



Contents lists available at ScienceDirect

# International Journal for Parasitology: Drugs and Drug Resistance

journal homepage: [www.elsevier.com/locate/ijpddr](http://www.elsevier.com/locate/ijpddr)

## 3-nitroimidazo[1,2-*b*]pyridazine as a novel scaffold for antiparasitics with sub-nanomolar anti-*Giardia lamblia* activity

Yang Zheng<sup>a,1</sup>, Joachim Müller<sup>b,1</sup>, Stefan Kunz<sup>a,b</sup>, Marco Siderius<sup>a</sup>, Louis Maes<sup>c</sup>, Guy Caljon<sup>c</sup>, Norbert Müller<sup>b</sup>, Andrew Hemphill<sup>b</sup>, Geert Jan Sterk<sup>a</sup>, Rob Leurs<sup>a,\*</sup>

<sup>a</sup> Amsterdam Institute for Molecules, Medicines and Systems, Division of Medicinal Chemistry, Faculty of Science, Vrije Universiteit Amsterdam, De Boelelaan 1108, 1081 HZ, Amsterdam, the Netherlands

<sup>b</sup> Vetsuisse Faculty, Institute of Parasitology, University of Bern, Länggass-Strasse 122, CH-3012, Bern, Switzerland

<sup>c</sup> Laboratory of Microbiology, Parasitology and Hygiene (LMPH), University of Antwerp, Universiteitsplein 1, Wilrijk, 2610, Belgium

### ARTICLE INFO

#### Keywords:

3-nitroimidazo[1,2-*b*]pyridazine

Synthesis

*Giardia lamblia*

3',5'-cyclic nucleotide phosphodiesterase

*In vitro*

### ABSTRACT

As there is a continuous need for novel anti-infectives, the present study aimed to fuse two modes of action into a novel 3-nitroimidazo[1,2-*b*]pyridazine scaffold to improve antiparasitic efficacy. For this purpose, we combined known structural elements of phosphodiesterase inhibitors, a target recently proposed for *Trypanosoma brucei* and *Giardia lamblia*, with a nitroimidazole scaffold to generate nitrosative stress. The compounds were evaluated *in vitro* against a panel of protozoal parasites, namely *Giardia lamblia*, *Trypanosoma brucei*, *T. cruzi*, *Leishmania infantum* and *Plasmodium falciparum* and for cytotoxicity on MRC-5 cells. Interestingly, selective sub-nanomolar activity was obtained against *G. lamblia*, and by testing several analogues with and without the nitro group, it was shown that the presence of a nitro group, but not PDE inhibition, is responsible for the low IC<sub>50</sub> values of these novel compounds. Adding the favourable drug-like properties (low molecular weight, cLogP (1.2–4.1) and low polar surface area), the key compounds from the 3-nitroimidazo[1,2-*b*]pyridazine series can be considered as valuable hits for further anti-giardiasis drug exploration and development.

### 1. Introduction

Antibiotic, antiviral, antifungal and antiprotozoal resistance is on the rise worldwide and poses a formidable challenge in achieving a Universal Health Coverage, as highlighted in a recent report from the Interagency Coordination Group on Antimicrobial resistance to the UN Secretary-General. In a worst-case scenario, this is projected to lead to 10 million deaths annually by 2050 (World Health Organisation, 2019). As such, there is a continuous need for novel antimicrobials, which are not only valid for bacterial infections but also for parasitic diseases. An estimated disease burden for sixteen of the most common parasitic infections, most of them on the WHO list of neglected tropical diseases, came to about 3 billion cases with 1 out of every 6 persons worldwide experiencing 1 or more infections each year. In DALYs (Disability Adjusted Life Years), this adds up to 25 million with over 500,000 deaths annually (Herrick et al., 2017). The high disease burden combined with the threat of drug resistance calls for urgent and continuous drug discovery efforts for parasitic diseases (Cui et al., 2015; Nabarro et al.,

2015; Vanaerschot et al., 2014).

Heterocyclic aromatic nitro-compounds (imidazoles, furans and thiazoles, Fig. 1) are known for their potential in treating parasitic infections and their mode-of-action is considered to be the radical damage caused by the reactive and toxic species that are obtained from reduction of the nitro group by parasitic reductases (Ang et al., 2017). Benzimidazole (1) and nifurtimox (2) are in clinical use for Chagas disease (Ribeiro et al., 2020). Fexinidazole (3) and nifurtimox combination with eflornithine (NECT), are used for treating the African sleeping sickness (de Koning, 2020) while metronidazole (4), tinidazole (5) and nitazoxanide (6) have a role in the treatment of giardiasis (Mørch and Hanevik, 2020). The compound DNDI-VL-2098 (7) is in development for visceral leishmaniasis (Gupta et al., 2015), while nitroimidazopyridazines (8) have been reported to be effective against a broad range of parasites growing under anaerobic or semi aerobic conditions (Tomcufcik et al., 1974).

The 3',5'-cyclic nucleotide phosphodiesterases (PDEs) have a critical role in cellular signal transduction: they hydrolyse the second messengers cyclic adenosine-3',5'-monophosphate (cAMP) and cyclic

\* Corresponding author.

E-mail address: [r.leurs@vu.nl](mailto:r.leurs@vu.nl) (R. Leurs).

<sup>1</sup> These authors contributed equally.

<https://doi.org/10.1016/j.ijpddr.2022.05.004>

Received 15 January 2022; Received in revised form 18 May 2022; Accepted 20 May 2022

Available online 26 May 2022

2211-3207/© 2022 The Authors. Published by Elsevier Ltd on behalf of Australian Society for Parasitology. This is an open access article under the CC BY license (<http://creativecommons.org/licenses/by/4.0/>).

guanosine-3',5'-monophosphate (cGMP) to AMP and GMP, respectively. So far, eleven families of class-I PDEs were found (Maurice et al., 2014) and have been described as potential targets for the treatment of several parasitic diseases such as malaria (Howard et al., 2015) and Chagas disease (Sharon et al., 2010). Moreover, this class of enzymes has been proposed as potential therapeutic targets for sleeping sickness (de Koning et al., 2012; Oberholzer et al., 2007) and more recently also for giardiasis (Kunz et al., 2017a). For these last two parasitic diseases, tetrahydrophthalazinones were identified as an interesting hit series (9 and 10, Fig. 2), and initial hit optimization strategies towards parasitic PDE-selective compounds have been reported (Blaazer et al., 2018; Kunz et al., 2017a).

Previous studies have shown hybrid antimalarial compounds as a promising strategy to overcome the emergence of resistant parasite strains (Oliveira et al., 2015). Moreover, nitro-drugs have been reported to suffer from parasite resistance (Patterson and Wyllie, 2014), whereas resistance for hybrid anti-parasitic drugs has not been reported for PDE inhibitor scaffolds so far. In this study, we hypothesized that by merging a tetrahydrophthalazinone scaffold with a nitroimidazopyridazine (8), compounds with a putative dual mode-of-action could be generated, making them potentially more effective against a range of parasites. To test this hypothesis, we synthesized a series of hybrid molecules that were tested against the protozoal parasites including *G. lamblia*, *T. brucei*, *T. cruzi*, *L. infantum* and *P. falciparum*.

## 2. Materials and methods

### 2.1. Synthetic procedures

All chemicals were obtained from commercial suppliers without further purification. Reaction progress was monitored using thin-layer chromatography (TLC) and LC-MS analysis. For TLC analysis, Merck F254 alumina silica plates were used and visualized using UV. Silicycle UltraPure silica gel was used for manual purification columns. Automatic columns were performed using Biotage equipment. LC-MS analysis was performed on a Shimadzu LC-20AD liquid chromatograph pump system, equipped with an Xbridge (C18) 5  $\mu$ m column (50 mm, 4.6 mm), connected to a Shimadzu SPD-M20A diode array detector, and MS detection using a Shimadzu LC-MS-2010EV mass spectrometer. The LC-MS conditions were as follows: solvent B (acetonitrile with 0.1% formic acid) and solvent A (water with 0.1% formic acid), flow rate of 1.0 mL/min, start with 5% B, linear gradient to 90% B in 4.5 min, then 1.5 min at 90% B, then linear gradient to 5% B in 0.5 min, then 1.5 min at 5% B; total run time of 8 min. Reverse-phase column chromatography purifications were performed using Buchi PrepChrom C-700 equipment with a discharge deuterium lamp ranging from 200 to 600 nm to detect compounds using solvent B (acetonitrile with 0.1% formic acid), solvent A (water with 0.1% formic acid), flow rate of 15.0 mL/min, and a gradient (start 95% A for 3.36 min, then linear gradient to 5% A in 30 min, then at 5% A for 3.36 min, then linear gradient to 95% A in 0.5 min, and then 1.5 min at 95% A). The purity of a compound was determined

by calculating the peak area percentage by UV detection at 254 nm and was in all cases >95% except for some intermediates (indicated in the characterization). A Bruker 500 or 600 MHz spectrometer was used to record  $^1\text{H}$ ,  $^{13}\text{C}$  and 2D NMR spectra. Chemical shifts ( $\delta$  in ppm) and coupling constants ( $J$  in Hz) are reported with residual solvent as internal standard ( $\delta$   $^1\text{H}$  NMR,  $\text{CDCl}_3$  7.26,  $\text{CD}_2\text{Cl}_2$  5.32,  $\text{DMSO-}d_6$  2.50;  $\delta$   $^{13}\text{C}$  NMR,  $\text{CDCl}_3$  77.16,  $\text{CD}_2\text{Cl}_2$  53.84,  $\text{DMSO-}d_6$  39.52). Abbreviations used for  $^1\text{H}$  NMR descriptions are as follows: s = singlet, d = doublet, t = triplet, q = quintet, hept = heptet, dd = doublet of doublets, dt = doublet of triplets, tt = triplet of triplets, m = multiplet, app = apparent, br = broad signal. For HRMS analysis, a Bruker micrOTOF mass spectrometer was used using ESI in positive ion mode. All reactions were carried out under an inert nitrogen atmosphere.

#### 2.1.1. General methods

Method A: A mixture of 6-chloro-3-nitroimidazo[1,2-*b*]pyridazine (2.1 mmol, 0.42 g), an amine (2.0 mmol) and  $\text{K}_2\text{CO}_3$  (4.7 mmol, 0.65 g) in DMF (20 mL) was heated for 2–24 h at 60 °C after which the mixture was poured into water. The aqueous mixture was extracted with DCM and the combined organic layers were dried over  $\text{MgSO}_4$  and evaporated. The residue was purified by silica gel column chromatography (eluent of EtOAc and *n*-heptane) or reverse phase chromatography.

Method B: A mixture of the benzyl chloride (1.0 mmol), 12c (1.0 mmol, 0.52 g) and  $\text{K}_2\text{CO}_3$  (2.0 mmol, 0.28 g) in DMF (15 mL) was heated for 4 h at 60 °C, after which the mixture was poured into water. The aqueous mixture was extracted with DCM and the combined organic layers were dried over  $\text{MgSO}_4$  and evaporated. The residue was purified by silica gel column chromatography (eluent of EtOAc and *n*-heptane).

2.1.1.1. The intermediates are presented as supplemental data. *cis*-6-(3,4-Dimethoxybenzoyl)cyclohex-3-ene-1-carboxylic acid (11a) and (1*R*,6*S*)-6-(3,4-dimethoxybenzoyl)cyclohex-3-ene-1-carboxylic acid (11a') Prepared as described (van der Mey et al., 2002).

(4*aS*,8*aR*)-4-(3,4-Dimethoxyphenyl)-2-(piperidin-4-yl)-4*a*,5,8*a*-tetrahydrophthalazin-1(2*H*)-one-HCl (11b) Prepared as described (Sterk et al., 2004a).

*cis*-6-(3-(Cyclopentyloxy)-4-methoxybenzoyl)cyclohex-3-ene-1-carboxylic acid (12a) Prepared as described (van der Mey et al., 2002).

*cis*-4-(3-(Cyclopentyloxy)-4-methoxyphenyl)-2-(piperidin-4-yl)-4*a*,5,8*a*-tetrahydrophthalazin-1(2*H*)-one-HCl (12b) Prepared from 12a (20 mmol, 6.9 g) and 4-hydrazinopiperidine-2HCl (20 mmol, 3.8 g) as described previously (van der Mey et al., 2002). Yield: 74%; LC-MS-ESI<sup>+</sup>  $m/z$  424 [M+H]<sup>+</sup>; purity 92%.

*cis*-4-(3-Hydroxy-4-methoxyphenyl)-2-(1-(3-nitroimidazo[1,2-*b*]pyridazin-6-yl)piperidin-4-yl)-4*a*,5,8*a*-tetrahydrophthalazin-1(2*H*)-one (12c) Prepared from 12 by hydrolysing the cyclopentyl ether with 4-toluene sulfonic acid in a Dean-Stark apparatus as described previously (van der Mey et al., 2002). Yield 74%; LC-MS-ESI<sup>+</sup>  $m/z$  518 [M+H]<sup>+</sup>; purity 94%.

4-(3,4-Dimethoxyphenyl)-4-oxobutanoic acid (18a) Prepared as described (van der Mey et al., 2001).

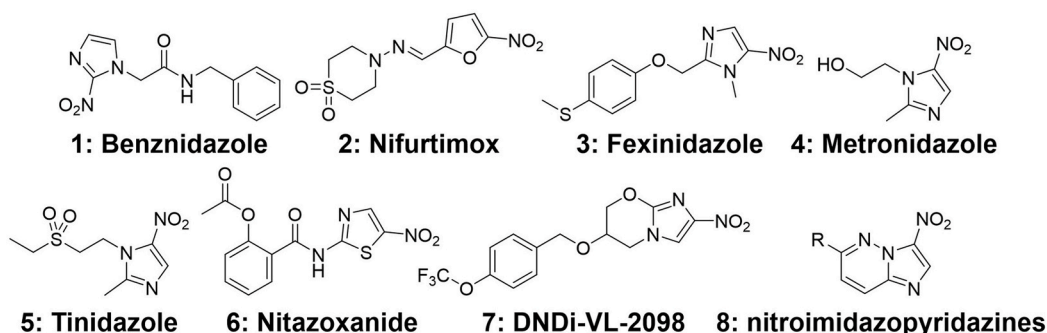


Fig. 1. Nitro-group containing heterocyclic aromatic compounds with antiparasitic activities.

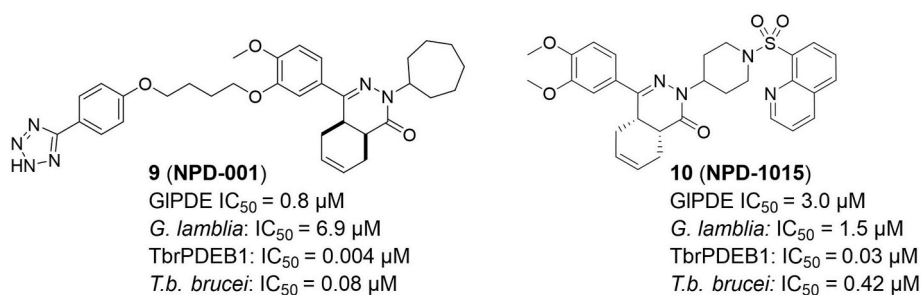


Fig. 2. Tetrahydrophthalazinones **9**, **10** and their reported IC<sub>50</sub> values against *G. lamblia* and *T. b. brucei* and relevant parasitic PDEs (Blaazer et al., 2018; Kunz et al., 2017b).

**6-(3,4-Dimethoxyphenyl)-2-(piperidin-4-yl)-4,5-dihydropyridazin-3(2H)-one-HCl (18b)** Prepared as described (Sterk et al., 2004b).

**4-(3-Chloro-4-methoxyphenyl)-4-oxobutanoic acid (19a)** Prepared from 2-chloroanisole and succinic anhydride as described for the 3,4-dimethoxy analogue (van der Mey et al., 2002). Yield: 84%; LC-MS-ESI<sup>+</sup> *m/z* 243 [M+H]<sup>+</sup>; purity 94%.

**6-(3-Chloro-4-methoxyphenyl)-2-(piperidin-4-yl)-4,5-dihydropyridazin-3(2H)-one-HCl (19b)** Prepared from 4-(3-chloro-4-methoxyphenyl)-4-oxobutanoic acid and 4-hydrazinopiperidine-2HCl as described for the 3,4-dimethoxy analogue (Sterk et al., 2004a); Yield: 67%; LC-MS-ESI<sup>+</sup> *m/z* 322 [M+H]<sup>+</sup>; purity 91%.

**2.1.1.2. Final compounds (chemical analysis spectra shown in Figs. S1–S66).** **(4aS,8aR)-4-(3,4-Dimethoxyphenyl)-2-(1-(3-nitroimidazo[1,2-b]pyridazin-6-yl)piperidin-4-yl)-4a,5,8,8a-tetrahydrophthalazin-1(2H)-one (11)** Prepared from **11b** (2.0 mmol, 0.81 g) by method A; Yield 72%. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.39 (s, 1H), 7.85 (d, *J* = 10.0 Hz, 1H), 7.39–7.35 (m, 1H), 7.29–7.24 (m, 1H), 7.16 (d, *J* = 10.1 Hz, 1H), 6.85 (d, *J* = 8.4 Hz, 1H), 5.81 (app d, *J* = 9.9 Hz, 1H), 5.70 (app d, *J* = 10.0 Hz, 1H), 4.96 (tt, *J* = 11.4, 3.9 Hz, 1H), 4.53–4.39 (m, 2H), 3.90 (s, 3H), 3.82 (s, 3H), 3.37–3.30 (m, 1H), 3.25–3.12 (m, 2H), 3.06–2.98 (m, 1H), 2.80 (app t, *J* = 5.8 Hz, 1H), 2.32–2.16 (m, 3H), 2.10–1.95 (m, 2H), 1.91–1.84 (m, 2H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.1, 155.7, 154.5, 151.0, 149.3, 137.7, 134.5, 134.3, 127.7, 126.5, 126.1, 124.0, 119.4, 113.9, 110.7, 108.4, 56.1, 56.0, 52.4, 45.61, 45.57, 34.9, 31.3, 29.5, 28.8, 23.5, 22.5. HR-MS: calc. for [M+H]<sup>+</sup>. 532.2303, found 532.2311.

**cis-4-(3-(Cyclopentylloxy)-4-methoxyphenyl)-2-(1-(3-nitroimidazo[1,2-b]pyridazin-6-yl)piperidin-4-yl)-4a,5,8,8a-tetrahydrophthalazin-1(2H)-one (12)** Prepared from **12b** (2.2 mmol, 0.92 g) by method A. Purified by crystallization from MeOH. Yield: 84%; <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.40 (s, 1H), 7.90 (d, *J* = 10.1 Hz, 1H), 7.36 (d, *J* = 2.0 Hz, 1H), 7.26–7.22 (m, 1H), 7.19 (d, *J* = 10.1 Hz, 1H), 6.84 (d, *J* = 8.4 Hz, 1H), 5.84–5.78 (m, 1H), 5.76–5.66 (m, 1H), 4.97 (tt, *J* = 11.6, 4.2 Hz, 1H), 4.77–4.69 (m, 1H), 4.52–4.40 (m, 2H), 3.86 (s, 3H), 3.31 (dt, *J* = 11.6, 5.8 Hz, 1H), 3.27–3.14 (m, 2H), 3.06–2.97 (m, 1H), 2.79 (app t, *J* = 5.9 Hz, 1H), 2.31–2.15 (m, 3H), 2.11–1.95 (m, 4H), 1.91–1.70 (m, 6H), 1.55–1.42 (m, 2H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.1, 155.6, 154.6, 152.1, 147.8, 137.3, 133.4, 127.5, 126.2, 126.1, 124.1, 119.2, 114.1, 112.3, 111.3, 80.9, 56.2, 52.3, 45.6, 34.9, 32.91, 32.88, 31.3, 29.6, 28.9, 24.19, 24.17, 23.5, 22.5. HR-MS: calc. for [M+H]<sup>+</sup>. 586.2772, found 586.2775.

**cis-4-(4-Methoxy-3-((3-nitrobenzyl)oxy)phenyl)-2-(1-(3-nitroimidazo[1,2-b]pyridazin-6-yl)piperidin-4-yl)-4a,5,8,8a-tetrahydrophthalazin-1(2H)-one (13)** Prepared from 3-nitrobenzylchloride (1.0 mmol, 0.17 g) by method B; Yield 45%. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.38 (s, 1H), 8.31 (s, 1H), 8.14 (d, *J* = 8.1 Hz, 1H), 7.89 (d, *J* = 10.0 Hz, 1H), 7.76 (d, *J* = 7.6 Hz, 1H), 7.55 (d, *J* = 7.9 Hz, 1H), 7.42 (s, 1H), 7.34–7.29 (m, 1H), 7.24 (d, *J* = 10.1 Hz, 1H), 6.91 (d, *J* = 8.5 Hz, 1H), 5.81 (app d, *J* = 9.3 Hz, 1H), 5.70 (app d, *J* = 9.5 Hz, 1H), 5.23 (s,

2H), 4.94 (ddd, *J* = 15.2, 11.2, 3.8 Hz, 1H), 4.53–4.40 (m, 2H), 3.95 (s, 3H), 3.29 (dt, *J* = 11.5, 5.7 Hz, 1H), 3.25–3.10 (m, 2H), 3.02 (app d, *J* = 17.1 Hz, 1H), 2.78 (app t, *J* = 5.8 Hz, 1H), 2.33–2.09 (m, 4H), 2.03–1.80 (m, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.0, 155.8, 154.0, 151.8, 148.5, 147.7, 139.3, 137.7, 134.5, 134.1, 133.4, 129.8, 127.7, 126.4, 126.1, 123.9, 123.2, 122.3, 120.6, 114.1, 112.0, 111.4, 70.4, 56.2, 52.3, 45.6, 45.5, 34.8, 31.2, 29.6, 28.9, 23.4, 22.4. HR-MS: calc. for [M+H]<sup>+</sup>. 653.2467, found 653.2477.

**cis-4-(4-Methoxy-3-((3-methoxybenzyl)oxy)phenyl)-2-(1-(3-nitroimidazo[1,2-b]pyridazin-6-yl)piperidin-4-yl)-4a,5,8,8a-tetrahydrophthalazin-1(2H)-one (14)** Prepared from 3-methoxybenzylchloride (1.0 mmol, 0.16 g) by method B; Yield 51%. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.39 (s, 1H), 7.93 (d, *J* = 9.8 Hz, 1H), 7.41 (d, *J* = 1.9 Hz, 1H), 7.29–7.21 (m, 3H), 7.00–6.93 (m, 2H), 6.88 (d, *J* = 8.4 Hz, 1H), 6.80 (d, *J* = 8.0 Hz, 1H), 5.84–5.78 (m, 1H), 5.72–5.66 (m, 1H), 5.17–5.09 (m, 2H), 4.94 (tt, *J* = 11.4, 4.1 Hz, 1H), 4.55–4.42 (m, 2H), 3.93 (s, 3H), 3.78 (s, 3H), 3.30–3.13 (m, 3H), 3.06–2.98 (m, 1H), 2.77 (app t, *J* = 6.0 Hz, 1H), 2.28–2.17 (m, 2H), 2.15–2.06 (m, 1H), 2.04–1.78 (m, 4H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.1, 160.0, 155.7, 154.2, 151.5, 148.2, 138.6, 137.5, 134.5, 133.7, 129.7, 127.5, 126.3, 126.0, 124.0, 119.7, 119.6, 114.1, 113.5, 113.0, 111.5, 111.1, 71.3, 56.2, 55.4, 52.2, 45.6, 45.5, 34.8, 31.2, 29.6, 28.9, 23.4, 22.4. HR-MS: calc. for [M+H]<sup>+</sup>. 638.2722, found 638.2731.

**cis-4-(4-Methoxy-3-(pyridin-3-ylmethoxy)phenyl)-2-(1-(3-nitroimidazo[1,2-b]pyridazin-6-yl)piperidin-4-yl)-4a,5,8,8a-tetrahydrophthalazin-1(2H)-one (15)** Prepared from 3-(chloromethyl)pyridine-HCl (1.0 mmol, 0.16 g) by method B; Yield 42%. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.71 (s, 1H), 8.56 (d, *J* = 6.2 Hz, 1H), 8.36 (s, 1H), 7.98 (d, *J* = 7.9 Hz, 1H), 7.83 (d, *J* = 10.0 Hz, 1H), 7.51–7.46 (m, 1H), 7.43 (d, *J* = 2.0 Hz, 1H), 7.32–7.27 (m, 2H), 6.89 (d, *J* = 8.5 Hz, 1H), 5.83–5.77 (m, 1H), 5.71–5.65 (m, 1H), 5.19 (s, 2H), 4.93 (tt, *J* = 11.4, 4.2 Hz, 1H), 4.54–4.41 (m, 2H), 3.91 (s, 3H), 3.32–3.26 (m, 1H), 3.23–3.12 (m, 2H), 3.05–2.98 (m, 1H), 2.77 (app t, *J* = 5.8 Hz, 1H), 2.27–2.11 (m, 3H), 2.06–1.92 (m, 3H), 1.88–1.82 (m, 1H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.0, 155.8, 153.9, 151.7, 147.6, 146.8, 146.1, 138.1, 137.9, 134.9, 134.6, 134.3, 127.7, 126.7, 126.1, 124.6, 123.9, 120.7, 114.1, 112.1, 111.4, 68.6, 56.2, 52.4, 45.6, 34.8, 31.2, 29.7, 28.9, 23.4, 22.4. HR-MS: calc. for [M+H]<sup>+</sup>. 609.2568, found 609.2587.

**cis-4-(4-Methoxy-3-((2-methoxybenzyl)oxy)phenyl)-2-(1-(3-nitroimidazo[1,2-b]pyridazin-6-yl)piperidin-4-yl)-4a,5,8,8a-tetrahydrophthalazin-1(2H)-one (16)** Prepared from 2-methoxybenzylchloride (1.0 mmol, 0.16 g) by method B; Yield 72%. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) δ 8.37 (s, 1H), 7.89 (d, *J* = 10.1 Hz, 1H), 7.40 (d, *J* = 7.4 Hz, 1H), 7.35 (s, 1H), 7.32 (d, *J* = 8.5 Hz, 1H), 7.25–7.16 (m, 2H), 6.92 (app t, *J* = 7.5 Hz, 1H), 6.90–6.82 (m, 2H), 5.81 (d, *J* = 9.7 Hz, 1H), 5.69 (d, *J* = 9.7 Hz, 1H), 5.26–5.17 (m, 2H), 4.93 (tt, *J* = 11.3, 3.8 Hz, 1H), 4.50 (app d, *J* = 13.4 Hz, 1H), 4.41 (app d, *J* = 13.3 Hz, 1H), 3.92 (s, 3H), 3.81 (s, 3H), 3.29–3.23 (m, 1H), 3.23–3.11 (m, 2H), 3.02 (app d, *J* = 16.7 Hz, 1H), 2.78 (app t, *J* = 5.8 Hz, 1H), 2.28–2.08 (m, 4H), 2.05–1.80 (m, 3H). <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) δ 167.0, 156.8, 155.6, 154.4, 151.5, 148.4, 137.5, 134.5, 133.8, 129.1, 128.9, 127.5, 126.3,



126.0, 125.1, 124.1, 120.8, 119.4, 113.8, 111.7, 111.3, 110.4, 66.2, 56.2, 55.6, 52.2, 45.41, 45.39, 34.8, 31.2, 29.5, 28.9, 23.4, 22.4. HR-MS: calc. for  $[M+H]^+$ . 638.2722, found 638.2737.

**cis-4-(4-Methoxy-3-(pyridin-2-ylmethoxy)phenyl)-2-(1-(3-nitroimidazo [1,2-*b*] pyridazin-6-yl)piperidin-4-yl)-4a,5,8,8a-tetrahydrophthalazin-1(2*H*)-one (17)** Prepared from 2-chloromethylpyridine-HCl (1.0 mmol, 0.16 g) by method B; Yield 62%.  $^1H$  NMR (500 MHz,  $CD_2Cl_2$ )  $\delta$  8.49 (d,  $J = 4.7$  Hz, 1H), 8.31 (s, 1H), 7.83 (d,  $J = 10.1$  Hz, 1H), 7.74 (app t,  $J = 6.5$  Hz, 1H), 7.54 (d,  $J = 6.8$  Hz, 1H), 7.47 (d,  $J = 2.0$  Hz, 1H), 7.30 (dd,  $J = 8.5, 2.0$  Hz, 1H), 7.23 (app d,  $J = 10.0$  Hz, 2H), 6.89 (d,  $J = 8.5$  Hz, 1H), 5.79–5.74 (m, 1H), 5.71–5.65 (m, 1H), 5.20 (s, 2H), 4.90 (tt,  $J = 11.5, 4.0$  Hz, 1H), 4.52–4.40 (m, 2H), 3.88 (s, 3H), 3.35–3.27 (m, 1H), 3.22–3.10 (m, 2H), 2.97–2.89 (m, 1H), 2.76 (app t,  $J = 5.9$  Hz, 1H), 2.26–2.08 (m, 3H), 2.04–1.89 (m, 3H), 1.84–1.78 (m, 1H).  $^{13}C$  NMR (126 MHz,  $CD_2Cl_2$ )  $\delta$  167.1, 157.1, 156.1, 154.1, 151.6, 149.0, 148.3, 138.5, 137.8, 134.9, 134.8, 128.0, 126.9, 126.2, 124.4, 123.3, 122.2, 120.1, 114.2, 111.44, 111.40, 71.6, 56.3, 52.5, 45.8, 45.7, 35.1, 31.3, 29.7, 29.1, 23.5, 22.7. HR-MS: calc. for  $[M+H]^+$ . 609.2568, found 609.2583.

**6-(3,4-Dimethoxyphenyl)-2-(1-(3-nitroimidazo [1,2-*b*] pyridazin-6-yl)piperidin-4-yl)-4,5-dihydropyridazin-3(2*H*)-one (18)** Prepared from **18b** (2.0 mmol, 0.71 g) by method A; Yield 72%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.39 (s, 1H), 7.83 (d,  $J = 10.1$  Hz, 1H), 7.32 (d,  $J = 2.3, 1H$ ), 7.22 (dd,  $J = 8.4, 2.3$  Hz, 1H), 7.16 (d,  $J = 9.9$  Hz, 1H), 6.85 (d,  $J = 8.5$  Hz, 1H), 4.94 (tt,  $J = 11.6, 4.2$  Hz, 1H), 4.46 (d,  $J = 13.4$  Hz, 2H), 3.90 (s, 3H), 3.82 (s, 3H), 3.24–3.13 (m, 3H), 2.97–2.86 (m, 2H), 2.61 (app t,  $J = 8.2$  Hz, 2H), 2.15 (app qd,  $J = 12.6, 4.4, 2H$ ), 1.93 (app dd,  $J = 13.5, 3.9$  Hz, 2H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$  165.3, 155.6, 150.9, 150.6, 149.1, 137.9, 134.7, 128.6, 126.6, 119.4, 113.8, 110.6, 108.6, 77.2, 56.1, 56.0, 52.1, 45.6, 29.2, 27.6, 22.4. HR-MS: calc. for  $[M+H]^+$ . 480.1990, found 480.1990.

**6-(3-Chloro-4-methoxyphenyl)-2-(1-(3-nitroimidazo [1,2-*b*] pyridazin-6-yl)piperidin-4-yl)-4,5-dihydropyridazin-3(2*H*)-one (19)** Prepared from **19b** (2.0 mmol, 0.72 g) by method A; Yield 72%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.40 (s, 1H), 7.89 (d,  $J = 10.0$  Hz, 1H), 7.78–7.74 (m, 1H), 7.56 (dd,  $J = 8.6, 1.7$  Hz, 1H), 7.19 (d,  $J = 10.1$  Hz, 1H), 6.91 (d,  $J = 8.7$  Hz, 1H), 4.95 (tt,  $J = 11.5, 3.8$  Hz, 1H), 4.48 (app d,  $J = 13.4$  Hz, 2H), 3.92 (s, 3H), 3.17 (app t,  $J = 12.5$  Hz, 2H), 2.89 (t,  $J = 8.2$  Hz, 2H), 2.62 (t,  $J = 8.2$  Hz, 2H), 2.15 (app td,  $J = 12.5, 3.9$  Hz, 2H), 1.92 (app d,  $J = 11.1$  Hz, 2H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$  165.0, 156.2, 155.5, 149.1, 137.4, 133.7, 132.5, 129.0, 127.7, 126.3, 125.5, 122.9, 113.9, 111.6, 56.3, 52.0, 45.3, 29.1, 27.3, 22.2. HR-MS: calc. for  $[M+H]^+$ . 484.1495, found 484.1500.

**6-(3,4-Dimethoxyphenyl)-2-(1-(imidazo [1,2-*b*] pyridazin-6-yl)piperidin-4-yl)-4,5-dihydropyridazin-3(2*H*)-one (20)** Prepared from **18b** (2.0 mmol, 0.71 g) and 6-chloroimidazo[1,2-*b*]pyridazine (2.1 mmol, 0.32 g) by method A, but heating at 60 °C for 24 h; Yield 19%.  $^1H$  NMR (600 MHz,  $CDCl_3$ )  $\delta$  7.89 (d,  $J = 9.9$  Hz, 1H), 7.69 (s, 1H), 7.58 (s, 1H), 7.35 (d,  $J = 1.9$  Hz, 1H), 7.24 (dd,  $J = 8.4, 2.0$  Hz, 1H), 6.97 (d,  $J = 10.0$  Hz, 1H), 6.87 (d,  $J = 8.4$  Hz, 1H), 4.94 (tt,  $J = 11.6, 4.1$  Hz, 1H), 4.29 (app d,  $J = 13.4$  Hz, 2H), 3.92 (s, 3H), 3.79 (s, 3H), 3.16–3.05 (m, 2H), 2.93 (t,  $J = 8.2$  Hz, 2H), 2.69–2.58 (m, 2H), 2.17 (app qd,  $J = 12.6, 4.1$  Hz, 2H), 1.94–1.85 (m, 2H).  $^{13}C$  NMR (151 MHz,  $CDCl_3$ )  $\delta$  165.3, 155.2, 150.9, 150.4, 149.2, 135.6, 129.9, 128.6, 125.3, 119.3, 116.7, 111.6, 110.7, 108.5, 56.1, 55.9, 52.2, 46.1, 29.1, 27.6, 22.3. HR-MS: calc. for  $[M+H]^+$ . 435.2139, found 435.2153.

***N,N*-Dimethyl-3-nitroimidazo [1,2-*b*] pyridazin-6-amine (21)** Prepared from dimethylamine hydrochloride by method A; Yield 71%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.37 (s, 1H), 7.82 (d,  $J = 10.0$  Hz, 1H), 7.03 (d,  $J = 10.0$  Hz, 1H), 3.22 (s, 6H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$  155.8, 137.7, 134.5, 134.2, 126.3, 112.8, 38.7. HR-MS: calc. for  $[M+H]^+$ . 208.0829, found 208.0829.

**3-Nitro-6-(pyrrolidin-1-yl)imidazo [1,2-*b*] pyridazine (22)** Prepared from pyrrolidine by method A; Yield 65%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.35 (s, 1H), 7.78 (d,  $J = 9.9$  Hz, 1H), 6.86 (d,  $J = 9.9$  Hz, 1H), 3.63–3.54 (m, 4H), 2.13–2.04 (m, 4H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$

153.8, 138.0, 134.4, 134.0, 126.2, 113.8, 47.4, 25.5. HR-MS: calc. for  $[M+H]^+$ . 234.0986, found 234.0996. Spectral data are in agreement with a previous report (Sridhar et al., 2017).

**3-Nitro-6-(piperidin-1-yl)imidazo [1,2-*b*] pyridazine (23)** Prepared from piperidine by method A; Yield 61%.  $^1H$  NMR (600 MHz,  $CDCl_3$ )  $\delta$  8.37 (s, 1H), 7.81 (d,  $J = 10.0$  Hz, 1H), 7.12 (d,  $J = 10.0$  Hz, 1H), 3.65 (app s, 4H), 1.72 (app s, 6H).  $^{13}C$  NMR (151 MHz,  $CDCl_3$ )  $\delta$  156.1, 138.0, 134.8, 134.4, 126.4, 114.2, 47.3, 25.7, 24.6. HR-MS: calc. for  $[M+H]^+$ . 248.1142, found 248.1149. Spectral data are in agreement with a previous report (Sridhar et al., 2017).

**6-(4-Methylpiperazin-1-yl)-3-nitroimidazo [1,2-*b*] pyridazine formate (24)** Prepared from *N*-methylpiperazine by method A; Yield 45%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.41 (s, 1H), 8.24 (s, 1H), 7.88 (d,  $J = 10.0$  Hz, 1H), 7.10 (d,  $J = 10.0$  Hz, 1H), 5.42 (s, 1H), 3.81 (t,  $J = 5.2$  Hz, 4H), 2.84 (t,  $J = 5.1$  Hz, 4H), 2.51 (s, 3H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$  165.5, 155.5, 138.0, 134.9, 133.9, 127.0, 113.4, 53.5, 45.0, 44.8. HR-MS: calc. for  $[M+H]^+$ . 263.1251, found 263.1257.

**3-Nitro-6-(4-(pyrrolidin-1-yl)piperidin-1-yl)imidazo [1,2-*b*] pyridazine formate (25)** Prepared from 4-(pyrrolidin-1-yl)-piperidine by method A; Yield 46%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.43 (s, 1H), 8.39 (s, 1H), 7.84 (d,  $J = 10.0$  Hz, 1H), 7.11 (d,  $J = 10.0$  Hz, 1H), 6.11 (s, 1H), 4.41 (app d,  $J = 13.6$  Hz, 2H), 3.17 (app s, 4H), 3.02 (app t,  $J = 12.1$  Hz, 3H), 2.20 (app d,  $J = 12.2$  Hz, 2H), 2.09–1.90 (m, 6H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$  167.9, 155.4, 138.0, 135.0, 134.6, 126.9, 113.7, 61.5, 50.6, 44.9, 28.5, 23.4. HR-MS: calc. for  $[M+H]^+$ . 317.1721, found 317.1732.

**3-Nitro-6-(4-phenylpiperidin-1-yl)imidazo [1,2-*b*] pyridazine (26)** Prepared from 4-phenylpiperidine by method A; Yield 46%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.41 (s, 1H), 7.87 (d,  $J = 10.0$  Hz, 1H), 7.35 (t,  $J = 7.6$  Hz, 2H), 7.27–7.23 (m, 3H), 7.20 (d,  $J = 10.1$  Hz, 1H), 4.50 (app d,  $J = 13.4$  Hz, 2H), 3.16 (app td,  $J = 13.1, 2.1$  Hz, 2H), 2.86 (tt,  $J = 12.2, 3.4$  Hz, 1H), 2.06 (app d,  $J = 13.1$  Hz, 2H), 1.90–1.79 (m, 2H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$  155.8, 145.1, 137.6, 134.5, 133.8, 128.8, 126.9, 126.8, 126.3, 114.2, 46.9, 42.6, 32.8. HR-MS: calc. for  $[M+H]^+$ . 324.1455, found 324.1440.

**3-Nitro-6-(4-phenylpiperazin-1-yl)imidazo [1,2-*b*] pyridazine (27)** Prepared from *N*-phenylpiperazine by method A; Yield 49%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.41 (s, 1H), 7.87 (d,  $J = 10.0$  Hz, 1H), 7.32 (app t,  $J = 7.8$  Hz, 2H), 7.16 (d,  $J = 10.0$  Hz, 1H), 7.02 (app d,  $J = 6.2$  Hz, 2H), 6.95 (t,  $J = 6.9$  Hz, 1H), 3.86 (app s, 4H), 3.41–3.33 (m, 4H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$  155.7, 150.9, 138.1, 134.9, 134.6, 129.5, 126.9, 120.8, 116.7, 113.4, 49.2, 45.9. HR-MS: calc. for  $[M+H]^+$ . 325.1408, found 325.1405. Spectral data are in agreement with a previous report (Sridhar et al., 2017).

**6-(4-Benzylpiperidin-1-yl)-3-nitroimidazo [1,2-*b*] pyridazine (28)** Prepared from 4-benzylpiperidine by method A; Yield 67%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.37 (s, 1H), 7.79 (d,  $J = 10.0$  Hz, 1H), 7.34–7.28 (m, 2H), 7.22 (t,  $J = 7.4$  Hz, 1H), 7.16 (d,  $J = 7.4$  Hz, 2H), 7.10 (d,  $J = 10.0$  Hz, 1H), 4.31 (app d,  $J = 13.6$  Hz, 2H), 3.02–2.90 (m, 2H), 2.59 (d,  $J = 6.8$  Hz, 2H), 1.91–1.78 (m, 3H), 1.40–1.28 (m, 2H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$  155.7, 139.9, 137.7, 134.2, 129.2, 128.5, 126.31, 126.26, 114.0, 77.2, 46.4, 43.1, 38.1, 31.6. HR-MS: calc. for  $[M+H]^+$ . 338.1612, found 338.1614.

**(1-(3-Nitroimidazo [1,2-*b*] pyridazin-6-yl)piperidin-4-yl) (phenyl)methanone (29)** Prepared from 4-benzoylpiperidine by method A; Yield 59%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.39 (s, 1H), 7.97 (d,  $J = 7.3$  Hz, 2H), 7.86 (d,  $J = 10.0$  Hz, 1H), 7.60 (t,  $J = 7.4$  Hz, 1H), 7.51 (t,  $J = 7.7$  Hz, 2H), 7.17 (d,  $J = 10.1$  Hz, 1H), 4.36 (app d,  $J = 13.5$  Hz, 2H), 3.60 (tt,  $J = 10.8, 3.8$  Hz, 1H), 3.32–3.20 (m, 2H), 2.12–2.02 (m, 2H), 2.00–1.87 (m, 2H).  $^{13}C$  NMR (126 MHz,  $CDCl_3$ )  $\delta$  201.7, 155.7, 137.8, 135.7, 134.5, 134.3, 133.5, 129.0, 128.4, 126.5, 114.0, 45.8, 43.1, 28.0. HR-MS: calc. for  $[M+H]^+$ . 352.1404, found 352.1417.

***N*-(4-Methoxybenzyl)-*N*-methyl-3-nitroimidazo [1,2-*b*] pyridazin-6-amine (30)** Prepared from *N*-(4-methoxybenzyl)methylamine by method A; Yield 62%.  $^1H$  NMR (500 MHz,  $CDCl_3$ )  $\delta$  8.38 (s, 1H), 7.79 (d,  $J = 10.0$  Hz, 1H), 7.28 (d,  $J = 8.5$  Hz, 2H), 6.99 (d,  $J = 10.0$  Hz, 1H),

6.87 (d,  $J = 8.5$  Hz, 2H), 4.74 (s, 2H), 3.79 (s, 3H), 3.26 (s, 3H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  159.3, 155.4, 137.7, 134.4, 134.1, 128.9, 128.6, 126.4, 114.3, 113.1, 55.4, 53.9, 37.3. HR-MS: calc. for  $[\text{M}+\text{H}]^+$ . 314.1248, found 314.1247.

**6-(Piperidin-1-yl)imidazo[1,2-*b*]pyridazine (31)** Prepared from piperidine and 6-chloroimidazo[1,2-*b*]pyridazine instead of 6-chloro-3-nitroimidazo[1,2-*b*]pyridazine by method A, but heating at 60 °C for 24 h; Yield 21%.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.68 (s, 1H), 7.66 (s, 1H), 7.51 (s, 1H), 6.81 (d,  $J = 9.9$  Hz, 1H), 3.47 (app s, 4H), 1.67 (app s, 6H).  $^{13}\text{C}$  NMR (126 MHz,  $\text{CDCl}_3$ )  $\delta$  155.4, 136.4, 131.7, 125.8, 116.6, 110.7, 47.6, 25.5, 24.6. HR-MS: calc. for  $[\text{M}+\text{H}]^+$ . 203.1291, found 203.1291.

## 2.2. In vitro tests

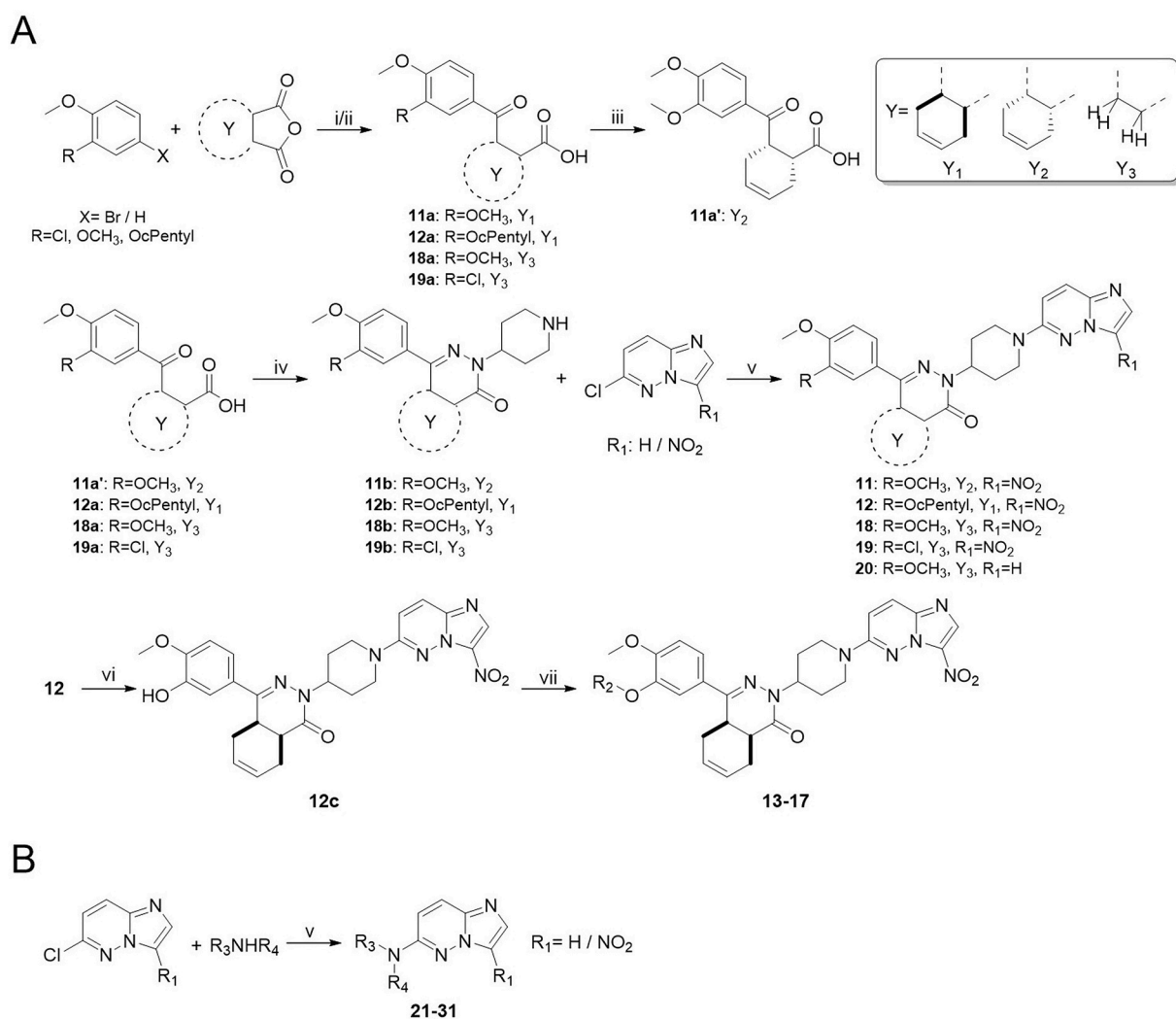
*G. lamblia* trophozoites (WBC6) were seeded into 96-well-plates in 0.2 mL Keister-medium containing  $10^3$  trophozoites per well in the presence of the compounds dissolved in dimethyl sulfoxide (DMSO, final concentration 0.25%) or with DMSO as a solvent control and incubated for 72 h at 37 °C in an anaerobic chamber (80%  $\text{N}_2$ , 10%  $\text{H}_2$ , 10%  $\text{CO}_2$ ). Then, plates were centrifuged (2 min, 1000 rpm), medium was removed, and cells were washed once with PBS containing 1 g/L glucose. Then, PBS-glucose containing 10 mg/L resazurin was added, and plates were incubated for up to 5 h at 37 °C. Fluorescence reading (ex 530 nm/em

590 nm) was performed at various time points, and the linear increase of fluorescence was used as a marker for cell viability.  $\text{IC}_{50}$  values were calculated using the logit-log algorithm and are given as mean values with 95%-confidence intervals (Müller et al., 2020; Müller and Hemp-hill, 2013; Müller and Müller, 2019). The assays for growth inhibition of *T. b. brucei*, *T. cruzi*, *L. infantum*, *P. falciparum* and MRC-5 (human lung fibroblasts MRC-5<sub>SV40</sub>) were previously described (Blaazer et al., 2014). PDE activity was determined exactly as described (de Heuvel et al., 2021; Kunz et al., 2005, 2017b). All assays were carried out in triplicates and no more than 20% substrate was hydrolyzed in all reactions. Compounds were dissolved in DMSO and the final DMSO concentration was 1% in all reaction mixes. Control reactions with DMSO alone were always included. Data were analyzed using the GraphPad Prism software package (v7.0, GraphPad, San Diego).

## 3. Results and discussion

### 3.1. Synthesis of 3-nitroimidazo[1,2-*b*]pyridazine analogues

Compounds **11-20** were prepared according to Scheme 1A. In the case R is MeO or Cl, X is H and the sequence started with a Friedel–Crafts acylation (step i) using succinic anhydride or tetrahydrophthalic anhydride. If R is a cyclopentyloxy group, X is Br and the sequence started



**Scheme 1.** Synthesis of analogues **11-31**. Reagents and conditions: i for **11a**, **18a**, **19a**:  $\text{AlCl}_3$ , DCM, rt, 4h; ii for **12a**: Mg, THF, rt followed by 2 h reflux; iii: crystallization ( $\times 3$ ) with (*S*)-(-)- $\alpha$ -methylbenzylamine from EtOAc; iv: 4-hydrazinopiperidine-2HCl, TEA, EtOH, reflux, 16 h; v: DMF,  $\text{K}_2\text{CO}_3$ , 60 °C, 2–24 h; vi: Dean-Stark, 4-toluenesulfonic acid, toluene, reflux, 4 h; vii:  $\text{R}_2\text{Cl}$ ,  $\text{K}_2\text{CO}_3$ , DMF, 60 °C, 4 h.

with a Grignard reaction (step ii). In both cases  $\gamma$ -ketocarboxylic acids were obtained. Enantiomeric acid **11a'** was obtained through resolution with (*S*)-(-)- $\alpha$ -methylbenzylamine in EtOAc from **11a** (step iii). The next step (iv), condensation with 4-hydrazinopiperidine, was performed in refluxing ethanol and resulted in the substituted piperidines, which were further *N*-substituted with an imidazopyridazine in DMF using  $K_2CO_3$  to scavenge the HCl. As described previously (van der Mey et al., 2002), the cyclopentyl group of **12** can be hydrolyzed selectively using 4-toluenesulfonic acid in a Dean-Stark apparatus and the resulting phenol **12c** was substituted with commercially available benzyl chlorides or pyridylmethyl chlorides under standard conditions. All final compounds were prepared as racemic mixtures of the *cis*-diastereomers (4*aS*,8*aR* and 4*aR*,8*aS*) except for compound **11** (4*aS*, 8*aR*). Scheme 1B depicts the synthesis of 6-amino-imidazopyridazine analogues (**21–31**) using conditions essentially identical to step v in Scheme 1A.

### 3.2. Antiparasitic activities of 3-nitroimidazo[1,2-*b*]pyridazine analogues

As PDE inhibitors and nitro heterocyclic compounds both have therapeutic potential in protozoal infections, we combined the tetrahydrophthalazinone scaffold with a nitroimidazopyridazine aiming for two modes of action into one hybrid molecule with improved efficacy. For this purpose, we first prepared the tetrahydrophthalazinones **11–17** with variations in  $R_2$ , the *meta* position of the phenyl ring, and tested them against the panel of protozoa and for cytotoxicity on the human MRC-5 cell line (using tamoxifen as positive control) together with the benchmark compounds metronidazole, miltefosine, chloroquine, benznidazole and suramin. Activities of control antiparasitic agents were within previously described ranges (Bénére et al., 2007; El Sayed et al., 2009; van Baelen et al., 2008; Venkatraj et al., 2014) and the results for **11–20** are summarized in Table 1 with representative dose-response

curves shown in Fig. S67. All compounds show excellent anti-*Giardia* activity and no or very weak toxicity (Table 1) with high selectivity over other parasites (Table S1). Notably, all these novel tetrahydrophthalazinones have much higher anti-*Giardia* activity than the clinically used metronidazole ( $IC_{50} = 0.8 \mu M$ ): a 40-fold (chlorine derivative **19**,  $IC_{50} = 19.0 \text{ nM}$ ) to an exceptional >1,000-fold (methoxy and 2-pyridyl-methoxy derivatives **11** and **17**,  $IC_{50} = 0.5 \text{ nM}$  for both).

As mentioned earlier, tetrahydrophthalazinones are known PDE inhibitors of selected human (e.g. human PDE4) and parasitic enzymes (e.g. GIPDE). To investigate if PDE inhibition is contributing to the high anti-*Giardia* activity of **11**, we determined its activity as a GIPDE inhibitor (Table 2). Moreover, in light of the known interaction of tetrahydrophthalazinones with human PDE4, also the activity against the off-target human PDE4 (Houslay et al., 2005; Manallack et al., 2005; Tenor et al., 2011) was measured. As can be seen in Table 2, **11** is a moderate inhibitor of GIPDE ( $K_i = 0.5 \mu M$ , Fig. S69), but is 3,000-fold more active against human PDE4 ( $K_i = 0.16 \text{ nM}$ , Fig. S68). As such potent human PDE4 inhibition will prevent any further development because of foreseen unwanted adverse effects (Souness et al., 2000; Tenor et al., 2011), we aimed at modifying the scaffold in such a way that the anti-*Giardia* activity would be retained while the potency against human PDE4 would be strongly reduced. Based on expert knowledge from previous structure-activity relationship (SAR) studies for PDE4 inhibition (van der Mey et al., 2001, 2002; Veerman et al., 2016), we decided to remove the cyclohexene ring  $Y_1/Y_2$  (Table 1) and replace one of the ethers ( $R_2$ , Table 1) by a chlorine, resulting in the pyridazinones **18** and **19**. Removing the cyclohexene ring as in **18** resulted in a 2.5-fold increase in anti-*Giardia* potency (Table 1,  $IC_{50} = 0.2 \text{ nM}$ ), while the hPDE4 affinity was 20-fold decreased (Table 2). An additional replacement of the *meta*-methoxy group by a chlorine (**19**) decreased the anti-*Giardia* activity 100-fold while the affinity for hPDE4B was 60-fold decreased

**Table 1**

Anti-protozoal activity and toxicity of pyridazines and tetrahydrophthalazinones.

Cmpd	$R_1$	$R_2$	Y	M.W.	cLogP <sup>a</sup>	tPSA <sup>a</sup>	IC <sub>50</sub> (nM)	
							G.L. <sup>b</sup>	MRC-5 <sup>c</sup>
<b>11</b>	NO <sub>2</sub>	MeO	Y <sub>1</sub>	531.6	3.3	127.7	0.5 ± 0.2	>6.4 × 10 <sup>4</sup>
<b>12</b>	NO <sub>2</sub>	cyclopentylloxy	Y <sub>2</sub>	585.7	4.6	127.7	9.3 ± 0.6	>6.4 × 10 <sup>4</sup>
<b>13</b>	NO <sub>2</sub>		Y <sub>2</sub>	652.7	5.0	170.8	1.2 ± 0.4	>6.4 × 10 <sup>4</sup>
<b>14</b>	NO <sub>2</sub>		Y <sub>2</sub>	637.7	4.9	136.9	1.0 ± 0.5	>6.4 × 10 <sup>4</sup>
<b>15</b>	NO <sub>2</sub>		Y <sub>2</sub>	608.7	3.8	140.6	6.8 ± 1.5	15,000 ± 11,000
<b>16</b>	NO <sub>2</sub>		Y <sub>2</sub>	637.7	4.9	136.9	2.8 ± 0.8	49,000 ± 21,000
<b>17</b>	NO <sub>2</sub>		Y <sub>2</sub>	608.7	3.9	140.6	0.5 ± 0.1	>6.4 × 10 <sup>4</sup>
<b>18</b>	NO <sub>2</sub>	MeO	Y <sub>3</sub>	479.5	2.1	127.7	0.2 ± 0.1	>6.4 × 10 <sup>4</sup>
<b>19</b>	NO <sub>2</sub>	Cl	Y <sub>3</sub>	483.9	2.9	118.5	19.0 ± 3.2	>6.4 × 10 <sup>4</sup>
<b>20</b>	H	MeO	Y <sub>3</sub>	434.5	2.1	84.5	930 ± 100	>6.4 × 10 <sup>4</sup>
<b>3 (MET)</b>	-	-	-	171.2	-0.5	81.2	800 ± 100	-
<b>Tamoxifen</b>	-	-	-	371.5	6.8	12.5	-	11,000 ± 5,000

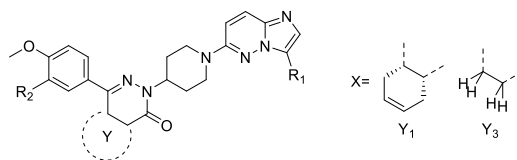
MET: metronidazole; G.L.: *Giardia lamblia*.

<sup>a</sup> cLogP, tPSA are calculated using Collaborative Drug Discovery (CDD) Vault.

<sup>b</sup> mean values ± standard errors are given for quadruplicates, the standard errors correspond to the 95% confidence intervals as calculated via the logit-log algorithm.

<sup>c</sup> mean values ± standard deviations,  $n \geq 2$ .

**Table 2**  
PDE inhibiting activity of selected compounds.



Cmpd	R <sub>1</sub>	R <sub>2</sub>	Y	M.W.	cLogP <sup>a</sup>	tPSA <sup>a</sup>	G.L. IC <sub>50</sub> (nM) <sup>b</sup>	GIPDE K <sub>i</sub> (nM) <sup>c</sup>	hpPDE4B K <sub>i</sub> (nM) <sup>c</sup>
11	NO <sub>2</sub>	MeO	Y <sub>1</sub>	531.6	3.3	127.7	0.5 ± 0.2	500 ± 300	0.16 ± 0.02
18	NO <sub>2</sub>	MeO	Y <sub>3</sub>	479.5	2.1	127.7	0.2 ± 0.1	>10 <sup>4</sup>	3.2 ± 0.4
19	NO <sub>2</sub>	Cl	Y <sub>3</sub>	483.9	2.9	118.5	19.0 ± 3.2	>10 <sup>6</sup>	200 ± 50
20	H	MeO	Y <sub>3</sub>	434.5	2.1	84.5	930 ± 100	>10 <sup>6</sup>	5.0 ± 1.0

<sup>a</sup> cLogP, tPSA are calculated using CDD Vault.

<sup>b</sup> mean values ± standard errors are given for quadruplicates, the standard errors correspond to the 95% confidence intervals as calculated via the logit-log algorithm.

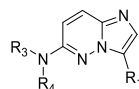
<sup>c</sup> mean values ± standard deviations, n ≥ 2.

compared to **18**. In this series of tetrahydrophthalazonones/pyridazinones, **18** is our most potent anti-*Giardia* compound with an IC<sub>50</sub> value on growth inhibition of ~200 pM.

For the compounds **11**, **18–20** we also measured their GIPDE

activities (Table 2). From these data, one can conclude that it is highly unlikely that inhibition of this PDE significantly contributes to the anti-parasitic efficacy of these compounds against *Giardia*. This is especially evident for our most potent compound **18**, with only a K<sub>i</sub> value of >10

**Table 3**  
Antiprotozoal activity and toxicity of 6-substituted imidazo[1,2-*b*]pyridazines.



Cmpd	R <sub>1</sub>	NR <sub>3</sub> R <sub>4</sub>	M.W.	cLogP <sup>a</sup>	tPSA <sup>a</sup>	IC <sub>50</sub> (nM)	
						G.L. <sup>b</sup>	MRC-5 <sup>c</sup>
21	NO <sub>2</sub>		207.2	1.4	76.6	22.0 ± 3.0	>6.4 × 10 <sup>4</sup>
22	NO <sub>2</sub>		233.2	1.8	76.6	2.3 ± 0.4	>6.4 × 10 <sup>4</sup>
23	NO <sub>2</sub>		247.3	2.2	76.6	3.6 ± 0.8	28,100 ± 1,400
24 <sup>d</sup>	NO <sub>2</sub>		308.3	1.2	79.8	144.0 ± 31.0	>6.4 × 10 <sup>4</sup>
25 <sup>d</sup>	NO <sub>2</sub>		316.4	1.8	79.8	5.8 ± 0.9	>6.4 × 10 <sup>4</sup>
26	NO <sub>2</sub>		323.4	3.7	76.6	8.2 ± 1.0	>6.4 × 10 <sup>4</sup>
27	NO <sub>2</sub>		324.3	3.1	79.8	0.7 ± 0.1	>6.4 × 10 <sup>4</sup>
28	NO <sub>2</sub>		337.4	4.1	76.6	0.5 ± 0.1	7,960 ± 210
29	NO <sub>2</sub>		351.4	3.2	93.6	1.1 ± 0.2	>6.4 × 10 <sup>4</sup>
30	NO <sub>2</sub>		313.3	2.9	85.8	10.3 ± 1.5	>6.4 × 10 <sup>4</sup>
31	H		202.3	2.2	33.4	>10 <sup>4</sup>	>6.4 × 10 <sup>4</sup>
2 (MET)	-	-	171.2	-0.5	81.2	800 ± 100	-

<sup>a</sup> cLogP, tPSA are calculated using CDD Vault.

<sup>b</sup> mean values ± standard errors are given for quadruplicates, the standard errors correspond to the 95% confidence intervals as calculated via the logit-log algorithm.

<sup>c</sup> mean values ± standard deviations, n ≥ 2.

<sup>d</sup> formate.



$\mu\text{M}$  on GIPDE, which is over 50,000-fold more than the  $\text{IC}_{50}$  value (0.2 nM) for *Giardia* growth inhibition.

To find out if this class of compounds has any other activity, independent of the nitro group, which might contribute to their anti-*Giardia* efficacy, we decided to prepare **20**, an analogue of **18** lacking the nitro group. As **20** is 4,500-fold less active than its nitro analogue **18**, we concluded that the main mode-of-action of **18** is directly related to the nitro group. The remaining anti-*Giardia* activity of **20** ( $\text{IC}_{50} = 0.93 \mu\text{M}$ ) that is equipotent to metronidazole is also not related to GIPDE inhibition, since also **20** shows no GIPDE inhibition (Table 2), indicating that PDE inhibition should not be regarded as an additional mode-of-action of these anti-*Giardia* compounds. Meanwhile, imidazo[1,2-*b*]pyridazines have been previously reported as kinase inhibitors for *P. falciparum* and *Toxoplasma gondii*, which could serve as a potential mechanism for the remaining activity of **20** (Garrido et al., 2021).

To better understand the potential of nitroimidazopyridazines in designing novel and potent compounds with anti-*Giardia* activity, the relatively “simple” analogues in Table 3 lacking the tetrahydrophthalazinone part associated with the off-target PDE4 inhibition were prepared and tested for antiparasitic efficacy and cytotoxicity (Table 3). Surprisingly, also in this series analogues with nano- to picomolar  $\text{IC}_{50}$  values against *G. lamblia* were identified. None of the analogues showed relevant activity against the other protozoa (Table S1). Anti-*Giardia* activities range from 144 nM (24,  $\text{NR}_{3\text{R}_4} = N\text{-methylpiperazine}$ ) to 500 pM (28,  $\text{NR}_{3\text{R}_4} = 4\text{-benzylpiperidine}$ ). Also for this series, the importance of the nitro group was investigated. Compound **31**, the non-nitro analogue of **23**, was prepared and proved to be inactive (Table 3). The difference between both compounds in inhibiting growth of *G. lamblia* is larger than 2,500 folds, providing strong evidence that the nitro group is of main importance for the activity of this scaffold. Like the compounds in Table 1, also the compounds in Table 3 have negligible toxicity on the human cell line MRC-5.

The importance of physicochemical properties of potential new drugs has become a mainstay in drug development since the formulation of the Lipinski rule of 5 (Ro5) (Lipinski et al., 1997). Still, the use of this Ro5 should be limited to oral availability of systemically active drugs. Moreover, it was clearly stated by McKerrow and Lipinski that this Ro5 is not applicable to potential anti-parasitic drugs (McKerrow and Lipinski, 2017). Despite these limitations of the Ro5, for our project we are convinced that physicochemical properties for targeting gastrointestinal parasites are important. Though the optimal values are not known, to our knowledge, it seems plausible that low lipophilicity (cLogP), low polar surface area (tPSA) and low molecular weight must be important, e.g. for proper solubility and membrane passage to enter parasites. Therefore, we believe that the result from this study, compounds which combine high anti-parasite potency with good calculated physicochemical properties (cLogP: 1.2–4.1; tPSA: 77–94 Å; molecular weight: 202.3–351.4 Dalton) warrants detailed evaluation of this class of compounds for their potential in the treatment of giardiasis. Importantly, the known nitro-resistance of a number of parasites, including *Giardia lamblia* (Nabarro et al., 2015), might limit the ultimate usefulness of this class of compounds and warrants future studies with resistant parasite strains.

In summary, we have identified novel nitroimidazopyrimidine compounds that have nanomolar to picomolar activity against *G. lamblia* without significant toxicity against human MRC-5 cells. Though this study was set up to find novel compounds with dual activity, their potency seems to rely mainly on the nitro group. In view of their good drug-like properties, especially low lipophilicity, low polar surface area and the low molecular weight, 3-nitroimidazo[1,2-*b*]pyridazines are promising candidates for further lead optimization studies.

#### Declaration of competing interest

The authors declared that there is no conflict of interest.

#### Acknowledgment

This work was supported by the European Commission 7th Framework Program FP7-HEALTH-2013-INNOVATION-1 under project reference 602666 “Parasite-specific cyclic nucleotide phosphodiesterase inhibitors to target Neglected Parasitic Diseases” (PDE4NPD). YZ acknowledges the China Scholarship Council (CSC) for funding (Grant No. 201506220185). The authors thank Hans Custers and Andrea van de Stolpe for their technical assistance.

#### Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.ijpddr.2022.05.004>.

#### References

- Ang, C.W., Jarrad, A.M., Cooper, M.A., Blaskovich, M.A.T., 2017. Nitroimidazoles: molecular fireworks that combat a broad spectrum of infectious diseases. *J. Med. Chem.* 60, 7636–7657. <https://doi.org/10.1021/acs.jmedchem.7b00143>.
- Bénére, E., da Luz, R.A.I., Vermeersch, M., Cos, P., Maes, L., 2007. A new quantitative in vitro microculture method for *Giardia duodenalis* trophozoites. *J. Microbiol. Methods* 71, 101–106. <https://doi.org/10.1016/J.MIMET.2007.07.014>.
- Blaazer, A.R., Orrling, K.M., Shanmugham, A., Jansen, C., Maes, L., Edink, E., Sterk, G.J., Siderius, M., England, P., Bailey, D., de Esch, I.J.P., Leurs, R., 2014. Fragment-based screening in tandem with phenotypic screening provides novel antiparasitic hits. *J. Biomol. Screen* 20, 131–140. <https://doi.org/10.1177/1087057114549735>.
- Blaazer, A.R., Singh, A.K., de Heuvel, E., Edink, E., Orrling, K.M., Veerman, J.J.N., van den Bergh, T., Jansen, C., Balasubramanian, E., Mooij, W.J., Custers, H., Sijm, M., Tagoe, D.N.A., Kalejaiye, T.D., Munday, J.C., Tenor, H., Matheussen, A., Wijtmans, M., Siderius, M., de Graaf, C., Maes, L., de Koning, H.P., Bailey, D.S., Sterk, G.J., de Esch, I.J.P., Brown, D.G., Leurs, R., 2018. Targeting a subpocket in trypanosoma brucei phosphodiesterase B1 (TbrPDEB1) enables the structure-based discovery of selective inhibitors with trypanocidal activity. *J. Med. Chem.* 61, 3870–3888. <https://doi.org/10.1021/acs.jmedchem.7b01670>.
- Cui, L., Mharakurwa, S., Ndiaye, D., Rathod, P.K., Rosenthal, P.J., 2015. Antimalarial drug resistance: literature review and activities and findings of the IECMR network. *Am. Soc. Trop. Med. Hyg.* 93, 57–68. <https://doi.org/10.4269/ajtmh.15-0007>.
- de Heuvel, E., Kooistra, A.J., Edink, E., van Klaveren, S., Stuijt, J., van der Meer, T., Sadek, P., Mabilde, D., Caljon, G., Maes, L., Siderius, M., de Esch, I.J.P., Sterk, G.J., Leurs, R., 2021. Discovery of diaryl ether substituted tetrahydrophthalazinones as TbrPDEB1 inhibitors following structure-based virtual screening. *Front. Chem.*
- de Koning, H.P., 2020. The drugs of sleeping sickness: their mechanisms of action and resistance, and a brief history. *Trop. Med. Infect. Dis.* 5, 14. <https://doi.org/10.3390/tropicalmed5010014>.
- de Koning, H.P., Gould, M.K., Sterk, G.J., Tenor, H., Kunz, S., Lugnbuehl, E., Seebeck, T., 2012. Pharmacological validation of trypanosoma brucei phosphodiesterases as novel drug targets. *J. Infect. Dis.* 206, 229–237. <https://doi.org/10.1093/infdis/jir857>.
- El Sayed, I., van der Veken, P., Steert, K., Dhooghe, L., Hostyn, S., van Baelen, G., Lemièrre, G., Maes, B.U.W., Cos, P., Maes, L., Joossens, J., Haemers, A., Pieters, L., Augustyns, K., 2009. Synthesis and antiparasitic activity of aminoalkylamino-substituted neocryptolepine derivatives. *J. Med. Chem.* 52, 2979–2988. <https://doi.org/10.1021/jm801490z>.
- Garrido, A., Vera, G., Delaye, P.O., Enguehard-Gueffier, C., 2021. Imidazo[1,2-*b*]pyridazine as privileged scaffold in medicinal chemistry: an extensive review. *Eur. J. Med. Chem.* 226, 113867 <https://doi.org/10.1016/J.EJMECH.2021.113867>.
- Gupta, S., Yardley, V., Vishwakarma, P., Shivahare, R., Sharma, B., Launay, D., Martin, D., Puri, S.K., 2015. Nitroimidazo-oxazole compound DNDI-VL-2098: an orally effective preclinical drug candidate for the treatment of visceral leishmaniasis. *J. Antimicrob. Chemother.* 70, 518–527. <https://doi.org/10.1093/jac/dku422>.
- Herricks, J.R., Hotez, P.J., Wanga, V., Coffeng, L.E., Haagsma, J.A., Basáñez, M.-G., Buckle, G., Budke, C.M., Carabin, H., Fèvre, E.M., Fürst, T., Halasa, Y.A., King, C.H., Murdoch, M.E., Ramaiah, K.D., Shepard, D.S., Stolk, W.A., Undurraga, E.A., Stanaway, J.D., Naghavi, M., Murray, C.J.L., 2017. The global burden of disease study 2013: what does it mean for the NTDs? *PLoS Neglected Trop. Dis.* 11, 1–21. <https://doi.org/10.1371/journal.pntd.0005424>.
- Houslay, M.D., Schafer, P., Zhang, K.Y.J., 2005. Keynote review: phosphodiesterase-4 as a therapeutic target. *Drug Discov. Today* 10, 1503–1519. [https://doi.org/10.1016/S1359-6446\(05\)03622-6](https://doi.org/10.1016/S1359-6446(05)03622-6).
- Howard, B.L., Harvey, K.L., Stewart, R.J., Azevedo, M.F., Crabb, B.S., Jennings, I.G., Sanders, P.R., Manallack, D.T., Thompson, P.E., Tonkin, C.J., Gilson, P.R., 2015. Identification of potent phosphodiesterase inhibitors that demonstrate cyclic nucleotide-dependent functions in apicomplexan parasites. *ACS Chem. Biol.* 10, 1145–1154. <https://doi.org/10.1021/cb501004q>.
- Kunz, S., Balmer, V., Sterk, G.J., Pollastri, M.P., Leurs, R., Müller, N., Hemphill, A., Spycher, C., 2017a. The single cyclic nucleotide-specific phosphodiesterase of the intestinal parasite *Giardia lamblia* represents a potential drug target. *PLoS Neglected Trop. Dis.* 11, 1–23. <https://doi.org/10.1371/journal.pntd.0005891>.
- Kunz, S., Balmer, V., Sterk, G.J., Pollastri, M.P., Leurs, R., Müller, N., Hemphill, A., Spycher, C., 2017b. The single cyclic nucleotide-specific phosphodiesterase of the



- intestinal parasite *Giardia lamblia* represents a potential drug target. *PLoS Neglected Trop. Dis.* 11, e0005891 <https://doi.org/10.1371/journal.pntd.0005891>.
- Kunz, S., Oberholzer, M., Seebeck, T., 2005. A FYVE-containing unusual cyclic nucleotide phosphodiesterase from *Trypanosoma cruzi*. *FEBS J.* 272, 6412–6422. <https://doi.org/10.1111/j.1742-4658.2005.05039.x>.
- Lipinski, C.A., Lombardo, F., Dominy, B.W., Feeney, P.J., 1997. Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings. *Adv. Drug Deliv. Rev.* 23, 3–25. [https://doi.org/10.1016/S0169-409X\(96\)00423-1](https://doi.org/10.1016/S0169-409X(96)00423-1).
- Manallack, D.T., Hughes, R.A., Thompson, P.E., 2005. The next generation of phosphodiesterase inhibitors: structural clues to ligand and substrate selectivity of phosphodiesterases. *J. Med. Chem.* 48, 3449–3462. <https://doi.org/10.1021/jm040217u>.
- Maurice, D.H., Ke, H., Ahmad, F., Wang, Y., Chung, J., Manganiello, V.C., 2014. Advances in targeting cyclic nucleotide phosphodiesterases. *Nat. Rev. Drug Discov.* 13, 290–314. <https://doi.org/10.1038/nrd4228>.
- McKerrow, J.H., Lipinski, C.A., 2017. The rule of five should not impede anti-parasitic drug development. *Int. J. Parasitol.: Drugs Drug Resist.* 7, 248–249. <https://doi.org/10.1016/j.ijpddr.2017.05.003>.
- Mørch, K., Hanevik, K., 2020. Giardiasis treatment: an update with a focus on refractory disease. *Curr. Opin. Infect. Dis.* 33.
- Müller, J., Braga, S., Uldry, A.-C., Heller, M., Müller, N., 2020. Comparative proteomics of three *Giardia lamblia* strains: investigation of antigenic variation in the post-genomic era. *Parasitology* 147, 1008–1018. <https://doi.org/10.1017/S0031182020000657>.
- Müller, J., Hemphill, A., 2013. New approaches for the identification of drug targets in Protozoan parasites. In: *International Review of Cell and Molecular Biology*. Academic Press, pp. 359–401. <https://doi.org/10.1016/B978-0-12-407704-1.00007-5>.
- Müller, J., Müller, N., 2019. Nitroreductases of bacterial origin in *Giardia lamblia*: potential role in detoxification of xenobiotics. *Microbiologyopen* 8, e904. <https://doi.org/10.1002/mbo3.904>.
- Nabarro, L.E.B., Lever, R.A., Armstrong, M., Chiodini, P.L., 2015. Increased incidence of nitroimidazole-refractory giardiasis at the hospital for tropical diseases, London: 2008–2013. *Clin. Microbiol. Infect.* 21, 791–796. <https://doi.org/10.1016/j.cmi.2015.04.019>.
- Oberholzer, M., Marti, G., Baresic, M., Kunz, S., Hemphill, A., Seebeck, T., 2007. The *Trypanosoma brucei* cAMP phosphodiesterases TbrPDEB1 and TbrPDEB2: flagellar enzymes that are essential for parasite virulence. *Faseb. J.* 21, 720–731. <https://doi.org/10.1096/fj.06-6818com>.
- Oliveira, R., Miranda, D., Magalhães, J., Capela, R., Perry, M.J., O'Neill, P.M., Moreira, R., Lopes, F., 2015. From hybrid compounds to targeted drug delivery in antimalarial therapy. *Bioorg. Med. Chem.* 23, 5120–5130. <https://doi.org/10.1016/j.BMC.2015.04.017>.
- Patterson, S., Wyllie, S., 2014. Nitro drugs for the treatment of trypanosomatid diseases: past, present, and future prospects. *Trends Parasitol.* 30, 289–298. <https://doi.org/10.1016/j.PT.2014.04.003>.
- Ribeiro, V., Dias, N., Paiva, T., Hagström-Bex, L., Nitz, N., Pratesi, R., Hecht, M., 2020. Current trends in the pharmacological management of Chagas disease. *Int. J. Parasitol.: Drugs Drug Resist.* 12, 7–17. <https://doi.org/10.1016/j.ijpddr.2019.11.004>.
- Sharon, K.-K., Minyong, L., Alyssa, S., Shilong, Z., Gurpreet, K., Xiaochuan, Y., Binghe, W., Roberto, D., 2010. Chemical validation of phosphodiesterase C as a chemotherapeutic target in *trypanosoma cruzi*, the etiological agent of Chagas' disease. *Antimicrob. Agents Chemother.* 54, 3738–3745. <https://doi.org/10.1128/AAC.00313-10>.
- Souness, J.E., Aldous, D., Sargent, C., 2000. Immunosuppressive and anti-inflammatory effects of cyclic AMP phosphodiesterase (PDE) type 4 inhibitors. *Immunopharmacology* 47, 127–162. [https://doi.org/10.1016/S0162-3109\(00\)00185-5](https://doi.org/10.1016/S0162-3109(00)00185-5).
- Sridhar, P., Alagumuthu, M., Ram, B., Arumugam, S., Reddy, S.R., 2017. Drugs against neurodegenerative diseases: design and synthesis of 6-Amino-substituted imidazo [1,2-b]pyridazines as acetylcholinesterase inhibitors. *ChemistrySelect* 2, 842–847. <https://doi.org/10.1002/slct.201601353>.
- Sterk, G.J., Hatzelmann, A., Barsig, J., Marx, D., Kley, H.-P., Christiaans, J.A.M., Menge, W.M.P.B., 2004a. Piperidine-pyridazones and Phthalazones as PDE4 Inhibitors. *PCT WO2004017974A1*.
- Sterk, G.J., Hatzelmann, A., Barsig, J., Marx, D., Kley, H.-P., Christiaans, J.A.M., Menge, W.M.P.B., 2004b. Pyridazinone-derivatives as PDE4 Inhibitors. *PCT WO2004018451A1*.
- Tenor, H., Hatzelmann, A., Beume, R., Lahu, G., Zech, K., Bethke, T.D., 2011. Pharmacology, Clinical Efficacy, and Tolerability of Phosphodiesterase-4 Inhibitors: Impact of Human Pharmacokinetics, Phosphodiesterases as Drug Targets. *Springer*. <https://doi.org/10.1007/978-3-642-17969-3.3>.
- Tomcufcik, A.S., Izzo, P.T., Fabio, P.F., 1974. 6-substituted 3-Nitroimidazo(1,2-B)pyridazines and Method of Preparing Same, US3828041A.
- van Baelen, G., Meyers, C., Lemièrre, G.L.F., Hostyn, S., Dommissie, R., Maes, L., Augustyns, K., Haemers, A., Pieters, L., Maes, B.U.W., 2008. Synthesis of 6-methyl-6H-indolo[3,2-c]isoquinoline and 6-methyl-6H-indolo[2,3-c]isoquinoline: two new unnatural isoquinoline isomers of the cryptolepine series. *Tetrahedron* 64, 11802–11809. <https://doi.org/10.1016/J.TET.2008.08.116>.
- van der Mey, M., Boss, H., Couwenberg, D., Hatzelmann, A., Sterk, G.J., Goubitz, K., Schenk, H., Timmerman, H., 2002. Novel selective phosphodiesterase (PDE4) inhibitors. 4. Resolution, absolute configuration, and PDE4 inhibitory activity of cis-tetra- and cis-hexahydrophthalazinones. *J. Med. Chem.* 45, 2523–2535. <https://doi.org/10.1021/jm0110338>.
- van der Mey, M., Hatzelmann, A., van der Laan, I.J., Sterk, G.J., Thibaut, U., Timmerman, H., 2001. Novel selective PDE4 inhibitors. 1. Synthesis, Structure–Activity relationships, and molecular modeling of 4-(3,4-dimethoxyphenyl)-2H-phthalazin-1-ones and analogues. *J. Med. Chem.* 44, 2511–2522. <https://doi.org/10.1021/jm010837k>.
- Vanaerschoot, M., Huijben, S., van den Broeck, F., Dujardin, J.-C., 2014. Drug resistance in vectorborne parasites: multiple actors and scenarios for an evolutionary arms race. *FEMS (Fed. Eur. Microbiol. Soc.) Microbiol. Rev.* 38, 41–55. <https://doi.org/10.1111/1574-6976.12032>.
- Veerman, J., van den Bergh, T., Orrling, K.M., Jansen, C., Cos, P., Maes, L., Chatelain, E., Ioset, J.-R., Edink, E.E., Tenor, H., Seebeck, T., de Esch, I., Leurs, R., Sterk, G.J., 2016. Synthesis and evaluation of analogs of the phenylpyridazinone NPD-001 as potent trypanosomal TbrPDEB1 phosphodiesterase inhibitors and in vitro trypanocidal. *Bioorg. Med. Chem.* 24, 1573–1581. <https://doi.org/10.1016/j.bmc.2016.02.032>.
- Venkatraj, M., Ariën, K.K., Heeres, J., Joossens, J., Dirie, B., Lyssens, S., Michiels, J., Cos, P., Lewi, P.J., Vanham, G., Maes, L., van der Veken, P., Augustyns, K., 2014. From human immunodeficiency virus non-nucleoside reverse transcriptase inhibitors to potent and selective antitrypanosomal compounds. *Bioorg. Med. Chem.* 22, 5241–5248. <https://doi.org/10.1016/J.BMC.2014.08.005>.
- World Health Organisation, 2019. No Time to Wait: Securing the Future from Drug-Resistant Infections. World Health Organisation, Geneva.