NDT Plus (2009) 2: 104–110 doi: 10.1093/ndtplus/sfn203 Advance Access publication 15 January 2009

In-Depth Clinical Review



Peritoneal function in clinical practice: the importance of follow-up and its measurement in patients. Recommendations for patient information and measurement of peritoneal function

Annemieke M. Coester¹, Watske Smit^{1,2}, Dirk G. Struijk^{1,2} and Raymond T. Krediet¹

¹Division of Nephrology, Department of Medicine, Academic Medical Centre University of Amsterdam and ²Dianet Foundation, Amsterdam, The Netherlands

Abstract

A review is given on peritoneal function, especially ultrafiltration and ultrafiltration failure followed by recommendations on how to translate pathophysiology into clinical practice. The subsequent consequences for management of peritoneal membrane function and for patient information are also included.

Keywords: long-term PD; peritoneal dialysis; peritoneal function; recommendations; ultrafiltration failure

Introduction

The survival of peritoneal dialysis (PD) patients, especially non-diabetics, is superior to that of haemodialysis (HD) patients during the first years of PD [1,2]. This is probably due to a better preservation of residual renal function [3,4]. The peritoneum is a biological membrane, in which alterations can develop in the long term in some patients. These may influence the initial survival advantage. The major morphological alterations include loss of mesothelial cells, neoangiogenesis and vasculopathy, and also submesothelial and interstitial fibrosis [5–8]. Ultrafiltration failure is the most important functional abnormality. This may lead to hypervolaemia, an important risk factor for cardiovascular death.

The objectives of the present review are to present a compact survey of our current knowledge on peritoneal function, and to give recommendations on how to translate pathophysiology into clinical practice. The following subjects will be discussed: physiology of peritoneal fluid transport, mechanisms of ultrafiltration failure, ultrafiltration failure during peritonitis and in long-term PD, definition and detection of ultrafiltration failure, the measurement

Correspondence and offprint requests to: Raymond T. Krediet, Professor of Nephrology, Room F4-215, Academic Medical Center, Division of Nephrology, Department of Medicine, PO Box 22700, 1100 DE Amsterdam, The Netherlands. Tel: +31-20-566 5990; Fax: +31-20-6914904; E-mail: C.N.deboer@amc.uva.nl

of peritoneal transport and reasons for elective discontinuation of PD. Also recommendations for patient information and management of peritoneal membrane function will be given.

Physiology of peritoneal fluid transport

Fluid transport during PD is determined by hydrostatic and osmotic pressure gradients, and also by uptake from the peritoneal cavity into the lymphatic system. Ultrafiltration in peritoneal capillaries (transcapillary ultrafiltration rate) is dependent on the water permeability (hydraulic permeability) of the peritoneum, the surface area for ultrafiltration and on the hydrostatic, colloid osmotic and crystalloid osmotic pressure gradients. Blood pressure in peritoneal capillaries averages 17 mmHg, but may be variable. The intraperitoneal pressure during CAPD averages 8 mmHg in the supine position [9], but can increase to 20 mmHg during walking [10]. The intraperitoneal pressure is also influenced by the dialysate volume [11]. The colloid osmotic pressure in peritoneal capillaries averages 21 mmHg [12]. The protein concentration in the dialysate is so low that its influence on the pressure gradient can be neglected.

The crystalloid osmotic pressure gradient is especially determined by the glucose concentration in the dialysis solution. The efficacy of glucose as osmotic agent is dependent on the resistance of the peritoneal membrane to its transport. This resistance is expressed as the reflection coefficient (sigma). Sigma can vary between 1 (no passage, ideal semipermeable membrane) and 0 (free passage, so no osmotic effect of glucose). One mosmol/kg H₂O induces an osmotic pressure of 19.3 mmHg when the reflection coefficient equals 1. The peritoneal reflection coefficient of glucose averages 0.03 [13]. The various pressure gradients are summarized in Table 1.

The peritoneum is a heterogeneous dialysis membrane consisting of different structures, that is the mesothelium, interstitial tissue and the endothelial cells of the microvascular wall. The latter is the main barrier for peritoneal fluid transport. Small interendothelial pores are most important

© The Author [2009]

The online version of this article has been published under an open access model. Users are entitled to use, reproduce, disseminate, or display the open access version of this article for non-commercial purposes provided that: the original authorship is properly and fully attributed; the Journal and Oxford University Press are attributed as the original place of publication with the correct citation details given; if an article is subsequently reproduced or disseminated not in its entirety but only in part or as a derivative work this must be clearly indicated. For commercial re-use, please contact journals.permissions@oxfordjournals.org

Table 1. Pressure gradients across the peritoneal membrane during dialysis

	Pressure in peritoneal capillaries	Pressure in the filled peritoneal cavity	Pressure gradient
Hydrostatic pressure (mmHg)	17	8 (recumbent)	9
Colloid osmotic pressure (mmHg)	21	0.1	-21
Osmolarity (mosmol/kg H ₂ O)	305	347 (1.36% glucose) 486 (3.86% glucose)	
Max. crystalloid osmotic pressure gradient (mmHg)		1.36% glucose	$(347 - 305) \times 0.03 \times 19.3 = 24$
1 0 0		3.86% glucose	$(486 - 305) \times 0.03 \times 19.3 = 105$
Max. net pressure gradient (mmHg)		C	12 ^a (1.36% glucose) 93 ^a (3.86% glucose)

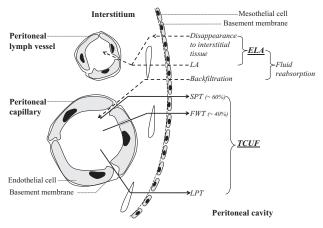
The reflection coefficient for glucose is 0.03.

for the transport of fluid and solutes according to the generally accepted three-pore theory. The number of large interendothelial pores is so small that their contribution to fluid transport can be neglected. Free water transport, that is water transport without solute transport, occurs through endothelial water channels, of which aquaporin-1 is the most important [15]. Free water transport explains the so-called sodium sieving, that is the decrease of the dialysate sodium concentration in the initial phase of dialysis with a strong hypertonic dialysis solution [16,17]. This phenomenon is absent in aquaporin-1 knock-out mice [18].

The peritoneal reflection coefficient of 0.03 consists of two components: one for the small pores (low value) and one for the water channels (1.0). This explains the capability of glucose as osmotic agent despite its small size. The overall efficacy of glucose for osmotic fluid transport can be expressed as the osmotic conductance. This is the product of the peritoneal ultrafiltration coefficient (hydraulic permeability × surface area) and the reflection coefficient sigma. The contribution of free water transport to total ultrafiltration during the first hour of a 3.86% glucose exchange averages 35-40%, but can vary between 15% and 80% [19,20]. Free water transport decreases to 20% after 4 h due to absorption of glucose [2]. The addition of a macromolecular marker to the dialysis solution makes it possible to estimate lymphatic absorption from the peritoneal cavity and peritoneal tissues by its disappearance rate. The clearance of the marker is indicated as the effective lymphatic absorption rate (ELAR). Although not universally accepted [21], the concept of the ELAR is useful for assessment of causes of ultrafiltration failure. The mean value is 1.5 mL/min [22] and is not dependent on the duration of the dialysis [23]. The ELAR is influenced by the intraperitoneal pressure [9]. A review of the use of the ELAR is given in [24]. The various pathways for fluid transport are illustrated in Figure 1 [25].

Mechanisms of ultrafiltration failure

Ultrafiltration failure may be present at the start of PD. In that situation, it is always associated with fast transport of low molecular weight solutes or with a high ELAR. The



 $NUF = \Delta IPV = TCUF - ELA$

Fig. 1. Transcapillary ultrafiltration (TCUF) is induced by the crystal-loid osmotic pressure gradient across the peritoneal membrane. It comprises water transport through small interendothelial pores (SPT) and ultrasmall transendothelial pores, the so-called free water transport (FWT). The amount of transported water across the large pores (LPT) is considered negligible. Changes in intraperitoneal volume (Δ IPV) result from TCUF and fluid reabsorption. Fluid reabsorption includes lymphatic absorption, disappearance to the interstitial tissues (together effective lymphatic absorption, ELA) and backfiltration into the capillaries. Adopted from reference [25] with permission from Oxford University Press.

fast transport of small solutes leads to a higher glucose absorption and thereby to a rapid disappearance of the osmotic gradient. This indicates an enlargement of the effective vascular peritoneal surface area, for instance because more vessels are perfused. A fast transport status is present in \sim 15% of new patients [26–28], and is probably caused by local release of vasoactive substances by macrophages or mesothelial cells. Cultured mesothelial cells synthesize various chemokines, prostaglandins and growth factors [29]. Some are produced constitutively, like vascular endothelial growth factor (VEGF) [30] and cancer antigen 125 (CA 125) [31]. Others, like interleukin-6 (IL-6) and IL-8, are produced after stimulation with chemokines [32,34]. All these substances can be detected in peritoneal effluent of PD patients. Effluent CA 125 can be considered as a marker for mesothelial cell mass [35]. It follows from these data that the number of mesothelial cells may be involved

^aThese pressure gradients decrease during the dwell due to glucose absorption (61% per 4 h on average).

indirectly in the regulation of the effective peritoneal vascular surface area.

Clinical judgement and the determination of the abovementioned mediators make it likely that two types of inherent fast transporters can be distinguished. Both types cause decreased ultrafiltration. One is associated with comorbidity [27,36] and is characterized by high plasma and dialysate concentrations of the inflammatory cytokine IL-6 and of VEGF [37,38]. The other type can be present in patients without marked comorbidity. This type is associated with a high effluent CA 125 concentration suggesting a link with mesothelial cell mass [39-41]. Only effluent VEGF was increased in patients with this type, and it appeared that the relationship between solute transport and CA 125 was probably mediated by VEGF [41]. This type disappears spontaneously [42], because the mesothelial cell mass decreases with the duration of PD, as judged from CA 125 values [43].

Ultrafiltration failure due to an inherent fast transport status or a high ELAR is usually not a clinical problem, because these patients will generally still produce urine. This urine production can be stimulated with high-dose loop diuretics [44,45]. Also the use of icodextrin for the long dwell is especially effective in fast transporters [12].

Ultrafiltration failure during peritonitis

Acute peritonitis causes ultrafiltration failure, which is usually reversible, due to an inflammation-induced enlargement of the effective peritoneal vascular surface area and increased peritoneal blood flow [46,47]. The latter leads to a fast transport status [46,47] causing a rapid disappearance of the osmotic gradient, and consequently ultrafiltration failure. Free water transport is not impaired [48]. The development of the fast transport status is probably mediated by IL-6 and tumour necrosis factor alpha (TNF- α) [49]. The alterations in a transport status are temporary and usually reversible within 1 or 2 weeks after the institution of antibiotic treatment. Similar to other situations with a fast transport status, the use of icodextrin is very effective in increasing ultrafiltration [50,51].

Ultrafiltration failure in long-term PD

Ultrafiltration failure, as defined by the guidelines of the International Society for Peritoneal Dialysis (ISPD), is present in 36% of patients treated with PD for more than 4 years [52]. Age, gender, duration of PD and peritonitis incidence were not different from patients without this complication. The associated fast transport status in this patient group with ultrafiltration failure is reflected in high values of the dialysate/plasma (D/P) ratio of creatinine, the mass transfer area coefficient (MTAC) of creatinine and a high glucose absorption. In addition, free water transport is often impaired (see below).

Table 2 shows a comparison between the causes of ultrafiltration failure in patients treated shorter than 2 years and longer than 4 years [53]. A combination of factors is often present, but impaired free water transport was especially

Table 2. Causes of early and late ultrafiltration failure, expressed as % of the patients in whom one or more causes could be identified [53]

Early UF failure (<2 years)	Late UF failure (>4 years)	
44	61	
68	30*	
8	43*	
	(<2 years) 44	

MTAC: mass transfer area coefficient; ELAR: effective lymphatic absorption rate.

*P < 0.01.

The total of the percentages is more than 100% because more than one cause was present in a number of patients.

found in long-term PD. Selective drop out of patients with a high ELAR could explain the lower value in the long-term group. These findings are in line with those of Davies, who reported a qualitative difference in the decrease of ultrafiltration compared to expected values based on D/P creatinine [54]. This underlines that long-term ultrafiltration failure is not only caused by fast solute transport rates. A study in 50 patients with ultrafiltration failure, in whom the effect of the duration of PD on transport could be analysed, showed an increase of the MTAC creatinine, but also a decrease of small-pore fluid transport and of free water transport. Also, the contribution of free water transport decreased, especially after 5 years, as well as the osmotic conductance to glucose [55]. In all time periods, a relationship between free water transport and the osmotic conductance to glucose was present. The development of ultrafiltration failure is associated with vascular abnormalities in the peritoneum. These are often accompanied by increased fibrosis [7]. Up to now, no substance is known that can be determined in peritoneal effluent and that could represent the amount of fibrosis. Loss of mesothelial cells may also occur [7], and is probably reflected in a decrease of effluent CA 125 [56].

In summary, ultrafiltration failure in long-term PD is most often due to a combination of a rapid disappearance of the osmotic gradient, together with a decrease in the osmotic conductance to glucose. The latter leads to a decrease of free water transport. It has not been elucidated whether the decreased osmotic conductance is the consequence of a decrease in aquaporin-1 function, or is due to a lower peritoneal ultrafiltration coefficient.

Ultrafiltration failure in long-term PD patients is a serious problem. It can easily cause overhydration, because those patients will often be anuric. Overhydration can lead to an increased risk for cardiovascular death. A relationship has been found between peritoneal ultrafiltration and death in two studies in anuric patients [57,58].

Figure 2 shows a scheme of the various types of fast transport that can cause ultrafiltration failure.

Definition and detection of ultrafiltration failure

Hypervolaemia or overhydration is a clinical diagnosis that can have a number of causes, like a decreased urine production combined with the usual oral fluid intake. The

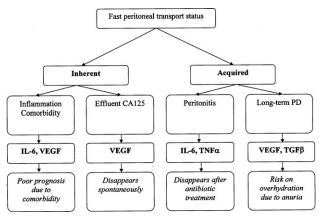


Fig. 2. A schematic representation of the potential pathogenic pathways for different types of fast transport as causes of ultrafiltration failure. Also, the prognosis and the presumed contribution of peritoneal cytokines/growth factors are indicated.

ultrafiltration volume is dependent on the status of the peritoneum. Its function should therefore be investigated during a standardized condition. The world-wide-used peritoneal equilibration test (PET) is useful for this, but only with some modifications [59]. The most important one is the use of a 3.86% glucose solution, instead of the original 2.27%. This allows better assessment of the drained volume. Also, free water transport can be estimated. The use of different glucose concentrations has no effect on D/P creatinine, but influences the D_4/D_0 ratio for dialysate glucose [60]. The higher the glucose concentration, the lower the D_4/D_0 ratio. The transition of a 2.27% PET to a 3.86% PET does not influence longitudinal follow-up of D/P creatinine in individual patients. It also does not matter whether the investigation is done with a conventional or more biocompatible dialysis solution [61].

The definition of ultrafiltration failure by the ISPD can be summarized as the 3×4 rule: (1) less net ultrafiltration than 400 mL, (2) after a dwell of 4 h, (3) of a 4% (3.86%/4.25%) glucose based dialysis solution [62]. The 400 mL limit is based on sparse data from literature [63,64] and clinical data of patients who fulfil this criterion. A cause for ultrafiltration failure could be identified in >95% of these patients [52]. The prevalence of ultrafiltration failure is higher with a limit of 500 mL, but a cause can be defined less often.

The 3.86% glucose PET (modified PET) provides information on net ultrafiltration after 4 h, D/P creatinine (or MTAC creatinine), D_4/D_0 glucose (or glucose absorption, expressed as percentage of the instilled quantity) just like the 2.27% PET, but also information on free water transport. The latter is calculated from the transport of sodium to the peritoneal cavity. Convection is the main mechanism of sodium transport [65] because the concentrations in plasma and dialysate are only slightly different. The importance of diffusion increases, however, in the presence of a relatively large gradient, like in hypernatraemia, or in the presence of a large diffusion area, as present in for instance fast transporters. Free water transport can be underestimated under these conditions when no diffusion correction for sodium is applied (see below).

Determination of D/P Na⁺ after 1 h is the simplest way to obtain information on free water transport. D/P Na⁺ after 4 h can also be used when a diffusion correction is applied [66]. A precise calculation of free water transport during the first hour of a PET is possible, assuming that the small pores offer no hindrance to sodium transport [67]. This requires the determination of the intraperitoneal volume after 1 h to allow the calculation of the quantity of sodium transported in this period. This amount, divided by the sodium concentration in the small pores (mean of plasma and dialysate sodium), yields the amount of fluid transported through the small pores. Free water transport is the difference between total net fluid transport and the volume transported through the small pores. The percentage-free water transport is the volume of free water transport divided by the total amount of fluid transported [19].

The 3.86% glucose mini-PET, in which the peritoneal cavity is drained after 1 h to determine the volume and dialysate Na⁺, is an elegant method for assessment of free water transport [67]. However, the interpretation of D/P ratios is difficult because MTACs are higher during the first hour of a 4-h dwell than in the period thereafter [13]. It is not possible to use a correction factor to translate to a 4-h dwell [68]. This limits its application for overall assessment of peritoneal function. Determination of the osmotic conductance to glucose is usually done by an estimation of sigma using kinetic modelling and calculation of LpA with a volume marker [54]. A simple alternative has recently been described by La Milia *et al.* using two mini-PETS: one with 1.36% glucose and one with 3.86% (double mini-PET) [69].

The modified PET and the mini-PET can be combined when the peritoneal cavity is drained after 1 h to determine the volume at that time by weighing. After taking a dialysate sample, the drained volume is reinfused and left for another 3 h. With this method, no differences were found for net ultrafiltration and small solute transport compared to those in a modified PET [70].

The personal dialysis capacity test (PDC) is performed by the patients themselves with five exchanges per 24 h [71]. These are different in dwell time and glucose concentration. Kinetic modelling is applied to estimate parameters like surface area available for diffusion, fluid absorption and large-pore flow. Superiority of the PDC over the PET has been claimed [72]. However, the PDC has a number of disadvantages like the risk of inaccuracies, a large number of laboratory investigations, the assumptions used for kinetic modelling and most importantly, the exclusion of sodium kinetics. This makes the PDC less suitable for the analysis of ultrafiltration failure, than the modified PET. A summary of the advantages and limitations of the various peritoneal function tests without the use of a volume marker is given in Table 3.

Assessment of the peritoneum should also include an effluent marker. Most experience has been obtained with CA 125. A single low value is difficult to interpret [73], but a downward trend with time suggests a decrease of mesothelial cell mass. The CA 125 concentration in effluent is dependent on the dwell time. It increases linearly during a dwell of 4 h and is not influenced by the dialysis solution used [55]. A linear increase is also present during

Table 3. Advantages and limitations of the various peritoneal function test without a volume marker

Test ^a	Advantages	Limitations
Original PET	Widely used	Limited information
	Gold standard for small solute transport	No Na ⁺ sieving
		No FWT
		No OC
Modified PET	Definition for UFF	No FWT
	Na ⁺ sieving D/P creatinine similar to original PET	No OC
Peritoneal dialysis capacity test	Large-pore flow using albumin	No Na ⁺ sieving
• •	Peritoneal absorption	No FWT
	Area parameter	No OC
Mini-PET	FWT	D/P creatinine difficult to compare with PET values
		No OC
Double mini-PET	FWT	D/P creatinine difficult to compare with PET values
	OC	Two tests
Modified PET with temporary drainage	Definition of UFF	No OC
r 7	Na ⁺ sieving FWT	
	D/P creatinine similar to original PET	

^aThe value of the test is increased when an effluent CA125 determination is added.

FWT: free water transport; OC: osmotic conductance.

longer dwells [74]. Therefore, it is recommended to express effluent CA 125 as its dialysate appearance rate, that is the product of the concentration and the drained volume, divided by the dwell time. An algorithm for the follow-up of patients is given in Figure 3.

Reasons for elective discontinuation of PD

Both clinical indications and those that are related to the transport function of the peritoneum can be reasons to switch from PD to HD. A Kt/V_{urea} < 1.7/week without clinical manifestations of underdialysis is not a reason for discontinuation of PD [75]. It is evident that untreatable overhydration caused by ultrafiltration failure is an indication to switch to HD. Yet, studies in patients with residual renal function have—with one exception [76]—not been able to show an effect of ultrafiltration on patient survival [77,78]. An effect of peritoneal ultrafiltration on survival is, however, present in patients without urine production [57,58]. Until now, it has been impossible to define a minimum ultrafiltration volume [79], because the occurrence of overhydration is dependent on oral fluid intake and peritoneal ultrafiltration.

At present, it is not possible to give fixed limits of any peritoneal transport parameter below or above which PD

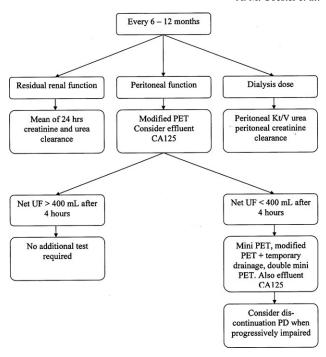


Fig. 3. A suggested algorithm for follow-up of patients on peritoneal dialysis.

should be discontinued, because longitudinal investigations on the time course of peritoneal function are only limited. Signs that indicate membrane damage are the development of a fast transport status with ultrafiltration failure, the development of a reduction in free water transport and a decrease of effluent CA 125 during the time course of PD.

Recommendations for patient information and measurement of peritoneal membrane function in PD patients

- 1. The following items should be discussed during the information to predialysis patients:
 - a. The survival of PD patients is better than that of HD patients during the first years of dialysis, probably because of better preservation of residual renal function.
 - Functional and morphological peritoneal abnormalities may occur in about one-third of patients treated for more than 4 years.
 - c. An elective switch to HD should be considered when the above-mentioned abnormalities develop to minimize the risk of encapsulating peritoneal sclerosis.
- Measurement of peritoneal membrane function should be incorporated in the follow-up of PD patients. The frequency is at least once a year.
- 3. The measurement should be done with the most hypertonic solution, glucose concentration around 4%,

- during a standardized dwell of 4 h. Ultrafiltration failure is defined as net ultrafiltration <400 mL.
- 4. Information should be obtained on the transport of creatinine, glucose and sodium. Net ultrafiltration should be measured and insight into free water transport should be obtained. Determination of effluent CA 125 gives an added value.

Conflict of interest statement. Watske Smit is a part time employee of Baxter, The Netherlands.

References

- Krediet RT, Boeschoten EW, Dekker FW. Why is the evidence favoring hemodialysis over peritoneal dialysis misleading? Semin Dial 2007; 20: 205–208
- Krediet RT. Advances in peritoneal dialysis. Minerva Urol Nephrol 2007; 59: 251–260
- Van Den Wall Bake AWL, Kooman JP, Lange JM et al. Adequacy of peritoneal dialysis and the importance of preserving residual renal function. Nephrol Dial Transplant 2006; 21(Suppl 2): 34–37
- Krediet RT. How to preserve residual renal function in patients with chronic kidney disease and on dialysis? *Nephrol Dial Transplant* 2006; 21(Suppl 2): 42–46
- Di Paolo N, Sacchi G. Peritoneal vascular changes in continuous ambulatory peritoneal dialysis (CAPD): an in vivo model for the study of diabetic microangiopathy. Perit Dial Int 1989; 9: 41–45
- Honda K, Nitta K, Horita S et al. Morphological changes in the vasculature of patients on CAPD with ultrafiltration failure. Nephron 1996; 72: 171–176
- Mateijsen MAM, Van Der Wal AC, Hendriks PMEM et al. Vascular and interstitial changes in the peritoneum of CAPD patients with peritoneal sclerosis. Perit Dial Int 1999; 19: 517–525
- Williams JD, Craig KJ, Topley N et al. Morphologic changes in the peritoneal membrane of patients with renal disease. J Am Soc Nephrol 2002; 13: 470–479
- Imholz ALT, Koomen GCM, Struijk DG et al. The effect of an increased intraperitoneal pressure on fluid and solute transport during CAPD. Kidney Int 1993; 44: 1078–1085
- Twardowski ZJ, Khanna R, Nolph KD et al. Intra-abdominal pressures during natural activities in patients treated with continuous ambulatory peritoneal dialysis. Nephron 1986; 44: 129–135
- Twardowski ZJ, Prowant BF, Nolph KD et al. High volume, low frequency continuous ambulatory peritoneal dialysis. Kidney Int 1983; 23: 64–70
- Ho-dac-Pannekeet MM, Schouten N, Langedijk MJ et al. Peritoneal transport characteristics with glucose polymer dialysate. Kidney Int 1996; 50: 979–986
- Imholz ALT, Koomen GCM, Struijk DG et al. Fluid and solute transport in CAPD patients using ultralow sodium dialysate. Kidney Int 1994; 46: 333–340
- Rippe B, Stelin G. Simultations of peritoneal solute transport during CAPD. Application of two-pore formatism. *Kidney Int* 1989; 35: 1234–1244
- Pannekeet MM, Mulder JB, Weening JJ et al. Demonstration of aquaporin-CHIP in peritoneal tissue of uremic and CAPD patients. Perit Dial Int 1996; 16(Suppl 1): S54–S57
- Nolph KD, Hano JE, Teschan PE. Peritoneal sodium transport during hypertonic peritoneal dialysis. *Ann Intern Med* 1969; 70: 931

 941
- Nolph KD, Twardowski ZJ, Popovich RP et al. Equilibiration of peritoneal dialysis solutions during long-dwell exchanges. J Lab Clin Med 1979: 93: 246–256
- Ni J, Verbavatz JM, Rippe A et al. Aquaporin-1 plays an essential role in water permeability and ultrafiltration during peritoneal dialysis. Kidney Int 2006; 69: 1518–1525

- Smit W, Struijk DG, Ho-dac-Pannekeet MM et al. Quantification of free water transport in peritoneal dialysis. Kidney Int 2004; 66: 849–854
- Parikova A, Smit W, Struijk DG et al. The contribution of free water transport and small pore transport to the total fluid removal in peritoneal dialysis. Kidney Int 2005; 68: 1849–1856
- Rippe B. Free water transport small pore transport and the osmotic pressure gradient three pore model of peritoneal transport. *Nephrol Dial Transplant* 2008; 23: 2147–2153
- Smit W, van Dijk P, Langedijk MJ et al. Peritoneal function and assessment of reference values using a 3.86% glucose solution. Perit Dial Int 2003; 23: 440–449
- Michels WM, Zweers MM, Smit W et al. Does lymphatic absorption change with the duration of peritoneal dialysis? Perit Dial Int 2004; 24: 347–352
- Krediet RT. The effective lymphatic absorption rate is an accurate and useful concept in the physiology of peritoneal dialysis. *Perit Dial Int* 2004; 24: 309–317
- Coester AM, Struijk DG, Smit W et al. The cellular contribution to effluent potassium and its relation with free water transport during peritoneal dialysis. Nephrol Dial Transplant 2007; 22: 3593–3600
- Churchill DN, Thorpe KE, Nolph KD et al. for the CANUSA Study Group. Increased peritoneal membrane transport is associated with decreased patient and technique survival for continuous peritoneal dialysis patients. J Am Soc Nephrol 1998; 9: 1285–1292
- Chung SH, Chu WS, Lee HA et al. Peritoneal transport characteristics, comorbid diseases and survival in CAPD patients. Perit Dial Int 2000; 20: 541–547
- Rumpsfeld M, McDonald SP, Purdie DM et al. Predictors of baseline peritoneal transport status in Australian and New Zealand peritoneal dialysis patients. Am J Kidney Dis 2004; 43: 492–501
- Topley N, Williams JD. Role of the peritoneal membrane in the control of inflammation in the peritoneal cavity. *Kidney Int* 1994; 46(Suppl 48): S71–S78
- Selgas R, del Peso G, Bajo M-A et al. Spontaneous VEGF production by cultured mesothelial cells from patients on peritoneal dialysis. Perit Dial Int 2000; 20: 798–801
- Visser CE, Brouwer-Steenbergen JJE, Betjes MGH et al. Cancer antigen 125: a bulk marker for the mesothelial mass in stable peritoneal dialysis patients. Nephrol Dial Transplant 1995; 10: 64–69
- Topley N, Jörres A, Luttmann W et al. Human peritoneal mesothelial cells synthesize IL-6: induction by IL-Iβ and TNFαKidney Int 1993;43:226–233
- Betjes MGH, Tuk CW, Struijk DG et al. Interleukin-8 production by human peritoneal mesothelial cells in response to tumor necrosis factor-α, interleukin-1 and medium conditions by macrophages cocultured with staphylococcus epidermidis. J Infect Dis 1993; 168: 1202–1210
- Ho-Dac-Pannekeet MM, Krediet RT. Inflammatory changes in vivo during CAPD. What can the effluent tell us? Kidney Int 1996; 50(Suppl 56): S12–S16
- Krediet RT. Dialysate cancer antigen 125 concentration as marker of peritoneal membrane status in patients treated with chronic peritoneal dialysis. *Perit Dial Int* 2001; 21: 560–567
- Davies SJ, Brian J, Phillips L et al. The predictive value of Kt/V and peritoneal solute transport in CAPD patients in dependent on the type of comorbidity present. Perit Dial Int 1990; 16(Suppl 1): S158

 S162
- Pecoits-Filho R, Araujo MRT, Lindholm B et al. Plasma and dialysate IL-6 and VEGF concentrations are associated with high peritoneal solute transport rate. Nephrol Dial Transplant 2002; 17: 1480– 1486
- Pecoits-Filho R, Carvalho MJ, Stenvinkel P et al. Systemic and intraperitoneal interleukin-6 system during the first year of peritoneal dialysis. Perit Dial Int 2006; 26: 53–63
- Rodrigues A, Almeida M, Fonseca I et al. Peritoneal fast transport in incident peritoneal dialysis patients is not consistently associated with systemic inflammation. Nephrol Dial Transplant 2006; 21: 763– 769

- Rodrigues AS, Martins M, Korevaar JC et al. Evaluation of peritoneal transport and membrane status in peritoneal dialysis: focus on incident fast transporters. Am J Nephrol 2007; 27: 84–91
- Van Esch S, Zweers MM, Jansen MAM et al. Determinants of peritoneal solute transport rates in newly started nondiabetic peritoneal dialysis patients. Perit Dial Int 2004; 24: 554

 –561
- Lo WK, Brendolan A, Prowant BF et al. Changes in the peritoneal equilibration test in selected chronic peritoneal dialysis patients. J Am Soc Nephrol 1994; 4: 1466–1474
- Ho-dac-Pannekeet MM, Hiralall JK, Struijk DG et al. Longitudinal follow-up of CA 125 in peritoneal effluent. Kidney Int 1997; 51: 888–893
- Van Olden RW, Guchelaar H-J, Struijk DG et al. Acute effects of high-dose furosemide on residual renal function in CAPD patients. Perit Dial Int 2003; 23: 339–347
- Medcalf JF, Harris KPG, Walls J. Role of diuretics in the preservation of residual renal function in patients on continuous ambulatory peritoneal dialysis. *Kidney Int* 2001; 59: 1128–1133
- Krediet RT, Zuyderhoudt FMJ, Boeschoten EW et al. Alterations in the peritoneal transport of water and solutes during peritonitis in continuous ambulatory peritoneal dialysis patients. Eur J Clin Invest 1987; 17: 43–52
- Douma CE, de Waart DR, Struijk DG et al. Are phospholipase A2 and nitric oxide involved in the alterations in peritoneal transport during CAPD peritonitis? J Lab Clin Med 1998; 132: 329–340.
- Smit W, Van Den Berg N, Schouten N et al. Free water transport in fast transport status: a comparison between CAPD peritonitis and long-term PD. Kidney Int 2004; 65: 298–303
- Zemel D, Koomen GCM, Hart AAM et al. Relationship of TNF-α, interleukin-6 and prostogladins to peritoneal permeability for macromolecules during longitudinal follow-up of peritonitis in continuous ambulatory peritoneal dialysis. J Lab Clin Med 1993; 122: 686–696
- Gokal R, Mistiy CD, Peers EM MIDAS Study Group. Peritonitis occurrence in a multicentre study of icodextrin and glucose in CAPD. Perit Dial Int 1994; 15: 226–230
- Posthuma N, Ter Wee PM, Donker AJM et al. Icodextrin use in CCPD patients during peritonitis: ultrafiltration and serum disacharide concentrations. Nephrol Dial Transplant 1998; 13: 2341–2344
- 52. Smit W, Schouten N, Van Den Berg N et al. for the Netherlands Ultrafiltration failure study group. Analysis of the prevalence and causes of ultrafiltration failure during long-term peritoneal dialysis: a cross sectional study. Perit Dial Int 2004; 24: 562–570
- Smit W, Parikova A, Struijk DG et al. The difference in causes of early and late ultrafiltration failure in peritoneal dialysis. Perit Dial Int 2005; 25(Suppl 3): S41–S45
- Davies SJ. Longitudinal relationship between solute transport and ultrafiltration capacity in peritoneal dialysis patients. *Kidney Int* 2004; 66: 2437–2445
- Parikova A, Smit W, Struijk DG et al. Analysis of fluid transport pathways and their determinants in peritoneal dialysis patients with ultrafiltration failure. Kidney Int 2006; 70: 1988–1994
- Ho-dac-Pannekeet MM, Hiralall JK, Struijk DG et al. Longitudinal follow-up of CA125 in peritoneal effluent. Kidney Int 1997; 51: 888– 893
- Brown EA, Davies SJ, Rutherford P et al. Survival of functionally anuric patients on automated peritoneal dialysis: the European APD Outcome study. J Am Soc Nephrol 2003; 14: 2948–2957
- Jansen MAM, Termorshuizen F, Korevaar JC et al. for the NECOSAD study group. Predictors of survival in anuric peritoneal dialysis patients. Kidney Int 2005; 68: 119–1205
- Twardowski ZJ, Nolph KD, Khanna R et al. Peritoneal equilibration test. Perit Dial Bull 1987; 7: 138–147
- Smit W, Langedijk MJ, Schouten N et al. A comparison between 1.36% and 3.86% glucose solutions for the assessment of peritoneal membrane function. Perit Dial Int 2000; 20: 734–741

- Parikova A, Struijk DG, Zweers MM et al. Does the biocompatibility of the PD solution matter for the assessment of peritoneal function? Perit Dial Int 2007; 27: 691–696
- 62. Nolph K, Gokal R, Mujais S *et al.* ISPD ad hoc committee on ultrafiltration management in peritoneal dialysis. *Perit Dial Int* 2000; 20(Suppl 4): S3–S4
- 63. Virga G, Amici G, da Rin G *et al.* Comparison of fast peritoneal equilibration test with 1.36% and 3.86% dialysis solution. *Blood Purif* 1994; 12: 113–120
- 64. Ho-dac-Pannekeet MM, Atasever B, Struijk DG et al. Analysis of ultrafiltration failure in peritoneal dialysis patients by means of the standard peritoneal permeability analysis. Perit Dial Int 1997; 17: 144–150
- Wang T, Waniewski J, Heimburger O et al. A quantitative analysis of sodium transport and removal during peritoneal dialysis. Kidney Int 1997; 52: 1609–1616
- 66. Westra WM, Smit W, Zweers MM et al. Diffusion correction of sodium sieving applicable in a peritoneal equilibration test. Adv Perit Dial 2003; 19: 6–9
- 67. La Milia V, Di Filippo S, Crepoldi M *et al*. Mini-peritoneal equilibration test: a simple and fast method to assess free water and small solute transport across the peritoneal membrane. *Kidney Int* 2005; 68: 840–846
- 68. Rodriquez AS, Silva S, Bravo F *et al.* Peritoneal membrane evaluation in routine clinical practice. *Blood Purif* 2007; 25: 497–504
- La Milia V, Limardo M, Virga G et al. Simultaneous measurement of peritoneal glucose and free water osmotic conductances. Kidney Int 2007; 72: 643–650
- Cnossen TT, Smit W, Konings CJA et al. Quantification of free water transport during the peritoneal equilibration test. Perit Dial Int; in press
- Haraldsson B. Assessing the peritoneal dialysis capacities of individual patients. Kidney Int 1995; 47: 1187–1198
- 72. Van Biesen W, Van Der Tol A, Veys N et al. The personal dialysis capacity test is superior to the peritoneal equilibration test to discriminate inflammation as the cause of fast transport status in peritoneal dialysis patients. Clin J Am Soc Nephrol 2006; 1: 269– 274
- Sanusi AA, Zweers MM, Weening JJ et al. Expression of cancer antigen 125 by peritoneal mesothelial cells is not influenced by duration of nitroneal dialysis. Perit Dial Int 2001; 21: 495
 500
- 74. Akman S, Van Westrhenen R, De Waart DR et al. The effect of dwell time on dialysate cancer antigen 125 appearance rates in patients on continuous ambulatory peritoneal dialysis. Adv Perit Dial 2003; 19: 24–27.8
- The EBPG Expert Group on Peritoneal Dialysis. European best practice guidelines for peritoneal dialysis. *Nephrol Dial Transplant* 2005; 20(Suppl 9): S24–S27
- Ates K, Nergizoglu G, Keven K et al. Effect of fluid and sodium removal on mortality in peritoneal dialysis patients. Kidney Int 2001; 60: 767–776
- Bargman JM, Thorpe KE, Churchill DN. for the CANUSA Peritoneal Dialysis Study Group. Relative contribution of residual renal function and peritoneal clearance to adequacy of dialysis: a reanalysis of the CANUSA study. *J Am Soc Nephrol* 2001; 12: 2158–2162
- 78. Termorshuizen F, Korevaar JC, Dekker FW et al. The relative importance of residual renal function compared with peritoneal clearance for patient survival and quality of life: an analysis of the Netherlands Cooperative Study on the Adequacy of Dialysis (NECOSAD)-2. Am J Kidney Dis 2003; 41: 1293–1302
- Lo WK, Bargman JM, Burkart J et al. Guideline on targets for solute and fluid removal in adult patients on chronic peritoneal dialysis. Perit Dial Int 2006; 26: 520–522

Received for publication: 27.5.08 Accepted in revised form: 15.12.08