Action in pairs Two tandem genes in the fish pathogen *Yersinia ruckeri* are virulence factors

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Yersinia ruckeri is a gram-negative rod-shaped bacterium belonging to the family Enterobacteriaceae. It is in the same genus as *Yersinia pestis*, the causative agent of the bubonic plaque and a category A select agent. The taxonomic assignment of *Y. ruckeri*, however, has raised controversy since it appears to have diverged from the rest of the Yersinia genus in phylogenetic analysis.¹

Y. ruckeri is the causative agent of enteric redmouth (ERM) disease in salmonids, known as yersiniosis, and can cause significant economic losses, particularly in the rainbow trout farming industry.² Clinical signs include hemorrhaging around the mouth, intestines and other organs. *Y. ruckeri* is naturally associated with the aquatic environment and is thought to initially infect via adhesion to the gill surface.

While most research effort has been focused on characterizing human pathogens, studies of their closely related non-pathogenic-to-human relatives have been largely overlooked although it has been recognized that many human pathogens have actually emerged very recently from protean environmental, commensal, or zoonotic populations, and there are similarities in the genomic, biochemical levels among these strains.^{3,4}

This is also the case for research on *Y. ruckeri* despite its importance in aquaculture. Most research attention in species of the Yersinia genus is focused on the three well-known human pathogens, *Y. pestis, Yersinia pseudotuberculosis*, and *Yersinia enterocolitica*. For example, the virulence mechanisms of *Y. pestis* have been well studied⁵ because it has caused approximately 200 million human deaths historically, with at least 2000 cases of plague reported annually by the World Health Organization (WHO). However, there are only a few pathogenic mechanisms of *Y. ruckeri* that have been described so far. Some of these mechanisms have been proven to be involved in virulence, such as the iron uptake mechanism via the siderophore natural product ruckerbatin⁶ and the YhlA hemolysin⁷ and a new type of twocomponent operon that contains an amino acid permease motif and an L-cysteine desulfidase motif.⁸ The operon was confirmed to be involved in the regulation of cysteine uptake, and knockout of this operon abolishes virulence of *Y. ruckeri* in fish. *Y. ruckeri* was also found to produce an antibiotic natural product holomycin,⁹ a founding member of a unique family of dithiolopyrrolone natural products.¹⁰ Interestingly, holomycin production was also found to have a connection with the regulation of cysteine uptake.¹¹ It remains to be shown whether holomycin production is associated with pathogenicity of *Y. ruckeri*.

Featured in the present issue of Virulence, Navais et al.¹² reported a potential new virulence factor that includes one pair of tandem genes *yrpA* and *yrpB*, encoding putative peptidases, in the chromosome of Y. ruckeri. Their studies demonstrated that these two genes may be transcribed together. The expression of the genes can be induced when the bacterium was cultured with peptone and under microaerobic culturing conditions. More importantly, inactivation of *yrpA*, resulting in the mutant strain $\Delta yrpA$, greatly reduced the infection process for this mutant strain. In silico analysis indicated that similar genes with the same genetic arrangement also exist in the genomes of other human-related pathogenic yersiniae, suggesting a possible new virulence mechanism in this genus. The study also highlights the potential for Y. ruckeri to be used as a surrogate model for Y. pestis because of such similarities at the genomic level between these two species.¹³ Since Y. ruckeri does not require the biosafety category 3 facilities needed for some of the resistant Y. pestis strains it is easier to study and to work with.

Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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References

- Kotetishvili M, Kreger A, Wauters G, Morris JG Jr., Sulakvelidze A, Stine OC. Multilocus sequence typing for studying genetic relationships among Yersinia species. J Clin Microbiol 2005; 43:2674-84; PMID:15956383; http://dx.doi.org/10.1128/ JCM.43.6.2674-2684.2005
- Harun NO, Wang T, Secombes CJ. Gene expression profiling in naïve and vaccinated rainbow trout after *Yersinia ruckeri* infection: insights into the mechanisms of protection seen in vaccinated fish. Vaccine 2011; 29:4388-99; PMID:21504776; http://dx.doi. org/10.1016/j.vaccine.2011.04.003
- van Baarlen P, van Belkum A, Summerbell RC, Crous PW, Thomma BP. Molecular mechanisms of pathogenicity: how do pathogenic microorganisms develop cross-kingdom host jumps? FEMS Microbiol Rev 2007; 31:239-77; PMID:17326816; http://dx.doi. org/10.1111/j.1574-6976.2007.00065.x
- Achtman M, Zurth K, Morelli G, Torrea G, Guiyoule A, Carniel E. Yersinia pestis, the cause of plague, is a recently emerged clone of Yersinia pseudotuberculosis. Proc Natl Acad Sci U S A 1999; 96:14043-8; PMID:10570195; http://dx.doi.org/10.1073/ pnas.96.24.14043
- Morelli G, Song Y, Mazzoni CJ, Eppinger M, Roumagnac P, Wagner DM, Feldkamp M, Kusecek B, Vogler AJ, Li Y, et al. Yersinia pestis genome sequencing identifies patterns of global phylogenetic diversity. Nat Genet 2010; 42:1140-3; PMID:21037571; http://dx.doi.org/10.1038/ng.705

- Fernández L, Márquez I, Guijarro JA. Identification of specific *in vivo*-induced (*ivi*) genes in *Yersinia ruckeri* and analysis of ruckerbactin, a catecholate siderophore iron acquisition system. Appl Environ Microbiol 2004; 70:5199-207; PMID:15345400; http://dx.doi. org/10.1128/AEM.70.9.5199-5207.2004
- Fernández L, Prieto M, Guijarro JA. The ironand temperature-regulated haemolysin YhlA is a virulence factor of *Yersinia ruckeri*. Microbiology 2007; 153:483-9; PMID:17259619; http://dx.doi. org/10.1099/mic.0.29284-0
- Méndez J, Reimundo P, Pérez-Pascual D, Navais R, Gómez E, Guijarro JA. A novel cdsAB operon is involved in the uptake of L-cysteine and participates in the pathogenesis of *Yersinia ruckeri*. J Bacteriol 2011; 193:944-51; PMID:21169490; http://dx.doi. org/10.1128/JB.01058-10
- Qin Z, Baker AT, Raab A, Huang S, Wang T, Yu Y, Jaspars M, Secombes CJ, Deng H. The fish pathogen Yersinia ruckeri produces holomycin and uses an RNA methyltransferase for self-resistance. J Biol Chem 2013; 288:14688-97; PMID:23572522; http://dx.doi.org/10.1074/jbc.M112.448415
- Qin Z, Huang S, Yu Y, Deng H. Dithiolopyrrolone natural products: isolation, synthesis and biosynthesis. Mar Drugs 2013; 11:3970-97; PMID:24141227; http://dx.doi.org/10.3390/md11103970

- Bouras N, Mathieu F, Sabaou N, Lebrihi A. Effect of amino acids containing sulfur on dithiolopyrrolone antibiotic productions by *Saccharothrix algeriensis* NRRL B-24137. J Appl Microbiol 2006; 100:390-7; PMID:16430516; http://dx.doi. org/10.1111/j.1365-2672.2005.02762.x
- Navais R, Méndez J, Pérez-Pascual D, Cascales D, Guijarro JA. The *yrpAB* operon of *Yersinia ruckeri* encoding two putative U32 peptidases is involved in virulence and induced under microaerobic conditions. Virulence 2014; 5:619-24; http://dx.doi. org/10.4161/viru.29363; PMID:24865652
- Chen PE, Cook C, Stewart AC, Nagarajan N, Sommer DD, Pop M, Thomason B, Thomason MPK, Lentz S, Nolan N, et al. Genomic characterization of the Yersinia genus. Genome Biol 2010; 11:R1; PMID:20047673; http://dx.doi.org/10.1186/ gb-2010-11-1-r1.