

RESEARCH ARTICLE

SEGEL: A Web Server for Visualization of Smoking Effects on Human Lung Gene Expression

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Abstract

Cigarette smoking is a major cause of death worldwide resulting in over six million deaths per year. Cigarette smoke contains complex mixtures of chemicals that are harmful to nearly all organs of the human body, especially the lungs. Cigarette smoking is considered the major risk factor for many lung diseases, particularly chronic obstructive pulmonary diseases (COPD) and lung cancer. However, the underlying molecular mechanisms of smoking-induced lung injury associated with these lung diseases still remain largely unknown. Expression microarray techniques have been widely applied to detect the effects of smoking on gene expression in different human cells in the lungs. These projects have provided a lot of useful information for researchers to understand the potential molecular mechanism (s) of smoke-induced pathogenesis. However, a user-friendly web server that would allow scientists to fast query these data sets and compare the smoking effects on gene expression across different cells had not yet been established. For that reason, we have integrated eight public expression microarray data sets from trachea epithelial cells, large airway epithelial cells, small airway epithelial cells, and alveolar macrophage into an online web server called SEGEL (Smoking Effects on Gene Expression of Lung). Users can query gene expression patterns across these cells from smokers and nonsmokers by gene symbols, and find the effects of smoking on the gene expression of lungs from this web server. Sex difference in response to smoking is also shown. The relationship between the gene expression and cigarette smoking consumption were calculated and are shown in the server. The current version of SEGEL web server contains 42,400 annotated gene probe sets represented on the Affymetrix Human Genome U133 Plus 2.0 platform. SEGEL will be an invaluable resource for researchers interested in the effects of smoking on gene expression in the lungs. The server also provides useful information for drug development against smoking-related

diseases. The SEGEL web server is available online at http://www.chengfeng.info/smoking_database.html.

Introduction

Data from the World Health Organization (WHO) show that cigarette smoking is one of the biggest threats to public health. Cigarette smoking caused 100 million deaths over the 20th century throughout the world [1]. About 19.5% of American adults are current tobacco users in the United States [2]. Cigarette smoking caused more than 440,000 fatalities per year [3], and smoke-related diseases cost 96 billion dollars in direct medical care expenses and 97 billion dollars in lost productivity in the United States [4]. In the human body, the primary organ that is affected by smoking is the lung. The toxins from cigarette smoke can directly injure lung structures and cause permanent airway damage. In present smokers, nearly 73% of smoking-related problems are serious lung diseases including COPD and lung cancer. Cigarette smoking is a major risk factor for COPD [5–7] that is the third leading cause of mortality and morbidity in the United States. Smoking is accountable for approximately 80–90% of COPD deaths and 90% of the lung cancer deaths in the United States.

Precise detection of changes in gene expression between smokers and nonsmokers is crucial to the understanding of the molecular mechanisms of smoking-induced lung diseases. Recently, high-throughput techniques such as expression microarray have been applied to detect smoking effects on gene expression in human lung cells. These research projects generated valuable data and provided new insights into the effects of tobacco smoke. However, a user-friendly web server that allows scientists to fast query the gene expression levels from these data sets and compare smoking effects on gene expression across different data sets had not been established.

In this paper, we report an online web server called SEGEL (Smoking Effects on Gene Expression of Lung) for easy and fast visualization of smoking effects on human lung gene expression. The web server integrates 362 samples collected from eight public expression microarray data sets from trachea epithelial cells, large airway epithelial cells, small airway epithelial cells, and alveolar macrophage. Gene expression patterns of regular smokers and nonsmokers across these cells can be queried by gene symbols. Sex difference in response to smoking is also shown. The correlation coefficients between the gene expression and cigarette smoking consumption (the number of packs of cigarettes consumed per year) were also calculated and are shown in the web server. The current version of SEGEL contains around 42,400 annotated gene probe sets represented on the Affymetrix Human Genome U133 Plus 2.0. SEGEL will be an invaluable tool and resource for scientists interested in the effects of smoking on lung gene expression. The web server can be used to identify reliable molecular signatures for drug discovery against smoking-related diseases. The SEGEL web server is available online at http://www.chengfeng.info/smoking_database.html

Materials and Methods

362 samples in eight published microarray sets (Shown in [Table 1](#)) from the NCBI Gene Expression Omnibus (GEO) repository were integrated in the SEGEL to compare gene expression of smokers and nonsmokers in alveolar macrophage, small airways epithelial cells, large airways epithelial cells, and trachea epithelial cells. Detailed information of these 362 samples is provided in the Supplementary Information ([S1 Table](#)).

Table 1. Eight microarray datasets integrated in the SEGEL web server.

Data set	Cell	NCBI GEO entry ID	#Nonsmokers	#Smokers	Cigarette intake (Packs-year)
Trachea	Trachea Epithelial Cells	GSE13933	26	19	28±16
LargeAirway1	Large Airway Epithelial Cells	GSE10006	9	20	36±19
LargeAirway2	Large Airway Epithelial Cells	GSE10135	14	18	18±13
SmallAirway1	Small Airway Epithelial Cells	GSE4498	12	10	33 ± 7
SmallAirway2	Small Airway Epithelial Cells	GSE13933	23	19	27 ± 15
SmallAirway3	Small Airway Epithelial Cells	GSE30063	60	73	NA
Macrophage1	Alveolar Macrophage	GSE8823	11	13	36 ± 6
Macrophage2	Alveolar Macrophage	GSE13896	13	22	22 ± 14

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Trachea epithelial cells

The trachea is a tube that connects mouth/nose to the lungs and is protected by an epithelial cell layer. The trachea epithelium is extremely vulnerable to cigarette smoking. One microarray data set GSE13933 was chosen to investigate gene expression of trachea epithelium [8]. This data set includes 26 healthy nonsmokers and 19 regular smokers (28±16 packs-year).

Large airway epithelial cells

Large airways, or bronchi, are airways that are divided from the trachea. Cigarette smoking could possibly be connected with the growth of cancer in the large air epithelium. Two data sets were evaluated in the large airways. GSE10006 [9] and GSE10135 [10] were chosen in SEGEL to evaluate gene expression in large airway epithelial cells. The first data set includes nine healthy nonsmokers and 20 regular smokers (36±19 packs-year). The second data set, GSE10135, includes 14 healthy nonsmokers and 18 regular smokers (18±13 packs-year).

Small airway epithelial cells

Small airway epithelium was considered as the first site of lung pathology induced by cigarette smoking. Three GEO data sets, GSE4498, GSE13933 [8] and GSE30063 [11], were selected in SEGEL. The first data set includes 12 healthy nonsmokers and 10 regular smokers (33 ± 7 packs-year). The second data set includes 26 healthy nonsmokers and 19 regular smokers (27 ± 15 pack/year). The third data set includes 60 healthy nonsmokers and 73 regular smokers.

Alveolar macrophage

An alveolar macrophage is a type of macrophage that is located on the epithelial surface area of bronchi alveoli. Alveolar macrophages supply initial protection against infectious, toxic, or sensitive contaminants from smoking. Alveolar macrophage appears to play an important role in initiating and sustaining COPD progression [12]. Two microarray data sets GSE8823 [13] and GSE13896 [14] were selected in SEGEL. In the first data set, alveolar macrophages were obtained from bronchoalveolar lavage of 11 nonsmokers and 13 regular smokers (36 ± 6 pack-year). In the second data set, alveolar macrophages were obtained from 13 nonsmokers and 22 regular smokers (22 ± 14 pack-year).

Data normalization and analysis

All 362 samples in SEGEL were profiled with the Affymetrix HG-U133 Plus 2.0 GeneChip microarray platform that features comprehensive coverage of the human transcriptome. The current version of SEGEL contains 42,400 annotated gene probe sets with official gene symbols. The RMAexpress program was used to normalize raw microarray data and to summarize expression of the probe sets using the default settings: RMA background correction, quantile normalization, and log₂-transformation. A boxplot (S1 Fig) was used to check whether the signal intensity distributions of samples from different data sets are similar.

Comparisons of the mean expression levels between smokers and nonsmokers were carried out by an unpaired t-test. All figures in SEGEL were plotted using R.

Results and Discussion

SEGEL is a php-based online web server for easy and fast visualization of smoking's effects on human lung gene expression. 362 samples in eight published microarray sets (shown in Table 1) from the NCBI Gene Expression Omnibus (GEO) repository were integrated in SEGEL to compare gene expression of smokers and nonsmokers in alveolar macrophage, small airways epithelial cells, large airways epithelial cells, and trachea epithelial cells. Analyzed microarray data are stored in a MySQL relational database.

Fig 1 shows the query interface for the web server access. The input to the system is official gene symbols, which could be looked up from the NCBI gene database (<http://www.ncbi.nlm.nih.gov/gene>). All probe sets representing the queried gene in the Affymetrix Gene-Chip Human Genome U133 Plus 2.0 Array platform will be shown. The gene expression patterns of each probe set are reported in one PDF file. The file includes graphs that show gene expression patterns of smokers and nonsmokers in different cells, sex difference in response to cigarette smoking, gene expression levels change with age, and the relationships between the gene expression and smoking intake. SEGEL provides some user-friendly functions to query the server. Users can enter a list of multiple gene symbols separated by spaces for the batch searching

Database

Smoking Effects on Gene Expression of Lungs (SEGEL)

Please input the gene symbol* (For example DUOX2, or HSPA6):

All genes included in the SEGEL database could be found [here](#).

*If you cannot find the genes you are interested in, please check their official gene symbols at www.genecards.com

Batch search

You may enter multiple official gene symbols separated by spaces for the batch searching of the TIBS database (for example "ADORA2B HSPA6 PPARG").

Fuzzy search

You may enter a SINGLE query string for the fuzzy searching of the TIBS database (For example, "ADOR" or "HSPA").

Fig 1. The query interface of the SEGEL web server. The input to the web server is official gene symbols. All probe sets representing the queried gene symbol will be shown; each probe set is reported in one PDF file. ADORA2B gene was used as an example to query SEGEL in this paper. The gene has one probe set in the Affymetrix Gene-Chip Human Genome U133 Plus 2.0 platform, *205891_at*.

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(for example, “PPARG HSPA6 ADORA2B”). Results of all genes in the list will be shown. Users can also enter a query string for the fuzzy searching (For example, “ADOR” or “PAR”). Any gene symbols containing the query string will be displayed.

The gene ADORA2B, adenosine A_{2B} receptor was used as an example to query the SEGEL. ADORA2B encodes the adenosine receptor A_{2B}, a member of the G protein-coupled receptors that stimulate adenylate cyclase activity in the presence of adenosine. The ADORA2B is generally defined as “the low affinity” adenosine receptor and requires high micromolar intracellular adenosine concentrations to be activated [15, 16]. In COPD, the lungs are constantly in a hypoxic and inflammatory environment, which causes increased intracellular concentration of adenosine [17]. Higher adenosine concentrations observed in COPD patients may increase A_{2B} receptor activation [17–21]. Evidences have suggested that ADORA2B contributes to the pathogenesis of COPD. First, the ADORA2B gene plays an established role in the inflammatory processes of the lung in smokers[12]. Secondly, the interactions among the adenosine receptors (A₁, A_{2A}, A_{2B} and A₃) can trigger signal events that can further contributes to the expression of other genes including IL-8, IL-1beta, CCL2) all of which have been previously described as COPD-related genes [12, 22, 23]. And thirdly, the engagement of the ADORA2B has been implicated in the disease progression and tissue remodeling [24–26].

ADORA2B has one gene probe set in the Affymetrix Gene-Chip Human Genome U133 Plus 2.0 Array platform, 205891_at. The output results of this probe set are shown in Figs 2–5.

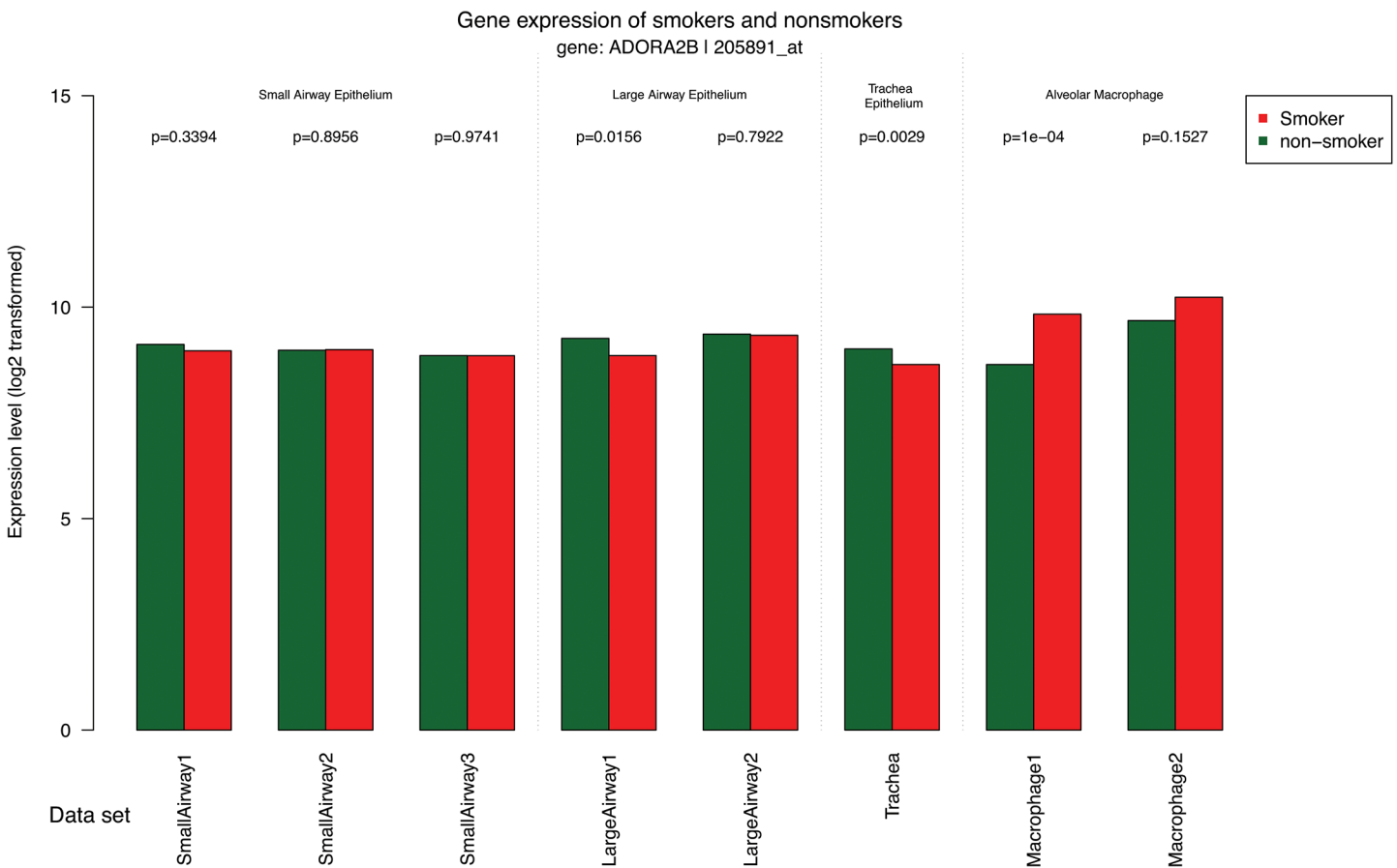


Fig 2. The expression levels of the gene ADORA2B in different cell types from smokers (red bars) and nonsmokers (green bars). These cell types include small airways epithelial cells, large airways epithelial cells, trachea epithelial cells, and alveolar macrophage.

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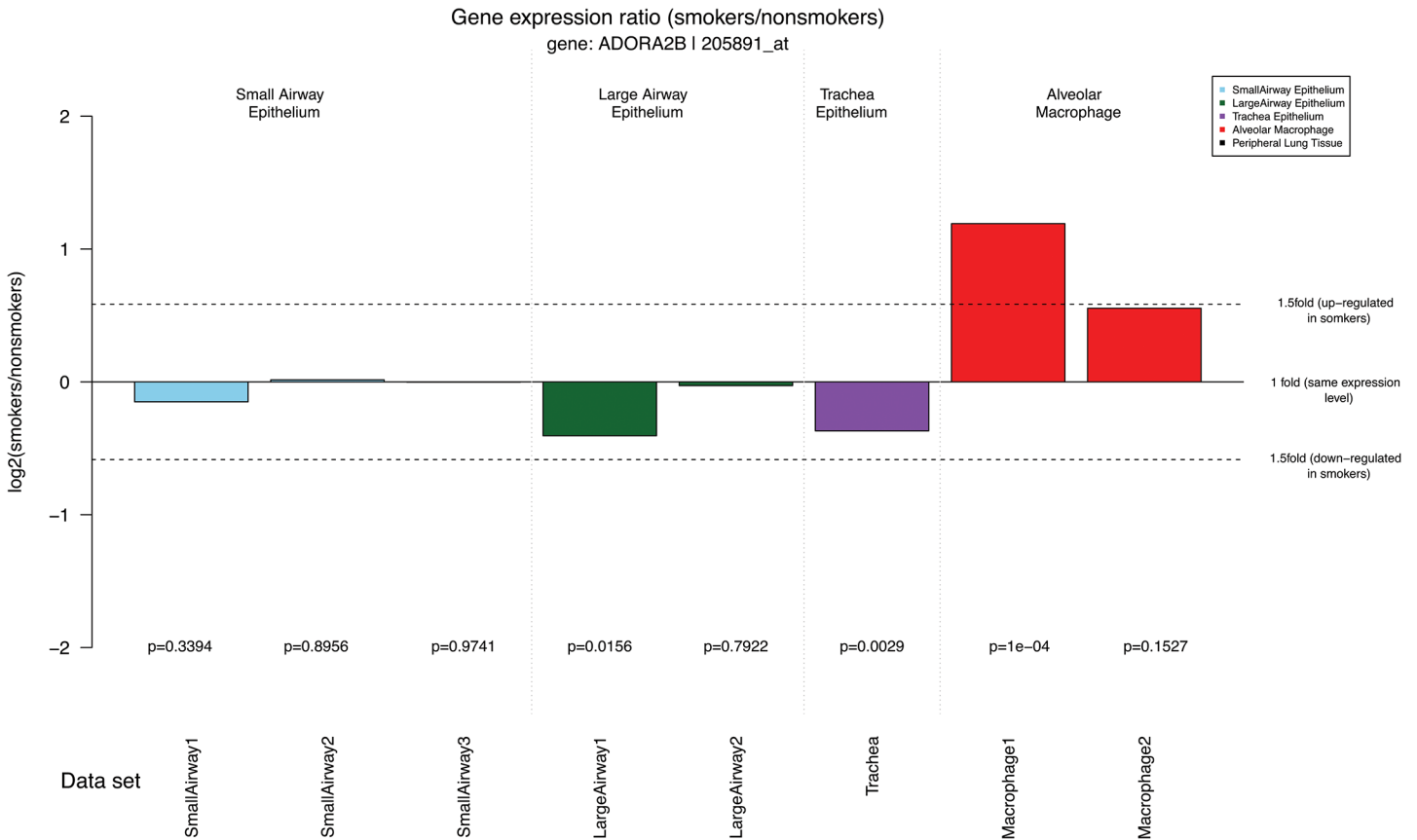


Fig 3. The ratio (log₂-transformed) of ADORA2B expression levels in smokers vs nonsmokers. Positive and negative values in the graph demonstrated up and down regulation of the gene in smokers. The dotted line represents 1,5 fold change.

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The first graph of the output PDF file (Fig 2) shows the spatial differences of ADORA2B gene expression in small airway epithelial cells, large airway epithelial cells, trachea epithelial cells, and alveolar macrophage from smokers (red bars) and nonsmokers (green bars). The ADORA2B gene is highly expressed in alveolar macrophage from smokers as compared to epithelial cells in small airways, large airways, and trachea. Furthermore, our result demonstrated this increase expression of ADORA2B was more prominent in alveolar macrophages from smokers compared to nonsmokers. On the contrary, there were no obvious differences between smokers and nonsmokers in other types of cell. To clearly display the differences of ADORA2B gene expression between smokers and nonsmokers, we calculated the gene expression ratio (log₂-transformed) of smokers vs nonsmokers and presented the values in the second graph of the output PDF file (Fig 3). Positive and negative values in the graph demonstrated up and down regulation of genes in smokers. The dotted line represents a 1.5 fold change in gene expression. Consistent with Fig 2, ADORA2B shows a significant difference in gene expression between smokers and nonsmokers in the first data set of alveolar macrophage. Gene expression differences in other cell types were lower than a 1.5 fold. Our data suggest that ADORA2B is up-regulated in the alveolar macrophage from smokers. Alveolar macrophages are the predominant defense cells in normal lung and during conditions associated with chronic inflammation i.e., COPD [27]. Recent studies revealed stimulation of ADORA2B promotes alternative macrophage activation, which have been implicated to contributing to airway remodeling and lung dysfunction [28, 29]. Results from SEGEL further substantiate these earlier findings.

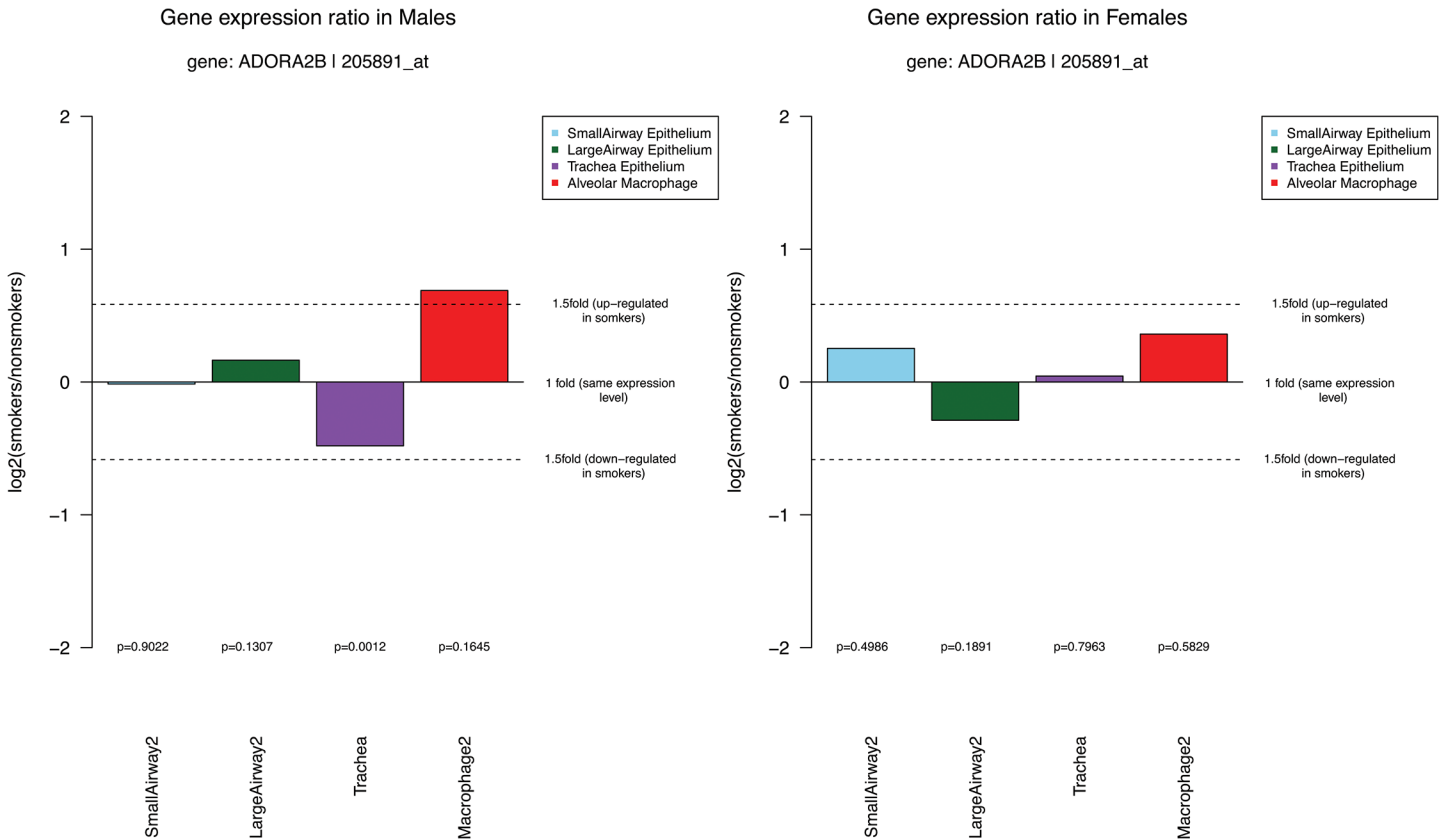


Fig 4. The ratio (log₂-transformed) of ADORA2B expression levels in (a) male smokers vs male nonsmokers (b) female smokers vs female nonsmokers. Positive and negative values in the graph demonstrated up and down regulation of the gene in smokers. The dotted line represents 1.5 fold change.

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Previous studies have showed that airway diseases including COPD are influenced by sex [30]. The third graph in the output PDF file (Fig 4) shows the sex differences in response to cigarette smoke among four microarray data sets from small airway epithelial cells, large airway epithelial cells, trachea epithelial cells, and alveolar macrophage. The other four data sets were not included because the sex information was lacking or there were not enough samples. As shown in the figure, ADORA2B was significantly down regulated in trachea epithelial cells in males but the difference is less than a 1.5 fold. We observed no significant differences in either females or males in the other cell types. The gene expression of males and females in different cells shown in the figure may provide the basis for understanding the molecular mechanisms underlying the sex differences in the prevalence and severity of some smoking-related lung diseases.

There is a close relationship between aging and chronic lung diseases, particularly COPD. COPD is known as a chronic inflammatory disease of the lung that progresses slowly and the majority of the patients are elderly. The classic epidemiology studies suggest death and disability from COPD is related to the rapid decline in lung function with time [31]. We employed SEGEL to track the gene expression change along with the age. The fourth graph in the output PDF file of the web server (Fig 5) demonstrates gene expression level in smokers/nonsmokers along with their age. Seven microarray data sets with the age information were included in this plot. As shown in the figure, Y-axis and X-axis represent the gene expression level

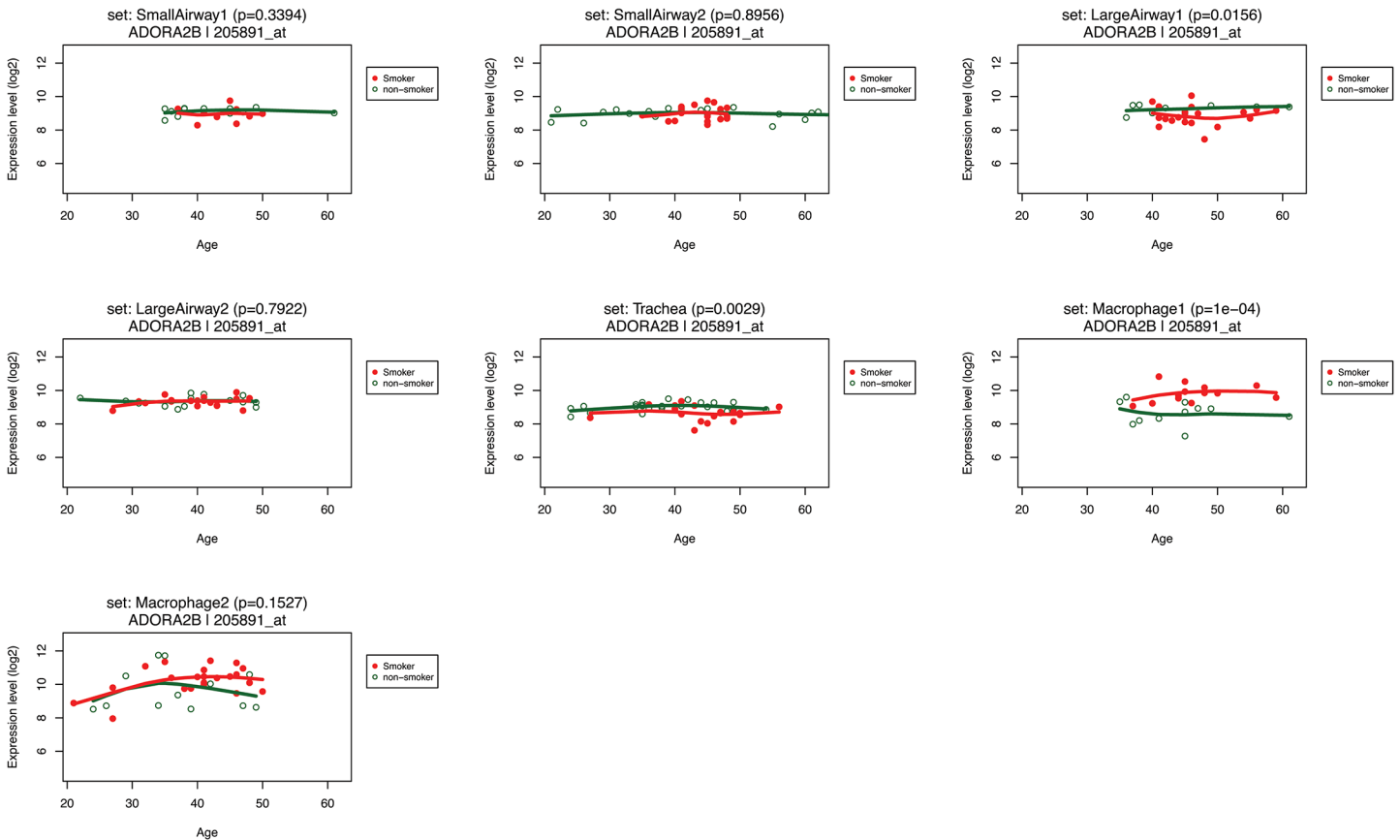


Fig 5. The ADORA2B expression level changes along with age in smokers (red) and nonsmokers (green).

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(log₂-transformed) and age respectively. Alveolar macrophage shows the greatest difference in ADORA2B expression between smokers (red lines) and nonsmokers (green lines), especially after 40 years old. ADORA2B showed a general increase in smokers in alveolar macrophage from 20 to 40 years old but showed constant expression in other types of cells from both smokers and nonsmokers. SEGEL can provide evidence in gene expression levels for numerous studies [32–34] that there is an association between age and smoking-related diseases.

As we all know, heavy smokers have a higher risk than light smokers in the smoking-related diseases. The cause of death in patients with 30 or more packs per year showed significant differences compared to patients with less than 30 packs per year. Smoking pack-years is an important clinical factor in evaluating the risk of lung diseases. However, very little is known if the number of smoking pack-years is related to the gene expression levels. In the fifth graph of the output PDF file (Fig 6), the smoking intake (packs per year) was considered in the analysis. The graph provides a quantitative measurement of the effects of smoking intake on lung gene expression by calculating the Pearson’s correlation coefficients (and corresponding p-values) between gene expression levels and tobacco intake. As shown in Fig 6, the gene ADORA2B expression shows a strong positive correlation with smoking exposure in the first data set of alveolar macrophage (correlation coefficient is greater than 0.5 and p-value = 0.0032). Our data suggest that greater smoking intake will increase the expression level of ADORA2B in the alveolar macrophage from older smokers.

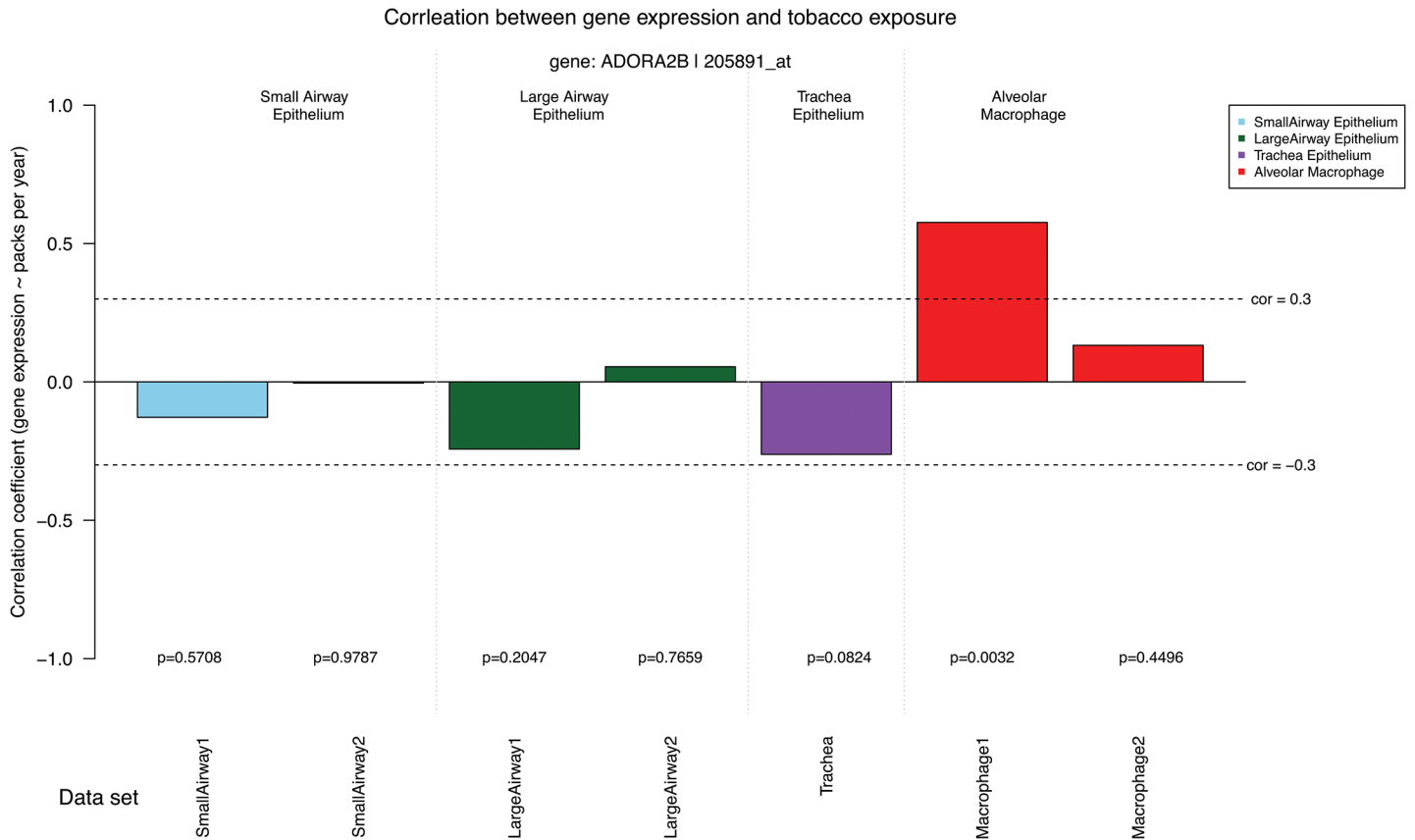


Fig 6. The Pearson's correlation coefficients between ADORA2B expression levels and smoking intake (packs per year).

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SEGEL is an important resource for cigarette smoking research. The web server is comprehensive in sample size, cell types and the number of genes. Previous transcriptome studies of the effects of smoking on lungs have used relatively small numbers of samples or only focused on one lung tissue or on specific genes. SEGEL combined more than 360 samples from eight public microarray data in four cell types (alveolar macrophage, small airways epithelial cells, large airways epithelial cells, and trachea epithelial cells). We also did a comprehensive analysis of more than 42,000 gene probe sets. The analysis includes the spatial difference among tissues/cells, temporal difference, sex difference in response of smoking, and the correlation between gene expression and smoking intake. It will provide deeper insights into the transcriptional foundations of smoking effects on the lungs. SEGEL will also provide useful information regarding the molecular mechanism of smoking-related lung diseases such as COPD, and aid in discovery of novel drugs. Researchers can freely access our server to check the gene expression profiles of disease-related genes, or identify which genes in which types of cells would be important for disease development.

We will expand our web server by adding more public microarray data sets in the future. All samples in our current server were profiled with Affymetrix HG-U133 Plus 2.0 GeneChip microarrays. In the next stage, we will include more samples from different platforms such as Affymetrix HG-U133A or Affymetrix Exon ST 1.0 Array. We believe information provided by the SEGEL web server on smoking effects on the lung will be more reliable with a larger sample size.

Supporting Information

S1 Fig. A boxplot of the signal intensity distributions of 362 samples from eight microarray datasets.

(PDF)

S1 Table. Detail information of these 362 samples integrated in the SEGEL web server.

(DOCX)

Author Contributions

Conceived and designed the experiments: YX BH SSA DAG FC. Performed the experiments: YX BH SSA YL FC. Analyzed the data: YX YL YH FC. Contributed reagents/materials/analysis tools: BH DAG FC. Wrote the paper: DAG FC.

References

1. Edwards R. The problem of tobacco smoking. *Bmj*. 2004; 328(7433):217–9. doi: [10.1136/bmj.328.7433.217](https://doi.org/10.1136/bmj.328.7433.217) PubMed PMID: [14739193](https://pubmed.ncbi.nlm.nih.gov/14739193/); PubMed Central PMCID: PMC318495.
2. King BA, Dube SR, Tynan MA. Current tobacco use among adults in the United States: findings from the National Adult Tobacco Survey. *American journal of public health*. 2012; 102(11):e93–e100. doi: [10.2105/AJPH.2012.301002](https://doi.org/10.2105/AJPH.2012.301002) PubMed PMID: [22994278](https://pubmed.ncbi.nlm.nih.gov/22994278/); PubMed Central PMCID: PMC3477973.
3. Richter P, Pechacek T, Swahn M, Wagman V. Reducing levels of toxic chemicals in cigarette smoke: a new Healthy People 2010 objective. *Public health reports*. 2008; 123(1):30–8. PubMed PMID: [18348477](https://pubmed.ncbi.nlm.nih.gov/18348477/); PubMed Central PMCID: PMC2099323.
4. Johnson BA, Messing RO, Charness ME, Crabbe JC, Goldman MS, Harris RA, et al. How Should Addiction-Related Research at the National Institutes of Health be Reorganized? *Frontiers in psychiatry*. 2011; 2:2. doi: [10.3389/fpsy.2011.00002](https://doi.org/10.3389/fpsy.2011.00002) PubMed PMID: [21629835](https://pubmed.ncbi.nlm.nih.gov/21629835/); PubMed Central PMCID: PMC3098615.
5. Dubar V, Gosset P, Aerts C, Voisin C, Wallaert B, Tonnel AB. In vitro acute effects of tobacco smoke on tumor necrosis factor alpha and interleukin-6 production by alveolar macrophages. *Experimental lung research*. 1993; 19(3):345–59. Epub 1993/05/01. PubMed PMID: [8319604](https://pubmed.ncbi.nlm.nih.gov/8319604/).
6. Mio T, Romberger DJ, Thompson AB, Robbins RA, Heires A, Rennard SI. Cigarette smoke induces interleukin-8 release from human bronchial epithelial cells. *American journal of respiratory and critical care medicine*. 1997; 155(5):1770–6. Epub 1997/05/01. doi: [10.1164/ajrccm.155.5.9154890](https://doi.org/10.1164/ajrccm.155.5.9154890) PubMed PMID: [9154890](https://pubmed.ncbi.nlm.nih.gov/9154890/).
7. Shoji S, Ertl RF, Koyama S, Robbins R, Leikauf G, Von Essen S, et al. Cigarette smoke stimulates release of neutrophil chemotactic activity from cultured bovine bronchial epithelial cells. *Clin Sci (Lond)*. 1995; 88(3):337–44. Epub 1995/03/01. PubMed PMID: [7736704](https://pubmed.ncbi.nlm.nih.gov/7736704/).
8. Turetz ML, O'Connor TP, Tilley AE, Strulovici-Barel Y, Salit J, Dang D, et al. Trachea epithelium as a "canary" for cigarette smoking-induced biologic phenotype of the small airway epithelium. *Clinical and translational science*. 2009; 2(4):260–72. doi: [10.1111/j.1752-8062.2009.00129.x](https://doi.org/10.1111/j.1752-8062.2009.00129.x) PubMed PMID: [20443905](https://pubmed.ncbi.nlm.nih.gov/20443905/); PubMed Central PMCID: PMC3875387.
9. Carolan BJ, Harvey BG, De BP, Vanni H, Crystal RG. Decreased expression of intelectin 1 in the human airway epithelium of smokers compared to nonsmokers. *Journal of immunology*. 2008; 181(8):5760–7. PubMed PMID: [18832735](https://pubmed.ncbi.nlm.nih.gov/18832735/); PubMed Central PMCID: PMC2651682.
10. Vanni H, Kazeros A, Wang R, Harvey BG, Ferris B, De BP, et al. Cigarette smoking induces overexpression of a fat-depleting gene AZGP1 in the human. *Chest*. 2009; 135(5):1197–208. doi: [10.1378/chest.08-1024](https://doi.org/10.1378/chest.08-1024) PubMed PMID: [19188554](https://pubmed.ncbi.nlm.nih.gov/19188554/); PubMed Central PMCID: PMC2679098.
11. Wang R, Ahmed J, Wang G, Hassan I, Strulovici-Barel Y, Salit J, et al. Airway epithelial expression of TLR5 is downregulated in healthy smokers and smokers with chronic obstructive pulmonary disease. *Journal of immunology*. 2012; 189(5):2217–25. doi: [10.4049/jimmunol.1101895](https://doi.org/10.4049/jimmunol.1101895) PubMed PMID: [22855713](https://pubmed.ncbi.nlm.nih.gov/22855713/); PubMed Central PMCID: PMC3579667.
12. Poliska S, Csanky E, Szanto A, Szatmari I, Mesko B, Szeles L, et al. Chronic obstructive pulmonary disease-specific gene expression signatures of alveolar macrophages as well as peripheral blood monocytes overlap and correlate with lung function. *Respiration; international review of thoracic diseases*. 2011; 81(6):499–510. doi: [10.1159/000324297](https://doi.org/10.1159/000324297) PubMed PMID: [21430361](https://pubmed.ncbi.nlm.nih.gov/21430361/).
13. Kazeros A, Harvey BG, Carolan BJ, Vanni H, Krause A, Crystal RG. Overexpression of apoptotic cell removal receptor MERTK in alveolar macrophages of cigarette smokers. *American journal of*

- respiratory cell and molecular biology. 2008; 39(6):747–57. doi: [10.1165/rcmb.2007-0306OC](https://doi.org/10.1165/rcmb.2007-0306OC) PubMed PMID: [18587056](https://pubmed.ncbi.nlm.nih.gov/18587056/); PubMed Central PMCID: PMC2586050.
14. Shaykhiev R, Krause A, Salit J, Strulovici-Barel Y, Harvey BG, O'Connor TP, et al. Smoking-dependent reprogramming of alveolar macrophage polarization: implication for pathogenesis of chronic obstructive pulmonary disease. *Journal of immunology*. 2009; 183(4):2867–83. doi: [10.4049/jimmunol.0900473](https://doi.org/10.4049/jimmunol.0900473) PubMed PMID: [19635926](https://pubmed.ncbi.nlm.nih.gov/19635926/); PubMed Central PMCID: PMC2873685.
 15. Feoktistov I, Biaggioni I. Adenosine A2B receptors. *Pharmacological reviews*. 1997; 49(4):381–402. Epub 1998/01/27. PubMed PMID: [9443164](https://pubmed.ncbi.nlm.nih.gov/9443164/).
 16. Beukers MW, den Dulk H, van Tilburg EW, Brouwer J, Ijzerman AP. Why are A(2B) receptors low-affinity adenosine receptors? Mutation of Asn273 to Tyr increases affinity of human A(2B) receptor for 2-(1-Hexynyl)adenosine. *Molecular pharmacology*. 2000; 58(6):1349–56. Epub 2000/11/28. PubMed PMID: [11093773](https://pubmed.ncbi.nlm.nih.gov/11093773/).
 17. Mohsenin A, Blackburn MR. Adenosine signaling in asthma and chronic obstructive pulmonary disease. *Current opinion in pulmonary medicine*. 2006; 12(1):54–9. Epub 2005/12/17. PubMed PMID: [16357580](https://pubmed.ncbi.nlm.nih.gov/16357580/).
 18. Spicuzza L, Di Maria G, Polosa R. Adenosine in the airways: implications and applications. *European journal of pharmacology*. 2006; 533(1–3):77–88. Epub 2006/02/07. doi: [10.1016/j.ejphar.2005.12.056](https://doi.org/10.1016/j.ejphar.2005.12.056) PubMed PMID: [16458886](https://pubmed.ncbi.nlm.nih.gov/16458886/).
 19. Sun CX, Zhong H, Mohsenin A, Morschl E, Chunn JL, Molina JG, et al. Role of A2B adenosine receptor signaling in adenosine-dependent pulmonary inflammation and injury. *The Journal of clinical investigation*. 2006; 116(8):2173–82. Epub 2006/07/15. doi: [10.1172/JCI27303](https://doi.org/10.1172/JCI27303) PubMed PMID: [16841096](https://pubmed.ncbi.nlm.nih.gov/16841096/); PubMed Central PMCID: PMC1501110.
 20. Chunn JL, Mohsenin A, Young HW, Lee CG, Elias JA, Kellems RE, et al. Partially adenosine deaminase-deficient mice develop pulmonary fibrosis in association with adenosine elevations. *American journal of physiology Lung cellular and molecular physiology*. 2006; 290(3):L579–87. Epub 2005/11/01. doi: [10.1152/ajplung.00258.2005](https://doi.org/10.1152/ajplung.00258.2005) PubMed PMID: [16258000](https://pubmed.ncbi.nlm.nih.gov/16258000/).
 21. Fredholm BB, Irenius E, Kull B, Schulte G. Comparison of the potency of adenosine as an agonist at human adenosine receptors expressed in Chinese hamster ovary cells. *Biochemical pharmacology*. 2001; 61(4):443–8. Epub 2001/02/28. PubMed PMID: [11226378](https://pubmed.ncbi.nlm.nih.gov/11226378/).
 22. Heguy A, O'Connor TP, Luettich K, Worgall S, Ciecuch A, Harvey BG, et al. Gene expression profiling of human alveolar macrophages of phenotypically normal smokers and nonsmokers reveals a previously unrecognized subset of genes modulated by cigarette smoking. *Journal of molecular medicine*. 2006; 84(4):318–28. doi: [10.1007/s00109-005-0008-2](https://doi.org/10.1007/s00109-005-0008-2) PubMed PMID: [16520944](https://pubmed.ncbi.nlm.nih.gov/16520944/).
 23. Ning W, Li CJ, Kaminski N, Feghali-Bostwick CA, Alber SM, Di YP, et al. Comprehensive gene expression profiles reveal pathways related to the pathogenesis of chronic obstructive pulmonary disease. *Proceedings of the National Academy of Sciences of the United States of America*. 2004; 101(41):14895–900. doi: [10.1073/pnas.0401168101](https://doi.org/10.1073/pnas.0401168101) PubMed PMID: [15469929](https://pubmed.ncbi.nlm.nih.gov/15469929/); PubMed Central PMCID: PMC522001.
 24. Caruso M, Holgate ST, Polosa R. Adenosine signalling in airways. *Current opinion in pharmacology*. 2006; 6(3):251–6. doi: [10.1016/j.coph.2006.02.002](https://doi.org/10.1016/j.coph.2006.02.002) PubMed PMID: [16584919](https://pubmed.ncbi.nlm.nih.gov/16584919/).
 25. Rennard SI. Inflammation and repair processes in chronic obstructive pulmonary disease. *American journal of respiratory and critical care medicine*. 1999; 160(5 Pt 2):S12–6. doi: [10.1164/ajrccm.160.supplement_1.5](https://doi.org/10.1164/ajrccm.160.supplement_1.5) PubMed PMID: [10556162](https://pubmed.ncbi.nlm.nih.gov/10556162/).
 26. Richter A, O'Donnell RA, Powell RM, Sanders MW, Holgate ST, Djukanovic R, et al. Autocrine ligands for the epidermal growth factor receptor mediate interleukin-8 release from bronchial epithelial cells in response to cigarette smoke. *American journal of respiratory cell and molecular biology*. 2002; 27(1):85–90. doi: [10.1165/ajrcmb.27.1.4789](https://doi.org/10.1165/ajrcmb.27.1.4789) PubMed PMID: [12091250](https://pubmed.ncbi.nlm.nih.gov/12091250/).
 27. Shapiro SD. The macrophage in chronic obstructive pulmonary disease. *American journal of respiratory and critical care medicine*. 1999; 160(5 Pt 2):S29–32. Epub 1999/11/11. doi: [10.1164/ajrccm.160.supplement_1.9](https://doi.org/10.1164/ajrccm.160.supplement_1.9) PubMed PMID: [10556166](https://pubmed.ncbi.nlm.nih.gov/10556166/).
 28. Gordon S. Alternative activation of macrophages. *Nature reviews Immunology*. 2003; 3(1):23–35. Epub 2003/01/04. doi: [10.1038/nri978](https://doi.org/10.1038/nri978) PubMed PMID: [12511873](https://pubmed.ncbi.nlm.nih.gov/12511873/).
 29. Csoka B, Selmecey Z, Koscsó B, Nemeth ZH, Pacher P, Murray PJ, et al. Adenosine promotes alternative macrophage activation via A2A and A2B receptors. *FASEB journal: official publication of the Federation of American Societies for Experimental Biology*. 2012; 26(1):376–86. Epub 2011/09/20. doi: [10.1096/fj.11-190934](https://doi.org/10.1096/fj.11-190934) PubMed PMID: [21926236](https://pubmed.ncbi.nlm.nih.gov/21926236/); PubMed Central PMCID: PMC3250237.
 30. Sorheim IC, Johannessen A, Gulsvik A, Bakke PS, Silverman EK, DeMeo DL. Gender differences in COPD: are women more susceptible to smoking effects than men? *Thorax*. 2010; 65(6):480–5. Epub 2010/06/05. doi: [10.1136/thx.2009.122002](https://doi.org/10.1136/thx.2009.122002) PubMed PMID: [20522842](https://pubmed.ncbi.nlm.nih.gov/20522842/).

31. Fletcher C, Peto R. The natural history of chronic airflow obstruction. *British medical journal*. 1977; 1 (6077):1645–8. Epub 1977/06/25. PubMed PMID: [871704](#); PubMed Central PMCID: PMC1607732.
32. Ito K, Lim S, Caramori G, Cosio B, Chung KF, Adcock IM, et al. A molecular mechanism of action of theophylline: Induction of histone deacetylase activity to decrease inflammatory gene expression. *Proceedings of the National Academy of Sciences of the United States of America*. 2002; 99(13):8921–6. Epub 2002/06/19. doi: [10.1073/pnas.132556899](#) PubMed PMID: [12070353](#); PubMed Central PMCID: PMC124399.
33. Tudor RM, Yoshida T, Arap W, Pasqualini R, Petrache I. State of the art. Cellular and molecular mechanisms of alveolar destruction in emphysema: an evolutionary perspective. *Proceedings of the American Thoracic Society*. 2006; 3(6):503–10. Epub 2006/08/22. doi: [10.1513/pats.200603-054MS](#) PubMed PMID: [16921129](#); PubMed Central PMCID: PMC2647641.
34. Vogelmeier C, Bals R. Chronic obstructive pulmonary disease and premature aging. *American journal of respiratory and critical care medicine*. 2007; 175(12):1217–8. Epub 2007/06/05. doi: [10.1164/rccm.200703-513ED](#) PubMed PMID: [17545454](#).