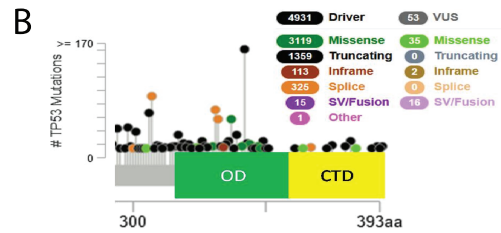
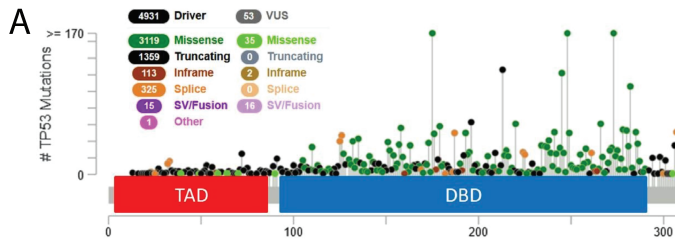


Supplementary Information

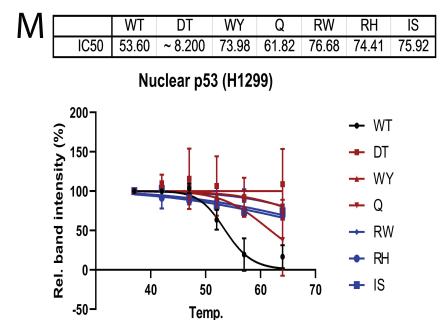
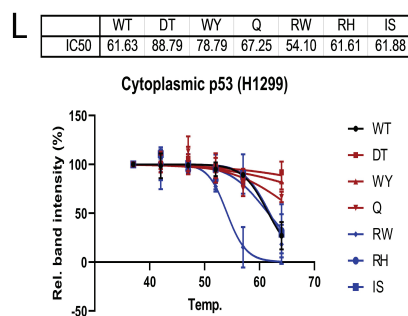
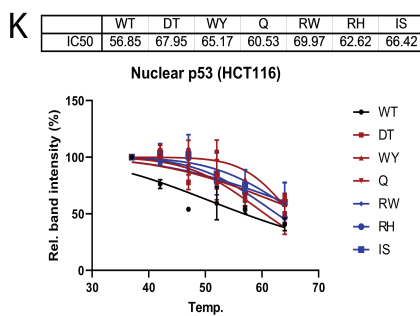
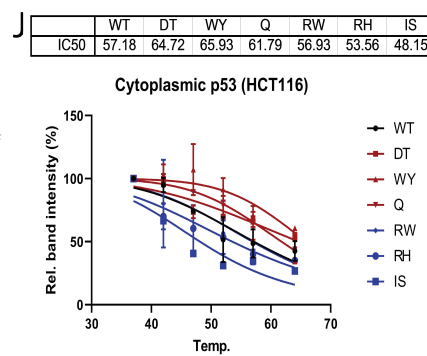
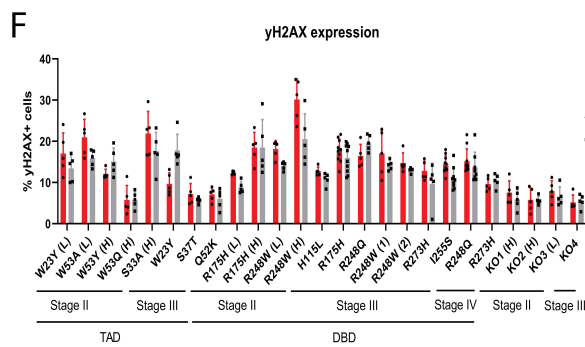
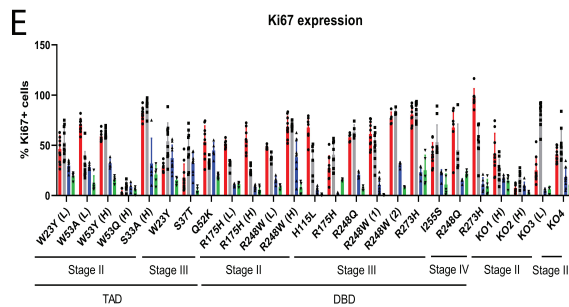
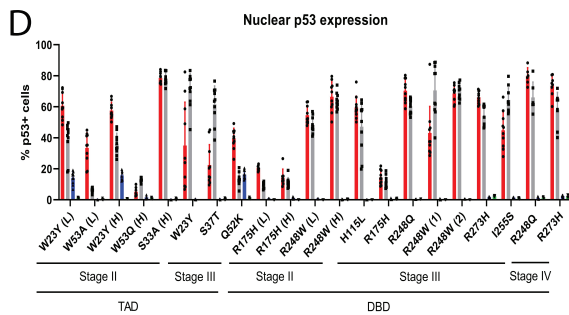
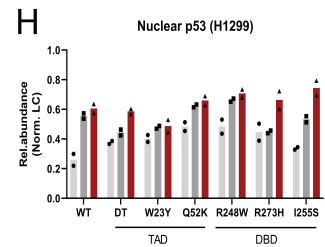
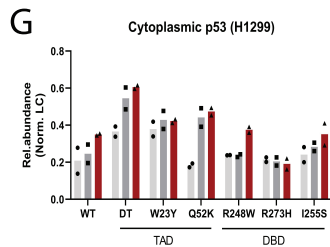
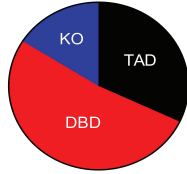
Domain-specific p53 mutants activate EGFR by distinct mechanisms exposing tissue-independent therapeutic vulnerabilities

Teresa Lai Fong Ho^{1,2}, May Yin Lee³, Hui Chin Goh², Germaine Yi Ning Ng⁴, Jane Jia Hui Lee³, Srinivasaraghavan Kannan⁵, Yan Ting Lim^{6,7}, Tianyun Zhao^{6,7}, Edwin Kok Hao Lim³, Cheryl Zi Jin Phua³, Yi Fei Lee³, Rebecca Yi Xuan Lim³, Perry Jun Hao Ng³, Ju Yuan³, Dedrick Kok Hong Chan^{8,9}, Bettina Lieske^{9,10}, Choon Seng Chong^{9,10}, Kuok Chung Lee⁹, Jeffrey Lum¹¹, Wai Kit Cheong⁹, Khay Guan Yeoh¹², Ker Kan Tan^{9,10}, Radoslaw M. Sobota^{6,7}, Chandra S. Verma^{5,13,14}, David P. Lane¹, Wai Leong Tam^{2,3,13,15,16} and Ashok R. Venkitaraman^{1,2,16,*}.



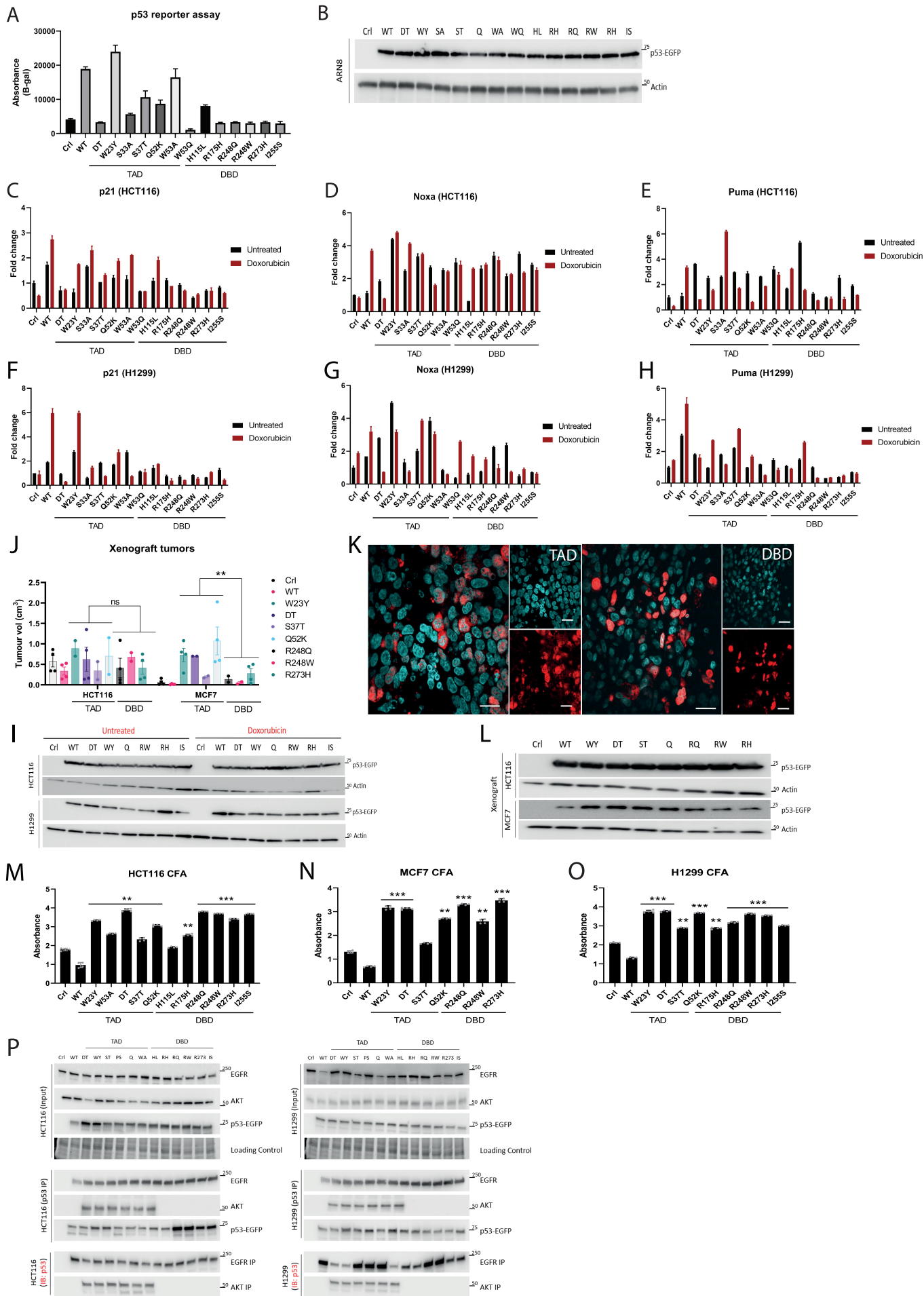
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| TAD | DBD |
|---------------------------|-------|
| W23Y | H115L |
| S33A | R175H |
| S37T | R248Q |
| Q52K | R248W |
| W53A | R273H |
| W53Q | I255S |
| DT (LW22/23QS, WF53/54QS) | |



Supplementary Fig. 1: Phenotypic differences between TAD and DBD *TP53* mutant cells in vivo and in vitro

(A-B) Frequency of various types of *TP53* mutations as reported in pan-cancer clinical samples. Data from MSK-IMPACT Clinical Sequencing Cohort consisting of 10336 patients and 10945 samples. (C) Proportion of colorectal tumors expressing either TAD or DBD mutations or lacking p53 (KO) in our patient cohort. Table lists the unique TAD and DBD mutations identified. (D-F) Quantification of the percentage of cells with (D) nuclear p53, (E) Ki67 and (F) γ H2AX staining in TAD and DBD mutant colorectal tumors from 10 fields of view (Mean \pm SEM). (TC: tumor core; TIF: tumor invasive front) and surrounding normal tissue (2cm and 5cm from the tumor invasive front). (G-H) p53 protein turnover in H1299 (G) cytosolic and (H) nuclear cellular fractions following cycloheximide treatment. n=2. (I) Protein levels of p53-EGFP (cytosolic and nuclear cellular fractions) in untreated and cycloheximide treated HCT116 and H1299 expressing WT and mutant p53. n=2. (J-M) CETSA-derived curves of cytosolic and nuclear WT and mutant p53 in untreated HCT116 and H1299 cells. IC50 values represent temperature at which 50% of the protein has degraded. Cytosolic TAD (n=3 independent mutants) versus DBD (n=3 independent mutants) p<0.001. n=2. Two-tailed unpaired Student's t-test was performed. (* p<0.05, ** p<0.01, *** p<0.001, **** p<0.0001). Source data are provided as a Source Data file.



Upregulated in TAD



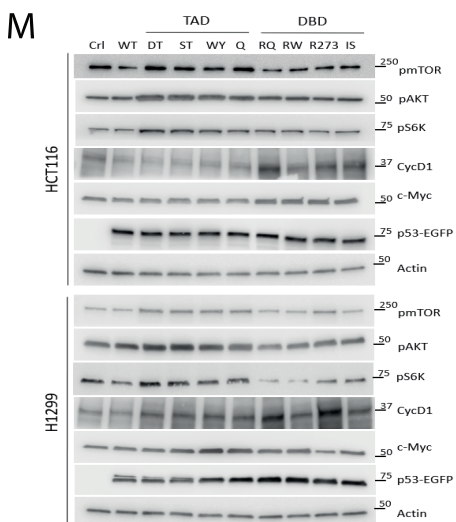
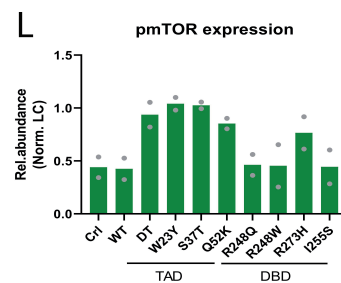
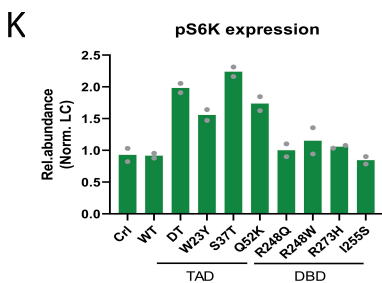
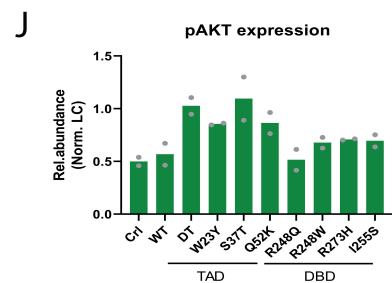
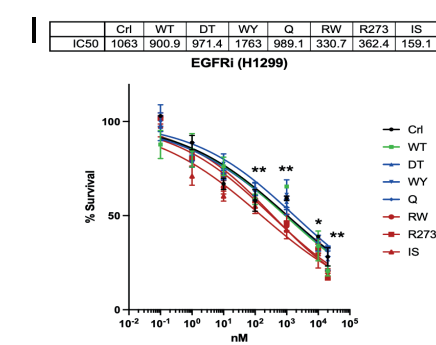
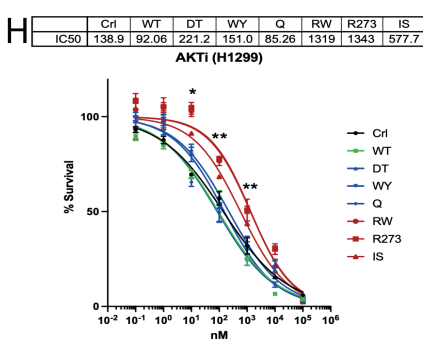
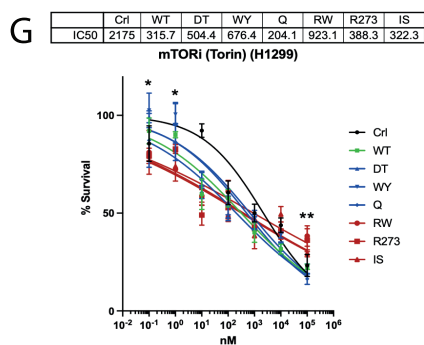
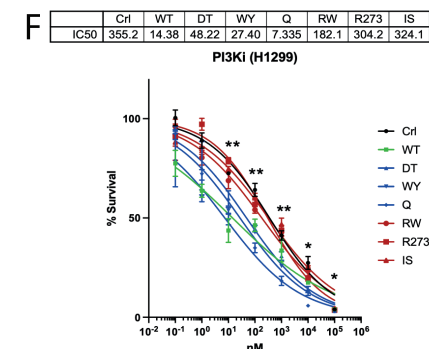
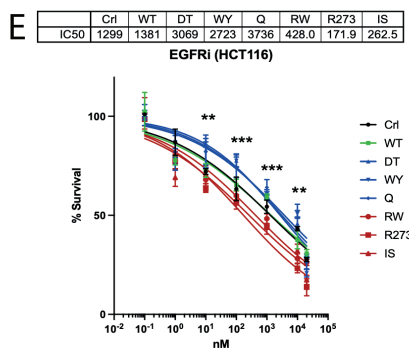
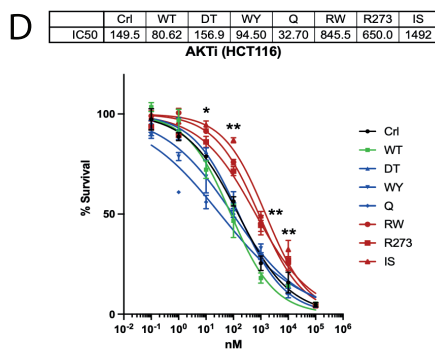
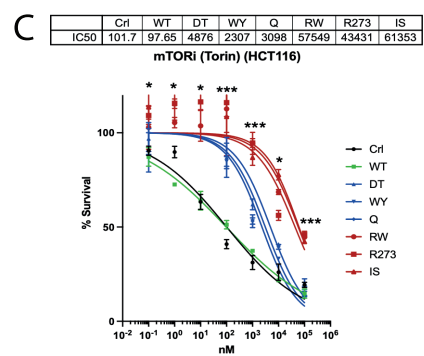
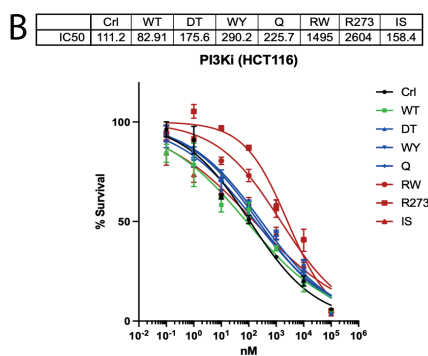
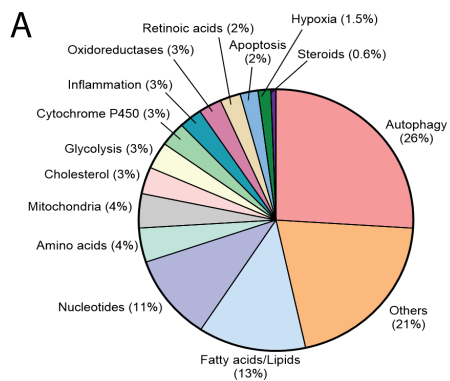
Homo sapiens ()

Upregulated in DBD



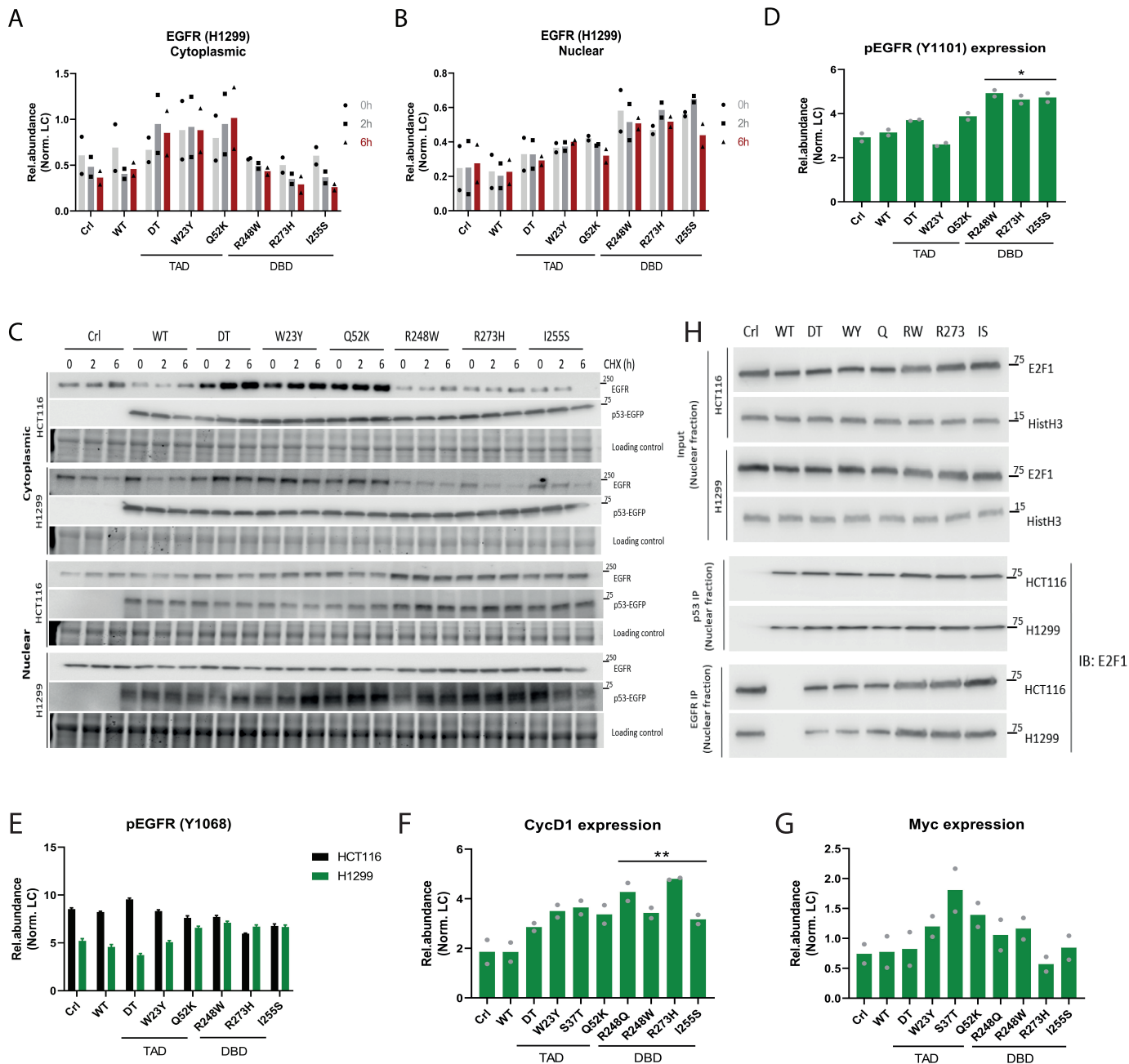
Supplementary Fig. 2: Phenotypic differences between TAD and DBD *TP53* mutant cells in vivo and in vitro

(A) Quantification of Beta-Galactosidase activity in p53 promoter assay with ARN8 cells expressing WT and mutant p53. n=12 independent replicates. (B) Protein levels of p53-EGFP in ARN8 cells transiently expressing WT and mutant p53. n=2. (C-H) Quantification of mRNA transcript levels of (C, F) *p21*, (D, G) *NOXA* and (E, H) *PUMA* in untreated and doxorubicin treated HCT116 and H1299 cells expressing WT and mutant p53. n=12 independent replicates. (I) Protein levels of p53-EGFP in untreated and doxorubicin treated HCT116 and H1299 cells expressing WT and mutant p53. n=2. (J) Quantification of tumor volume of xenograft tumors derived from HCT116 and MCF7 cells expressing WT and mutant p53. n=2 mice; n=4 tumors per mutant. Individual data points denote individual tumors. Statistical tests performed on TAD versus DBD. (K) Representative immunofluorescence staining of p53 (red) in DBD and TAD mutant xenograft tumor tissue. DAPI staining shows nuclei. n=2. Scale bar = 30μm. (L) Protein levels of p53-EGFP in lines derived from respective HCT116 and MCF7 xenograft tumors. n=1-2 mice, n=2-4 tumors per mutant. (M-O) Quantification of crystal violet intensity from colony formation assays using HCT116, MCF7 and H1299 cells constitutively expressing WT and mutant p53. Statistical tests performed on TAD/DBD (n=3 independent mutants each) versus Control. n=4 independent experiments. (P) Protein levels of EGFR, AKT and p53-EGFP in input fractions from HCT116 and H1299 cells expressing WT and mutant p53. P53 IP and reverse IP validation of EGFR and AKT. n=2. (Q-R) Reactome plots of protein interactors enriched in (Q) TAD- and (R) DBD- mutant expressing HCT116 cells respectively indicating the molecular and functional pathways that they are involved in. Two-tailed unpaired Student's t-test was performed. (* p<0.05, ** p<0.01, *** p<0.001, **** p<0.0001). Source data are provided as a Source Data file.



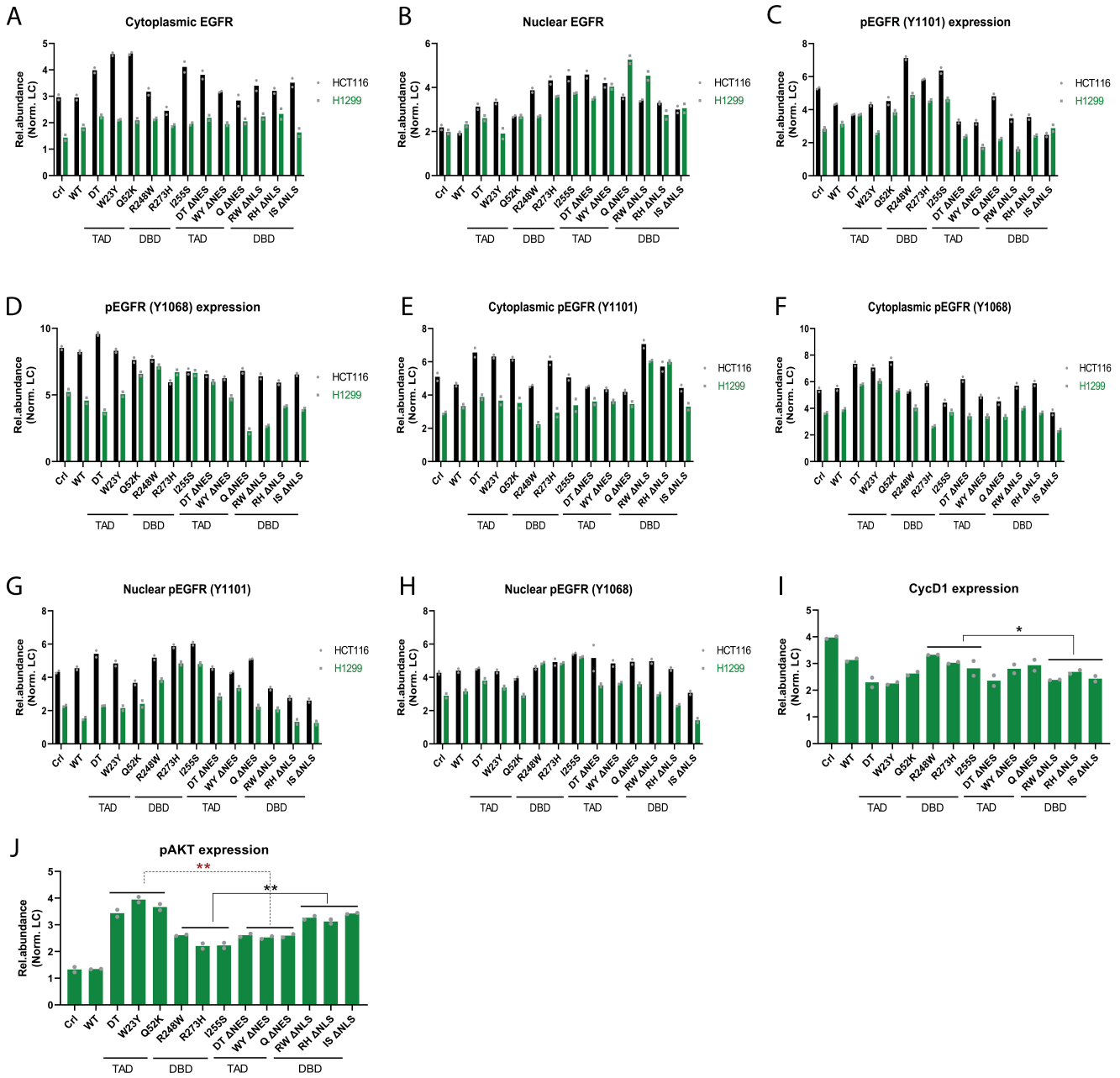
Supplementary Fig. 3: Differential response to small molecule inhibitors

(A) Summary of other signaling and metabolic pathways and/or molecular functions that TAD and DBD mutant cells are differentially dependent on based on sensitivity to small molecular inhibitor treatment. (B-I) Dose response curves of HCT116 and H1299 cells expressing WT and mutant p53 treated with inhibitors targeting (B, F) PI3K, (C, G) mTOR, (D, H) AKT and (E, I) EGFR. n=3 independent experiments. (J-L) Quantification of protein levels of (J) pAKT, (K) pS6K and (L) pmTOR in H1299 cells expressing WT and mutant p53. n=2. (M) Protein levels of pmTOR, pAKT, pS6K, pERK, Cyclin D1 and c-Myc in HCT116 and H1299 expressing WT and mutant p53. n=2. Two-tailed unpaired Student's t-test was performed. (* $p<0.05$, ** $p<0.01$, *** $p<0.001$, **** $p<0.0001$). Source data are provided as a Source Data file.



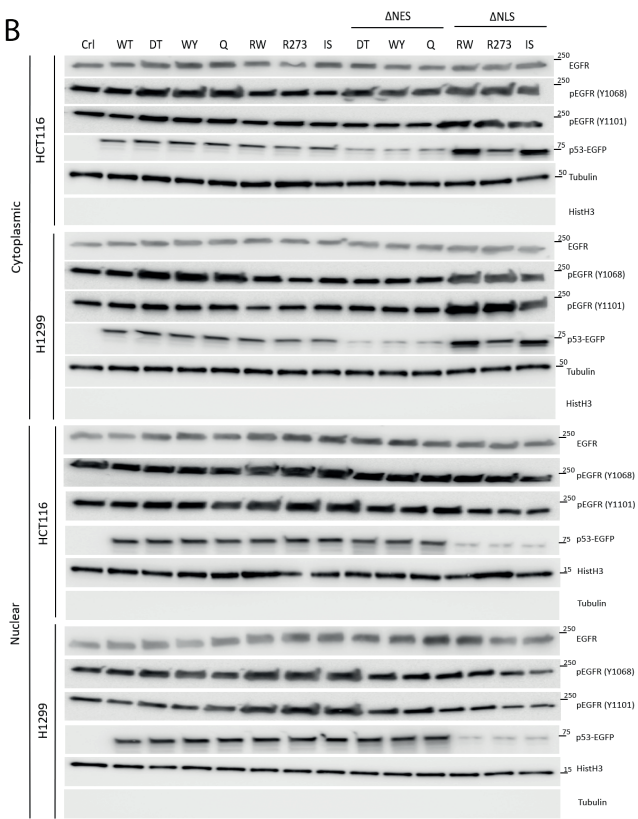
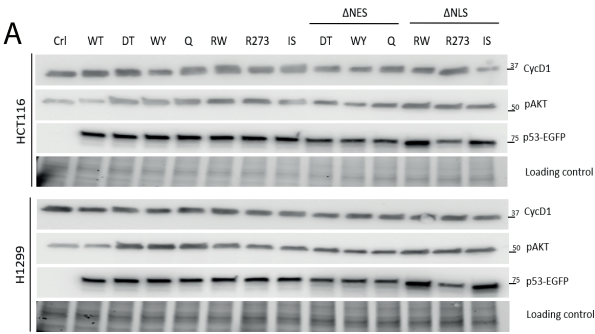
Supplementary Fig. 4: Comparison of EGFR stability and signaling in TAD and DBD mutant cells

(A-B) Quantification of EGFR protein turnover in H1299 (A) cytosolic and (B) nuclear cellular fractions following treatment with cycloheximide. n=2. (C) Protein levels of cytosolic and nuclear EGFR in HCT116 and H1299 cells expressing WT and mutant p53 treated with cycloheximide. n=2. (D) Quantification of pEGFR (Y1101) protein levels in H1299 cells expressing WT and mutant p53. Statistical tests performed on TAD (n=3 independent mutants) versus DBD (n=3 independent mutants). n=2. (E) Quantification of pEGFR (Y1068) protein levels in HCT116 and H1299 cells expressing WT and mutant p53. n=2. (F-G) Quantification of (F) Cyclin D1 and (G) c-Myc protein levels in H1299 cells expressing WT and mutant p53. Statistical tests performed on TAD (n=3 independent mutants) versus DBD (n=3 independent mutants). n=2. (H) Protein levels of nuclear E2F1 in input fractions from HCT116 and H1299 cells expressing WT and mutant p53. P53 and EGFR IP of nuclear fractions. n=2. Two-tailed unpaired Student's t-test was performed. (* p<0.05, ** p<0.01, *** p<0.001, **** p<0.0001). Source data are provided as a Source Data file.



Supplementary Fig. 5: Differential stabilization of EGFR in the cytosol and nucleus in TAD and DBD mutant cells

(A-D) Quantification of (A) cytosolic EGFR, (B) nuclear EGFR, (C) pEGFR (Y1101) and (D) pEGFR (Y1068) protein levels in HCT116 and H1299 cells expressing WT and mutant p53. n=2. (E-H) Quantification of (E, F) cytosolic and (G, H) nuclear pEGFR protein levels in HCT116 and H1299 cells expressing WT and mutant p53. n=2. (I-J) Quantification of (I) Cyclin D1 and (J) pAKT protein levels in HCT116 cells expressing WT and mutant p53. Statistical tests performed on TAD (n=3 independent mutants) versus DBD (n=3 independent mutants). n=2. Two-tailed unpaired Student's t-test was performed. (* p<0.05, ** p<0.01, *** p<0.001, **** p<0.0001). Source data are provided as a Source Data file.

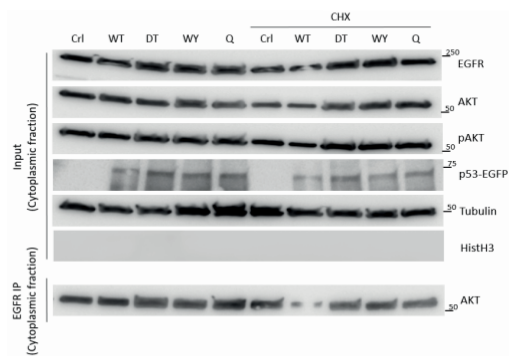


Supplementary Fig. 6: EGFR activation in DBD nuclear localization and TAD nuclear export defective mutants

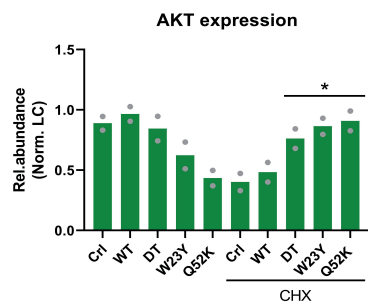
(A) Protein levels of various factors in HCT116 and H1299 cells expressing WT and mutant p53. n=2.

(B) Protein levels of various factors in cytosolic and nuclear fractions of HCT116 and H1299 cells expressing WT and mutant p53. n=2. Source data are provided as a Source Data file.

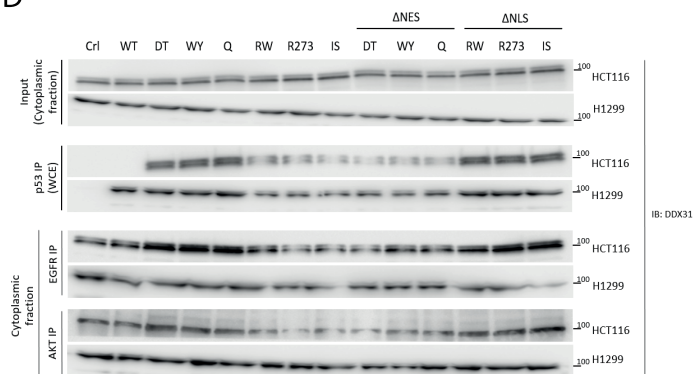
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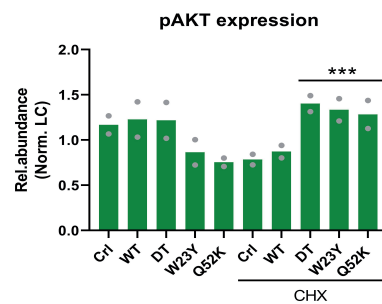
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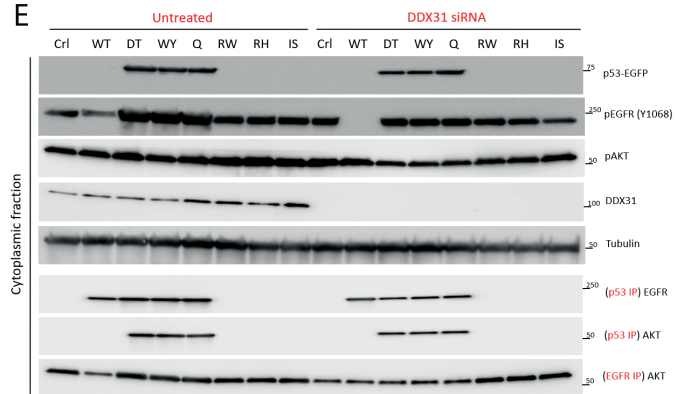
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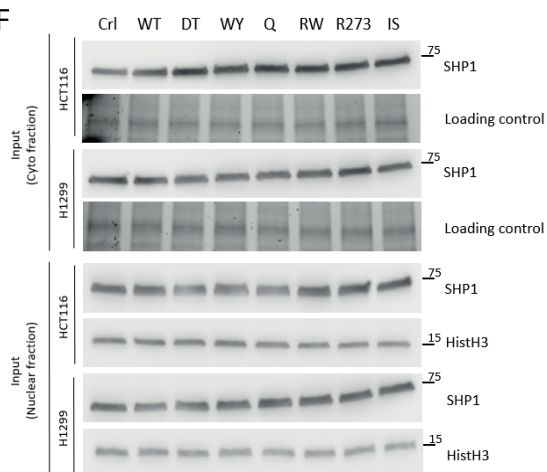
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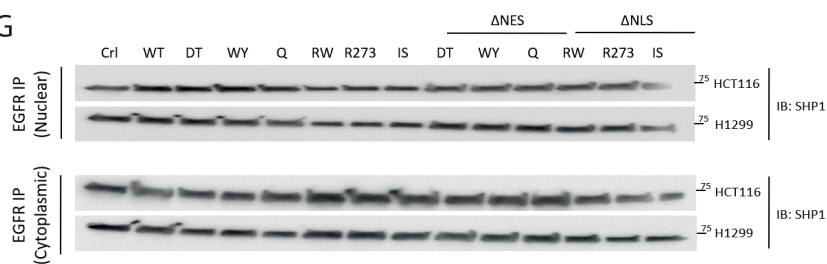
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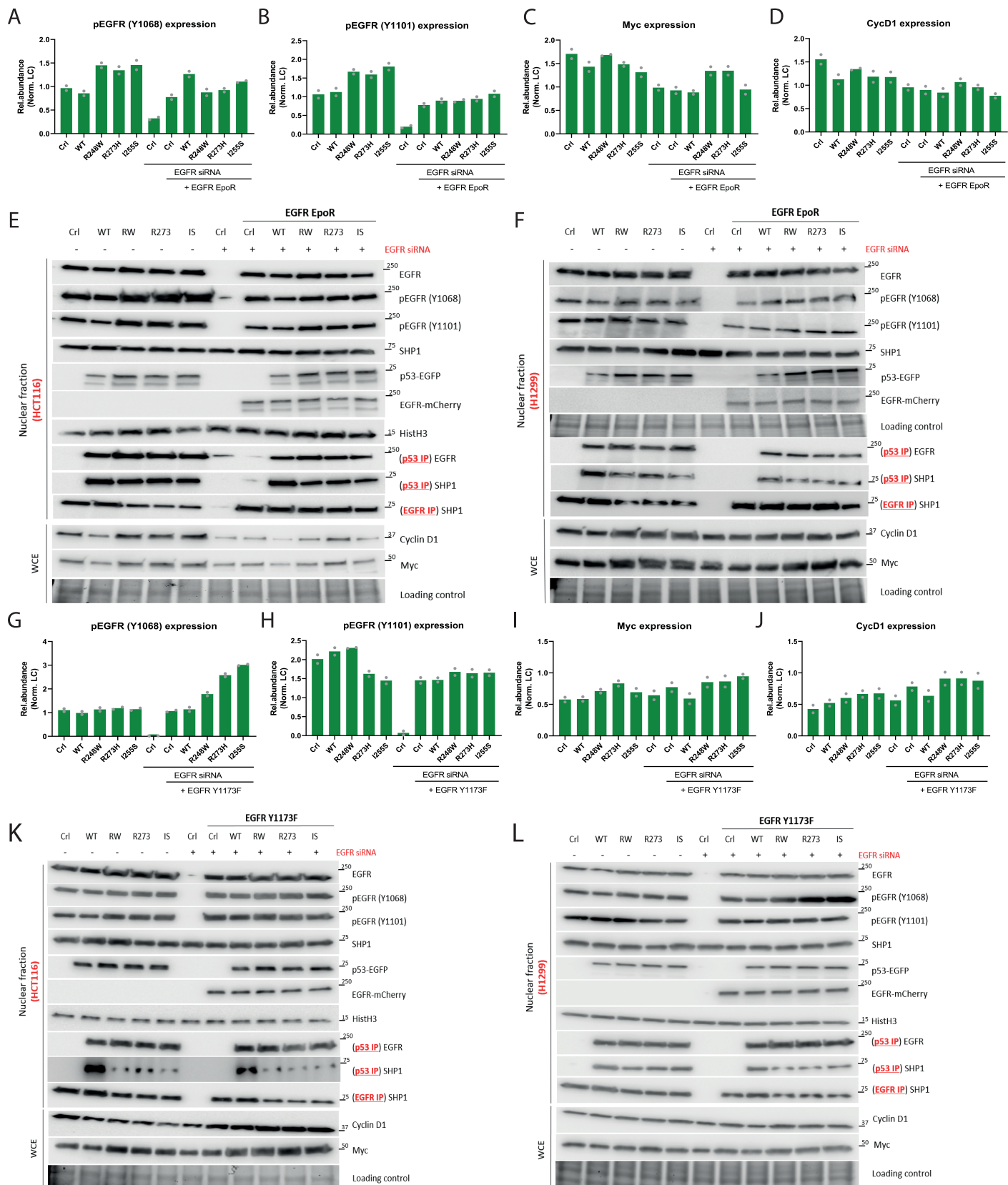


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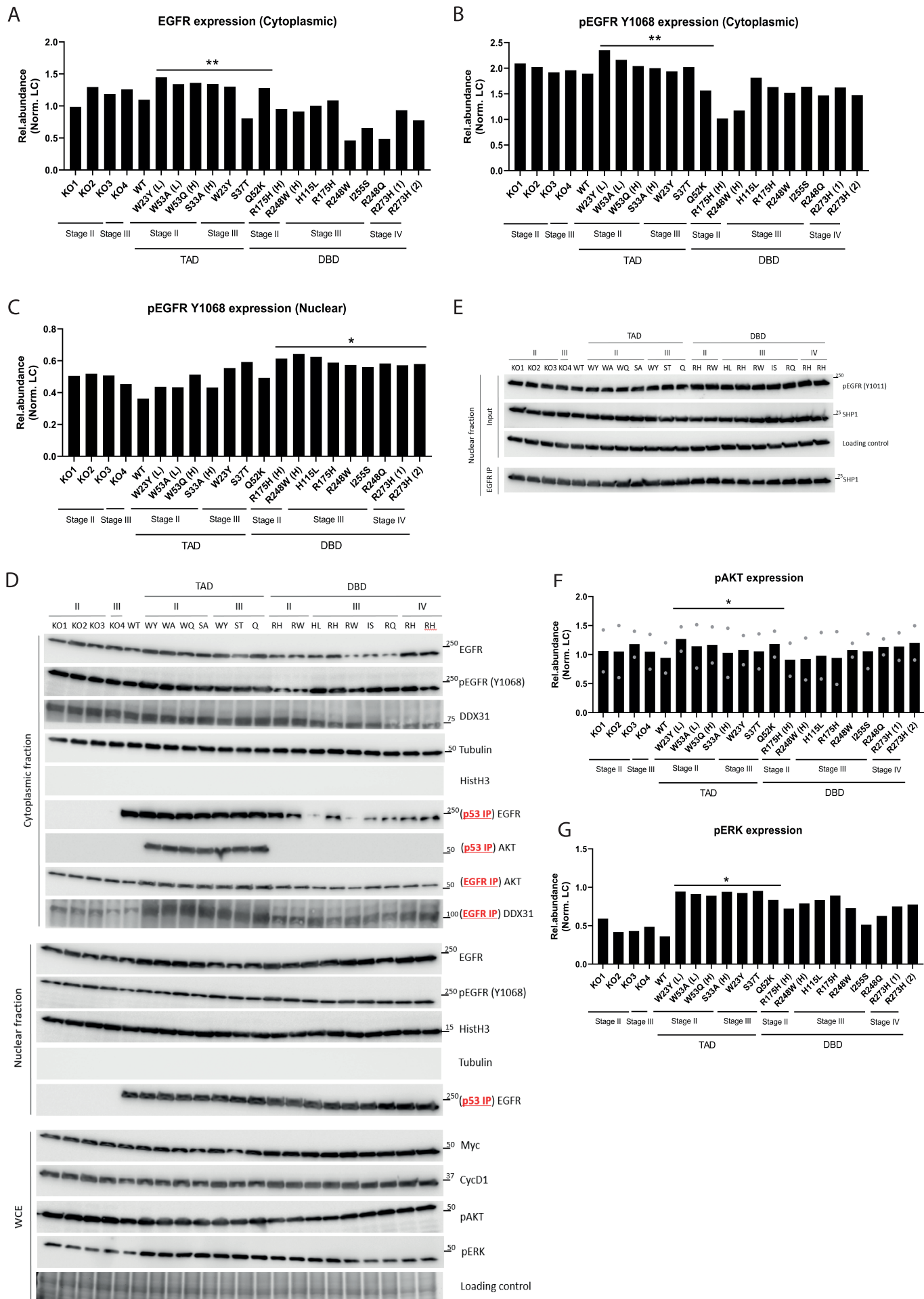
Supplementary Fig. 7: Stabilization of AKT in TAD mutants and disruption of SHP1-EGFR interaction in DBD mutants

(A) Protein levels of EGFR, AKT and pAKT in input cytosolic fractions from H1299 cells expressing TAD mutants under untreated and cycloheximide-treated conditions. Protein levels of AKT pulled down with EGFR. n=2. (B-C) Quantification of (B) AKT and (C) pAKT protein levels in H1299 cells expressing TAD mutants under untreated and cycloheximide-treated conditions. Statistical tests performed on TAD (n=3 independent mutants) versus TAD+CHX (n=3 independent mutants). n=2. (D) Protein levels of DDX31 in input cytosolic fractions from HCT116 and H1299 cells expressing WT and mutant p53. Protein levels of DDX31 pulled down with p53, EGFR and AKT respectively. n=2. (E) Protein levels of various proteins (input and after pulldown with p53 or EGFR) in HCT116 cells expressing WT and mutant p53 under untreated and DDX31 siRNA treated conditions. n=2. (F) Protein levels of SHP1 in cytosolic and nuclear fractions from HCT116 and H1299 cells expressing WT and mutant p53. n=2. (G) Protein levels of SHP1 pulled down with EGFR in cytosolic and nuclear fractions from HCT116 and H1299 cells expressing WT and mutant p53. n=2. Two-tailed unpaired Student's t-test was performed. (* p<0.05, ** p<0.01, *** p<0.001, **** p<0.0001). Source data are provided as a Source Data file.



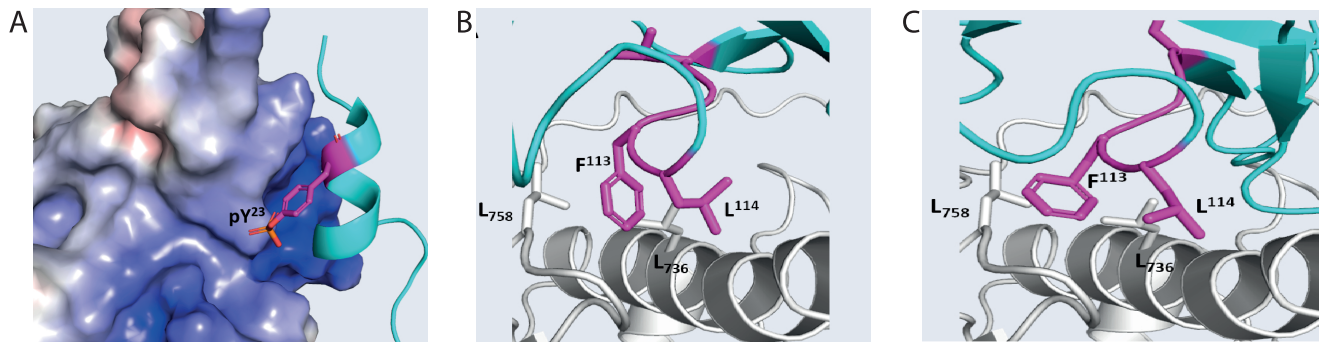
Supplementary Fig. 8: EGFR mutants validate DBD mutant effects on EGFR-SHP1 interaction

(A-D) Quantification of (A, B) pEGFR, (C) c-Myc and (D) Cyclin D1 protein levels in H1299 cells expressing DBD mutants. Cells were also treated with EGFR siRNA and expressed EGFR EpoR (high affinity for SHP1). n=2. (E-F) Protein levels of EGFR, pEGFR and SHP1 in input nuclear fractions and Cyclin D1 and c-Myc in whole-cell extract from HCT116 and H1299 cells expressing DBD mutants. Cells were also treated with EGFR siRNA and expressed EGFR EpoR (high affinity for SHP1). Protein levels of EGFR and SHP1 pulled down with p53 and/or EGFR. n=2. (G-J) Quantification of (G, H) pEGFR, (I) c-Myc and (J) Cyclin D1 protein levels in H1299 cells expressing DBD mutants. Cells were also treated with EGFR siRNA and expressed EGFR Y1173F (reduced affinity for SHP1). n=2. (K-L) Protein levels of EGFR, pEGFR and SHP1 in input nuclear fractions and Cyclin D1 and c-Myc in whole-cell extract from HCT116 and H1299 cells expressing DBD mutants. Cells were also treated with EGFR siRNA and expressed EGFR Y1173F (reduced affinity for SHP1). Protein levels of EGFR and SHP1 pulled down with p53 and/or EGFR. n=2. Source data are provided as a Source Data file.

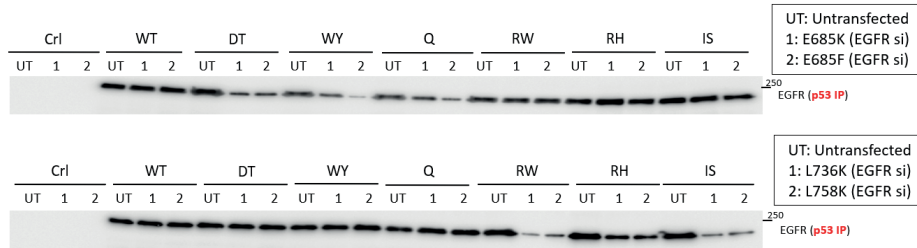


Supplementary Fig. 9: Mechanistic validation of mutant p53 activation of EGFR signaling in patient tumors

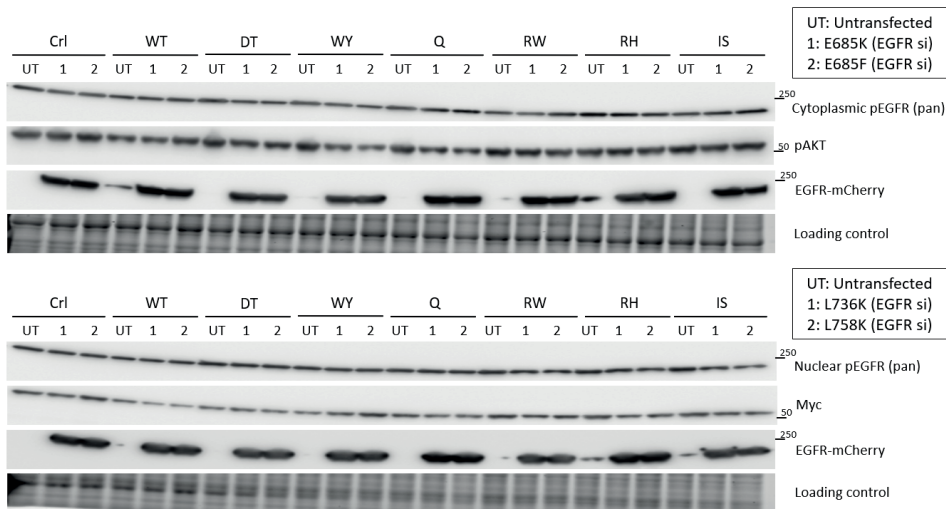
(A-C) Quantification of (A) cytosolic EGFR and (B) cytosolic and (C) nuclear pEGFR (Y1068) protein levels in p53 null and TAD and DBD mutant patient colorectal tumors. (D) Protein levels of various factors in cytosolic and nuclear fractions from p53 null and TAD and DBD mutant patient colorectal tumors. HCT116 cells expressing WT p53 as a control. Protein levels of EGFR, AKT and DDX31 pulled down with p53 or EGFR. (E) Protein levels of pEGFR (Y1011) and SHP1 in nuclear fractions from p53 null and DBD mutant patient colorectal tumors. HCT116 cells expressing WT p53 as a control. Protein levels of SHP1 pulled down with EGFR. (F-G) Quantification of (F) pAKT and (G) pERK protein levels in p53 null and TAD and DBD mutant patient colorectal tumors. HCT116 cells expressing WT p53 as a control. All statistical tests performed on TAD (n=7 independent patients) versus DBD (n=9 independent patients). Two-tailed unpaired Student's t-test was performed. (* $p<0.05$, ** $p<0.01$, *** $p<0.001$, **** $p<0.0001$). Source data are provided as a Source Data file.



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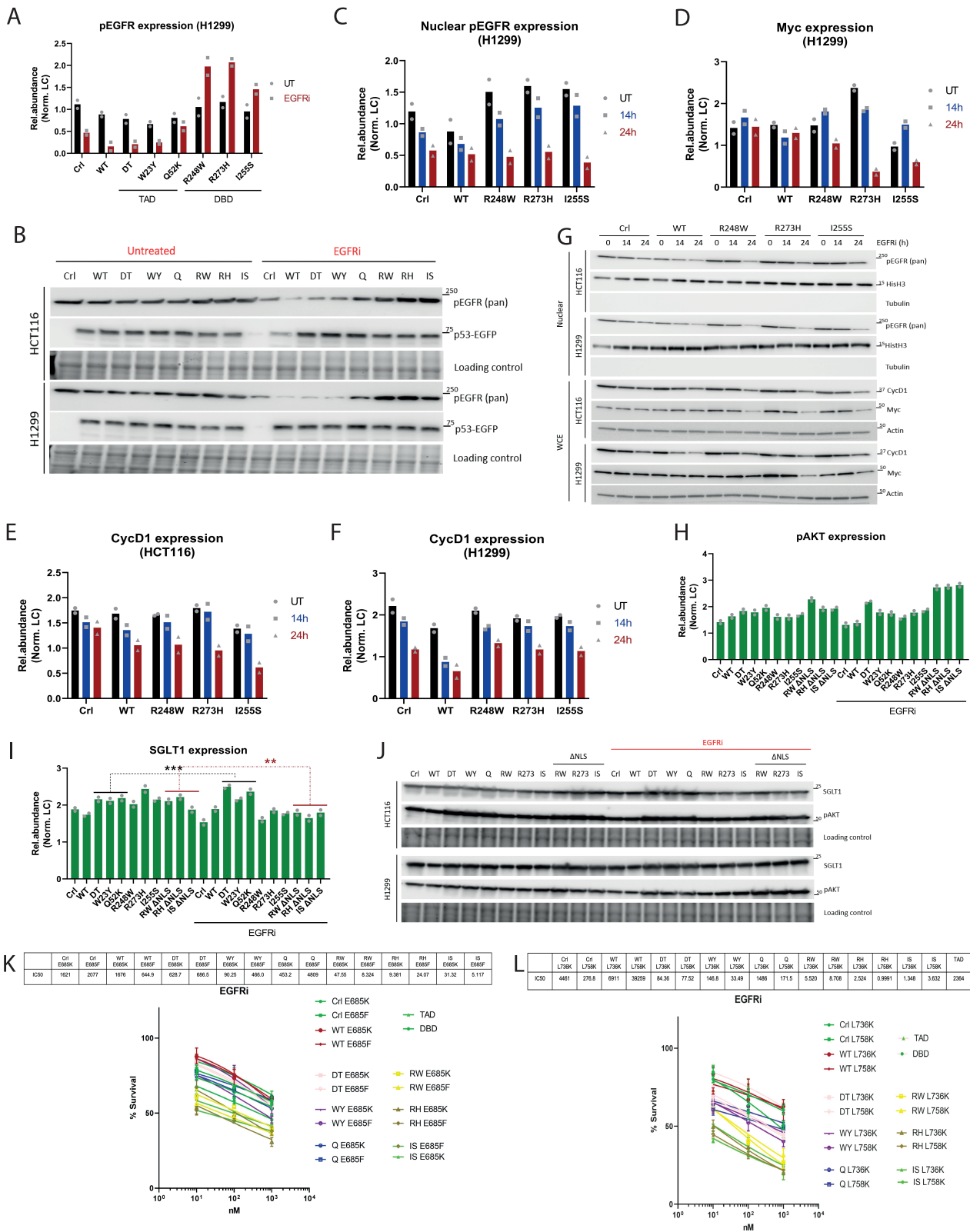


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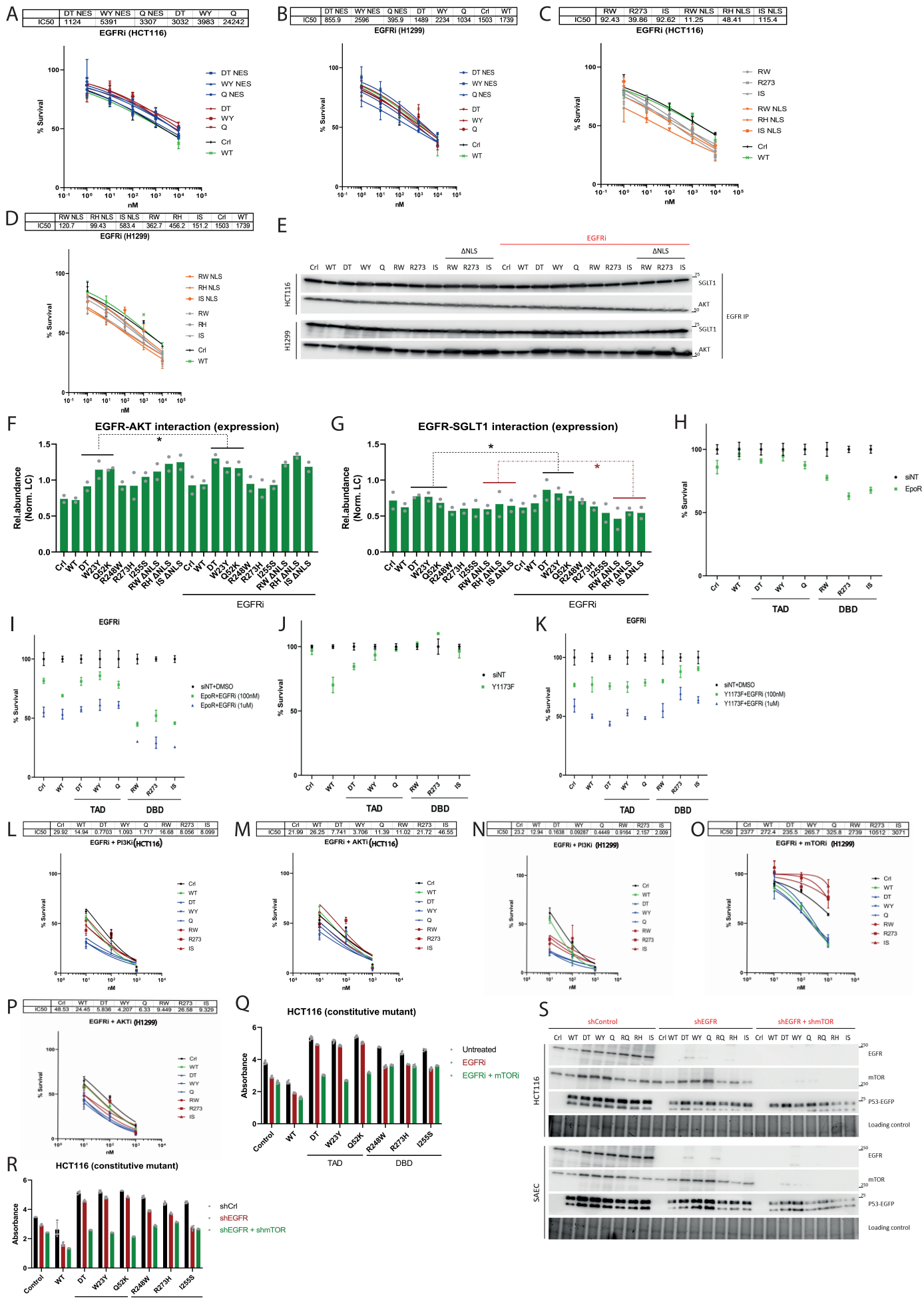
Supplementary Fig. 10: Validation of structural EGFR mutants that disrupt interaction with mutant p53

(A) Structure of EGFR-p53TADW23Y complex. EGFR is shown with its surface colored according to the electrostatic potential (with red to blue color corresponding to -5kcal/mol to + 5kcal/mol) and the p53TAD1 peptide is shown as cartoon (Cyan). Residue pY23 (phosphorylated tyrosine 23) from TAD1 is shown as sticks. Phosphorylation status of W23Y mutant was predicted using the webserver NetPhos-3.1. (B-C) Structures of EGFR-p53DBDI255S and EGFR-p53DBDR248W complexes. EGFR is shown with its surface colored according to the electrostatic potential (with red to blue color corresponding to -5kcal/mol to + 5kcal/mol) and (B) p53DBD-I255S and (C) p53DBD-R248W shown as cartoon (Cyan) with the solvent exposed hydrophobic patch highlighted in magenta. Residues F113, L114 from p53 DBD and L736, L758 from EGFR shown as sticks. Disruption of hydrophobic interactions due to L736K and L758K mutations. (D) Protein levels of EGFR pulled down with p53 in HCT116 cells expressing WT and mutant p53. Cells were also treated with EGFR siRNA and expressed the EGFR mutants that disrupt TAD binding (E685K, E685F) and mutants that disrupt DBD binding (L736K, L758K). n=2. (E) Protein levels of pEGFR, pAKT and c-Myc in HCT116 cells expressing WT and mutant p53. Cells were also treated with EGFR siRNA and expressed the EGFR mutants that disrupt TAD binding (E685K, E685F) and mutants that disrupt DBD binding (L736K, L758K). n=2. Source data are provided as a Source Data file.



Supplementary Fig. 11: Mutant p53 confers resistance to EGFR inhibition via different mechanisms

(A) Quantification of pEGFR (pan) protein levels in H1299 cells expressing WT and mutant p53 treated with EGFR inhibitor. n=2. (B) Protein levels of pEGFR (pan) and p53-EGFP in HCT116 and H1299 cells expressing WT and mutant p53 treated with EGFR inhibitor. n=2. (C-F) Quantification of (C) nuclear pEGFR (pan), (D) whole-cell c-Myc in H1299 cells and (E,F) Cyclin D1 protein levels in HCT116 and H1299 cells expressing WT and mutant p53 treated with EGFR inhibitor. n=2. (G) Protein levels of nuclear pEGFR (pan), whole-cell c-Myc and Cyclin D1 in HCT116 and H1299 cells expressing WT and mutant p53 treated with EGFR inhibitor. n=2. (H-I) Quantification of (H) pAKT and (I) SGLT1 protein levels in H1299 cells expressing WT and mutant p53. Cells were also treated with EGFRi. Statistical tests performed on TAD (n=3 independent mutants) versus DBDΔNLS (n=3 independent mutants).n=2. (J) Protein levels of SGLT1 and pAKT in HCT116 and H1299 cells expressing WT and mutant p53. Cells were also treated with EGFRi. n=2. (K-L) Dose response curves of HCT116 cells expressing WT and mutant p53 treated with EGFR inhibitor. Cells were also treated with EGFR siRNA and expressed the EGFR mutants that disrupt (K) TAD binding (E685K, E685F) and mutants that disrupt (L) DBD binding (L736K, L758K). n=3 independent experiments. Two-tailed unpaired Student's t-test was performed. (* p<0.05, ** p<0.01, *** p<0.001, **** p<0.0001). Source data are provided as a Source Data file.



Supplementary Fig. 12: Mutant p53 confers resistance to EGFR inhibition via different mechanisms

(A-D) Dose response curves of (A, C) HCT116 and (B, D) H1299 cells expressing WT and mutant p53 treated with EGFR inhibitor. n=3 independent experiments. (E) Protein levels of SGLT1 and AKT in HCT116 and H1299 cells expressing WT and mutant p53 and treated with EGFRi. Protein levels of SGLT1 and AKT pulled down with EGFR. n=2. (F-G) Quantification of (F) AKT (after EGFR pulldown) and (G) SGLT1 (after EGFR pulldown) protein levels in HCT116 cells expressing WT and mutant p53 and treated with EGFRi. n=2. (H-K) Survival plots various cancer cell lines and HCT116 expressing WT and mutant p53 treated with EGFR inhibitor. Cells were treated with EGFR siRNA and expressed (H, I) EGFR EpoR (high affinity for SHP1) or (J, K) EGFR Y1173F (reduced affinity for SHP1). n=3 independent experiments. (L-M) Dose response curves of HCT116 cells expressing WT and mutant p53 treated with EGFR inhibitor in combination with (L) PI3Ki and (M) AKTi. n=3 independent experiments. (N-P) Dose response curves of H1299 cells expressing WT and mutant p53 treated with EGFR inhibitor in combination with (N) PI3Ki, (O) mTORi, (P) AKTi. n=3 independent experiments. (Q-R) Quantification of crystal violet intensity from colony formation assays using HCT116 cells constitutively expressing WT and mutant p53. Cells were treated with (Q) EGFRi or EGFRi+mTORi and (R) inducible expression of shControl or shEGFR and/or shmTOR. n=4 independent experiments. Error bars denote Mean \pm SEM of technical replicates. $p < 0.0001$ for DBD untreated vs DBD EGFRi/DBD shEGFR and TAD untreated vs TAD EGFRi+mTORi/shEGFR+shmTOR (n=3 mutants each). (S) Protein levels of EGFR, mTOR and p53-EGFP in HCT116 and SAECK cells constitutively expressing WT and mutant p53 and doxycycline-induced expression of shEGFR and shmTOR. Two-tailed unpaired Student's t-test was performed. (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$, **** $p < 0.0001$). Source data are provided as a Source Data file.