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Construction of an immune-related IncRNA signature pair for predicting oncologic outcomes and the sensitivity of immunosuppressor in treatment of lung adenocarcinoma

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Abstract

Background: Although immunotherapy has shown clinical activity in lung adenocarcinoma (LUAD), LUAD prognosis has been a perplexing problem. We aimed to construct an immune-related lncRNA pairs (IRLPs) score for LUAD and identify what immunosuppressor are appropriate for which group of people with LUAD.

Methods: Based on The Cancer Genome Atlas (TCGA)-LUAD cohort, IRLPs were identified to construct an IRLPs scoring system by Cox regression and validated in the Gene Expression Omnibus (GEO) dataset using log-rank test and the receiver operating characteristic curve (ROC). Next, we used spearman's correlation analysis, *t*-test, signaling pathways analysis and gene mutation analysis to explore immune and molecular characteristics in different IRLP subgroups. The "pRRophetic" package was used to predict the sensitivity of immunosuppressant.

Results: The IRLPs score was constructed based on eight IRLPs calculated as $2.12 \times (MIR31HG|RRN3P2) + 0.43 \times (NKX2-1-AS1|AC083949.1) + 1.79 \times (TMPO-AS1|LPP-AS2) + 1.60 \times (TMPO-AS1|MGC32805) + 1.79 \times (TMPO-AS1|PINK1-AS) + 0.65 \times (SH3BP5-AS1|LINC01137) + 0.51 \times (LINC01004|SH3PXD2A-AS1) + 0.62 \times (LINC00339|AGAP2-AS1). Patients with a lower IRLPs risk score had a better overall survival (OS) (Log-rank test <math>P_{TCGA train dataset} < 0.001, P_{TCGA test dataset} = 0.017, P_{GEO dataset} = 0.027)$ and similar results were observed in the AUCs of TCGA dataset and GEO dataset (AUC $_{TCGA train dataset} = 0.777$, AUC $_{TCGA test dataset} = 0.685$, AUC $_{TCGA total dataset} = 0.733$, AUC $_{GEO dataset} = 0.680$). Immune score (Cor = -0.18893, P < 0.001), stoma score (Cor = -0.24804, P < 0.001), and microenvironment score (Cor = -0.22338, P < 0.001) were significantly decreased in the patients with the higher IRLP risk score. The gene set enrichment analysis found that high-risk group enriched in molecular changes in DNA and chromosomes signaling pathways, and in this group the tumor mutation burden (TMB) was higher than

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in the low-risk group (P = 0.0015). Immunosuppressor methotrexate sensitivity was higher in the high-risk group (P = 0.0052), whereas parthenolide (P < 0.001) and rapamycin (P = 0.013) sensitivity were lower in the high-risk group.

Conclusions: Our study established an IRLPs scoring system as a biomarker to help in the prognosis, the identification of molecular and immune characteristics, and the patient-tailored selection of the most suitable immunosuppressor for LUAD therapy.

Keywords: LUAD, IRLPs signature, Biomarker, Prognosis, Immunosuppressant

Background

Lung cancer is one of the most common malignant tumors globally, with a high incidence of 11.4% in 2020. Approximately 40% of the primary lung tumors are lung adenocarcinomas (LUAD) [1, 2]. LUAD, which is common in females and non-smokers, is characterized by high mortality and metastasis rates [3]. Although great improvement has been made in the clinical diagnosis and treatment, the 5-year survival rate in LUAD patients is only 18% [4]. Therefore, the identification of new biomarkers to help in the prognosis of LUAD is of great significance.

Chemotherapeutic is one of the most effective ways in the treatment of advanced non-small cell lung cancer. However, drug resistance is a is a major problem that puzzles researchers. Immunosuppressants recently were found to be potential anti-cancer drugs in LUAD. Previous studies have shown that rapamycin and parthenolide will result some beneficial modulation in lung cancer chemotherapy [5–7]. Recently, the emergence of immunotherapy has brought unprecedented levels of survival to lung cancer patients, especially those with advanced or metastatic LUAD [8, 9]. However, immunotherapy brings not only considerable therapeutic effects but also immune-related adverse events (ir AEs). Corticosteroid therapy can successfully treat most ir AEs, but a combination of immunosuppressors is needed to combat more serious adverse reactions [10, 11]. Immunosuppressant methotrexate were used to treat rheumatic ir AEs [11], and high-dose methotrexate can be applied to immunochemotherapy in many type of cancers [12, 13]. However, since it is unclear whether a patient can undergo immunosuppressive therapy of these immunosuppressors safely, there is an urgent need to find some biomarkers to predict the drug sensitivity of immunosuppressants.

LUAD is an immune-sensitive cancer, studies have shown that the immunotherapy response may be predicted by tumor-immune cell infiltration and an immune score[14]. Long non-coding RNAs (lncRNAs) are RNAs without protein-coding capacity and greater than 200 nucleotides in length[15]. Studies showed that lncRNAs can regulate the immune response and immune cell development[16, 17]. Several studies have

proposed immune-related lncRNA signatures to help in the prognostic of LUAD. However, the results cannot be directly generalized to all patients due to the use of different chip sequencing protocols, different platforms, and different testing times for gene expression [18–20]. These shortcomings could be overcome by combining two or more biomarkers, which work better than a single prediction criterion in cancer prediction models, and immune-related gene pairs (IRLPs) were reported to have accurately predicted the LUAD prognoses [21, 22]. However, these studies have focused on mRNAs rather than lncRNAs, which play an important role in the immune system. Therefore, the clinical relevance and prognostic significance of immune-related lncRNAs pairs (IRLPs) are currently unclear.

In this study, we constructed an individualized signature of IRLPs that works as an independent and predictive factor of overall survival (OS) for LUAD patients. Furthermore, the IRLP model also helps distinguishing the LUAD patients responsive to immunotherapy and predicts the sensitivity of immunosuppressors used in treatment of LUAD.

Materials and methods

Data source

The RNA-seq data of 515 LUAD cases (including 535 tumor samples and 59 normal samples) and 569 LUAD cases of genetic alteration data were downloaded from The Cancer Genome Atlas (TCGA)-LUAD cohort (https://portal.gdc.cancer.gov/) (October 10th, 2020). In addition, the normalized data of RNA expression matrix of GSE30219, GSE37745, and GSE50081 (we selected the RNA expression datasets which were normalized and measured from the same platform after searched LUAD RNA expression data) were downloaded from Gene Expression Omnibus (GEO https:// www.ncbi.nlm.nih.gov/geo/); the platform of these datasets was GPL570 (Affymetrix Human Genome U133 Plus 2.0 Array). The relevant clinical characteristics of patients were also downloaded, and the patients without the information on survival time and survival status were excluded from our study.

Identification of immune-related IncRNAs

Two thousand four hundred ninety-eight immune-related genes were downloaded from the ImmPort Portal (https://www.immport.org/). Then, immune-related lncRNAs were identified by Pearson's correlation analysis between immune-related genes and lncRNA expression levels ($|correlation\ coefficient| > 0.6$ and p < 0.001).

Construction of a prognostic IRLP signature

To ensure that the immune-related lncRNAs could be measured on all platforms included in this study, the intersect function was used to identify the common immune-related lncRNAs in the TCGA and GEO datasets. We only selected the lncRNAs with a relatively high variation in expression levels (median absolute deviation > 0.5). Next, the immune-related lncRNAs were paired randomly to construct a collection of lncRNA pairs. For each LUAD sample, the IRLPs were computed by pairwise comparison of the expression level. The output is one if the expression of the first lncRNA is higher than that of the second one; otherwise, the output is zero. We screened out overlapping IRLPs in TCGA and GEO dataset, after removing IRLPs with small variation and imbalanced distribution (MAD=0), the remaining ones were selected as candidate IRLPs. The TCGA dataset was randomly divide into train dataset and test dataset and we performed univariate Cox regression analysis and LASSO regression analysis with tenfold cross-validation to find out OS-related IRLPs in TCGA train dataset. Finally, multivariate Cox regression analysis was carried out to identify top OS-related IRLPs and to establish the final model of an IRLP risk score to predict the prognosis of LUAD. The IRLP risk score was calculated using the following formula:

$$IRLPriskscore = \sum_{i=1}^{n} coef_i \times x_i$$

where $coef_i$ is the coefficient and x_i is the output of pairwise comparison of expression level in each sample. (methods in selection of 8 IRLPs step by step were shown in the supplement).

Validation of IRLPs signature in the GEO data set

The IRLP model was further evaluated in the LUAD patients from the GEO dataset by the log-rank test. We also accessed the prognosis value of the IRLP risk score based on other clinical factors in univariate and multivariate Cox regression analysis. The receiver operating characteristic curve (ROC) was used to evaluate the

predicting accuracy of this signature by calculating the area under ROC (AUC).

Comprehensive analysis of immune characteristic and molecular variation in different IRLP risk score subgroups

Data on the infiltration of immune cells found in the TCGA dataset were downloaded from TIMER2.0 (http://timer.comp-genomics.org), a website that provides four modules for investigating the associations between immune infiltrates and genetic or clinical features, and four modules for exploring cancer-related associations in the TCGA cohorts [23]. Spearman's correlation analysis was performed to analyze the relationship between the immune cell infiltrates and the IRLPs risk score. In addition, Student's *t*-test was used to compare the different levels of immune cell infiltrates between the high-risk and low-risk groups defined by the IRLP risk score.

Differential expression analysis was performed on all genes between the high-risk group and low-risk group of TCGA samples. In addition, gene set enrichment analysis was used to determine the signaling pathways based on the Kyoto Encyclopedia of Genes and Genomes (KEGG) and Gene Ontology (GO) gene set (GSEA software).

In the gene mutation analysis, gene mutation quantity and quality were analyzed in two subgroups of LUAD patients (Maftools package). In addition, we also analyzed the relationship between tumor mutation burden (TMB) and IPLPs risk score subgroup using a Student's *t*-test.

Predicting the drug sensitivity of the immunosuppressors

pRRophetic is an R package used to predict clinical chemotherapeutic response from tumor gene expression level, a ridge regression model was applied. Genes (ruled out genes with very low variability across samples) as predictors and the drug sensitivity (IC50) values (of the drug of interest) as the outcome variable[24]. pRRophetic package included prediction of drug sensitivity in immunosuppressants (methotrexate, parthenolide, rapamycin). In order to identify which immunosuppressant might be useful, we used the "pRRophetic" R package to predict drug sensitivity from tumor gene expression levels.

Statistical analysis

All statistical analyses were performed in the R 4.0.5 software. Student's t-test was used to compare the differences between two subgroups. The Kaplan–Meier method was used to analyze the differences in survival curves using the log-rank test.

Results

Dataset of LUAD patients

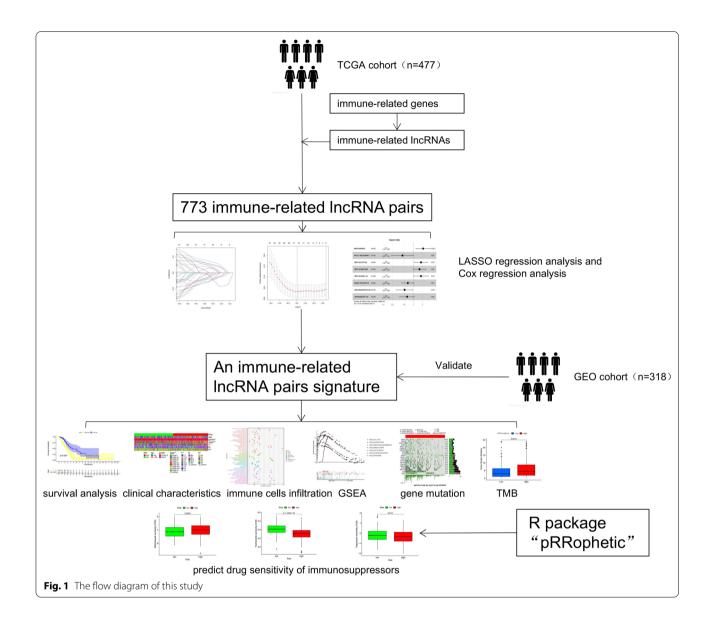
After excluding the patients for whom the survival status and survival time were missing, a total of 795 patients (TCGA-LUAD: 477 cases; GEO: 318 cases) were included in our study. All clinical characteristics (age, gender, stage, TNM grade) of TCGA and GEO dataset were present at Table 1. The flow diagram of this study was shown in Fig. 1.

Construction and validation of a prognostic IRLP signature A total of 105 immune-related lncRNAs were found in all platforms of the dataset, and 773 IRLPs were paired. First, univariate Cox regression identified 53 IRLPs that

were related to the OS of LUAD patients in the TCGA train dataset (P<0.01). Then, the Least Absolute Shrinkage and Selection Operator (LASSO) regression analysis with iteration=1000 selected 18 IRLPs (Fig. 2a) for the multivariate Cox regression analysis, and, finally, eight IRLPs were identified to calculate the IRLP risk score (Fig. 2b). The IRLPs risk score was calculated as 2.12 × (MIR31HG|RRN3P2) + 0.43 × (NKX2-1-AS1 | AC083949.1) + 1.79 × (TMPO-AS1 | LPP-AS2) + 1.60 × (TMPO-AS1 | MGC32805) + 1.79 × (TMPO-AS1 | PINK1-AS) + 0.65 × (SH3BP5-AS1|LINC01137) + 0.51 × (LINC01004|SH3PXD2A-AS1) + 0.62 × (LINC00339|AGAP2-AS1). Furthermore,

Table 1 Clinical characteristics of TCGA and GEO dataset

Variable	TCGA-LUAD dataset (N = 477) N(%)	GEO LUAD dataset (N = 318)		
		GSE30219 N(%)	GSE37745 N(%)	GSE50081 N(%)
Age				
< 68	261 (54.7)	61 (73.5)	71 (67.0)	48 (37.2)
≥68	216 (45.3)	22 (26.5)	35 (33.0)	81 (62.8)
Gender				
Female	257 (53.9)	18 (21.7)	60 (56.6)	62 (48.1)
Male	220 (46.1)	65 (78.3)	46 (43.4)	67 (51.9)
Stage				
I	4 (0.8)	NA	NA	NA
IA	124 (26.0)	NA	NA	37 (28.7)
IB	125 (26.2)	NA	NA	56 (43.4)
II	191 (40.0)	NA	NA	NA
IIA				7 (5.4)
IIB				29 (22.5)
III	NA	NA	NA	0 (0)
IV	25 (5.2)	NA	NA	0 (0)
Unknow	8 (1.7)	NA	NA	0 (0)
Т				
T ₁	159 (33.3)	69 (83.1)	NA	44 (34.1)
T_2	254 (53.2)	12 (14.5)	NA	83 (64.3)
T ₃	43 (9.0)	2 (2.4)	NA	2 (1.6)
T_4	18 (3.8)	0 (0)	NA	0 (0)
T_X	3 (0.6)	0 (0)	NA	0 (0)
N				
N_0	307 (64.4)	80 (96.4)	NA	95 (73.6)
N_1	90 (18.9)	3 (3.6)	NA	34 (26.4)
N_2	67 (14.0)	0 (0)	NA	0 (0)
N_X	2 (0.4)	0 (0)	NA	0 (0)
Unknow	1 (0.2)	0 (0)	NA	0 (0)
М				
M_0	313 (65.6)	83 (100)	NA	129 (100)
M_1	24 (5.0)	0 (0)	NA	0 (0)
M_X	136 (28.5)	0 (0)	NA	0 (0)



we compared the survival curves of the TCGA train dataset (P<0.001), TCGA test dataset (P=0.017), and GEO dataset (P=0.027) (Fig. 3a–c). These results all showed that high-risk LUAD patients exhibited a poorer prognosis than low-risk LUAD patients.

Assessing the value of the IRLP signature to predict the prognostic of overall survival

We took advantage of the univariate and multivariate Cox regression model to analyze the predictive value of IRLP risk score and clinical parameters. The univariable Cox regression analysis indicated that the IRLPs risk score was an important factor for patients' prognosis (TCGA total dataset: HR = 1.077 (95% CI: 1.047 - 1.107), P < 0.001; TCGA train dataset: HR = 1.428 (95% CI: 1.312 - 1.555),

 $P{<}0.001;\ TCGA$ test dataset: $HR{=}1.054$ (95% $CI{:}$ $1.014{-}1.097),\ P{=}0.009;\ GEO$ dataset: $HR{=}1.095$ (95% $CI{:}$ $1.053{-}1.140),\ P{<}0.001;\ Fig. 4a{-}d).$ The multivariable Cox regression indicated that the IRLPs risk score was an independent predictive indicator for the OS of LUAD patients (TCGA total dataset: $HR{=}1.078$ (95% $CI{=}1.045{-}1.112),\ P{<}0.001;\ TCGA$ train dataset: $HR{=}1.380$ (95% $CI{=}1.265{-}1.507),\ P{<}0.001;\ TCGA$ test dataset: $HR{=}1.050$ (95% $CI{=}1.001{-}1.100),\ P{=}0.043;$ GEO dataset: $HR{=}1.070$ (95% $CI{=}1.023{-}1.119),\ P{=}0.003;\ Fig. 4a{-}d).$ The ROC curves also revealed that the IRLPs risk score plays an important role in predicting LUAD prognosis (TCGA total dataset: AUC = 0.733; TCGA train dataset: AUC = 0.777; TCGA test dataset: AUC = 0.685; GEO dataset: AUC = 0.680 Fig. 5a{-}d).

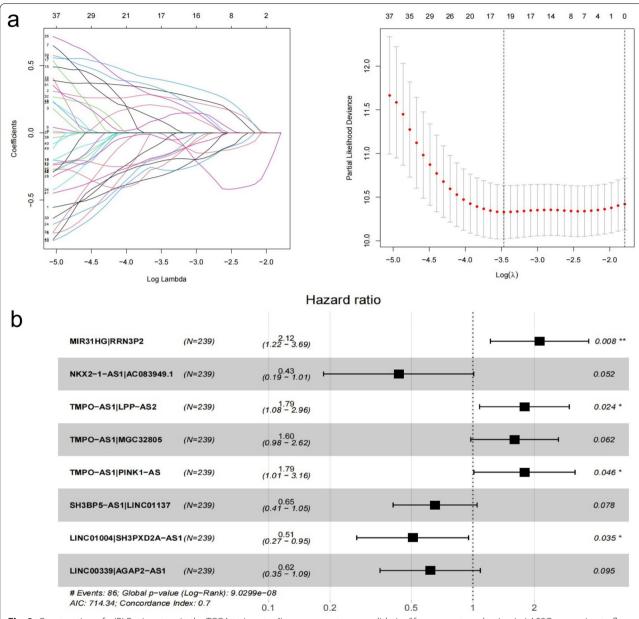


Fig. 2 Construction of a IRLPs signature in the TCGA train set. a "Leaveone-out-cross-validation" for parameter selection in LASSO regression to flter out 18 IRLPs. b The forest map of multivariate Cox regression analysis to establish a IRLPs signature with 8 IRLPs in TCGA train dataset

Relationship between the IRLP subgroups and clinical characteristics

We evaluated the correlation between the IRLP subgroups and clinical characteristics by heatmap. The results showed that the distributions of gender, stage, T stage, and N stage were significantly different between the high-risk and low-risk groups of the TCGA total dataset (Fig. 6a), and the GEO dataset showed that the distributions of the T and M stage were significantly different (Fig. 6b).

Relationship between the IRLPs and immune cell infiltrates

Immune score reflect immune cells infiltrate situation, and stoma score reflect the stromal cells infiltrate. While microenvironment score reflect a a comprehensive situation of the former two scores, represents the overall picture of the immune microenvironment. Spearman's rank correlation analysis showed that the immune score (Cor = -0.18893, P < 0.001), stoma score (Cor = -0.24804, P < 0.001), and microenvironment score (Cor = -0.22338, P < 0.001) were significantly

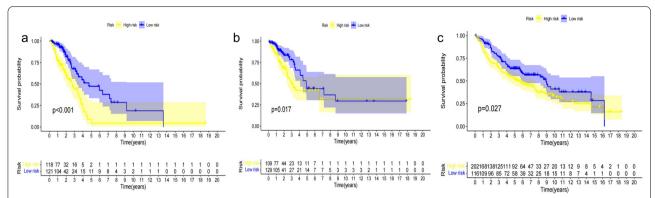


Fig. 3 Kaplan–Meier survival curves of LUAD in IRLPs high-risk and low-risk group a) in the TCGA train dataset, b in the TCGA test dataset. c in the GEO dataset

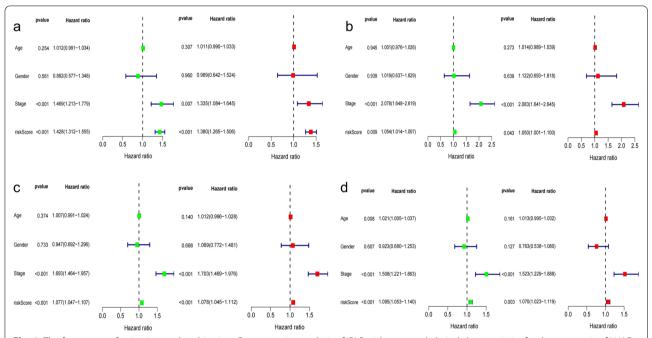


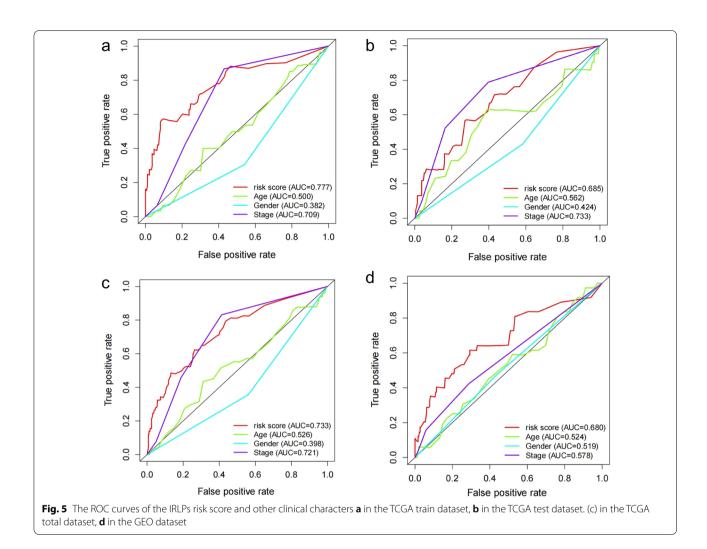
Fig. 4 The forest map of univariate and multivariate Cox regression analysis of IRLPs risk score and clinical characteristics for the prognosis of LUAD patients. **a** in the TCGA train dataset, **b** in the TCGA test dataset. **c** in the TCGA total dataset, **d** in the GEO dataset

decreased in the group with the higher IRLP risk score (Fig. 7). The distributions of immune cell infiltrates were different in the two IRLP subgroups (Fig. 8).

Molecular characteristics of different IRLP subgroups

The gene sets of the high-IRLP subgroup were most enriched in KEGG_DNA_REPLICATION (enrich score = 0.76), GOBP_ATTACHMENT_OF_MITOTIC_SPINDLE_MICROTUBULES_TO_KINETOCHORE (enrich score = 0.92), GOMF_SINGLE_STRANDED_DNA_HELICASE_ACTIVITY (enrich score = 0.86), and GOCC_CONDENSED_NUCLEAR_CHROMO-

SOME_KINETOCHORE (enrich score = 0.83) (Fig. 9a). The gene sets of low-IRLPs subtype were most enriched in KEGG_ASTHMA (enrich score = 0.70), GOMF_ATP_DEPENDENT_MICROTUBULE_MOTOR_ACTIV-ITY_MINUS_END_DIRECTED (enrich score = 0.81), GOCC_AXONEMAL_DYNEIN_COMPLEX (enrich score = 0.79), and GOBP_SODIUM_ION_EXPORT_ACROSS_PLASMA_MEMBRANE (enrich score = 0.76) (Fig. 9b). GO analysis showed that the differentially expressed genes between the IRLP subgroups were enriched in neutrophil activation involved in immune response (BP), the cell-cell junction (CC), and metal



ion transmembrane transporter activity (MF; Fig. 9c-e). KEGG analysis showed that the differentially expressed genes were enriched in Herpes simplex virus 1 infection (Fig. 9f). Then, we analyzed gene mutations to gain further biological insight into the immunological nature of the IRLPs subgroups. The high-risk group had the highest mutation rate (the top 20 genes), and missense variations were the most common mutation type in the two subgroups (Fig. 10a, b). The TTN mutation was the highest in the high-risk group. Next, we explored the relationship between the TTN mutation and IRLP subgroups. The TTN mutation was significantly more frequent in the high-risk group (Fig. 10c). Finally, we compared the tumor mutation burden (TMB) between these two subgroups. As a result, TMB was higher in the IRLPs of the high-risk group (Fig. 10d).

Prediction of drug sensitivity of immunosuppressors

We identified three immunosuppressors (methotrexate, parthenolide, and rapamycin) in the "pRRophetic"

R package. Methotrexate had higher sensitivity in the IRLP high-risk group (P=0.0052, Fig. 11a), whereas parthenolide (P<0.001) and rapamycin (P=0.013) showed lower sensitivity in IRLP high-risk group (Fig. 11b, c).

Discussion

With the development of sequencing technology, more and more attention in cancer research has been paid to bioinformatics methods. In our present study, we constructed a risk scoring system based on eight IRLPs in the TCGA dataset, and the patients were divided into high-risk and low-risk groups according to the cut-off of risk score. Survival analysis showed that the high-risk group had a poor prognosis. The IRLP-risk score was an independent risk factor in our Cox regression analysis combined with clinical characteristics (age, gender, and stage). These results were also proven in the GEO dataset. Furthermore, our results also showed that the IRLP risk score was related to immune cell infiltration. Next, we explored the gene functional enrichment and gene

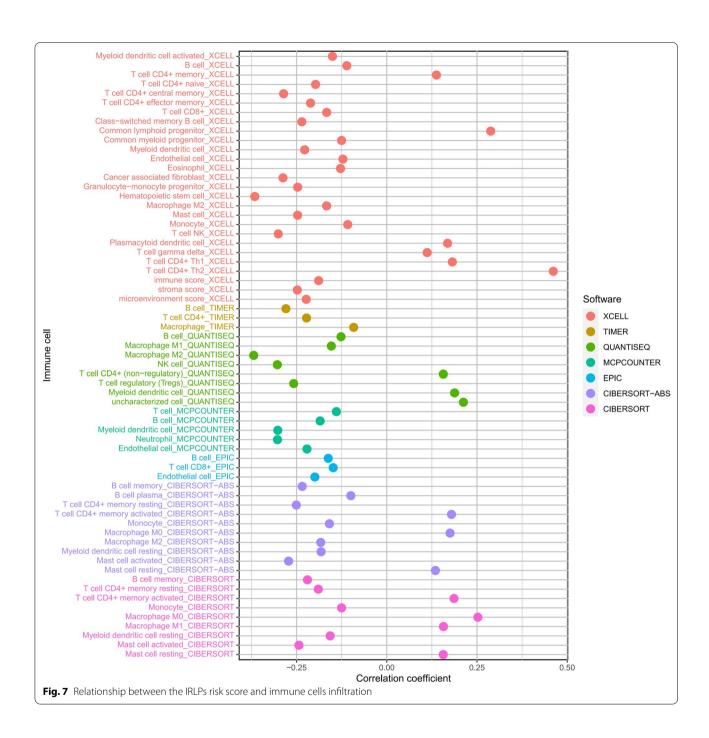


Fig. 6 Heat map for relationship between IRLPs subgroups and clinical characters * means P < 0.05; ** means P < 0.01; *** means P < 0.001 **a** in the TCGA total dataset. **b** in the GEO dataset

mutation in two IRLPs subgroups. The high-risk group was found to be enriched in molecular changes in DNA and chromosomes, and to have a higher TMB than the low-risk group. Finally, the drug sensitivity of immunosuppressors was predicted to find the most suitable drugs for each group.

There are several studies developed prognostic signatures of LUAD based on RNA-seq or microarray expression in recent years [25, 26]. We built a prognostic IRLPs signature based on 8 IRLPs in our study, our results showed that patients with higher IRLPs risk

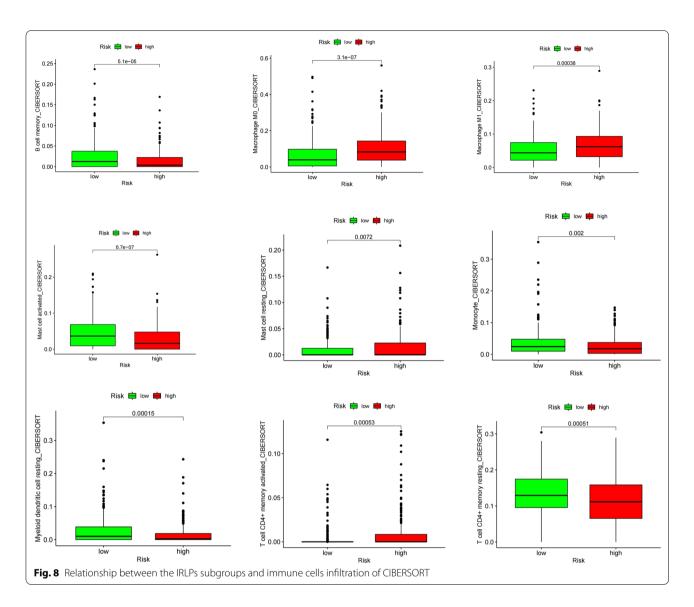
score related to a worse prognosis and also could predict overall survival time (AUC=0.777). What's more, both internal and external queues were used to validate the IRLPs risk score system we built. All of the valid dataset indicated that our IRLPs signature was reliable (AUC $_{\rm TCGA\ test\ dataset}$ =0.685, AUC $_{\rm GEO\ dataset}$ =0.680). A previous study have built an immune-related prognostic signature directly with the expression level of genes based on RNA-seq for LUAD in TCGA dataset, the AUC for overall survival time is 0.662 [27], inferior to the AUCs in our study. The IRLPs risk score we



constructed had a important prognostic significance, it may become a novel biomarker of LUAD.

The tumor microenvironment (TME) is correlated with cancer prognosis, supports cancer cells to replicative proliferation, and affects the malignant phenotypes [28, 29]. Many immune cells are present in the TME, modulating tumor cell migration, invasion, metastasis, and anticancer drug sensitivity [30]. The relationship between the IRLP score and infiltrating immune cells was analyzed in

our study, and we found that they were significantly correlated. These results indicated that our IRLP risk score might allow the prognosis of LUAD by being sensitive to the functional status of immune cells. The immune score reflected the infiltration of immune cells in the tumor tissue based on the algorithm. A study found that patients with medium and high immune scores had a longer OS time than those in the low immune score group in lung cancer[31]. This means that a higher immune score may



be beneficial for survival in lung cancer patients. The IRLP risk score was found to be negatively correlated with the immune score in our current results. These results demonstrated that a high immune activity might play an important role in the increased survival time of LUAD patients.

To gain further biological insight into the IRLP subgroups, we studied the functional enrichment and gene mutations in these two subgroups. Functional enrichment analysis found that molecular changes in DNA and chromosomes were most enriched in the high-risk subgroup. As previously reported, our results also showed that missense mutations are the most common type of mutations in LUAD[32]. The TTN mutation was found to be more frequent in the high-risk group than in the low-risk group and showed a significant difference between the high-risk and low-risk groups. The TTN mutation

was reported as a potential biomarker associated with a better response to immune checkpoint blockade in solid tumors [33]. A study based on the TCGA dataset reported that the TTN missense mutation correlates with favorable prognosis in lung squamous cell carcinoma (LUSC) but not in LUAD [34]. Our results are also in agreement with the notion that TTN mutation plays a different role in LUAD.

Next, the relationship between the IRLP score and TMB was explored. Not only a high TMB was found to reflect worse clinical outcomes in non-small cell lung cancer [35], but also patients with high TMB (TMB-H) achieved good results in immunotherapy of solid tumors [36]. In this study, the high-risk subgroup had the higher TMB. Thus, the TMB may explain why IRLPs are correlated with the prognosis of LUAD, and the IRLP score may also help explain the immunotherapy response.

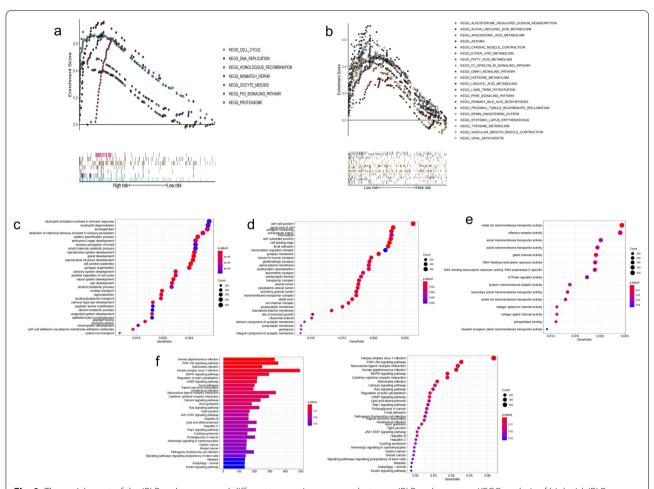


Fig. 9 The enrichment of the IRLPs subgroups and different expression gene set between IRLPs subgroups **a** KEGG analysis of high-risk IRLPs subgroup, **b** KEGG analysis of low-risk IRLPs subgroup, **c** GO-BP analysis of different expression gene set between IRLPs subgroups, **d** GO-CC analysis of different expression gene set between IRLPs subgroups, **f** KEGG analysis of different expression gene set between IRLPs subgroups, **f** KEGG analysis of different expression gene set between IRLPs subgroups

However, other possible mechanisms involved in this relationship still need to be further studied.

With immune checkpoint inhibitors effectively improve the OS time in various cancers, immunotherapy-mediated ir AEs were frequently reported because of their specificity and severity [37, 38]. In the clinic, immunosuppressors can be used to treat severe ir AEs [39, 40] and play a relevant role as anticancer agents in recent decades [41, 42]. Since the immune systems of different individuals are not equally sensitive to drugs, immunosuppressors should further selected for LUAD therapy. Hence, we explored the drug sensitivity of three immunosuppressors: methotrexate, parthenolide, and rapamycin. Methotrexate usually used for autoimmune disease therapy, studies have reported that methotrexate has a good curative effect in rheumatic ir AEs [43-47], and fatal myositis can be successful cured by high-dose corticosteroids and methotrexate [48]. Our findings showed

that irAEs in high-risk groups may be more sensitive to methotrexate treatment. Parthenolide is one of the biologicals that play an anti-inflammatory role by inhibiting nuclear factor kappa B (NF-κB) and cytokine tumor necrosis factor (TNF)- α [49, 50]. Parthenolide can induce lung cancer cells apoptosis and inhibit human lung cancer cell growth [51-53], demonstrated anticancer activity in the treatment of lung cancer. Rapamycin, an mTOR inhibitor, has broad anti-proliferative activity across NSCLC cells [54]. What's more, studies have reported that combined application of rapamycin and other chemotherapy drugs would enhance the efficacy [55, 56]. In our exploration, the drug sensitivity of rapamycin and parthenolide were higher in low-risk group. These immunosuppressors (methotrexate, parthenolide, and rapamycin) have a different mechanism of action, and patients also had different drug sensitivities. Thus, our IRLPs scores may help identify the patients who would benefit

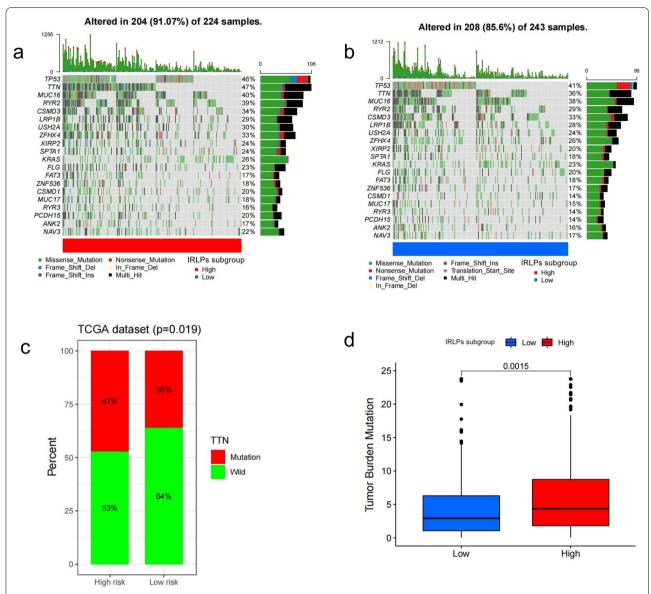
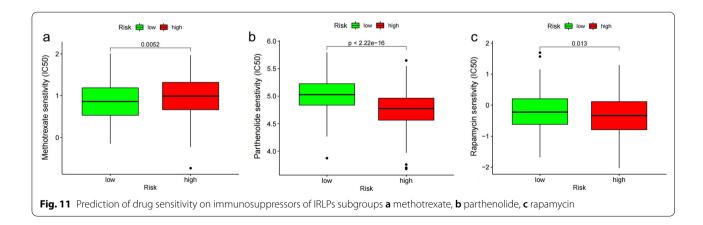


Fig. 10 Gene mutations of different IRLPs subgroups **a** Top 20 genes mutation of high-risk IRLPs subgroup, **b** Top 20 genes mutation of low-risk IRLPs subgroup, **c** relationship between TTN mutation and IRLPs subgroups



from immunosuppressors therapy, but the mechanisms of drug action in these two subgroups still need to be clarified.

Although we have constructed an IRLP risk scoring system that showed a good predictive performance for LUAD patients and overcame the inconsistent sequencing platforms, there were still some noteworthy limitations. First, the patients included in the training set were downloaded from TCGA, which mainly includes white race patients. Thus, other ethnic groups still need to be evaluated. However, the results showed that the IRLP score constructed in TCGA also applies to the Asian GEO dataset. Second, we intersected the lncRNAs from two public datasets to overcome the differences in the sequencing platforms, and some important lncRNAs may have been ignored or contributed to selection bias. Third, our prediction on drug sensitivities was not validated in a cohort. There is no complete cohort data at present because of the difficulty in developing clinical immunosuppressive therapy experiment. Finally, we used the "pRRophetic" R package to explore the drug sensitivity of immunosuppressors, which includes a limited set of drugs and did not allow us to address the sensitivity of many commonly used drugs.

Conclusion

In summary, we built a risk model based on IRLPs. This signature had a good predictive accuracy and effectiveness for LUAD. Furthermore, our IRLPs score significantly correlated with TME and TMB, indicating that these molecular changes might explain the different clinical outcomes. Importantly, our IRLPs may enhance the identification of the patients who can benefit from immunosuppressive therapy.

Abbreviations

LUAD: Lung adenocarcinoma; LUSC: Lung squamous cell carcinoma; IRLP: Immune-related IncRNA pair; ir AEs: Immune-related adverse events; TCGA: The cancer genome atlas; GEO: Gene expression omnibus database; OS: Overall survival; TMB: Tumor mutation burden; IncRNAs: Long non-coding RNAs; KEGG: Kyoto encyclopedia of genes and genomes; GO: Gene ontology; GSEA: Gene set enrichment analysis; TME: Tumor microenvironment.

Supplementary Information

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Additional file 1. Methods in selection of 8 IRLPs.

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Contribution to the field statement

Immunotherapy has shown clinical activity in lung adenocarcinoma (LUAD), but immunotherapy-mediated ir AEs such as pneumonitis and thyroid

dysfunction were frequently reported because of their specificity and severity. Based on The Cancer Genome Atlas (TCGA) LUAD dataset, IRLPs were identified to construct an IRLPs scoring system by Cox regression and were validated in the Gene Expression Omnibus (GEO) dataset. Next, immune and molecular characteristics were explored in different IRLP subgroups. The "pRRophetic" package was used to predict the sensitivity of drugs used to treat ir AEs. Our study established an IRLPs scoring system as a biomarker to help in the prognosis, the identification of molecular and immune characteristics, and the patient-tailored selection of the most suitable drugs for ir AEs therapy.

Author contributions

FH designed the study. JZ, ZC, ML and XX analyzed the data. ZH and FH contributed to interpretation of the results. SY and YL searched the dataset. JZ, ZC and JC drafted the manuscript. FH contributed to critical revision of the manuscript for important intellectual content. FH and ZH approved the final version of the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

The original contributions presented in the study are included in the article/ Supplementary Material. Further inquiries can be directed to the corresponding authors.

Declarations

Ethics approval and consent to participate

Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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References

- Freddie SHFJSRLLMSIJAB. Global cancer statistics 2020: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2021;71(3):209–49.
- Siegel R, Miller K, Jemal A. Cancer statistics, 2019. CA Cancer J Clin. 2019;69(1):7–34.
- Luo W, Tian P, Wang Y, Xu H, Chen L, Tang C, et al. Characteristics of genomic alterations of lung adenocarcinoma in young never-smokers. Int J Cancer. 2018:143(7):1696–705.
- 4. Lin J, Cardarella S, Lydon C, Dahlberg S, Jackman D, Jänne P, et al. Fiveyear survival in EGFR-mutant metastatic lung adenocarcinoma treated with EGFR-TKIs. J Thoracic Oncol. 2016;11(4):556–65.
- Talib W, Al Kury L. Parthenolide inhibits tumor-promoting effects of nicotine in lung cancer by inducing P53—dependent apoptosis and inhibiting VEGF expression. Biomed Pharmacother. 2018;107:1488–95.

- Lin M, Bi H, Yan Y, Huang W, Zhang G, Zhang G, et al. Parthenolide suppresses non-small cell lung cancer GLC-82 cells growth via B-Raf/ MAPK/Erk pathway. Oncotarget. 2017;8(14):23436–47.
- Yang Z, Lei Z, Li B, Zhou Y, Zhang G, Feng Z, et al. Rapamycin inhibits lung metastasis of B16 melanoma cells through down-regulating alphav integrin expression and up-regulating apoptosis signaling. Cancer Sci. 2010;101(2):494–500.
- 8. Mostafa A, Morris D. Immunotherapy for lung cancer: has it finally arrived? Front Oncol. 2014;4:288.
- Garon E, Hellmann M, Rizvi N, Carcereny E, Leighl N, Ahn M, et al. Five-Year overall survival for patients with advanced non-small-cell lung cancer treated with pembrolizumab: results from the phase I KEYNOTE-001 study. J Clin Oncol. 2019;37(28):2518–27.
- Verspohl S, Holderried T, Behning C, Brossart P, Schäfer V. Prevalence, therapy and tumour response in patients with rheumatic immunerelated adverse events following immune checkpoint inhibitor therapy: a single-centre analysis. Ther Adv Musculoskelet Dis. 2021. https://doi.org/10.1177/1759720X211006963.
- Zhong H, Zhou J, Xu D, Zeng X. Rheumatic immune-related adverse events induced by immune checkpoint inhibitors. Asia Pac J Clin Oncol. 2020. https://doi.org/10.1111/ajco.13346.
- Miyakita Y, Ohno M, Takahashi M, Muragaki Y, Katai H, Narita Y. Immunochemotherapy using rituximab (RTX) and high-dose methotrexate (HD-MTX): an evaluation of the addition of RTX to HD-MTX in recurrent primary central nervous system lymphoma (PCNSL). Jpn J Clin Oncol. 2017;47(10):919–24.
- Stang A, Schwärzler P, Schmidtke S, Tolosa E, Kobbe R. Successful immunochemotherapy for burkitt lymphoma during pregnancy as a bridge to postpartum high-dose methotrexate therapy: a case report and review of the literature. Clin Lymphoma Myeloma Leuk. 2020;20(6):e284–90.
- Alexandrov L, Kim J, Haradhvala N, Huang M, Tian Ng A, Wu Y, et al. The repertoire of mutational signatures in human cancer. Nature. 2020;578(7793):94–101.
- Yao R, Wang Y, Chen L. Cellular functions of long noncoding RNAs. Nat Cell Biol. 2019;21(5):542–51.
- 16. Chen Y, Satpathy A, Chang H. Gene regulation in the immune system by long noncoding RNAs. Nat Immunol. 2017;18(9):962–72.
- 17. Atianand M, Caffrey D, Fitzgerald K. Immunobiology of long noncoding RNAs. Annu Rev Immunol. 2017;35:177–98.
- Li J, Zhang C, Zhang C, Wang H. Construction of immune-related and prognostic IncRNA clusters and identification of their immune and genomic alterations characteristics in lung adenocarcinoma samples. Aging. 2020;12(10):9868–81.
- Jin D, Song Y, Chen Y, Zhang P. Identification of a seven-IncRNA immune risk signature and construction of a predictive nomogram for lung adenocarcinoma. Biomed Res Int. 2020;2020:7929132.
- Leek J, Scharpf R, Bravo H, Simcha D, Langmead B, Johnson W, et al. Tackling the widespread and critical impact of batch effects in highthroughput data. Nat Rev Genet. 2010;11(10):733–9.
- Wu C, Hu Q, Ma D. Development of an immune-related gene pairs signature for predicting clinical outcome in lung adenocarcinoma. Sci Rep. 2021;11(1):3611.
- Xu F, Zhan X, Zheng X, Xu H, Li Y, Huang X, et al. A signature of immune-related gene pairs predicts oncologic outcomes and response to immunotherapy in lung adenocarcinoma. Genomics. 2020;112(6):4675–83.
- 23. Li T, Fu J, Zeng Z, Cohen D, Li J, Chen Q, et al. TIMER20 for analysis of tumor-infiltrating immune cells. Nucleic Acids Res. 2020;48:W509–14.
- Paul G, Nancy C, Huang RS, Barbour JD. pRRophetic: an R package for prediction of clinical chemotherapeutic response from tumor gene expression levels. PLoS ONE. 2014;9(9):e107468.
- Guo D, Wang M, Shen Z, Zhu J. A new immune signature for survival prediction and immune checkpoint molecules in lung adenocarcinoma. J Transl Med. 2020;18(1):123.
- Lei Z, Zhe Z, Zhenglun Y. Identification of a novel glycolysis-related gene signature for predicting metastasis and survival in patients with lung adenocarcinoma. J Transl Med. 2020;17(1):423.
- 27. Li N, Wang J, Zhan X. Identification of immune-related gene signatures in lung adenocarcinoma and lung squamous cell carcinoma. Front Immunol. 2021;12: 752643.

- Thakkar S, Sharma D, Kalia K, Tekade R. Tumor microenvironment targeted nanotherapeutics for cancer therapy and diagnosis: a review. Acta Biomater. 2020:101:43–68.
- Mony J, Schuchert M. Prognostic implications of heterogeneity in intratumoral immune composition for recurrence in early stage lung cancer. Front Immunol. 2018;9:2298.
- Li L, Yu R, Cai T, Chen Z, Lan M, Zou T, et al. Effects of immune cells and cytokines on inflammation and immunosuppression in the tumor microenvironment. Int Immunopharmacol. 2020;88: 106939.
- Han K, Qian K, Zhao T, Liu X, Zhang Y. Prediction of prognosis of patients with lung cancer in combination with the immune score. 2021. Biosci Rep. https://doi.org/10.1042/BSR20203431.
- 32. Chen J, Yang H, Teo A, Amer L, Sherbaf F, Tan C, et al. Genomic landscape of lung adenocarcinoma in East Asians. Nat Genet. 2020;52(2):177–86.
- Yang Y, Zhang J, Chen Y, Xu R, Zhao Q, Guo W. MUC4, MUC16, and TTN genes mutation correlated with prognosis, and predicted tumor mutation burden and immunotherapy efficacy in gastric cancer and pancancer. Clin Transl Med. 2020;10(4): e155.
- Cheng X, Yin H, Fu J, Chen C, An J, Guan J, et al. Aggregate analysis based on TCGA: TTN missense mutation correlates with favorable prognosis in lung squamous cell carcinoma. J Cancer Res Clin Oncol. 2019;145(4):1027–35.
- Chae Y, Davis A, Agte S, Pan A, Simon N, lams W, et al. Clinical implications
 of circulating tumor DNA tumor mutational burden (ctDNA TMB) in nonsmall cell lung cancer. Oncologist. 2019;24(6):820–8.
- Halbert B, Einstein D. Hot or not: tumor mutational burden (TMB) as a biomarker of immunotherapy response in genitourinary cancers. Urology. 2021:147:119–26
- Osorio J, Ni A, Chaft J, Pollina R, Kasler M, Stephens D, et al. Antibodymediated thyroid dysfunction during T-cell checkpoint blockade in patients with non-small-cell lung cancer. Ann Oncol. 2017;28(3):583–9.
- Nishino M, Chambers E, Chong C, Ramaiya N, Gray S, Marcoux J, et al. Anti-PD-1 inhibitor-related pneumonitis in non-small cell lung cancer. Cancer Immunol Res. 2016;4(4):289–93.
- 39. Weber J, Dummer R, de Pril V, Lebbé C, Hodi F. Patterns of onset and resolution of immune-related adverse events of special interest with ipilimumab: detailed safety analysis from a phase 3 trial in patients with advanced melanoma. Cancer. 2013;119(9):1675–82.
- Melissaropoulos K, Klavdianou K, Filippopoulou A, Kalofonou F, Kalofonos H, Daoussis D. Rheumatic manifestations in patients treated with immune checkpoint inhibitors. Int J Mol Sci. 2020. https://doi.org/10.3390/ijms2 1003380
- 41. Romano S, Di Pace A, Sorrentino A, Bisogni R, Sivero L, Romano M. FK506 binding proteins as targets in anticancer therapy. Anticancer Agents Med Chem. 2010;10(9):651–6.
- Fan Y, Hou H, Tay H, Guo W, Berggren P, Loo S. Preservation of anticancer and immunosuppressive properties of rapamycin achieved through controlled releasing particles. AAPS PharmSciTech. 2017;18(7):2648–57.
- Kostine M, Rouxel L, Barnetche T, Veillon R, Martin F, Dutriaux C, et al. Rheumatic disorders associated with immune checkpoint inhibitors in patients with cancer-clinical aspects and relationship with tumour response: a single-centre prospective cohort study. Ann Rheum Dis. 2018;77(3):393–8.
- Kuswanto W, MacFarlane L, Gedmintas L, Mulloy A, Choueiri T, Bermas B. Rheumatologic symptoms in oncologic patients on PD-1 inhibitors. Semin Arthritis Rheum. 2018;47(6):907–10.
- 45. Cappelli L, Gutierrez A, Baer A, Albayda J, Manno R, Haque U, et al. Inflammatory arthritis and sicca syndrome induced by nivolumab and ipilimumab. Ann Rheum Dis. 2017;76(1):43–50.
- Cappelli L, Brahmer J, Forde P, Le D, Lipson E, Naidoo J, et al. Clinical presentation of immune checkpoint inhibitor-induced inflammatory arthritis differs by immunotherapy regimen. Semin Arthritis Rheum. 2018;48(3):553–7.
- Ornstein M, Calabrese C, Wood L, Kirchner E, Profusek P, Allman K, et al. Myalgia and arthralgia immune-related adverse events (irAEs) in patients with genitourinary malignancies treated with immune checkpoint inhibitors. Clin Genitourin Cancer. 2019;17(3):177–82.
- Liu Y, Liu Z, Zeng X, Bai C, Chen L, Lin S, et al. Fatal myositis and spontaneous haematoma induced by combined immune checkpoint inhibitor treatment in a patient with pancreatic adenocarcinoma. BMC Cancer. 2019;19(1):1193.

- Park J, Kang Y, Hwang S, Oh S, Byun J. Parthenolide has negative effects on in vitro enhanced osteogenic phenotypes by inflammatory cytokine TNF-α via inhibiting JNK signaling. Int J Mol Sci. 2020;21(15):5433.
- Karam L, Abou Staiteieh S, Chaaban R, Hayar B, Ismail B, Neipel F, et al. Anticancer activities of parthenolide in primary effusion lymphoma preclinical models. Mol Carcinog. 2021. https://doi.org/10.1002/mc.23324.
- Zhao X, Liu X, Su L. Parthenolide induces apoptosis via TNFRSF10B and PMAIP1 pathways in human lung cancer cells. J Exp Clin Cancer Res. 2014:33:3
- 52. Sun L, Yuan W, Wen G, Yu B, Xu F, Gan X, et al. Parthenolide inhibits human lung cancer cell growth by modulating the IGF-1R/PI3K/Akt signaling pathway. Oncol Rep. 2020;44(3):1184–93.
- 53. Li X, Huang R, Li M, Zhu Z, Chen Z, Cui L, et al. Parthenolide inhibits the growth of non-small cell lung cancer by targeting epidermal growth factor receptor. Cancer Cell Int. 2020;20(1):561.
- 54. Bi Y, Jiang Y, Li X, Hou G, Li K. Rapamycin inhibits lung squamous cell carcinoma growth by downregulating glypican-3/Wnt/β-catenin signaling and autophagy. J Cancer Res Clin Oncol. 2021;147(2):499–505.
- 55. Kawabata S, Chiang C, Tsurutani J, Shiga H, Arwood M, Komiya T, et al. Rapamycin downregulates thymidylate synthase and potentiates the activity of pemetrexed in non-small cell lung cancer. Oncotarget. 2014;5(4):1062–70.
- Niu H, Wang J, Li H, He P. Rapamycin potentiates cytotoxicity by docetaxel possibly through downregulation of Survivin in lung cancer cells. J Exp Clin Cancer Res. 2011;30:28.

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