

# 

**Citation:** Silva BRA, Rodrigues RS, Rufino R, Costa CH, Vilela VS, Levy RA, et al. (2018) Computed tomography trachea volumetry in patients with scleroderma: Association with clinical and functional findings. PLoS ONE 13(8): e0200754. https://doi.org/10.1371/journal.pone.0200754

Editor: Masataka Kuwana, Keio University, JAPAN

Received: February 23, 2018

Accepted: July 1, 2018

Published: August 1, 2018

**Copyright:** © 2018 Silva et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All data are available from the Open Science Framework (<u>https://osf.io/</u>ue7mn/).

Funding: This research was supported by the Brazilian Council of Scientific and Technological Development (Conselho Nacional de Desenvolvimento Científico e Tecnológico - CNPq] and the Carlos Chagas Filho Foundation for Research Support of the State of Rio de Janeiro (Fundação Carlos Chagas Filho de Amparo à Pesquisa do Estado do Rio de Janeiro - FAPERJ). The funders had no role in study design, data **RESEARCH ARTICLE** 

# Computed tomography trachea volumetry in patients with scleroderma: Association with clinical and functional findings

Bruno Rangel Antunes Silva<sup>1</sup><sup>®</sup>, Rosana Souza Rodrigues<sup>2</sup><sup>®</sup>, Rogério Rufino<sup>1</sup><sup>®</sup>, Cláudia Henrique Costa<sup>1</sup><sup>®</sup>, Veronica Silva Vilela<sup>1</sup><sup>®</sup>, Roger Abramino Levy<sup>1</sup><sup>®</sup>, Alan Ranieri Medeiros Guimarães<sup>3</sup><sup>®</sup>, Alysson Roncally Silva Carvalho<sup>3,4</sup><sup>®</sup>, Agnaldo José Lopes<sup>1</sup><sup>®</sup>\*

1 Postgraduate Programme in Medical Sciences, School of Medical Sciences, State University of Rio de Janeiro, Rio de Janeiro, Brazil, 2 Department of Radiology, Federal University of Rio de Janeiro, Rio de Janeiro, Brazil, 3 Laboratory of Respiration Physiology, Carlos Chagas Filho Institute of Biophysics, Federal University of Rio de Janeiro, Rio de Janeiro, Brazil, 4 Laboratory of Pulmonary Engineering, Biomedical Engineering Programme, Alberto Luiz Coimbra Institute of Post-Graduation and Research in Engineering, Federal University of Rio de Janeiro, Rio de Janeiro, Brazil

So These authors contributed equally to this work.

## Abstract

## Background

In scleroderma, excessive collagen production can alter tracheal geometry, and computed tomography (CT) volumetry of this structure may aid in detecting possible abnormalities. The objectives of this study were to quantify the morphological abnormalities in the tracheas of patients with scleroderma and to correlate these findings with data on clinical and pulmonary function.

### Methods

This was a cross-sectional study in which 28 adults with scleroderma and 27 controls matched by age, gender and body mass index underwent chest CT with posterior segmentation and skeletonization of the images. In addition, all participants underwent pulmonary function tests and clinical evaluation, including the modified Rodnan skin score (mRSS).

### Results

Most patients (71.4%) had interstitial lung disease on CT. Compared to controls, patients with scleroderma showed higher values in the parameters measured by CT trachea volume-try, including area, eccentricity, major diameter, minor diameter, and tortuosity. The tracheal area and equivalent diameter were negatively correlated with the ratio between forced expiratory flow and forced inspiratory flow at 50% of forced vital capacity (FEF<sub>50%</sub>/FIF<sub>50%</sub>) (r = -0.44, p = 0.03 and r = -0.46, p = 0.02, respectively). The tracheal tortuosity was negatively correlated with peak expiratory flow (r = -0.51, p = 0.008). The mRSS showed a positive correlation with eccentricity (r = 0.62, p < 0.001) and tracheal tortuosity (r = 0.51, p = 0.007),

<sup>\*</sup> alopes@souunisuam.com.br

collection and analysis, decision to publish, or preparation of the manuscript.

PLOS ONE

**Competing interests:** The authors have declared that no competing interests exist.

while the presence of anti-topoisomerase I antibody (ATA) showed a positive correlation with tracheal tortuosity (r = 0.45, p = 0.03).

#### Conclusions

In a sample composed predominantly of scleroderma patients with associated interstitial lung disease, there were abnormalities in tracheal geometry, including greater eccentricity, diameter and tortuosity. In these patients, abnormalities in the geometry of the trachea were associated with functional markers of obstruction. In addition, tracheal tortuosity was correlated with cutaneous involvement and the presence of ATA.

#### Introduction

Scleroderma or systemic sclerosis (SSc) is a chronic progressive autoimmune disease of connective tissue characterized by microvascular involvement, activation of the immune system, and increased deposition of extracellular matrix in the skin and internal organs by excess collagen fibers, leading to fibrosis [1–4]. Scleroderma is eight times more common in females than in males; its reported prevalence is approximately 10 cases per 100,000 person, and this rate is probably underestimated [5]. The disease can affect several organs and systems; the skin is the most frequently affected site, followed by the lungs, kidneys, musculoskeletal system, cardiovascular system, and gastrointestinal tract. The presence of multiple affected sites worsens the prognosis [1].

In scleroderma, thoracic involvement is observed mainly as diffuse fibrosis or pulmonary hypertension; these conditions are associated with limited (lc-SSc) and diffuse cutaneous (dc-SSc) forms of scleroderma, respectively [1]. In an autopsy study, parenchymal involvement was seen in up to 100% of patients with scleroderma [2]. Although interstitial lung involvement is subclinical and asymptomatic at early stages in most patients, interstitial lung disease (ILD) associated with scleroderma (ILD-SSc) is observed in approximately 40% of cases and is a major cause of morbidity and mortality [6]. Despite the constellation of thoracic manifestations that occur in patients with scleroderma, little is known about the involvement of the trachea in these patients. Ooi et al. [7] reported that scleroderma affected small and large airways in 45–100% of patients. In this context, the trachea has been a 'forgotten zone' in the study of several diseases because the pathological processes involving this structure have often not received the necessary clinical recognition.

Computed tomography (CT) has become an important part of the detection and evaluation of routine thoracic involvement in scleroderma, and the abnormalities observed by this method are closely correlated with the observed physiological parameters [2]. In the last two decades, various computer tools that can be used to automatically slice the chest using CT images have been developed. These include multiplanar reformation, regional lung attenuation analysis of lung tissue and quantification of anatomical images, including the area and volume of airways and lungs [2,7]. In scleroderma, computer-assisted tomography analysis is highly efficient and, in combination with physiological and patient-centered measurements, may provide a means to accurately assess and monitor lung disease progression and response to therapy [2,8,9].

An understanding of the acquisition, processing and analysis of CT scans and how these processes affect the imaging of the trachea is essential for assessing the accuracy of the measurements and making effective use of newly available tools. In the study of the trachea, the cross-sectional area and diameter are the most commonly measured dimensions; in the

context of assessing possible obstruction, length and caliber are important [10–13]. More recently, the process of skeletonization and volumetry of the airways through CT in normal individuals and in individuals with some clinical conditions has been described [10–15]. In this method, the digital imaging component is transformed into a subset of the original component [10]. After automated segmentation of CT images, the skeletonization algorithm allows the extraction of a tracheal centerline and facilitates the reconstruction of CT data orthogonal to the reduction of the effects of partial volume averages [11]. However, no study has explored the use of this resource in the evaluation of the tracheas of patients with scleroderma.

We hypothesized that the excessive production of collagen that occurs in scleroderma alters the geometry of the trachea and that the skeletonization of this structure would aid in the detection of possible abnormalities. Thus, the present study aimed to identify and quantify the morphological abnormalities in the tracheas of patients with scleroderma and, secondarily, to correlate these findings with data on clinical and pulmonary function.

### Methods

#### Patients

This cross-sectional study was conducted between February 2016 and September 2017 in 43 consecutive patients with scleroderma who were  $\geq 18$  years of age, of both sexes, and were seen regularly at the Piquet Carneiro Polyclinic of the State University of Rio de Janeiro, Rio de Janeiro, Brazil. The patients included in the study had been diagnosed with scleroderma by a rheumatologist according to the parameters set forth by the American College of Rheumatology/European League Against Rheumatism [1]. The following exclusion criteria were used: clinical instability; history of respiratory infection in the last three weeks; history of previous or current smoking; evidence of overlapping of scleroderma with other connective tissue diseases; report of previous tracheal or pleuropulmonary disease not related to scleroderma; and inability to perform pulmonary function tests (PFTs). Regarding cutaneous involvement, patients were classified as having lc-SSc (thickening of the skin distal to the elbows and knees and proximal to the clavicles, including the face) or dc-SSc (thickening of the proximal skin as well as of the skin distal to the elbows and knees and including the trunk and face) [16]. The modified Rodnan skin score (mRSS) was used to assess skin damage in patients with scleroderma. In this system, a score of 0 (no thickening), 1 (light thickening), 2 (moderate thickening) or 3 (severe thickening) is assigned to each area, resulting in a total score ranging from 0 (best) to 51 (worst) [17]. The presence of autoantibodies, including anti-topoisomerase I and anti-centromere, was also investigated.

We also evaluated a control group of 27 individuals aged  $\geq$  18 years of both sexes. The subjects in the control group were asked to perform the PFTs after undergoing chest CT scanning in our service for the following reasons: investigation of contact with tuberculosis patients (n = 9); staging of neoplasms outside the thorax (n = 7); evaluation of trauma (n = 6); and evaluation of fever of unknown origin (n = 5). The following criteria were used to select subjects for the control group: no previous history of smoking or chronic tracheal or pleuropulmonary diseases; chest CT scans without abnormalities; and lung function parameters within the normal range (*i.e.*, no value below the lower limit of normal or above the upper limit of normal in relation to the predicted value). All CT scans and PFTs for the control group were performed using the same equipment that was used for the scleroderma group.

The protocol was approved by the Research Ethics Committee of the State University of Rio de Janeiro under the number CAAE- 50752615.9.0000.5259, and it complied with the current national and international standards. All individuals signed an informed consent form.

#### **Pulmonary function testing**

The PFTs performed were spirometry, body plethysmography, and diffusion capacity for carbon monoxide (DLco). The exams were performed on an HDpft 3000 (nSpire Health, Inc., Longmont, CO, USA) according to the standards set forth by the American Thoracic Society [18]. The Brazilian reference values [19,20] were used, and the results are expressed as percentages of the predicted values. An increase in the ratio between forced expiratory flow and forced inspiratory flow at 50% of forced vital capacity-FVC (FEF<sub>50%</sub>/FIF<sub>50%</sub>) > 1.50 was used as an indicator of extrathoracic airway obstruction [21].

#### CT scan interpretation and protocol

CT scans were performed on a 64-channel multislice Brilliance 40 scanner (Philips Medical Systems, Cleveland, OH, USA) that was capable of performing volumetric acquisitions with subsequent multiplanar reconstructions. The acquisitions were performed in the axial plane with patients in the supine position using the technical parameters 120 kV and 458 mA (these parameters varied according to the biotype of the patient), slice thickness 2 mm, and pitch 2 mm from the jugular notch to the xiphoid process in maximal inspiration and expiration. After acquisition of the images, a high-resolution reconstruction with a matrix of 512 × 512 was performed using a high-frequency algorithm, a window width of 1200 HU, and a level centered at -800 HU.

Parenchymal abnormalities on CT were interpreted by two independent readers (R.S.R. and G.B.C.) who were blinded to patient history and physiological results; a consensus opinion was reached in cases in which there was disagreement. CT scans were reviewed at five levels, as follows: 1) origin of major vessels; 2) carina; 3) confluence of pulmonary veins; 4) halfway between the third and the fifth sections; 5) 1 cm above the right hemidiaphragm [22,23]. The total extent of ILD was estimated to the nearest five percent in each of the five levels, with global extent of disease on CT as the mean of the scores [22–24]. The coarseness of pulmonary fibrosis was evaluated as follows: 0, ground-glass opacification alone; 1, fine intralobular fibrosis; 2, microcystic honeycombing comprising air spaces  $\leq$  4 mm in diameter; and 3, macrocystic honeycombing comprising air spaces > 4 mm in diameter. The total coarseness score for each patient was derived by summing the scores at the five levels (range 0 to 15) [22,23]. For each patient, the total extension of ILD and the coarseness of pulmonary fibrosis were derived by averaging the scores at each level assessed by the two independent readers. Finally, the extent of ILD-SSc was classified as limited (lung parenchyma involvement <20%) or extensive (>20%). For indeterminate cases, ILD-SSc was considered extensive if FVC <70% and limited if FVC >70% [22,25].

#### Imaging processing

The airways were segmented using 3DSlicer version 4.4.0 (http://slicer.org) [26] with the aid of its AirwaySegmentation extension. At the end of processing, a nearly raw raster data (.nrrd) file was saved. Using AirwayProcessing, two distinct processes were executed: 1) the .nrrd file was read and transformed into a binary matrix of data; in this way, the process of skeletonization was initiated, and a skeleton composed of several points was then produced; 2) using the skeleton and its coordinates (X, Y and Z axes), cross-sectional planes were generated. Individual processing of all points was performed using the slice command, and the processing angles were defined by a normal line between the point in question and the next five points. The plane of square cross-section was formed by a square grid with 70 pixels on the side. Using MATLAB 2014a (MathWorks Inc., Natick, MA, USA), information obtained individually by the slice command was catalogued using the 'regionprops' command.

Fig 1 shows the 3D reconstruction, the tracheal skeleton, the cross-sectional planes and their geometric parameters [9,10]. The scheme show in Fig 1 was chosen for illustrative purposes and accurately represents the region of the trachea in a 2D environment. In Fig 1, the axial sections indicated by specific letters represent area (\*A), perimeter (\*B), eccentricity (\*C), equivalent diameter (\*D), major diameter (\*E) and minor diameter (\*F). It is emphasized that the segments shown in red and blue in Fig 1 were enlarged for visualization purposes.

Thus, the following measures were calculated for each study participant:



**Fig 1. 3D reconstruction, cross-sectional planes, skeleton and indication of the plane used for representation.** The letters A to F and the values for the selected plane indicate, respectively: (\*A) area (182.4 mm<sup>2</sup>), which is the product of the pixel size and the number of pixels in the region (in this figure, the area is shown in yellow and includes the red line that demarcates the perimeter); (\*B) perimeter (47.3 mm), which is the length of the line indicated in red; (\*C) eccentricity (0.35), which is indicated by the equivalent ellipse drawn in blue; (\*D) equivalent diameter (15.2 mm), which is the circle of area equivalent to the cross-sectional area; (\*E) major diameter (15.8 mm), which is the longest segment of the equivalent ellipse (indicated by the inner line); and (\*F) minor diameter (14.8 mm), which is the smallest segment (indicated by the inner line).

https://doi.org/10.1371/journal.pone.0200754.g001

1) Area: the measured area of the tracheal lumen intersected by the cross-sectional plane. The area was calculated by multiplying the pixel area value by the number of pixels present, as shown in Eq.1,

$$Area = ps * n \tag{1}$$

where *ps* is the individual area of each pixel and *n* is the number of pixels.

- 2) Perimeter: the length of the outer margin of the tracheal lumen.
- 3) Eccentricity: a parameter associated with the ellipsoidal shape of a given region. It is the ratio of the focus of the ellipse to its largest diameter. The result is a value between 0 and 1, where 0 is the representation of a circle and 1 is the representation of a line. Fig 1\*C illustrates eccentricity as the equivalent ellipse (shown in blue); the outer edge of the tracheal lumen is shown in red. The eccentricity was calculated according to Eq 2,

$$Eccentricity = 2 * \frac{\sqrt{\frac{(MaAL)^2}{2} - \frac{(MiAL)^2}{2}}}{MaAL}$$
(2)

where MaAL represents the largest diameter and MiAL the smallest diameter of the ellipse.

4) Equivalent diameter: a circle of the same area as the region intersected by the cross-sectional plane. In Fig 1\*D, the equivalent diameter is shown as a blue circle with an area equivalent to that of the tracheal lumen in the same region. The equivalent diameter was calculated using Eq 3,

$$Equivalent \ diameter = \frac{\sqrt{4 * area}}{\pi} \tag{3}$$

5) Major diameter: the length of the longest segment of the ellipse equivalent to the region. Similarly, the minor diameter represents the length of the smallest segment of the ellipse. Fig 1\*E and 1\*F denote the perimeter of the tracheal lumen (red) along with its ellipse (blue); the center lines indicate the largest and the smallest segment, respectively, of each region.

We also calculated the tortuosity (sinuosity index), which is the deviation of the central axis of the trachea considering its proximal and distal extremities. Mathematically, the tortuosity was determined from the sum of the Euclidean distance of each segment of the trachea, with a distance equivalent to the thickness of each cross-section of the original CT image, divided by the Euclidean distance of the end points of the trachea. The tortuosity may be expressed succinctly using the formula described in Eq. 4,

$$Tortuosity = \frac{L}{\nu d}$$
(4)

where *L* is the length of the trachea and *vd* is the vectorial distance between the points at the extremities (Fig 2). The tortuosity is expressed as a value  $\geq 1$ ; the higher the index, the greater the tortuosity of the trachea [27].

Fig 3 exemplifies the skeletonization process of the trachea of a study participant. The figure shows the three-dimensional reconstruction images of the trachea in two different planes.

There is inherent variability in the measures used to determine the quantitative metrics of eccentricity and tortuosity. For eccentricity, there is a value for each section plane of the trachea of a given individual (Fig 3). We use the median to represent the value of each individual; therefore, there are n medians, where n is the number of elements in each group of variables.



scheme, L is the length of the trachea (the total length of 'ab', shown in blue) and vd is the vectorial distance between the points at the extremities (the length of the shortest possible path between 'a' and 'b', shown in red).

https://doi.org/10.1371/journal.pone.0200754.g002

For tortuosity, only one value was calculated for each trachea; it depends on the extreme values (proximal and distal), as shown in Fig 2.

#### Statistical analysis

To better evaluate the effect of the extent of ILD-SSc on the clinical variables, PFTs, and CT trachea volumetric parameters, we divided the participants into three subgroups as follows: 1) a control group; 2) a scleroderma group with no pulmonary involvement/limited ILD; and 3) a scleroderma group with extensive ILD. ANOVA followed by the Bonferroni post hoc test was used to compare the results obtained for the patients in these three groups. Comparison of the clinical variables, PFTs, and CT trachea volumetric parameters of the SSc patients without pulmonary involvement with those of control subjects and with those of SSc patients with no pulmonary involvement/limited ILD-SSc and SSc patients with extensive ILD-SSc was performed using Student's t-test for independent samples in the case of numerical data and using the chi-square or Fisher's exact test in the case of categorical data. To evaluate the associations between the numerical variables of PFTs and CT trachea volumetric parameters, the Pearson correlation coefficient (r) was used. The criterion for determining significance was 5%. Statistical analysis was performed using SAS 6.11 software (SAS Institute, Inc., Cary, NC, USA).

### Results

Of the 43 patients included in the study, 15 were excluded for the following reasons: reported being a smoker (n = 8); history of previous pleuropulmonary disease not associated with

Α



**Fig 3. Skeletonization process of the trachea.** Three-dimensional reconstruction of the trachea of one study participant in two planes (A and B). The median values showed in the ten planes of square cross-sections are as follows: area =  $184.5 \text{ mm}^2$ ; perimeter = 47.6 mm; eccentricity = 0.51; equivalent diameter = 15.3 mm; major diameter = 15.9 mm; minor diameter = 14.4 mm; and tortuosity = 1.027.

https://doi.org/10.1371/journal.pone.0200754.g003

PLOS ONE

scleroderma (n = 3); inability to perform PFTs (n = 2); scleroderma-polymyositis overlap syndrome (n = 1); and scleroderma-rheumatoid arthritis overlap syndrome (n = 1). In the total sample of patients, the mean age of subjects with scleroderma was  $52.5 \pm 11$  years, and 25 (89.3%) were women. The disease duration was  $4.21 \pm 2.50$  years from the onset of non-Raynaud's phenomenon and  $9.65 \pm 5.13$  years from the onset of Raynaud's phenomenon symptoms. Nineteen patients (67.9%) had lc-SSc, and nine (32.1%) had dc-SSc; of the latter, six (21.4%) had a mRSS > 18. Anti-topoisomerase I antibody (ATA), anticentromere antibody and anti-RNA polymerase III were positive in 13 (46.4%), six (21.4%) and three (10.7) patients, respectively; no autoantibodies were identified in six (21.4%) patients. Seven patients (25%) had an FEF<sub>50%</sub>/ FIF<sub>50%</sub> ratio > 1.50. Comparisons of clinical data, pulmonary function parameters and computed tomography scores in the control group, the scleroderma group with no pulmonary involvement/limited ILD, and the scleroderma group with extensive ILD are shown in Table 1. In addition, comparisons of lung function parameters in the control group and in patients without pulmonary involvement on CT did not show any significant difference (p > 0.05 for all).

On thorax CT, 16 patients (57.1%) were classified as having no pulmonary involvement (n = 8) or limited pulmonary involvement (n = 8), while 12 (42.9%) were classified as having extensive pulmonary involvement. In the measurements obtained through CT trachea volumetry, patients with scleroderma presented higher values for the following parameters: area, eccentricity, major diameter, minor diameter, and tortuosity. Table 2 compares the CT trachea volumetry findings for control subjects, patients in the scleroderma group with no pulmonary involvement/limited ILD, and patients in the scleroderma group with extensive ILD.



Variable Control group Scleroderma group with no pulmonary involvement/limited Scleroderma group with extensive p value (n = 27)ILD ILD (n = 16)(n = 12)Demographic data Females 23 (85.2) 14 (87.5) 11 (91.7) 0.41  $49 \pm 12.6$  $51.7 \pm 10$  $52.8 \pm 11.2$ 0.28 Age (years) BMI (kg/m<sup>2</sup>)  $27.2 \pm 5.87$  $24.2 \pm 5.22$  $25.1\pm5.30$ 0.09 Type of scleroderma lc-SSc 7 (58.3) 0.34 12 (75) \_ dl-SSc 4(25)5 (41.7) mRSS \_  $10.3 \pm 7.44$  $11.8 \pm 7.60$ 0.12 Type of autoantibody Anti-topoisomerase I antibody -5 (31.2) 8 (66.7) 0.28 Anticentromere antibody 5 (31.2) 1 (8.33) Anti-RNA polymerase III 2 (12.5) 1 (8.33) Autoantibody not identified 4 (25) 2 (16.7) Lung function FVC(L)  $3.08 \pm 0.81$  $2.62 \pm 0.72^{*}$  $2.25 \pm 0.87^{*}$ † 0.008 FVC (% predicted)  $102.4 \pm 19.8$  $85 \pm 19.3^*$  $68 \pm 21.5^{*}$ † 0.005  $FEV_1(L)$  $2.51\pm0.75$  $2.11 \pm 0.63^{\circ}$  $1.80 \pm 0.59^{*}$ † 0.009 FEV1 (% predicted)  $100.7 \pm 18.5$  $83.1 \pm 16^{*}$  $69.2 \pm 17.8^{*}$ † 0.006 FEV<sub>1</sub>/FVC (%)  $78 \pm 12.6$  $80 \pm 12$  $86\pm11.3$ 0.11 PEF (L/s)  $7.75 \pm 2.14$  $6.25 \pm 2.11$  $5.26 \pm 1.97^{*}$ 0.032 PEF (% predicted)  $110 \pm 31.7$  $86 \pm 30.5$  $75.5 \pm 29.2^*$ 0.025 FEF50%/FIF50% (%)  $1.28 \pm 0.72^{*}$  $1.34 \pm 0.70^{*}$ 0.046  $1.04 \pm 0.62$ DLco (mL/min/mmHg)  $20.6 \pm 3.44$  $18 \pm 4.22^{*}$ 9.27 ± 3.23\*† 0.003 DLco (% predicted)  $97 \pm 21.6$  $70.3 \pm 18.3^{*}$  $60\pm17.8^*\dagger$ 0.001 Raw (cm H<sub>2</sub>O/L/s)  $1.58\pm0.75$ 0.32  $1.78 \pm 0.77$  $1.93 \pm 0.78$ SGaw (L/s/cm H<sub>2</sub>O/L)  $0.210 \pm 0.078$  $0.245 \pm 0.089$  $0.257 \pm 0.103$ 0.09 Computed tomography scores ILD-SSc extent (%  $9.63 \pm 8.80$  $28.1 \pm 10.4$ < 0.0001 parenchyma) Coarseness of pulmonary  $6.35 \pm 4.20$ < 0.0001  $2.15 \pm 1.44$ fibrosis

Table 1. Demographic characteristics, clinical data, pulmonary function and computed tomography scores of patients with scleroderma and of patients in the control group.

The values shown are means  $\pm$  SD or number (%). Bold type indicates significant differences.

\*Significantly different from control group.

 $\pm$  Significantly different from scleroderma group with no pulmonary involvement/limited ILD-SSc. ILD-SSc = interstitial lung disease associated with scleroderma. BMI = body mass index; lc-SSc = limited cutaneous form; dc-SSc = diffuse cutaneous form; mRSS = modified Rodnan skin score; FVC = forced vital capacity; FEV<sub>1</sub> = forced expiratory volume in one second; PEF = peak expiratory flow; FEF<sub>50%</sub>/FIF<sub>50%</sub> = ratio between the forced expiratory flow and forced inspiratory flow at 50% of forced vital capacity; DLco = diffusing capacity for carbon monoxide; Raw: airway resistance; SGaw: specific airway conductance.

https://doi.org/10.1371/journal.pone.0200754.t001

We also compared patients without pulmonary involvement on CT (n = 8) with controls. In this evaluation, patients without pulmonary involvement on CT showed higher values, with significant differences in the following CT tracheal volumetric findings: area (210.7 ± 35 vs. 207.6 ± 32.6 mm<sup>2</sup>, p = 0.031; eccentricity ( $0.50 \pm 0.05$  vs.  $0.46 \pm 0.04$ , p = 0.018); major diameter ( $17.5 \pm 1.70$  vs.  $17.1 \pm 2.22$  mm, p = 0.039); minor diameter ( $14.9 \pm 1.68$  vs.  $14.5 \pm 2.11$  mm, p = 0.044); and tortuosity ( $1.039 \pm 0.017$  vs.  $1.023 \pm 0.014$ , p = 0.01).



Variable	Control group (n = 27)	Scleroderma group with no pulmonary involvement/limited ILD (n = 16)	Scleroderma group with extensive ILD (n = 12)	p value
Area (mm <sup>2</sup> )	$207.6 \pm 32.6$	$211.3 \pm 38.2^*$	$214.6 \pm 43.6^*$	0.028
Perimeter (mm)	$45.3 \pm 4.12$	49.2 ± 4.45	51.5 ± 4.67	0.12
Eccentricity	$0.46 \pm 0.04$	$0.51 \pm 0.06^{*}$	$0.54 \pm 0.06^{*}$	0.013
Equivalent diameter (mm)	$16.1 \pm 1.24$	$16.2 \pm 1.55$	$16.5 \pm 1.60$	0.82
Major diameter (mm)	$17.1 \pm 2.22$	$17.6 \pm 1.92^*$	$18.2 \pm 2.01^{*}$	0.039
Minor diameter (mm)	$14.5 \pm 2.11$	$15 \pm 1.72^*$	$15.3 \pm 1.85^*$	0.043
Tortuosity	$1.023 \pm 0.014$	$1.039 \pm 0.014^{*}$	$1.058 \pm 0.015^{*}$ †	0.009

#### Table 2. Variables of CT trachea volumetry according to group.

The values shown are means  $\pm$  SD or number (%). Bold type indicates significant differences.

\*Significantly different from control group.

+Significantly different from scleroderma group with no pulmonary involvement/limited ILD-SSc. ILD-SSc = interstitial lung disease associated with scleroderma.

https://doi.org/10.1371/journal.pone.0200754.t002

We evaluated the correlations between the parameters provided by CT trachea volumetry and the pulmonary function indices (absolute values) (Table 3). The tracheal area was negatively correlated with FEF<sub>50%</sub>/FIF<sub>50%</sub> (r = -0.44, p = 0.03), while the eccentricity was negatively correlated with FVC (r = -0.57, p = 0.002) (Fig 4) and forced expiratory volume in one second (r = -0.50, p = 0.009). The equivalent diameter was negatively correlated with FEF<sub>50%</sub>/FIF<sub>50%</sub> (r = -0.46, p = 0.02), while tortuosity was negatively correlated with peak expiratory flow (PEF) (r = -0.51, p = 0.008) (Fig 5). A positive correlation of tortuosity with coarseness of pulmonary fibrosis in CT was also found (r = 0.45, p = 0.02).

We also evaluated the correlations between the clinical findings and the parameters provided by CT trachea volumetry. The mRSS showed a positive correlation with eccentricity (r = 0.62, p < 0.001) and tortuosity (r = 0.51, p = 0.007). In turn, the presence of anti-topoisomerase I antibody showed a positive correlation with tortuosity (r = 0.45, p = 0.03).

### Discussion

The main findings of the present study were that in a sample composed predominantly of scleroderma patients with associated ILD, the trachea showed greater area, eccentricity, and

Table 3. Pearson's correlation coefficients for CT trachea volumetry, pulmonary fibrosis and pulmonary function.

	FVC	FEV <sub>1</sub>	FEV <sub>1</sub> /FVC	PEF	FEF <sub>50%</sub> /FIF <sub>50%</sub>	DLco	Raw	SGaw	Coarseness of fibrosis
Area	-0.13	-0.11	-0.06	0.24	-0.44*	-0.24	0.12	-0.19	0.25
Perimeter	-0.09	-0.07	-0.08	0.25	-0.47†	-0.18	0.13	-0.21	0.17
Eccentricity	-0.57‡	-0.50†	-0.12	-0.07	-0.17	-0.35	-0.11	-0.04	0.26
Equivalent diameter	-0.11	-0.10	-0.07	0.25	-0.46†	-0.27	0.11	-0.21	0.18
Major diameter	-0.04	-0.06	-0.08	0.25	-0.42*	-0.16	0.07	-0.20	0.22
Minor diameter	-0.22	-0.19	-0.03	0.21	-0.47†	-0.11	0.06	-0.17	0.25
Tortuosity	-0.17	-0.15	-0.26	-0.51†	-0.28	-0.12	0.09	-0.15	0.45*

 $FVC = forced vital capacity; FEV_1 = forced expiratory volume in one second; PEF = peak expiratory flow; FEF_{50\%}/FIF_{50\%} = ratio between the forced expiratory flow and forced inspiratory flow at 50% of forced vital capacity; DLco = diffusing capacity for carbon monoxide; Raw = airway resistance; SGaw = specific airway conductance. Bold type indicates significant differences.$ 

\*p < 0.05

†p < 0.01

p < 0.005

https://doi.org/10.1371/journal.pone.0200754.t003



Fig 4. Relationship between forced vital capacity (FVC) and eccentricity of the trachea (r = -0.57, p = 0.002). Scleroderma group (1) = scleroderma group with no pulmonary involvement/limited interstitial lung disease; scleroderma group (2) = scleroderma group with extensive interstitial lung disease.

#### https://doi.org/10.1371/journal.pone.0200754.g004

tortuosity. In these patients, a greater tracheal tortuosity led to a smaller airflow; in addition, the lower the tracheal diameter, the greater the degree of airway obstruction. Furthermore, tracheal tortuosity was associated with the presence of pulmonary fibrosis, the presence of ATA, and greater mRSS. To our knowledge, this is the first study to show abnormalities in the structure and function of the trachea as well as its correlations with clinical findings in patients with scleroderma.

In recent years, several CT image enhancement techniques have been developed in an attempt to find a method that is both quantitative and reliable and allows more accurate assessment than conventional visual reading. Compared to the traditional visual interpretation of





https://doi.org/10.1371/journal.pone.0200754.g005

CT findings, computer-based automatic evaluation can improve the objectivity, sensitivity, and repeatability of quantitative chest imaging analyses [2,10]. In this study, we applied a computer-assisted method to evaluate the tracheas of patients with scleroderma in an unprecedented way considering that computerized evaluation has previously only been used in the study of ILD-SSc [2,28]. Our results indicate that CT trachea volumetry and subsequent skeletonization of the images provide a number of interesting measures for the best evaluation of patients with scleroderma. We observed that patients with scleroderma presented greater area, eccentricity, and tortuosity of the trachea. In scleroderma, myofibroblasts constitutively secrete components of the extracellular matrix and exert excessive cicatrization of the skin and internal organs [3,29]. Thus, we think that the cicatricial changes that occur at the level of the neck can directly impact the geometry of the trachea and alter the measures of diameter and tortuosity of this structure.

In our study, the deviations of the trachea were measured by means of the tortuosity (sinuosity index) after the skeletonization process. The skeletonization process has been used previously by us to obtain the skeletons of specific structures by thinning [10]. Interestingly, we found an association between tracheal tortuosity and the pulmonary fibrosis score obtained by semi-quantitative CT reading. This association between structural alterations of the trachea and the lung, despite being evaluated by different techniques in our study, suggests that the deformity of the trachea in scleroderma may be an extension of the fibrotic disease that occurs at the pulmonary level. However, it is worth mentioning that in our study, patients without pulmonary involvement showed higher values in tracheal volumetry than control subjects. This finding suggests that factors other than pulmonary fibrosis are involved in the tracheal abnormalities that occur in SSc patients. Consistent with this observation, several studies have shown impairment of anatomic structures above the trachea in scleroderma patients; laryngeal dysfunction is relatively common, with pathologic findings demonstrating fibrinoid degeneration and an increase in collagen fibers [30,31]. Considering the promising advent of user-friendly software, evaluation of the trachea in patients with scleroderma may be an interesting approach both in clinical practice and in trials. Thus, precise characterization of the trachea in scleroderma may offer an additional tool for the follow-up of these patients and may be helpful in the evaluation of clinical treatment.

The presence of obstructive abnormalities in the cervical trachea can be detected by the flowvolume loop of spirometry even when there is no clinical suspicion, with the most used index being the ratio  $\text{FEF}_{50\%}/\text{FIF}_{50\%}$  [32]. In the present study, 25% of our patients had an  $\text{FEF}_{50\%}/$  $FIF_{50\%} > 1.50$ , and we found negative correlations between  $FEF_{50\%}/FIF_{50\%}$  and several indices measured by CT trachea volumetry, including area, perimeter, equivalent diameter, major diameter, and minor diameter. Consistent with our findings, Miranda et al. [33] used the forced oscillation technique in patients with scleroderma and found an increase in mean resistance, a parameter that reflects changes in the most central airways. Another study using impulse oscillometry observed an increase in the resistive and reactive properties of the respiratory system in patients with scleroderma, and these alterations were correlated with the findings of fibrosis in CT [34]. Interestingly, we observed an association between tracheal tortuosity and PEF (which reflects flow through large airways), supporting the notion that tracheal deviation is a key contributor to the reduction of airflow in the large airways of patients with scleroderma. However, quantitative measurement of eccentricity was not correlated with FEF<sub>50%</sub>/FIF<sub>50%</sub>. One possible explanation for this finding is that increased tracheal area in scleroderma patients may at least partially counterbalance the effects of reduced inspiratory flow in these patients.

In our study, patients with extensive ILD displayed greater tracheal area than patients without pulmonary involvement and patients with limited ILD; all of these patients, in turn, displayed greater tracheal area than the control subjects. Several recent studies have shown that larger esophageal diameter is associated with higher pulmonary fibrosis and worse lung function in

individuals with scleroderma [35,36]. Since microaspiration secondary to gastro-esophageal reflux is associated with the progression of pulmonary fibrosis in scleroderma [37,38], we think that the tracheal pathology may also contribute to this phenomenon. Longitudinal and controlled studies of larger numbers of patients are required to evaluate whether or not tracheal pathology is involved in the progression of lung fibrosis in patients with scleroderma.

In the present study, the mean value of FVC was higher in the control group and lower in the scleroderma group with extensive ILD and showed an intermediate value in the scleroderma group with no pulmonary involvement (or limited ILD) ( $102.4 \pm 19.8\%$  vs.  $85 \pm 19.3\%$  vs.  $68 \pm 21.5\%$  predicted, p = 0.005). Although the restrictive pattern in scleroderma is largely explained by the presence of ILD, a reduced compliance of the respiratory system may also be due to chest wall tightening from skin thickening, pleural disease, cardiac involvement, and respiratory muscle weakness [39-41]. Since no significant difference was observed in FVC between the control group and patients without pulmonary involvement in CT, we believe that ILD is the main contributor to the restrictive damage in our sample. However, we observed a significant correlation between FVC and the quantitative measurement of tracheal eccentricity. A possible explanation for this association is that overproduction of collagen and deposition of connective tissue, which are primary pathophysiological mechanisms of SSc, cause both abnormal tracheal geometry and reduced lung volume in these patients.

In scleroderma, cutaneous involvement occurs due to the increased thickness and hardness of the skin, which leads to shrinking of the skin in deeper structures; although cutaneous involvement is usually most prominent on the face and hands, these abnormalities may extend to the upper chest [42,43]. Some studies have shown that subclinical involvement of the upper chest is detectable by high-frequency ultrasound even with normal palpation [44,45]. In our study, the mRSS (a key measure in the clinical evaluation of patients with scleroderma) was associated with the eccentricity values provided by CT trachea volumetry. Since eccentricity measures the deviation of a conical structure in relation to its circumference [46], we think that skin thickening that occurs at the neck level may negatively impact the geometry of the trachea in patients with scleroderma [47]. Consistent with our findings, Kim et al. [48] found an association between the fibrosis score detected by CT using a computer-assisted method and the mRSS of patients with scleroderma both at baseline and during a 12-month follow-up period. This reinforces the idea that the measure of eccentricity may be a marker of the severity of the process of collagen hyalinization and abnormalities in the elastic tissues that surround the neck and may extend to the intrathoracic structures of patients with scleroderma [42]. Considering that future directions for the management of patients with scleroderma, including epigenetic modulation and antifibrotic or biological therapy, are being discussed [49,50], we think that the parameters provided by the measurement of tracheal geometry can contribute to the evaluation of the outcomes.

The identification of reliable and consistent biomarkers that can be used to predict the course of specific diseases is necessary for better stratification and management of patients and would be of great use in the treatment of a multifaceted disease such as scleroderma [3]. In this context, several attempts have been made in recent years to correlate possible serum biomarkers with clinical features distinct from scleroderma. Several investigators have shown an association between ATA and the risk of development and progression of pulmonary fibrosis in patients with scleroderma [51,52]. A recent study has shown that topoisomerase I peptide-loaded dendritic cells induce not only the autoantibody response but also cutaneous and pulmonary fibrosis [52]. In the present study, we observed a positive correlation between the presence of ATA and tracheal tortuosity. This significant correlation must be viewed cautiously since we did not evaluate predicting values. However, we think that this finding could serve as a starting point for longitudinal studies designed to evaluate a possible contribution of ATA to the management of tracheal pathology in patients with scleroderma.

A critical analysis of the limitations of the present study is pertinent. First, the sample size is small, and the sample is representative of only one center. Second, the sample is composed predominantly of scleroderma patients with associated ILD. Although we observed significant differences in the abnormalities of trachea volumetry between control subjects and patients without pulmonary involvement on CT, a greater number of patients in this group could allow for more robust conclusions. Third, CT pulmonary densitovolumetry could have aided in the study of correlations between tracheal changes and those observed in the lungs and lower airways. Fourth, an assessment comparing patients with ILD-SSc with patients with ILD due to other causes (e.g., idiopathic pulmonary fibrosis, rheumatoid arthritis-ILD) might better define the role of scleroderma in the development of tracheal abnormalities. Finally, the evaluation of other biomarkers could have helped us better understand the tracheal disease in scleroderma and might have opened new horizons in the field of precision medicine. In fact, new laboratory markers (including TGF $\beta$ 1, IL-6, sPD-1, sPD-L2, and CXCL4) have been associated with higher mRSS score and more pronounced changes in thoracic CT [3]. Despite these limitations, the quantitative analysis of the trachea through computer software offers a discriminant method that can help produce an objective measure and obtain prognostic information in scleroderma.

In conclusion, the present study shows that, in a sample composed predominantly of scleroderma patients with associated ILD, there were abnormalities in the geometry of the trachea in eccentricity, diameter, and tortuosity. In these patients, abnormalities in the geometry of the trachea were associated with markers of functional obstruction at the level of the cervical trachea. In addition, the measurement of tracheal tortuosity was correlated with cutaneous involvement, the degree of pulmonary fibrosis, and the presence of ATA. Although the encouraging data presented here require further validation in prospective studies, we believe that skeletonization and CT trachea volumetry may improve the ability of radiologists and rheumatologists to accurately assess the tracheas of patients with scleroderma both in clinical practice and in trials.

### **Author Contributions**

- **Conceptualization:** Bruno Rangel Antunes Silva, Rosana Souza Rodrigues, Rogério Rufino, Cláudia Henrique Costa, Veronica Silva Vilela, Roger Abramino Levy, Alan Ranieri Medeiros Guimarães, Alysson Roncally Silva Carvalho, Agnaldo José Lopes.
- Data curation: Bruno Rangel Antunes Silva.
- Formal analysis: Alan Ranieri Medeiros Guimarães, Alysson Roncally Silva Carvalho, Agnaldo José Lopes.

Funding acquisition: Agnaldo José Lopes.

- Methodology: Bruno Rangel Antunes Silva, Rogério Rufino, Cláudia Henrique Costa, Veronica Silva Vilela, Roger Abramino Levy, Alan Ranieri Medeiros Guimarães, Alysson Roncally Silva Carvalho, Agnaldo José Lopes.
- **Resources:** Rogério Rufino, Cláudia Henrique Costa, Alysson Roncally Silva Carvalho, Agnaldo José Lopes.
- Software: Alan Ranieri Medeiros Guimarães, Alysson Roncally Silva Carvalho.
- Supervision: Rosana Souza Rodrigues, Cláudia Henrique Costa, Roger Abramino Levy, Alysson Roncally Silva Carvalho, Agnaldo José Lopes.

Validation: Alan Ranieri Medeiros Guimarães, Alysson Roncally Silva Carvalho.

Writing – original draft: Bruno Rangel Antunes Silva, Alysson Roncally Silva Carvalho, Agnaldo José Lopes.

Writing – review & editing: Bruno Rangel Antunes Silva, Rosana Souza Rodrigues, Rogério Rufino, Cláudia Henrique Costa, Veronica Silva Vilela, Roger Abramino Levy, Alan Ranieri Medeiros Guimarães, Alysson Roncally Silva Carvalho, Agnaldo José Lopes.

#### References

- van den Hoogen F, Khanna D, Fransen J, Johnson SR, Baron M, Tyndall A, et al. 2013 classification criteria for systemic sclerosis: an American College of Rheumatology/European League Against Rheumatism collaborative initiative. Arthritis and Rheumatism. 2013; 65(11):2737–47. https://doi.org/10.1002/ art.38098 PMID: 24122180; PubMed Central PMCID: PMC3930146.
- Salaffi F, Carotti M, Di Donato E, Di Carlo M, Ceccarelli L, Giuseppetti G. Computer-aided tomographic analysis of interstitial lung disease (ILD) in patients with systemic sclerosis (SSc): correlation with pulmonary physiologic tests and patient-centred measures of perceived dyspnea and functional disability. PLoS One. 2016; 11(3):e0149240. https://doi.org/10.1371/journal.pone.0149240 PMID: 26930658; PubMed Central PMCID: PMC4773230.
- Barsotti S, Bruni C, Orlandi M, Della Rossa A, Marasco E, Codullo V, et al. One year in review 2017: systemic sclerosis. Clinical and Experimental Rheumatology. 2017; 35(Suppl 106 4):3–20. PMID: 29035173.
- Barnes J, Mayes MD. Epidemiology of systemic sclerosis: incidence, prevalence, survival, risk factors, malignancy, and environmental triggers. Current Opinion in Rheumatology. 2012; 24(2):165–70. https://doi.org/10.1097/BOR.0b013e32834ff2e8 PMID: 22269658.
- Sticherling M. Systemic sclerosis-dermatological aspects. Part 1: Pathogenesis, epidemiology, clinical findings. Journal der Deutschen Dermatologischen Gesellschaft. 2012; 10(10):705–18. https://doi.org/ 10.1111/j.1610-0387.2012.07999.x PMID: 22913330.
- Silva BRA, Rufino R, Costa CH, Vilela VS, Levy RA, Lopes AJ. Ventilation distribution and small airway function in patients with systemic sclerosis. Portuguese Journal of Pulmonology. 2017; 23(3):132–8. https://doi.org/10.1016/j.rppnen.2017.01.004 PMID: 28258938.
- Ooi GC, Mok MY, Tsang KWT, Wong Y, Khong PL, Fung PCW, et al. Interstitial lung disease in systemic sclerosis: an HRCT-clinical correlative study. Acta Radiologica. 2003; 44(3):258–64. PMID: 12751995
- Kim HG, Tashkin DP, Clements PJ, Li G, Brown MS, Elashoff R, et al. A computer-aided diagnosis system for quantitative scoring of extent of lung fibrosis in scleroderma patients. Clinical and Experimental Rheumatology. 2010; 28(5 Suppl 62):S26–35. PMID: 21050542; PubMed Central PMCID: PMC3177564.
- Ariani A, Carotti M, Gutierrez M, Bichisecchi E, Grassi W, Giuseppetti GM, et al. Utility of an opensource DICOM viewer software (OsiriX) to assess pulmonary fibrosis in systemic sclerosis: preliminary results. Rheumatology International. 2014; 34(4):511–6. https://doi.org/10.1007/s00296-013-2845-6 PMID: 23949623.
- Camilo GB, Carvalho ARS, Guimarães ARM, Kasuki L, Gadelha MR, Mogami R, et al. Computed tomography airway lumen volumetry in patients with acromegaly: Association with growth hormone levels and lung function. Journal of Medical Imaging and Radiation Oncology. 2017; 61(5):591–9. https:// doi.org/10.1111/1754-9485.12598 PMID: 28217888.
- Williamson JP, James AL, Phillips MJ, Sampson DD, Hillman DR, Eastwood PR. Quantifying tracheobronchial tree dimensions: methods, limitations and emerging techniques. The European Respiratory Journal. 2009; 34(1):42–55. https://doi.org/10.1183/09031936.00020408 PMID: 19567601.
- Yamashiro T, Tsubakimoto M, Nagatani Y, Moriya H, Sakuma K, Tsukagoshi S, et al. Automated continuous quantitative measurement of proximal airways on dynamic ventilation CT: initial experience using an ex vivo porcine lung phantom. International Journal of Chronic Obstructive Pulmonary Disease. 2015; 10:2045–54. <u>https://doi.org/10.2147/COPD.S87588</u> PMID: <u>26445535</u>; PubMed Central PMCID: PMC4590570.
- Zhang C, Wang H, Cao J, Li C, Mi W, Yang L, et al. Measurement and analysis of the tracheobronchial tree in Chinese population using computed tomography. PLoS One. 2015; 10(6):e0130239. https://doi. org/10.1007/s00117-003-0990-8
- Sorantin E, Halmai C, Erdohelyi B, Palágyi K, Nyúl LG, Ollé K, et al. 3D cross section of the laryngotracheal tract: a new method for visualization and quantification of tracheal stenosis. Radiologe. 2003; 43 (12):1056–68. https://doi.org/10.1007/s00117-003-0990-8 PMID: 14668994.
- Yamashiro T, Moriya H, Tsubakimoto M, Matsuoka S, Murayama S. Continuous quantitative measurement of the proximal airway dimensions and lung density on four-dimensional dynamic-ventilation CT in smokers. International Journal of Chronic Obstructive Pulmonary Disease. 2016; 11:755–64. https://doi.org/10.2147/COPD.S100658 PMID: 27110108; PMCID Central PMC4835141.

- LeRoy EC, Black C, Fleischmajer R, Jablonska S, Krieg T, Medsger TA Jr, et al. Scleroderma (systemic sclerosis): classification, subsets and pathogenesis. The Journal of Rheumatology. 1988; 15(2):202–5. PMID: 3361530.
- Clements P, Lachenbruch P, Siebold J, White B, Weiner S, Martin R, et al. Inter and intraobserver variability of total skin thickness score (modified Rodnan TSS) in systemic sclerosis. The Journal of Rheumatology. 1995; 22(7):1281–5. PMID: 7562759.
- Culver BH, Graham BL, Coates AL, et al. Recommendations for a standardized pulmonary function report. An official American Thoracic Society technical statement. American Journal of Respiratory and Critical Care Medicine. 2017; 196(11):1463–72. https://doi.org/10.1164/rccm.201710-1981ST PMID: 29192835.
- Pereira CAC, Sato T, Rodrigues SC. New reference values for forced spirometry in white adults in Brazil. Jornal Brasileiro de Pneumologia. 2007; 33(4):397–406. https://doi.org/10.1164/rccm.200706-877OC
- Neder JA, Andreoni S, Peres C, Nery LE. Reference values for lung function tests. III. Carbon monoxide diffusing capacity (transfer factor). Brazilian Journal of Medical and Biological Research. 1999; 32 (6):729–37. https://doi.org/10.1590/S0100-879X1999000600008 PMID: 10412551.
- Morewood DJ, Belchetz PE, Evans CC, Whitehouse GH. The extrathoracic airway in acromegaly. Clinical Radiology. 1986; 37(3):243–6. PMID: <u>3709048</u>.
- Goh NS, Desai SR, Veeraraghavan S, Hansell DM, Copley SJ, Maher TM, et al. Interstitial lung disease in systemic sclerosis: a simple staging system. American Journal of Respiratory and Critical Care Medicine. 2008; 177(11):1248–54. https://doi.org/10.1164/rccm.200706-877OC PMID: 18369202.
- Le Gouellec N, Duhamel A, Perez T, Hachulla AL, Sobanski V, Faivre JB, et al. Predictors of lung function test severity and outcome in systemic sclerosis-associated interstitial lung disease. PLoS One. 2017; 12(8):e0181692. https://doi.org/10.1371/journal.pone.0181692 PMID: 28763468; PubMed Central PMCID: PMC5538660.
- Lopes AJ, Capone D, Mogami R, Lanzillotti RS, Melo PL, Jansen JM. Severity classification for idiopathic pulmonary fibrosis by using fuzzy logic. Clinics (Sao Paulo). 2011; 66(6):1015–9. https://doi.org/ 10.1590/S1807-59322011000600016 PMID: 21808868; PubMed Central PMCID: PMC3129967.
- 25. Khanna D, Nagaraja V, Tseng CH, Abtin F, Suh R, Kim G, et al. Predictors of lung function decline in scleroderma-related interstitial lung disease based on high-resolution computed tomography: implications for cohort enrichment in systemic sclerosis-associated interstitial lung disease trials. Arthritis Research and Therapy. 2015; 17:372. https://doi.org/10.1186/s13075-015-0872-2 PMID: 26704522; PubMed Central PMCID: PMC4718035.
- Fedorov A, Beichel R, Kalpathy-Cramer J, Finet J, Fillion-Robin J-C, Pujol S, et al. 3D Slicer as an image computing platform for the quantitative imaging network. Magnetic Resonance Imaging. 2012; 30(9):1323–41. https://doi.org/10.1016/j.mri.2012.05.001 PMID: 22770690
- Zámolyi A, Székely B, Draganits E, Timár G. Neotectonic control on river sinuosity at the western margin of the Little Hungarian Plain. Geomorphology 2010; 122: 231–43. <u>https://doi.org/10.1155/2015/834262</u>; PubMed Central PMCID: PMC4299560.
- Salaffi F, Carotti M, Bosello S, Ciapetti A, Gutierrez M, Bichisecchi E, et al. Computer-aided quantification of interstitial lung disease from high resolution computed tomography images in systemic sclerosis: correlation with visual reader-based score and physiologic tests. BioMed Research International. 2015; 2015:834262. https://doi.org/10.1155/2015/834262 PMID: 25629053; PubMed Central PMCID: PMC4299560.
- Bouros D, Wells AU, Nicholson AG, Colby TV, Polychronopoulos V, Pantelidis P, et al. Histopathologic subsets of fibrosing alveolitis in patients with systemic sclerosis and their relationship to outcome. American Journal of Respiratory and Critical Care Medicine. 2002; 165(12):1581–6. https://doi.org/10.1164/ rccm.2106012 PMID: 12070056.
- Pepper JP, Kupfer RA, McHugh JB, Hogikyan ND. Histopathologic findings and clinical manifestations in a patient with dysphonia and vocal fold involvement by systemic sclerosis. Archives of Otolaryngology Head & Neck Surgery. 2011; 137(8):816–9. <u>https://doi.org/10.5114/aoms.2016.57582</u>; PubMed Central PMCID: PMC4754368.
- Ramos HV, Pillon J, Kosugi EM, Fujita R, Pontes P. Laryngeal assessment in rheumatic disease patients. Brazilian Journal of Otorhinolaryngology. 2005; 71(4):499–503. S0034-72992005000400017 PMID: 16446967.
- Camilo GB, Guimarães FS, Mogami R, Faria AC, Melo PL, Lopes AJ. Functional changes are associated with tracheal structural abnormalities in patients with acromegaly. Archives of Medical Science. 2016; 12(1):78–88. https://doi.org/10.5114/aoms.2016.57582 PMID: 26925121; PubMed Central PMCID: PMC4754368.
- Miranda IA, Dias Faria AC, Lopes AJ, Jansen JM, Lopes de Melo P. On the respiratory mechanics measured by forced oscillation technique in patients with systemic sclerosis. PLoS One 2013; 8(4):e61657. https://doi.org/10.1371/journal.pone.0061657 PMID: 23637877; PubMed Central PMCID: PMC3637442.

- Aronsson D, Hesselstrand R, Bozovic G, Wuttge DM, Tufvesson E. Airway resistance and reactance are affected in systemic sclerosis. European Clinical Respiratory Journal. 2015; 2:28667. https://doi. org/10.3402/ecrj.v2.28667 PMID: 26672963; PubMed Central PMCID: PMC4653312.
- 35. Winstone TA, Hague CJ, Soon J, Sulaiman N, Murphy D, Leipsic J, et al. Oesophageal diameter is associated with severity but not progression of systemic sclerosis-associated interstitial lung disease. Respirology. 2018. [Epub ahead of print]. https://doi.org/10.1111/resp.13309 PMID: 29641847.
- Richardson C, Agrawal R, Lee J, Almagor O, Nelson R, Varga J, et al. Esophageal dilatation and interstitial lung disease in systemic sclerosis: a cross-sectional study. Seminars in Arthritis and Rheumatism. 2016; 46(1):109–14. https://doi.org/10.1016/j.semarthrit.2016.02.004 PMID: 27033049; PubMed Central PMCID: PMC5500283.
- Savarino E, Ghio M, Marabotto E, Zentilin P, Sammito G, Cittadini G, et al. Possible connection between gastroesophageal reflux and interstitial pulmonary fibrosis in patients with systemic sclerosis. Recenti Progressi in Medicina. 2009; 100(11):512–6. PMID: 20066883.
- Lock G, Pfeifer M, Straub RH, Zeuner M, Lang B, Schölmerich J, et al. ssociation of esophageal dysfunction and pulmonary function impairment in systemic sclerosis. The American Journal of Gastroenterology. 1998; 93(3):341–5. https://doi.org/10.1111/j.1572-0241.1998.00341.x PMID: 9517636.
- van Laar JM, Stolk J, Tyndall A. Scleroderma lung: pathogenesis, evaluation and current therapy. Drugs. 2007; 67(7):985–96. PMID: 17488144.
- Farrokh D, Abbasi B, Fallah-Rastegar Y, Mirfeizi Z. The extrapulmonary manifestations of systemic sclerosis on chest high resolution computed tomography. Tanaffos. 2015; 14(3):193–200. PMID: 26858765; PubMed Central PMCID: PMC4745188.
- Lopes AJ, Justo AC, Ferreira AS, Guimaraes FS. Systemic sclerosis: association between physical function, handgrip strength and pulmonary function. Journal of Bodywork and Movement Therapies. 2017; 21(4): 972–77. https://doi.org/10.1016/j.jbmt.2017.03.018 PMID: 29037654.
- 42. Hasan O, Jessar M, Ashar M, Noordin S, Ahmad T. Systemic sclerosis: clinical manifestations, anesthetic and orthopedic considerations in a patient. International Journal of Surgery Case Reports. 2017; 42:24–8. https://doi.org/10.1016/j.ijscr.2017.11.051 PMID: 29207307; PubMed Central PMCID: PMC5724744.
- Steen VD. Clinical manifestations of systemic sclerosis. Seminars in Cutaneous Medicine and Surgery. 1998; 17(1):48–54. PMID: 9512107.
- 44. Sulli A, Ruaro B, Smith V, Paolino S, Pizzorni C, Pesce G, et al. Subclinical dermal involvement is detectable by high frequency ultrasound even in patients with limited cutaneous systemic sclerosis. Arthritis Research and Therapy. 2017; 19(1): 61. https://doi.org/10.1186/s13075-017-1270-8 PMID: 28320447; PubMed Central PMCID: PMC5360023.
- Hesselstrand R, Scheja A, Wildt M, Akesson A. High-frequency ultrasound of skin involvement in systemic sclerosis reflects oedema, extension and severity in early disease. Rheumatology. 2008; 47 (1):84–7. https://doi.org/10.1093/rheumatology/kem307 PMID: 18077496.
- 46. Yue Y, Fan Z, Yang W, Pang J, Deng Z, McKenzie E, et al. Geometric validation of self-gating k-spacesorted 4D-MRI vs 4D-CT using a respiratory motion phantom. Medical Physics. 2015; 42(10):5787–97. https://doi.org/10.1118/1.4929552 PMID: 26429253; PubMed Central PMCID: PMC4575318.
- Adnan ZA. Diagnosis and treatment of scleroderma. Acta Medica Indonesiana. 2008; 40(2):109–12. PMID: 18560030.
- Kim HJ, Tashkin DP, Gjertson DW, Brown MS, Kleerup E, Chong S, et al. Transitions to different patterns of interstitial lung disease in scleroderma with and without treatment. Annals of the Rheumatic Diseases. 2016; 75(7):1367–71. https://doi.org/10.1136/annrheumdis-2015-208929 PMID: 26757749.
- O'Reilly S. Epigenetic modulation as a therapy in systemic sclerosis. Rheumatology. 2018. [Epub ahead of print]. https://doi.org/10.1093/rheumatology/key071 PMID: 29579252.
- George PM, Wells AU. Disease staging and sub setting of interstitial lung disease associated with systemic sclerosis: impact on therapy. Expert Review of Clinical Immunology. 2018; 14(2):127–135. https://doi.org/10.1002/art.39166
- Hoffmann-Vold AM, Aaløkken TM, Lund MB, Garen T, Midtvedt Ø, Brunborg C, et al. Predictive value of serial high-resolution computed tomography analyses and concurrent lung function tests in systemic sclerosis. Arthritis & Rheumatology. 2015; 67(8):2205–12. https://doi.org/10.1002/art.39166 PMID: 25916462.
- Mehta H, Goulet PO, Nguyen V, Pérez G, Koenig M, Senécal JL, et al. Topoisomerase I peptide-loaded dendritic cells induce autoantibody response as well as skin and lung fibrosis. Autoimmunity. 2016; 49 (8):503–13. https://doi.org/10.1080/08916934.2016.1230848 PMID: 27808577.