mol Cell Endocrinol*. 2019;493:110424.
103. Fan L, Li H, Wang W. Long non-coding RNA PRRT3-AS1 silencing inhibits prostate cancer cell proliferation and promotes apoptosis and autophagy. *Exp Physiol*. 2020;105(5):793–808.
104. Ma X, Ren H, Zhang Y, Wang B, Ma H. lncRNA RHPN1-AS1 inhibition induces autophagy and apoptosis in prostate cancer cells via the miR-7-5p/EGFR/PI3K/AKT/mTOR signaling pathway. *Environ Toxicol*. 2022;37(12):3013–3027.
105. Yang F, Peng ZX, Ji WD, et al. lncRNA CCAT1 upregulates ATG5 to enhance autophagy and promote gastric cancer development by absorbing miR-140-3p. *Dig Dis Sci*. 2022;67(8):3725–3741.
106. Lin H, Zhang X, Feng N, et al. lncRNA LCPAT1 mediates smoking/particulate matter 2.5-induced cell autophagy and epithelial-mesenchymal transition in lung cancer cells via RCC2. *Cell Physiol Biochem*. 2018;47(3):1244–1258.
107. Han X, Liu Z. Long non-coding RNA JPX promotes gastric cancer progression by regulating CXCR6 and autophagy via inhibiting miR-197. *Mol Med Rep*. 2021;23(1):60.
108. Cheng Z, Liu G, Huang C, Zhao X. KLF5 activates lncRNA DANCER and inhibits cancer cell autophagy accelerating gastric cancer progression. *NPJ Genom Med*. 2021;6(1):75.