

# **Techniques of assessing small airways dysfunction**

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The small airways are defined as those less than 2 mm in diameter. They are a major site of pathology in many lung diseases, not least chronic obstructive pulmonary disease (COPD) and asthma. The small airways are frequently involved early in the course of these diseases, with significant pathology demonstrable often before the onset of symptoms or changes in spirometry and imaging. Despite their importance, they have proven relatively difficult to study. This is in part due to their relative inaccessibility to biopsy and their small size which makes their imaging difficult. Traditional lung function tests may only become abnormal once there is a significant burden of disease within them. This has led to the term 'the quiet zone' of the lung. In recent years, more specialised tests have been developed which may detect these changes earlier, perhaps offering the possibility of earlier diagnosis and intervention. These tests are now moving from the realms of clinical research laboratories into routine clinical practice and are increasingly useful in the diagnosis and monitoring of respiratory diseases. This article gives an overview of small airways physiology and some of the routine and more advanced tests of airway function.

Keywords: chronic obstructive pulmonary disease; asthma; lung function; small airways; impulse oscillometry; multiple breath nitrogen washout; imaging

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The airways consist of approximately 23 generations of dichotomously branching tubes from the trachea to the alveoli (1) (Fig. 1). The main function of the airways is to ventilate the gas exchanging units of the lung. They also play a role in the conditioning of inhaled air, removal of particulate matter, and immune defence within the lung.

The first 15 generations of airways are called the conducting airways and take no part in gas exchange. They constitute the anatomical dead space, which is approximately 100–150 ml in a human adult (2). Beyond this region lie the respiratory bronchioles which have occasional alveoli budding from them. These continue to divide until they reach the alveolar sacs with a total surface area of 70–80 m<sup>2</sup> (3). These airways take part in gas exchange and comprise the acinar airways.

The small airways refer to those airways less than 2 mm in diameter (4). These occur from approximately generation 8 and include a portion of the conducting airways as well as all the acinar airways. They have important structural and physiological differences from large airways. First, they lack the cartilaginous support seen in large airways and lack mucous glands. They are lined by surfactant which reduces surface tension and helps prevent them from closing on expiration and at low lung volumes (5).

Throughout successive airway generations, there is a reduction in the length and diameter of the airway. Because of the exponential increase in airway numbers, there is a rapid increase in cross-sectional area with each subsequent generation. This has two major effects on airway physiology. First, for any given flow, the velocity of gas transit within the lung decreases with increasing airway generation. The result of this is high velocity flow in the proximal airways which is turbulent and hence density dependent. In the small airways of the lung, flow is laminar and therefore independent of gas density (6). At the interface of the conducting and acinar airways, there is a change from bulk convective flow to diffusion down a concentration gradient. However the distance for diffusion is small, approximately 0.2 mm (7). Second, the resistance to airflow in the small airways is low in health, comprising between 10 and 25% of total airways resistance (8, 9). However, small airways resistance is significantly increased in disease (10). Small airways resistance is largely independent of lung volume whilst large airways resistance is altered significantly with change in lung volumes (8). These arrangements in the human lung help to achieve as equitable ventilation to lung units as possible, whilst maintaining low airflow resistance and minimal work of breathing.

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Fig. 1. Airway generations (adapted from ref. 1).

#### The small airways in disease

Both in chronic obstructive pulmonary disease (COPD) and asthma, the small airways have been shown to be the major site of airflow obstruction (9, 11, 12). The small airways may be more prone to pathology because of their size. Small inhaled particles and pathogens may be deposited here and pathological changes in airways disease make the small airways susceptible to occlusion. Therefore, small airways may require inhaled therapeutic aerosols of smaller size to be able to penetrate the airways tree and reach the distal lung region (13). Pouseille's law states that the resistance to flow is inversely proportional to the fourth power of the radius. Hence, airway obstruction can have profound effects on lung physiology. The obstruction of small airways can occur through a number of mechanisms, including luminal occlusion by mucus, reduction in luminal diameter from inflammatory infiltrates, smooth muscle hypertrophy, or airway wall thickening. In addition, loss of structural airway supports may enhance collapsibility of airways.

#### Asthma

In asthma, the small airways are thickened with a chronic inflammatory infiltrate affecting all layers of the airway (14). Inflammatory changes are present throughout the

airways, although differences in the extent and composition of the inflammatory infiltrate exist between large and small airways. The small airways are the major site of inflammation in asthma (15, 16) with a chronic inflammatory infiltrate consisting of eosinophils, Tlymphocytes, neutrophils, and macrophages. In addition, there is smooth muscle thickening and luminal occlusion by mucus (17–19). In small airways, the density of the lymphocytes and eosinophils is greater in the outer walls compared to large airways where more central airway wall inflammation predominates (15, 20). Mast cells are found more commonly in the periphery of the lung (21) than the central airways and more marked neutrophilic inflammation may be seen in the peribronchiolar lung parenchyma in fatal asthma (15). The severity of inflammatory changes correlates with lung function in nocturnal asthma (22), severe asthma (21), and is more marked in patients with fatal asthma compared to non-fatal asthma (23).

#### Chronic obstructive pulmonary disease

COPD is characterised predominately by neutrophilic and lymphocytic small airway infiltration along with the presence of (24–26). Lymphocytic infiltration and smooth muscle hypertrophy are more prominent in COPD than in asymptomatic smokers (25). In addition, there is airway remodelling with peribronchial fibrosis, smooth muscle hypertrophy, and luminal occlusion from mucus (27, 28). The extent of airway inflammation correlates with disease severity in COPD (24, 29, 30). However, it is airway wall thickness, rather than the severity of inflammatory changes, that is more strongly associated with disease progression in COPD (30). This suggests that regulation of the remodelling pathways through tissue growth factors may be altered in susceptible patients. Interestingly, smoking has been shown to increase tissue levels of growth factors that promote airway remodelling prior to the onset of inflammatory changes (31). Emphysematous destruction of lung tissue may also affect the small airways by disruption of the elastic fibres supporting airway walls. The extent of airway inflammation correlates with the degree of disruption (32) suggesting that peribronchiolar inflammation may drive the proteasemediated disruption of airway attachments. Indeed, small airways disease may precede emphysematous changes identified by computed tomography (CT) (33).

Inflammatory small airways disease may exacerbate small airways injury and dysfunction through mechanical stresses of cyclic opening and closing of airways during tidal breathing.

#### Physiological assessment of the small airways

Small airways obstruction may lead to a reduction in airflow, increased airways resistance, gas trapping, and inhomogeneity of ventilation. Consequently, physiological tests measuring these variables can detect and quantify small airways disease (34). Table 1 summarises the techniques available for the assessment of small airways disease.

#### Spirometry

Spirometry is the most widely used lung function test both in the diagnosis and stratification of severity of lung disease. A diagnosis of obstructive lung disease is made when the ratio of the Forced Expiratory Volume in 1 sec (FEV<sub>1</sub>) to Forced Vital Capacity (FVC) is less than 70% (35). Whilst a reduction in FEV<sub>1</sub> may reflect airflow obstruction, it is also dependent on lung volumes, elastic recoil, respiratory muscle strength, and patient effort (36). In health, the main site of airways resistance occurs in the 4th–8th airway generations. Thus, FEV<sub>1</sub> largely reflects large airways obstruction, and a significant amount of small airways disease must accumulate before FEV<sub>1</sub> becomes abnormal.

Examination of the mid-portion of expiratory flow may offer more information on small airway pathology. The Forced Expiratory Flow between 25 and 75% of the FVC (FEF<sub>25-75</sub>) is one of the most commonly cited measures of small airways pathology. McFadden and Linden postulated that the latter part of the vital capacity was affected by increased resistance in small airways as lung volume fell. Pathology in these airways causes excessive airway narrowing and collapse at an earlier time and closer to the alveolus during exhalation. This results in a reduction in the maximum expiratory flow that can be achieved (37). However,  $FEF_{25-75}$  is dependent on the FVC and therefore changes in FVC will affect the portion of the flow-volume curve examined. If FEF<sub>25-75</sub> is not adjusted for lung volume, there is poor reproducibility (38). Another disadvantage is the sensitivity of the FEF<sub>25-75</sub>, as it is frequently normal if the FEV<sub>1</sub>/FVC ratio is >75% (39). In addition, there is poor correlation with other markers of small airways disease such as gas trapping (40) and histological evidence of small airways inflammation (41). The Forced Expiratory Volume in 3 sec (FEV<sub>3</sub>) to FVC ratio has been suggested as an alternative measure of small airways disease. The fraction of air not expired in the first 3 sec (1-FEV<sub>3</sub>/FVC) is also calculated to estimate the growing proportion of long time constant lung units. As FEV<sub>1</sub>/FVC falls, the FEV<sub>3</sub>/ FVC falls and the 1-FEV<sub>3</sub>/FVC rises. These measures have a better accuracy than FEF<sub>25-75</sub>, particularly in advancing age (42).

Gibbons et al. (43) suggested that the change in FVC following a histamine provocation is a better measure of small airway dysfunction in asthmatic patients than the fall in FEV<sub>1</sub>. A fall in FVC suggests small airway closure and gas trapping. Other spirometric markers that have been suggessted for assessment of small airways disease have included the ratio of the FVC to slow vital capacity (SVC) (44).

#### Plethysmography

Plethysmographic assessment of lung volumes provides a sensitive measure of gas trapping and lung hyperinflation. Hyperinflation may be defined as an abnormal elevation of lung volumes at the end of expiration (45). It is a function of airflow limitation, lung elastic recoil, and chest wall compliance. Airway narrowing results in a prolonged time constant for expiration, and airways may close resulting in gas trapping. The residual volume (RV) is an important measure of small airways dysfunction and may be raised before the onset of abnormal spirometry in asthma (46, 47). The RV correlates with the degree of inflammatory changes in small airways in COPD (24) and with peripheral airway resistance in asthma (48). Indeed, improvement in asthma symptoms following treatment with monteleukast correlated with the reduction in RV but not spirometric parameters (49).

The residual volume/total lung capacity (RV/TLC) ratio may be a more useful marker of gas trapping as the TLC is frequently raised in obstructive lung disease. Sorkness et al. demonstrated that the RV/TLC ratio is higher in patients with severe asthma compared to non-severe

	Measures	Pros	Cons
Lung function			
Spirometry	FEV <sub>1</sub> , FEF <sub>25-75</sub> , FEV <sub>1</sub> /FVC, FEV <sub>3</sub> /FVC, FEV/SVC	Widely available Reproducible Standardised criteria	Relatively insensitive to early disease and subtle changes Effort dependent Not specific to small airways changes
Plethysmography	RV, RV/TLC, airways resistance	Widely available Reproducible Relatively easy to perform Sensitive to early change	Not specific for small airways disease Effort dependent Relatively time consuming
IOS	Z, R <sub>rs</sub> , X <sub>rs</sub>	Non-invasive and easy to perform Effort independent Reproducible Intra-breath analysis	Equipment not widely available Interference from swallowing and upper airway artefact
Inert gas washout	Closing capacity and closing volume Phase III slope: S <sub>III</sub> , S <sub>acin</sub> , S <sub>cond</sub>	Sensitive to early change Can distinguish between distal and proximal airways disease	Difficult to perform, requiring specialist equipment Restricted to research settings
Exhaled nitric oxide	FE <sub>NO</sub>	Easy and quick to perform Hand-held analysers available Sensitive to changes with treatment in asthma	Unclear role in COPD Affected by smoking status
Imaging			
High resolution computed tomography	Assessment of airway changes Assessment of gas trapping (MLD <sub>E/I</sub> )	Widely available Quick and easy to perform	Unable to visualise small airways directly Specialist software may be required No standardised measurements Radiation dose
Hyperpolarised magnetic resonance imaging	Apparent diffusion co-efficient Regional ventilation defects	Allows assessment of heterogeneity in distribution of disease No radiation dose	Expensive Limited to research applications
Nuclear medicine (scintigraphy, SPECT, and PET)	Ventilation Inhaled drug or receptor distribution	Allows assessment of heterogeneity in distribution of disease Can help target drugs to site of lung Can be tailored to study individual drugs or receptors	Radiation dose Difficult to identify small airways Some isotopes can be expensive SPECT and PET not yet widely available

#### Table 1. Summary of physiological and imaging techniques for assessing the small airways

 $FEV_1 =$ forced expiratory volume in 1 sec;  $FEV_3 =$ forced expiratory volume in 3 sec; FVC =forced vital capacity;  $R_{rs} =$ respiratory system resistance;  $X_{rs} =$ respiratory system reactance; Z =impedance; SVC =slow vital capacity; RV =residual volume; TLC =total lung capacity;  $FE_{25-75} =$ forced expiratory flow at 25–75% of vital capacity;  $FE_{NO} =$ fractional expired nitric oxide;  $S_{acin} = DCDI$  contribution to  $S_{nIII}$ ;  $S_{cond} = CDI$  contribution to  $S_{nIII} =$ slope of phase III;  $MLD_{E/I} =$ expiratory to inspiration mean lung density; SPECT =single photon emission computed tomography; PET =Positron emission tomography.

asthma and correlates inversely with FVC (40). However, the upper limit of normal value varies with age and sex and therefore the predicted value may provide a better measure of gas trapping than the absolute value.

Airways resistance  $(R_{aw})$  may also be measured by assessing pressure and flow at the mouth during body

plethysmography. Airways resistance is increased in obstructive lung diseases and is more sensitive to changes than spirometry in detecting bronchodilation (50). However, it is not specific for the small airways which limits its application in diagnosing and monitoring distal airways disease (51).

#### Impulse oscillometry

Impulse oscillometry (IOS) applies oscillating pressure variations in the form of random noise to the respiratory system in order to determine the mechanical properties of the lung. The multiple frequencies between 3 and 20 Hz are applied over normal tidal breathing from a loudspeaker. The resulting pressure and flow changes are measured at the mouth and analysed in a Fourier transformation to determine the impedance (Z) of the respiratory system. This is composed of the in-phase or 'real' part of the impedance, known as resistance  $(R_{rs})$ , and the out of phase or 'imaginary' part, called reactance (X<sub>rs</sub>). In health,  $R_{rs}$  is independent of oscillation frequency but becomes frequency dependent in the presence of airways obstruction. Reactance is determined by the elastic and the inertial properties of the lung and is frequency dependent. At low frequencies, X<sub>rs</sub> is negative and largely represents the elastic forces within the lung. At high frequencies, X<sub>rs</sub> is positive and is determined by inertiance within the lung resulting from acceleration of airflow. At a point where the elastance and inertiance are equal and opposite,  $X_{rs}$  is 0. This is known as the resonant frequency (Fres) and occurs between 8 and 12 Hz in healthy patients (52).

Higher frequency signals (>15 Hz) are absorbed by the respiratory system before reaching the small airways and hence reflect the contribution of large airways. Low frequencies (5 Hz) penetrate deep into the lung and therefore represent the whole lung. The contribution of the distal airways may be determined by the difference between R5 and R20, and therefore can give insight into small airways pathology. However, the anatomical location of the transition between the small and large airways has not been determined (53). Despite this, there is evidence that low frequency resistance and reactance measurements correlate strongly with transpulmonary resistance measured by oesophageal manometry (54) and other traditional small airways measures (55).

When airway obstruction is present,  $R_{rs}$  becomes frequency dependent with a predominant increase in low frequency resistance. This has been shown to identify patients with asthma (56–59) and COPD (56, 60, 61). Whilst  $R_{rs}$  does increase in early stage COPD (60), reactance measures are better at identifying severity of disease (62) and are more closely associated with other parameters including FEV<sub>1</sub> and measures of hyperinflation (61). Dyspnoea scores and health status correlate significantly with  $R_{5-20}$  and  $X_5$  quality of life in stable COPD and are sensitive to improvements following exacerbations (63).

IOS also allows for the discrimination of inspiratory and expiratory resistance and reactance. Inspiratory minus expiratory reactance at 5 Hz ( $\Delta X_5$ ) has been shown to help discriminate between asthma and COPD (64). In addition, it has also been shown to be a sensitive, non-invasive method of detecting expiratory flow limitation (EFL) in COPD. Expiratory reactance falls when EFL is present as the pressure signals cannot pass the choke point with in the airway (65, 66). This is likely to be due to the enhanced collapsibility of airways in expiration and is a major factor in the development of dynamic hyperinflation. Indeed, recent studies using  $R_{5-20}$  as an index of distal airway abnormality have shown the presence of small airways dysfunction even in patients with mild-moderate asthma (67).

Studies examining the effect of inhaled therapies on lung mechanics have demonstrated that IOS is sensitive to bronchodilation in both COPD (50, 68) and asthma (69, 70). It has also been used in the assessment of lung transplant recipients for bronchiolitis obliterans (71) and following environmental exposure to dusts (72, 73).

IOS has the advantage of being simple to use and is effort independent. It provides continuous measurement of pulmonary mechanics giving a high temporal resolution allowing intra-breath analysis. As IOS does not rely on forced manoeuvres, it may be more suitable for patients who cannot perform these easily such as children or those with severe lung diseases. This may also reduce the effects of premature airway closure seen during forced spirometry manoeuvres. Interference from upper airways artefacts such as tongue movement or swallowing can make assessment difficult. Patients undergoing IOS do need some coaching for accurate measures to be made.

#### Inert gas washout

Gas washout techniques were introduced in the 1950s as a way of measuring the efficiency of gas mixing within the lungs. This is dependent on the structure of both the large and small airways and hence information regarding these can be inferred from the tests. The most commonly employed technique is the single breath nitrogen washout (SBNW) and more recently the multiple breath nitrogen washout (MBNW). Other gases may be used including helium and sulphur hexafluoride (SF<sub>6</sub>) whose physical properties determine gas flow within the lung.

#### Single breath nitrogen washout

The SBNW is performed by inhaling 100% oxygen from RV to TLC followed by a SVC exhalation. The exhaled volume and nitrogen concentration is measured and the resulting trace can be broken down into four distinct phases.

In phase I, the nitrogen concentration is close to 0% as this represents anatomical dead space where there is no gas mixing. During phase II, there is a sharp rise in the expired nitrogen concentration as dead space gas mixes with resident alveolar gas. Phase III represents alveolar gas and the expired nitrogen concentration begins to plateau, although there is a slight rise from the start to finish of this phase due to ventilation heterogeneity.

This occurs whenever two lung units are ventilated to a different degree and the best ventilated unit will empty preferentially before a less well-ventilated lung unit. In health, this occurs to a degree because of asymmetry in lung structure and due to the effects of gravity on the base of the lung resulting in longer time constants for emptying. Finally, in phase IV, there is a steep rise in expired N<sub>2</sub> concentration as the most poorly ventilated areas (with little  $O_2$  mixing) empty. This is also the point at which the small airways start to close as a result of gravity-dependent collapse and is known as the closing volume (CV). The CV and RV together are known as the closing capacity (CC). Normally, small airways closure occurs close to RV. However, small airways disease may cause premature airway collapse resulting in an increased CV and gas trapping. CV may be expressed as a ratio of VC and should not exceed 25% (74). The CC may be expressed as a ratio of TLC and is useful in obstructive lung diseases.

Analysis of the slope of phase III (SIII) provides information on the ventilation heterogeneity in the lung. Airways diseases do not affect the lung uniformly and this results in disparities in the ventilation of individual subunits. This may occur in the conducting airways where gas flows by convection (convection-dependent ventilation inhomogeneity, CDI) and results from narrowing of airways or increased stiffness in the subtended lung units. It may also occur in the very distal acinar airways where the diffusion-convection front arises (diffusion-convection-dependent inhomogeneity, DCDI). Here, it occurs as a result of structural asymmetry between lung units (75). Thus, where airways disease occurs, those affected lung units mix less well with the inspired oxygen (and thus have a higher nitrogen concentration) and empty more slowly. This causes an increase in S<sub>III</sub>.

SBNW indices have been used in the assessment and response to treatment in both asthma and COPD. Asthmatic patients with a normal FEV<sub>1</sub> have increased CV and phase III slope compared to healthy controls. In addition, the frequency of exacerbations correlates with S<sub>III</sub> suggesting it may be a sensitive measure of patients with poor control (76). Indeed, increased CV in patients with severe asthma has been shown to be a risk factor for predicting an exacerbation (77). Levels of exhaled markers of airway inflammation including nitric oxide correlate with S<sub>III</sub> and CC/TLC ratio in asthma (78, 79). Furthermore, severe, steroid-dependent asthmatic patients have more marked changes in SBNW indices than patients with mild to moderate asthma (79). These markers have also been used to assess changes following both inhaled and oral therapies for asthma (80-83).

Over 35 years ago, the  $S_{III}$  of the SBNW was recognised as being more closely related to histological small airways inflammation in COPD than FEF<sub>25-75</sub> (29). Further evidence of its association with small airways inflammation in COPD came from the examination of bronchial biopsies and bronchoalveolar lavage (BAL) specimens (84). COPD severity may also be predicted by changes in SBNW indices as the  $S_{III}$  correlates with FEV<sub>1</sub> (85) and TL<sub>CO</sub> in alpha-1 antitrypsin deficiency (86).

SBNW is sensitive to early changes in airways in smokers with an increase in CV (87), but its use is controversial in COPD. Buist et al. demonstrated that many smokers with normal spirometry, but abnormal small airway indices, did not go on to develop obstructive spirometry over a 9–11 year follow-up. However, of those that did, the CC/TLC ratio predicted the rate of decline in FEV<sub>1</sub> suggesting it may be useful in identifying at risk smokers (88). Stănescu et al. similarly found that in a group of smokers and ex-smokers with normal spirometry, over half had abnormal small airway indices, yet most still had normal spirometry 13 years later. In their cohort, a high S<sub>III</sub> predicted accelerated decline in FEV<sub>1</sub> (89).

Despite its sensitivity, the SBNW is not specific to small airways pathology. Changes in any of the generations of the conducting airways will also affect the slope of phase III. Thus, whilst it is possible to infer that a normal  $S_{\rm III}$  indicates no small airways disease, the test is unable to locate the anatomical site of the pathology (34).

#### Multiple breath nitrogen washout

The MBNW is a modification of the single breath technique. The patient inhales 100% O<sub>2</sub> from FRC with a fixed tidal volume and respiratory rate to wash out the resident nitrogen from the lungs. The test continues until the exhaled nitrogen is less than 1/40th of the original concentration (approximately 2%) for three successive breaths. The speed and efficiency of gas mixing is determined by tidal volume, breath frequency, and ventilation heterogeneity. Thus, by keeping breath frequency and tidal volume relatively constant, inferences about ventilation heterogeneity can be made (90). Figure 2 demonstrates the nitrogen washout curves from a MBNW test.

This technique allows for measurement of the efficiency of gas mixing in the whole lung through the lung clearance index (LCI). It is defined as the number of lung turnovers (FRC equivalents) required to wash out the tracer gas to 1/40th of the original concentration. This is calculated by measuring the cumulative expired volume (CEV) required to washout the resident nitrogen and dividing it by FRC:

$$LCI = \frac{CEV}{FRC}$$

FRC may be calculated during the MBNW from the following formula, whereby the volume of tracer gas (i.e.  $N_2$ ) is divided by the end-tidal concentration of the tracer



*Fig. 2.* (a) Multiple breath nitrogen washout curve with individual breaths demonstrating Phase III slope (SnIII) from 1st (b) and 10th (c) breaths.

gas in the first breath minus the end-tidal concentration of the tracer gas in the last breath:

$$FRC = rac{V_{[tracer]}}{C_{int} - C_{end}}$$

The LCI has been used extensively as a measure of airways function in cystic fibrosis and asthma, particularly in the paediatric population (91).

As a MBNW progresses, the  $S_{III}$  of each breath changes throughout the test, becoming steeper with successive breaths. In order to compare breaths within a test, the slopes must be normalised for the mean expired nitrogen concentration for each breath ( $S_{nIII}$ ). In normal lungs, the DCDI is the major determinant of the  $S_{nIII}$  and reaches its maximum at approximately 1.5 lung turnovers. After this, the increase in  $S_{nIII}$  is diffusion independent and hence reflects CDI (92). This allows for the quantification of the contribution of the CDI component, referred to as  $S_{cond}$ , and the DCDI component, referred to as  $S_{acin}$  (75). Thus, these indices have the ability to anatomically locate the site of the airway pathology that result in ventilation inhomogeneity.

These indices have proven very sensitive, becoming abnormal in smokers with more than a 10-year pack history. In contrast, spirometric abnormalities only become abnormal after a 20 pack-year history of smoking. For smokers with a > 30 pack-year history and TL<sub>CO</sub> < 60% predicted, there were proportionately larger changes in S<sub>acin</sub> than S<sub>cond</sub>, reflecting parenchymal destruction (93). Smokers without COPD who were able to stop smoking showed sustained reversibility in S<sub>cond</sub> (94). This supports the hypothesis that the major site of pathology in smoking-related lung disease starts in the peripheral airways.

These abnormalities have been further described in both asthma and COPD. Verbanck et al. demonstrated in COPD patients that both Scond and Sacin are raised yet reflect different pathologies. Scond correlated with airways measures such as FEV<sub>1</sub> and specific airways resistance whilst Sacin was more closely associated with diffusing capacity (95). Asthmatic patients also have raised S<sub>cond</sub> and Sacin, although acinar ventilation heterogeneity is less pronounced than in COPD, presumably reflecting the degree of parenchymal destruction in COPD. In addition, asthmatic patients demonstrated bronchodilator reversibility in both Sacin and Scond, whilst COPD patients did not (96). In asthma, Sacin is more closely associated with airway inflammation (97) and severity in unstable patients (98). It has recently been shown that measures of ventilation heterogeneity are associated with levels of asthma control and may also predict the response to inhaled therapy (99, 100). With their sensitivity to small airways disease, they have been used in a variety of research settings. These include the assessment of inhaled

treatments in both asthma (101, 102) and COPD (103), assessment of airway hyper-responsiveness (104, 105), and monitoring of lung transplant recipients (106). However, they are not yet used in routine clinical practice as there are few commercially available machines, and interpretation of results can be difficult. Theoretically, abnormalities in any of the conducting airways from the first generation can cause abnormalities in S<sub>cond</sub> and therefore it is not specific to small airways. Interpreting the results with information from spirometry will help clarify this. In addition, theoretical modelling for localisation of airways disease was performed in normal subjects. It is possible that the convection–diffusion front is different in disease states and hence anatomical localisations may not be precise.

Helium and Sulphur hexafluoride washout tests

Other inert gasses including helium and  $SF_6$  may be used in small concentrations as tracer gasses. These require a wash-in period and specialised analytical equipment. However, they have the added benefit that the physiochemical properties can be exploited to gain further information from the  $S_{III}$ . The diffusion front of helium lies more proximally than  $SF_6$  and therefore changes in the helium  $S_{III}$  compared to  $SF_6 S_{III}$  suggest more proximal acinar changes. Where both  $S_{III}$  change so that the difference between them is still the same, the possibilities are either a change in the conducting airways or concomitant effects in the proximal and distal parts of the acinus (34). There are fewer clinical studies reporting  $SF_6$ as a tracer gas and these have largely been performed in children with cystic fibrosis (107–109).

#### Exhaled nitric oxide

Nitric oxide is produced in both the resident airway cells and the inflammatory cells in the lung and has a role in the regulation of airway function. Fractional exhaled nitric oxide ( $FE_{NO}$ ) may be measured in a single exhalation during tidal breathing. It reflects levels of inflammation, particularly eosinophillic inflammation, within the lung (110). Exhaled nitric oxide (eNO) exhibits flow rate dependency, with an inverse correlation between flow rate and  $FE_{NO}$  (111). This reflects both the transit time of exhaled gas and diffusion from the tissue as well as the compartment of the lung from which the NO was produced. Under low flow conditions, FE<sub>NO</sub> largely reflects central airways and at higher flows it represents alveolar NO (112-114). This may help to localise the site of inflammation within the lung. Indeed, Lehtimäki et al. demonstrated that patients with alveolitis had higher levels of alveolar NO than asthmatic patients, who in turn have higher bronchial NO. In patients with alveolitis, alveolar NO correlated with transfer factor and alveolar volume, whilst bronchial NO correlated with airways'

hyper-responsiveness in asthmatic patients. Both groups of patients showed an improvement in FE<sub>NO</sub> with steroid treatment, suggesting it is responsive to intervention (115). However, back-diffusion of NO between the alveolar and airway compartments complicates the interpretation of results. It has been recognised that NO will diffuse from the airways down a concentration gradient into the alveoli, thus elevating alveolar NO and reducing measured airway NO (116, 117). Models to correct this have been developed, however, in disease states where airways are narrowed or occluded; less NO can backdiffuse, resulting in higher FE<sub>NO</sub> and lower alveolar concentrations (118). It should also be noted that current smoking reduces FE<sub>NO</sub> levels and thus the smoking status of a patient needs to be taken into account when interpreting results (110).

FE<sub>NO</sub> has been used extensively in asthma clinical research and practice. Central airways appear to be the major sites of production of NO in asthma both in stable populations and during exacerbations (119). Alveolar NO concentrations are raised in severe asthmatics where they correlate with alveolar eosinophillic inflammation (120) and other measures of small airways dysfunction (121). Recently, it has also been shown that alveolar NO is also raised in patients with mild asthma (122).  $FE_{NO}$  is improved by both oral (123, 124) and inhaled corticosteroids (ICS) (125) and a raised FENO level before ICS treatment predicts an improvement in asthma control (126). This has made  $FE_{NO}$  an attractive prospect for adding to asthma treatment algorithms. However, the results of studies assessing impact of measuring  $FE_{NO}$ have been mixed. Meta-analyses suggest no overall benefit to asthma control and quality of life, but there is a reduction in ICS use in adults although an increase in ICS use in children (127, 128).

The role of  $FE_{NO}$  in COPD is less clear.  $FE_{NO}$  may be raised in COPD (129–131), although it is lower compared to asthmatic patients. An inverse correlation with  $FEV_1$ , transfer factor, and oxygen saturations has been reported (129). Contrary to this, Gelb et al. found no difference in baseline alveolar or airway NO levels between healthy controls and aged-matched COPD patients. Despite this, the addition of salmeterol 50 mcg/fluticasone 250 mcg combination inhaler significantly reduced airway, but not alveolar NO. There was no correlation between emphysema score and exhaled NO parameters (119). Higher  $FE_{NO}$  levels may help predict a clinical response to ICS as assessed by  $FEV_1$  reversibility (132) and this is associated with a higher sputum eosinophil count (133).

#### Imaging of the small airways

Imaging already plays an extensive role in the management of airways disease and can be used as a non-invasive measure of small airways function. Where global measures of lung function such as spirometry may classify patients of the same severity, imaging is useful in separating different phenotypes and localising heterogeneity. However, direct measurement of small airways is difficult as they are largely beyond the resolution of CT and MRI scanners. Nevertheless, both large airways have been assessed directly and the smaller airways by their impact on gas trapping and ventilation distribution. This provides both anatomical and functional information to the physician.

## **High resolution CT**

The small airways are beyond the resolution of CT scanners and difficult to assess directly (134). Airways as small as 2-2.5 mm in diameter can be visualised. McDonough et al. found fewer of these airways in patients with COPD undergoing CT lung cancer screening. The reduction in airway number worsened as COPD severity increased by stage, consistent with pathological findings in lung specimens (135). However, the accuracy of measurement of smaller airways may be problematic due to measurement error and artefact from breathing or cardiogenic oscillations. Nakano et al. demonstrated that measurement of intermediate-sized airways could predict the small airway dimensions measured by histology (136); thus, assessment may still prove useful in estimating the extent of small airways disease. Quantitative assessment of more proximal airway luminal diameter and airway wall thickening measured by CT correlate with lung function in COPD (137-140), with the strength of correlation increasing for more distal airways (139).

Small airways disease results in gas trapping and may be seen as areas of low attenuation distal to the site of obstruction. Mosaic attenuation reflects localised areas of gas trapping and suggests heterogeneous distribution of airways disease. It may be seen in both asthma (141) and COPD (142). However, gas trapping is best assessed on expiratory scans and may provide an indirect measure of small airways function (143). Comparing the mean lung density between expiratory and inspiratory CT provides a quantitative measure called  $MLD_{E/I}$ . In asthma,  $MLD_{E/I}$ correlates strongly with FEV<sub>1</sub>, FEV<sub>1</sub>/FVC ratio, FEF<sub>25-75</sub>, and RV/TLC, suggesting that it reflects small airways disease (144). In children, gas trapping has been shown to be associated with improvements in post-bronchodilator  $R_5$  and  $X_5$  measured by IOS (145). Gas trapping is more marked during acute exacerbations of asthma and shows responsiveness to steroids (146). Asthmatic patients with gas trapping are more likely to have had asthma-related hospital admissions, intensive care treatment, high levels of airway neutrophils, and more severe airflow obstruction than those without (147). ICS have been shown to improve gas trapping in asthma, although in these small studies no significant benefit in lung function or spirometry was seen (148, 149).

The assessment of gas trapping in COPD is more complicated as both small airways disease and emphysema give rise to low attenuation areas. Visual estimation of emphysema provides better correlation with lung function measures such as TL<sub>CO</sub>, whereas MLD<sub>E/I</sub> correlates well with lung function measures of gas trapping (150). In a large cohort of patients in the COPDGene study. expiratory scans with a threshold of -856 Hounsfield units (HU) for assessment of gas trapping had a stronger correlation with airflow obstruction than emphysema scores measured by the area of lung with -950 HU on inspiratory scans. The volume change between inspiratory and expiratory scans also reduced as COPD severity increased, reflecting more severe airway obstruction and gas trapping (151). Using MLD<sub>E/I</sub> scores to assess gas trapping and the 15th percentile of lung density to assess emphysema, Hartley et al. demonstrated that small airways disease contributes more strongly than emphysema to severity of COPD (152). MLD<sub>E/I</sub> also correlates with inflammatory changes measured by sputum neutrophils, adding further support to the inflammatory nature of small airways disease in COPD. MLD<sub>E/I</sub> is sensitive to early small airways changes and correlates with S<sub>III</sub> in SBNW in a group of asymptomatic, non-smokers (153). However, MLDE/I can show considerable variation between scans in individual patients and therefore may be difficult to use as a marker of response to treatment. Recently, in the research arena, static images obtained with CT have been made more functional by means of computational fluid dynamics (154) and biomarkers based on CT imaging have been developed that allow an assessment of functional small airways disease (155).

Whilst CT is a useful, non-invasive tool for indirectly assessing small airways function, it has a number of limitations. The exposure to radiation means that repeated assessment for monitoring is not feasible. There is no standardised measure of gas trapping at present and different authors have used different density thresholds for assessing gas trapping, making comparison more difficult. Gas trapping is not diagnostic for specific airways diseases and patterns such as mosaic attenuation are also seen in pulmonary vascular disease (156).

# Hyperpolarised helium magnetic resonance imaging

Hyperpolarised helium magnetic resonance imaging (<sup>3</sup>He MRI) allows for the assessment of distribution of ventilation and morphometry of the distal airways and lung parenchyma without exposure to ionising radiation (157). Diffusion imaging visualises the movement of <sup>3</sup>He in the peripheral airspaces, bound by alveolar and airways walls. This is calculated as the apparent diffusion co-efficient (ADC) and gives insight into the microstructure of the distal airspaces (158). ADC is increased in healthy smokers with normal lung function and correlates

with smoking history (159, 160), suggesting it is a sensitive marker of early damage. It is increased further in COPD where it correlates with lung function (161) and emphysematous destruction (162). Indeed, ADC correlates well with CT-derived emphysema scores and more strongly with TL<sub>CO</sub> than CT-derived emphysema scores (163). In COPD patients observed over 26 months, ADC and other parameters derived from <sup>3</sup>He MRI have been shown to decline whilst FEV<sub>1</sub> remained stable, suggesting it is also sensitive to change over time (164). Measuring diffusion over longer periods allows the assessment of collateral ventilation in emphysema (165, 166).

Quantification of regional ventilation can be achieved by both static and dynamic assessment of <sup>3</sup>He distribution within the lung. Ventilation defects are present in asthma (167, 168) and COPD (169, 170), resulting from airway narrowing or obstruction and uneven ventilation. In a group of asthmatic patients, the areas of ventilation defects were persistent or recurred in the same locations over time (169). Inflammatory cells obtained at bronchoalveolar lavage were more numerous in lobes with higher ventilation defects that those without, suggesting that the defects are the result of inflammatory airway narrowing (171). Dynamic ventilation is a more recent advance that allows imaging and assessment of ventilation with a high spatial and temporal resolution over the course of a respiratory cycle (161, 172). In asthma, areas of differential gas clearance have been observed that corroborat with evidence of airflow obstruction and gas trapping on CT (173).

Hyperpolarised MRI has the ability to assess regional lung function which makes it a useful tool in assessing airways diseases which have a heterogeneous distribution. However, the technique is still largely restricted to research applications and its role in the clinical management of airways disease is not yet clear.

# Nuclear medicine techniques

### Two-dimensional gamma scintigraphy

Two-dimensional (2-D) gamma scintigraphy has been in use for several decades. Gamma-emitting radionucleides deposited within the lung can be imaged as they decay. This allows for an assessment of the overall lung deposition and to some extent, regional differences in deposition. Incorporating radionucleide into drug compounds is challenging and hence an isotope bound to the drug such as <sup>99m</sup>Tc is more commonly used. These techniques must be validated to ensure that the addition of a radiolabel does not significantly change the behaviour of the drug (174).

2-D gamma scintigraphy has been used to assess the effect of particle size on deposition within the lungs. Usmani et al. studied three particle sizes of radiolabelled salbutamol and found that whilst small particle (1.5 micron) salbutamol was associated with a higher total

lung dose and more peripheral deposition, it was the large particles (6 micron) deposited in the more proximal airways that had the greatest effect on bronchodilation (13). In addition, the effect of late inhalation of a dry powder demonstrated that a higher proportion reaches the periphery of the lung without a change in the total lung dose (175).

Whilst scintigraphy does involve exposure to ionising radiation, the dose is low and is estimated at 0.15 mSv per study (176). However, 2-D imaging does not allow precise localisation of drug deposition as both central and small airways as well as alveolar distribution may contribute to gamma counts for any given area. Whilst assessment of deposition is a useful marker of drug distribution, it does not itself provide an assessment of clinical or physiological response. Hence, these studies must be assessed along with clinical and physiological data in order to evaluate efficacy.

#### Single photon emission computed tomography

Single photon emission computed tomography (SPECT) is a 3-D imaging modality using multiple gamma detectors that rotate around a supine patient. Reconstruction of the images can demonstrate the radionucleide distribution in three dimensions, thereby offering superior assessment of regional lung ventilation or particle distribution. SPECT may be combined with X-ray CT to relate the radionuclide distribution to anatomical information (177, 178). SPECT can be used to image ventilation using either radiolabelled gasses or ultrafine particles such as Technegas<sup>®</sup>. This is an ultrafine carbon particle labelled with <sup>99m</sup>Tc that has been shown to have a similar inhaled distribution in healthy patients to gases (179). This allows measurement of the extent of regional distribution of airflow. In healthy patients, airway closure measured with SPECT correlates with CC measured by SBNW. However, in asthmatic patients this correlation is lost, possibly due to regional heterogeneity in airway closure (180). Technegas SPECT has also been shown to identify regional EFL in asthma even when flow measurements or negative expiratory pressure techniques are insensitive to it (181). In COPD, Technegas SPECT can identify regional differences in emphysema which correlates with lung function and emphysema scores (182, 183). The technique can be combined with perfusion imaging to assess ventilationperfusion relationships in the lung (184).

SPECT has proven a useful tool in defining the deposition of inhaled drugs and can allow for treatments to be more specifically targeted to areas of the lung. Ciclesonide has been shown to have good peripheral lung deposition, with low oropharyngeal deposition in both health (185) and asthma (186). Limitations of SPECT scanning include higher radiation doses to patients and a longer acquisition time. This limits the assessment of deposition of molecules with a fast clearance. However, fast SPECT protocols have been developed with image

acquisition times under 1 min. This allows for assessment of both deposition and clearance of tracers (187).

#### Positron emission tomography

Positron emission tomography (PET) is an emerging technique for assessment of airways disease. It can be used to assess drug deposition (188), inflammation (189), and ventilation perfusion relationships in the lung (190). However, PET scanners and the facilities to produce radioisotopes are expensive. The radioisotopes used often have a short half-life and incorporating them into drugs is a complex process. However, it has the ability to produce higher resolution images than SPECT and allows for targeting of radioisotopes to specific receptors and targets within the lung. Therefore, it is likely to be an area of exciting research in the assessment of small airways disease and treatment (191).

#### Conclusions

An understanding of the role of small airways in COPD and asthma is increasingly important as it becomes necessary to distinguish individual phenotypes of the diseases. This will lead to a more tailored approach to assessment and treatment of patients with the aim of improving symptoms and function (192). It may also allow us to reduce unnecessary exposure to treatments that carry significant side effects. Given the anatomical, functional, and physiological information that can be obtained from these different tests, it is likely that a combination of investigations will be required to give the clearest picture of an individual's phenotype. At present, however, many of these investigations remain in the realm of the research laboratory and further work is required to understand their significance and interpretation in the management of these diseases.

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#### References

- Weibel ER. Principles and methods for the morphometric study of the lung and other organs. Lab Invest. 1963; 12: 131– 55.
- Tatsis G, Horsfield K, Cumming G. Distribution of dead space volume in the human lung. Clin Sci (Lond). 1984; 67: 493–7.

- Wiebe BM, Laursen H. Human lung volume, alveolar surface area, and capillary length. Microsc Res Tech. 1995; 32: 255–62.
- Ranga V, Kleinerman J. Structure and function of small airways in health and disease. Arch Pathol Lab Med. 1978; 102: 609–17.
- Macklem PT, Proctor DF, Hogg JC. The stability of peripheral airways. Respir Physiol. 1970; 8: 191–203.
- Baraldo S, Turato G, Saetta M. Pathophysiology of the small airways in chronic obstructive pulmonary disease. Respiration. 2012; 84: 89–97.
- Weibel ER, Sapoval B, Filoche M. Design of peripheral airways for efficient gas exchange. Respir Physiol Neurobiol. 2005; 148: 3–21.
- Macklem PT, Mead J. Resistance of central and peripheral airways measured by a retrograde catheter. J Appl Physiol. 1967; 22: 395–401.
- Hogg JC, Macklem PT, Thurlbeck WM. Site and nature of airway obstruction in chronic obstructive lung disease. N Engl J Med. 1968; 278: 1355–60.
- Wagner EM, Liu MC, Weinmann GG, Permutt S, Bleecker ER. Peripheral lung resistance in normal and asthmatic subjects. Am Rev Respir Dis. 1990; 141: 584–8.
- Yanai M, Sekizawa K, Ohrui T, Sasaki H, Takishima T. Site of airway obstruction in pulmonary disease: direct measurement of intrabronchial pressure. J Appl Physiol. 1992; 72: 1016–23.
- Wagner EM, Bleecker ER, Permutt S, Liu MC. Direct assessment of small airways reactivity in human subjects. Am J Respir Crit Care Med. 1998; 157: 447–52.
- Usmani OS, Biddiscombe MF, Barnes PJ. Regional lung deposition and bronchodilator response as a function of beta2-agonist particle size. Am J Respir Crit Care Med. 2005; 172: 1497–504.
- Hamid Q. Pathogenesis of small airways in asthma. Respir Int Rev Thorac Dis. 2012; 84: 4–11.
- De Magalhães Simões S, dos Santos MA, da Silva Oliveira M, Fontes ES, Fernezlian S, Garippo AL, et al. Inflammatory cell mapping of the respiratory tract in fatal asthma. Clin Exp Allergy. 2005; 35: 602–11.
- Hamid Q, Song Y, Kotsimbos TC, Minshall E, Bai TR, Hegele RG, et al. Inflammation of small airways in asthma. J Allergy Clin Immunol. 1997; 100: 44–51.
- James AL, Paré PD, Hogg JC. The mechanics of airway narrowing in asthma. Am Rev Respir Dis. 1989; 139: 242–6.
- Saetta M, Di Stefano A, Rosina C, Thiene G, Fabbri LM. Quantitative structural analysis of peripheral airways and arteries in sudden fatal asthma. Am Rev Respir Dis. 1991; 143: 138–43.
- Carroll N, Elliot J, Morton A, James A. The structure of large and small airways in nonfatal and fatal asthma. Am Rev Respir Dis. 1993; 147: 405–10.
- Haley KJ, Sunday ME, Wiggs BR, Kozakewich HP, Reilly JJ, Mentzer SJ, et al. Inflammatory cell distribution within and along asthmatic airways. Am J Respir Crit Care Med. 1998; 158: 565–72.
- Balzar S, Chu HW, Strand M, Wenzel S. Relationship of small airway chymase-positive mast cells and lung function in severe asthma. Am J Respir Crit Care Med. 2005; 171: 431–9.
- Kraft M, Martin RJ, Wilson S, Djukanovic R, Holgate ST. Lymphocyte and eosinophil influx into alveolar tissue in nocturnal asthma. Am J Respir Crit Care Med. 1999; 159: 228–34.
- Carroll N, Cooke C, James A. The distribution of eosinophils and lymphocytes in the large and small airways of asthmatics. Eur Respir J. 1997; 10: 292–300.
- Turato G, Zuin R, Miniati M, Baraldo S, Rea F, Beghé B, et al. Airway inflammation in severe chronic obstructive pulmonary

disease: relationship with lung function and radiologic emphysema. Am J Respir Crit Care Med. 2002; 166: 105–10.

- Saetta M, Di Stefano A, Turato G, Facchini FM, Corbino L, Mapp CE, et al. CD8+ T-lymphocytes in peripheral airways of smokers with chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 1998; 157: 822–6.
- Cosio MG, Hale KA, Niewoehner DE. Morphologic and morphometric effects of prolonged cigarette smoking on the small airways. Am Rev Respir Dis. 1980; 122: 265–21.
- Hogg JC. Pathophysiology of airflow limitation in chronic obstructive pulmonary disease. Lancet. 2004; 364: 709–21.
- Hogg JC, Timens W. The pathology of chronic obstructive pulmonary disease. Annu Rev Pathol. 2009; 4: 435–59.
- Cosio M, Ghezzo H, Hogg JC, Corbin R, Loveland M, Dosman J, et al. The relations between structural changes in small airways and pulmonary-function tests. N Engl J Med. 1978; 298: 1277–81.
- Hogg JC, Chu F, Utokaparch S, Woods R, Elliott WM, Buzatu L, et al. The nature of small-airway obstruction in chronic obstructive pulmonary disease. N Engl J Med. 2004; 350: 2645–53.
- Churg A, Tai H, Coulthard T, Wang R, Wright JL. Cigarette smoke drives small airway remodeling by induction of growth factors in the airway wall. Am J Respir Crit Care Med. 2006; 174: 1327–34.
- 32. Saetta M, Ghezzo H, Kim WD, King M, Angus GE, Wang NS, et al. Loss of alveolar attachments in smokers. A morphometric correlate of lung function impairment. Am Rev Respir Dis. 1985; 132: 894–900.
- 33. Hogg JC, McDonough JE, Sanchez PG, Cooper JD, Coxson HO, Elliott WM, et al. Micro-computed tomography measurements of peripheral lung pathology in chronic obstructive pulmonary disease. Proc Am Thorac Soc. 2009; 6: 546–9.
- Verbanck S. Physiological measurement of the small airways. Respiration. 2012; 84: 177–88.
- 35. Rabe KF, Hurd S, Anzueto A, Barnes PJ, Buist SA, Calverley P, et al. Global strategy for the diagnosis, management, and prevention of chronic obstructive pulmonary disease GOLD executive summary. Am J Respir Crit Care Med. 2007; 176: 532–55.
- 36. Pride NB. Tests of forced expiration and inspiration. Clin Chest Med. 2001; 22: 599–622.
- McFadden Jr. ER, Linden DA. A reduction in maximum midexpiratory flow rate: a spirographic manifestation of small airway disease. Am J Med. 1972; 52: 725–37.
- Boggs PB, Bhat KD, Vekovius WA, Debo MS. Volumeadjusted maximal mid-expiratory flow (Iso-volume FEF25– 75%): definition of "Significant" responsiveness in healthy, normal subjects. Ann Allergy. 1982; 48: 137–8.
- Gelb AF, Williams AJ, Zamel N. SPirometry. fev1 vs. fef25–75 percent. Chest J. 1983; 84: 473–4.
- Sorkness RL, Bleecker ER, Busse WW, Calhoun WJ, Castro M, Chung KF, et al. Lung function in adults with stable but severe asthma: air trapping and incomplete reversal of obstruction with bronchodilation. J Appl Physiol. 2008; 104: 394–403.
- Sutherland ER, Martin RJ, Bowler RP, Zhang Y, Rex MD, Kraft M. Physiologic correlates of distal lung inflammation in asthma. J Allergy Clin Immunol. 2004; 113: 1046–50.
- Hansen JE, Sun X-G, Wasserman K. Discriminating measures and normal values for expiratory obstruction\*. Chest J. 2006; 129: 369–77.
- Gibbons WJ, Sharma A, Lougheed D, Macklem PT. Detection of excessive bronchoconstriction in asthma. Am J Respir Crit Care Med. 1996; 153: 582–9.

- 44. Cohen J, Postma DS, Vink-Klooster K, van der Bij W, Verschuuren E, Ten Hacken NHT, et al. FVC to slow inspiratory vital capacity ratio: a potential marker for small airways obstruction. Chest. 2007; 132: 1198–203.
- 45. Burgel P-R, Bourdin A, Chanez P, Chabot F, Chaouat A, Chinet T, et al. Update on the roles of distal airways in COPD. Eur Respir Rev. 2011; 20: 7–22.
- Jain VV, Abejie B, Bashir MH, Tyner T, Vempilly J. Lung volume abnormalities and its correlation to spirometric and demographic variables in adult asthma. J Asthma. 2013; 50: 600–5.
- Perez T, Chanez P, Dusser D, Devillier P. Small airway impairment in moderate to severe asthmatics without significant proximal airway obstruction. Respir Med. 2013; 107: 1667–74.
- Kraft M, Pak J, Martin RJ, Kaminsky D, Irvin CG. Distal lung dysfunction at night in nocturnal asthma. Am J Respir Crit Care Med. 2001; 163: 1551–6.
- Kraft M, Cairns CB, Ellison MC, Pak J, Irvin C, Wenzel S. Improvements in distal lung function correlate with asthma symptoms after treatment with oral montelukast\*. Chest J. 2006; 130: 1726–32.
- Borrill ZL, Houghton CM, Woodcock AA, Vestbo J, Singh D. Measuring bronchodilation in COPD clinical trials. Br J Clin Pharmacol. 2005; 59: 379–84.
- Pellegrino R. Interpretative strategies for lung function tests. Eur Respir J. 2005; 26: 948–68.
- 52. Oostveen E, MacLeod D, Lorino H, Farré R, Hantos Z, Desager K, et al. The forced oscillation technique in clinical practice: methodology, recommendations and future developments. Eur Respir J. 2003; 22: 1026–41.
- Goldman MD, Saadeh C, Ross D. Clinical applications of forced oscillation to assess peripheral airway function. Respir Physiol Neurobiol. 2005; 148: 179–94.
- Johnson MK, Birch M, Carter R, Kinsella J, Stevenson RD. Use of reactance to estimate transpulmonary resistance. Eur Respir J. 2005; 25: 1061–9.
- 55. Hira H, Munjal J, Zachariah S, Chauhan M, Singh A. The site of airway obstruction among patients of emphysema: role of impulse oscillometry. Lung India. 2008; 25: 8–13.
- Grimby G, Takishima T, Graham W, Macklem P, Mead J. Frequency dependence of flow resistance in patients with obstructive lung disease. J Clin Invest 1968; 47: 1455–65.
- Cavalcanti JV, Lopes AJ, Jansen JM, Melo PL. Detection of changes in respiratory mechanics due to increasing degrees of airway obstruction in asthma by the forced oscillation technique. Respir Med. 2006; 100: 2207–19.
- Kaminsky DA, Irvin CG, Lundblad L, Moriya HT, Lang S, Allen J, et al. Oscillation mechanics of the human lung periphery in asthma. J Appl Physiol (1985) 2004; 97: 1849–58.
- Goldman MD, Carter R, Klein R, Fritz G, Carter B, Pachucki P. Within- and between-day variability of respiratory impedance, using impulse oscillometry in adolescent asthmatics. Pediatr Pulmonol. 2002; 34: 312–9.
- Di Mango AMGT, Lopes AJ, Jansen JM, Melo PL. Changes in respiratory mechanics with increasing degrees of airway obstruction in COPD: detection by forced oscillation technique. Respir Med. 2006; 100: 399–410.
- Kolsum U, Borrill Z, Roy K, Starkey C, Vestbo J, Houghton C, et al. Impulse oscillometry in COPD: identification of measurements related to airway obstruction, airway conductance and lung volumes. Respir Med. 2009; 103: 136–43.
- Timmins SC, Diba C, Farrow CE, Schoeffel RE, Berend N, Salome CM, et al. The relationship between airflow obstruction, emphysema extent, and small airways function in COPD. Chest. 2012; 142: 312–9.

- Stevenson NJ, Walker PP, Costello RW, Calverley PMA. Lung mechanics and dyspnea during exacerbations of chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2005; 172: 1510–6.
- 64. Paredi P, Goldman M, Alamen A, Ausin P, Usmani OS, Pride NB, et al. Comparison of inspiratory and expiratory resistance and reactance in patients with asthma and chronic obstructive pulmonary disease. Thorax. 2010; 65: 263–7.
- 65. Dellacà RL, Santus P, Aliverti A, Stevenson N, Centanni S, Macklem PT, et al. Detection of expiratory flow limitation in COPD using the forced oscillation technique. Eur Respir J. 2004; 23: 232–40.
- Dellacà RL, Duffy N, Pompilio PP, Aliverti A, Koulouris NG, Pedotti A, et al. Expiratory flow limitation detected by forced oscillation and negative expiratory pressure. Eur Respir J. 2007; 29: 363–74.
- Anderson WJ, Zajda E, Lipworth BJ. Are we overlooking persistent small airways dysfunction in community-managed asthma? Ann Allergy Asthma Immunol. 2012; 109: 185–9.e2.
- 68. Houghton CM, Woodcock AA, Singh D. A comparison of plethysmography, spirometry and oscillometry for assessing the pulmonary effects of inhaled ipratropium bromide in healthy subjects and patients with asthma. Br J Clin Pharmacol. 2005; 59: 152–9.
- Park J-W, Lee Y-W, Jung Y-H, Park S-E, Hong C-S. Impulse oscillometry for estimation of airway obstruction and bronchodilation in adults with mild obstructive asthma. Ann Allergy Asthma Immunol. 2007; 98: 546–52.
- Houghton CM, Woodcock AA, Singh D. A comparison of lung function methods for assessing dose-response effects of salbutamol. Br J Clin Pharmacol. 2004; 58: 134–41.
- Hamakawa H, Sakai H, Takahashi A, Zhang J, Okamoto T, Satoda N, et al. Forced oscillation technique as a non-invasive assessment for lung transplant recipients. Adv Exp Med Biol. 2010; 662: 293–8.
- Skloot G, Goldman M, Fischler D, Goldman C, Schechter C, Levin S, et al. Respiratory symptoms and physiologic assessment of ironworkers at the World Trade Center disaster site. Chest. 2004; 125: 1248–55.
- Mauer MP, Cummings KR, Hoen R. Long-term respiratory symptoms in World Trade Center responders. Occup Med. 2010; 60: 145–51.
- Becklake MR, Leclerc M, Strobach H, Swift J. The N2 closing volume test in population studies: sources of variation and reproducibility. Am Rev Respir Dis. 1975; 111: 141–7.
- Verbanck S, Paiva M. Gas mixing in the airways and airspaces. Comprehensive Physiology. John Wiley & Sons; 2011. Available from: http://onlinelibrary.wiley.com/doi/10.1002/cphy. c100018/abstract [cited 8 August 2013].
- Bourdin A, Paganin F, Préfaut C, Kieseler D, Godard P, Chanez P. Nitrogen washout slope in poorly controlled asthma. Allergy. 2006; 61: 85–9.
- In 't Veen JC, Beekman AJ, Bel EH, Sterk PJ. Recurrent exacerbations in severe asthma are associated with enhanced airway closure during stable episodes. Am J Respir Crit Care Med. 2000; 161: 1902–6.
- Battaglia S, den Hertog H, Timmers MC, Lazeroms SPG, Vignola AM, Rabe KF, et al. Small airways function and molecular markers in exhaled air in mild asthma. Thorax. 2005; 60: 639–44.
- van Veen IH, Sterk PJ, Schot R, Gauw SA, Rabe KF, Bel EH. Alveolar nitric oxide versus measures of peripheral airway dysfunction in severe asthma. Eur Respir J. 2006; 27: 951–6.
- Corda L, Gardenghi GG, Modina D, Montemurro LT, Novali M, Tantucci C. Effects on small airway obstruction of long-term treatments with beclomethasone/formoterol

hydrofluoroalkane (metered-dose inhaler) versus fluticasone/ salmeterol (dry-powder inhaler) in asthma: a preliminary study. Allergy Asthma Proc. 2011; 32: 29–34.

- Langley SJ, Houghton CM, Singh SD. Bronchoprotective and bronchodilator effects of an HFA pMDI vs. a CFC pMDI and a DPI containing formoterol in asthma patients. Respir Int Rev Thorac Dis. 2005; 72(Suppl 1): 35–7.
- Cohen J, Douma WR, ten Hacken NHT, Vonk JM, Oudkerk M, Postma DS. Ciclesonide improves measures of small airway involvement in asthma. Eur Respir J. 2008; 31: 1213–20.
- Zeidler MR, Kleerup EC, Goldin JG, Kim HJ, Truong DA, Simmons MD, et al. Montelukast improves regional airtrapping due to small airways obstruction in asthma. Eur Respir J. 2006; 27: 307–15.
- 84. Lapperre TS, Willems LNA, Timens W, Rabe KF, Hiemstra PS, Postma DS, et al. Small airways dysfunction and neutro-philic inflammation in bronchial biopsies and BAL in COPD. Chest J. 2007; 131: 53–9.
- Gennimata S-A, Palamidas A, Karakontaki F, Kosmas EN, Koutsoukou A, Loukides S, et al. Pathophysiology of evolution of small airways disease to overt COPD. COPD. 2010; 7: 269–75.
- Fregonese L, van Veen HP, Sterk PJ, Stolk J. Ventilation inhomogeneity in alpha1-antitrypsin-deficient emphysema. Eur Respir J. 2006; 28: 323–9.
- Corbin RP, Loveland M, Martin RR, Macklem PT. A fouryear follow-up study of lung mechanics in smokers. Am Rev Respir Dis. 1979; 120: 293–304.
- Buist AS, Vollmer WM, Johnson LR, McCamant LE. Does the single-breath N2 test identify the smoker who will develop chronic airflow limitation? Am Rev Respir Dis. 1988; 137: 293– 301.
- Stănescu D, Sanna A, Veriter C, Robert A. Identification of smokers susceptible to development of chronic airflow limitation: a 13-year follow-up. Chest. 1998; 114: 416–25.
- Robinson PD, Goldman MD, Gustafsson PM. Inert gas washout: theoretical background and clinical utility in respiratory disease. Respiration. 2009; 78: 339–55.
- Horsley A. Lung clearance index in the assessment of airways disease. Respir Med. 2009; 103: 793–9.
- Crawford AB, Makowska M, Paiva M, Engel LA. Convectionand diffusion-dependent ventilation maldistribution in normal subjects. J Appl Physiol 1985; 59: 838–46.
- Verbanck S, Schuermans D, Meysman M, Paiva M, Vincken W. Noninvasive assessment of airway alterations in smokers: the small airways revisited. Am J Respir Crit Care Med. 2004; 170: 414–9.
- Verbanck S, Schuermans D, Paiva M, Meysman M, Vincken W. Small airway function improvement after smoking cessation in smokers without airway obstruction. Am J Respir Crit Care Med. 2006; 174: 853–7.
- Verbanck S, Schuermans D, Van Muylem A, Melot C, Noppen M, Vincken W, et al. Conductive and acinar lung-zone contributions to ventilation inhomogeneity in COPD. Am J Respir Crit Care Med. 1998; 157: 1573–7.
- Verbanck S, Schuermans D, Noppen M, Van Muylem A, Paiva M, Vincken W. Evidence of acinar airway involvement in asthma. Am J Respir Crit Care Med. 1999; 159: 1545–50.
- Verbanck S, Schuermans D, Vincken W. Inflammation and airway function in the lung periphery of patients with stable asthma. J Allergy Clin Immunol. 2010; 125: 611–6.
- Thompson BR, Douglass JA, Ellis MJ, Kelly VJ, O'Hehir RE, King GG, et al. Peripheral lung function in patients with stable and unstable asthma. J Allergy Clin Immunol. 2013; 131: 1322–8.

- Farah CS, King GG, Brown NJ, Peters MJ, Berend N, Salome CM. Ventilation heterogeneity predicts asthma control in adults following inhaled corticosteroid dose titration. J Allergy Clin Immunol. 2012; 130: 61–8.
- 100. Farah CS, King GG, Brown NJ, Downie SR, Kermode JA, Hardaker KM, et al. The role of the small airways in the clinical expression of asthma in adults. J Allergy Clin Immunol. 2012; 129: 381–7, 387.e1.
- Verbanck S, Schuermans D, Paiva M, Vincken W. The functional benefit of anti-inflammatory aerosols in the lung periphery. J Allergy Clin Immunol. 2006; 118: 340–6.
- Verbanck S, Schuermans D, Paiva M, Vincken W. Nonreversible conductive airway ventilation heterogeneity in mild asthma. J Appl Physiol (1985). 2003; 94: 1380–6.
- 103. Verbanck S, Schuermans D, Vincken W. Small airways ventilation heterogeneity and hyperinflation in COPD: response to tiotropium bromide. Int J Chron Obstruct Pulmon Dis. 2007; 2: 625–34.
- 104. Downie SR, Salome CM, Verbanck S, Thompson B, Berend N, King GG. Ventilation heterogeneity is a major determinant of airway hyperresponsiveness in asthma, independent of airway inflammation. Thorax. 2007; 62: 684–9.
- 105. King GG, Downie SR, Verbanck S, Thorpe CW, Berend N, Salome CM, et al. Effects of methacholine on small airway function measured by forced oscillation technique and multiple breath nitrogen washout in normal subjects. Respir Physiol Neurobiol. 2005; 148: 165–77.
- Van Muylem A, Verbanck S, Estenne M. Monitoring the lung periphery of transplanted lungs. Respir Physiol Neurobiol. 2005; 148: 141–51.
- 107. Aurora P, Kozlowska W, Stocks J. Gas mixing efficiency from birth to adulthood measured by multiple-breath washout. Respir Physiol Neurobiol. 2005; 148: 125–39.
- 108. Gustafsson PM, Aurora P, Lindblad A. Evaluation of ventilation maldistribution as an early indicator of lung disease in children with cystic fibrosis. Eur Respir J. 2003; 22: 972–9.
- 109. Amin R, Subbarao P, Lou W, Jabar A, Balkovec S, Jensen R, et al. The effect of dornase alfa on ventilation inhomogeneity in patients with cystic fibrosis. Eur Respir J. 2011; 37: 806–12.
- 110. Barnes PJ, Dweik RA, Gelb AF, Gibson PG, George SC, Grasemann H, et al. Exhaled nitric oxide in pulmonary diseases: a comprehensive review. Chest J. 2010; 138: 682–92.
- 111. Silkoff PE, McClean PA, Slutsky AS, Furlott HG, Hoffstein E, Wakita S, et al. Marked flow-dependence of exhaled nitric oxide using a new technique to exclude nasal nitric oxide. Am J Respir Crit Care Med. 1997; 155: 260–7.
- Tsoukias NM, George SC. A two-compartment model of pulmonary nitric oxide exchange dynamics. J Appl Physiol (1985). 1998; 85: 653–66.
- 113. Eckel SP, Linn WS, Berhane K, Rappaport EB, Salam MT, Zhang Y, et al. Estimation of parameters in the two-compartment model for exhaled nitric oxide. PLoS One. 2014; 9: e85471.
- 114. Paredi P, Kharitonov SA, Meah S, Barnes PJ, Usmani OS. A novel approach to partition central and peripheral airway nitric oxide. Chest. 2014; 145: 113–9.
- 115. Lehtimäki L, Kankaanranta H, Saarelainen S, Hahtola P, Järvenpää R, Koivula T, et al. Extended exhaled NO measurement differentiates between alveolar and bronchial inflammation. Am J Respir Crit Care Med. 2001; 163: 1557– 61.
- Shin H-W, George SC. Impact of axial diffusion on nitric oxide exchange in the lungs. J Appl Physiol (1985). 2002; 93: 2070– 80.
- 117. Shin H-W, Condorelli P, Rose-Gottron CM, Cooper DM, George SC. Probing the impact of axial diffusion on nitric

oxide exchange dynamics with heliox. J Appl Physiol (1985). 2004; 97: 874–82.

- Verbanck S, Kerckx Y, Schuermans D, Vincken W, Paiva M, Van Muylem A. Effect of airways constriction on exhaled nitric oxide. J Appl Physiol (1985). 2008; 104: 925–30.
- 119. Gelb AF, George SC, Silkoff PE, Krishnan A, Fraser C, Taylor CF, et al. Central and peripheral airway/alveolar sites of exhaled nitric oxide in acute asthma. Thorax. 2010; 65: 619– 25.
- 120. Berry M, Hargadon B, Morgan A, Shelley M, Richter J, Shaw D, et al. Alveolar nitric oxide in adults with asthma: evidence of distal lung inflammation in refractory asthma. Eur Respir J. 2005; 25: 986–91.
- 121. Van Veen IH, Sterk PJ, Schot R, Gauw SA, Rabe KF, Bel EH. Alveolar nitric oxide versus measures of peripheral airway dysfunction in severe asthma. Eur Respir J. 2006; 27: 951–6.
- Scichilone N, Battaglia S, Taormina S, Modica V, Pozzecco E, Bellia V. Alveolar nitric oxide and asthma control in mild untreated asthma. J Allergy Clin Immunol. 2013; 131: 1513–7.
- 123. Massaro AF, Gaston B, Kita D, Fanta C, Stamler JS, Drazen JM. Expired nitric oxide levels during treatment of acute asthma. Am J Respir Crit Care Med. 1995; 152: 800–3.
- 124. Yates DH, Kharitonov SA, Robbins RA, Thomas PS, Barnes PJ. Effect of a nitric oxide synthase inhibitor and a glucocorticosteroid on exhaled nitric oxide. Am J Respir Crit Care Med. 1995; 152: 892–6.
- 125. Silkoff PE, McClean PA, Slutsky AS, Caramori M, Chapman KR, Gutierrez C, et al. Exhaled nitric oxide and bronchial reactivity during and after inhaled beclomethasone in mild asthma. J Asthma. 1998; 35: 473–9.
- 126. Malinovschi A, Van Muylem A, Michiels S, Michils A. FeNO as a predictor of asthma control improvement after starting inhaled steroid treatment. Nitric Oxide. 2014; 40: 110–6.
- 127. Petsky HL, Cates CJ, Li A, Kynaston JA, Turner C, Chang AB. Tailored interventions based on exhaled nitric oxide versus clinical symptoms for asthma in children and adults. Cochrane Database Syst Rev. 2009; (4): CD006340.
- 128. Petsky HL, Cates CJ, Lasserson TJ, Li AM, Turner C, Kynaston JA, et al. A systematic review and meta-analysis: tailoring asthma treatment on eosinophilic markers (exhaled nitric oxide or sputum eosinophils). Thorax. 2012; 67: 199– 208.
- 129. Ansarin K, Chatkin JM, Ferreira IM, Gutierrez CA, Zamel N, Chapman KR. Exhaled nitric oxide in chronic obstructive pulmonary disease: relationship to pulmonary function. Eur Respir J. 2001; 17: 934–8.
- Maziak W, Loukides S, Culpitt S, Sullivan P, Kharitonov SA, Barnes PJ. Exhaled nitric oxide in chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 1998; 157: 998–1002.
- Corradi M, Majori M, Cacciani GC, Consigli GF, de' Munari E, Pesci A. Increased exhaled nitric oxide in patients with stable chronic obstructive pulmonary disease. Thorax. 1999; 54: 572–5.
- 132. Kunisaki KM, Rice KL, Janoff EN, Rector TS, Niewoehner DE. Exhaled nitric oxide, systemic inflammation, and the spirometric response to inhaled fluticasone propionate in severe chronic obstructive pulmonary disease: a prospective study. Ther Adv Respir Dis. 2008; 2: 55–64.
- 133. Papi A, Romagnoli M, Baraldo S, Braccioni F, Guzzinati I, Saetta M, et al. Partial reversibility of airflow limitation and increased exhaled NO and sputum eosinophilia in chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2000; 162: 1773–7.

- Hackx M, Bankier AA, Gevenois PA. Chronic obstructive pulmonary disease: CT quantification of airways disease. Radiology. 2012; 265: 34–48.
- 135. McDonough JE, Yuan R, Suzuki M, Seyednejad N, Elliott WM, Sanchez PG, et al. Small-airway obstruction and emphysema in chronic obstructive pulmonary disease. N Engl J Med. 2011; 365: 1567–75.
- 136. Nakano Y, Wong JC, de Jong PA, Buzatu L, Nagao T, Coxson HO, et al. The prediction of small airway dimensions using computed tomography. Am J Respir Crit Care Med. 2005; 171: 142–6.
- 137. Nakano Y, Muro S, Sakai H, Hirai T, Chin K, Tsukino M, et al. Computed tomographic measurements of airway dimensions and emphysema in smokers. Correlation with lung function. Am J Respir Crit Care Med. 2000; 162: 1102–8.
- Berger P, Perot V, Desbarats P, Tunon-de-Lara JM, Marthan R, Laurent F. Airway wall thickness in cigarette smokers: quantitative thin-section CT assessment. Radiology. 2005; 235: 1055–64.
- 139. Hasegawa M, Nasuhara Y, Onodera Y, Makita H, Nagai K, Fuke S, et al. Airflow limitation and airway dimensions in chronic obstructive pulmonary disease. Am J Respir Crit Care Med. 2006; 173: 1309–15.
- 140. Ohara T, Hirai T, Sato S, Sato A, Nishioka M, Muro S, et al. Comparison of airway dimensions in different anatomic locations on chest CT in patients with COPD. Respirol Carlton Vic. 2006; 11: 579–85.
- 141. Laurent F, Latrabe V, Raherison C, Marthan R, Tunon-de-Lara JM. Functional significance of air trapping detected in moderate asthma. Eur Radiol. 2000; 10: 1404–10.
- 142. Gupta P, Yadav R, Verma M, Agarwal D, Kumar M. Correlation between high-resolution computed tomography features and patients' characteristics in chronic obstructive pulmonary disease. Ann Thorac Med. 2008; 3: 87.
- 143. Hersh CP, Washko GR, Estépar RSJ, Lutz S, Friedman PJ, Han MK, et al. Paired inspiratory-expiratory chest CT scans to assess for small airways disease in COPD. Respir Res. 2013; 14: 42.
- 144. Ueda T, Niimi A, Matsumoto H, Takemura M, Hirai T, Yamaguchi M, et al. Role of small airways in asthma: investigation using high-resolution computed tomography. J Allergy Clin Immunol. 2006; 118: 1019–25.
- 145. Jain N, Covar RA, Gleason MC, Newell JD, Gelfand EW, Spahn JD. Quantitative computed tomography detects peripheral airway disease in asthmatic children. Pediatr Pulmonol. 2005; 40: 211–8.
- 146. Mitsunobu F, Ashida K, Hosaki Y, Tsugeno H, Okamoto M, Nishida N, et al. Decreased computed tomographic lung density during exacerbation of asthma. Eur Respir J. 2003; 22: 106–12.
- 147. Busacker A, Newell JD, Keefe T, Hoffman EA, Granroth JC, Castro M, et al. A multivariate analysis of risk factors for the air-trapping asthmatic phenotype as measured by quantitative CT analysis. Chest. 2009; 135: 48–56.
- 148. Goldin JG, Tashkin DP, Kleerup EC, Greaser LE, Haywood UM, Sayre JW, et al. Comparative effects of hydrofluoroalkane and chlorofluorocarbon beclomethasone dipropionate inhalation on small airways: assessment with functional helical thinsection computed tomography. J Allergy Clin Immunol. 1999; 104: S258–67.
- 149. Tunon-de-Lara J-M, Laurent F, Giraud V, Perez T, Aguilaniu B, Meziane H, et al. Air trapping in mild and moderate asthma: effect of inhaled corticosteroids. J Allergy Clin Immunol. 2007; 119: 583–90.
- 150. Eda S, Kubo K, Fujimoto K, Matsuzawa Y, Sekiguchi M, Sakai F. The relations between expiratory chest CT using

helical CT and pulmonary function tests in emphysema. Am J Respir Crit Care Med. 1997; 155: 1290–4.

- 151. Schroeder JD, McKenzie AS, Zach JA, Wilson CG, Curran-Everett D, Stinson DS, et al. Relationships between airflow obstruction and quantitative CT measurements of emphysema, air trapping, and airways in subjects with and without chronic obstructive pulmonary disease. Am J Roentgenol. 2013; 201: W460–70.
- 152. Hartley R, Barker B, Edwards K, Finch J, Shelley M, Parker S, et al. Quantitative CT in COPD MAP: emphysema and small airways disease independently contribute to FEV1. Eur Respir J. 2013; 42(Suppl 57): P2269.
- 153. Bommart S, Marin G, Bourdin A, Molinari N, Klein F, Hayot M, et al. Relationship between CT air trapping criteria and lung function in small airway impairment quantification. BMC Pulm Med. 2014; 14: 29.
- 154. De Backer JW, Vos WG, Vinchurkar SC, Claes R, Drollmann A, Wulfrank D, et al. Validation of computational fluid dynamics in CT-based airway models with SPECT/CT. Radiology. 2010; 257: 854–62.
- 155. Galbán CJ, Han MK, Boes JL, Chughtai KA, Meyer CR, Johnson TD, et al. Computed tomography-based biomarker provides unique signature for diagnosis of COPD phenotypes and disease progression. Nat Med. 2012; 18: 1711–5.
- 156. Wijesuriya S, Chandratreya L, Medford AR. Chronic pulmonary emboli and radiologic mimics on CT pulmonary angiography: a diagnostic challenge. Chest. 2013; 143: 1460– 71.
- 157. Van Beek EJR, Wild JM. Hyperpolarized 3-helium magnetic resonance imaging to probe lung function. Proc Am Thorac Soc. 2005; 2: 528–32.
- 158. Van Beek EJR, Hoffman EA. Imaging in COPD. Imaging Decis MRI. 2009; 13: 11–7.
- 159. Kauczor HU, Ebert M, Kreitner KF, Nilgens H, Surkau R, Heil W, et al. Imaging of the lungs using 3He MRI: preliminary clinical experience in 18 patients with and without lung disease. J Magn Reson Imaging. 1997; 7: 538–43.
- 160. De Lange EE, Mugler JP, Brookeman JR, Knight-Scott J, Truwit JD, Teates CD, et al. Lung air spaces: MR imaging evaluation with hyperpolarized 3He gas. Radiology. 1999; 210: 851–7.
- 161. Salerno M, de Lange EE, Altes TA, Truwit JD, Brookeman JR, Mugler JP. Emphysema: hyperpolarized helium 3 diffusion MR imaging of the lungs compared with spirometric indexes initial experience. Radiology. 2002; 222: 252–60.
- 162. Yablonskiy DA, Sukstanskii AL, Leawoods JC, Gierada DS, Bretthorst GL, Lefrak SS, et al. Quantitative in vivo assessment of lung microstructure at the alveolar level with hyperpolarized 3He diffusion MRI. Proc Natl Acad Sci. 2002; 99: 3111–6.
- 163. Diaz S, Casselbrant I, Piitulainen E, Magnusson P, Peterson B, Wollmer P, et al. Validity of apparent diffusion coefficient hyperpolarized 3He-MRI using MSCT and pulmonary function tests as references. Eur J Radiol. 2009; 71: 257–63.
- 164. Kirby M, Mathew L, Wheatley A, Santyr GE, McCormack DG, Parraga G. Chronic obstructive pulmonary disease: longitudinal hyperpolarized 3He MR imaging. Radiology. 2010; 256: 280–9.
- 165. Woods JC, Yablonskiy DA, Choong CK, Chino K, Pierce JA, Hogg JC, et al. Long-range diffusion of hyperpolarized 3He in explanted normal and emphysematous human lungs via magnetization tagging. J Appl Physiol. 2005; 99: 1992–7.
- 166. Marshall H, Deppe MH, Parra-Robles J, Hillis S, Billings CG, Rajaram S, et al. Direct visualisation of collateral ventilation in COPD with hyperpolarised gas MRI. Thorax. 2012; 67: 613–7.

- 167. De Lange EE, Altes TA, Patrie JT, Parmar J, Brookeman JR, Mugler III JP, et al. The variability of regional airflow obstruction within the lungs of patients with asthma: assessment with hyperpolarized helium-3 magnetic resonance imaging. J Allergy Clin Immunol. 2007; 119: 1072–8.
- 168. Tustison NJ, Altes TA, Song G, de Lange EE, Mugler JP, Gee JC. Feature analysis of hyperpolarized helium-3 pulmonary MRI: a study of asthmatics versus nonasthmatics. Magn Reson Med. 2010; 63: 1448–55.
- 169. Parraga G, Ouriadov A, Evans A, McKay S, Lam WW, Fenster A, et al. Hyperpolarized 3He ventilation defects and apparent diffusion coefficients in chronic obstructive pulmonary disease: preliminary results at 3.0 Tesla. Invest Radiol. 2007; 42: 384–91.
- 170. Mathew L, Evans A, Ouriadov A, Etemad-Rezai R, Fogel R, Santyr G, et al. Hyperpolarized 3He magnetic resonance imaging of chronic obstructive pulmonary disease: reproducibility at 3.0 tesla. Acad Radiol. 2008; 15: 1298–311.
- 171. Fain SB, Gonzalez-Fernandez G, Peterson ET, Evans MD, Sorkness RL, Jarjour NN, et al. Evaluation of structurefunction relationships in asthma using multidetector CT and hyperpolarized He-3 MRI. Acad Radiol. 2008; 15: 753–62.
- 172. Wild JM, Paley MNJ, Kasuboski L, Swift A, Fichele S, Woodhouse N, et al. Dynamic radial projection MRI of inhaled hyperpolarized 3He gas. Magn Reson Med. 2003; 49: 991–7.
- 173. Holmes JH, Korosec FR, Du J, O'Halloran RL, Sorkness RL, Grist TM, et al. Imaging of lung ventilation and respiratory dynamics in a single ventilation cycle using hyperpolarized He-3 MRI. J Magn Reson Imaging. 2007; 26: 630–6.
- 174. Newman SP, Pitcairn GR, Hirst PH. A brief history of gamma scintigraphy. J Aerosol Med. 2001; 14: 139–45.
- 175. Bondesson E, Bengtsson T, Borgström L, Nilsson L-E, Norrgren K, Olsson B, et al. Dose delivery late in the breath can increase dry powder aerosol penetration into the lungs. J Aerosol Med. 2005; 18: 23–33.
- 176. Newman SP. Lung distribution of inhaled drugs. Br J Clin Pharmacol. 2001; 52: 716–7.
- 177. Fleming J, Conway J, Majoral C, Tossici-Bolt L, Katz I, Caillibotte G, et al. The use of combined single photon emission computed tomography and X-ray computed tomography to assess the fate of inhaled aerosol. J Aerosol Med Pulm Drug Deliv. 2011; 24: 49–60.
- 178. Roach PJ, Schembri GP, Bailey DL. V/Q scanning using SPECT and SPECT/CT. J Nucl Med. 2013; 54: 1588–96.
- 179. Amis TC, Crawford AB, Davison A, Engel LA. Distribution of inhaled 99mtechnetium labelled ultrafine carbon particle aerosol (Technegas) in human lungs. Eur Respir J. 1990; 3: 679–85.
- Crawford AB, Davison A, Amis TC, Engel LA. Intrapulmonary distribution of 99mtechnetium labelled ultrafine carbon aerosol (Technegas) in severe airflow obstruction. Eur Respir J. 1990; 3: 686–92.
- 181. Pellegrino R, Biggi A, Papaleo A, Camuzzini G, Rodarte JR, Brusasco V. Regional expiratory flow limitation studied with Technegas in asthma. J Appl Physiol (1985). 2001; 91: 2190–8.
- 182. Nagao M, Murase K, Ichiki T, Sakai S, Yasuhara Y, Ikezoe J. Quantitative analysis of technegas SPECT: evaluation of regional severity of emphysema. J Nucl Med. 2000; 41: 590–5.
- 183. Nagao M, Murase K. Measurement of heterogeneous distribution on Technegas SPECT images by three-dimensional fractal analysis. Ann Nucl Med. 2002; 16: 369–76.
- 184. King GG. Cutting edge technologies in respiratory research: lung function testing. Respirology. 2011; 16: 883–90.

- 185. Leach CL, Bethke TD, Boudreau RJ, Hasselquist BE, Drollmann A, Davidson P, et al. Two-dimensional and threedimensional imaging show ciclesonide has high lung deposition and peripheral distribution: a nonrandomized study in healthy volunteers. J Aerosol Med. 2006; 19: 117–26.
- 186. Newman S, Salmon A, Nave R, Drollmann A. High lung deposition of 99mTc-labeled ciclesonide administered via HFA-MDI to patients with asthma. Respir Med. 2006; 100: 375–84.
- Eberl S, Chan H-K, Daviskas E. SPECT imaging for radioaerosol deposition and clearance studies. J Aerosol Med. 2006; 19: 8–20.
- 188. Dolovich MB, Bailey DL. Positron emission tomography (PET) for assessing aerosol deposition of orally inhaled drug

products. J Aerosol Med Pulm Drug Deliv. 2012; 25(Suppl 1): S52–71.

- 189. Jones HA, Marino PS, Shakur BH, Morrell NW. In vivo assessment of lung inflammatory cell activity in patients with COPD and asthma. Eur Respir J. 2003; 21: 567–73.
- 190. Harris RS, Winkler T, Tgavalekos N, Musch G, Melo MFV, Schroeder T, et al. Regional pulmonary perfusion, inflation, and ventilation defects in bronchoconstricted patients with asthma. Am J Respir Crit Care Med. 2006; 174: 245–53.
- 191. Biddiscombe MF, Usmani OS. The importance of imaging and physiology measurements in assessing the delivery of peripherally targeted aerosolized drugs. Ther Deliv. 2012; 3: 1329–45.
- Usmani OS. Treating the small airways. Respir Int Rev Thorac Dis. 2012; 84: 441–53.