

# Lentiviral transduction of rat Sertoli cells as a means to modify gene expression

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Primary cell culture is an established and widely used technique to study Sertoli cell function in vitro. However, the relative difficulty of stably overexpressing or knocking down genes in Sertoli cell culture has limited progress in the field. In this technical report, we present a method to transduce 20 dpp rat Sertoli cell cultures with VSV-G pseudotyped lentiviral based vectors at a high rate (~80%), with stable reporter gene expression. Although high transgene expression is desirable, it was noted that at transduction rates > 60% inter-Sertoli cell tight junction integrity and, hence, Sertoli cell function, were transiently compromised. We envisage that this optimized procedure has the potential to stimulate Sertoli cell research, and motivate the use of Sertoli cells in various cell therapy applications.

## Introduction

The somatic Sertoli cell of the testis provides structural and nutritional support to germ cells in the process of spermatogenesis. The specialized “nurse” functions of the Sertoli cell has been studied extensively in vitro since the first isolation and culture of primary cells in the 1970s.<sup>1-3</sup> Various techniques to manipulate Sertoli cell gene expression, including transfection have been used,<sup>4,5</sup> but the efficiency nor longevity of these methods is rarely reported as they do not lead to stable genomic integration.

Recently, adenovirus vectors have been used to transduce Sertoli cells in vivo,<sup>6,7</sup> although low rates of transduction and cellular toxicity were noted. An alternative virus of the *retroviridae* family, *lentivirus*, possess the inherent capacity to transduce and stably integrate into non-dividing cells.<sup>8,9</sup> Recently, retroviral transduction was used to overexpress a panel of neuronal transcription factors in proliferative phase Sertoli cells, directing the cells toward an induced neural stem/progenitor lineage.<sup>10</sup> This study demonstrated the viability and potential use of retroviral vector transgene expression in primary juvenile Sertoli cells. Lentiviral based vectors have also been used to transduce Sertoli cells in an elegant in vivo study, in which the overexpression *kit* ligand in *c-kit* ligand null mice was able to restore fertility.<sup>6</sup>

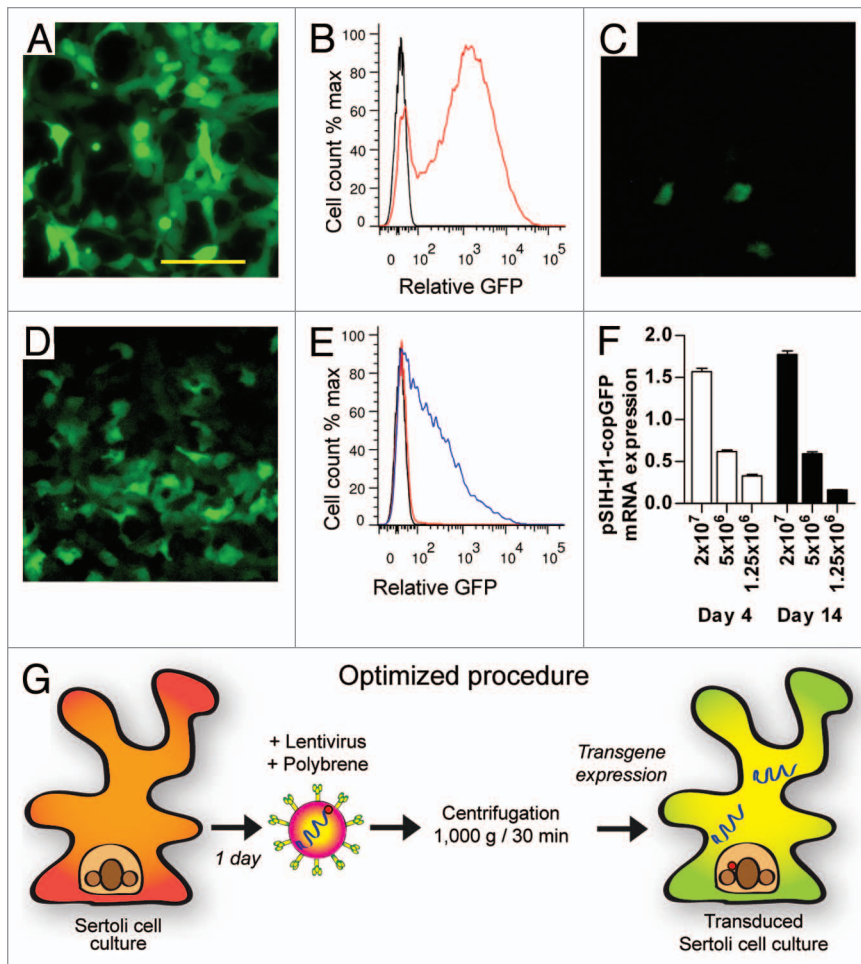
Our laboratory has extensive experience in primary rat Sertoli cell culture as a model to study the regulation of cell junctions,<sup>11,12</sup> growth factor signaling<sup>13</sup> and gene expression,<sup>14</sup> and we are currently utilizing a lentiviral based vector approach to manipulate Sertoli cell gene function in vitro. In early experiments, we noted that rat Sertoli cell monolayers were relatively inert to retroviral transduction. Herein, we describe a modified inoculation and

centrifugation protocol (“spinoculation”),<sup>15,16</sup> which considerably improves Sertoli cell transduction in vitro.

## Results

**Optimization of Sertoli cell transduction procedure.** To confirm that the lentiviral preparations could express transgene-GFP in a model cell line, HEK293T cells were transduced with limiting dilutions of a GFP-virus. Using a titer of  $5 \times 10^6$  TU/ml, robust GFP fluorescence was observed by live cell imaging (Fig. 1A) and quantitative flow cytometry (Fig. 1B) after 48 h. When a similar protocol was employed in primary Sertoli cells 72 h after isolation, using an identical viral titer of GFP-virus, we determined that transduction efficiency of primary Sertoli cells was very low by live cell imaging of reporter GFP fluorescence (Fig. 1C), which equated to 3% GFP+ cells by flow cytometry (Fig. 1E). To improve transduction efficiency in primary Sertoli cell cultures, we performed viral transduction in combination with centrifugation of the culture plate at 1,000 *g* for 30 min at room temperature in a procedure known as ‘spinoculation’.<sup>15</sup> This procedure initially improved transduction efficiency by 9-fold, as measured by genomic incorporation of the lentiviral GFP transfer vector by real-time PCR (Table 1,  $p < 0.01$ ). We next performed this viral transduction in combination with the poly-cationic reagent, polybrene (4  $\mu\text{g}/\text{ml}$ , Sigma), at the time of viral addition. The addition of polybrene resulted in a further improvement of transduction and genomic incorporation by 1.6-fold, to 14-fold above the initial level (Table 1,  $p = \text{ns}$ ). During primary Sertoli cell culture, tight junctions become established within 72 h of culture,<sup>12,13</sup> and we reasoned that tight junction formation may limit transduction efficiency. We subsequently

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**Figure 1.** Transduction of primary Sertoli cells using lentivirus vector. (A) Live reporter GFP fluorescence of HEK293T cells transduced at  $5 \times 10^6$  TU/ml (B) Flow cytometric analysis of control (no virus, black) and lentivirus transduced HEK293T cells (red). (C) Live GFP fluorescence of Sertoli cells transduced with  $5 \times 10^6$  TU/ml prior to and (D) after optimization. (E) Flow cytometric analysis of Sertoli cells prior to (red) and after (blue) optimization. (F) Expression of pSIH transfer vector normalized to  $\beta$ -actin in primary Sertoli cells 4 d (white) and 14 d (black) after transduction. (G) Experimental outline of optimized lentiviral vector transduction procedure in primary Sertoli cell culture. Scale bar = 50  $\mu$ m.

repeated viral transduction 24 h after commencement of cell culture, rather than after 72 h of culture, and noted a further improvement in genomic incorporation by 12.4-fold, with a total improvement in transduction efficiency of 222-fold above basal conditions (Table 1,  $p < 0.01$ ). Using alternative combinations of the optimization steps outlined above, we consistently observed that the combination of centrifugation, polybrene and addition of lentiviral vectors 24 h after the initial culture produced the greatest enhancement in viral transduction (data not shown). Live cell imaging (Fig. 1D) and flow cytometry (Fig. 1E) confirmed that the increased genomic transduction under optimized conditions also resulted in a significant improvement in the proportion of reporter GFP-fluorescent cells (from 3 to 64%, Fig. 1E).

Stability of DNA incorporation was determined by examining the expression of the lentiviral transfer vector in transduced primary Sertoli cells at 3 and 14 d (Fig. 1F). Using a range of viral titers, expression was dependent upon lentiviral

titer administration, but remained consistent across this time course. An outline of the optimized transduction procedure is presented in Figure 1G.

**Effect of high transduction rate upon tight junction integrity.** Flow cytometric analysis of reporter GFP fluorescence indicated that our optimized procedure resulted in Sertoli cell transduction rates  $\sim 80\%$  at high titer ( $2 \times 10^7$  TU/ml, Fig. 2A). We noted that increased lentiviral vector titer allowed for a parallel increase in reporter gene expression (Fig. 2A). As a surrogate measure of Sertoli cell function, we assessed the integrity of tight junctions following lentiviral vector transduction by real-time measurement of transepithelial electrical resistance (TER).<sup>13</sup> Interestingly, monolayers with a high titer of virus and reporter GFP fluorescence exhibited reduced tight junction integrity (Fig. 2A). When tight junction function was assessed over an eight-day period, TER function only recovered to control levels with titers  $\leq 5 \times 10^6$  TU/ml after one week (Fig. 2B).

Sertoli cell tight junctions are composed of several transmembrane proteins, including Claudin-11 (Cldn11), bound to intracellular plaque proteins, such as zona occludens-1 (Tjp1). To determine the effect of lentiviral vector transduction upon Sertoli cell tight junctions, cell monolayers were then visualized by the co-localization of reporter GFP fluorescence with the somatic nuclear marker, Gata4, and the Sertoli cell tight junction-related proteins Tjp1 and Cldn11. Reporter gene intensity appeared highly variable between adjacent Sertoli cells (Fig. 2C–F), consistent with a broad range of GFP reporter fluorescence observed by flow cytometry (Fig.

1E). Immunocytochemical co-localization with Tjp1 indicated a normal localization, consistent with inter-Sertoli cell tight junctions (Fig. 2C and D). However, Cldn11 immunostaining appeared reduced in cells treated with high titer lentiviral vector (Fig. 2E and F). Of particular note, Cldn11 was reduced at inter-Sertoli cell contacts between transduced (GFP, green) cells (Fig. 2F), suggesting that the reduced TER following high titer lentiviral vector administration (Fig. 2A and B) may be mediated through Cldn11.

## Discussion

**Use of lentiviral vector transduction to study Sertoli cell gene function.** The data presented in this technical report validate lentiviral based vectors as a feasible methodology to manipulate rat Sertoli cell gene function in vitro. Using this optimized procedure, we achieve robust and stable expression of a GFP transgene

**Table 1.** Fold increase in genomic incorporation of lentiviral transfer vector by qPCR

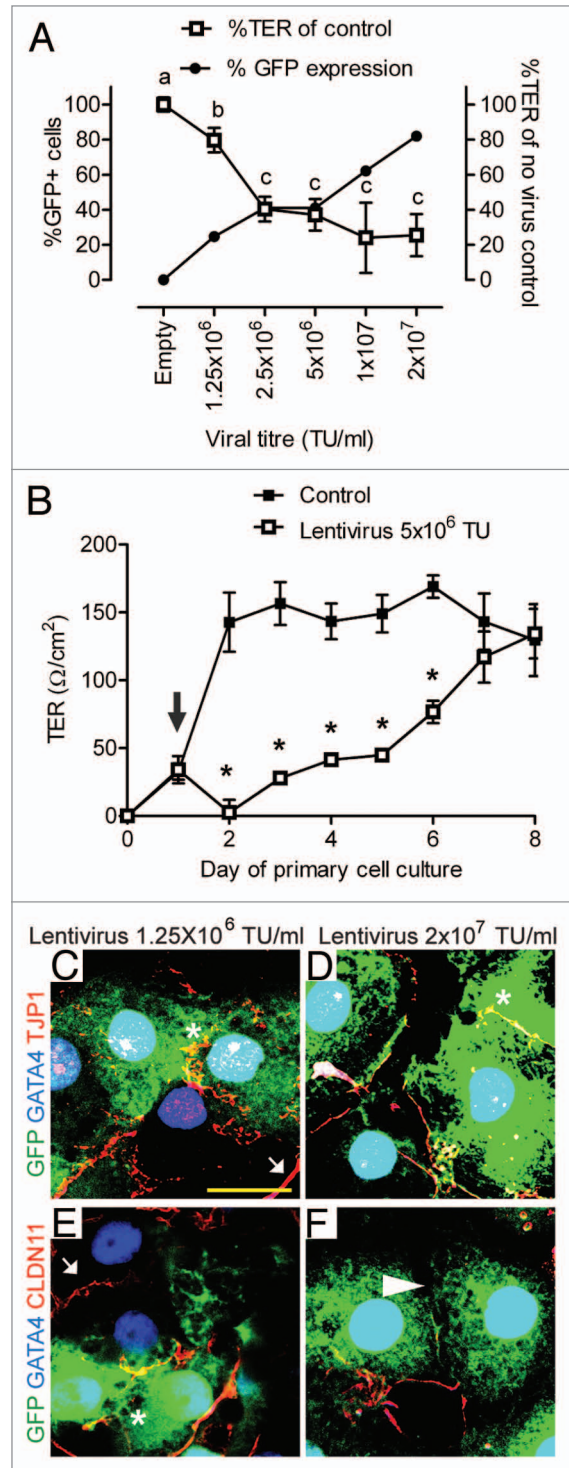
	Basal conditions (at 72 h)	+ Spin (spinoculation) (at 72 h)	+ Polybrene + Spin (at 72 h)	+ Polybrene + Spin (at 24 h)
Total fold improvement	1.0 <sup>a</sup>	9.0 ± 4.0 <sup>b</sup>	14.0 ± 4.5 <sup>b</sup>	222 ± 85.0 <sup>c</sup>

Optimization and fold improvement of lentiviral based vector transduction by real-time PCR using primers specific to the transfer plasmid and normalized to genomic  $\beta$ -actin. A significant improvement in transduction is observed under optimized conditions. Mean  $\pm$  SD,  $n > 3$ , letters denote significant difference  $p < 0.001$ .

**Figure 2.** Effect of *lentivirus* administration on inter-Sertoli cell tight junction. **(A)** Transepithelial electrical resistance (%TER of control monolayers, white boxes) taken 48 h after Sertoli cell transduction with between  $1.25 \times 10^6$  and  $2 \times 10^7$  TU/ml. GFP reporter fluorescence was determined by flow cytometry (% GFP+ cells, flow cytometry, black circles). Increased *lentivirus* addition (and increased %GFP+ cells) is associated with loss of TER. TER data are mean,  $\pm$  SD,  $n = 3$ , letters denote significant difference  $p < 0.01$ . **(B)** Daily TER of control (black boxes) and *lentivirus* transduced cells ( $5 \times 10^6$  TU/ml, white boxes) over 8 d culture period. Black arrow indicates *lentivirus* addition 24 h after primary cell isolation. TER data are mean,  $\pm$  SD,  $n = 3$ , \*  $p < 0.01$  between control and transduced cells. **(C–F)** Co-localization of endogenous reporter gene expression (GFP, green), Gata4 (blue) and tight junction protein (red); **(C and D)** Tjp1 and **(E and F)** Cldn11 with  $1.25 \times 10^6$  or  $2 \times 10^7$  TU/ml. White arrow indicates TJ co-localization between non-transduced cells, \* indicates TJ localization between transduced (GFP) cells, and white arrowhead in **(F)** indicates reduced Cldn11 localization between adjacent transduced cells after high viral titer addition. Scale bar = 20  $\mu$ m.

in primary Sertoli cells through two weeks of culture. Our optimized procedure likely results from an enhanced interaction between lentivirus vectors and Sertoli cells,<sup>16</sup> with multiple vectors likely transducing each cell at high titer. Given that lentiviral based vectors are susceptible to gene silencing,<sup>23</sup> the increased rate of incorporation achieved with high titer likely overcomes silencing phenomena to allow for more consistent gene expression across individual cells.

The improved efficiency of viral transduction described following “spinoculation” represents a significant improvement upon basal transduction rates. Consistent with this, previous reports have described the necessity to undertake multiple rounds of infection of primary Sertoli cells to ensure high rates of transduction.<sup>10</sup> The proportion of transduced cells using this optimized methodology is much greater than previous reports which describe low rates of primary Sertoli cell transfection (25–38%) for overexpression of genes or luciferase assays.<sup>5,24</sup> We envisage that this optimized method could be exploited to overexpress genes implicated in Sertoli cell development and dysfunction in primary cell culture. This method may be particularly useful as an alternative to immortalized Sertoli-like cell lines, which do not fully recapitulate Sertoli cell gene expression and function.<sup>25</sup> Previous reports indicate that Sertoli cells express the gene silencing machinery required for siRNA mediated gene knockdown,<sup>4</sup> and consequently, the protocol outlined in this report could also be utilized to express an shRNA transcript to knockdown gene function in a differentiated Sertoli cell. Furthermore, miRNAs are highly expressed and hormonally-regulated in Sertoli cells, but their functional significance is largely unknown.<sup>14</sup> The protocol





described here could be easily adapted to overexpress specific miRNA hairpins, or miRNA inhibitors such as “sponges”<sup>26</sup> to define the role of these non-coding RNAs in Sertoli cell function.

Given that Sertoli cell tight junction function appears compromised at high viral titer, we aim to transduce 60% of Sertoli cells in culture to maintain optimal Sertoli cell function.

**Sertoli cell therapy.** Sertoli cell transplantation into diseased organs has been suggested as a cell therapy approach to chronic disease.<sup>27</sup> Analogous to their “nurse” cell function within the testis, transplanted Sertoli cells provide immunoprotective, and structural support functions within host tissues.<sup>28</sup> In a model of type I diabetes in mice, Sertoli cell therapy has been utilized to prevent, or revert the disease.<sup>29,30</sup> Furthermore, Sertoli cell transplantation into the striatum of the brain has been observed to reduce neuronal hyperactivity in a rodent model of early Huntington’s disease,<sup>31</sup> and have trophic effects upon dopaminergic neurons in Parkinson disease.<sup>32</sup>

The ability to engineer Sertoli cells using a retroviral approach may significantly improve the scope and feasibility of Sertoli cell therapy.<sup>27,33</sup> This approach was recently demonstrated with the induction of Sertoli cells into a functional, induced neuronal stem/progenitor cell population using virally engineered cells.<sup>10</sup> Furthermore, the integration of lentiviral based vectors into post-mitotic cells (such as differentiated Sertoli cells) offers significant safety advantage when compared with proliferative cells, including reduced integration into protein coding, and potentially oncogenic, regions of the genome.<sup>34</sup>

**Safety considerations.** Use of HIV-based vectors falls within the NIH biosafety Level 2, and as such, appropriate technical and physical safeguards should be put into practice to protect researchers. Due to the broad tropism of *lentivirus*, there is the potential for human transmission, and as such, it should be used under strict safety requirements specific to the host institution. Our laboratory utilizes a ‘3rd generation’ vector system, with several enhanced safety features compared with previous generations. Significantly, these vectors encode a self-inactivating (SIN) construct in the 3’ LTR, which prevents the production of wildtype viral particles by infected cells.<sup>35</sup> Novel commercially available lentiviral based vector plasmids now frequently utilize inducible promoters, which have enhanced safety and experimental benefits.

**Summary.** The generation of transgenic mice to selectively knock out Sertoli cell gene products is hampered by financial and time constraints, fertility defects, and by the limited access to inducible Sertoli cell transgenic mouse lines.<sup>36</sup> The technique presented in this technical report provides a novel, in-expensive, and rapid method to assess gene function in primary Sertoli cells. The ability to manipulate gene function in Sertoli cells in vitro is an important technical development, which will allow researchers to realize new advances in the field of Sertoli cell biology.

Using this transduction approach, we have stably expressed GFP in primary Sertoli cell cultures, and in un-published observations, confirmed that these cells express the translational and processing “machinery” for shRNA mediated gene knockdown. Finally, this lentiviral based approach could be used in a range

of applications, including engineering Sertoli cells for potential cell-based gene therapy.

## Methods

**Lentiviral vector production.** To produce VSV-G-pseudotyped lentiviral based vectors capable of expressing the green fluorescent protein (GFP) reporter gene, HEK293T cells were transfected with the minimal *lentivirus* genomic requirements, divided across four DNA plasmids, to produce lentiviral based vectors capable of cell transduction.<sup>17</sup> The transfer plasmid encoding reporter GFP protein used in these experiments was pSIH-H1-copGFP, with GFP expression under the control of a CMV promoter (System Biosciences). The remainder of the viral genome was encoded by an HIV *gag/pol* packaging plasmid, a *rev* expression plasmid, and the *VSV-G* expression plasmid.<sup>17</sup> Transfection of HEK293T cells at 60% confluence proceeded for 24 h, with subsequent medium replacement and virus collection 48 h later. Supernatant (50 ml) was filtered through 0.2  $\mu\text{m}$  filters, and virus concentrated by ultracentrifugal pelleting at 20,000  $g$  for 16 h at 4°C. The lentiviral based vector was resuspended in 100  $\mu\text{l}$  PBS and stored at -80°C. The functional titer of viral aliquots was assayed by serial limiting dilution on HEK293T cells and measurement of reporter gene expression (GFP) by flow cytometry to determine the number of transducing units (TU/ml) as previously described.<sup>18</sup> This viral production procedure regularly gave yields of  $\geq 1 \times 10^9$  TU/ml when assayed on 293T cells. The pSIH transfer vector used in these experiments generates lentiviral based vectors with an insert size of 4,267 bp.

**Primary cell culture and transduction.** Sertoli cells were isolated as previously described.<sup>13</sup> Briefly, Sertoli cells from 20 dpp Sprague Dawley rats were plated at  $6.13 \times 10^5$  cells/cm<sup>2</sup> at 37°C in 24-well culture plates (Nalge Nunc) or into Millicell PCF bicameral chambers (12mm diameter, 0.4  $\mu\text{m}$  pore size, 0.6 cm<sup>2</sup> surface area; Millipore) pre-coated with Matrigel (BD Biosciences), for measurement of transepithelial electrical resistance (TER). Cell cultures proceeded in DMEM/Hams F12 (Gibco) supplemented with non-essential amino acids (10 mM, Gibco), dialysed bovine serum albumin (1% w/v, Sigma), HEPES (10 mM, Gibco), insulin (5  $\mu\text{g/ml}$ , Novo-Nordisk), transferrin (5  $\mu\text{g/ml}$ , Sigma), sodium selenite (50 ng/ml, Sigma), penicillin (200 U/ml)-streptomycin (200  $\mu\text{g/ml}$ )-fungizone (0.5  $\mu\text{g/ml}$ ) (CSL), testosterone (28 ng/ml, Sigma) and FSH (150 mIU/ml, Puregon, N.V. Organon). After 72 h, Sertoli cell cultures were hypotonically shocked with 10% DMEM/F12 in distilled H<sub>2</sub>O for 45 sec to remove contaminating germ cells, and then replaced with culture medium. Sertoli cell cultures using this method are typically 92% pure after 5 d, with the contaminant cells predominantly being peritubular myoid cells, or residual germ cells.<sup>19</sup> In initial experiments,  $5 \times 10^6$  TU/ml lentiviral based vectors were applied directly to established Sertoli cell monolayers immediately following hypotonic shock (72 h after commencement of culture). The virus containing media was subsequently replaced 24 h later with fresh medium. Cells were then cultured for at least a further 48 h to allow for reporter gene expression. Transgene expression was confirmed by live imaging of GFP fluorescence. Cells were removed from cell culture for DNA extraction and

flow cytometry (see below) using a combination of trypsin, collagenase and hyaluronidase (2 mg/ml each, Sigma) for 20 min at 37°C, in an orbital shaker at 180 r.p.m. The enzymatic activity was then quenched using 10% FCS, and the cells washed and pelleted by centrifugation at 500 *g* at 4°C for 5 min.

Centrifugation and inoculation (“spinoculation”) of primary Sertoli cell cultures with *lentivirus*-based vectors was performed using an Eppendorf plate centrifuge at 1,000 *g* at room temperature for 30 min.

**Assessment of inter-Sertoli cell tight junction integrity.** Tight junction function was assessed by measurement of transepithelial electrical resistance (TER) of cells plated in bicameral chambers (EMD Millipore) on a daily basis at 37°C using a Millicell-electrical resistance system (EMD Millipore), as previously described.<sup>13</sup>

**Genomic DNA extraction, RNA extraction and qPCR.** DNA was extracted from Sertoli cell pellets using an isopropanol precipitation method.<sup>20,21</sup> RNA was extracted using Trizol following the manufacturer’s instructions (Life Technologies). Contaminating DNA was removed using a DNase free kit (Life Technologies), RNA concentration quantified using a nanodrop spectrophotometer (Thermo Scientific) and reverse transcribed using SuperScript-VILO (Life Technologies). Quantitative real-time PCR analysis was performed using the Roche Lightcycler 380 (Roche) with the FastStart DNA Master SYBR-Green 1 polymerase (Roche). Oligonucleotide primers specific to the GFP transfer vector (pSIH-cop-GFP, F: 5'-AAT GTC TTT GGA TTT GGG AAT CTT AT-3', R: 5'-TGG TCT AAC CAG AGA GAC CCA GTA-3') were used to determine viral DNA incorporation, or RNA expression, by comparing expression against reference DNA ( $\beta$ -actin, F: 5'-CCG TAA AGA CCT CTA TGC CAA CA-3', R: 5'-GAT TAC TGC CCT GGC TCC TAG C-3'), using previously published conditions.<sup>22</sup>

**Flow cytometry.** Following cell harvesting, cells were washed once in 0.5% BSA/PBS (%w/v), and fixed in 4% paraformaldehyde for 10 min at room temperature. Cells were subsequently washed and resuspended in 0.5% BSA/PBS. Cell population and reporter GFP expression were analyzed using a LSRII flow cytometer (BD Biosciences) and summary data of at least 30,000 cells per data point collated using FlowJo software (Tree Star Inc.).

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**Immunocytochemistry and microscopy.** Sertoli cell monolayers cultured on Millicell bicameral chambers (EMD Millipore) were washed once in PBS, and then fixed in 4% paraformaldehyde in PBS for 30 min at room temperature. Cells were then washed in PBS, and permeabilised with 0.05% (v/v) Triton X-100 in PBS for 5 min on ice. The chamber was then washed in PBS and stored at 4°C until use. Immunocytochemistry was performed by first blocking non-specific binding sites with CAS block (Zymed) containing 10% normal goat serum (Vector Laboratories) for 20 min. Primary antiserum (rat anti-Gata4 1:500, C#14-9980, eBioscience; rabbit anti-Cldn11 1:200 C#36-4500 and rabbit-Tjp1 1:200 #61-7300, Zymed Laboratories) was applied and incubated for 16 h at 4°C. Cell monolayers were then washed, and secondary antisera (goat anti-rat AlexaFluor-568, goat anti-rabbit AlexaFluor-647, Molecular Probes) applied. Confocal analysis was performed using an Olympus Fluoview FV300 confocal system attached to an Olympus IX 81 inverted microscope.

**Statistics.** All measurements were conducted with replicates from which the mean and SD were calculated. Statistics were performed using SigmaStat version 3.5 (Systat Software, Inc.). Before analysis, homogeneity of variance was confirmed, and assessed using a one-way ANOVA, followed by the Student-Newman-Keuls post hoc multiple group comparisons test for significance. A *p*-value of  $\leq 0.05$  was used as a measure of statistical significance for all experiments.

## Disclosure of Potential Conflicts of Interest

No potential conflicts of interest were disclosed.

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