

# Prognostic significance of mutated genes in megakaryocytic disorders

Ali Amin Asnafi, Mohammad bagher Mohammadi, Hadi Rezaeeyan, Nader Davari, Najmaldin Saki

Thalassemia and Hemoglobinopathy Research Center, Research Institute of Health, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

## Abstract

Megakaryopoiesis is a process during which platelets that play a major role in hemostasis are produced due to differentiation and maturation of megakaryocytic precursors. Several genes, including oncogenes and tumor suppressor genes, play a role in the regulation of this process. This study was conducted to investigate the oncogenes and tumor suppressor genes as well as their mutations during the megakaryopoiesis process, which can lead to megakaryocytic disorders. Relevant literature was identified by a PubMed search (1998-2019) of English language papers using the terms 'Megakaryopoiesis', 'Mutation', 'oncogenes', and 'Tumor Suppressor'. According to investigations, several mutations occur in the genes implicated in megakaryopoiesis, which abnormally induce or inhibit megakaryocyte production, differentiation, and maturation, leading to platelet disorders. GATA-1 is one of the important genes in megakaryopoiesis and its mutations can be considered among the factors involved in the incidence of these disorders. Considering the essential role of these genes (such as GATA-1) in megakaryopoiesis and the involvement of their mutations in platelet disorders, study and examination of these changes can be a positive step in the diagnosis and prognosis of these diseases.

Correspondence: Najmaldin Saki, Thalassemia and Hemoglobinopathy Research Center, Research Institute of Health, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran. Tel.: +98.6113738317 - Fax: +98.6113738330. E-mail: najmaldinsaki@gmail.com

Acknowledgements: this study is part of an MSc thesis from Seyed Mohammad Bagher Mohammadi. Special thanks to Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran for their financial support.

Contributions: NS conceived the manuscript and revised it; AAA, MBM, HR and ND wrote the manuscript and prepared the tables.

Conflict of interest: the authors declare no conflict of interest.

Received for publication: 6 December 2018. Revision received: 27 June 2019. Accepted for publication: 28 June 2018.

This work is licensed under a Creative Commons Attribution NonCommercial 4.0 License (CC BY-NC 4.0).

©Copyright: the Author(s), 2019 Licensee PAGEPress, Italy Oncology Reviews 2019; 13:408 doi:10.4081/oncol.2019.408

## Introduction

Megakaryopoiesis is as a process in which bone marrow (BM) progenitors are progressively differentiated into megakaryocytes and finally platelets.<sup>1</sup> Several genes, including oncogenes and tumor suppressors, are involved in the process of megakaryopoiesis.2 Oncogenes and tumor suppressors exert their effects via regulating the expressions of cell surface receptors, growth factors, cytokines, signaling molecules, transcription factors, as well as controlling the cell cycle and apoptosis.<sup>3</sup> Oncogenes are the genes that increase cell proliferation and decrease or even inhibit cell apoptosis, so that any mutation and dysfunction in these genes may lead to excessive proliferation of malignant cells and potentiate the activity of these cells. In contrast, tumor suppressors are the genes that reduce proliferation of cells, increase their differentiation and induce apoptosis in a normal situation, which can lead to elimination or decreased activity of tumor cells.<sup>2</sup> However, any disturbance in the expression or function of these genes (e.g. mutation) may interfere with the production of various cells, including hematopoietic cells. Impaired production of hematopoietic cells disrupts the generation of cells differentiated from them, including erythrocytes, leukocytes, and platelets.<sup>3</sup> It is worth noting that mutations occur in each of these tumor suppressor genes, which contribute to the diversity of their function as well as effect on the process of megakaryopoiesis, predisposing people to certain diseases.<sup>4,5</sup> In this review, we examine the mutations in genes effective upon megakaryopoiesis, their role as tumor suppressors or oncogenes, and their effect upon platelet production process and related diseases.

# Oncogenes in normal and mutated situation and their effect on megakaryopoiesis

# GATA-1

GATA-1 is an important transcription factor in maturation and differentiation processes of different hematopoietic lineages, especially megakaryocytes.<sup>6</sup> It has two zinc finger motifs, one at C-terminal and the other at N-terminal. The latter plays a role in GATA-1 binding to DNA, as well as in the interaction with Friend Of GATA-1 (FOG-1) cofactor and the formation of GATA-1: FOG-1 complex.<sup>7,8</sup> FOG-1 is a protein with zinc finger motif interacting with GATA-1 to induce the maturation and differentiation of GATA-1 or FOG-1, *i.e.* impaired interaction of these two molecules can be a function of mutation in either of these two factors (especially GATA-1), leading to megakaryocytic disorders such as X-linked thrombocytopenia.<sup>10</sup> Several variants occur due to a

mutation in GATA-1, each somehow affecting GATA-1 function and thus platelet function, including V205M that results from a missense mutation in C-terminal of GATA-1 (Val205Met).11 Val205 is a highly conserved amino acid, which plays an essential role in binding and interaction of GATA-1: FOG-1; therefore, mutation in this amino acid reduces GATA-1 binding affinity to FOG-1 and eventually impairs the production, maturation, and differentiation of megakaryocytic lineage via inhibiting the normal function of GATA-1. V205M mutation also leads to platelet disorders such as macrothrombocytopenia, which indicates the important role of GATA-1: FOG-1 complex in the production of platelets.10,11 Another variant, namely D218G that leads to macrothrombocytopenia is a function of mutation in C-terminal GATA-1, which reduces GATA-1 affinity to FOG-1but does not inhibit GATA-1: DNA binding.<sup>12,13</sup> Asp218 amino acid may be converted to Tyr due to mutation, which leads to D218Y variant and is quite similar to D218G in terms of mutation site. The difference between the two mutations lies in the severity of clinical symptoms among patients, so that the severity of clinical complications is higher in D218Y because GATA-1 affinity to FOG-1 is lower in D218Y than D218G but is similar to V205M. It is worth mentioning that macrothrombocytopenia resulting from D218Y is more severe than D218G.<sup>11,13</sup> The next variant, which is a function of mutation in codon 208 of C-terminal GATA-1, is called G208S and leads to the conversion of Gly to Ser.<sup>7</sup> Following this mutation, the interaction of FOG-1: GATA-1 is also disrupted due to the reduction of GATA-1 affinity to its cofactor because Gly208 plays a crucial role in binding to FOG-1 and is therefore essential for the production of normal platelets.<sup>9</sup> This mutation also leads to macrothrombocytopenia.14 Another mutation may occur in codon 208 converting Gly to Arg and resulting in the incidence of G208R. The difference between G208S and G208R mutations is that the latter interferes with and reduces GATA-1: DNA binding in addition to decreasing GATA-1 affinity to FOG-1. It follows from the above explanations that the clinical symptoms of patients harboring G208R mutation are more severe than those of G208S, and these patients will be afflicted with severe thrombocytopenia of macrothrombocytopenia type.7 R216Q is another variant of GATA-1 that is derived from a missense mutation in C-terminal GATA-1 in which Gln replaces Arg.<sup>8,15</sup> Unlike previous mutations, in this case GATA-1: FOG-1 interaction is normal and only GATA-1 binding to its site on DNA is disrupted. Patients bearing this mutation have beta-thalassemia as well as macrothrombocytopenia.11 Indeed, R216Q mutation is associated with X-Linked Thrombocytopenia with Thalassemia (XLTT).<sup>16</sup> The binding mechanism of GATA-1: FOG-1 from a molecular perspective is as follows. In normal conditions, the amino acid of interest (e.g. Val205) is phosphorylated by AKT (a serine-threonine kinase), GATA-1 binding affinity to FOG-1 is increased, and megakaryocytic lineage is normally proliferated. However, following a mutation in GATA-1 (such as V205M), the amino acid is not phosphorylated by AKT, resulting in disrupted formation of GATA-1: FOG-1 complex<sup>17</sup> (Table 1<sup>6-35</sup>). As discussed, GATA-1 plays a central role in megakaryopoiesis, and mutation in GATA-1 can lead to the incidence of megakaryocytic disorders. The question that arises here is whether GATA-1 can be used for the diagnosis or prognosis of megakaryocytic disorders (Figure 1).

#### Myeloproliferative leukemia protein

[page 98]

Myeloproliferative leukemia protein (MPL) is a member of Cytokine Receptor Superfamily, which plays an essential role in platelet production by inducing proliferation and maturation along with its ligand of Thrombopoietin (TPO).<sup>18,19</sup> To date, a majority of MPL gene mutations have affected the amino acids in



Transmembrane or Extracellular Domains (such as Ser or Arg). W508S is a new MPL variant, which, unlike what stated up to now, is the result of a mutation in Intracellular Domain leading to Ser substitution for Trp. Intracellular signaling pathways like JAK-STAT and PI3K-AKT-Bad are activated because of this mutation.<sup>18</sup> As JAK-STAT (JAK2 and STAT-3 and STAT-5) signaling pathway is involved in the induction of proliferation and maturation of megakaryocytes,<sup>20</sup> the occurrence of W508S mutation may lead to increased production of platelets and thrombocytosis. Megakaryocytic progenitors harboring MPL-W508S are only able to proliferate but cannot differentiate.<sup>18</sup> S505N is another variant resulting from a point mutation converting Ser into Asn. The incidence of this variant relaxes the inhibitory effect on TPO gene and hence increases the translation and expression of TPO gene, leading to increased serum TPO and thus enhanced platelet production.<sup>19</sup> R102P is another mutation with a negative inhibitory effect on megakaryopoiesis, which has been identified in Congenital Amegakaryocytic Thrombocytopenia<sup>21</sup> (Table 1).

#### Friend leukemia virus integration-1

Friend leukemia virus integration-1 (FLI-1) is a transcription factor and member of E-twenty-six (ETS) (11q24.3) family.<sup>22,23</sup> Normal expression of FLI-1 plays an important role in the production and evolution of megakaryocytes; in other words, it induces the expression of some genes involved in megakaryocyte maturation and differentiation, including GP6, c-mpl, GP9, and ITGA2B.24,25 Although a heterozygous mutation has been recently discovered in FLI-1 leading to thrombocytopenia,<sup>22</sup> most disorders causing the impairment and defective function of FLI-1 are a function of a homozygous mutation or deletion in 11q chromosome. This heterozygous mutation is a missense mutation, which commonly occurs in codon970 of the gene and leads to the conversion of Arg to Trp in FLI-1 DNA-Binding domain.<sup>26</sup> As a result, the transcriptional activity of FLI-1 target genes such as GP6, c-mpl, GP9, and ITGA2B decreases and subsequently leads to impaired production of platelets.<sup>24</sup> In general, mutation in FLI-1 causing impairment or loss of its function leads to Inherited Thrombocytopenia, especially Jacobson Syndrome and Paris-Trousseau Syndrome. As noted above, FLI-1 is located on 11g chromosome<sup>23</sup> and most patients with Paris Trousseau Syndrome lack FLI-1;24 therefore, to achieve targeted therapy for differential diagnosis of this syndrome or to find the genetic defect of this disease, chromosome 11 (especially its long arm) can be assessed, or this molecule can be considered as a diagnostic factor of the disease.

#### Other oncogenes

Myeloblastosis (myb) is a gene downregulating the platelet production process.<sup>32</sup> Also, thrombopoietin (TPO) gene induces megakaryopoiesis, which leads to platelet production in physiologic conditions.<sup>33</sup> One of the effective protooncogenes in megakaryopoiesis is C-myc, which downregulates megakaryopoiesis and any mutation in this gene can lead to platelet disorders such as thrombosis.<sup>34</sup> The last protooncogene we discuss is LNK (SH2B3) that inhibits cell proliferation by downregulating cytokine signaling<sup>35</sup> (Table 1).

Tumor suppressor genes in normal and mutated situation and their effect on megakaryopoiesis

P53

P53 is an essential tumor suppressor transcription factor, which





is activated in conditions such as stress or hypoxia in the cell, leading to the cessation of cell cycle, inhibition of cell proliferation, and finally apoptosis. The activity of P53 is increased in later stages of megakaryocyte differentiation, leading to endomitosis arrest via suppression of polyploidization and thus the onset of apoptosis. Therefore, no megakaryocytic apoptosis occurs due to mutation in the gene of this transcription factor, which can lead to aneuploidy or polyploidy.<sup>27-29</sup> Therefore, it may be concluded that the absence or dysfunction of P53 results in unchecked production and proliferation of platelets, which can lead to platelet disorders.

# **ETS variant 6**

ETS Variant 6 (ETV6) is a transcription factor that inhibits the

Table 1. Commo	n mutated ge	enes that lea	d to platel	et disorders.
----------------	--------------	---------------	-------------	---------------

controlling the activity of ETS, including FLI-1. Mutation in ETV6 results in impaired platelet function and problems such as thrombocytopenia or megakaryocyte hyperplasia.<sup>24,25,30</sup> In one study, after examining three groups of patients with thrombocytopenia, a missense mutation was observed in ETV6, which led to three types of amino acid translocations in ETS binding domain of ETV6 gene, including Arg399Cys, Arg369Gln, and Pro214Leu. In the first two variants, amino acids playing a role in DNA binding change and thus interfere with the binding and interaction of ETV6 with DNA, and in the third one, Internal Linker Domain involved in DNA binding is disrupted. In other words, this mutation in ETV6 impairs its binding to DNA and reduces the effect of tran-

proliferation, differentiation, and maturation of megakaryocytes by

		-	-			
Gene	Chro.	Variant	Function	Disease	Method	Ref.
GATA-1	Xp11.23	V205M D218G D218Y G208S G208R R216Q	GATA-1:FOG-1 interaction GATA-1:DNA binding	Macrothrombocytopenia Macrothrombocytopenia Macrothrombocytopenia Thrombocytopenia Thrombocytopenia XLTT	Nested PCR PCR	6,7,10,11,13,14,16
MPL	1p34.2	W508S S505N R102P	Activates the signal transduction pathways Removes the inhibition on TPO Decreases megakaryopoiesis	Thrombosis Excessive platelets production CAMT	PCR	18,19,21
Myb	6q23.3	-	Downregulation of megakaryopoiesis	-	RT-qPCR	32
TPO	2p25.3	-	Upregulation of megakaryopoiesis	Thrombocytopenia	-	33
C-myc	8q24.21	-	Downregulation of megakaryopoiesis	Thrombosis	RT-PCR	34
LKN (SH2B3)	12q24.12	R262W	Downregulation of cytokine signaling and cell proliferation	Excessive platelets production	RT-qPCR	35

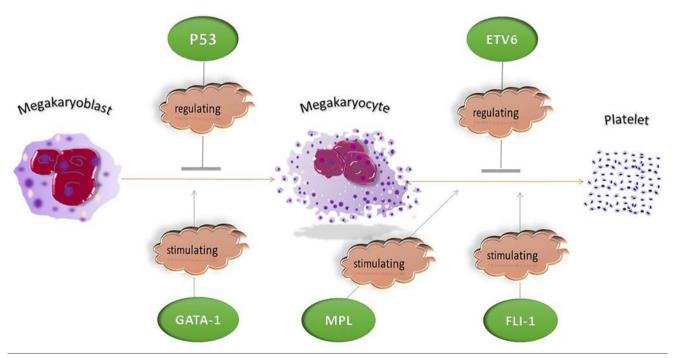


Figure 1. GATA-1 is an important transcription factor in megakaryopoiesis. It stimulates MKs maturation and differentiation through binding to its cofactor called FOG-1. GATA-1: FOG-1 interaction causing multipotent progenitor cell differentiation to committed megakaryocyte progenitor cell and then platelet production. Several variants occur due to mutation in GATA-1, each somehow affecting GATA-1 function and thus platelet function, including V205M, D218G, D218Y, G208S, G208R and R216Q. The first five variants cause macrothrombocytopenia and the last one cause XLTT. MKs, megakaryocytes; FOG-1, friend Of GATA-1; V, valine; M, methionine; D, aspartic Acid; G, glycine; Y, tyrosine; S, serine; R, arginine; Q, glutamine; XLTT, X-linked thrombocytopenia with thalassemia.

scriptional inhibitors, leading to disruption of hematopoiesis (Figure 2). $^{31}$ 

# Discussion

Several genes control the process of megakaryopoiesis, some of which have oncogenic properties (such as GATA-1 and MPL) and others are tumor suppressive (e.g. P53 and ETV-6). However, any mutation in such genes leading to increased or decreased expression of them affects megakaryopoiesis and thus platelet production.<sup>36</sup> GATA-1 is one of the most important genes involved in megakaryopoiesis, which interact with FOG-1 to induce megakarvocvtic differentiation and maturation, so that any mutation and emergence of different variants upset megakaryopoiesis and platelet production. V205M, G208R, G208S, and R216Q are among these variants. V205M is a missense mutation decreasing GATA-1 affinity to FOG-1, G208S variant disrupts the formation of GATA-1: FOG-1, whereas G208R decreases GATA-1 binding to DNA in addition to the reduction of GATA-1: FOG-1 binding, and finally R216Q variant leads to platelet disorders through disruption in GATA-1: DNA binding.<sup>7,11</sup> It is notable that V205M, G208S and G208R cause X-Linked Thrombocytopenia (XLT) with increased Mean Platelet Volume (MPV) and that R216O causes X-Linked Thrombocytopenia with Thalassemia (XLTT).<sup>37</sup> MPL is another gene that plays an essential role in megakaryopoiesis. In contrast to GATA-1, mutations of MPL cause disorders associated with



ACCESS

increased platelet counts (thrombosis).<sup>18</sup> Considering the content discussed in this paper, it is likely to hypothesize that since the variants of genes involved in megakaryopoiesis (especially GATA-1) play a crucial role in the incidence of platelet disorders, the study of these variants in patients (even when the patient has other symptoms such as erythroid disorders as well as platelet disorders) is useful and contributes to the diagnosis and prognosis of these diseases.

# **Conclusions and future perspective**

The current study reveals the importance of detection and investigation of genes mutations such as oncogenes (*e.g.* GATA-1 and MPL) and tumor suppressor genes (*e.g.* P53 and ETV-6) for diagnosis and prognosis of platelet disorders. Thus, according to what we discussed in this paper, we may take a few steps forward in diagnosis and treatment of these patients; however, further studies are need to reach that goal.

#### References

- 1. Wen Q, Goldenson B, Crispino JD. Normal and malignant megakaryopoiesis. Exp Rev Mol Med 2011;13:32.
- 2. Lou X, Zhang J, Liu S, et al. The other side of the coin: The tumor-suppressive aspect of oncogenes and the oncogenic

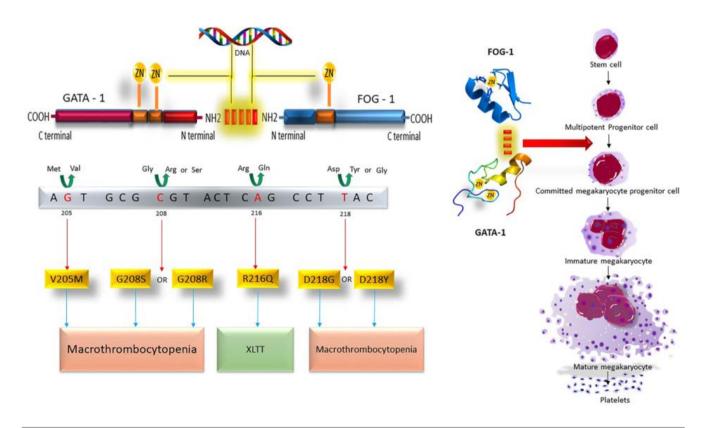


Figure 2. Oncogenes and tumor suppressor genes can up- or downregulate the process of platelet production. Some of them are called oncogenes, which stimulate and increase the production of platelets such as GATA-1, MPL, and FLI-1, while the tumor suppressor genes like P53 and ETV-6 regulate and decrease platelet production. MPL, myelo proliferative leukemia protein; FLI-1, friend leukemia virus integration-1; ETV-6, E-twenty-six variant 6.



aspect of tumor-suppressive genes, such as those along the CCND-CDK4/6-RB axis. Cell Cycle 2014;13:1677-93.

- 3. Kavianpour M, Ahamadzadeh A, Shahrabi S, Saki N. Significance of oncogenes and tumor suppressor genes in AML prognosis. Tumor Biol 2016;37:10041-52.
- Rezaeeyan H, Jaseb K, Alghasi A, et al. Association between gene polymorphisms and clinical features in idiopathic thrombocytopenic purpura patients. Blood Coagul Fibrinol 2017;28:617-22.
- El Aziz SA, El Ghonemy MS, S Aref, et al. Impact of serum immunoglobulins level and IL-18 promoter gene polymorphism among Egyptian patients with idiopathic thrombocytopenic purpura. Hematology 2017;22:99-104.
- Krebs DL, Mielke LA, Metcalf D, et al. A mutation in the translation initiation codon of Gata-1 disrupts megakaryocyte maturation and causes thrombocytopenia. Proc Natl Acad Sci 2006;103:14146-51.
- 7. Del Vecchio GC, Giordani L, De Santis A, De Mattia D. Dyserythropoietic anemia and thrombocytopenia due to a novel mutation in GATA-1. Acta Haematol 2005;114:113-6.
- Yu C, Niakan KK, Matsushita M, et al. X-linked thrombocytopenia with thalassemia from a mutation in the amino finger of GATA-1 affecting DNA binding rather than FOG-1 interaction. Blood 2002;100:2040-5.
- 9. Mehaffey MG, Newton AL, Gandhi MJ, et al. X-linked thrombocytopenia caused by a novel mutation of GATA-1. Blood 2001;98:2681-8.
- Nichols KE, Crispino JD, Poncz M, et al. Familial dyserythropoietic anemia and thrombocytopenia due to an inherited mutation in GATA1. Nat Genet 2000;24:266-70.
- 11. Freson K, Matthijs G, Thys C et al. Different substitutions at residue D218 of the X-linked transcription factor GATA1 lead to altered clinical severity of macrothrombocytopenia and anemia and are associated with variable skewed X inactivation. Hum Mol Genet 2002;11:147-52.
- 12. Freson K, Devriendt K, Matthijs G, et al. Platelet characteristics in patients with X-linked macrothrombocytopenia because of a novel GATA1 mutation. Blood 2001;98:85-92.
- 13. Molecular cloning and characterization of the GATA1 cofactor human FOG1 and assessment of its binding to GATA1 proteins carrying D218 substitutions. Hum Genet 2003;112:42-9.
- White JG, Nichols WL, Steensma DP. Platelet pathology in sex-linked GATA-1 dyserythropoietic macrothrombocytopenia I ultrastructure. Platelets 2007;18:273-83.
- 15. Hughan SC, Senis Y, Best D, et al. Selective impairment of platelet activation to collagen in the absence of GATA1. Blood 2005;105:4369-76.
- 16. Tubman VN, Levine JE, Campagna DR, et al. X-linked gray platelet syndrome due to a GATA1 Arg216Gln mutation. Blood 2007;109:3297-9.
- Kadri Z, Lefevre C, Goupille O, et al. Erythropoietin and IGF-1 signaling synchronize cell proliferation and maturation during erythropoiesis. Genes Develop 2015;29:2603-16.
- Abe M, Suzuki K, Inagaki O, et al. A novel MPL point mutation resulting in thrombopoietin-independent activation. Leukemia 2002;16:1500-6.
- Liu K, Martini M, Rocca B, et al. Evidence fora founder effect of the MPL-S505N mutation in eight Italian pedigrees with hereditary thrombocythemia. Haematologica 2009;94:1368-74.
- 20. Yang Q, Crispino JD, Wen QJ. Kinase signaling and targeted therapy for primary myelofibrosis. Exper Hematol 2017;48:32-8.

- Ballen-Chantelot C, Mosca M, Marty C, et al. Identification of MPL R102p mutation in hereditary thrombocytosis. Front Endocrinol 2017;8:235.
- Vo KK, Jarocha DJ, Lyde RB, et al. FLI1 level during megakaryopoiesis affects thrombopoiesis and platelet biology. Blood 2017;129:3486-94.
- Songdej N, Koneti Rao A. Inherited platelet dysfunction and hematopoietic transcription factor mutations. Platelets 2017;28:20-6.
- 24. Di Paola J. Paris-Trousseau: evidence keeps pointing to FLI1. Blood 2015;126:1973-4.
- 25. Noris P, Valli R, Pecci A, et al. Clonal chromosome anomalies affecting FLI1 mimic inherited thrombocytopenia of the Paris-Trousseau type. Eur J Haematol 2012;89:345-9.
- 26. Stevenson WS, Rabbolini DJ, Beutler L, et al. Paris-Trousseau thrombocytopenia is phenocopied by the autosomal recessive inheritance of a DNA-binding domain mutation in FLI1. Blood 2015;126:2027-30.
- Apostolidis PA, Lindsey S, Miller WM, Papoutsakis ET. Proposed megakaryocytic regulon of p53: the genes engaged to control cell cycle and apoptosis during megakaryocytic differentiation. Physiol Genom 2012;44:638-50.
- Fuhrken PG, Apostolidis PA, Lindsey S, et al. Tumor suppressor protein p53 regulates megakaryocytic polyploidization and apoptosis. J Biol Chem 2008;283:15589-600.
- 29. Bates S, Phillips AC, Clark PA, et al. p14ARF links the tumor suppressors RB and p53. Nature 1998;395:124-5.
- Zhang MY, Churpek JE, Keel SB, et al. Germline ETV6 mutations in familial thrombocytopenia and hematologic malignancy. Nature Genet 2015;47:180-5.
- Poggi M, Canault M, Favier M, et al. Germline variants in ETV6 underlie reduced platelet formation, platelet dysfunction and increased levels of circulating CD34+ progenitors. Haematologica 2017;102:282-94.
- 32. Bianchi E, Bulgarelli J, Ruberti S, et al. MYB controls erythroid versus megakaryocyte lineage fate decision through the miR-486-3p-mediated downregulation of MAF. Cell Death Different 2015;22:1906-21.
- 33. Allen AJ, Gale RE, Harrison CN, et al. Lack of pathogenic mutations in the 5-untranslated region of the thrombopoietin gene in patients with non-familial essential thrombocythemia. Eur J Haematol 2001;67:232-7.
- Guo Y, Niu C, Breslin P, et al. c-Myc-mediated control of cell fate in megakaryocyte-erythrocyte progenitors. Blood 2009;114:2097-106.
- 35. Wang W, Tang Y, Wang Y, et al. LNK/SH2B3 loss of function promotes atherosclerosis and thrombosis. Circul Res 2016;116:308955.
- Eto K, Kunishima S. Linkage between the mechanisms of thrombocytopenia and thrombopoiesis. Blood 2016;127:1234-41.
- Millikan PD, Balamohan SM, Raskind WH, Kacena MA. Inherited thrombocytopenia due to GATA-1 mutations. Semin Thromb Hemost 2011;37:682-9.