



Lower androgen levels promote abnormal cartilage development in female patients with adolescent idiopathic scoliosis

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Background: Adolescent idiopathic scoliosis (AIS) is a disease characterized by changes in the three-dimensional structure of the spine. Studies have shown that the development of AIS might be associated with genetic, biomechanics, endocrine factors and abnormal bone or cartilage development.

Methods: Blood samples collected from 301 female patients (161 females with AIS and 140 females without AIS) were used for genotyping. Forty-eight serum samples from 161 females with AIS and 40 serum samples from 140 females without AIS were subjected to enzyme-linked immunosorbent assays (ELISAs). We also evaluated 32 facet joints (18 females with AIS and 14 females without AIS from the 301 female patients) using immunohistochemistry, Western blotting, and isolation of human primary chondrocytes, among other methods. We treated the AIS primary chondrocytes with dihydrotestosterone (DHT) to verify the relationship among androgen, the androgen receptor (AR), and its downstream pathway proteins.

Results: The serum androgen level in the AIS group was significantly decreased (1.94 ± 0.09 vs. 2.284 ± 0.103) compared with that in the non-AIS (control) group. The single nucleotide polymorphism genotyping results showed that the mutation rates of rs6259 between the AIS and control groups were significantly different (G/G genotype: 48.4% vs. 42.1%, G/A genotype: 40.4% vs. 35.7%, $P < 0.05$). The levels of interleukin (IL)-6 and metalloproteinase (MMP)-13 were increased in the cartilage of AIS patients, and these patients also exhibited decreased AR levels. The cell experiment results showed that androgen reduced the degree of abnormal cartilage development in female AIS patients through the AR/IL-6/signal transducer and activator of transcription 3 (STAT3) signaling pathway.

Conclusions: Our study provides a new perspective on the pathogenesis of AIS and indicates that decreased androgen levels in female AIS patients play a potential role in the development of AIS via the AR/IL-6/STAT3 signaling pathway.

Keywords: Adolescent idiopathic scoliosis (AIS); androgen; interleukin (IL)-6; signal transducer and activator of transcription 3 (STAT3); cartilage

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Introduction

Adolescent idiopathic scoliosis (AIS) is a disease characterized by changes in the three-dimensional structure of the spine. According to epidemiological statistics, the prevalence of scoliosis is between 1% and 4% (1). The pathogenesis of AIS remains unclear, but recent studies have shown that the development of AIS might be associated with genetics (2), hormone and endocrine factors, and biomechanics (3). Many scholars have shown that patients with AIS abnormally secrete many types of hormones, such as estrogen, leptin, and ghrelin (4-6), and changes in the levels of these hormones can induce abnormal bone and cartilage development and might eventually result in AIS. These results suggest that hormone abnormalities might be an important pathogenic factor in the development of AIS.

Clinical presentations of AIS are not evident in childhood. The disease usually develops in adolescence, and many patients experience rapid disease progression. Specifically, the secretion levels of various hormones increase rapidly during adolescence, AIS is more common in females, and the overall ratio of girls to boys with scoliosis is 11:1 (7), indicating that sex hormones might be key factors for the pathogenesis and progression of AIS caused by hormone abnormalities. Many studies have demonstrated that estrogen can promote the development of AIS (4,8). However, few studies are available on the function of androgen in the pathogenesis of AIS. The term androgen is derived from the ancient Greek word “andro”, and androgen is usually defined as a male hormone that is secreted by testicular interstitial cells; however, the adrenal cortex and female ovary can also synthesize a small amount of androgen (9). Because the organs that secrete androgen in males and females are different and the individual contents show wide variations, direct comparisons are not suitable. Furthermore, this disease mainly occurs in female; therefore, only female patients were used as participants in this study. Serum samples from 88 adolescent female patients (48 females with AIS and 40 females without AIS) were subjected to enzyme-linked immunosorbent assays (ELISAs), and the results showed that androgen levels in patients with AIS were decreased compared with those in females without AIS (control), indicating that serum androgen levels might be associated with AIS. Recent studies confirmed that androgen is closely associated with bone metabolism, skeletal muscle growth, and protein synthesis and metabolism. Almeida *et al.* showed that

androgen is involved in trabecular bone maintenance and cortical bone growth (10). The androgen receptor (AR) is extensively distributed throughout the body, and in bone tissue, the AR is mainly distributed in growth sites where proliferation and maturation are promoted during endochondral ossification and in bone remodeling sites, demonstrating that androgen directly acts on cartilage and bone (11). A study conducted by Cicuttini *et al.* indicated that androgen is positively correlated with the male tibial cartilage volume (12). In addition, a study performed by Steffens *et al.* showed that androgen regulates experimental bone loss through the AR (13). Although androgen is closely associated with bone development, the role of androgen in the development of AIS and whether androgen induces abnormal cartilage or bone development in patients with AIS have not been reported. Furthermore, serum androgen levels are affected by many factors. Recent studies confirmed that single nucleotide polymorphism (SNP) loci, including rs12150660, rs727428, rs6259, rs5934505, rs10822184, and rs6258, are associated with serum androgen levels (14-16). Therefore, we enrolled 161 females with AIS and 140 females without AIS and performed genotyping to determine the causes of the reductions in the androgen level observed in patients with AIS.

Our previous studies showed that interleukin (IL)-6 expression is increased in the cartilage of patients with AIS, suggesting that IL-6 might be associated with the development of AIS. A study conducted by Cho DC *et al.* showed that androgen blocks IL-6 to promote cortical bone formation in mice (17), indicating that IL-6 expression in patients with AIS might be correlated with androgen abnormalities. In addition, numerous studies have indicated that IL-6 is closely associated with cartilage abnormalities in patients with osteoarthritis (18-20). As a member of the interleukin family, IL-6 can be secreted by many types of cells and can induce the proliferation and differentiation of many cell types. A study conducted by Yamaguchi *et al.* showed that chondrocytes in patients with hip synovitis secrete IL-6 and stimulate synovial cell proliferation (21). Previous studies have confirmed that IL-6 promotes the differentiation of mesenchymal stem cells into chondrocytes and the self-repair of cartilage through the IL-6/STAT3 pathway (22). However, whether IL-6 is associated with the development and progression of AIS has not been reported. Therefore, this study investigated whether androgen and IL-6 abnormalities induce abnormal cartilage development in patients with AIS.

Table 1 Clinical data of the enrolled female AIS patients

Items	AIS	Control	P value
Number	161	140	–
Age	12.6±3.5	11.2±4.2	0.15
Main curve Cobb angle	37.5±12.4	–	–

Student's *t*-test was used for the statistical analysis, and a *P* value >0.05 indicated no significant difference between the two groups.

Methods

Subjects

All of the specimens were collected from female AIS patients between 10 and 17 years of age and matched patients without AIS (*Table 1*). The criterion used for selection of the AIS patients was a minimum Cobb angle greater than 10, and a clinical assessment was conducted every 3 months. Patients with a congenital spinal deformity, Marfan syndrome, neuromuscular diseases, or spinal deformities caused by other syndromes were excluded from this research. The inclusion criteria for the control group were age-matched non-AIS patients, including healthy individuals and patients with lumbar herniation or spine fracture at our medical examination center. The patients in the control group were subjected to a comprehensive X-ray examination to preclude any spinal deformity.

This study was approved by the Ethics Committee of Xiangya Hospital of Central South University (No. 201703358). Written informed consent was provided by all the subjects and their legal guardians prior to participation in the study, and the study was performed in accordance with the tenets of the Declaration of Helsinki (as revised in 2013) and its amendments.

Isolation and culture of human primary chondrocytes

We collected 32 facet joints from 18 AIS patients and 14 patients without AIS during spinal surgery; the enrolled non-AIS patients were generally patients with spinal fracture or lumbar disc herniation who needed surgery. All of the cartilage was peeled from the facet joints, cut into slices with a thickness of approximately 0.5 mm, washed with phosphate-buffered saline (PBS), and treated with 0.25% trypsin (Gibco, USA) for 30 min and 1 ng/mL collagenase type II (Sigma-Aldrich, USA) for 4 h at 37 °C. The fragment was then plated into 6-cm culture dishes.

All chondrocytes were cultured in DMEM/high-glucose medium (Gibco, USA) supplemented with 10% fetal bovine serum (Gibco, USA) and 1% penicillin/streptomycin (Gibco, USA) at 37 °C in an incubator with 5% CO₂. All the cells were passaged twice before use.

Enzyme-linked immunosorbent assay (ELISA)

The serum androgen level and the secretion of IL-6 from chondrocytes into the cell culture medium were measured using a commercial ELISA kit (Cusabio Biotech, China) in accordance with the manufacturer's recommended protocols.

Genotyping

We used the SQ blood DNA Kit II (Omega Bio-TEK, USA) to extract genomic DNA from peripheral blood, and SNP genotyping was performed using the improved multi-link detection reaction (iMLDR) technique developed by Genesky Biotechnology Company (China). Different allele-specific oligonucleotide probes were used to identify the alleles for each SNP, and different extension lengths at the 3' end were used to further distinguish different SNPs. We set up two negative controls, namely, double-distilled water as the template and a DNA sample without primers, while keeping the other conditions unchanged. Random samples accounting for 5% of the total DNA samples were sequenced using Big Dye-Terminator version 3.1 and an ABI 3730XL automatic sequencer (a biological application system) to confirm the iMLDR results.

Cell proliferation assay

After intervention, chondrocytes were seeded into 96-well plates at a density of 3,000 cells per well. At 12, 24, 48, and 72 h after seeding, cell viability was measured using the Cell Counting Kit-8 (CCK-8) system (Dojindo, Japan) according to the manufacturer's instructions. The results were detected at 450 nm.

siRNA transfection

First, the cells were seeded into 24-well plates at a density of 1×10⁴ cells per well. The cells were then transfected with siRNAs against AR using riboFECTTM CP reagent at final siRNA concentrations of 50 nM following the manufacturer's recommended protocols. The siRNA

sequences used to silence AR expression are shown in [Table S1](#). Forty-eight hours after transfection, the expression level of AR protein was verified by fluorescence microscopy and Western blotting (WB).

Real-time quantitative PCR

The experimental method used for qRT-PCR was based on a previously described method (18). The following primers were used in this study: 18S, CCCTCCAATGGATCCTCGTT (forward) and AGAAACGGCTACCACATCCA (reverse); IL-6, AGGAGACTTGCCTGGTGAAA (forward) and CAGGGGTGGTTATTGCATCT (reverse); and AR, GACGCTTCTACCAGCTCACC (forward) and GCTTCACTGGGTGTGAAAT (reverse).

Western blotting

We collected total protein from chondrocytes using RIPA lysis buffer (CW BIO, China) containing phosphatase inhibitor and PMSE, and the protein was then denatured in a metal bath thermostat. The proteins were separated by 10% SDS-PAGE and then transferred to PVDF membranes (Bio-Rad, Hercules, USA). The membranes were subsequently blocked with 5% skim milk for 1 h and incubated overnight with primary antibodies against GAPDH (1:2,000, # 5174S, RRID:AB_10622025, Cell Signaling Technology, CST, USA), AR (1:200, ab198394, RRID:AB_2861275, Abcam, USA), IL-6 (1:800, ab6672, RRID:AB_2127460, Abcam), metalloproteinase (MMP)-13 (MMP13) (1:800, # 94808, RRID:AB_2800235, CST), STAT3 (1:800, # 12640S, RRID:AB_2629499, CST), and p-STAT3 (1:500, # 9134S, RRID:AB_331589, CST). The membranes were then incubated with secondary antibodies (1:8,000) at room temperature for 1 h. The results were then detected using the Chemiluminescent Protein Detection Module (Thermo Scientific, USA).

Immunohistochemistry (IHC)

The tissues collected during surgery were fixed with formalin for 24 h and decalcified in 10% EDTA for 15 days. The tissues were cut into 5-mm-thick slices, and the paraffinized sections were treated with xylene and rehydrated with a graded alcohol series (100%; 95% \times 2, 85% \times 2, and 70% for 3 min each) and double-distilled water for 5 min. The slices were then pretreated with 3%

hydrogen peroxide for 10 min, blocked with 5% bovine serum albumin (BSA) for 30 min, and washed with PBS. The sections were incubated with primary antibodies against AR (1:200, ab198394, RRID:AB_2861275, Abcam) and IL-6 (1:800, ab6672, RRID:AB_2127460, Abcam) overnight at 4 °C and subsequently with biotinylated goat anti-rabbit immunoglobulins for 30 min and a streptavidin-horseradish peroxidase solution (ZSGBBIO, China) containing the secondary antibody for 45 min. The sections were then stained with 3,30-diaminobenzidine (DAB) for 30 s and counterstained with hematoxylin (Servicebio, China) for 1 min. Pictures were acquired via using a microscope (Leica, Germany). The results were detected quantitatively via using ImageJ software (Bethesda, USA).

Immunofluorescent staining

AIS primary chondrocytes after incubation with IL-6 were first washed with PBS, fixed with 4% paraformaldehyde for 25 min, incubated with 0.5% Triton X-100 for 10 min, and then blocked with 5% BSA in PBS for 30 min. Chondrocytes were then incubated overnight with primary antibody against Ki67 (1:500; # 9449; RRID: AB_2797703; CST) at 4 °C. After washing with PBS, the chondrocytes were incubated with goat anti-rabbit secondary antibody and Alexa Fluor 488-conjugated antibody (1:300; Sigma, USA) for 1 h. Finally, the cells were incubated with DAPI (Solarbio, China) for 5 min. Pictures were acquired by immunofluorescence microscopy (Leica, Germany). The results were detected quantitatively using ImageJ software (Bethesda, USA).

Statistics

The data obtained in the genetic association study were analyzed by the Chi-square (and Fisher's exact) test. One-way ANOVA and Student's *t*-test were used to analyze the remaining data, and the quantitative data are expressed as the means \pm standard deviations (SDs). All analyses were performed using GraphPad Prism software (GraphPad Prism Software, USA), and P values <0.05 indicated statistical significance.

Results

Serum levels of androgen in females with AIS and females without AIS

To detect the serum level of androgen in AIS patients, we

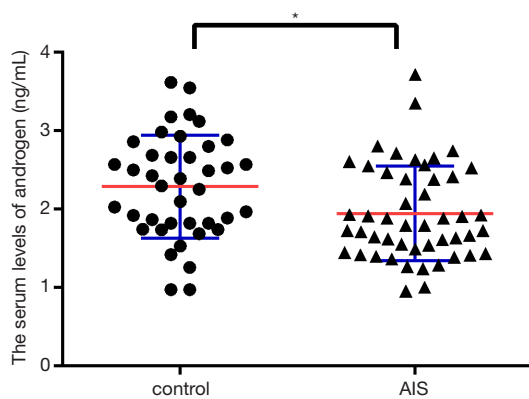


Figure 1 Serum levels of androgen in female AIS and non-AIS patients. The serum androgen level in the AIS group was lower than that in the control group (1.94 ± 0.09 vs. 2.284 ± 0.103). *indicates a significant difference compared with the control group ($P < 0.05$).

analyzed serum samples from 48 females with AIS and 40 age-matched females without AIS by ELISA. The clinical information for all the patients is provided in *Table 1*. No differences in age or body weight were found between the AIS and control groups. The ELISA results showed that the serum androgen level in the AIS group was lower than that in the control group (1.94 ± 0.09 vs. 2.284 ± 0.103) (*Figure 1*).

SNP genotype frequency distributions in AIS and non-AIS patients

We genotyped eight SNPs that were previously shown to be related to the serum androgen levels. Specifically, a total of 161 AIS patients and 140 controls were successfully genotyped and subjected to statistical analysis. The eight genotyped SNPs in the two groups were analyzed using the Chi-square (and Fisher's exact) test. The genotype distribution frequencies are listed in *Table 2*.

A significant difference in the presence of rs6259 was detected between the two groups ($P < 0.05$). Overall, the frequencies of the G/G and G/A genotypes in the AIS group were significantly higher than those in the non-AIS group (G/G genotype: 48.4% vs. 42.1%, G/A genotype: 40.4% vs. 35.7%, and A/A genotype: 11.2% vs. 22.2%; $P < 0.05$).

Increased IL-6 expression and reduced AR expression in the cartilage of AIS patients

To confirm that androgen expression is associated with

cytokines and proteins in the cartilage tissue of patients with AIS, the cartilage tissue separated from the facet joint tissue of the patients during surgery was analyzed by WB and IHC. WB analysis of total proteins extracted from the tissues showed that the cartilage of patients with AIS exhibited higher protein expression levels of IL-6 and MMP-13 and a lower protein expression level of the AR compared with the cartilage of non-AIS patients (*Figure 2A,B*). In addition, the IHC results showed that patients with AIS exhibited increased IL-6 expression and decreased AR expression in cartilage compared with the control group (*Figure 2C,D,E,F*).

Androgen inhibits the secretion of IL-6 in AIS primary chondrocytes

To confirm the effect of androgen on primary chondrocytes from patients with AIS, AIS primary chondrocytes were treated with 50 nM dihydrotestosterone (DHT), whereas the control group was treated with PBS. After 48 h of treatment, the culture medium from the chondrocytes was collected for ELISA, and the results showed that the IL-6 level in the culture medium after DHT treatment was decreased compared with that in the control group, indicating that androgen inhibited the secretion of IL-6 from chondrocytes (*Figure 3A*). Total RNA and protein were extracted from AIS primary chondrocytes. PCR analysis showed that the mRNA level of IL-6 was decreased (*Figure 3B,C*), and WB revealed a decreased IL-6 protein level and increased AR expression (*Figure 3D,E,F*). Furthermore, the expression of MMP13 did not significantly change after androgen treatment (*Figures S1,S2*).

The function of androgen in blocking IL-6 expression requires AR activation

We further investigated the mechanism underlying the regulation of IL-6 expression by androgen. Therefore, we transfected AIS primary chondrocytes with siAR or siNC for 48 h and then treated the chondrocytes with 50 nM DHT or PBS (control group). The siRNA transfection efficiency was verified by fluorescence microscopy and WB (*Figures S3,S4*). After treatment, total RNA and protein were collected, and WB analysis showed that IL-6 expression in the DHT + siAR group was significantly increased compared with that in the DHT + siNC group and was significantly different from that in the control group (*Figure 4A,B*). In addition, although the mRNA level

Table 2 Frequency distributions of allele SNPs

Single nucleotide polymorphism	Group AIS number (%)	Group control number (%)	P value (AIS vs. control)
rs6259 (SHBG)			
G/G	78 (48.4)	59 (42.1)	0.03
G/A	65 (40.4)	50 (35.7)	
A/A	18 (11.2)	31 (22.2)	
rs10822184 (REEP3)			
C/C	20 (12.4)	19 (13.6)	0.85
C/T	88 (54.7)	72 (51.4)	
T/T	53 (32.9)	49 (35.0)	
rs12150660 (SHBG)			
G/G	57 (35.4)	46 (32.9)	0.89
G/A	79 (49.1)	72 (51.4)	
A/A	25 (15.5)	22 (15.7)	
rs11642015 (FTO)			
C/C	121 (75.2)	101 (72.2)	0.48
C/T	39 (24.2)	36 (25.7)	
T/T	1 (0.6)	3 (2.1)	
rs266719 (EIF4A2)			
C/C	133 (82.6)	112 (80.0)	0.51
C/T	28 (17.4)	27 (19.3)	
T/T	0	1 (0.7)	
rs5934505 (FAM9B)			
C/C	25 (15.5)	24 (17.1)	0.77
C/T	35 (21.7)	26 (18.6)	
T/T	101 (62.8)	90 (64.3)	
rs7639352 (FTO)			
C/C	75 (46.6)	73 (52.1)	0.62
C/T	75 (46.6)	59 (42.2)	
T/T	11 (6.8)	8 (5.7)	
rs1799941 (SHBG)			
G/G	160 (99.3)	140 (100.0)	1
G/A	1 (0.7)	0	
A/A	0	0	

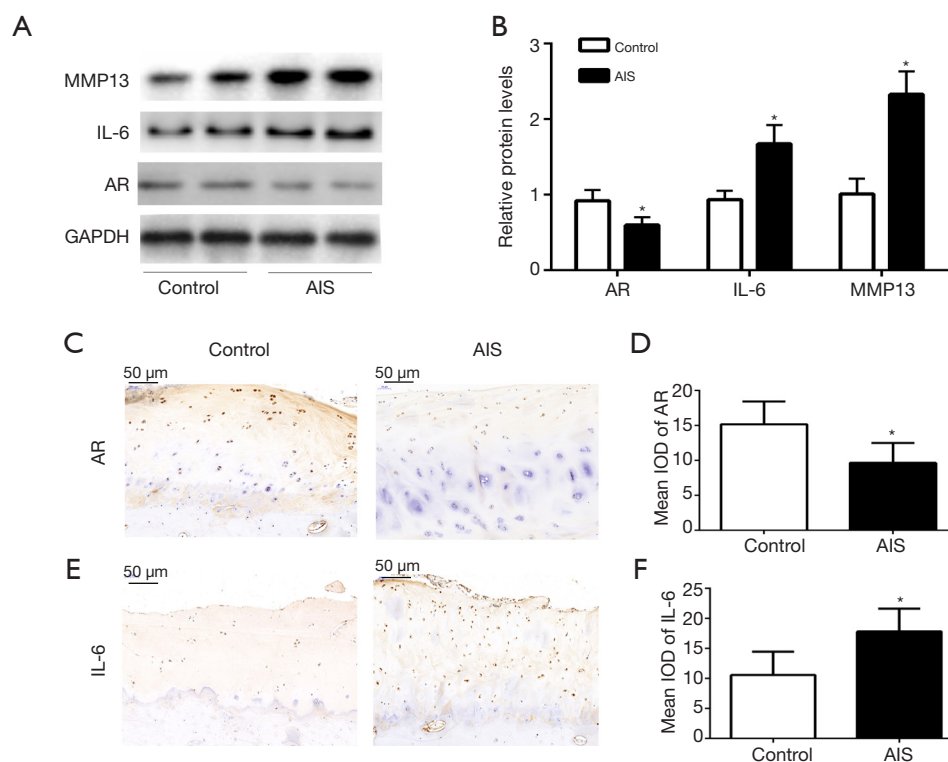


Figure 2 Increased IL-6 expression and reduced AR expression in the cartilage of AIS patients. (A,B) The protein levels of AR, IL-6, and MMP13 in the cartilage of female AIS and non-AIS patients. (C,D,E,F) AR and IL-6 levels detected by IHC in the cartilage of AIS and non-AIS patients; scale bar: 50 μ m. The data are shown as the means \pm SDs. *indicates $P < 0.05$ vs. the control group.

of IL-6 in the siAR-transfected group was not significantly different from that in the siNC group, its expression level was higher than that in the siNC group (Figure 4C). These results indicate that the function of androgen in blocking IL-6 expression requires AR activation.

Promotion of MMP13 expression and inhibition of chondrocyte proliferation by IL-6

To study the effect of IL-6 overexpression on cartilage tissue, primary chondrocytes from patients with AIS were directly treated with 20 nM IL-6 or PBS (control group). Total protein from the chondrocytes was collected for WB detection, and the results showed that the protein expression of MMP13 in chondrocytes was significantly increased after IL-6 treatment (Figure 5A,B). In addition, chondrocyte proliferation was assessed with Ki67 12, 24, 48, and 72 h after IL-6 treatment using the CCK-8 assay, and the results showed that the proliferation ability of chondrocytes from AIS patients decreased after treating with 20 nM IL-6 for 24 h (Figure 5C). Furthermore, the proliferation of primary

cells from both the AIS and non-AIS groups was assessed. Interestingly, the proliferation rate obtained for the AIS group was slightly slower than that found for the non-AIS group, but the results were not significantly different (Figure 5D). Ki67 immunofluorescence staining was also used to determine AIS chondrocyte proliferation after IL-6 intervention and the chondrocytes proliferation in cartilage from AIS and non-AIS patients. The results showed that IL-6 reduced the proliferation ability of chondrocytes (Figure 5E,F), but tissue staining results showed that there was no significant difference in chondrocytes proliferation between AIS patients and non-AIS patients (Figure S5).

Effects of IL-6 on chondrocytes from AIS patients mediated by STAT3 signaling pathway proteins

Our results showed that the phosphorylation level of STAT3 protein in chondrocytes from AIS patients was higher than that in chondrocytes from non-AIS patients (Figure 6A,B). STAT3 is the most specific downstream protein in the IL-6 pathway. To confirm that the effect of IL-6 on chondrocytes

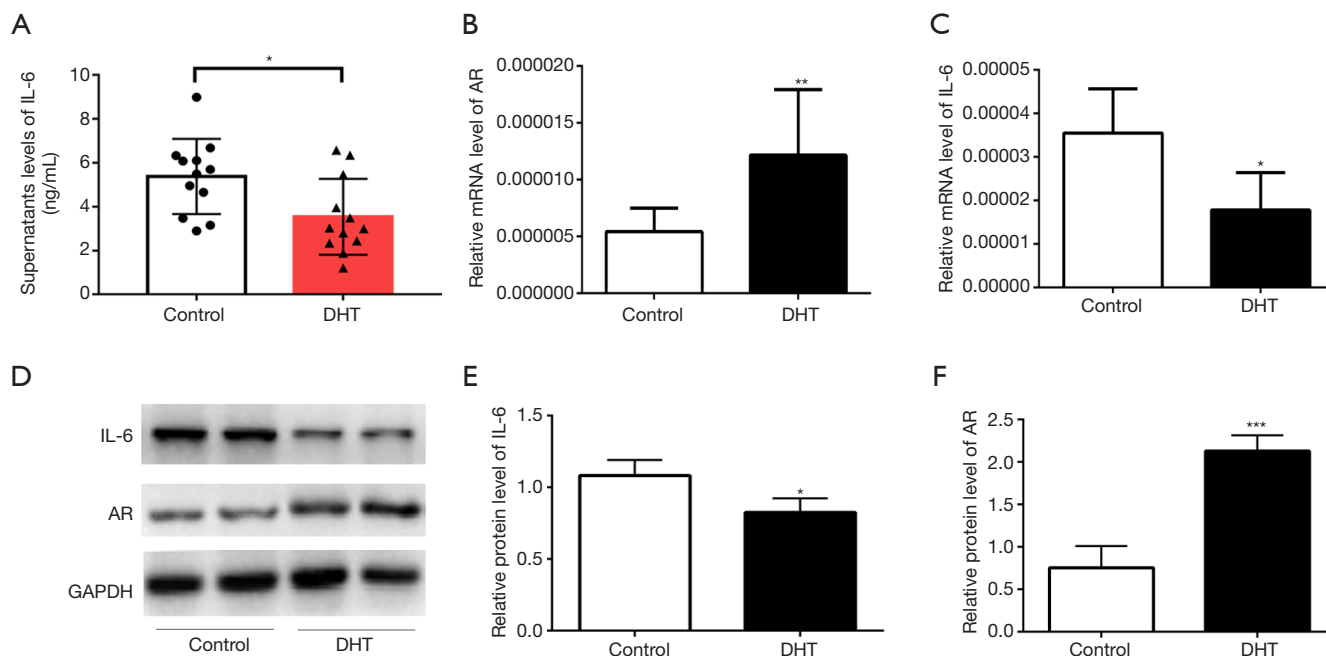


Figure 3 Androgen inhibited the secretion of IL-6 in AIS primary chondrocytes. (A) The IL-6 level in the AIS primary chondrocyte culture medium after DHT treatment. (B,C) The relative mRNA levels of AR and IL-6 in AIS primary chondrocytes after the DHT intervention. (D,E,F) The protein levels of AR and IL-6 in AIS primary chondrocytes after DHT treatment. The data are shown as the means ± SDs. *indicates $P < 0.05$ vs. the control group.

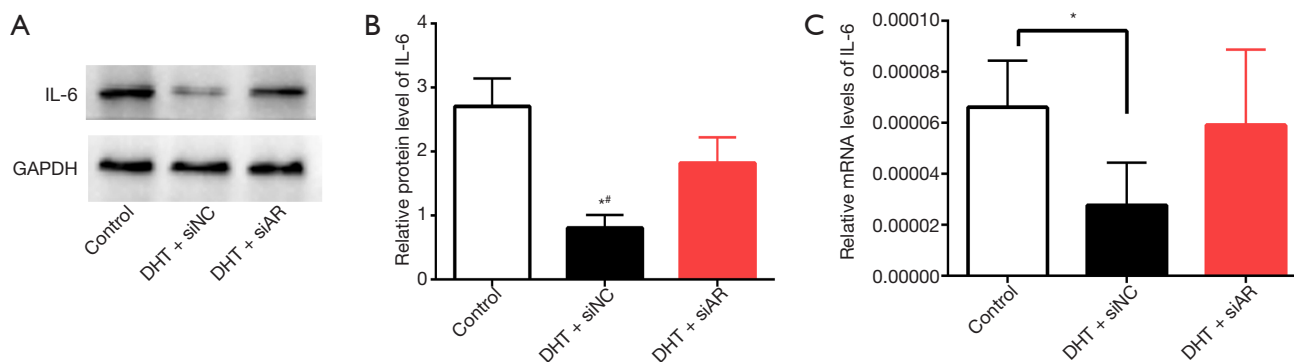


Figure 4 (A,B) The protein level of IL-6 in AIS primary chondrocytes after transfection. (C) The relative mRNA level of IL-6 in AIS primary chondrocytes after transfection. The data are shown as the means ± SDs. *indicates $P < 0.05$ vs. the control group; #indicates $P < 0.05$ vs. the DHT + siAR group.

from AIS patients was achieved through activation of STAT3 protein, we cultured AIS primary chondrocytes with Stattic (MCE, USA), a nonpeptidic small molecule that specifically inhibits STAT3. The results showed that the level of phosphorylated STAT3 protein increased after treating with 20 nM IL-6 for 48 h, and the expression of

the STAT3 downstream protein MMP13 also increased. However, STAT3 phosphorylation was inhibited in the IL-6 + Stattic group, and accordingly, the protein expression of MMP13 was also decreased in this group (Figure 6C,D). These results indicate that the regulatory function of IL-6 on chondrocytes is achieved through STAT3 protein

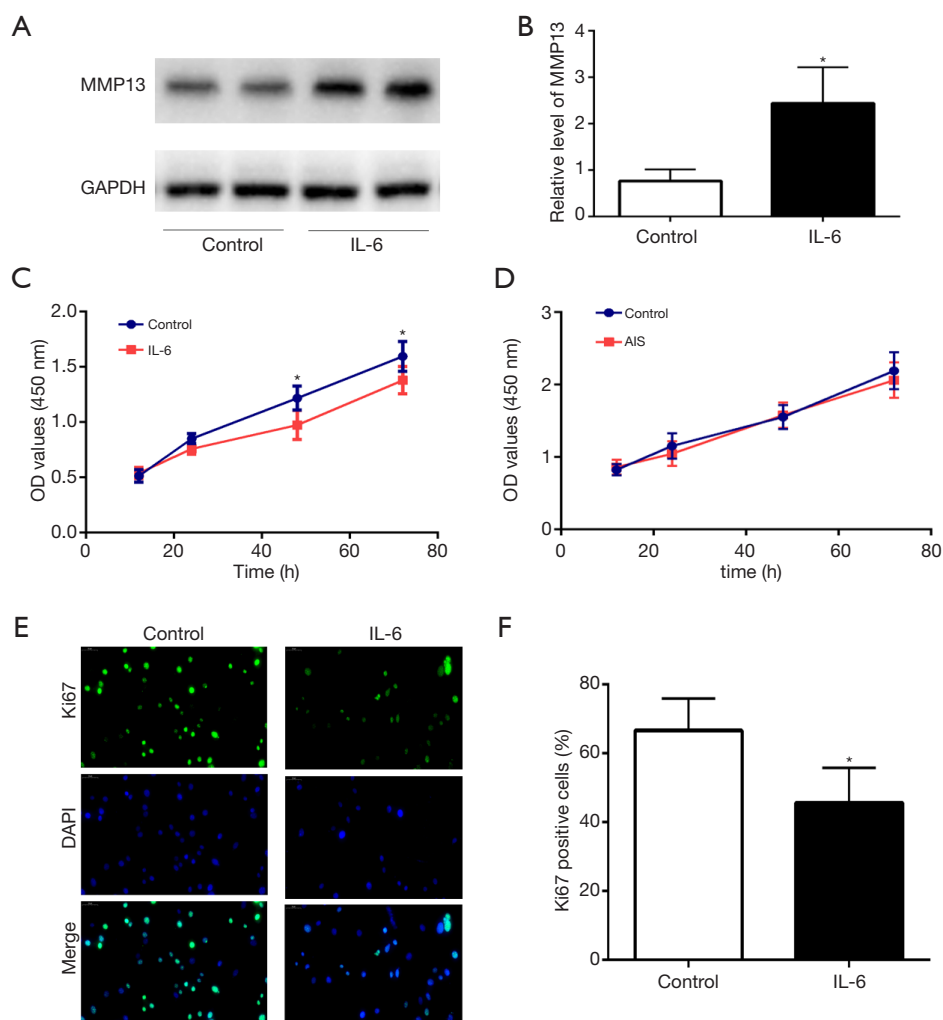


Figure 5 Promotion of MMP13 protein expression and inhibition of chondrocyte proliferation by IL-6. (A,B) The protein level of MMP13 in AIS primary chondrocytes after the intervention. (C) Chondrocyte proliferation was measured 12, 24, 48, and 72 h after treatment. (D) The proliferation of chondrocytes from AIS patients and non-AIS patients. (E,F) The proliferation of AIS primary chondrocytes after treatment with 20 nM IL-6 for 48 h, green signal, 400 \times . The data are shown as the means \pm SDs. *indicates $P < 0.05$ vs. the control group.

activation.

Discussion

AIS causes three-dimensional deformation of the spine. Although its pathogenesis remains unclear, numerous experiments have suggested that AIS development is closely associated with genetic factors (2,23) and hormone secretion (4,24). In addition, previous studies have confirmed that chondrocyte activity on the AIS patients and non-AIS patients is different (25) and that endochondral ossification is a very important process in the human body.

These factors might cause unbalanced growth between the convex and concave sides of the spine, which would eventually result in deformity (26). Therefore, we collected facet joint tissue during surgery for IHC and WB analyses, and the results showed that AR expression and the levels of IL-6, MMP9 and MMP13 were significantly increased in the cartilage tissue of AIS patients, which might be important factors inducing abnormal development or chondrocyte degradation in patients with AIS.

High levels of a number of cellular and soluble factors, including inflammatory cytokines, such as IL-6, are present in AIS cartilage (18). IL-6 causes cartilage damage by acting

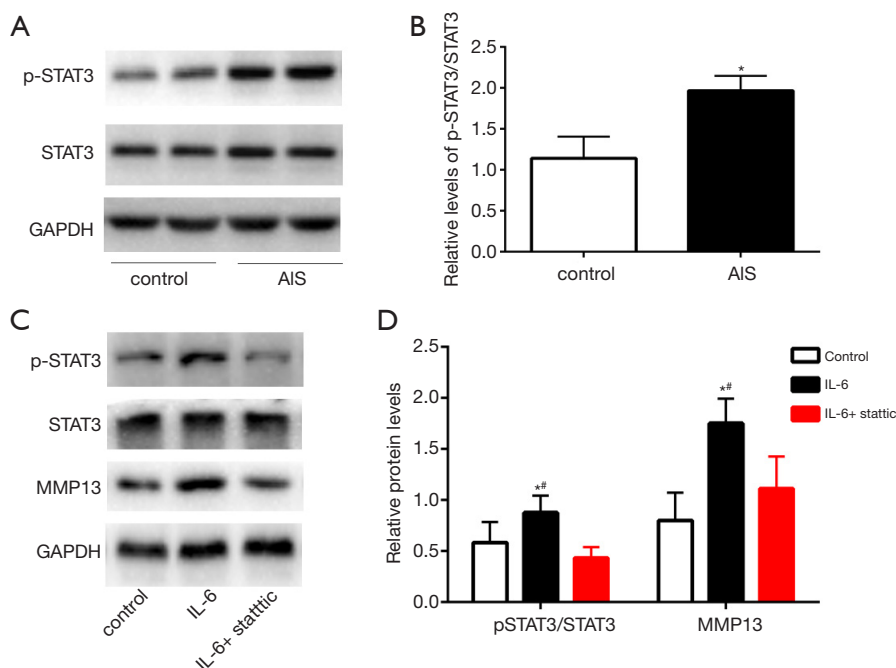


Figure 6 Effects of IL-6 on chondrocytes from AIS patients mediated by STAT3 signaling pathway proteins. Total and phosphor-STAT3 protein levels in AIS and non-AIS patients (A,B). Total, phosphor-STAT3, and MMP13 protein levels in AIS primary chondrocytes after the intervention (C,D). The data are shown as the means \pm SDs. *indicates $P < 0.05$ vs. the control group; #indicates $P < 0.05$ vs. the IL-6 + Stattic group.

on both anabolic and catabolic mechanisms of cartilage physiology (27). After the initial finding of abnormal IL-6 levels in cartilage tissue, these results were further validated through cell culture experiments. In this research, primary chondrocytes from AIS patients were stimulated with 20 nM IL-6, and the results showed that IL-6 promoted MMP13 protein expression and inhibited chondrocyte proliferation. IL-6 was previously found to be a crucial mediator of the MMP13 levels in chondrocytes (28), and other studies have shown that IL-6 inhibits the expression of type II collagen (29). The most specific downstream protein in the IL-6 pathway is STAT3, and numerous studies have shown that IL-6 activates STAT3 protein in chondrocytes to regulate the expression levels of downstream proteins (22,30,31). Our results also showed that IL-6 exerted its function in chondrocytes from patients with AIS through STAT3 protein activation. IL-6 and MMP13 are usually associated with cartilage catabolism, Ryu *et al.* found that IL-6 induced experimental osteoarthritis cartilage destruction in mice via regulation of MMP13 levels (32). Bouaziz *et al.* indicated the lack of MMP13 protects the articular cartilage from degradation

and osteoarthritis in mice (33). Therefore, the detected increase in IL-6 expression in patients with AIS might induce an inflammatory reaction in chondrocytes to accelerate degradation and thus induced unbalanced vertebral growth.

Previous studies have confirmed that androgen can promote cortical bone formation through IL-6, which indicated that androgen plays a very important regulatory role in bone formation and growth (17). A study conducted by ElBaradie *et al.* demonstrated that DHT regulates growth plate chondrocytes in a sex-specific manner via rapid signaling pathways (34). In an earlier study, Raz *et al.* found that rat costochondral growth plate chondrocytes exhibit sex-specific and cell maturation-dependent responses to testosterone. In addition, these researchers proposed that the specific response of chondrocytes to hormones requires further metabolism of testosterone to DHT (35). Therefore, in our study, primary chondrocytes from AIS patients were treated with 5×10^{-7} M DHT, and the results showed that androgen played an important role in cartilage through IL-6. Specifically, DHT inhibited IL-6 expression through AR activation in chondrocytes of AIS patients. In

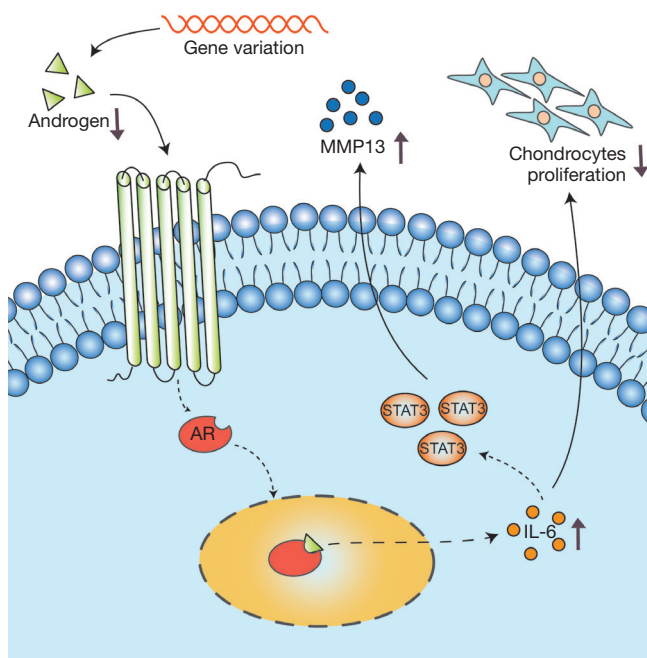


Figure 7 Schematic diagram of androgen in the abnormal development of cartilage in female AIS patients.

addition, the serum androgen levels in patients with AIS were lower than those in the non-AIS group, which resulted in a weakened inhibitory function of androgen on IL-6 in patients with AIS. These results suggest that androgen might play a protective role in the development of AIS to inhibit IL-6-induced abnormal chondrocyte development.

We investigated the reasons for the lower serum androgen levels in patients with AIS. A previous study on the relationship between estrogen and AIS showed that patients with AIS have low serum androgen levels in the body, but no further studies have been conducted (8). In addition, Raczkowski found increased testosterone levels above the reference values in girls with AIS, though that study only included 27 samples in the AIS group and seven samples in the control group, which can easily lead to errors in the experiment (36). Based on studies of the gene sequence of androgen, we speculated that many allelic SNPs might affect the androgen secretion levels. Recent studies confirmed that SNP loci, including rs12150660, rs727428, rs6259, rs5934505, and rs10822184, are associated with androgen levels (14,15). Therefore, 161 females with AIS and 140 females without AIS were recruited for genotyping. The results showed that the mutation rates of rs6259 were significantly different between the AIS and

non-AIS groups, and the G/G and G/A allelic mutation rates were significantly higher in the AIS group (G/G genotype: 48.4% *vs.* 42.1%, G/A genotype: 40.4% *vs.* 35.7%, and A/A genotype: 11.2% *vs.* 22.2%; $P < 0.05$). A study performed by Martínez-García MÁ *et al.* showed that rs6259 is a nonsynonymous mutation, and this SNP reduces the expression of androgen through an increase in the sex hormone-binding globulin gene (SHBG) (16), which is consistent with our results. Many studies have also revealed that rs6259 can increase the risk of polycystic ovary syndrome (37), diabetes mellitus (38), and metastatic prostate cancer (39), among others, but no research on the relationship between rs6259 and AIS has been reported.

This study also has some limitations. First, we could not confirm whether androgen was the most important factor in AIS development. Androgen might affect abnormal cartilage development in patients with AIS in combination with other hormones or factors. Second, the chondrocytes extracted in this study were all obtained from the cartilage tissue of the facet joint of the spine, which is not representative of the different cartilage tissues throughout the body. Third, the experimental samples included in this study were limited and cannot fully reflect differences in sex, Cobb angle, and race.

In conclusion, this study provides a new perspective for studying the pathogenesis of AIS. Our results show that androgen can reduce abnormal cartilage development in patients with AIS through AR/IL-6/STAT3 pathway inhibition (*Figure 7*). The data suggest that decreased androgen levels in female AIS patients might play a potential role in the occurrence of AIS.

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Footnote

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Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/atm-20-3171>). The authors have no conflicts of interest to declare.

Ethical Statement: All authors are accountable for all

aspects of the work in ensuring that questions related to the integrity or accuracy of the work are appropriately investigated and resolved. This study was approved by the Ethics Committee of Xiangya Hospital of Central South University (No. 201703358). Written informed consent was provided by all the subjects and their legal guardians prior to participation in the study, and the study was performed in accordance with the tenets of the Declaration of Helsinki (as revised in 2013) and its amendments.

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