



Sexual Dimorphism of Immune Responses: A New Perspective in Cancer Immunotherapy

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Nowadays, several types of tumors can benefit from the new frontier of immunotherapy, due to the recent increasing knowledge of the role of the immune system in cancer control. Among the new therapeutic strategies, there is the immune checkpoint blockade (ICB), able to restore an efficacious antitumor immunity and significantly prolong the overall survival (OS) of patients with advanced tumors such as melanoma and non-small cell lung cancer (NSCLC). Despite the impressive efficacy of these agents in some patients, treatment failure and resistance are frequently observed. In this regard, the signaling governed by IFN type I (IFN-I) has emerged as pivotal in orchestrating host defense. This pathway displays different activation between sexes, thus potentially contributing to sexual dimorphic differences in the immune responses to immunotherapy. This perspective article aims to critically consider the immune signals, with particular attention to IFN-I, that may differently affect female and male antitumor responses upon immunotherapy.

Keywords: cancer, immunotherapy, sexual dimorphism, interferon, immune response, immune checkpoint inhibitors, therapeutic vaccines

In the last decade the increasing knowledge of the role of the immune system in cancer control has led to the development of the immune checkpoint inhibitors (ICIs), that targeting proteins acting as negative regulators of T-cell activation, reverse the tumor-induced immune tolerance.

The main actors of this scene are two key molecules called cytotoxic T-lymphocyte-associated protein 4 (CTLA-4) and programmed cell death receptor 1 (PD-1) (1). CTLA-4 binds B7 receptors on antigen-presenting cells (APCs), thus blocking T-cell activation. Likewise, PD-1 expressed on activated T lymphocytes, upon interaction with its ligands PD-L1 and PD-L2 on APCs or tumor cells, limits their activity delivering negative signals. PD-1 is also expressed by regulatory T cells (Tregs), enhancing their function. The clinical use of ICIs has extraordinarily increased overall survival (OS) in patients with cancer, suggesting that targeting the immune checkpoint blockade (ICB) is a privileged strategy for fighting cancer (2, 3). A crucial part of this game is played by tumor-infiltrating T cells (4) and in many cases, the effectiveness of ICIs is limited by the lack of adequate antitumor immunity in the tumor microenvironment (TME) (5). To overcome this limitation, new combination therapies are being investigated (6). In this landscape, a new perspective for improving the efficacy of immunotherapy is to take into account sexually dimorphic differences of immune responses (7).

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COMPONENTS OF ANTITUMOR IMMUNITY CRITICAL FOR IMMUNOTHERAPY

In the era of ICIs, T-cell immunity remains central for tumor regression (8). This is far from being simple and occurs only when diverse elements coexist either during cancer immunosurveillance or immunotherapeutic treatments (9). Cancer cells carry tumor antigens, and in particular neoantigens, that are the most capable of inducing an effective T-cell immunity. These antigens, mainly present in tumors with high mutational rate, generate T-cell responses upon processing and presention by DCs. Accordingly, CD8+ T cells infiltrates at high frequency these tumors and this associates with a patient survival advantage (10). Nevertheless, in specific conditions tumor-infiltrating DCs lack their immunostimulatory function and acquire immunosuppressive activity (11). This relies on different phenotypes and functions of DC subsets that provide additional variability to the onset of antitumor responses. Within human tumors, rare BDCA3+ DCs were found extremely competent in processing and cross-presenting antigens, driving the expansion of tumor-specific cytotoxic T cells (CTLs) (12). These DCs are the equivalent of the mouse CD103+ DCs, depending uniquely on DC-lineage committing transcription factors such as interferon regulatory factor 8 (IRF-8) (13). Tumor-infiltrating CD103+/ BDCA3+ DCs represent privileged players for responses upon ICB (14, 15). However, in TME the most abundant DCs show immunosuppressive activity, including plasmacytoid DC (pDCs). Activation by toll-like receptor 7 (TLR7) ligand reverses pDC immunosuppressive function to such an extent that their administration to melanoma patients induces tumor-specific CD4+ and CD8+ T-cell responses (16, 17). PDCs are the main producers of IFN- α (18), a well-known cytokine linking innate and adaptive immunity and endowed with potent direct and immunomediated antitumor effects (19). Endogenous IFN-I is required to initiate antitumor response in the elimination phase of cancer immunoediting (20), and IFN-I production is also essential for tumor rejection by DC-stimulated activated T cells (21). IFN-I specifically improves the ability of CD8a+ CD103+ DCs to cross-prime tumor specific CD8+ T cells (22-24). However, although IFN-I is crucial for the outcome of ICB, its activation may produce an opposite role in a time-dependent manner (25). In melanoma, while early IFN-I activation correlates with an effective PD-1 blockade (5, 26), prolonged IFN-I signaling seems to favor resistance (27). This apparent discrepancy might not be surprising since, during persistent viral infections, IFN-I displays protective effects in early stages and becomes detrimental upon continued signaling activation in the chronic phase (28). Interestingly, CML patients in remission who stopped IFN-a treatment because of good clinical response developed higher protective T-cell memory response than patients who continued therapy (29). In TME, while IFN-I signaling is critical for survival and full activation of CD8+ T cells, the continuous generation of terminally differentiated CD8+ T cells might determine the failure of ICB by reducing progenitors able to respond (30). Thus, the generation of exhausted CD8+ T cells may be the results of complex immunosuppressive interactions including prolonged IFN-I signaling (31). Indeed, IFN-I is critical for the induction of immune checkpoints or coinhibitory receptors, such as PD-1 and T-cell immunoglobulin- and mucin-containing molecule-3 (Tim-3) (32, 33), whose persistent overexpression characterizes exhausted T cells within TME (34). Moreover, acute and chronic IFN-I expression impacts Tregs differently. While IFN-α abrogates the suppressive activity of CD4+ CD25+ Foxp3+ Tregs, thus hampering tumor evasion (35, 36), IFN-I signal blockade boosts the ICB-induced antitumor response by favoring the effector T cells to Tregs ratio (37). Of interest, Treg-mediated immune tolerance may occur through the control of IFN-I on indoleamine 2,3-dioxygenase (IDO) (38, 39), being melanoma peritumoral IDO expression by pDCs an early marker of resistance (40). Noteworthy, both Tregs and IFN-I also suppress the function of natural killer (NK) cells whose antigen-independent cytotoxicity is the second effector mechanism responsible for an efficacious antitumor response (41, 42). Altogether, these immune components build a functional framework for innate and acquired resistance to PD-1 blockade (43), where the balance between inflammation and suppression determined by the fine-tune regulation of IFN-I is crucial. This central achievement occurs through the modulation of different transcriptional programs involving the Janus kinase (JAK)-signal transducer and activator of transcription (STAT) and IFN regulatory factors (IRFs) families, as well as proteins of the phosphatidyl inositide3-kinase (PI3K) and mitogen-activated protein kinases (MAPK) pathways (44). Moreover, epigenetic signals, such as histone modification, DNA methylation, and microRNAs (miRs), represent a second key layer of regulation (45). Indeed, exhausted CD8+ T cells within tumors display a distinct epigenetic profile and the limited remodeling may represent a pivotal component of resistance to PD-L1 blockade (46). Likewise, miRs modulate the expression of PD-L1 potentially accounting for innate resistance (47). Finally, it is worthy to mention drug toxicity as one of the major cause of reduced dosage, delayed drug administration and therapy discontinuation (48). Immune-related adverse events (irAEs), such as the loss of the protective function of intestinal barriers and changes occurring in the microbiota composition, represent the most frequent ICI-associated toxicities to which dysregulated activation of the IFN-I signaling may contribute (49, 50).

SEXUAL DIMORPHISM OF THE IMMUNE COMPONENTS OF HOST RESPONSE TO IMMUNOTHERAPY

Sexual dimorphism of the immune functions is a crucial element that has so far been largely ignored in the field of immunotherapy (51). These differences affect both innate and adaptive immune responses, leading to a considerable functional diversity between females and males (7) (**Figure 1**). Sex variations include the number and activity of cells as well as intracellular and extracellular signals orchestrating the two branches of immunity. In the innate context, females own APCs that perform antigen presentation more vigorously, have neutrophils and macrophages endowed



with higher phagocytic activity, and show a higher frequency of both progenitors and mature group 2 innate lymphoid cells (ILCs), key regulators of type-2 inflammatory responses (52). On the contrary, males exhibit enhanced numbers of NK cells (7). In the adaptive context, females exhibit higher CD4+ T cell counts associated with an increased CD4+/CD8+ T-cell ratio, along with Th2 prevalence, and greater proliferation and cytotoxicity of T cells. In contrast, males have higher CD8+ and Treg cell counts associated with Th1 dominance, lower B cell numbers and basal immunoglobulin levels along with weaker antibody responses (53). Both hormonal and genetic differences concur in the sexual dimorphism of the immune system. The 17β-estradiol (E2) –estrogen receptor α (ER) axis is a key regulator of innate immune populations. E2 reduces mobility and inflammatory activity of neutrophils (54), and female mMDSCs seem to be more suppressive than the male counterpart (55). Importantly, the E2–ERα axis exerts a tight control on the functional responses of diverse DC subsets, also by modulating IFN-I production. Moreover, high levels of E2 promote epigenetic changes in DC precursors of females driving DC differentiation and robust IFN-I production (56) (Figure 2). PDCs are the DC with major differences between the sexes. Their activity is driven by TLR7, whose gene located on X chromosome and under the E2-ERa signaling (57, 58) promotes high IFN- α production through enhanced expression of IRF-5 (59) (Figure 2). Nevertheless, the development and function of other DC subsets are also affected by the E2–ER α axis (60). In female mice, CD103+ conventional DCs (cDCs) are represented at very high levels in the cutaneous lymph nodes and CD103– cDCs show high expression of ER α in the lung (61, 62). Furthermore, E2 treatment has been reported to enhance IRF-4+ DC capability to stimulate Th17 in a murine

model of HSV-2 infection (63), while it induces IFN- α and IL-6 production as well as CD40, CD86, and MHCII expression in cDCs. In contrast, tolerogenic FOXO3-expressing DCs display lower frequency and reduced function in tumors from females compared with their male counterpart (64). Altogether, these evidences confirm a positive regulatory feedback loop between the E2–ER α and IFN-I signals in regulating the phenotype and function of DC populations (65). DC differentiation and function are also affected by other hormones, such as prolactin, progesterone and glucocorticoids, driving either a pro-inflammatory or a tolerogenic phenotype (66). In this light, the sex-specific components of the innate immunity become important in improving cancer immunotherapy. Many elements of the adaptive immunity are regulated by the E2-ERa axis as well. Low levels of estrogen, as during the luteal phase, favor Th1 polarization of CD4+ T cells associated with increased production of IFN-y, responsiveness to IL-12 through STAT-4 activation, and T proliferation. Conversely, high levels of estrogens, found in the follicular phase and during pregnancy, sustain Th2-mediated immunity characterized by IRF-1-mediated reduction of IFN-y, IL-4 induction, and PD-1-overexpressing Tregs associated with reduced Th17 response (67). Nevertheless, estrogen together with other factors may directly stimulate Th17 cells inducing IL-17 production (68). Thus, females exhibit activated and proliferating CD4+ and CD8+ T cells, characterized by preferential production of IFNy and high-cytotoxicity activity, whereas males exhibit IL-17-producing T cells (69). This underlines how sex hormones become crucial in determining the efficacy of some therapies. B16 melanoma-bearing female mice, more than males, benefit from ICB, partially due to a greater PD-L1 blockade-mediated reduction of Treg function (70).



FIGURE 2 | Sex-biased regulation of T-cell response through E2-induced IFN type I (IFN-I) production. The E2–ER α axis controls the functional responses of diverse DC subsets along with IFN-I production. In females, upon 17 β -estradiol (E2) stimulation plasmacytoid DC (pDCs) express enhanced interferon regulatory factor 5 (IRF-5) and reduced indolearnine 2,3-dioxygenase (IDO) leading to transient production of high levels of IFN- α that, in turn, stimulate CD8+ T-cell activity and downregulate regulatory T (Treg) cells. CD8+ T cell can also be stimulated by activation of BDCA3+ DCs, whose activation is interferon regulatory factor 8 (IRF-8) dependent. Moreover, the activity of CD8+ cells is directly modulated by the E2–ER α axis in a hormone dosage-dependent manner. In this context, IFN-I maybe the signal which, in some conditions such as the onset of the antitumor response restored by immune checkpoint inhibitor, drives a more powerful inflammatory cell-mediated immune response in female.

The molecular point of view of the sexual immune dimorphism is also extremely intriguing. Many key immune-related genes, such as FOXP3 and CD40L, are located on the X chromosome (71, 72), and numerous X-linked genes of T cells carry the estrogen response elements (EREs) in their promoter. Hence, T cells from women may display a sex-biased signature characterized by inflammatory/cytotoxic effector genes such as IFN-y, granulysin (GNLY), granzyme A (GZMA), RIGI, LTβ, IL12Rβ2, OAS1, IF16, X3CL1, CX3CL2, and the cytokines IL-15 and IL-16 (73). Therefore, the X chromosome may be responsible, at least in part, for the immunological advantage of females, whose signals are potentially more suitable to be activated by immunotherapies. This assumption is confirmed by the finding that the X chromosome is highly enriched in miRs as about 118 are located in this position, whereas only two miRs have been identified on the Y chromosome (74). MiRs sex-specific expression may have an enormous regulatory power on immune responses, through the control of signals and function of specific immune cell populations. For instance, CD4+ T cells from female lupus patients have been found to overexpress 18 X chromosome-linked miRs with respect to the male counterpart (75). The PD-L1 expression itself, directly or through the control on trascription factors, appears to be modulated by several X-linked miRs, including miR-221, miR-222, miR-106b, miR-20b, and miR-513 (47); of interest, the X-linked miR-424 targets both PD-L1 and CD80 regulating concurrently the PD-L1/PD-1 and CD80/CTLA-4 pathways (76). Notably, a close crosstalk has been reported between the E2-ERa axis estrogen activity and epigenetic regulation by X-linked miRs, since molecules such as miR-221 and miR222 bind and regulate the ER α transcript (74).

The impact of sex differences on antitumor immunity is critical. In melanoma patients, partially exhausted cytotoxic CD8+ T lymphocytes (peCTLs), upon increased expression and engagement of CTLA-4 and PD-1, drive the dysregulation of the host response. Therefore, while both sexes with high peCTLs show a similar objective response rate (ORR) following anti-PD-1 monotherapy as well as combination therapy, females with low peCTLs exhibit higher ORR after combination therapy (77). In the sex-biased immune landscape the complex interplay between microbiome, host immune system, and tumor is pivotal (49). The composition of gut microbiome affects responsiveness to ICIs, as melanoma patients with a greater variety of gut bacteria have high frequence of tumor-infiltrating CTLs predicting a better prognosis (78). In a murine melanoma model, the commensal Bifdobacterium was shown to improve ICB through DC activation associated with higher tumor-infiltrating CD8+ T cells (79). Also, irAEs, such as diarrhea and colitis due to gut bacteria composition changes and intestinal barrier dysfunction (49), may have a sex-biased occurrence as gut microbiome is under a hormone-dependent control (80).

THE IMPLEMENTATION OF IMMUNOTHERAPY ON A SEX-BASED PERSPECTIVE

Vaccines was one of the first cancer immunotherapy approach. Despite most of them elicit antigen-specific immune responses in several clinical settings, they have largely failed in achieving a survival benefit (81), due to both tumor-cell intrinsic and

extrinsic factors circumventing immune recognition and creating a suppressive TME (82).

Among therapeutic cancer vaccines, DCs have been regarded as a promising approach (83) and have been tested on wide range of tumor types (84). A comprehensive meta-analysis has demonstrated tumor-specific T-cell response in 77% of prostate and 61% of Renal Cell Carcinoma (RCC) patients (85). Although a correlation between DC-induced antigen-specific immune responses and prolonged patient survival has been confirmed in many studies (86, 87), the ORR does not exceed 15-20% (88). Moreover, most of these studies are phase-I/-II trials, involving few patients, whose immune and clinical responses were not evaluated taking into account patient characteristics, including sex. Future promising areas of investigation will be the use of personalized vaccines targeting neoantigens and their combination with immunomodulatory agents limiting the inhibitory signals in the TME (82, 89). In this context, due also to the hormonal modulation of many immune populations, including DCs (66), it would be extremely beneficial to consider how to take advantage of the sex-specific immune components.

A major paradigm shift in cancer immunotherapy was the use of antibodies targeting the immune-inhibitory pathways to unleash anticancer T-cell responses (90). Ipilimumab, targeting CTLA-4, was the first antibody clinically approved for treating patients with advanced unresectable melanoma (91). Soon after, nivolumab and pembrolizumab were developed to target PD-1. Today, ICIs are in the front line of immunotherapy of various advanced cancers; partial or complete objective responses have been obtained in patients with melanoma (31-44%) (92), NSCLC (20%) (93), and RCC (22-25%) (94). However, while such therapies exhibit clinical efficacy in many patients, the lack of response in a significant fraction of them remains the major concern. In this regard, intratumoral-infiltrating T cells correlate with a favorable outcome in melanoma patients, to the extent that analyses of type, functional orientation, density, and spatial location of these cells have been developed into a predictive immune scoring system (95). Accordingly, ICI efficacy seems to correlate with an ongoing cellular immune response and patients who do not respond to therapy often present metastatic lesions poorly infiltrated with immune cells (96).

Nevetheless, other crucial components concur to ICI treatment failure, including DC breakdown in proper Ag presentation to effector T cells, persistence of exhausted T cells, and lack of T-cell memory in TME (4, 9). Upon therapy, this dysfunctional state can be reversed or not. In this light, IFN-I signaling has been identified as an important factor for both response and resistance to ICI therapy (25). In fact, IFN-I may have immuno-suppressive or stimulatory activities depending on the magnitude, the timing and the duration of the activation of the signaling. Therefore, the activation of the IFN system may be crucial for the initial response to anti-PD-1, potentially favored in females who have an enhanced aptitude to activate this signal. It may, however, be detrimental if prolonged, promoting escape and resistance in patients who first responded (26, 27). In this light, the implementation of

ICI therapies should take into account also the timing of the activation of the sex-specific components of the antitumor response.

Recently, a meta-analysis evaluating sex-related differences in the response to ICIs was carried out (97). The study assessed the progression free survival (PFS) and the OS in selected 36 phase II/III clinical trials on patients with solid tumors, including melanoma, NSCLC, RCC, head and neck, and urotheial carcinoma, and treated in the first or second line with ipilimumab and/or anti-PD-1 antibodies. Overall, 3,274 patients, of which 2,007 males (61.3%) and 1,267 females (38.7%), were analyzed. The results showed a better OS associated with ipilimumab in males compared with females. Not statistically significant results were observed with anti-PD-1 neither for OS nor for PFS. It should be emphasized that this analysis presents some weaknesses and limitations depending on the heterogeneity of the trials, the different cancer types considered, the variability of treatment regimens including patients treated or not with previous therapies, and the absence of information about hormonal status and on PD-L1 expression according to sex.

CONCLUSION

Research in cancer therapeutics has largely focused on two distinct approaches. One is based on the characterization of mutations causing tumorigenesis pivotal for the use of drugs targeting altered proteins in cancer cells; the other treats cancer "indirectly," exploiting the activation of the immune system to generate an antitumor immunity. While the first approach elicits impressive, but often not durable, tumor responses, the second accomplishes durable clinical responses but only in a subset of patients, a fraction of which experiences relapse after initial encouraging responses. Mechanisms of primary or acquired resistance are the major obstacles in both cases. Sex-associated factors correlate with cancer incidence, outcome, and response to therapy, underlining that sex differences are critical in tumorhost interaction. In this framework, sex disparity in immunity have been recently "re-discovered" and IFN-I signal could play a pivotal role in this scenario. Thus, the sexual dimorphism of the immune signals, including the IFN-I ones, may be a new attractive perspective for optimizing immunotherapy. Moreover, this critical challenge could represent a future opportunity to better integrate immunotherapies with other conventional as well as targeted therapies.

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All authors contributed to the writing and editing of the manuscript.

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REFERENCES

- Dyck L, Mills KHG. Immune checkpoints and their inhibition in cancer and infectious diseases. *Eur J Immunol* (2017) 47(5):765–79. doi:10.1002/ eji.201646875
- Ugurel S, Rohmel J, Ascierto PA, Flaherty KT, Grob JJ, Hauschild A, et al. Survival of patients with advanced metastatic melanoma: the impact of novel therapies. *Eur J Cancer* (2016) 53:125–34. doi:10.1016/j.ejca.2015. 09.013
- Fusi A, Dalgleish A. The importance for immunoregulation for long-term cancer control. *Future Oncol* (2017) 13:1619–32. doi:10.2217/fon-2017-0085
- Zarour HM. Reversing T-cell dysfunction and exhaustion in cancer. Clin Cancer Res (2016) 22(8):1856–64. doi:10.1158/1078-0432.CCR-15-1849
- Wang Q, Wu X. Primary and acquired resistance to PD-1/PD-L1 blockade in cancer treatment. *Int Immunopharmacol* (2017) 46:210–9. doi:10.1016/ j.intimp.2017.03.015
- Kyi C, Postow MA. Immune checkpoint inhibitor combinations in solid tumors: opportunities and challenges. *Immunotherapy* (2016) 8(7):821–37. doi:10.2217/imt-2016-0002
- Klein SL, Flanagan KL. Sex differences in immune responses. Nat Rev Immunol (2016) 16(10):626–38. doi:10.1038/nri.2016.90
- Hegde PS, Karanikas V, Evers S. The where, the when, and the how of immune monitoring for cancer immunotherapies in the era of checkpoint inhibition. *Clin Cancer Res* (2016) 22(8):1865–74. doi:10.1158/1078-0432.CCR-15-1507
- Vasaturo A, Verdoes M, de Vries J, Torensma R, Figdor CG. Restoring immunosurveillance by dendritic cell vaccines and manipulation of the tumor microenvironment. *Immunobiology* (2015) 220(2):243–8. doi:10.1016/ j.imbio.2014.11.007
- Brown SD, Warren RL, Gibb EA, Martin SD, Spinelli JJ, Nelson BH, et al. Neoantigens predicted by tumor genome meta-analysis correlate with increased patient survival. *Genome Res* (2014) 24(5):743–50. doi:10.1101/gr.165985.113
- Veglia F, Gabrilovich DI. Dendritic cells in cancer: the role revisited. Curr Opin Immunol (2017) 45:43–51. doi:10.1016/j.coi.2017.01.002
- Broz ML, Krummel MF. The emerging understanding of myeloid cells as partners and targets in tumor rejection. *Cancer Immunol Res* (2015) 3(4):313–9. doi:10.1158/2326-6066.CIR-15-0041
- Collin M, Bigley V. Human dendritic cell subsets: an update. *Immunology* (2018). doi:10.1111/imm.12888
- Salmon H, Idoyaga J, Rahman A, Leboeuf M, Remark R, Jordan S, et al. Expansion and activation of CD103(+) dendritic cell progenitors at the tumor site enhances tumor responses to therapeutic PD-L1 and BRAF inhibition. *Immunity* (2016) 44(4):924–38. doi:10.1016/j.immuni.2016.03.012
- Roberts EW, Broz ML, Binnewies M, Headley MB, Nelson AE, Wolf DM, et al. Critical role for CD103(+)/CD141(+) dendritic cells bearing CCR7 for tumor antigen trafficking and priming of T cell immunity in melanoma. *Cancer Cell* (2016) 30(2):324–36. doi:10.1016/j.ccell.2016.06.003
- Le Mercier I, Poujol D, Sanlaville A, Sisirak V, Gobert M, Durand I, et al. Tumor promotion by intratumoral plasmacytoid dendritic cells is reversed by TLR7 ligand treatment. *Cancer Res* (2013) 73(15):4629–40. doi:10.1158/0008-5472.CAN-12-3058
- Tel J, Aarntzen EH, Baba T, Schreibelt G, Schulte BM, Benitez-Ribas D, et al. Natural human plasmacytoid dendritic cells induce antigen-specific T-cell responses in melanoma patients. *Cancer Res* (2013) 73(3):1063–75. doi:10.1158/0008-5472.CAN-12-2583
- Swiecki M, Colonna M. The multifaceted biology of plasmacytoid dendritic cells. Nat Rev Immunol (2015) 15(8):471–85. doi:10.1038/nri3865
- Ivashkiv LB, Donlin LT. Regulation of type I interferon responses. Nat Rev Immunol (2014) 14(1):36–49. doi:10.1038/nri3581
- Parker BS, Rautela J, Hertzog PJ. Antitumour actions of interferons: implications for cancer therapy. Nat Rev Cancer (2016) 16(3):131–44. doi:10.1038/ nrc.2016.14
- Burnette BC, Liang H, Lee Y, Chlewicki L, Khodarev NN, Weichselbaum RR, et al. The efficacy of radiotherapy relies upon induction of type i interferondependent innate and adaptive immunity. *Cancer Res* (2011) 71(7):2488–96. doi:10.1158/0008-5472.CAN-10-2820
- 22. Fuertes MB, Kacha AK, Kline J, Woo SR, Kranz DM, Murphy KM, et al. Host type I IFN signals are required for antitumor CD8+ T cell responses through

CD8{alpha}+ dendritic cells. J Exp Med (2011) 208(10):2005–16. doi:10.1084/ jem.20101159

- Schiavoni G, Mattei F, Gabriele L. Type I interferons as stimulators of DC-mediated cross-priming: impact on anti-tumor response. *Front Immunol* (2013) 4:483. doi:10.3389/fimmu.2013.00483
- 24. Corrales L, Matson V, Flood B, Spranger S, Gajewski TF. Innate immune signaling and regulation in cancer immunotherapy. *Cell Res* (2017) 27(1):96–108. doi:10.1038/cr.2016.149
- Minn AJ, Wherry EJ. Combination cancer therapies with immune checkpoint blockade: convergence on interferon signaling. *Cell* (2016) 165(2):272–5. doi:10.1016/j.cell.2016.03.031
- Zaretsky JM, Garcia-Diaz A, Shin DS, Escuin-Ordinas H, Hugo W, Hu-Lieskovan S, et al. Mutations associated with acquired resistance to PD-1 blockade in melanoma. N Engl J Med (2016) 375(9):819–29. doi:10.1056/ NEJMoa1604958
- Benci JL, Xu B, Qiu Y, Wu TJ, Dada H, Twyman-Saint Victor C, et al. Tumor interferon signaling regulates a multigenic resistance program to immune checkpoint blockade. *Cell* (2016) 167(6):1540–54.e12. doi:10.1016/ j.cell.2016.11.022
- 28. Teijaro JR. Pleiotropic roles of type 1 interferons in antiviral immune responses. *Adv Immunol* (2016) 132:135–58. doi:10.1016/bs.ai.2016.08.001
- Ilander M, Kreutzman A, Rohon P, Melo T, Faber E, Porkka K, et al. Enlarged memory T-cell pool and enhanced Th1-type responses in chronic myeloid leukemia patients who have successfully discontinued IFN-alpha monotherapy. *PLoS One* (2014) 9(1):e87794. doi:10.1371/journal.pone.0087794
- Katlinski KV, Gui J, Katlinskaya YV, Ortiz A, Chakraborty R, Bhattacharya S, et al. Inactivation of interferon receptor promotes the establishment of immune privileged tumor microenvironment. *Cancer Cell* (2017) 31(2):194–207. doi:10.1016/j.ccell.2017.01.004
- Snell LM, McGaha TL, Brooks DG. Type I interferon in chronic virus infection and cancer. Trends Immunol (2017) 38(8):542–57. doi:10.1016/j.it.2017.05.005
- 32. Terawaki S, Chikuma S, Shibayama S, Hayashi T, Yoshida T, Okazaki T, et al. IFN-alpha directly promotes programmed cell death-1 transcription and limits the duration of T cell-mediated immunity. *J Immunol* (2011) 186(5):2772–9. doi:10.4049/jimmunol.1003208
- 33. Schwartz JA, Clayton KL, Mujib S, Zhang H, Rahman AK, Liu J, et al. Tim-3 is a marker of plasmacytoid dendritic cell dysfunction during HIV infection and is associated with the recruitment of IRF7 and p85 into lysosomes and with the submembrane displacement of TLR9. *J Immunol* (2017) 198(8):3181–94. doi:10.4049/jimmunol.1601298
- Tsai HF, Hsu PN. Cancer immunotherapy by targeting immune checkpoints: mechanism of T cell dysfunction in cancer immunity and new therapeutic targets. J Biomed Sci (2017) 24(1):35. doi:10.1186/s12929-017-0341-0
- Yu Y, Huang R, Zong X, He X, Mo W. INFalpha-2b inhibitory effects on CD4(+)CD25(+)FOXP3(+) regulatory T cells in the tumor microenvironment of C57BL/6 J mice with melanoma xenografts. *BMC Cancer* (2016) 16:397. doi:10.1186/s12885-016-2473-0
- Bacher N, Raker V, Hofmann C, Graulich E, Schwenk M, Baumgrass R, et al. Interferon-alpha suppresses cAMP to disarm human regulatory T cells. *Cancer Res* (2013) 73(18):5647–56. doi:10.1158/0008-5472.CAN-12-3788
- Fend L, Yamazaki T, Remy C, Fahrner C, Gantzer M, Nourtier V, et al. Immune checkpoint blockade, immunogenic chemotherapy or IFN-alpha blockade boost the local and abscopal effects of oncolytic virotherapy. *Cancer Res* (2017) 77(15):4146–57. doi:10.1158/0008-5472.CAN-16-2165
- Munn DH, Sharma MD, Johnson TS, Rodriguez P. IDO, PTEN-expressing Tregs and control of antigen-presentation in the murine tumor microenvironment. *Cancer Immunol Immunother* (2017) 66(8):1049–58. doi:10.1007/ s00262-017-2010-2
- Lemos H, Mohamed E, Huang L, Ou R, Pacholczyk G, Arbab AS, et al. STING promotes the growth of tumors characterized by low antigenicity via IDO activation. *Cancer Res* (2016) 76(8):2076–81. doi:10.1158/0008-5472. CAN-15-1456
- Chevolet I, Speeckaert R, Haspeslagh M, Neyns B, Kruse V, Schreuer M, et al. Peritumoral indoleamine 2,3-dioxygenase expression in melanoma: an early marker of resistance to immune control? *Br J Dermatol* (2014) 171(5):987–95. doi:10.1111/bjd.13100
- 41. Rautela J, Huntington ND. IL-15 signaling in NK cell cancer immunotherapy. *Curr Opin Immunol* (2017) 44:1–6. doi:10.1016/j.coi.2016.10.004

- Paolini R, Bernardini G, Molfetta R, Santoni A. NK cells and interferons. *Cytokine Growth Factor Rev* (2015) 26(2):113–20. doi:10.1016/ j.cytogfr.2014.11.003
- O'Donnell JS, Long GV, Scolyer RA, Teng MW, Smyth MJ. Resistance to PD1/ PDL1 checkpoint inhibition. *Cancer Treat Rev* (2017) 52:71–81. doi:10.1016/ j.ctrv.2016.11.007
- Schreiber G. The molecular basis for differential type I interferon signaling. J Biol Chem (2017) 292(18):7285–94. doi:10.1074/jbc.R116.774562
- Chen K, Liu J, Cao X. Regulation of type I interferon signaling in immunity and inflammation: a comprehensive review. J Autoimmun (2017) 83:1–11. doi:10.1016/j.jaut.2017.03.008
- Pauken KE, Sammons MA, Odorizzi PM, Manne S, Godec J, Khan O, et al. Epigenetic stability of exhausted T cells limits durability of reinvigoration by PD-1 blockade. *Science* (2016) 354(6316):1160–5. doi:10.1126/science. aaf2807
- Grenda A, Krawczyk P. New dancing couple: PD-L1 and microRNA. Scand J Immunol (2017) 86:130–4. doi:10.1111/sji.12577
- Linardou H, Gogas H. Toxicity management of immunotherapy for patients with metastatic melanoma. *Ann Transl Med* (2016) 4(14):272. doi:10.21037/ atm.2016.07.10
- Botticelli A, Zizzari I, Mazzuca F, Ascierto PA, Putignani L, Marchetti L, et al. Cross-talk between microbiota and immune fitness to steer and control response to anti PD-1/PDL-1 treatment. *Oncotarget* (2017) 8(5):8890–9. doi:10.18632/oncotarget.12985
- Kotredes KP, Thomas B, Gamero AM. The protective role of type I interferons in the gastrointestinal tract. *Front Immunol* (2017) 8:410. doi:10.3389/ fimmu.2017.00410
- Mirandola L, Wade R, Verma R, Pena C, Hosiriluck N, Figueroa JA, et al. Sexdriven differences in immunological responses: challenges and opportunities for the immunotherapies of the third millennium. *Int Rev Immunol* (2015) 34(2):134–42. doi:10.3109/08830185.2015.1018417
- Laffont S, Blanquart E, Savignac M, Cenac C, Laverny G, Metzger D, et al. Androgen signaling negatively controls group 2 innate lymphoid cells. J Exp Med (2017) 214(6):1581–92. doi:10.1084/jem.20161807
- Pennell LM, Galligan CL, Fish EN. Sex affects immunity. J Autoimmun (2012) 38(2–3):J282–91. doi:10.1016/j.jaut.2011.11.013
- Zilio S, Serafini P. Neutrophils and granulocytic MDSC: the Janus god of cancer immunotherapy. Vaccines (Basel) (2016) 4(3):E31. doi:10.3390/ vaccines4030031
- Lu C, Redd PS, Lee JR, Savage N, Liu K. The expression profiles and regulation of PD-L1 in tumor-induced myeloid-derived suppressor cells. *Oncoimmunology* (2016) 5(12):e1247135. doi:10.1080/2162402X.2016.1247135
- Kovats S. Estrogen receptors regulate innate immune cells and signaling pathways. *Cell Immunol* (2015) 294(2):63–9. doi:10.1016/j.cellimm.2015.01.018
- 57. Seillet C, Laffont S, Tremollieres F, Rouquie N, Ribot C, Arnal JF, et al. The TLR-mediated response of plasmacytoid dendritic cells is positively regulated by estradiol in vivo through cell-intrinsic estrogen receptor alpha signaling. *Blood* (2012) 119(2):454–64. doi:10.1182/blood-2011-08-371831
- Laffont S, Rouquie N, Azar P, Seillet C, Plumas J, Aspord C, et al. X-Chromosome complement and estrogen receptor signaling independently contribute to the enhanced TLR7-mediated IFN-alpha production of plasmacytoid dendritic cells from women. *J Immunol* (2014) 193(11):5444–52. doi:10.4049/jimmunol.1303400
- Griesbeck M, Ziegler S, Laffont S, Smith N, Chauveau L, Tomezsko P, et al. Sex differences in plasmacytoid dendritic cell levels of IRF5 drive higher IFNalpha production in women. *J Immunol* (2015) 195(11):5327–36. doi:10.4049/ jimmunol.1501684
- Laffont S, Seillet C, Guery JC. Estrogen receptor-dependent regulation of dendritic cell development and function. *Front Immunol* (2017) 8:108. doi:10.3389/fimmu.2017.00108
- Bajana S, Roach K, Turner S, Paul J, Kovats S. IRF4 promotes cutaneous dendritic cell migration to lymph nodes during homeostasis and inflammation. *J Immunol* (2012) 189(7):3368–77. doi:10.4049/jimmunol.1102613
- Heng TS, Painter MW; Immunological Genome Project Consortium. The Immunological Genome Project: networks of gene expression in immune cells. *Nat Immunol* (2008) 9(10):1091–4. doi:10.1038/ni1008-1091
- Anipindi VC, Bagri P, Roth K, Dizzell SE, Nguyen PV, Shaler CR, et al. Estradiol enhances CD4+ T-cell anti-viral immunity by priming vaginal DCs

to induce Th17 responses via an IL-1-dependent pathway. *PLoS Pathog* (2016) 12(5):e1005589. doi:10.1371/journal.ppat.1005589

- 64. Thompson MG, Peiffer DS, Larson M, Navarro F, Watkins SK. FOXO3, estrogen receptor alpha, and androgen receptor impact tumor growth rate and infiltration of dendritic cell subsets differentially between male and female mice. *Cancer Immunol Immunother* (2017) 66(5):615–25. doi:10.1007/ s00262-017-1972-4
- Panchanathan R, Shen H, Zhang X, Ho SM, Choubey D. Mutually positive regulatory feedback loop between interferons and estrogen receptor-alpha in mice: implications for sex bias in autoimmunity. *PLoS One* (2010) 5(5):e10868. doi:10.1371/journal.pone.0010868
- Mackern-Oberti JP, Jara EL, Riedel CA, Kalergis AM. Hormonal modulation of dendritic cells differentiation, maturation and function: implications for the initiation and progress of systemic autoimmunity. *Arch Immunol Ther Exp* (2017) 65(2):123–36. doi:10.1007/s00005-016-0418-6
- 67. Fish EN. The X-files in immunity: sex-based differences predispose immune responses. *Nat Rev Immunol* (2008) 8(9):737–44. doi:10.1038/nri2394
- Arsenovic-Ranin N, Kosec D, Pilipovic I, Nacka-Aleksic M, Bufan B, Stojic-Vukanic Z, et al. Sex and age as determinants of rat T-cell phenotypic characteristics: influence of peripubertal gonadectomy. *Mol Cell Biochem* (2017) 431(1–2):169–85. doi:10.1007/s11010-017-2989-x
- 69. Zhang MA, Rego D, Moshkova M, Kebir H, Chruscinski A, Nguyen H, et al. Peroxisome proliferator-activated receptor (PPAR)alpha and -gamma regulate IFNgamma and IL-17A production by human T cells in a sex-specific way. *Proc Natl Acad Sci U S A* (2012) 109(24):9505–10. doi:10.1073/ pnas.1118458109
- Lin PY, Sun L, Thibodeaux SR, Ludwig SM, Vadlamudi RK, Hurez VJ, et al. B7-H1-dependent sex-related differences in tumor immunity and immunotherapy responses. *J Immunol* (2010) 185(5):2747–53. doi:10.4049/ jimmunol.1000496
- Markle JG, Fish EN. SeXX matters in immunity. *Trends Immunol* (2014) 35(3):97–104. doi:10.1016/j.it.2013.10.006
- 72. Xu A, Wang R, Freywald A, Stewart K, Tikoo S, Xu J, et al. CD40 agonist converting CTL exhaustion via the activation of the mTORC1 pathway enhances PD-1 antagonist action in rescuing exhausted CTLs in chronic infection. *Biochem Biophys Res Commun* (2017) 484(3):662–7. doi:10.1016/ j.bbrc.2017.01.172
- Hewagama A, Patel D, Yarlagadda S, Strickland FM, Richardson BC. Stronger inflammatory/cytotoxic T-cell response in women identified by microarray analysis. *Genes Immun* (2009) 10(5):509–16. doi:10.1038/gene.2009.12
- Dai R, Ahmed SA. Sexual dimorphism of miRNA expression: a new perspective in understanding the sex bias of autoimmune diseases. *Ther Clin Risk Manag* (2014) 10:151–63. doi:10.2147/TCRM.S33517
- Hewagama A, Gorelik G, Patel D, Liyanarachchi P, McCune WJ, Somers E, et al. Overexpression of X-linked genes in T cells from women with lupus. *J Autoimmun* (2013) 41:60–71. doi:10.1016/j.jaut.2012.12.006
- Smolle MA, Calin HN, Pichler M, Calin GA. Noncoding RNAs and immune checkpoints-clinical implications as cancer therapeutics. *FEBS J* (2017) 284(13):1952–66. doi:10.1111/febs.14030
- Loo K, Tsai KK, Mahuron K, Liu J, Pauli ML, Sandoval PM, et al. Partially exhausted tumor-infiltrating lymphocytes predict response to combination immunotherapy. *JCI Insight* (2017) 2(14):93433. doi:10.1172/jci. insight.93433
- Gut microbes may up PD-1 inhibitor response. *Cancer Discov* (2017) 7(5):448. doi:10.1158/2159-8290.CD-NB2017-039
- Sivan A, Corrales L, Hubert N, Williams JB, Aquino-Michaels K, Earley ZM, et al. Commensal *Bifidobacterium* promotes antitumor immunity and facilitates anti-PD-L1 efficacy. *Science* (2015) 350(6264):1084–9. doi:10.1126/ science.aac4255
- Markle JG, Frank DN, Adeli K, von Bergen M, Danska JS. Microbiome manipulation modifies sex-specific risk for autoimmunity. *Gut Microbes* (2014) 5(4):485–93. doi:10.4161/gmic.29795
- Melero I, Gaudernack G, Gerritsen W, Huber C, Parmiani G, Scholl S, et al. Therapeutic vaccines for cancer: an overview of clinical trials. *Nat Rev Clin Oncol* (2014) 11(9):509–24. doi:10.1038/nrclinonc.2014.111
- van der Burg SH, Arens R, Ossendorp F, van Hall T, Melief CJ. Vaccines for established cancer: overcoming the challenges posed by immune evasion. *Nat Rev Cancer* (2016) 16(4):219–33. doi:10.1038/nrc.2016.16

- Steinman RM. Decisions about dendritic cells: past, present, and future. Annu Rev Immunol (2012) 30:1–22. doi:10.1146/annurev-immunol-100311-102839
- Bracci L, Capone I, Moschella F, Proietti E, Belardelli F. Exploiting dendritic cells in the development of cancer vaccines. *Expert Rev Vaccines* (2013) 12(10):1195–210. doi:10.1586/14760584.2013.836905
- Draube A, Klein-Gonzalez N, Mattheus S, Brillant C, Hellmich M, Engert A, et al. Dendritic cell based tumor vaccination in prostate and renal cell cancer: a systematic review and meta-analysis. *PLoS One* (2011) 6(4):e18801. doi:10.1371/journal.pone.0018801
- Sheikh NA, Petrylak D, Kantoff PW, Dela Rosa C, Stewart FP, Kuan LY, et al. Sipuleucel-T immune parameters correlate with survival: an analysis of the randomized phase 3 clinical trials in men with castration-resistant prostate cancer. *Cancer Immunol Immunother* (2013) 62(1):137–47. doi:10.1007/ s00262-012-1317-2
- GuhaThakurta D, Sheikh NA, Fan LQ, Kandadi H, Meagher TC, Hall SJ, et al. Humoral immune response against nontargeted tumor antigens after treatment with Sipuleucel-T and its association with improved clinical outcome. *Clin Cancer Res* (2015) 21(16):3619–30. doi:10.1158/1078-0432.CCR-14-2334
- Anguille S, Smits EL, Lion E, van Tendeloo VF, Berneman ZN. Clinical use of dendritic cells for cancer therapy. *Lancet Oncol* (2014) 15(7):e257–67. doi:10.1016/S1470-2045(13)70585-0
- Yarchoan M, Johnson BA III, Lutz ER, Laheru DA, Jaffee EM. Targeting neoantigens to augment antitumour immunity. *Nat Rev Cancer* (2017) 17(4):209–22. doi:10.1038/nrc.2016.154
- Littman DR. Releasing the brakes on cancer immunotherapy. Cell (2015) 162(6):1186–90. doi:10.1016/j.cell.2015.08.038
- Wolchok JD, Hodi FS, Weber JS, Allison JP, Urba WJ, Robert C, et al. Development of ipilimumab: a novel immunotherapeutic approach for the treatment of advanced melanoma. *Ann N Y Acad Sci* (2013) 1291:1–13. doi:10.1111/nyas.12180

- Robert C, Schachter J, Long GV, Arance A, Grob JJ, Mortier L, et al. Pembrolizumab versus ipilimumab in advanced melanoma. N Engl J Med (2015) 372(26):2521–32. doi:10.1056/NEJMoa1503093
- Garon EB, Rizvi NA, Hui R, Leighl N, Balmanoukian AS, Eder JP, et al. Pembrolizumab for the treatment of non-small-cell lung cancer. N Engl J Med (2015) 372(21):2018–28. doi:10.1056/NEJMoa1501824
- Motzer RJ, Rini BI, McDermott DF, Redman BG, Kuzel TM, Harrison MR, et al. Nivolumab for metastatic renal cell carcinoma: results of a randomized phase II trial. J Clin Oncol (2015) 33(13):1430–7. doi:10.1200/JCO.2014.59.0703
- Erdag G, Schaefer JT, Smolkin ME, Deacon DH, Shea SM, Dengel LT, et al. Immunotype and immunohistologic characteristics of tumor-infiltrating immune cells are associated with clinical outcome in metastatic melanoma. *Cancer Res* (2012) 72(5):1070–80. doi:10.1158/0008-5472.CAN-11-3218
- Galon J, Fox BA, Bifulco CB, Masucci G, Rau T, Botti G, et al. Immunoscore and immunoprofiling in cancer: an update from the melanoma and immunotherapy bridge 2015. J Transl Med (2016) 14:273. doi:10.1186/s12967-016-1029-z
- Botticelli A, Onesti CE, Zizzari I, Cerbelli B, Sciattella P, Occhipinti M, et al. The sexist behaviour of immune checkpoint inhibitors in cancer therapy? Oncotarget (2017) 8(59):99336–46. doi:10.18632/oncotarget.22242

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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