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OPEN Application of iridium(III) complex in label-free and non-enzymatic electrochemical detection of hydrogen peroxide based on a novel "on-off-on" switch platform

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We herein report a label-free and non-enzymatic electrochemical sensor for the highly sensitive detection of hydrogen peroxide (H₂O₂) based on a novel "on-off-on" switch system. In our design, MB was used as an electron mediator to accelerate the electron transfer while AuNPs was used to amplify the electrochemical signal due to its excellent biocompatibility and good conductivity. The "switch-off" state was achieved by introducing the guanine-rich capture probe (CP) and an iridium complex onto the electrode surface to form a hydrophobic layer, which then hinders electron transfer. Upon addition of H_2O_2 , fenton reaction occurs and produces OH in the presence of Fe^{2+} . The OH cleaves the CP into DNA fragments, thus resulting in the release of CP and iridium complex from the sensing interface, recovering the electrochemical signal to generate a "switch-on" state. Based on this novel switch system, a detection limit as low as 3.2 pM can be achieved for H₂O₂ detection. Moreover, satisfactory results were obtained by using this method for the detection of H₂O₂ in sterilized milk. To the best of our knowledge, this is the first G-quadruplex-based electrochemical sensor using an iridium(III) complex.

Hydrogen peroxide (H_2O_2) is not only an important compound in food, bioanalysis and environmental analyses¹, but also plays an important role in cellular reactions catalyzed by many oxidases. Moreover, H₂O₂ is also involved in the function and signal transduction of the cell^{2,3}. Thus, the sensitive and simple detection of H_2O_2 is of great interest in chemical, biological, clinical and environmental fields. In the past decades, several kinds of strategies such as electrochemical⁴⁻⁷, fluorescent⁸⁻¹⁰ and colorimetric¹¹⁻¹³ methods have been developed for the detection of H₂O₂. Among these methods, electrochemical sensing has attracted great attention due to its merits such as high sensitivity, rapid response, and simple operation.

Enzyme-based detection methods for H_2O_2 have been developed due to number of the advantages of enzymatic reactions, including catalytic activity, high efficiency and good selectivity¹⁴. Unfortunately, natural enzymes are expensive and easily denatured by environmental changes, thus the enzymes used in these sensing methods are susceptible to potential interfering substances^{15,16}. Therefore, non-enzymatic electrochemical sensors have received great attention in the development of H₂O₂ sensors. Imlay, Linn and co-workers reported that the antibacterial property of H₂O₂ is due to DNA damage resulting from Fenton-like reactions in the presence of metal ions such as Fe^{2+i7} , it was reported that H_2O_2 could react with certain transition metal ions in low oxidation states and thus produce HO, which may cleave single-stranded DNA (ssDNA) to DNA fragments¹⁸. Based on this principle, many biosensors have been developed for biomolecular detection¹⁹⁻²¹.

Guanine-rich (G-rich) nucleic acid sequences, existing in the region of gene promoter and chromosome telomeres, can fold into a unique G-quadruplex structure in the presence of metal ion, such as potassium ion (K⁺)^{22,23}. Based on these characteristics, many G-quadruplex-based sensors have been developed for the detection of metal ions²⁴⁻²⁶, DNA^{27,28}, small molecules^{29,30} and protein biomarkers³¹. Such K⁺-mediated

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Figure 1. Scheme diagram for the preparation of H₂O₂ sensor.

G-quadruplex-based sensing platforms are convenient, low-cost and relatively quick. Previously, our group has synthesized different iridium(III) complexes and discovered that some of them have highly specific binding properties to G-quadruplex, and a number of detection platforms were subsequently constructed based on the specific interaction between iridium(III) complexes and G-quadruplex DNA³²⁻³⁷, through detecting the luminescent enhancement of iridium(III) complexes.

Recently, we discovered that some of the iridium(III) complexes can hinder the electron transfer between the electrode surface and the electrolyte mainly because of the hydrophobicity and the large molecular structure of them. Based on of this unique property, we designed the application of one type of the iridium(III) that specific to G-quadruplex (the structure of it were shown in Fig. 1) in label-free and non-enzymatic electrochemical sensing platform for H_2O_2 by using a novel "on-off-on" switch system. As shown in Fig. 1, the first "switch on" state was achieved based on the immobilize of Nafion (Nf) and methylene blue (Nf@MB) composite membrane as inner-layer, and self-assembled positively charged gold nanoparticles ((+)AuNPs) as outer-layer. Successively, the capture probe (CP, guanine-rich nucleic acid) was modified onto the (+)AuNPs surface through an Au-S bond, and the CP would change to G-quadruplex structure in the presence of K⁺, followed by the specific binding with an iridium(III) complex using a simple and label-free method³⁸⁻⁴⁴. As a consequence, a hydrophobic and large molecule structural layer wad formed on the electrode surface, and resulted in a "switch off" state. Then, upon incubation of the sensor with H_2O_2 and Fe^{2+} , Fenton reaction would happen and produced tremendous OH• to cleave CP into DNA fragments, leading to the release of such fragments and the iridium(III) complex from the sensing interface, recovering the electrochemical signal and achieving a "switch on" state. Importantly, the electrochemical signal change was proportional to H2O2 concentration. Thus, H2O2 could be detected by using such proposed method based on monitoring the signal change of MB. There are several advantages for such a proposed method: Firstly, due to the good stability of negatively charged Nf membrane on the electrode surface, abundant positively charged and highly stable Nf@MB membrane could formed on the electrode, which can greatly improve the stability of the sensor. In addition, MB is highly conductive as a redox indicator, which can great enhance the sensitivity of the sensor. Meantime, about 4 nm of (+)AuNPs, with excellent biocompatibility and good conductivity, was successfully self-assembly onto the Nf@MB membrane, which can greatly enlarge the electrode surface for more CP immobilization. Moreover, to the best of our knowledge, this is the first application of the iridium(III) complex and (+)AuNPs in electrochemical sensor for H₂O₂ analysis. We envision that this "on-off-on" platform could also provide new opportunities for biosensor development.

Results and Discussion

To investigate the conductivity of the iridium(III) complex, several modified electrodes were characterized by cyclic voltammetry experiments (CVs) in the presence of $5 \text{ mM Fe}(\text{CN})_6^{3-/4-}$. As seen from curve a in Fig. 2A, a stable and well-defined redox peak was obtained when the bare gold electrode was scanned in $5 \text{ mM of Fe}(\text{CN})_6^{3-/4-}$. After the immobilization of CP on the electrode surface, an obvious decrease of the peak current was observed, which was attributed to the fact that DNA hinders electron transfer (curve b)^{45,46}. Then, G-quadruplex structures were formed after the CP-modified electrode was incubated in tris-buffer containing 100 mM of K⁺. Subsequently, the interaction of iridium(III) complex with the G-quadruplex led to a further decrease in peak current, because of the high resistance of the electrode interface induced by the adsorption of iridium(III) complex and the formation of a hydrophobic layer on the electrode surface (curve c). Since other



Figure 2. (A) Conductivity investigation of the iridium(III) complex and other organic dyes that specific to G-quadruplex in 5 mM of Fe(CN)^{3-/4-}: (a) Gold electrode; (b) CP/gold electrode; (c) iridium(III) complex/CP/ gold electrode; (d) CV/CP/gold electrode. (e) Rhodamine/CP/gold electrode. (B) DPV of the different modified electrodes in 10 mM tris-buffer solution (pH 7.4): (a) Gold electrode; (b) Nf@MB/gold electrode; (c) (+) AuNPs/Nf@MB/gold electrode; (d) iridium(III) complex/CP/(+)AuNPs/Nf@MB/gold electrode; (e) (d) after the incubation with 1.5 nM of H₂O₂. (C) Circular dichroism of CP before (a) and after (b) the incubation of it with tris-buffer that contained 100 mM of K⁺. (D) Circular dichroism of G-quadruplex structure before (a) and after (b) the selective interaction of it with iridium(III) complex, and after the cleavage with H₂O₂ (c).

organic dyes such as crystal violet (CV) and rhodamine can also specifically bind to G-quadruplex^{47,48}, we also investigated the conductivity of CV and rhodamine after they bound to the G-quadruplex that immobilized on the electrode. The results demonstrated that CV and rhodamine would accelerate the electron transfer (curve d and e). Therefore, to obtain the step of the "switch off" state, we selected iridium(III) complex as the hydrophobic layer.

In order to confirm the successful fabrication of the proposed sensing platform, the fabrication process of the sensor was characterized by DPV experiments. As shown in Fig. 2B, no oxidation peak was observed at the bare gold electrode while an obvious oxidation peak appeared after electro-polymerization of MB on the electrode surface, which is mainly due to the high conductivity of MB as an electron mediator (curve b). Then, the peak current increased after the electro-deposition of AuNPs onto the MB film on account of the excellent conductivity of AuNPs (curve c). However, after the interaction of iridium(III) complex with G-quadruplex form of CP in the presence of K⁺, the peak current obviously decreased, since iridium(III) complex would form a hydrophobic and large molecule structural layer on the electrode surface and hinder the electron transfer (curve d). However, upon incubation of the sensor with 1.5 nM of H_2O_2 and Fe^{2+} , a dramatic increase of the peak current was observed (curve e). This increase could be ascribed to the release of iridium(III) complex and CP fragments from the electrode surface after the cleavage of CP by Fenton reaction in the presence of Fe^{2+} and H_2O_2 .

To validate the mechanism of the assay, circular dichroism (CD) spectroscopy was performed. As shown in Fig. 2C, there was no obvious peak for CP alone, while a positive Cotton effect peak at around 264 nm and a negative Cotton effect peak at around 236 nm appeared after CP was incubated with tris-buffer containing 100 mM of K⁺, indicating the formation of the classic G-quadruplex structure in accordance with literature⁴⁹. No significant change was observed upon the selective interaction of the iridium(III) complex with the G-quadruplex structure of CP (Fig. 2D, curve a and b). However, the intensity at the peak of 264 nm and 236 nm decreased dramatically when the CP was incubated with H₂O₂, these results might be attributed to the cleavage of CP (curve c).

Experimental conditions including the self-assembly time of (+)AuNPs, the concentration of Fe²⁺ and the cleavage time of CP by H₂O₂ were optimized. The self-assembly time of (+)AuNPs is an important factor that might affect the properties of the sensor. As shown in Fig. 3A, the current intensity was proportional to the self-assembly time of (+)AuNPs from 0 to 30 min and then reached a plateau. In view of the sensitivity of the sensor, an electro-deposition time of 30 min was chosen for all the experiments.

The role of Fe^{2+} in the Fenton reaction is to catalyze the cleavage of DNA. Thus, Fe^{2+} concentration is an important factor that can affect the degree of CP cleavage. Figure 3B shows that the current signal increased along with the increase of Fe^{2+} concentration up to $1.5 \,\mu$ M in the presence of $1.2 \,n$ M H₂O₂, indicating that CP could be



Figure 3. Effect of the self-assembly time of (+)AuNPs on the sensor properties (**A**) Effect of the concentration of Fe²⁺ (**B**), the pH value (**C**) and the incubation time for CP cleavage in the presence of $1.2 \text{ nM H}_2\text{O}_2$ (**D**).

cleaved effectively upon addition of $1.5 \,\mu$ M of Fe²⁺. Thus, we selected $1.5 \,\mu$ M as the optimal concentration of Fe²⁺ in the experiments.

DNA structure depends on the pH environment, so the cleavage activity of H_2O_2 and Fe^{2+} may depend on the pH as well. As shown in Fig. 3C, the current signal increased with the pH over the range from 5.0 to 6.5 in the absence of H_2O_2 , and reached a plateau over the range of 6.5–7.4. However, when the pH was higher than 7.4, the current signal decreased dramatically, such result might be due to the fact that an acidic environment can stabilize the Fe²⁺ions, and higher pH can induce the oxidation of ferrous ions to ferric ions under atmosphere (O₂). Considering the sensitivity of the sensor, we selected 7.4 for further experiments.

Since the second "switch-on" state depends on the cleavage of CP, the cleavage time of CP in the presence of $1.2 \text{ nM H}_2\text{O}_2$ was investigated in Fig. 3D. It was found that the current signal increased when the cleavage time increased from 0 to 25 min, and then reached a plateau. To achieve the effective cleavage of CP, 30 min was chosen as the cleavage time of CP.

To evaluate the effect of H_2O_2 concentration on current signal, the proposed sensor was incubated with tris-buffer that contained different concentration of H_2O_2 . As shown in Fig. 4A, the DPV signal increased along with the increase of H_2O_2 concentration. According to the calibration plot of Fig. 4B, the linear regression equation was I = 11.18 + 7.87c (c: nM) in a dynamic range from 8.0 pM to 2.0 nM with a correlation coefficient of $R^2 = 0.994$. The detection limit was 3.2 pM calculated by the three-signal method. Here, the electro-deposition of AuNPs onto MB film can greatly enlarge the electrode surface for CP immobilization, and accordingly improve the sensitivity of the sensor. The detection limit of our "on-off-on" detection platform for H_2O_2 detection was lower than other AuNPs^{49,50–53} or MB^{54–57} based electrochemical methods, which can satisfy the demand for H_2O_2 detection in real samples such as food or industrial products. Thus, our proposed sensor could be used to quantify H_2O_2 for food safety and clinical diagnosis.

The stability of the sensor was investigated by CV. After continuous scanning for 20 cycles, the sensor retained 98.1% of its initial response, indicating acceptable stability. Meantime, the selectivity of such sensor for H_2O_2 was evaluated against other interferences including ascorbic acid, purine trione, glucose and mixed metal ions (contained Pb²⁺, Cu²⁺, K⁺, Ca²⁺ and Mg²⁺). From the results in Fig. 5, it could be seen that the current intensity change (Δ I) upon addition of ascorbic acid (150 nM), purine trione (150 nM), glucose (150 nM) and metal ions (150 nM of Pb²⁺, Cu²⁺, K⁺, Ca²⁺ and Mg²⁺) were much lower than that of H_2O_2 (1.5 nM). Moreover, the Δ I of the sensor after the incubation of it with a solution containing both H_2O_2 and the interferences was almost the same with that of H_2O_2 (1.5 nM) only. Such high selectivity could be attributed to the highly specific cleavage of DNA based on the Fenton reaction in the presence of H_2O_2 and Fe²⁺.

 H_2O_2 is used as a stabilizer in milk in some European and American countries. However, residual H_2O_2 may cause adverse effects to the human body. Thus, the sensitive and rapid detection of H_2O_2 in milk samples is of



Figure 4. (A) DPV experiments for H_2O_2 detection in 10 mM of tris-buffer solution (pH 7.4); (B) Calibration curve for the sensor.



Figure 5. Selectivity of the sensor for H_2O_2 (1.5 nM) detection against interferences including ascorbic acid (150 nM), purine trione (150 nM), glucose (150 nM) and metal ions (150 nM).

Sample	Added (nM)	Found (nM)	Recovery %	RSD %
	0.10	0.11	110	2.06
Milk	0.30	0.29	96.7	1.38
	0.50	0.48	96.0	3.16
	1.00	1.09	109	2.12

Table 1. Recovery experiments for H₂O₂ in milk samples.

great importance. To evaluate the robustness of the system for H_2O_2 detection in milk, milk samples were collected and centrifuged at 13600 rpm to remove fat. After corroborating the absence of hydrogen peroxide, such milk samples were diluted ten-fold with tris-buffer (pH 7.4) for recovery experiments. As shown in Table 1, a good recovery ranging from 96.0% to 109% and a relative standard deviation (RSD) between 1.38% and 3.16% were obtained (each sample was tested 3 times). This result indicated that our sensor has potential application for the quantitative determination of H_2O_2 level in real samples.

In conclusion, a novel and label-free "on-off-on" switch system was successfully developed for the sensitive detection of H_2O_2 . The first "switch on" state was achieved based on the immobilization of Nf@MB and (+) AuNPs. Here, MB was used as the current signal and (+)AuNPs were used to enlarge the surface of the electrode and improve the sensitivity of the sensor. Then, the "switch off" state was obtained based on the formation of a massive hydrophobic layer that contained G-quadruplex structures and iridium(III) complex on the electrode surface. After the incubation of the sensor with H_2O_2 , CP would be cleaved into DNA fragments and released from the electrode surface, followed by signal recovering as the second "switch on" state. The novel concept of this "on-off-on" platform was successfully used for H_2O_2 detection with a detection limit down to 3.2 pM. Moreover, experiments proved that such strategy could be applied effectively for H_2O_2 detection in milk samples.

Methods

Materials and reagents. Nafion, Methylene blue (MB), rhodamine, thiazole orange (TO), 6-mercapto-1-hexanol (MCH) and gold chloride (HAuCl₄) were purchased from Sigma Aldrich (St. Louis, MO). The milk sample (Hokkaido Specially Select 3.6 Milk, UHT Processed whole milk, fat: 4.2 g per 100mL, carbohydrates: 4.8 g per 100 mL, sodium: 41 mg per 100 mL, calcium: 115 mg per 100 mL) was purchased from supermarket in Hong Kong. The iridium(III) complex used in the experiment that specific to G-quadruplex DNA was selected and synthesized according to our previous method⁵⁸. Other chemicals and solvents were of analytical grade and double distilled water was used throughout this study. 10.0 mM of tris-buffer (pH 7.4) was used for H₂O₂ detection and 10.0 mM of tris-buffer (pH 7.4, with 100 mM K⁺) was used for G-quadruplex formation. The capture probe (CP) was designed according to the literature based on the principle that there are preferential sequences for iron-mediated DNA cleavage. CP was synthesized by Techdragon Inc. (Hong Kong, China) with the sequence of 5'-GTG₃TAG₃CG₃T₂G₂-(CH₂)₆-SH-3'.

Apparatus. Electrochemical measurements were monitored by a CHI 630C electrochemical workstation (CH Instruments, Inc. U.S.A.). A conventional three-electrode system consisted of a modified working electrode, a platinum wire counter electrode and an Ag/AgCl reference electrode was used in the experiment. CD spectros-copy measurements were constructed by using an Olis 17/UV/VIS/NIR spectropolarimeter at room temperature.

H₂O₂ sensing protocol. Gold electrodes were pretreated according to our previous method. Then, 0.5 mg of MB was added into 2 mL of 0.5% Nf ethanol solution and sonicated to obtain a homogeneous suspension contained Nf@MB. Next 5µL of Nf@MB was cast on the pretreated electrode and dried in the air to obtain an Nf@MB film. After that, 4 nm of (+)AuNPs prepared according to literature⁵⁹ was self-assembly onto the Nf@ MB film for 30 min, based on the electrostatic adsorption to enlarge the electrode surface for CP immobilization, Subsequently, the modified electrodes were incubated in 10 mM of tris-buffer (pH 7.4) that contained 2.0µM of CP for 12 h at room temperature (prior to modification, the disulfide bond at the 3' end of CP was cleaved with tris(2-carboxyethyl)phosphine (TCEP)), followed by reaction with 2.0µM of MCH for 1 h to block nonspecific sites, and incubated in 10 mM of tris-buffer (pH 7.4) that contained 3.0µM of iridium(III) complex for another 1 h to induce the formation of a hydrophobic layer on the electrode surface. Then, the prepared sensor was incubated with different concentrations of H₂O₂ at 37 °C for 30 min in the presence of Fe²⁺. Finally, the electrochemical characteristics of the sensor were investigated in tris-buffer by using differential pulse experiments (DPV) from -500 mV to 0.0 mV at room temperature.

References

- Baghayeri, M., Zare, E. N. & Lakouraj, M. M. A simple hydrogen peroxide biosensor based on a novel electro-magnetic poly(pphenylenediamine)@Fe₃O₄ nanocomposite. *Biosens. Bioelectron.* 55, 259–265 (2014).
- 2. Chen, X. et al. Advances in enzyme-free electrochemical sensors for hydrogen peroxide, glucose, and uric acid. Microchim. Acta 181, 689–705 (2014).
- 3. Terai, T. & Nagano, T. Small-molecule fluorophores and fluorescent probes for bioimaging. Arch. Eur. Phys. 465, 347-359 (2013).
- Liu, Q. et al. NiO nanoparticles modified with 5,10,15,20-tetrakis(4-carboxyl pheyl)-porphyrin: Promising peroxidase mimetics for H₂O₂ and glucose detection. Biosens. Bioelectron. 64, 147–153 (2014).
- Zhang, Y. et al. Fabrication of 2D ordered mesoporous carbon nitride and its use as electrochemical sensing platform for H₂O₂, nitrobenzene, and NADH detection. Biosens. Bioelectron. 53, 250–256 (2014).
- 6. Lin, S. *et al.* Luminescence switch-on detection of protein tyrosine kinase-7 using a G-quadruplex-selective probe. *Chem. Sci.* **6**, 4284–4290 (2015).
- Wu, S. H. et al. Amplified electrochemical hydrogen peroxide reduction based on hemin/G-quadruplex DNAzyme as electrocatalyst at gold particles modified heated copper disk electrode. Biosens. Bioelectron. 73, 41–46 (2015).
- Bortolozzi, R., Gradowsk, S. & Ihmels, H. Selective ratiometric detection of H₂O₂ in water and in living cells with boronobenzo[b] quinolizinium derivatives. *Chem. Commun.* 50, 8242–8245 (2014).
- Marks, P. et al. Highly efficient detection of hydrogen peroxide in solution and in the vapor phase via fluorescence quenching. Chem. Commun. 51, 7061–7064 (2015).
- Chang, H. C. & Ho, J. A. Gold nanocluster-assisted fluorescent detection for hydrogen peroxide and cholesterol based on the inner filter effect of gold nanoparticles. *Anal. Chem.* 87, 10362–10367 (2015).
- 11. Ge, S. G. *et al.* Colorimetric detection of the flux of hydrogen peroxide released from living cells based on the high peroxidase-like catalytic performance of porous PtPd nanorods. *Biosens. Bioelectron.* **71**, 456–462 (2015).
- 12. Chen, S. *et al. In situ* growth of silver nanoparticles on graphene quantum dots for ultrasensitive colorimetric detection of H₂O₂ and glucose. *Anal. Chem.* **86**, 6689–6694 (2014).
- 13. Wang, G. L. *et al.* Ultrasensitive and dual functional colorimetric sensors for mercury(II) ions and hydrogen peroxide based on catalytic reduction property of silver nanoparticles. *Biosens. Bioelectron.* **31**, 337–342 (2012).
- 14. Privett, B. J., Shin, J. H. & Schoenfisch, M. H. Electrochemical sensors. Anal. Chem. 82, 4723-4241 (2010).
- 15. Zhang, B. *et al.* Anodic-stripping voltammetric immunoassay for ultrasensitive detection of low-abundance proteins using quantum dot aggregated hollow microspheres. *Chem. Eur. J.* **19**, 2496–2503 (2013).
- Gao, Z. et al. High-resolution colorimetric assay for rapid visual readout of phosphatase activity based on gold/silver core/shell nanorod. ACS Appl. Mater. Interfaces, 6, 18243–18250 (2014).
- 17. Imlay, J. & Linn, S. DNA damage and oxygen radical toxicity. Science 240, 1302–1309 (1988).
- Imiay, J., Chin, S. & Linn, S. Toxic DNA damage by hydrogen peroxide through the Fenton reaction *in vivo* and *in vitro*. Science 240, 640–642 (1988).
- Li, H. & Rothberg, L. Colorimetric detection of DNA sequences based on electrostatic interactions with unmodified gold nanoparticles. Proc. Natl. Acad. Sci. USA 101, 14036–14039 (2004).
- Huang, W. T. et al. A simple and facile strategy based on Fenton-induced DNA cleavage for fluorescent turn-on detection of hydroxyl radicals and Fe²⁺. J. Mater. Chem. 22, 1477–1481 (2012).
- Zhang, L. P. et al. Fenton reaction-triggered colorimetric detection of phenols in water samples using unmodified gold nanoparticles. Sens. Actuat. B 225, 593–599 (2016).
- Hu, R. et al. An efficient fluorescent sensing platform for biomolecules based on fenton reaction triggered molecular beacon cleavage strategy. Biosens. Bioelectron. 41, 442–445 (2013).
- Moye, A. L. et al. Telomeric G-quadruplexes are a substrate and site of localization for human telomerase. Nat. Commun. 6, 7643, doi: 10.1038 /ncomms8643 (2015).

- 24. Aizen, R. et al. G-Quadruplex-Stimulated optical and electrocatalytic DNA Switches. Small 11, 3654–3658 (2015).
- Hao, Y. L. et al. Amplified colorimetric detection of mercuric ions through autonomous assembly of G-quadruplex DNAzyme nanowires. Biosens. Bioelectron. 52, 261–264 (2014).
- Xu, L. J. et al. Turn-on and label-free fluorescence detection of lead ions based on target-induced G-quadruplex formation. Chem. Commun. 51, 8165–8168 (2015).
- Liu, M. et al. A SERS/fluorescence dual-mode nanosensor based on the human telomeric G-quadruplex DNA: Application to mercury(II) detection. Biosens. Bioelectron. 69, 142–147 (2015).
- Li, Y. et al. A novel electrochemical DNA biosensor based on HRP-mimicking hemin/G-quadruplex wrapped GOx nanocomposites as tag for detection of *Escherichia coli* O157:H7. *Biosens. Bioelectron.* 63, 1–6 (2015).
- Shi, L. J. et al. A label-free hemin/G-quadruplex DNAzyme biosensor developed on electrochemically modified electrodes for detection of a HBV DNA segment. RSC Adv. 5, 11541–11548 (2015).
- 30. Chen, Q. G. *et al.* An enzyme-free and label-free fluorescent biosensor for small molecules by G-quadruplex based hybridization chain reaction. *Talanta* **138**, 15–19 (2015).
- Xue, Q. W., Wang, L. & Jiang W. A novel label-free cascade amplification strategy based on dumbbell probe-mediated rolling circle amplification-responsive G-quadruplex formation for highly sensitive and selective detection of NAD⁺ or ATP. Chem. Commun. 49, 2640–2642 (2013).
- 32. Lin, Y. P. et al. Non-enzymatic sensing of hydrogen peroxide using a glassy carbon electrode modified with a nanocomposite made from carbon nanotubes and molybdenum disulphide. *Microchim. Acta* 182, 1803–1809 (2015).
- 33. He, H. Z. et al. G-quadruplexes for luminescent sensing and logic gates. Nucleic Acids Res. 41, 4345–4359 (2013).
- 34. Ma, D. et al. Simple DNA-based logic gates responding to biomolecules and metal ions. Chem. Sci. 4, 3366–3380 (2013).
- 35. Leung, C. et al. Luminescent detection of DNA-binding proteins. Nucleic Acids Res. 40, doi:10.1093 (2012).
- 36. Wang, M. *et al.* Conjugating a groove-binding motif to Ir(III) complex for the enhancement of G-quadruplex probe behavior. *Chem. Sci.* **7**, 2516–2523 (2016).
- Ma, D. et al. Group 9 organometallic compounds for therapeutic and bioanalytical applications. Acc. Chem. Res. 47, 3614–3631 (2014).
- Leung, K. H. et al. Label-free luminescence switch-on detection of hepatitis C virus NS3 helicase activity using a G-quadruplexselective probe. Chem. Sci. 6, 2166–2171 (2015).
- 39. Lu, L. H. *et al.* Detection of nicking endonuclease activity using a G-quadruplex-selective luminescent switch-on probe. *Chem. Sci.* 5, 4561–4568 (2014).
- Liu, J. et al. Iridium(III) complex-coated nanosystem for ratiometric upconversion luminescence bioimaging of cyanide anions. J. Am. Chem. Soc. 133, 15276–15279 (2011).
- 41. Yu, M. *et al.* Cationic iridium(III) complexes for phosphorescence staining in the cytoplasm of living cells. *Chem. Commun.* 2115–2117 (2008).
- Ma, D. L. et al. A luminescent cocaine detection platform using a split G-Quadruplex-selective iridium(III) complex and a three-Way DNA junction architecture. ACS Appl. Mater. Interfaces 7, 19060–19067 (2015).
- Xiong, C. *et al.* Sensitive detection of H₂O₂ and H₂O₂-related reactant with Ru(bipy)₂(7,8-dimethyl-dipyridophenazine)²⁺ and oligodeoxyribonucleotide. *Analyst* 137, 4428–4434 (2012).
- 44. Rai, P., Wemmer, D. E. & Linn, S. Preferential binding and structural distortion by Fe²⁺ at RGGG-containing DNA sequences correlates with enhanced oxidative cleavage at such sequences. *Nucleic Acids Res.* 33, 497–510 (2005).
- Tuukkanen, S. et al. Dielectrophoresis of nanoscale double-stranded DNA and humidity effects on its electrical conductivity. Appl. Phys. Lett. 87, 183102, doi: 10.1063/1.2117626 (2005).
- Cai, L. T., Tabata, H. & Kawai, T. Self-assembled DNA networks and their electrical conductivity. *Appl. Phys. Lett.* 77, 3105–3106 (2000).
- 47. Wang, M. *et al.* Label-free luminescent detection of LMP1 gene deletion using an intermolecular G-quadruplex-based switch-on probe. *Biosens. Bioelectron.* **70**, 338–344 (2015).
- Lu, L. H. et al. Label-Free luminescent switch-on probe for ochratoxin a detection using a G-quadruplex-selective iridium(III) complex. ACS Appl. Mater. Interfaces 7, 8313–8318 (2015).
- Zhu, J., Zhang, L. & Wang, E. Measurement of the base number of DNA using a special calliper made of a split G-quadruplex. Chem. Commun. 48, 11990–11992 (2012).
- 50. Guo, S., Xu, L. & Xu B. A ternary nanocomposite electrode of polyoxometalate/carbon nanotubes/gold nanoparticles for electrochemical detection of hydrogen peroxide. *Analyst* **140**, 820–826 (2015).
- 51. Zhang, Y., Liu, Y. & He, J. Electrochemical behavior of graphene/Nafion/Azure I/Au nanoparticles composites modified glass carbon electrode and its application as nonenzymatic hydrogen peroxide sensor. *Electrochim. Acta* **90**, 550–555 (2013).
- Hong, J., Yang, W. & Zhao, Y. Catalase immobilized on a functionalized multi-walled carbon nanotubes–gold nanocomposite as a highly sensitive bio-sensing system for detection of hydrogen peroxide. *Electrochim. Acta* 89, 317–325 (2013).
- Huang, K., Niu, D. & Liu, X. Direct electrochemistry of catalase at amine-functionalized graphene/gold nanoparticles composite film for hydrogen peroxide sensor. *Electrochim. Acta* 56, 2947–2953 (2011).
- Roushani, M., Bakyas, K. & Dizajdizi, B. Development of sensitive amperometric hydrogen peroxide sensor using a CuNPs/MB/ MWCNT-C₆₀-Cs-IL nanocomposite modified glassy carbon electrode. *Mat. Sci. Eng.* 64, 54–60 (2016).
- 55. Gu, T. & Hasebe, Y. Peroxidase and methylene blue-incorporated double stranded DNA-polyamine complex membrane for electrochemical sensing of hydrogen peroxide. *Anal. Chim. Acta* **525**, 191–198 (2004).
- Lin, X., Chen, J. & Chen, Z. Amperometric biosensor for hydrogen peroxide based on immobilization of horseradish peroxidase on methylene blue modified graphite electrode. *Electroanal.* 12, 306–310 (2000).
- Lin, K., Yin, C. & Chen, S. An electrochemical biosensor for determination of hydrogen peroxide using nanocomposite of poly(methylene blue) and FAD hybrid film. Sens. Actuat. B 157, 202–210 (2011).
- Ma, D. L. et al. A luminescent cocaine detection platform using a split G-Quadruplex-selective iridium(III) complex and a three-way DNA junction architecture. ACS Appl. Mater. Interfaces 7, 19060–19067 (2015).
- Li, Z. B. et al. Enhanced electrochemical recognition of double-stranded DNA by using hybridization chain reaction and positively charged gold nanoparticles. Biosens. Bioelectron. 74, 687–690 (2015).

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Author Contributions

X.-M.M. and C.Y. carried out the experiments, performed the data analysis and wrote the manuscript. C.-H.L. and D.-L.M. designed the experiments and analyzed the results.

Additional Information

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