

Evaluation of accuracy and time taken to make an open tray implant impressions with two techniques

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ABSTRACT

Implant dentistry's emergence and acceptance have provided physicians with a wide range of new options for fixed and removable rehabilitation. The eventual replacement of the lost tooth is the ultimate success, and this can be accomplished using recognized prosthodontic methods. The present study aims to evaluate the time and dimensional accuracy of implant definitive models with an open tray impression with two different techniques. Impression was made in two different techniques and the cast was poured and checked for the time taken and the accuracy of the impression. In IBM SPSS software version 23.0 all the results were statistically analyzed. An Independent *t*-test was performed for the parameters. No statistically significant difference was present ($P > 0.05$) when comparing the accuracy and time taken between the two groups. It is concluded that time taken and the misfit is less for modified open tray impression techniques when compared to the regular open tray impression.

Key words: Accuracy, discrepancy, implants, innovation, open tray impression, time

INTRODUCTION

For making a final prosthesis, a good impression is required, which replicates the intra-oral structures.^[1,2] The accuracy of the master cast is also determined by the accuracy of the impression. Incorrect recording of impressions leads to mechanical and/or biological complications such as implant fracture, implant abutment fractures, screw loosening, screw fracture, and occlusal discrepancies,^[3,4] increased plaque accumulation due to undesirable hard and soft-tissue reactions.^[5-8]

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Obtaining a passive fit prostheses is optimal for the long-term success of any implant treatment. Long-term implant stability will be compromised by an ill-fitting prosthesis.^[9] According to the literature supporting this hypothesis, clinically good fitting prosthesis will create a significant amount of misfit load but no loss of osseointegration.^[10,11] Gaps $<60 \mu\text{m}$ are difficult to measure clinically.^[12,12] Basic impression techniques for implants are implant level impressions (open tray impression, closed tray impression), abutment level, and digital impression.

The aim of this *in vitro* study is to compare two impression techniques for evaluating the time and dimensional accuracy of implant definitive models with open tray impressions. Our team has extensive knowledge and research experience that has translated into high-quality publications.^[1,2,13-37]

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MATERIALS AND METHODS

Model preparation

A mandibular edentulous styrofoam model was taken and three implants placed in between the mental foramen region [Figure 1]. The tentative implant locations should be marked on the cast based on the visible healing abutments. A single wax chimney was created around the implant sites. An acrylic special tray should be made to cover the entire edentulous ridge and also over the walls of the wax chimney [Figures 2 and 3]. Open tray impression copings should be placed on the implants and tightened (torqued) to 20 N-cm.

The copings should be splinted together with 23 gauge SS wire or dental floss and reinforced with pattern resin and an IOPA taken to verify [Figures 4-8]. The tray chimney should be sealed with wax such that the imprints of the implant copings indent the wax. Monophase material should be loaded in the tray and also between the copings it should be injected and the tray should be seated such that the copings and the marks made on the wax over the chimney align with one another. The wax roof over the chimney should be removed and the excess material inside the coping screw channel should be excavated with an explorer. The impression coping should be removed from the implant and tightened onto the lab analog. To achieve the right emergence profile in the restoration, a soft tissue mask and a silicone soft tissue replica are necessary. A low expansion type 3 dental stone has been used to make models for implant restoration.

Whereas in the new impression technique, a modified special tray should be made in which it covers the open tray implant copings and a small rectangular slot is placed on the labial side of the special tray [Figure 3]. Here the copings are not splinted to each other, Monophase impression is injected between the copings and impression is made and impression material which is covering the top portion of the impression copings should be scraped/removed and then the splinting of the impression copings to impression tray with the help of pattern resin. The pattern resin should encircle the copings and it is extended to the labial surface of the tray and is engaged into the rectangular slot. After setting the impression material, the coping screws are removed, a soft tissue mask is placed and a cast is poured.

The time taken for making these 2 impressions are recorded with the help of a stopwatch and the accuracy of the impression is evaluated by doing the jig verification of the control cast with the 2 experimental casts and checking for discrepancy under stereo electronic microscope in 3 areas [Figure 9].

RESULTS

The mean time for making a regular open tray impression time is 8.80 mins and the modified open tray impression



Figure 1: Implants placed in a styrofoam model with implant open tray copings



Figure 2: Conventional open tray made with polytray

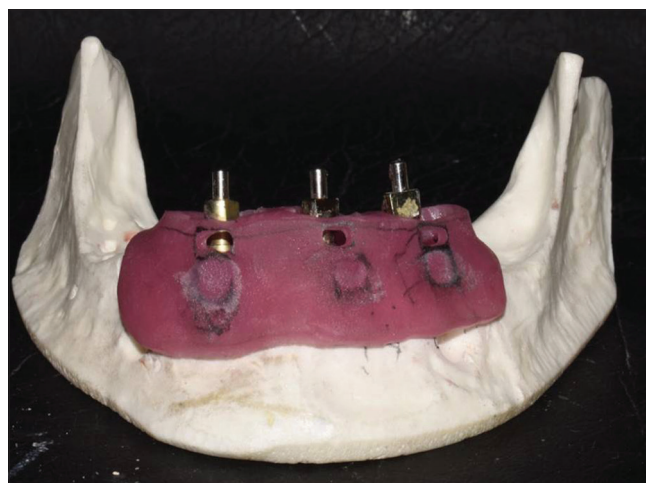


Figure 3: Modified impression tray made with polytray

technique is 6.88 mins and the P -value(>0.05) is found not statistically significant [Tables 1 and 2].

When discrepancy was evaluated on mesial, buccal and distal aspects, the mean discrepancy for regular open tray impression are .0980, .0686, .0986 and for modified open tray impression technique are .0880, .0350 and .0500 respectively and the *P*-value (>0.05) is found not statistically significant [Table 3 and Figure 8].

DISCUSSION

In the present study, the accuracy and time taken to make two open tray impression techniques were evaluated. The time taken for making a modified impression technique is less compared to the regular open tray impression; however, the statistics showed no significant differences between groups.

The results of this study, as measured by the vertical fit discrepancy, in both techniques showed 10 µm difference in medians. This is presumably of very little clinical

consequence, mainly as it has been demonstrated that measuring gaps shorter than 60 µm is challenging clinically. In comparison to the original model, models made from an

Table 1: The time taken for making implant impressions for both the techniques

Conventional	New technique
8:51.85	7:11.67
9:11.54	7:56.13
8:9.49	6:63.72
8:91.21	6:81.41
9:05.67	6:86.84
8:65.81	7:03.72
8:98.56	6:75.84
8:66.79	6:66.71
8:65.26	6:71.44
8:43.19	6:76.57



Figure 4: Splinting of impression copings with floss



Figure 5: Splinting of impression copings to the custom tray with pattern resin

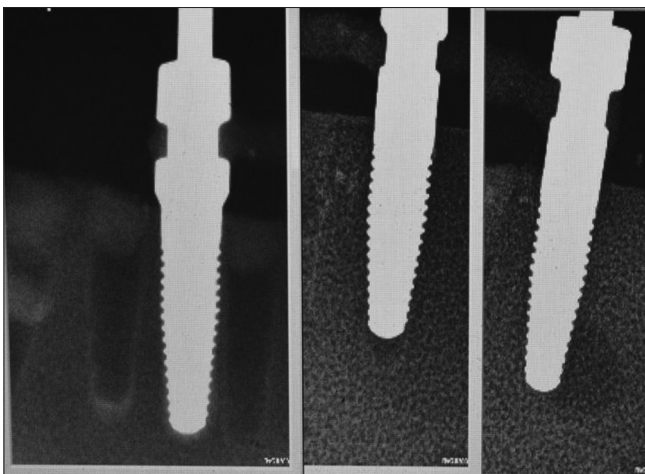


Figure 6: IOPA of the control model. IOPA: Intra Oral Periapical radiographs

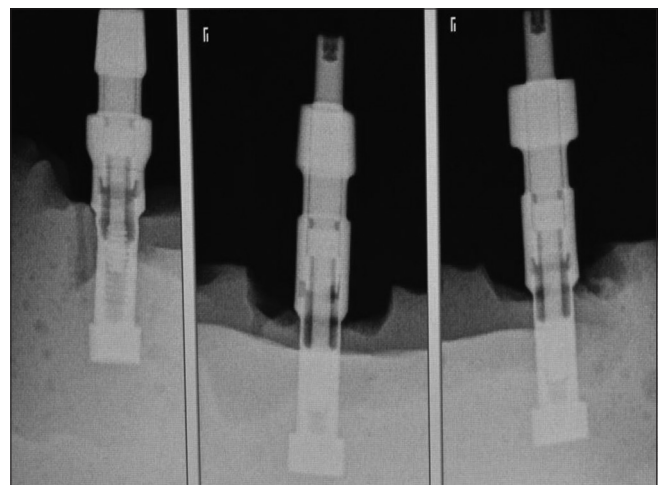


Figure 7: IOPA of the conventional implant open tray impression after pouring the cast. IOPA: Intra Oral Periapical radiographs

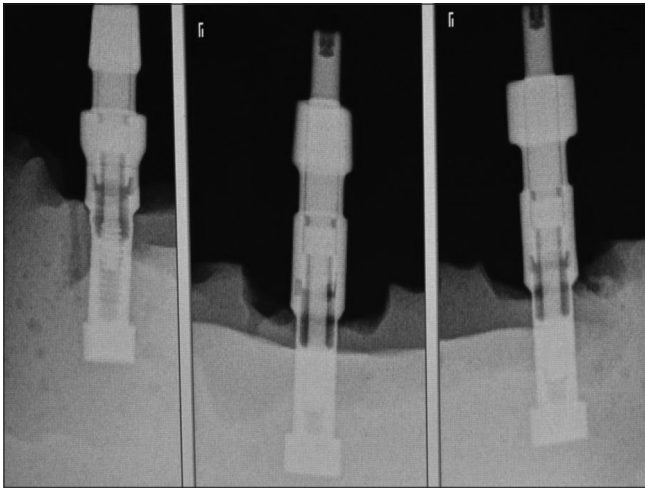


Figure 8: IOPA of the modified implant open tray impression after pouring the cast. IOPA: Intra Oral Periapical radiographs

Table 2: The average time taken for making both the impressions and the $P > 0.05$ is found not to be statistically significant

Type of impression	Mean	SD	P
Regular open tray impression	8.80	13.664	0.803
Modified open tray impression	6.88	17.240	

SD: Standard deviation

Table 3: The marginal discrepancy of two different techniques on three different surfaces and the $P > 0.05$ is found not to be statistically significant

Location	Groups	Mean	SD	P
Mesial	Regular impression	0.0980	0.01687	0.100
	Modified impression	0.0880	0.10983	
Buccal	Regular impression	0.0686	0.01345	0.478
	Modified impression	0.0350	0.00972	
Distal	Regular impression	0.0986	0.02035	0.104
	Modified impression	0.0500	0.01054	

SD: Standard deviation

open tray impression show a variation in distances between analogs in samples. The impression tray withdrawal path/pattern, which puts greater stress on the impression materials, could be the cause of increased distortion.

Despite the lack of a consistent finding for higher accuracy with one impression technique over the other impression technique, splinting or nonsplinting. Many studies indicate that for accurate implant impressions, splinting of copings technique is preferred than the nonsplinting technique. Some authors have mentioned problems such as distortion of materials used for splinting and fracture of the link between splinting material and implant open tray impression coping when using the splinting technique.^[38] Some examined the implant impression accuracy in a variety of lab processes

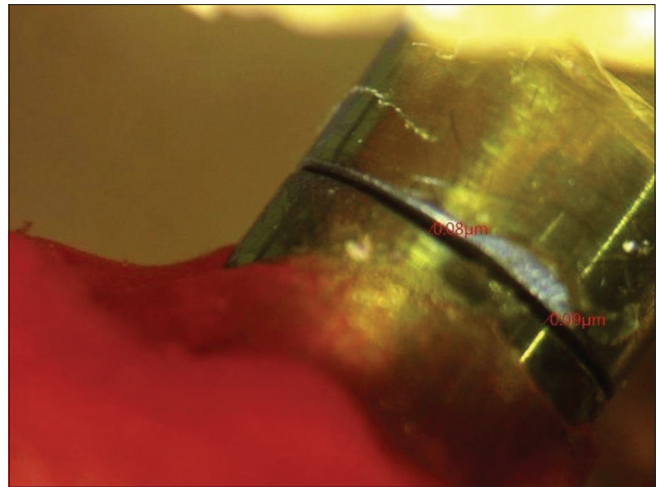


Figure 9: Marginal discrepancy is seen between the implant and the impression coping under the stereo electronic microscope of the cast, which is made of conventional open tray technique

and discovered that the nonsplinting technique was more precise during the impression-making process.^[39,40]

CONCLUSION

The present *in vitro* study concludes that the time taken and accuracy for making an open tray implant impression is faster in the new modified technique when compared to the regular open tray impression.

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Conflicts of interest

There are no conflicts of interest.

REFERENCES

1. Agarwal S, Ashok V, Maiti S. Open- or closed-tray impression technique in implant prosthesis: A dentist's perspective. *J Long Term Eff Med Implants* 2020;30:193-8.
2. Merchant A, Maiti S, Ashok V, Ganapathy DM. Comparative analysis of different impression techniques in relation to single tooth impression. *Bioinformation* 2020;16:1105-10.
3. Burguete RL, Johns RB, King T, Patterson EA. Tightening characteristics for screwed joints in osseointegrated dental implants. *J Prosthet Dent* 1994;71:592-9.
4. Eckert SE, Meraw SJ, Cal E, Ow RK. Analysis of incidence and associated factors with fractured implants: A retrospective study. *Int J Oral Maxillofac Implants* 2000;15:662-7.

5. Lindhe J, Berglundh T, Ericsson I, Liljeborg B, Marinello C. Experimental breakdown of peri-implant and periodontal tissues. A study in the beagle dog. *Clin Oral Implants Res* 1992;3:9-16.
6. Augthun M, Conrads G. Microbial findings of deep peri-implant bone defects. *Int J Oral Maxillofac Implants* 1997;12:106-12.
7. Leonhardt A, Renvert S, Dahlén G. Microbial findings at failing implants. *Clin Oral Implants Res* 1999;10:339-45.
8. Kan JY, Rungcharassaeng K, Bohsali K, Goodacre CJ, Lang BR. Clinical methods for evaluating implant framework fit. *J Prosthet Dent* 1999;81:7-13.
9. Ponnanna AA, Maiti S, Rai N, Jessy P. Three-dimensional-printed Malo Bridge: Digital fixed prosthesis for the partially edentulous maxilla. *Contemp Clin Dent* 2021;12:451-3.
10. Smedberg JI, Nilner K, Rangert B, Svensson SA, Glantz SA. On the influence of superstructure connection on implant preload: A methodological and clinical study. *Clin Oral Implants Res* 1996;7:55-63.
11. Kushali R, Maiti S, Girija SA, Jessy P. Evaluation of microbial leakage at implant abutment interface for different implant systems: An *in vitro* study. *J Long Term Eff Med Implants* 2022;32:87-93.
12. McLean JW, von Fraunhofer JA. The estimation of cement film thickness by an *in vivo* technique. *Br Dent J* 1971;131:107-11.
13. Avinash K, Malaippan S, Dooraiswamy JN. Methods of isolation and characterization of stem cells from different regions of oral cavity using markers: A systematic review. *Int J Stem Cells* 2017;10:12-20.
14. Pratha AA, Thenmozhi MS. A study of occurrence and morphometric analysis on Meningo orbital foramen. *Res J Pharm Technol* 2016;9:880-2.
15. Nair M, Jeevanandan G, Vignesh R. Comparative evaluation of post-operative pain after pulpectomy with k-files, kedo-s files and mtwo files in deciduous molars-a randomized clinical trial. *Braz Dent J* 2018;21(4). Available from: <https://doi.org/10.14295/bds.2018.v21i4.1617>.
16. Kannan R, Thenmozhi MS. Morphometric study of styloid process and its clinical importance on Eagle's syndrome. *Res J Pharm Technol* 2016;9:1137-9.
17. Merchant A, Maiti S, Rajaraman V, Velayudhan A, Ganapathy DM. Comparative analysis of pink and white esthetics of anterior full veneer crown: Indian scenario. *J Adv Pharm Technol Res* 2022;13:5282-7.
18. Viswanath A, Ramamurthy J, Dinesh SP, Srinivas A. Obstructive sleep apnea: Awakening the hidden truth. *Niger J Clin Pract* 2015;18:1-7.
19. Dinesh SP, Arun AV, Sundari KK, Samantha C, Ambika K. An indigenously designed apparatus for measuring orthodontic force. *J Clin Diagn Res* 2013;7:2623-6.
20. Varghese SS, Thomas H, Jayakumar ND, Sankari M, Lakshmanan R. Estimation of salivary tumor necrosis factor-alpha in chronic and aggressive periodontitis patients. *Contemp Clin Dent* 2015;6:S152-6.
21. Priyanka S, Kaarthikeyan G, Nadathur JD, Mohanraj A, Kavarthapu A. Detection of cytomegalovirus, Epstein-Barr virus, and Torque Teno virus in subgingival and atheromatous plaques of cardiac patients with chronic periodontitis. *J Indian Soc Periodontol* 2017;21:456-60.
22. Panda S, Jayakumar ND, Sankari M, Varghese SS, Kumar DS. Platelet rich fibrin and xenograft in treatment of intrabony defect. *Contemp Clin Dent* 2014;5:550-4.
23. Aparna J, Maiti S, Jessy P. Polyether ether ketone – As an alternative biomaterial for Metal Richmond crown-3-dimensional finite element analysis. *J Conserv Dent* 2021;24:553-7.
24. Kasabwala H, Maiti S, Ashok V, Sashank K. Data on dental bite materials with stability and displacement under load. *Bioinformation* 2020;16:1145-51.
25. Agarwal S, Maiti S, Ashok V. Correlation of soft tissue biotype with pink aesthetic score in single full veneer crown. *Bioinformation* 2020;16:1139-44.
26. Gopal TM, Rohinikumar S, Thiyaneswaran N, Maiti S. Effect of submandibular fossa on implant length in the posterior mandibular region. *J Long Term Eff Med Implants* 2020;30:219-26.
27. Rupawat D, Maiti S, Nallaswamy D, Sivaswamy V. Aesthetic outcome of implants in the anterior zone after socket preservation and conventional implant placement: A retrospective study. *J Long Term Eff Med Implants* 2020;30:233-9.
28. Muthukrishnan S, Krishnaswamy H, Thanikodi S, Sundaresan D, Venkatraman V. Support vector machine for modelling and simulation of heat exchangers. *Therm Sci* 2020;24 (1 part B):499-503.
29. Nandhini NT, Rajeshkumar S, Mythili S. The possible mechanism of eco-friendly synthesized nanoparticles on hazardous dyes degradation. *Biocatal Agric Biotechnol* 2019;19:101138.
30. Ezhilarasan D. Oxidative stress is bane in chronic liver diseases: Clinical and experimental perspective. *Arab J Gastroenterol* 2018;19:56-64.
31. Rajagopal R, Padmanabhan S, Gnanamani J. A comparison of shear bond strength and debonding characteristics of conventional, moisture-insensitive, and self-etching primers *in vitro*. *Angle Orthod* 2004;74:264-8.
32. Neelakantan P, Sharma S, Shemesh H, Wesselink PR. Influence of irrigation sequence on the adhesion of root canal sealers to dentin: A fourier transform infrared spectroscopy and push-out bond strength analysis. *J Endod* 2015;41:1108-11.
33. Sahu D, Kannan GM, Vijayaraghavan R. Carbon black particle exhibits size dependent toxicity in human monocytes. *Int J Inflamm* 2014;2014:827019.
34. Jose J, Ajitha P, Subbaiyan H. Different treatment modalities followed by dental practitioners for Ellis class 2 fracture – A questionnaire-based survey. *Open Dent J* 2020;14:59-65.
35. Wu F, Zhu J, Li G, Wang J, Veeraraghavan VP, Krishna Mohan S, et al. Biologically synthesized green gold nanoparticles from *Siberian ginseng* induce growth-inhibitory effect on melanoma cells (B16). *Artif Cells Nanomed Biotechnol* 2019;47:3297-305.
36. Dua K, Wadhwa R, Singhvi G, Rapalli V, Shukla SD, Shastri MD, et al. The potential of siRNA based drug delivery in respiratory disorders: Recent advances and progress. *Drug Dev Res* 2019;80:714-30.
37. Patil SB, Durairaj D, Suresh Kumar G, Karthikeyan D, Pradeep D. Comparison of extended nasolabial flap versus buccal fat pad graft in the surgical management of oral submucous fibrosis: A prospective pilot study. *J Maxillofac Oral Surg* 2017;16:312-21.
38. Burawi G, Houston F, Byrne D, Claffey N. A comparison of the dimensional accuracy of the splinted and unsplinted impression techniques for the Bone-Lock implant system. *J Prosthet Dent* 1997;77:68-75.
39. Kim S, Nicholls JJ, Han CH, Lee KW. Displacement of implant components from impressions to definitive casts. *Int J Oral Maxillofac Implants* 2006;21:747-55.
40. Baig MR. Accuracy of impressions of multiple implants in the edentulous arch: A systematic review. *Int J Oral Maxillofac Implants* 2014;29:869-80.